

One Model to Rule Them All?

Modelling approaches across OneHealth, for human, animal and plant epidemics.

Adam Kleczkowski¹, Andy Hoyle² and Paul McMenemy²

¹ Department of Mathematics and Statistics, University of Strathclyde G1 1XH, UK

² Computing Science and Mathematics, University of Stirling FK9 4LA, UK

Keywords

Infectious disease; OneHealth; compartmental models; bioeconomic models; plant pathogens; epidemiological data;

Abstract

One hundred years after the 1918 influenza outbreak, are we ready for the next pandemic? This paper addresses the need to identify and develop collaborative, interdisciplinary, and cross-sectoral approaches to modelling of infectious diseases including the fields of, not only human and veterinary medicine, but also plant epidemiology. Firstly, the paper explains the concepts on which the most common epidemiological modelling approaches are based: namely the division of a host population into Susceptible, Infected and Removed (SIR) classes, and the proportionality of the infection rate to the size of the susceptible and infected populations. It then demonstrates how these simple concepts have been developed into a vast and successful modelling framework which has been utilised in predicting and controlling disease outbreaks for over one hundred years. Secondly, it considers the compartmental models based on the SIR paradigm within the broader concept of a 'disease tetrahedron' (comprised of Host, Pathogen, Environment and Man) and uses it to review the similarities and differences amongst the fields comprising the 'OneHealth' approach. Finally, the paper advocates interactions between all fields and explores the future challenges facing modellers.

Introduction

Infectious diseases of humans, animals and plants have adversely affected humanity throughout recorded history. This was recognised over 3000 years ago in the Bible, which identified threats to human, animal and plant health: "Cursed shall be the fruit of thy body [human], and the fruit of thy land [plant], the increase of thy kine (cows) [animals], and the flocks of thy sheep [animals]." (Deuteronomy 28:18, King James Version).

Please cite as: Kleczkowski A, Hoyle A & McMenemy P (2019) One model to rule them all? Modelling approaches across OneHealth for human, animal and plant epidemics. *Philosophical Transactions B: Biological Sciences*, 374 (1775), Art. No.: 20180255.

In the wake of World War I, the 1918 influenza epidemic caused approximately 50 million deaths and was one of the biggest pandemics in the history of humankind (1). Twenty five years later, while another World War was raging, a 'perfect storm' (2) of environmental and political disasters led to a series of disease outbreaks in India. In October 1942, a fungal infection, *Cochliobolus miyabeanus* (Brown Spot), spread through rice fields in Bengal. The impact of the disease was intensified by tropical storms which widely distributed the fungal spores, causing estimated yield losses of up to 91% of the key staple crop, rice (3). The local population faced severe food shortages which were further magnified by the concurrent political situation, leading to massive starvation and a decrease in resistance to diseases. Meanwhile, the weather also created conditions conducive to mosquito breeding leading to an outbreak of malaria, causing twice as many cases in 1943 than those reported in the preceding years (4). In combination with other human diseases like cholera and smallpox, which also thrived in an already affected population, an estimated 2-3 million people died as a result. More suffering and displacement followed in what became known as the Bengal Famine.

These events, as well as other similar epidemics such as Asian flu (5), Black Death (6), Foot-and-mouth (7) and Irish potato famine (8), clearly demonstrate the interconnectedness between the health of humans, animals, plants and ecosystems: the underlying principle of the OneHealth concept (9).

If we are to avoid future outbreaks such as the 1918 influenza pandemic or the 1942-43 Bengal famine, we need to create an approach that is collaborative, interdisciplinary and cross-sectoral, which includes human and veterinary medicine, plant epidemiology, as well as mathematical modelling. Mathematical models have extensively been used to understand, predict and control disease outbreaks with many successes (10,11). However, a continuing need exists to facilitate the collaboration between the modellers working across these different fields, enabling knowledge exchange and the development of approaches that take into account more than one system.

This paper attempts to facilitate this dialogue by reviewing the ways in which different research communities apply the compartmental models based on the Susceptible-Infected-Removed (*SIR*) paradigm. Our approach then embeds the *SIR* framework in the even broader concept of a 'disease tetrahedron', originating in plant epidemiology. By doing so, we show how the *SIR* model family can be adapted to address particular modelling needs which are both specific to the individual areas and generic across all fields of epidemiology. Finally, we look at some future directions of further model development.

This review is necessarily brief, and further detail is available from many books and papers that illustrate these developments, from a historical point of view and from mathematical and epidemiological angles. For medical epidemiology, the textbook of Anderson & May (10) is still one of the best options, with Keeling & Rohani (12) providing a more recent overview. Anderson & May (13) and Kao (14) contain additional references for animal diseases, and Madden et al. (11) and Gilligan (15) for compartmental models of plant pests. The three volumes of the 1993 programme at the Isaac Newton Institute, Cambridge, UK (16,17,18), and the follow up in *Epidemics* a decade later (19), also provide a valuable overview of past, current and future challenges.

Key components of an epidemic

For both an outbreak to be initiated and the pathogen to spread, a number of conditions need to be fulfilled. Van der Plank (20) and Zadoks & Schein (21) summarised these conditions in the form of a disease tetrahedron, with corners comprised of Host, Pathogen, Environment, and Man (Fig. 1). Firstly, the host and the pathogen need to be in a state that is conducive to the disease and able to interact at the same time and in the same location (Fig. 1; thick arrows). The host and pathogen states are, in turn, affected by the environmental drivers (Fig. 1; thin arrows). All three elements are influenced by human actions, including prevention, interference and control (Fig. 1; broken arrows).

The model

Compartmental models provide a widely used framework for describing the interaction between host and pathogen. The key feature of the generic *SIR* model is that the population of N individuals is split into three classes: susceptible (S), infected and infectious (I), and recovered or removed (R), Fig. 2. This reflects the assumption that there are no internal dynamics in the status of an infected individual and in its capacity to pass on the infection (a microparasite infection as defined in (10)). The models based on this paradigm may have different mathematical structures and we review some here, starting with a continuous-time and deterministic model,

$$\begin{aligned}
 \frac{dS}{dt} &= b(S, I, R) - f_p(S, V) - f_s(S, I)I - d_s S \\
 \frac{dI}{dt} &= f_p(S, V) + f_s(S, I)I - gI - d_I I \\
 \frac{dR}{dt} &= gI - d_R R.
 \end{aligned} \tag{1}$$

Above, $b(S, I, R)$ represents the influx into the susceptible class, either from births or from a transition back from the removed class, minus potential vaccination. The rates, d_x (with $x = S, I, R$), quantify removal either from natural or disease-induced causes; g represents the removal of infected individuals into the R class which can be interpreted as removed, recovered and immune. The term $b(S, I, R)$ is often constant, linear, or logistic in its dependence on the population variables; the death and removal terms are typically linear, corresponding to exponentially distributed life span and infectious periods (22). However, gamma-distributed infectious periods have been found to characterise the epidemiology of a number of diseases more accurately (23,24). The two functions describing the primary or vector-based infection, $f_p(S, V)$, and the secondary or direct infection, $f_s(S, I)$, are discussed below.

The basic SIR model has been modified to suit many different systems. Other compartments, such as a latent class (infectious but asymptomatic) or a detected class (25,26,10), have been proposed to provide more fine-grained division. Age-structured models have also been introduced, either by utilising sub-classes for the susceptible and infected individuals (27) and using a Who-Acquired-Infection-From-Whom matrix (10); or by adapting equations (1) into integro-differential equations by separating time and age variables (28).

Other extensions include difference, rather than differential, equations which use discrete time steps, rather than continuous time, with time interval often equal to the infectious period (29). Discrete time is often assumed for stochastic and network models, and for the purpose of parameter estimation (30).

Different stochastic approaches have existed since the early days of mathematical epidemiology (31,32), and their correspondence to the deterministic case was put on firm footing by Kurtz and others (33,34). Variation in the infection rate can also be used to account for environmental and demographic stochasticity (35,36).

Hosts and pathogens

Before the model can be used to describe a real-life system, its key elements must be populated. The three host groups (humans, animals and plants) have different requirements and hence require different approaches while preserving the basic structure of the SIR family of models.

Hosts. In medical epidemiology, there is only one primary host: the human being. This assumption, however, hides a possibility of alternative hosts and living stages of a pathogen, as it is now recognised that many human diseases are of zoonotic origin (37). Veterinary epidemiology deals with two types of hosts: domestic (livestock and companion animals) and wild animals, with only a few species classed as important from a societal standpoint. The situation is very different for plants, where there exists a large number of domesticated and wild plant genera (both often in proximity to each other). Despite this, most models consider only a single host species.

The basic modelling unit is typically a single individual (human, animal or plant), although in the continuous approach summarised in equations (1), S and I are often densities per unit area. For animal diseases (14), and even more often for plant diseases (11), S and I can represent groups of individuals such as flocks or fields (14).

Pathogens. The *SIR* paradigm describes the dynamics of microparasites, as defined by Anderson & May (10). An alternative framework describing macroparasitic infection, where an individual load of infection is important, has attracted significant attention in medical and veterinary epidemiology (10,38), but less so for plant diseases (39); these are not covered in this review.

Many human diseases (e.g., Ebola (40)) and animal diseases (e.g., bovine tuberculosis (41)) involve alternative hosts whose dynamics could be incorporated into the models. Malaria and dengue are examples of vector-borne diseases of humans (10,42), whereas bluetongue is an example of a vector-borne disease of animals (43). Others, such as a measles virus (44), are confined to one host and directly transmitted. Few human and animal fungal disease outbreaks have been described by the *SIR* paradigm (45,46); in contrast, the UK Plant Health Risk Register (47) currently lists 180 fungal pathogens and related species, but only 57 bacteria and 116 viruses. It is the sheer number of plant pathogens, many of which have a broad range of hosts, e.g., *Xylella* spp. (48), which makes their impact difficult to model. Despite this, compartmental models have been successfully used to describe the spread and control of plant and forest pathogens (11,49,50).

Host-pathogen axis

The functions $f_p(S, V)$ and $f_s(S, I)$ capture different aspects of host-pathogen interactions. The term $f_p(S, V)$ describes an external source of infection, with V representing the external force of infection coming either from the wider environment (e.g., primary inoculum for soil- or water-borne plant diseases (39)), from alternative hosts (e.g., bats (37)), or an indirect transmission mediated by vectors (e.g., mosquitoes (29)). There will

often be a separate equation describing the dynamics of V and its link to the infectious population, I (13).

The $f_s(S, I)$ term describes the direct transmission component that depends upon the number of currently infected individuals. Different forms have been proposed which attempt to capture various aspects of the interaction. The most common approach (51,52) assumes that the rate at which new infections are produced is proportional to (i) the number of existing susceptible individuals, $S(t)$; (ii) the proportion of existing infectious individuals, $I(t)/N$; (iii) a factor capturing the rate of contacts between individuals, $C(N)$, which is often dependent on the total population size, N ; and (iv) a per-contact probability of infection, β . This results in

$$f_s(S, I)I = \beta C(N) \frac{I}{N} S, \quad (2)$$

with $C(N) = N$ for density-dependent transmission, and $C(N)$ independent of N for the frequency-dependent transmission (27,53,49).

The simplest SIR model removes the birth, death and primary infection terms from equations (1), and assumes a fixed population size, N , and a linear secondary infection term, equation (2). The resultant model describes the characteristic rise-and-fall, bell-shaped curve for $I(t)$ often seen in medical and veterinary science (44,54). When there is no discerning recovery or removal, the basic SIR model with frequency-dependent transmission can be rewritten as an SI model equivalent to a logistic growth model; this is often used in plant epidemiology (55).

The effective and basic reproduction rates

Another way of applying an SIR model uses the idea of an effective (or apparent) reproduction rate, R_t (20). It is defined as the number of newly infected individuals produced by a single infected individual during its infectious period, at a given time t during the epidemic (56). The rate R_t typically decreases as the susceptible individuals are removed from the population; its maximum value is denoted by R_0 and forms one of the most important concepts in epidemiology (10,51). Surprisingly, for the 1918 influenza pandemic, the R_0 value estimate was only in the range of 2-3 (57), but it can be much higher for other diseases, e.g. 10-18 for measles (10).

The main advantage of characterising an epidemic by the basic reproduction rate lies in simplicity and predictive power of R_0 . The number of infected individuals will never increase if $R_0 < 1$, and so the outbreak can be prevented or stopped by lowering the value of R_0 . This condition is the foundation of many successful vaccination and control programmes (58). The threshold dependence of the disease dynamics is well known in demography and ecology and has been applied to vector-borne diseases (29), to directly transmitted human diseases (54), and to plant-pathogen systems (20,59). The pathogen's evolutionary history has also been linked to the value of the reproduction rate (60).

Parameter estimation

Although qualitative results, like the threshold behaviour, can be obtained from studying the mathematical structure of the *SIR* equations, any potential to predict the size of the new epidemic, or impact of control measures, requires estimation of parameters (61). In practice, it is often necessary to attempt to construct a predictive model based on insufficient and noisy data which limits the capacity of models to capture all details of the real world processes (14). Among the parameters, the infection functions, $f_p(S, V)$ and $f_s(S, I)$ are most difficult to identify as they attempt to capture many complex processes (62). Two broad approaches can be distinguished, with estimation based directly upon the outbreak data using different proxies (56) including observed cases (63); or through secondary data, e.g., serological studies for estimating the proportion of individuals having a contact with disease in the past (64,65), or sociological studies yielding the contact patterns between individuals (66). The two methods often produce different results, as the observation of cases can be biased by under-reporting (30).

Different statistical and modelling techniques (62) can be employed to estimate parameters in presence of considerable within- and between-sample variation and to augment data, and can reveal different aspects of the disease dynamics (cf. (55) for least-squares method, (32) for a maximum likelihood method, and (67) for a full Bayesian treatment of the same data set).

Environment: temporal heterogeneity

The functions $f_p(S, V)$ and $f_s(S, I)$ can depend on temperature, humidity and hence on time (35), thus making R_0 temporally variable (65,68). This variation can be short-term (e.g., weather-driven) or long-term (e.g., climate-driven). Models have successfully been used to study the effect of environment on cholera outbreaks in humans (69,70). The periodicity of school holidays has been shown to lead to complex dynamics in, for example,

measles epidemics (71). Less attention has been given to short-term weather effects on animal diseases (65). Plant pathogens often have complex life cycles driven by environmental conditions (e.g., temperature, rainfall); this can be captured in compartmental models by varying rates in space and time (72,73).

The effects of climate change can be captured by variations in compartmental model parameters on a time-scale longer than the epidemic itself (74,69). Vector-borne diseases have received particular attention due to their dependence on insect populations which are characterised by narrower ranges of environmental factors that are conducive to their survival and spread (75).

Environment: spatial heterogeneity

Basic compartmental models effectively assume that each susceptible individual can be equally affected by any infected individual in the population, a property often called 'homogeneous mixing' (51). This is often not true as some individuals can be separated by great distance. While humans and animals can travel large distances, their movements are often geographically and socially stratified, concentrated around schools, work and transportation routes. This is even more important for plants which generally do not move (if we ignore trade), although the pathogens and their spores can travel large distances, either from wind or vectors. Mathematical epidemiologists have approached this problem in different ways, with some approaches more common in different areas.

Some plant epidemiologists (20,11) concentrate on independent pathogen movement (e.g., spores transported by wind or water) and thus convert the *SIR* differential equations into integro-differential ones (11), allowing transmission rates to be dependent on spatial distance (76).

Alternatively, by assuming that susceptible and/or infected individuals move randomly, the *SIR* model turns into reaction-diffusion equations, such as those describing spread of rabies (77) or plant pathogens (78). Again, stochastic versions of the model have been developed, dealing with some artefacts of the continuous model (79).

Another solution, a metapopulation model, splits a geographic space into sub-compartments then assumes that individuals fully mix within each compartment. The additional force of infection comes from other compartments, either from the nearest neighbours or with some distance-dependence (80,73).

In the early 1980s a new paradigm was proposed, first by physicists (81,82) and later by epidemiologists (83). Individuals are represented by vertices of a graph (network), and transmission is only possible if two vertices are connected by an edge. These network *SIR*

models are currently at the forefront of epidemiological modelling for human diseases (12), animal diseases (84) and plant pathogens (85,86,87).

In the late 1990s, an approximation to network models was developed which captured some of the contact structure by measuring not only the average values of epidemiological variables (first moments), but also correlations between them (higher moments) (88). This leads to modified *SIR* equations which may be easier to analyse than the spatially-structured models.

Adding a human factor

The simple model shown above, equations (1) and (2), effectively ignores various ways in which our actions affect the infection dynamics and, conversely, the impact of disease on our behaviour. However, these feedbacks are an essential factor determining our ability to predict and control outbreaks (7,89,90). Accordingly, models are continually being adapted to include the impact of human influence on disease transmission. Firstly, different groups of individuals can behave in different ways and hence experience different forces of infection. Thus, the division into *SIR* compartments can be expanded to capture different behaviours, such as hospitalisation or avoidance of contacts (possibly due to fear or spread of rumours) (91). The *SIR* compartments can also be subdivided into components representing location, social status or different risk groups (e.g., for sexually transmitted diseases). The infection term, here $f_s(S, I)$, can be modified accordingly through the inclusion of, for example, a simple spatial structure (92), a city-satellites model (30) or various household models (93).

Secondly, the assumption that the combined infection term, $f_s(S, I)I$, is linear in the number of susceptible and infected individuals can be relaxed (27). The power function is often used in this context (94,95,96), e.g., to account for the heterogeneity of contacts between hosts and was found to capture the dynamics of measles (94) and Ebola (95).

Conclusions: from genes to the globe

So, are we prepared for the next pandemic (97)? Like the 1918 influenza, and many other outbreaks, any future epidemics would most likely be associated with a 'perfect storm' of events. A combination of genetic (host and pathogen), environmental, economic and socio-political factors must be studied to understand how a pathogen can emerge and spread globally (9).

The world is now faced with new challenges which can combine again into a 'perfect storm' associated with issues such as climate change, the rise of antimicrobial resistance (AMR), and globalisation which allows hosts and pathogens to travel long distances quickly.

The call to epidemiology is to bridge all scales, from a genetic to a global level. The catalyst for this interdisciplinary approach is the application of mathematical modelling. Here we have argued that compartmental models can be, and indeed have been, used successfully to capture the complexities of the host-pathogen-environment interactions. However, we still need to improve our ability to work across different disciplines and fields comprising the OneHealth approach.

Three major opportunities are identified here as immediate areas for development. Firstly, we are now equipped with an unprecedented capacity to gather and analyse 'big data'. Advances in data collection and computing power allow modellers to carry out simulations in which agents are traced as they move and interact, as vectors fly, or spores move with the wind. These individual-based models have been used, e.g., to predict the threat posed by influenza in SE Asia (98), and the spread of sudden oak death in the USA (73). Much attention in recent years has been given to social networks which explicitly incorporate the granularity of individual interactions and the heterogeneity of their behaviour, including animal and plant movements by trade (80,84). We can now include the detailed genetic information about hosts and pathogens to augment modelling techniques to improve traceability of the progress of epidemics (99). Linking detailed weather and climate change models with epidemic models allows us to predict any future outbreaks in more detail (69).

Secondly, human, animal and plant pathogens are in a constant race with their hosts, with evolution driving the resistance to any treatments or immunological responses. AMR is a rapidly rising problem which has the potential to enable epidemics not experienced since the pre-antibiotic era (100). The environment is known to act as a 'mixing bowl' for water (containing antimicrobials) from humans, agriculture, plants and aquaculture (101). However, it is still not known to what extent AMR genes transfer between species (e.g., livestock and humans) via the environment (102). An *SIR* framework has been used to model AMR either at a host/patient level (103) or even at a cellular/within-host level (101). The novelty of the latter approach is that *S* represents the antibiotic-sensitive bacterial cells, and *I* represents the antibiotic-resistant bacterial cells ('infected' with a resistant gene). Resistance has also been a major problem in plant diseases, leading to significant advances in modelling (104); this is one area where cross-fertilisation between plant and human/animal epidemiology can be very beneficial.

Thirdly, epidemiological models increasingly need to be merged with an economic and behavioural framework (105). Epi- or bio-economic models (106) capture the individual or the corporate decision to engage in particular actions, such as changing the contact structure (107,108), engaging with prevention (e.g., vaccination) (109), or control by culling (25). Recently, it has been argued that compartmental models can be extended to explicitly include adaptive behavioural responses to disease risk (110). Thus, instead of *a priori* specifying the infection term $f_s(S, I)$ in terms of S and I , the new approach uses explicit behavioural models describing how individuals formulate predictions on the effects of their decision whether to engage in a contact that might lead to infection. This prediction then informs their decision-making process which often involves maximisation of their profit or utility (110); the infection process emerges from such considerations. Such models (108) can be used to explain the multiple waves of the 1918 influenza pandemic (111).

So, are we equipped to deal with the next pandemic? According to Blackburn et al. (97), the answer currently is a qualified “no”. However, the tools and processes may already be available to allow a more emphatic “yes” to be the answer. Our future success in preventing and combating pandemics requires close collaboration across disciplines and systems. As demonstrated by their long and successful history, the *SIR* paradigm and the disease tetrahedron concept are essential tools facilitating such an approach.

Competing interests

We have no competing interests.

Authors' Contributions

A.K. conceived the design and wrote the first draft of the document; A.H. and P.M. contributed to the text; all authors revised the manuscript.

Funding

This work was partly funded by the Tree Health and Plant Biosecurity Initiative (phase 2) grant by BBSRC, Defra, ESRC, Forestry Commission, NERC and Scottish Government, BB/L012561/1, the BBSRC project BB/M008894/1 and the NSF Grant 1414374 as part of the joint NSF-NIH-USDA Ecology and Evolution of Infectious Diseases program.

References

1. Taubenberger JK, Morens DM. 1918 Influenza: the Mother of All Pandemics. *Emerg.Infect.Dis.* 2006; 12(1): p. 15-22.doi:10.3201/eid1201.050979.
2. Godfray CH, Beddington JR, Crute IR, Lawrence D, Muir JF, Pretty J, et al. Food Security: The Challenge of Feeding 9 Billion People. *Science.* 2010 January; 327: p. 812-818.doi:10.1126/science.1185383.
3. Padmanabhan SY. The Great Bengal Famine. *Annu.Rev.Phytopathol.* 1973; 11(1): p. 11-24.doi:10.1146/annurev.py.11.090173.000303.
4. Learmonth ATA. Some Contrasts in the Regional Geography of Malaria in India and Pakistan. *Trans. (Pap.) Inst.Br.Geogr.* 1957; 23: p. 37-59.doi:10.2307/621155.
5. Dowdle WR. The origin of pandemic influenza viruses. *Science.* 1984; 223(4603): p. 1402-1403.
6. Byrne JP. *Encyclopedia of the Black Death, Volume 1: ABC-CLIO; 2012.*
7. Woods A. *A Manufactured Plague: The History of Foot-and-mouth Disease in Britain: Routledge; 2004.*
8. Salaman R. *The History and Social Influence of the Potato. 2nd ed. Cambridge: Cambridge Univ. Press; 2010.*
9. Zinsstag J, Schelling E, Waltner-Toews D, Tanner M. From “one medicine” to “one health” and systemic approaches to health and well-being. *Prev.Vet.Med.* 2011; 101(3-4): p. 148-156.doi:10.1016/j.prevetmed.2010.07.003.
10. Anderson RM, May RM. *Infectious Diseases of Humans: Dynamics and Control Oxford: Oxford University Press; 1992.*
11. Madden LV, Hughes G, van den Bosch F. *The Study of Plant Disease Epidemics: Phytopathology; 2007.*
12. Keeling MJ, Rohani P. *Modeling Infectious Diseases in Humans and Animals Princeton: Princeton University Press; 2008.*
13. Anderson RM, May RM. Population Biology of Infectious Diseases: Part I,II. *Nature.* 1979; 280: p. 361-367,455-461.
14. Kao RR. The role of mathematical modelling in the control of the 2001 FMD epidemic in the UK. *Trends Microbiol.* 2002; 10(6): p. 279-286.doi:10.1016/S0966-842X(02)02371-5.
15. Gilligan CA. An epidemiological framework for disease management. *Adv.Bot.Res.* 2002; 38: p. 1-64.
16. Mollison D. *Epidemic Models: the Structure and Relation to Data Cambridge: Cambridge University Press; 1995.*

17. Grenfell BT, Dobson AP. Ecology of Infectious Diseases in Natural Populations Cambridge: Cambridge University Press; 1995.
18. Isham V, Medley G. Models for Infectious Human Diseases: Their Structure and Relation to Data Cambridge: Cambridge University Press; 1996.
19. Lloyd-Smith JO, Mollison D, Metcalf CJ, Klepac P, Heesterbeek JAP. Challenges in Modelling Infectious Disease Dynamics: Preface. *Epidemics*. 2015; 10: p. iii-iv.[doi:10.1016/j.epidem.2015.02.001](https://doi.org/10.1016/j.epidem.2015.02.001).
20. Van der Plank JE. Plant Diseases: Epidemics and Control: Academic Press; 1963.
21. Zadoks JC, Schein RD. Epidemiology and Plant Disease Management: Oxford University Press; 1979.
22. Keeling MJ, Grenfell BT. Effect of variability in infection period on the persistence and spatial spread of infectious diseases. *Math.Biosci.* 1998; 147(2): p. 207-226.[doi:10.1016/S0025-5564\(97\)00101-6](https://doi.org/10.1016/S0025-5564(97)00101-6).
23. Wearing HJ, Rohani P, Keeling MJ. Appropriate Models for the Management of Infectious Diseases. *PLOS Medicine*. 2005; 2(7): p. e174.
24. Thompson RN, Gilligan CA, Cunniffe NJ. Detecting Presymptomatic Infection Is Necessary to Forecast Major Epidemics in the Earliest Stages of Infectious Disease Outbreaks. *PLOS Computational Biology*. 2016; 12(4): p. e1004836.
25. Kleczkowski A, Oles K, Gilligan CA. Searching for the most cost-effective strategy for controlling epidemics spreading on regular and small-world networks. *J.R.Soc. Interface*. 2011; 9(66): p. 158-169.[doi:10.1098/rsif.2011.0216](https://doi.org/10.1098/rsif.2011.0216).
26. Thompson RN, Hart WS. Effect of confusing symptoms and infectiousness on forecasting and control of Ebola outbreaks. *Clin.Infect.Dis.* 2018;; p. [doi:10.1093/cid/ciy248](https://doi.org/10.1093/cid/ciy248).
27. McCallum H, Barlow N, Hone J. How should pathogen transmission be modelled? *Trends Ecol.Evol.* 2001; 16(6): p. 295-300.[doi:10.1016/S0169-5347\(01\)02144-9](https://doi.org/10.1016/S0169-5347(01)02144-9).
28. Busenberg S, Castillo-Chavez C. A general solution of the problem of mixing of subpopulations and its application to risk- and age-structured epidemic models for the spread of AIDS. *Math.Med.Biol.* 1991; 8(1): p. 1-29.[doi:10.1093/imammb/8.1.1](https://doi.org/10.1093/imammb/8.1.1).
29. Ross R. The Prevention of Malaria. 2nd ed. London: Murray; 1911.
30. Grenfell BT, Bjoernstadt ON, Kappey JN. Travelling waves and spatial hierarchies in measles epidemics. *Nature*. 2001; 414: p. 716-723.[doi:10.1038/414716a](https://doi.org/10.1038/414716a).
31. Renshaw E. Modelling Biological Populations in Space and Time (Cambridge Studies in Mathematical Biology) Cambridge: Cambridge University Press; 1991.
32. Gibson GJ, Gilligan CA, Kleczkowski A. Predicting variability in biological control of a plant-pathogen system using stochastic controls. 1999; 266: p. 1743-1753.[doi:10.1098/rspb.1999.0841](https://doi.org/10.1098/rspb.1999.0841).

33. Kurtz TG. Limit theorems for sequences of jump Markov processes approximating ordinary differential processes. *J.Appl.Probab.Stat.* 1971; 8(2): p. 344-356.doi:10.2307/3211904.
34. Diekmann O, Heesterbeek JAP. *Mathematical Epidemiology of Infectious Diseases: Model Building, Analysis and Interpretation* Chichester: John Wiley; 2000.
35. Nisbet RM, Gurney WSC. *Modelling Fluctuating Populations*: Wiley-Blackwell; 1982.
36. Kleczkowski A, Gilligan CA. Parameter estimation and prediction for the course of a single epidemic outbreak of a plant disease. *J.R.Soc. Interface.* 2007; 4(16): p. 865--877.doi:10.1098/rsif.2007.1036.
37. Lloyd-Smith JO, George D, Pepin KM, Pitzer VE, Pulliam JR, Dobson AP, et al. Epidemic dynamics at the human-animal interface. *Science.* 2009 December; 326: p. 1362-1367.doi:10.1126/science.1177345.
38. Isham V. Stochastic models of host-macroparasite interaction. *Ann.Appl.Probab.* 1995; 5(3): p. 720-740.
39. Gilligan CA, Kleczkowski A. Population dynamics of botanical epidemics involving primary and secondary infections. *Phil.Trans.Roy.Soc.B.* 1997; 352(1353): p. 591-608.doi:10.1098/rstb.1997.0040.
40. Chowell G, Nishiura H. transmission dynamics and control of Ebola virus disease (EVD): a review. *BMC Medicine.* 2014; 12(196): p. doi:10.1186/s12916-014-0196-0.
41. Renwick AR, White PC, Bengis RG. Bovine tuberculosis in southern African wildlife: a multi-species host–pathogen system. *Epidemiol.Infect.* 2007; 135(4): p. 529-540.doi:10.1017/S0950268806007205.
42. Pandey A, Mubayi A, Medlock J. Comparing vector–host and SIR models for dengue transmission. *Math.Biosci.* 2013; 246: p. 252-259.doi:10.1016/j.mbs.2013.10.007.
43. Szmaraagd C, Wilson AJ, Carpenter S, Wood JLN, Mellor PS, Gubbins S. A modeling framework to describe the transmission of bluetongue virus within and between farms in Great Britain. *PLOS ONE.* 2009; 4(11): p. doi:10.1371/journal.pone.0007741.
44. Cliff A, Haggett P, Smallman-Raynor M. *Measles: An Historical Geography* Oxford: Blackwell; 1993.
45. Schleicher J, Conrad T, GM, Cedersund G, Guthke R, Linde J. Facing the challenges of multiscale modelling of bacterial and fungal pathogen–host interactions. *Brief.Funct. Genomics.* 2017; 16(2): p. 57-69.doi:10.1093/bfgp/elv064.
46. Fisher MC, Henk DA, Briggs CJ, Brownstein JS, Madoff LC, McCraw SL. Emerging fungal threats to animal, plant and ecosystem health. *Nature.* 2012; 484: p. 186-194.
47. Baker RHA, Anderson H, Bishop S, MacLeod A, Parkinson N, Tuffen MG. The UK Plant Health Risk Register: a tool for prioritizing actions. *Bull.OEPP.* 2014 June 26; 44(2): p. 187-194.
48. Hopkins DL. *Xylella fastidiosa*: xylem-limited bacterial pathogen of plants. *Ann.Rev.Phytopathol.*

- 1989; 27: p. 271-290.doi:10.1146/annurev.py.27.090189.001415.
49. Macpherson MF, Kleczkowski A, Healey JR, Quine CP, Hanley N. The effects of invasive pests and pathogens on strategies for forest diversification. *Ecol.Modell.* 2017; 350: p. 87-99.doi:10.1016/j.ecolmodel.2017.02.003.
 50. Cunniffe NJ, Cobb RC, Meentemeyer RK, Rizzo DM, Gilligan CA. Modeling when, where, and how to manage a forest epidemic, motivated by sudden oak death in California. *Proceedings of the National Academy of Sciences.* 2016; 113(20): p. 5640-5645.
 51. Heesterbeek JAP. A brief history of R_0 and a recipe for its calculation. *Acta Biotheor.* 2002; 50(3): p. 189-204.doi:10.1023/A:1016599411804.
 52. Serfling RE. Historical review of epidemic theory. *Human Biology.* 1952; 24(3): p. 145-166.
 53. Begon M, Bennett M, Bowers RG, French NP, Hazel SM, Turner J. A clarification of transmission terms in host-microparasite models: numbers, densities and areas. *Epidemiol.Infect.* 2002; 129(01): p. 147-153.doi:10.1017/S0950268802007148.
 54. Kermack WO, McKendrick AG. Contributions to the mathematical theory of epidemics, part 1. *Proc.R.Soc.Lond. A.* 1927; 115: p. 700-721.doi:10.1098/rspa.1927.0118.
 55. Kleczkowski A, Bailey DJ, Gilligan CA. Dynamically generated variability in plant-pathogen systems with biological control. *Proc.R.Soc.Lond.B: Biol.Sci.* 1996; 263(1371): p. 777-783.doi:10.1098/rspb.1996.0116.
 56. Marmara V, Cook A, Kleczkowski A. Estimation of force of infection based on different epidemiological proxies: 2009/2010 Influenza epidemic in Malta. *Epidemics.* 2014; 9: p. 52-61.doi:10.1016/j.epidem.2014.09.010.
 57. Mills CE, Robins JM, Lipsitch M. Transmissibility of 1918 pandemic influenza. *Nature.* 2004; 432(7019): p. 904-906.doi:10.1038/nature03063.
 58. Anderson RM, May RM. Directly transmitted infections diseases: control by vaccination. *Science.* 1982; 215(4536): p. 1053-1060.doi:10.1126/science.7063839.
 59. Jeger MJ, van den Bosch F. Threshold criteria for model plant disease epidemics. 1. Asymptotic results. *Phytopathology.* 1994; 84: p. 24-27.
 60. Nowak MA, May RM. Superinfection and the evolution of parasite virulence. *Proceedings of the Royal Society of London. Series B: Biological Sciences.* 1994; 255(1342): p. 81-89.
 61. Nsoesie EO, Brownstein JS, Ramakrishnan N, Marathe MV. A systematic review of studies on forecasting the dynamics of influenza outbreaks. *Influenza Other Respir.Viruses.* 2014; 8(3): p. 309-316.
 62. Becker NG. *Analysis of Infectious Disease Data.* 1st ed. New York: Routledge; 1989.
 63. Bobashev G, Ellner SP, Nychka D, Grenfell BT. Reconstructing susceptible and recruitment dynamics from measles epidemic data. *Mathematical Population Studies.* 2000; 8(1).

64. Gay N, Hesketh L, Morgan-Capner P, Miller E. Interpretation of serological surveillance data for measles using mathematical models: Implications for vaccine strategy. *Epidemiology and Infection*. 1995; 115(1): p. 139-156.
65. Haydon DT, Woolhouse ME, Kitching RP. An analysis of foot-and-mouth-disease epidemics in the UK. *IMA.J.Math.Appl.Med.Biol*. 1997; 14(1): p. 1-9.doi:10.1093/imammb/14.1.1.
66. Bayham J, Kuminoff NV, Gunn Q, Fenichel EP. Measured voluntary avoidance behaviour during the 2009 A/H1N1 epidemic. *Proc.R.Soc.B*. 2015; 282: p. 20150814.
67. Gibson GJ, Kleczkowski A, Gilligan CA. Bayesian analysis of botanical epidemics using stochastic compartmental models. *PNAS*. 2004; 101(33): p. 12120-12124.
68. Mordecai EA, Paaijmans KP, Johnson LR, Balzer C, Ben-Horin T, Moor E, et al. Optimal temperature for malaria transmission is dramatically lower than previously predicted. *Ecol.Lett*. 2013; 16(1): p. 22-30.
69. Lipp EK, Huq A, Colwell RR. Effects of global climate on infectious disease: the cholera model. *Clin.Microbiol.Rev*. 2002; 15(4): p. 757-770.doi:10.1128/CMR.15.4.757-770.2002.
70. Grad YH, Miller JC, Lipsitch M. Cholera modeling: challenges to quantitative analysis and predicting the impact of interventions. *Epidemiology*. 2012; 23(4): p. 523-530.doi:10.1097/EDE.0b013e3182572581.
71. Grenfell BT. Chance and Chaos in Measles Dynamics. *J.R.Stat.Soc. Series B: Stat.Methodol*. 1992; 54(2): p. 383-398.doi:10.2307/2346134.
72. Skelsey P, Dancey SR, Lees AK, Cooke DEL. Forecasting the spread of aerially transmitted crop diseases. *Plant.Pathol*. 2018; 67(4): p. 920-928.doi:10.1111/ppa.12808.
73. Meentemeyer RK, Cunniffe NJ, Cook AR, Filipe JAN, Hunter RD, Rizzo DM, et al. Epidemiological modeling of invasion in heterogeneous landscapes: spread of sudden oak death in California (1990–2030). *Ecosphere*. 2011; 2(2): p. 1-24.doi:10.1890/ES10-00192.1.
74. Pautasso M, Doring TF, Garbelotto M, Pellis L, Jeger MJ. Impacts of climate change on plant diseases—opinions and trends. *Eur.J.Plant Pathol*. 2012; 133(1): p. doi:10.1007/s10658-012-9936-1295-313.
75. Semenza JC, Suk JE. Vector-borne diseases and climate change: a European perspective. *FEMS Microbiol.Lett*. 2018; 365(2): p. doi:10.1093/femsle/fnx244.
76. Keeling MJ, Brooks SP, Gilligan CA. Using conservation of pattern to estimate spatial parameters from a single snapshot. *PNAS*. 2004; 101(24): p. 9155-9160.doi:10.1073/pnas.0400335101.
77. Murray JD, Stanley EA, Brown DL. On the spatial spread of rabies among foxes. *Proc.R.Soc.Lond.B: Biol.Sci*. 1986; 229: p. 111-150.
78. van den Bosch F, Zadoks JC, Metz JAJ. Focus Expansion in Plant Disease. I: The Constant Rate of Focus Expansion. *Phytopathology*. 1988; 78: p. 54-58.
79. Mollison D. Spatial Contact Models for Ecological and Epidemic Spread. *J.R.Stat.Soc.Ser.B*:

- Methodol. 1977; 39: p. 283-326.
80. Harwood TD, Xu X, Pautasso M, Jeger MJ, Shaw MW. Epidemiological risk assessment using linked network and grid based modelling: *Phytophthora ramorum* and *Phytophthora kernoviae* in the UK. *Ecol.Modell.* 2009; 220(23): p. 3353-3361.doi:10.1016/j.ecolmodel.2009.08.014.
 81. Grassberger P. On the critical behavior of the general epidemic process and dynamical percolation. *Math.Biosci.* 1983; 63: p. 157-172.doi:10.1016/0025-5564(82)90036-0.
 82. Newman MEJ. Spread of epidemic disease on networks. *Phys.Rev.E.* 2002; 66(1): p. 016128.doi:10.1103/PhysRevE.66.016128.
 83. Kleczkowski A, Grenfell BT. Mean-field-type equations for spread of epidemics: the 'small world' model. *Physica A.* 1999; 274(1-2): p. 355-360.doi:10.1016/S0378-4371(99)00393-3.
 84. Kao RR, Danon L, Green DM, Kiss IZ. Demographic structure and pathogen dynamics on the network of livestock movements in Great Britain. *Proc.Roy.Soc.B.* 2006; 273: p. 1999-2007.doi:10.1098/rspb.2006.3505.
 85. Jeger MJ, Pautasso M, Holdenrieder O, Shaw MW. Modelling disease spread and control in networks: implications for plant sciences. *New Phytol.* 2007; 174(2): p. 279-297.doi:10.1111/j.1469-8137.2007.02028.x.
 86. Shaw MW, Pautasso M. Networks and Plant Disease Management: Concepts and Applications. *Annu.Rev.Phytopathol.* 2014; 52(1): p. 477-493.
 87. Garrett KA, Alcalá-Briseño RI, Andersen KF, C.E. B, Choudhury RA, Fulton JC, et al. Network Analysis: A Systems Framework to Address Grand Challenges in Plant Pathology. *Annu.Rev.Phytopathol.* 2018; 56: p. 559-580.
 88. House T, Keeling MJ. Insights from unifying modern approximations to infections on networks. *J.R.Soc. Interface.* 2010;: p. doi:10.1098/rsif.2010.0179.
 89. Ferguson N. Capturing human behaviour. *Nature.* 2007; 446: p. 733.doi:10.1038/446733a.
 90. Funk S, Salathe M, Jansen VAA. Modelling the influence of human behaviour on the spread of infectious diseases: a review. *J.Roy.Soc. Interface.* 2010;: p. doi:10.1098/rsif.2010.0142.
 91. Epstein JM, Parker D, Cummings D, Hammond RA. Coupled contagion dynamics of fear and disease: mathematical and computational explorations. *PLOS ONE.* 2008; 3(12): p..doi:10.1371/journal.pone.0003955e3955.
 92. Bartlett MS. Measles Periodicity and Community Size. *J.Roy.Stat.Soc. A.* 1957; 120(1): p. 48-70.doi:10.2307/2342553.
 93. Ball F, Mollison D, Scalia-Tomba G. Epidemics with two levels of mixing. *Ann.Appl.Probab.* 1997; 7(1): p. 46-89.doi:10.1214/aoap/1034625252.
 94. Finkenstaedt BF, Grenfell BT. Time series modelling of childhood diseases: a dynamical systems approach. *J.R.Stat.Soc.Ser. C, Appl.Stat.* 2002; 49(2): p. 187-205.doi:10.1111/1467-9876.00187.

95. Chowell G, Viboud C, Hyman JM, Simonsen L. The Western Africa Ebola Virus Disease Epidemic Exhibits Both Global Exponential and Local Polynomial Growth Rates. *PLoS Curr.* 2015;; p. doi:10.1371/currents.outbreaks.8b55f4bad99ac5c5db3663e916803261.
96. Gubbins S, Gilligan C, Kleczkowski A. Population dynamics of plant-parasite interactions: thresholds for invasion. *Theor.Popul.Biol.* 2000; 57(3): p. 219-233.doi:10.1006/tpbi.1999.1441.
97. Blackburn CC, Natsios AS, Parker GW. *Global Leadership at a Crossroads: Are We prepared for the Next Pandemic?* Scowcroft Institute of International Affairs; 2018.
98. Ferguson NM, Cummings DAT, Cauchemez S, Fraser C, Riley S, Meeyai A, et al. Strategies for containing an emerging influenza pandemic in Southeast Asia. *Nature.* 2005; 437(7056): p. 209-214.doi:10.1038/nature04017.
99. Kao RR, Haydon DT, Lycett SJ, Murcia PR. Supersize me: how whole-genome sequencing and big data are transforming epidemiology. *Trends Microbiol.* 2014; 22(5): p. 282-291.doi:10.1016/j.tim.2014.02.011.
100. Ventola CL. The Antibiotic Resistance Crisis: Part 1: Causes and Threats. *P.T.* 2015; 40(4): p. 277-283.
101. Taylor NGH, Verner-Jeffreys DW, Baker-Austin C. Aquatic systems: maintaining, mixing and mobilising antimicrobial resistance? *Trends Ecol.Evol.* 2011; 26(6): p. 278-284.doi:10.1016/j.tree.2011.03.004.
102. Woolhouse M, Ward M, van Bunnik B, Farrar J. Antimicrobial resistance in humans, livestock and the wider environment. *Philos.Trans.R.Soc.Lond., B, Biol.Sci.* 2015; 370(1670): p. doi:10.1098/rstb.2014.0083.
103. Paterson I, Hoyle A, Ochoa G, Baker-Austin C, Taylor N. Optimising Antibiotic Usage to Treat Bacterial Infections. *Sci.Rep.* 2016; 6: p. 37853.doi:10.1038/srep37853.
104. van den Bosch F, Gilligan CA. Models of fungicide resistance dynamics. *Annu.Rev.Phytopathol.* 2008; 46: p. 123-147.
105. Horan RD, Fenichel EP, Wolf CA, Gramig BM. Managing Infectious Animal Disease Systems. *Annual Review of Resource Economics.* 2010; 2(1): p. 101-124.
106. Philipson T. Economic epidemiology and infectious diseases. *Handbook of Health Economics.* 2000; 1(B): p. 1761-1799.doi:10.3386/w7037.
107. Gross T, D'Lima CJD, Blasius B. Epidemic Dynamics on an Adaptive Network. *Phys.Rev.Lett.* 2006; 96(20): p. doi:10.1103/PhysRevLett.96.208701.
108. Kleczkowski A, Maharaj S, Rasmussen S, Williams L, Cairns N. Spontaneous social distancing in response to a simulated epidemic: A virtual experiment. *BMC Public Health.* 2015; 15(1): p. 973.doi:10.1186/s12889-015-2336-7.
109. Galvani AP, Reluga TC, Chapman CB. Long-standing influenza vaccination policy is in accord with individual self-interest but not with the utilitarian optimum. *PNAS.* 2007; 104(13): p. 5692-

5697.doi:10.1073/pnas.0606774104.

110. Fenichel EP, Castillo-Chavez C, Ceddia MG, Chowell G, Parra PAG, Hickling GJ, et al. Adaptive human behavior in epidemiological models. PNAS. 2011; 108(15): p. 6036-6311.doi:10.1073/pnas.1011250108.
111. Caley P, Philp DJ, McCracken K. Quantifying social distancing arising from pandemic influenza. J.Roy.Soc. Interface. 2008; 5: p. 631-639.doi:10.1098/rsif.2007.1197.

Figure captions

Fig. 1: Disease tetrahedron; modified, after Zadoks and Schein (21). Nodes correspond to the main determinants of a disease outbreak: Host, Pathogen, Environment and Man. The thick double arrow represents the host-pathogen axis. Thin solid arrows indicate the influence of environment on the host-pathogen combination. Broken arrows represent the human factor affecting all components.

Fig. 2: A basic SIR model with primary and secondary infections, births, deaths, culling and natural- and disease-induced death, as represented in equations (1). Solid arrows represent the flows of individuals between classes within the population (Susceptible, S; Infected, I; Removed/recovered, R). Dashed lines represent the dependence of the infection rate on external and internal factors. Vaccination at birth effectively removes a proportion of individuals before they enter the population.