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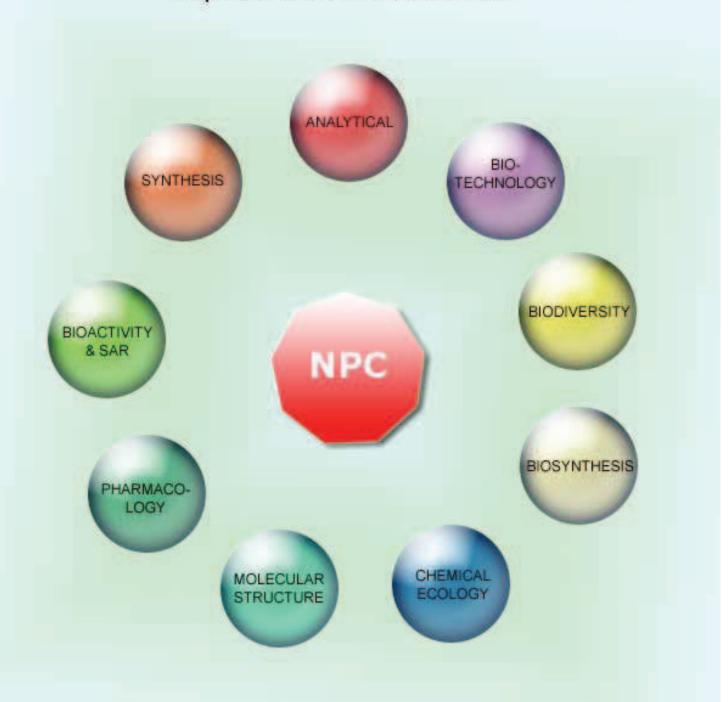
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Potential Phytopharmacy and Food Applications of *Capsicum* spp.: A Comprehensive Review

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Capsicum genus (Solanaceae) is native to the Americas. Today, it is an important agricultural crop cultivated around the world, not only due to its economic importance, but also for the nutritional value of the fruits. Among their phytochemical constituents, capsaicinoids are characteristic and responsible of the pungency of sharp-tasting cultivars. Moreover, Capsicum and capsaicinoids (mainly, capsaicin) have been largely studied because of their health benefits. Thus, this study reviews the scientific knowledge about Capsicum spp. and their phytochemicals against cancer, diabetes, gastrointestinal diseases, pain, and metabolic syndrome, as well as their antioxidant and antimicrobial activity. These bioactivities can be the basis of the formulation of functional ingredients and natural preservatives containing Capsicum extracts or isolated compounds.

Keywords: Capsicum, Capsaicin, Pepper, Metabolic syndrome, Natural preservative.

Introduction

Medicinal plants are considered as significant natural remedies for the treatment of various diseases [1-7]. Natural products of higher plants be potential provide a new source of curative agents with maybe novel mechanisms of activity [8-11].

Capsicum spp. are popular vegetables grown and consumed throughout the world [12]. They belong to the family Solanaceae that comprises 90 genera and 2000 species. This plant family is native to the Americas, and includes vegetables such as pepper, tomato, and potato [13]. In fact, chili peppers (also chile pepper, chilli pepper, or simply chilli) are the oldest crops cultivated by Native Americans between 5200 and 3400 BC [14]. Moreover, Capsicum spp. have been used in traditional medicinal practices since pre-Hispanic times from Aztecs and Mayans. Today, this genus is cultivated in various regions of the world including tropical, subtropical and temperate regions of Africa, Asia, and America, as well as the Mediterranean basin, thanks to its economic importance, and the nutritional value of the fruits (Table 1). Among

the five recognized cultivated species of this genus are: C. chinense, C. annuum, C. pubescens, C. baccatum, and C. frutescens [15, 16]. Peppers are a good source of vitamins C and E, provitamin A, carotenoids, as well as phenolic compounds; all these components are related to the total antioxidant activity of this plant food, and its bioactive properties [13]. The most characteristic phenolic derivatives found in pepper fruits is capsaicinoids (vanillylamine linked with a branched-chain fatty acid), such as capsaicin and dihydrocapsaicin (Figure 1), which are responsible for 90% of the pungency of peppers [17]. The degree of pungency depends on the Capsicum species and cultivar. The contents of these compounds may vary between undetectable in certain non-pungent cultivars to 664 mg/100 g, in pungent ones, respectively [18]. The capsinoids capsiate and dihydrocapsiate, two non-pungent analogues of capsaicin and dihydrocapsaicin respectively, have also been described in Capsicum spp. [19]. Other phenolic compounds are flavonols and flavone glycosides as well as hydroxycinnamic acids

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Some of the health benefits related to *Capsicum* ssp. consumption as part of a normal diet or related to its bioactive metabolites are shown in Figure 1. These bioactivities are reviewed here in the subsequent sections.

Table 1: Nutritional composition of *Capsicum* spp. fruit (per 100 g of edible portion) [20].

	Spices,	Peppers,	Peppers,	Peppers,
Nutrient ^a	pepper,	sweet,	hot chili,	sweet,
Nutrient	red or	green,	green,	yellow,
	cayenne ^b	raw ^c	raw ^d	raw ^c
Water (g)	8.05	93.89	87.74	92.02
Protein (g)	12.01	0.86	2.00	1.00
Energy (kcal)	318	20	40	27
Carbohydrate (g)	56.63	4.64	9.46	6.32
Calcium (mg)	148	10	18	11
Phosphorus (mg)	293	20	46	24
Selenium (µg)	8.8	0.0	0.5	0.3
Iron (mg)	7.80	0.34	1.20	4
Sodium (mg)	30	3	7	2
Copper (mg)	0.373	0.066	0.30	0.107
Potassium (mg)	2014	175	340	212
Fatty acids, total saturated (g)	3.260	0.058	0.021	0.031
Fatty acids, total monounsaturated (g)	2.750	0.008	0.011	
Total lipids (fat) (g)	17.27	0.17	0.20	0.21
Ash (g)	6.04	0.43	0.60	0.45
Niacin (mg)	8.701	0.480	0.950	0.890
Vitamin C, total ascorbic (mg)	76.4	80.4	242.5	183.5
Thiamin (mg)	0.328	0.047	0.090	0.028
Vitamin B-6 (mg)	2.450	0.224	0.278	0.168
Carotene, beta (mg)	21.84	0.208	0.671	0.120
Cryptoxanthin, beta (mg)	6.252	0.007	0.050	

^aComposition data obtained from the National Nutrient Database for Standard Reference Release 28, USDA Food Composition database (https://ndb.nal.usda.gov/ndb/; accesed 18/11/2017);

Scientific name: Capsicum frutescens or Capsicum annuum.

^dScientific name: *Capsicum annuum*. ^dScientific name: *Capsicum frutescens*.

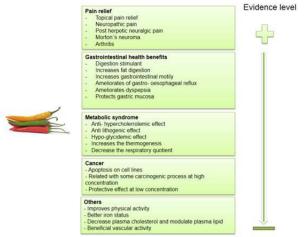


Figure 1. Example of health benefits of the consumption of Capsicum spp

Capsaicin is the most studied bioactive compound of *Capsicum*. It is considered safe in a normal daily intake. Its lethal dose (LD₅₀) estimated in humans is between 0.5–5 g capsaicin/kg (around 50,000 mg per person) [21]. However, it is known that capsaicin present in hot varieties can cause skin irritation and edema. Also, it can produce eye irritation and cause corneal lesions in rats and mice. Inhalation causes bronchoconstriction, coughing, nausea, and loss of coordination in the upper body. Exposure to capsaicinoids may result in temporary blindness, lacrimation, burning sensation, pain and skin redness, nasal irritation as well as dyspnea. These effects can be avoided using non-pungent peppers.

Cultivation of Capsicum species

Capsicum is a cool season crop, but Capsicum can be grown throughout the year under controlled temperature and moisture conditions. The average day temperature requirement is 25-30°C and night temperature 18-20°C with relative humidity of 50-60%. This crop is temperature sensitive if the temperature rises above 35°C or falls below 12°C, it will have adverse effects [22, 23]. The yield of green capsicum depends upon variety and season, generally it yields 20-40 tons per hectare in about 4-5 months, whereas in green house this yield is 80-100 tons per hectare but the duration of crop is 7-10 months [12]. For good production it is very important that the soil is properly plowed to provide a fine tilth. Raised beds of size 90-100 cm wide and 15-22 cm height are formed. Moreover, organic manure at the rate of 20-25 kg/m² must be mixed with soil before cultivation. One application is sufficient to grow three capsicum crops successively [12].

Productivity of capsicum is affected by soil solarization, organic fertilizer and endomycorrhizae [24, 25]. Proper development of plants, flowers, and fruits can be achieved by adding organic fertilizer [26]. The addition of organic fertilizer not only reduced the cost of chemical fertilizer but also resulted in the production of superior quality fruit [27, 28]. The addition of organic fertilizer in capsicum field resulted in significant increase in soil carbon, nitrogen, pH, cation exchange capacity, and exchangeable Ca, Mg, and K which invariably enhance crop yield and productivity of pepper [29]. However, combination of sunlight and organic fertilizers has positive effect on crop productivity. The pH requirement is 6-7 for getting the high production. The yield of several crops is enhanced by vesicular arbuscular mycorrhizal fungi [30-32]. They are important in ecological agriculture because of the benefits they provide to the majority of cultivars and the conservation of the environment by acting as biofertilizers, biological protectors, and biological control agents [33].

The most important issue in cultivation of *Capsicum* is the high susceptibility of the crop to a number of microorganisms, viruses and insects [34]. The common diseases of *Capsicum* are root rot, bacterial leaf spot, leaf curl, mosaic, seedling damping off and pests attacking fruits. The susceptibility of the plants varies with climate, location and time. Susceptibility of *Capsicum* and mosaic incidence reported from 70-90 %. However, leaf curl is reported to be the most devastating disease for *Capsicum*. The damage of crops due to pathogen attack can cause complete loss [35]. Hence, the areas with high rainfall and humidity are not suitable for its cultivation because they promote foliar diseases. Moreover the areas with high wind velocity are not suitable because they could enhance maintenance cost of the structure. The most recommended soil for capsicum is well drained sandy loam soil with good percolation [36].

Traditional methods are recommended to combat the sequential or simultaneous infectious diseases caused by microbes and pathogen treatment with chemicals starting with seed, soil and periodic spraying during growth, maturation and ripening [37, 38]. Identification of resistant varieties and breeding for disease-resistant varieties is in practice worldwide for *Capsicum* cultivation. The pure line progeny selection has been employed for genetic upgrading of economic attributes of *Capsicum* [35].

Phytochemical composition of Capsicum plants

Pepper (*C. annuum* L.) has been known to cure various degenerative human disorders due to the presence of bioactive compounds [39, 40]. Bell peppers (also known as sweet pepper or pepper) are a significant source of phenolic compounds such as quercetin, luteolin, and capsaicinoids [41-43]. These phenolic

compounds provide protection against cancer insurgence, diabetes, oxidative stress, cardiovascular diseases, and neural disorders such as Parkinson's and Alzheimer's disease [39, 41, 44].

C. annuum can be consumed fresh or processed, as preserves, sauces, or dehydrated powder. It is an important source of nutrients, such as provitamin A (carotene), vitamin C (ascorbic acid), chlorophyll, carotenoids, tocopherols and calcium in the human diet [45, 46]. Capsaicinoids (Figure 2) are exhibits several applications in food and pharmacy. Additional capsaicinoids consist of dihydrocapsaicin, homodihydrocapsaicin, nordihydrocapsaicin, norcapsaicin, homocapsaicin, and nornorcapsaicin Capsaicinoids are compounds characteristic in pungent pepper, which is normally consumed in South America. Nevertheless, a recent study on three different non-pungent varieties, 'Italian Sweet' (green pepper), 'Lamuyo' (yellow pepper), and 'California wonder' (red pepper) showed that the main phenolic classes are glycoside derivatives of flavonoids, such as quercetin 3,7-di-O-α-Lrhamnopyranoside and narigenin-7-O-β-D-(3"-p-coumaroyl)glucopyranoside [13] . Other compounds of non-pungent cultivars of C. annuum are capsiate, dihydrocapsiate, and nordihydrocapsiate [48] (Figure 2).

Figure 2. Examples of chemical structures of capsaicinoids and capsinoids: capsaicin (1), dihydrocapsaicin (2), capsiate (3), and dihydrocapsiate (4).

Bell pepper changes its color from green to yellow and red, while ripening. Chlorophyll and carotenoids are the major phytochemicals responsible for green color [49], while compounds such as α - and β -carotene, zeaxanthin, luteolin and β -cryptoxanthin are responsible for the yellow color in bell pepper [19]. The red color of bell pepper is mainly due to the presence of capsanthin, capsorubin and capsanthin-5,6-epoxide [50]. The concentrations of bioactive compounds in *C. annuum* is significantly affected by different maturity stages [49, 50].

There are different types of metabolites present in C. annuum such as four quercetin glycosides namely quercetin 3-O-rhamnoside-7-Oglucoside, quercetin 3-O-rhamnoside, and quercetin 3-O-glucoside-7-O-rhamnoside with rhamnoside-glucoside attached either at the C-3 or C-7 position. Furthermore, five luteolin C-glycosides, luteolin 8-C-hexoside, luteolin 6-C-hexoside, luteolin 6-Chexoside-8-C-pentoside, luteolin 6-C-pentoside-8-C-hexoside, and luteolin 6,8-di-C-hexoside along with two luteolin O-glycosides, luteolin 7-O-(2-apiosyl)-glucoside, and luteolin (apiosyl-acetyl)glucoside have also been identified in the pericarp of pepper fruits. In addition, two apigenin C-glycosides were also identified as apigenin 6-C-pentoside-8-C-hexoside and apigenin 6,8-di-Chexoside [51]. Structures of C-glycosides isolated from pepper fruit (C. annuum L., var. Capel Hot) are luteolin 6-C-glucoside, luteolin 6,8-di-C-glucoside and apigenin 6-C-glucoside-8-C-arabinoside [52]. The composition of polyphenols mainly varied with genetics, developmental stage and environmental conditions [53].

Traditional medicinal use of Capsicum genus

Native from Mexico and Central America, Capsicum spp. have been used in traditional medicine practices since pre-Hispanic times from Aztecs and Mayans, which is well documented in several codices. The most important of them is considered Libellus de Medicinabilus Indorum Herbtis (Little book of the medicinal herbs of the Indians) written by the indigenous medicine man Martin de la Cruz in 1522 [54]. At the beginning of the 20th century, studies of the botanical pharmacopeia of the indigenous Mayan inhabitants of Mesoamerica reported approximately 32 different health-related uses of Capsicum spp. [55], including treatment for arthritis, rheumatism, stomach aches, skin rashes as well as for alleviating dog and snake bites. However, the use of Capsicum fruits is not exclusively from Latin America, and their medicinal uses have been spread worldwide along with their consumption and cultivation. Hence, Capsicum fruits are mentioned in a classic text of the Tibetan medical tradition, the "Blue Beryll" to increase the digestive warmth of the stomach and as medication for the alleviation of edema, hemorrhoids, parasitic protozoa, and leprosy [56]. Furthermore, in Africa they are considered as antispasmodic, pulmonary disinfectant, counter-irritant and antitussive agents [57]. In general, Capsicum fruits are applied topically in pain disorders, neuropathy, cluster headache, migraine, psoriasis, trigeminal neuralgia and herpes zoster. Also, it has been used to treat dyspepsia, loss of appetite, flatulence, atherosclerosis, stroke, heart disease, and muscle tension [58]. Today, the fruits, fresh or dried, are consumed daily by a quarter of the global population as spices, food supplements, and additives.

Food preservative applications of plants of the genus Capsicum

Today's consumers are focused on the types of ingredients that are used in foods for flavoring, coloring and preservation [59-65]. To avoid the stigma of complex chemical names, food companies are seeking alternatives to the use of complex preservatives. Consumers want "clean labels" and products that do not contain traditional chemical preservatives.

There is an increasing trend to use natural bioingredients in food for preservation, extension of shelf-life, and microbial safety [66-70]. Spices have found application in a wide range of foods due to the extensive array of phytochemicals they possess. Ginger, allspice, pepper, nutmeg, cloves, celery, leaves, chives, and pepper are just a few spices produced through the world. Capsaicin, the major chemical compound found in pungent peppers, is reported to have antimicrobial activity against Gram-negative and Gram-positive spoilage and pathogenic bacteria [71]. As indicated elsewhere in this review, capsaicin contributes to the pungency of peppers, which can limit application in certain food systems.

The antimicrobial activity of peppers was thought to be strictly associated with capsaicin and dihydrocapsaicin, although research suggests this may not be the case [72]. Antimicrobial activity of capsaicin, dihydrocapsaicin and chrysoeriol isolated from the acetonitrile extract of C. frutescens was tested against seven microorganisms: Escherichia coli, Pseudomonas aeruginosa, Klebsiella pneumoniae, Enterococcus faecalis, Bacillus subtilis, Staphylococcus aureus, and Candida albicans. Surprisingly, under the conditions evaluated, chrysoeriol showed the greatest activity against all microorganisms evaluated with an MIC of ≤1 µg/mL [73]. In a further effort to address this issue, subfractions of crude jalapeño extract were collected and evaluated for antimicrobial activity against Listeria monocytogenes, Salmonella enterica Baildon, and E. coli O157:H7. Of the fractions collected they consistently inhibited growth of L. monocytogenes, had no effect on E. coli O157:H7 and only one fraction inhibited Salmonella [74].

The fractions that exhibited the greatest activity were matched to capsianosides, which are acyclic diterpene glycosides. These studies are not entirely contradictory and demonstrate that several compounds produced by *Capsicum* plants exhibit antimicrobial activity.

Rather than investigate the antimicrobial action of specific components, three habañero chili sauces were used directly in a well-dilution assay [75]. *E. coli, Bacillus thuringiensis, Yersinia enterocolitica*, and *Salmonella enterica* subsp. *typhimurium* were inoculated into wells containing chili sauce and Mueller-Hinton broth. At the highest concentration (1:3 dilution) of chili sauce, *Y. enterocolitica* and *S. enterica* were completely inhibited, while the growth of *E. coli* and *B. thuringiensis* was not influenced.

Research clearly suggests that various phytochemicals of peppers exhibit antimicrobial activity. The coupling of capsaicin, dihydrocapsaicin and chrysoeriol with antimicrobial compounds derived from other plants or microbiological sources may prove more effective and mitigate some negative aspects of Capsicum plant-derived compounds linked to pungency. Moreover, novel methods of food processing, including high pressure processing (HPP), pulsed light, and UV-light, are becoming more common to control pathogens. Utilization of Capsicum derived compounds with processing methods that do not influence organoleptic qualities of a food must be considered. Examples of this type of hurdle technology processing are rich in literature, but using other type of natural products. As an example, the compound nisin was used in combination with HPP to control the thermoacidophilic and sporeforming bacterium, Alicyclobacillus acidoterrestris, in apple juice [76]. The investigators demonstrated that using pressure of 200 MPa for 45 min with a nisin concentration of 250 IU/mL enabled total spore inactivation (over 6 log). Although the time requirement of the process would be prohibitive for commercial use, the study demonstrates the proof-of-concept.

A major global concern is the prevalence of antibiotic resistant foodborne pathogens. Some consumers are concerned that the use of food preservatives, sanitizers and others types of antimicrobials by the food industry contributes to the development of antibiotic resistant bacteria. Researchers investigated the synergistic effect of capsaicin and ciprofloxacin on *S. aureus* [77] showing that the MIC of ciprofloxacin was reduced 2 to 4 fold in the presence of capsaicin.

The antimicrobial action of *Capsicum* phytochemicals is typically evaluated under *in vitro* conditions. However, these studies may not translate to activity in a complex food matrix. Thus, research investigated the inhibitory effect of extracts from *Capsicum* against *S. typhimurium* and *P. aeruginosa* inoculated in minced beef. A concentration of ml/100 g of meat showed a bactericidal effect for *P. aeruginosa*, whereas mL/100g of meat inactivated *Salmonella*. Interestingly, combining the *Capsicum* extract with sodium chloride (1%, w/w) was only beneficial against *P. aeruginosa* [78]. The antimicrobial characteristics of sweet and hot pepper extracts were evaluated for controlling spoilage and pathogenic bacteria associated with Fassiekh (wet-salted fermented fish) products. The research demonstrates that extracts from hot pepper were significantly (P < 0.001) more effective in reducing the population of spoilage bacteria on Fassiekh over a 4-day period [79].

The antimicrobial mode of action of *Capsicum*-derived compounds is not clearly elucidated. Moreover, the accumulation of pyruvic acid and the reduction of ATP demonstrated that the extract influences cell membrane permeability, and bacterial respiratory

metabolism, which ultimately leads to death. Nevertheless, modeof-action studies need to be expanded to include a greater variety of microorganisms (bacteria, yeast and molds).

Complexing compounds such as capsaicin with other molecules to make more efficient delivery systems (e.g., microcapsules) may broaden food application potential [80], especially for products in which pungency is not a desirable attribute. Additionally, Capsicum-derived extracts could be used in active packaging systems. Achieving greater use of Capsicum-derived extracts for food preservation by the food industry requires greater research demonstrating efficacy in complex food systems and demonstration of cost effectiveness.

Biological activities of *Capsicum* plant extracts and their constituents: Potential as a functional ingredient

Antioxidant activity

Among Capsicum phytochemicals, carotenoids works as coloring compounds in peppers and exhibits a significant role to provide protection against cells and tissues from harmful reactive oxygen species (ROS), acting as scavengers of singlet molecular oxygen, peroxyl radicals, and reactive nitrogen species (RNS). Nonetheless, much of the total antioxidant activity of Capsicum spp. is related to its phenolic content, not only to its vitamin and carotenoid content [13]. Generally, in C. annuum, C. frutescens, and C. chinense, the concentration of antioxidants constituents (carotenoids, flavonoids, phenolic acids, and ascorbic acid) increases during fruit maturation together with the antioxidant activity determined in vitro [81]. Moreover, the antioxidant activity of Capsicum is higher compared to other vegetables. In this sense, a recent study by Morales-Soto et al. (2013) showed that, among 44 cultivars of fruits and vegetables grown in Andalusia (Spain), red 'California' pepper (4.550-13.810 mmol eq. Trolox/100 g), 'Fino' lemon (11.560-14.340 mmol eq. FeSO₄/100 g), and red onion (2.921-13.830 mmol eq. Trolox/100 g) exhibited the highest antioxidant capacity in the TEAC (trolox equivalent antioxidant capacity), FRAP (ferric ion reducing antioxidant power), and ORAC (oxygen radical absorbance capacity) assays, respectively. As a note, this red pepper is a nonpungent and its main constituents are hydroxycinnamic acids, and flavonoid glycosides [13]. However, a potential pro-oxidant effect of Capsicum flavonoids may occur in the presence of metal ions in some experiments [81]. Capsaicin has also shown an antioxidant potential comparable to that of butylhydroxyanisole (BHA), being able to protect the oxidation of human low density lipoprotein (LDL), inhibit cooper ion-induced lipid peroxidation decreasing the formation of thiobarbituric acid reactive substance (TBARS) [82]. Another study suggested that the main antioxidant compounds from red hot pepper fruit (C. annuum) are sinapoyl and feruloyl glycosides, and the main compound from green pepper is quercetin-3-O-L-rhamnoside, together with capsaicin and dihydrocapsaicin [53].

Several studies have examined the antioxidant activity of *Capsicum* spp. on human cells. In this context, lipid peroxidation induced by ascorbic acid and ferrous sulfate in erythrocyte membranes was found to be inhibited by active principles of *C. annuum*, suggesting a defensive mechanism against free radicals and a protective effect on erythrocyte integrity [83]. The phenolic glycosides isolated from *C. annuum* fruit and their aglycones were examined according to their antioxidant activities and their protective abilities against X-radiation on human cell lymphocytes, showing that phenolic glycosides present higher X-ray radioprotective activities than their aglycones [84]. Another study examined the antioxidant properties of different fraction of methanolic extract of unripe fruit part of the

C. annuum var. conoides plant [85]. This study showed a dose-responsive induction for antioxidant properties of some fractions, which may be potent natural antioxidant supplements for anti-oral cancer cell treatment through their observed effect on decreasing cell viability of human oral cancer cells (Ca9-22) [85]. Another study showed that capsaicin directly inhibits the growth of human leukemic cells through oxidative stress, involving phosphorylation of p53 at Ser-15 residue by ROS [86]. The binding and cytotoxic activities of extracted polyphenols from bioactive compounds of C. annuum correlates with their concentrations [87]. Methanolic extracts from C. annuum have been reported to inhibit 4-hydroxy-2-nonenal (HNE)-induced and H₂O₂-induced DNA damage on human leucocytes and human colorectal adenocarcinoma cells (HT-29), suggesting a role as antioxidant and cancer prevention [88].

Concerning the antioxidant activity of *Capsicum in vivo*, a recent study by Morittu *et al.* [89] evaluated the oxidative status of *C. annuum* cv. Fiesta at two ripening stage in CD-1 mice. Their results showed that, at the dose reported by the European Commission as the mean daily consumption of capsaicin, the extracts were well tolerated, independently from the ripening stage. No beneficial effect was seen in terms of antioxidant activity, but at the same time a non-significant increase of ROS production was observed. On the contrary, capsaicin may improve the antioxidant status indirectly [89]. As an example, it was evaluated against carbon tetrachloride-induced liver injury in rats treated for 8 weeks, showing a hepatoprotective effect *via:* decrease of the generation of malondialdehyde, induction of antioxidant systems (superoxide dismutase, catalase, and glutathione-*S*-transferase) and inhibition of active caspase-3 [90].

Chemopreventive activity

Capsaicin has been largely studied due to its potential to induce apoptosis in many different types of cancer cell lines including pancreatic, colonic, prostatic, liver, esophageal, bladder, skin, leukemia, lung and endothelial cells while normal cells are unharmed [91]. However, its role in tumorigenesis remains controversial because both cancer prevention and promotion have been proposed [92]. The promoter effect seems to be related to the consumption of high amounts of capsaicin in the diet. In this sense, a meta-analysis from 2014 suggested a moderate capsaicin consumption due to this dual role [93].

In general, its anticancer activity is through targeting multiple signaling pathways and cancer associated genes at different tumor stages [94]. These mechanisms have been studied *in vitro* and in animal models, as commented below.

A dose-dependent apoptotic effect of capsaicin on human pancreatic cancer cells was assessed in a study performed using in vitro and in vivo systems [95]. This study suggested the involvement of endoplasmic reticulum stress in the induction of apoptosis in human pancreatic cancer cells by its [95]. Studies performed on tumors formed by human prostate cancer cell cultures grown in mouse models reported that it is able to cause apoptosis and kill prostate cancer cells [96]. Moreover, another mechanism to induce apoptosis of human pancreatic cells is through the decrease of thioredoxin expression and dissociation of thioredoxin/apoptosis signalregulating kinase 1 (ASK1) [97]. Capsaicin reduces the viability, propagation, and induces apoptotic death in pancreatic neuroendocrine tumor (NET) cells. In addition, it also causes loss of mitochondrial membrane potential, inhibits ATP synthesis, reduces mitochondrial Bcl-2 protein production, increases cytochrome c levels, and minimizes ROS generation [98]. Another study shows

that capsaicin leads to induction of apoptosis in human lung cancer cells as well [99].

A study conducted by Amantini et al. [100] showed that administration of capsaicin prevented the proliferation of basal carcinoma (BC) cells lines via stimulation of the autophagic process, altering redox homeostasis, inducing mitochondrial depolarization, changing adenosine diphosphate/adenosine triphosphate (ADP/ATP) ratio along with activating activated protein kinase (AMPK) pathway. This enhanced capsaicin-induced cell death, demonstrating that capsaicin-induced autophagy acts as a pro-survival process in BC cells. Moreover, capsaicin-treated BC cells displayed typical mesenchymal features of the epithelial mesenchymal transition (EMT) as elongated shape and overexpression of vimentin, α5 and β1 integrin subunits, integrin-like kinase and the anti-apoptotic Bcl-2 proteins. It stimulates upregulation of Dhh/Ptch2/Zeb2 members of the Hedgehog signaling pathway, increases CD24, vascular endothelial growth factor-A (VEGFA) and tissue inhibitors of metalloproteinases-1 (TIMP-1) and decreases CD44 and activates leukocyte cell adhesion molecule (ALCAM) mRNA expression levels [100].

A case-control study conducted in Italy revealed a protective role of chili against stomach cancer [101]. Capsicum is considered a medicinal plant effective against gastric cancer in Mexico [102]. The fruit of C. annuum present elevated accumulation of arsenic, which has been reported to show anticancer activity [103]. Several studies have demonstrated dose-dependent inhibitory effects of capsaicin against stomach [104, 105] and colorectal human cancers [106]. In human gastric cell lines, capsaicin induces cell death via bcl-2-sensitive apoptotic pathway [104], overexpression of p53 and/or c-myc genes [107], activation of caspase-3 [108], and inhibits cell proliferation by cytochrome c release [105]. Moreover, capsaicin enhances the potential effectiveness of other chemotherapeutic agents on Korean human gastric cancer cell line, increasing apoptotic cell death of cisplatin-resistant gastric cancer cells after co-treatment with cisplatin [109]. In human colorectal cancer cells in vitro, capsaicin suppressed cell proliferation involving the suppression of transcriptional activity of β-catenin [106]. Moreover, capsaicin provokes apoptosis cell death on human colon cancer thought the activations of caspase-3, -8 and -9 and down-regulation of anti-apoptotic bel-2 and up-regulation of proapoptotic bax proteins [110]. Furthermore, capsaicin also inhibits human colon cancer via inducing cell cycle G0/G1 phase arrest and apoptosis, stabilizing and activating p53 [111].

Furthermore, carotenoids isolated from the fruit of red paprika *C. annuum* showed potent *in vitro* anti-tumor-promoting activity through inhibitory effects on Epstein-Barr virus early antigen activation [112]. A protein fraction containing DING proteins, which have a great biological importance due to their ability to inhibit carcinogenic cell growth, is present in *C. chinense* [113, 114]. Moreover, these proteins also inhibit the growth of several plant and human pathogenic bacteria [114].

Anti-diabetic potential

Many studies have reported both *in vitro* and *in vivo* different effects of *Capsicum* on glucose metabolism. Selected pungent and non-pungent *Capsicum* varieties have shown high antioxidant activity and an excellent inhibitory profile on carbohydrate-degrading enzymes such as α -glycosidase, which is related to glucose absorption. In this study, the most anti-diabetic potential was shown by the sweet varieties, not for the pungent ones [115]. Nevertheless, the combination of different chilis (as is habitual in the diet) seems to increase these activities [116]. Regular

consumption of hot *Capsicum* spp. may improve postprandial glucose, insulin, and energy metabolism [117]. Others studies have found that *C. chinense* (habañero) showed higher activity against α -amylase than α -glucosidase [118]. The crude extract from different varieties of *Capsicum* oleoresin showed potent antioxidant and antidiabetic activities [119]. In this study, the antidiabetic effect was assessed against α -amylase inhibitory activity [119]. *C. baccatum* seeds also inhibited mammalian α -amylase activity [120]. Another species of *Capsicum*, *C. frutescens*, presents antioxidant and antiglycation properties [121]; the accumulation of advanced glycation end products in the body is associated with diabetes mellitus.

In humans, the effect of 5 g of C. frutescens on plasma glucose level was studied in 12 healthy volunteers by performing the oral glucose tolerance test [122]. The results of this study showed that the Capsicum dose decreased plasma glucose levels at 30 and 45 minutes and, moreover, increased plasma insulin levels at 60, 75, 105, and 120 minutes. This study concludes that C. frutescens might have clinical implications in the management of type 2 diabetes mellitus decreasing plasma glucose and maintaining insulin levels [122]. Similar effects of 5 g ingestion of C. frutescens has been reported on plasma glucose in healthy Thai women [123]. Moreover, this study evaluated the effect of chili pepper on metabolic rate in 12 women, showing an immediately increase of metabolic rate after ingestion and sustained up to 30 min [123]. However, a randomized controlled study performed with 100 mL juice of C. annuum var. grossum for 4-consecutive days, on 15 subjects with type 2 diabetes mellitus under yoga therapy, showed no difference in blood glucose level (fasting or postprandial) between control and experimental groups [124]. In the case of chili, its consumption provokes a decrease of insulin resistance reflecting a postprandial homeostasis model assessment of insulin resistance in a Chinese population [125]. The supplementation of 5 mg/d of capsaicin for 4 weeks improved postprandial hyperglycemia, hyperinsulinemia and fasting lipid metabolic disorders in women with gestational diabetes mellitus [126]. Overproduction of insulin after meals could result in insulin resistance; chili intake reduces this overproduction and may reduce the risk in insulin resistance [127]. Moreover, the type of chilis may influence insulin resistance more than their quantity [124].

Gastroprotective potential: anti-ulcer and antimicrobial activity

Gastrointestinal diseases may be caused by various factors such as alcohol, acetic acid, nonsteroidal anti-inflammatory drugs (e.g., aspirin, indomethacin and ibuprofen), *Helicobacter pylori*, gastric acid, pepsin and inflammatory cytokines. Moreover, ROS are known to be involved in gastrointestinal diseases [128].

In general, spices have been recognized for their digestive stimulant action because they can intensify salivary flow and gastric juice secretion and help digestion when the nerve centers are stimulated by the sense of smell and by the presence of pungent principles such as capsaicin [129]. The hot *Capsicum* digestive stimulatory action is supposed to be related to the stimulation of saliva and bile secretion as well as digestive enzymes activities of the pancreas and small intestine. It also stimulates the saliva production and the salivary amylase activity, which helps to digest starch, as well as the mucous membrane production in mouth, throat and gastrointestinal tract. Animal studies have found that hot *Capsicum* enhances the fat digestion and absorption in high fat fed animals through the stimulation of the liver to secrete bile rich in bile acids [58].

Chili extracts, capsaicin and its other phytochemicals regulate the intestinal transport system for various food nutrients, increasing the

permeability of intestinal epithelial cells [130], and enhance the rate of gastric emptying [131]. In view of irritant and likely acidsecreting nature provoked by chili ingestion, it is being avoided in persons with ulcers. C. frutescens extract present concentrationdependent cytopathic effect on oral mucosal fibroblasts [132]. However, various studies with chili extract and capsaicin have revealed protective roles versus injurious influence on gastric mucosa [128]. Therefore, the mechanism involved in gastric mucosa protection by chili and its phytochemical constituents may involve the activation of gastrointestinal transient receptor potential vanilloid subtype 1 (TRPV1), antioxidant enzymes and inhibition of inflammatory factors [133]. Capsaicin inhibits acid secretion, prevents and heals ulcers by disposing of acid from the stomach by stimulation of alkali, mucus, bicarbonate secretions and gastric mucosal blood [134-136]. Treatment with red pepper and capsaicincontaining red pepper decreases dyspeptic symptoms, epigastric pain, fullness, nauseas and heartburn in dyspeptic and heartburn patients [137-139]. Moreover, capsaicin decreases esophageal symptoms in different gastroesophageal reflux disease patients such as Barrett's esophagus [140].

An epidemiological study, performed with 103 Chinese patients with peptic ulcer and 87 Malaysians and Indians controls, has found that chili use has a protective effect against peptic ulcer disease [141]. A prospective study of 84 healthy human subjects has evaluated the gastroprotective effect of capsaicin against two models of induced gastric mucosal damage, by ethanol or indomethacin [142]. The results of this study show that small doses of capsaicin (1-8 µg/100 mL) have gastroprotective effect against injures by ethanol or indomethacin and suggest that is attributed to stimulation of the sensory nerve endings [142]. Other study, performed on 18 healthy volunteers, reported that the administration of 20 g chili orally with 200 mL water decreases the severity of acute aspirin-induced gastroduodenal mucosal injury [143]. Moreover, capsaicin has been described as a new potential gastroprotective therapeutic drug in healthy humans, in patients with mucosal damage and in diseases requiring treatment with nonsteroidal anti-inflammatory drugs [136].

Capsaicin may induce motility in the gastric antrum, duodenum, proximal jejunum and colon which helps to diminish functional dyspepsia. However, clinical studies on its efficacy in gastro-oesophageal reflux disease and dyspepsia are limited, and there is limited information regarding the effect of chronic capsaicin ingestion in both diseases. So, a randomized double-blinded study in 30 functional dyspepsia patients using 1.75 mg of capsaicin ingestion for 5 weeks demonstrated that it significantly improved overall symptoms of epigastric pain, fullness, and nausea but not epigastric burning, bloating and belching compared to placebo [144].

Furthermore, various phytochemicals of *Capsicum* spp. present antimicrobial activity and have been described as protectors against gastric pathogens [133, 145-147]. Chili inhibited the growth of foodborne gastrointestinal pathogens such as *Salmonella typhimurium*, *Listeria monocytogenes*, *Bacillus cereus* [148], *Proteus mirabilis*, *Pseudomonas aeruginosa*, *S. aureus*, *E. coli* [133, 145, 149-151] and *Vibrio cholerae* [152, 153]. Moreover, isolated compounds from chili have been found effective against few types of pathogenic yeasts [120, 151, 154] and fungi [155, 156]. *H. pylori* is an important causal factor in gastric and duodenal ulcers and is associated with the development of gastric cancers. *In vitro* studies showed that capsaicin inhibited the growth of this pathogenic bacteria [133, 157] suggesting that chili could prove to be a therapeutic agent for *H. pylori*-induced gastrointestinal disease.

Pain relief

Natural capsaicinoids of chili peppers have received considerable attention as topical pain relievers [128, 158]. This unique property of capsaicin has been exploited by the use of creams, ointments, and patches to treat many types of pain, especially the neuropathic type [159]. Two trials performed with topical application of creams containing 0.075% [160] or 0.025% [161] of capsaicin showed analgesic effect in 21% and 70% patients with osteoarthritis and rheumatoid arthritis, respectively. In a study involving investigators of 12 sites and 219 men and women with painful diabetic neuropathy, topical application of 0.075% capsaicin reduced pain with subsequent improvement in daily activities, enhancing the quality of the patient's life [162]. The repeated application of a low dose capsaicin cream (0.075%), or a single application of a high dose patch (8%), seems to provide a degree of chronic neuropathic pain relief in adults [163, 164]. A single application of capsaicin 8% patch for 60 minutes produced effective pain relief for up to 12 weeks in patients with neuropathic pain, suggesting that highconcentration capsaicin path have the advantages of longer duration of effect and low risk for systemic effects [164]. Moreover, capsaicin creams are used to treat psoriasis, reducing inflammation and itching [165, 166]. The use of 8% capsaicin patch has demonstrated its efficient and sustained pain relief in post-herpetic neuralgia pain, a painful condition that occurs after reactivation of the dormant herpes zoster virus [167]. However, the side effects are consistently higher in all studies about neuropathic relief with capsaicin, but the significance of safety data was not quantified, and it is not clear if topical capsaicin should be used as a first-line treatment [168]. In focal pain conditions, capsaicin may be given successfully by injection to knock out nociceptors and achieve pain control in single doses applications to alleviate the pain condition for months in Morton's neuroma and arthritis, but its major weakness is the immediate burning sensation lasting minutes to hours [169].

The mechanisms of pain relief action of topical capsaicin involve the transient receptor potential cation channel subfamily V member 1 (TRPV1), also known as the capsaicin receptor and the vanilloid receptor [158, 170]. Capsaicin selectively activates TRPV1, Ca²⁺ permeable cationic ion channel which is enriched at the terminals of certain nociceptors. Activation is followed by a prolonged decrease response to noxious stimuli [169].

Potential role in the metabolic syndrome

Metabolic syndrome is referred to the clustering of insulin resistance, hypertension, dyslipidemia, and obesity, and this condition has been associated with increased risk of cardiovascular diseases and type 2 diabetes in adults. *Capsicum* spp. and their constituents may be of interest to prevent or treat some of these conditions, being revised in this section.

The risk of having metabolic syndrome is closely linked to overweight and obesity. In humans, the potential benefits of capsaicinoids for weight managements are the increase of the energy expenditure, the enhancement of lipid oxidation and the appetite reduction [171]. The number of studies on weight management is large, while the mechanism of action for appetite reduction has been less studied. In this sense, a randomized double-blind placebo-controlled study, performed with 91 moderately overweight subjects, showed that capsaicin (135 mg/day) increased fat oxidation during weight maintenance which seems to lead to a decrease in appetite as well as a decrease in food intake [172]. A study that investigated the effects of regular consumption of chili on *in vitro* serum lipoprotein oxidation in healthy adult men and women conclude that regular consumption of chili resists serum

lipoprotein oxidation [173]. Conversely, another study performed on long distance male runners 18-23 years of age reported an increased carbohydrate oxidation at rest and during exercise [174].

In studies conducted in humans it was observed that red pepper induced a reduction of ad libitum food intake and an increased postprandial energy expenditure and lipid oxidation [123, 175-178]. After a chili meal, a reduced energy expenditure was noted in adult men and women with body mass index ≥ 26 [173] and postprandial respiration quotient was reduced by 30% [179, 180]. This reduction on respiration quotient was observed in dietary consumption of chili among habitual users and non-users of chili [181]. In a study performed with 12 men and 12 women, the exposure to capsaicin, oral and gastrointestinal, increased satiety and reduced caloric as well as fat intake [182]. Moreover, oral exposure provoked a stronger reduction suggesting that capsaicin has sensory effects [182]. Capsaicin has been shown to be effective on obesity management in some short-term studies, however, the long-term use of capsaicin may be limited by its strong pungency [183]. A larger increase in energy expenditure was observed in men after a meal containing 10 g of red pepper, and the author of this study suggest that it was caused by beta-adrenergic stimulation [177]. Chili consumption increases diet-induced thermogenesis in a high fat and carbohydrate diet [179, 184], but was more pronounced in irregular consumers of chili than habitual consumers [181]. In a 2-weeks study, a supplement containing 0.4 mg capsaicin, 625 mg green tea extract and 800 mg essence of chicken reduced body fat content of free-living healthy human subjects, approximately 460 g, and increases the resting energy expenditure [185].

A possible solution to avoid the pungency of capsaicin in long-term studies is use of non-pungent analogs. The effect of CH-19 sweet pepper, a non-pungent cultivar of red pepper, was investigated on oxygen consumption and body temperature in 11 healthy volunteers. As a note, it presents capsiate, which has a structure similar to capsaicin but without pungency [186]. The results of this study showed that this type of pepper increased thermogenesis, forehead, body surface and neck temperature, and energy consumption [186]. The effects of 9 mg capsaicinoids ingestion, extracted from pepper fruit variety CH-19 sweet, were examined in a study on energy expenditure to analyze its relation on brown adipose tissue in 18 healthy men [187]. The results suggested that capsaicinoids increase energy expenditure through the activation of brown adipose tissue [187].

However, the efficacy of these bioactive compounds in weight loss is not conclusive. In fact, no solid evidence shows that capsaicin ingestion provokes weight loss, but there is a positive correlation between capsaicin and a decrease in weight regain [128]. Mean short-term studies have demonstrated an increase in diet-induced thermogenesis and a decrease in the respiratory quotient immediately after a meal in which capsaicin was supplemented (Yoshioka *et al.*, 1995). Long-term studies (three months) showed a limited effect on weight regain after a model of weight loss [188]. The effect on energy expenditure is marginal, around 50 kcal /day [171]. These results were the basis of the European Food Safety Agency report [189] to assume that the consumption of capsaicin has no effect on body weight maintenance after weight loss.

Both capsaicin and capsaicinoids are capable of increasing the whole-body energy expenditure and reduce the body fat *via* brown adipose tissue stimulation through the specific receptor TRPV1, increasing the stamina consumption and decreasing body fat modestly but consistently [190]. Besides other factors, the *trans*-differentiation of mesenchymal stem cells into adipocytes is an

added detrimental factor that may cause the intensification of obesity. Thus, Ibrahim et al. [191] have recently determined whether capsaicin is capable of inhibiting the differentiation of bone marrow mesenchymal stem cells (BMSCs) to adipocytes in vitro. BMSCs were exposed to different capsaicin concentrations for a period of 6 days following 2 days of adipogenic induction. The capsaicin exposed cells were collected at three different time points (2, 4 and 6 days) and subjected to various analyses. After capsaicin exposure, dose- and time-dependent reduction in cell viability and proliferation was observed in BMSCs. Interestingly, although capsaicin is considered an antioxidant compound, it increased the production of ROS and RNS in order to induce cell cycle arrest at G0-G1 along with increased apoptosis. Capsaicin exposure significantly inhibited the early adipogenic differentiation, lipogenesis and maturation of adipocytes with concomitant repression of PPARγ, C-enhancer-binding proteins (C/EBPα), fatty aid binding protein-4 (FABP4) and stearoyl-CoA desaturase-1 (SCD-1). Capsaicin-based stimulation for ROS and RNS production potentially inhibits the adipogenic differentiation of mesenchymal stem cells via many different pathways, i.e., anti-proliferative, apoptotic and cell cycle arrest [191].

Several studies have documented the beneficial influence of on lipid metabolism, especially capsaicin antihypercholesterolemic agent, and anti-diabetic properties, commented before. In the obese diabetic model ob/ob mice, capsaicin (0.01-0.02% in the diet for 6 weeks) significantly suppressed the enhancement in fasting blood glucose, as well as also increased the insulin levels. Moreover, capsaicin significantly increased the fecal butyrate and plasma total GLP-1 levels, but decreased plasma total ghrelin, TNF-α, IL-1β, and IL-6 levels [192]. Capsaicin also reduced body weight via activating transient receptor potential vanilloid 1 (TRPV1) cation channel, improving glucose tolerance, up-regulating the expression of Mucin 2 gene Muc2 and antimicrobial protein gene Reg3g in the intestine of C57BL/6 mice [193]. In atherosclerotic plaque mice, treatment dihydrocapsaicin supplementation momentously showed significant reduction in cellular cholesterol content, enhancement in apoA1mediated cholesterol efflux. In addition, it also lowered the low density lipoprotein, plasma triglyderides, very low density lipoprotein, interleukin-6 (IL-6), interleukin-beta (IL-1\beta), Creactive protein (CRP), and tumor necrosis factor (TNF-α) levels, whereas it enhanced high density lipoprotein [194]. Moreover, capsaicin has shown to be hypotriglyceridemic, thus preventing accumulation of fat in liver under adverse situations by enhancing triglyceride transport out of the liver also in animal models [195]. Also, it has been reported the capacity of capsaicin to reduce adiposity in rats.

Concerning cardiovascular effects, a recent study performed on a rat model of metabolic syndrome suggested that capsaicin (0.5-1 mg/kg body weight for 14 d) promoted global improvement of cardiovascular autonomic control. It determined heart variability improvement, increased vascular sympathetic drive and increased α -index (spontaneous baroreflex sensitivity). However, this treatment did not improve lipid and glucose abnormalities [196] .

Other potential health benefits

Capsaicinoids and other bioactive compounds from *Capsicum* seem to have other health benefits when assayed in animal models:

- A supplementation with capsaicin may improve physical activities, including grid, strength and endurance performance by increasing live glycogen content. Also, it can ameliorate some exercise-induced related fatigue parameters in animal studies [197].
- The presence of hot chilies in the normal diet has been related to a better iron status in the population [198].
- Capsaicinoids but not capsinoids may decrease the total plasma cholesterol, reduce the formation of atherosclerotic plaque and relax the aortic artery increasing the fecal excretion of acidic sterols in hamsters fed with diets containing capsaicinoids. Moreover, it may modulate plasma lipids and possess beneficial vascular activity [199].
- A study aimed to examine various treatments for knee osteoarthritis on older humans reported that capsaicin, orally or locally, reduces rheumatoid arthritis pain, inflammatory heat and noxious chemical hyperalgesia [200]. In human polymorphonuclear leukocytes cells it is reported that capsaicin modulate 5-lipoxygenase, the key enzyme involved in the biosynthesis of the inflammatory mediators leukotrienes [201].

Conclusions

This review highlights that Capsicum and dietary capsaicinoids have intriguing potential for health promotion. In this sense, several scientific studies have provided evidence for their beneficial effects in vitro, in vivo and in humans against cancer, diabetes, gastrointestinal diseases, and pain. Moreover, the multifunctional properties of Capsicum and capsacin seem to be promising for the prevetion/treatment of the metabolic syndrome. This includes properties, management antidiabetic hypocholesterolemic and hypotriglyceridemic activity, and beneficial cardiovascular effects. Although most of these studies are still not conclusive and only focused on capsaicin, they may be the basis of the formulation of new pharmacological active ingredients to be used in pharmaceuticals and functional foods. This also requires the definition of the dose and target population.

The application of phytochemicals derived from *Capsicum* plants for food preservation and safety is also promising. Coupling for example capsaicin and dihydrocapsaicin with non-thermal processing methods, with other natural antimicrobials or salt, may provide to consumers with safer products, which do not contain chemical preservatives. Nevertheless, the required doses, the processing time of the applied non-thermal method and the sensory properties should be evaluated in each food. This is an avenue of future research of other non-pungent *Capsicum* constituents that deserve further studies.

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References

- [1] Sharifi-Rad M, Varoni EM, Iriti M, Martorell M, Setzer WN, del Mar Contreras M, Salehi B, Soltani-Nejad A, Rajabi S, Tajbakhsh M, Sharifi-Rad J. (2018) Carvacrol and human health: A comprehensive review. *Phytotherapy Research*, 32, 1675-1687.
- [2] Salehi B, Mishra AP, Shukla I, Sharifi-Rad M, Contreras MdM, Segura-Carretero A, Fathi H, Nasrabadi NN, Kobarfard F, Sharifi-Rad J. (2018) Thymol, thyme, and other plant sources: Health and potential uses. *Phytotherapy Research*, 32, 1688-1706.
- [3] Salehi B, Anil Kumar NV, Şener B, Sharifi-Rad M, Kılıç M, Mahady GB, Vlaisavljevic S, Iriti M, Kobarfard F, Setzer WN, Ayatollahi SA, Ata A, Sharifi-Rad J. (2018) Medicinal plants used in the treatment of human immunodeficiency virus. *International Journal of Molecular Sciences*, 19(5), 1459.

- [4] Bagheri G, Mirzaei M, Mehrabi R, Sharifi-Rad J. (2016) Cytotoxic and antioxidant activities of *Alstonia scholaris*, *Alstonia venenata* and *Moringa oleifera* plants from India. *Jundishapur Journal of Natural Pharmaceutical Products*, 11(3), e31129.
- [5] Sahraie-Rad M, Izadyari A, Rakizadeh S, Sharifi-Rad J. (2015) Preparation of strong antidandruff shampoo using medicinal plant extracts: A clinical trial and chronic dandruff treatment. *Jundishapur Journal of Natural Pharmaceutical Products*, 10(4), e21517.
- [6] Stojanović-Radić Z, Pejčić M, Stojanović N, Sharifi-Rad J, Stanković N. (2016) Potential of *Ocimum basilicum* L. and *Salvia officinalis* L. essential oils against biofilms of *P. aeruginosa* clinical isolates. *Cellular and Molecular Biology*, 62, 27-32.
- [7] Mishra AP, Sharifi-Rad M, Shariati MA, Mabkhot YN, Al-Showiman SS, Rauf A, Salehi B, Župunski M, Sharifi-Rad M, Gusain P. (2018) Bioactive compounds and health benefits of edible *Rumex* species-A review. *Cellular and Molecular Biology*, 64, 27-34.
- [8] Sharifi-Rad J, Salehi B, Stojanović-Radić ZZ, Fokou PVT, Sharifi-Rad M, Mahady GB, Sharifi-Rad M, Masjedi MR, Lawal TO, Ayatollahi SA, Masjedi J, Sharifi-Rad R, Setzer WN, Sharifi-Rad M, Kobarfard F, Rahman AU, Choudhary MI, Ata A, Iriti M. (2017) Medicinal plants used in the treatment of tuberculosis Ethnobotanical and ethnopharmacological approaches. *Biotechnology Advances*, early view (https://doi.org/10.1016/j.biotechadv.2017.07.001).
- [9] Sharifi-Rad J, Fallah F, Setzer WN, Entezari Heravi R, Sharifi-Rad M. (2016) *Tordylium persicum* Boiss. & Hausskn extract: A possible alternative for treatment of pediatric infectious diseases. *Cellular and Molecular Biology*, 62, 20-26.
- [10] Sharifi-Rad J, Soufi L, Ayatollahi SAM, Iriti M, Sharifi-Rad M, Varoni EM, Shahri F, Esposito S, Kuhestani K, Sharifi-Rad M. (2016) Anti-bacterial effect of essential oil from Xanthium strumarium against shiga toxin-producing Escherichia coli. Cellular and Molecular Biology, 62, 69-74
- [11] Mishra A, Saklani S, Salehi B, Parcha V, Sharifi-Rad M, Milella L, Iriti M, Sharifi-Rad J, Srivastava M. (2018) Satyrium nepalense, a high altitude medicinal orchid of Indian Himalayan region: Chemical profile and biological activities of tuber extracts. Cellular and Molecular Biology (Noisy-le-Grand, France), 64, 35.
- [12] Hebbar SS, Balakrishna B, Prabhakar M, Srinivas V, Nair AK, Ravikumar GS, Ganeshan G, Sharma D, Sudhakar Rao DV, Doijode SD, Hegde MR. Rao MS. (2011) Protected cultivation of *Capsicum. Indian Institute of Horticultural Research, Technical Bulletin*, 22.
- [13] Morales-Soto A, Gómez-Caravaca AM, García-Salas P, Segura-Carretero A and Fernández-Gutiérrez A. (2013) High-performance liquid chromatography coupled to diode array and electrospray time-of-flight mass spectrometry detectors for a comprehensive characterization of phenolic and other polar compounds in three pepper (Capsicum annuum L.) samples. Food Research International, 51, 977-984.
- [14] Meghvansi M, Siddiqui S, Khan MH, Gupta V, Vairale M, Gogoi H, Singh L. (2010) Naga chilli: a potential source of capsaicinoids with broad-spectrum ethnopharmacological applications. *Journal of Ethnopharmacology*, 132, 1-14.
- [15] Igbokwe G, Aniakor G, Anagonye C. (2013) Determination of β-carotene & vitamin C content of fresh green pepper (*Capsicum annum*), fresh red pepper (*Capsicum annum*) and fresh tomatoes (*Solanum lycopersicum*). *Bioscientist*, 1, 89-93.
- [16] Kehie M, Kumaria S, Tandon P. (2014) Manipulation of culture strategies to enhance capsaicin biosynthesis in suspension and immobilized cell cultures of *Capsicum chinense* Jacq. cv. Naga King Chili. *Bioprocess and Biosystems Engineering*, 37, 1055-1063.
- [17] de Jesús Ornelas-Paz J, Martínez-Burrola JM, Ruiz-Cruz S, Santana-Rodríguez V, Ibarra-Junquera V, Olivas GI, Pérez-Martínez JD. (2010) Effect of cooking on the capsaicinoids and phenolics contents of Mexican peppers. *Food Chemistry*, 119, 1619-1625.
- [18] Ammar S, del Mar Contreras M, Belguith-Hadrich O, Segura-Carretero A, Bouaziz M. (2015) Assessment of the distribution of phenolic compounds and contribution to the antioxidant activity in Tunisian fig leaves, fruits, skins and pulps using mass spectrometry-based analysis. *Food & Function*, 6, 3663-3677.
- [19] Howard LR, Wildman RE. (2007) Antioxidant vitamin and phytochemical content of fresh and processed pepper fruit (*Capsicum annuum*), In: Wildman REC (Ed), *Nutraceuticals and Functional Foods*, 2nd Ed. CRC Press, Boca Raton, Florida, 165-191.
- [20] Fathima N. (2015) A Systemic Review on Phytochemistry and Pharmacological Activities of Capsicum annuum. International Journal of Pharmacy and Pharmaceutical Research, 4, 51-68.
- [21] Jang J-J, Devor D, Logsdon D, Ward J. (1992) A 4-week feeding study of ground red chilli (*Capsicum annuum*) in male B6C3F1 mice. *Food and Chemical Toxicology*, 30, 783-787.
- [22] Kurt S and Emir B. (2004) Effect of soil solarization, chicken litter and viscera on populations of soilborne fungal pathogens and pepper growth. Plant Pathology Journal, 3, 118-124.
- [23] Cimen I, Turgay B, Pirinc V. (2010) Effect of solarization and vesicular arbuscular mychorrizal on weed density and yield of lettuce (*Lactuca sativa L.*) in autumn season. *African Journal of Biotechnology*, 9, 3520-3526.
- [24] Horouwitz M, Reger Y, Herzlingar G. (1983) Solarization for weed control. *Journal of Weed Science*, 31, 170-179.
- [25] Abdallah M. (1991) Control of different weed species at different soil depth with soil solarization. *Egypt Journal of Agronomy, Special Issue*, 81-88.
- [26] da Silva TRB, Bortoluzzi T, da Silva CAT, Arieira CRD. (2012) A comparison of poultry litter applied like organic fertilizer and that applied like chemical fertilizer in corn development. *African Journal of Agricultural Research*, 7, 194-197.
- [27] Delate K, Cambardella CA. (2004) Agroecosystem performance during transition to certified organic grain production. *Agronomy Journal*, 96, 1288-1298.
- [28] Farhad W, Saleem M, Cheema M, Hammad H. (2009) Effect of poultry manure levels on the productivity of spring maize (*Zea mays L.*). *Journal of Animal & Plant Sciences*, 19, 122-125.
- [29] Zayed MS, Hassanein M, Esa NH, Abdallah M. (2013) Productivity of pepper crop (Capsicum annuum L.) as affected by organic fertilizer, soil solarization, and endomycorrhizae. Annals of Agricultural Sciences, 58, 131-137.
- [30] Thanuja T, Hegde RV, Sreenivasa M. (2002) Induction of rooting and root growth in black pepper cuttings (*Piper nigrum L.*) with the inoculation of arbuscular mycorrhizae. *Scientia Horticulturae*, 92, 339-346.
- [31] Durgapal A, Pandey A, Palni LMS. (2002) The use of rhizosphere soil for improved establishment of conifers at nursery stage for application in plantation programmes. *Journal of Sustainable Forestry*, 15, 57-73.
- [32] Smith S. (1997) Vesicular-arbuscular mycorrhizas in agriculture and horticulture, In: Smith SE, Reed DJ (Eds), *Mycorrhizal Symbiosis*, 2nd Ed. Academic Press, London, 453-469.
- [33] Azcón-Aguilar C, Jaizme-Vega M, Calvet C. (2002) The contribution of arbuscular mycorrhizal fungi to the control of soil-borne plant pathogens, In: Gianinazzi S, Schüepp H, Barea JM, Haselwandter K (Eds), *Mycorrhizal Technology in Agriculture*, Springer, Basel, Switzerland, 187-197.
- [34] Stanton WJ, Futrell CM. (1987) Fundamentals of Marketing. McGraw-Hill, New York.
- [35] Peter KV, Peethamparan CK, Gopalakrishnan PK. (1980) Breeding for disease resistance in chilli. All India Workshop on Chillies. Hyderabad, India
- [36] Singh B. (1998) Vegetable production under protected conditions: Problems and prospects. *Indian Society of Vegetable Science Souvenir: Silver Jubilee, National Symposium Dec, 12*(14).
- [37] Seetharam Raju KaSR, G. (1980) Diseases control for yield and quality of chillies in Andhra Prades. *All India Workshop on Chillies*. Hyderabad, India.
- [38] Rao M. (1980) Insect pests and their control in chillies for stepping up production. All India Workshop on Chillies. Hyderabad, India.

- [39] Blanco-Ríos AK, Medina-Juárez LÁ, González-Aguilar GA, Gámez-Meza N. (2013) Antioxidant activity of the phenolic and oily fractions of different sweet bell peppers. *Journal of the Mexican Chemical Society*, 57, 137-143.
- [40] Chen C-Y, Yeh Y-T, Yang W-L. (2011) Amides from the stem of Capsicum annuum. Natural Product Communications, 6, 227-229.
- [41] Oboh G, Rocha J. (2007) Distribution and antioxidant activity of polyphenols in ripe and unripe tree pepper (*Capsicum pubescens*). *Journal of Food Biochemistry*, 31, 456-473.
- [42] Liu Y, Nair MG. (2010) Capsaicinoids in the hottest pepper Bhut Jolokia and its antioxidant and antiinflammatory activities. *Natural Product Communications*, 5, 91-94.
- [43] Si W, Man SW, Chenb Z, Chung HY. (2014) Stability of capsaicinoid content at raised temperatures. *Natural Product Communications*, 9, 985-988.
- [44] Salehi B, Zucca P, Sharifi-Rad M, Pezzani R, Rajabi S, Setzer WN, Varoni EM, Iriti M, Kobarfard F, Sharifi-Rad J. (2018) Phytotherapeutics in cancer invasion and metastasis. *Phytotherapy Research*, early view (https://doi.org/10.1002/ptr.6087).
- [45] Shetty A, Magadum S, Managanvi K. (2013) Vegetables as sources of antioxidants. Journal of Food and Nutrition Disorders, 2(1), 1000104.
- [46] de Moraes LP, da Paz MF, Argandoña EJS, da Silva LR, de Oliveira Zago T. (2013) Compostos fenólicos e atividade antioxidante de molho de pimenta "dedo-de-moça" fermentado. BBR-Biochemistry and Biotechnology Reports, 1, 33-38.
- [47] Prasad BCN, Gururaj HB, Kumar V, Giridhar P, Ravishankar GA. (2006) Valine pathway is more crucial than phenyl propanoid pathway in regulating capsaicin biosynthesis in *Capsicum frutescens* Mill. *Journal of Agricultural and Food Chemistry*, 54, 6660-6666.
- [48] Sutoh K, Kobata K, Yazawa S, Watanabe T. (2006) Capsinoid is biosynthesized from phenylalanine and valine in a non-pungent pepper, *Capsicum annuum* L. cv. CH-19 sweet. *Bioscience, Biotechnology, and Biochemistry*, 70, 1513-1516.
- [49] Marín A, Ferreres F, Tomás-Barberán FA, Gil MI. (2004) Characterization and quantitation of antioxidant constituents of sweet pepper (*Capsicum annuum* L.). *Journal of Agricultural and Food Chemistry*, 52, 3861-3869.
- [50] Sun T, Xu Z, Wu CT, Janes M, Prinyawiwatkul W, No H. (2007) Antioxidant activities of different colored sweet bell peppers (*Capsicum annuum* L.). *Journal of Food Science*, 72, S98-S102.
- [51] Wahyuni Y, Ballester A-R, Sudarmonowati E, Bino RJ, Bovy AG. (2011) Metabolite biodiversity in pepper (*Capsicum*) fruits of thirty-two diverse accessions: Variation in health-related compounds and implications for breeding. *Phytochemistry*, 72, 1358-1370.
- [52] Materska M. (2015) Flavone C-glycosides from Capsicum annuum L.: Relationships between antioxidant activity and lipophilicity. European Food Research and Technology, 240, 549-557.
- [53] Materska M, Perucka I. (2005) Antioxidant activity of the main phenolic compounds isolated from hot pepper fruit (*Capsicum annuum L.*). *Journal of Agricultural and Food Chemistry*, 53, 1750-1756.
- [54] Domínguez F, Alonso-Castro AJ, Anaya M, González-Trujano ME, Salgado-Ceballos H, Orozco-Suárez S. (2015) Mexican traditional medicine: Traditions of yesterday and phytomedicines for tomorrow. In: Duarte MCT, Rai M (Eds), *Therapeutic Medicinal Plants: From Lab to the Market*, CRC Press, Boca Raton, Florida, 10-46.
- [55] Cichewicz RH, Thorpe PA. (1996) The antimicrobial properties of chile peppers (*Capsicum* species) and their uses in Mayan medicine. *Journal of Ethnopharmacology*, 52, 61-70.
- [56] Basu SK, De AK, De A. (2003) Capsicum: Historical and botanical perspectives. In: De AK (Ed), Capsicum: The genus Capsicum, Taylor & Francis, New York, 1-15.
- [57] Elujoba AA, Odeleye O, Ogunyemi C. (2006) Traditional medicine development for medical and dental primary health care delivery system in Africa. African Journal of Traditional, Complementary and Alternative Medicines (AJTCAM), 2, 46-61.
- [58] Maji AK, Banerji P. (2016) Phytochemistry and gastrointestinal benefits of the medicinal spice, Capsicum annuum L. (Chilli): A review. Journal of Complementary and Integrative Medicine, 13, 97-122.
- [59] Arimboor R, Natarajan RB, Menon KR, Chandrasekhar LP, Moorkoth V. (2015) Red pepper (*Capsicum annuum*) carotenoids as a source of natural food colors: Analysis and stability—a review. *Journal of Food Science and Technology*, 52, 1258-1271.
- [60] Billing J, Sherman PW. (1998) Antimicrobial functions of spices: why some like it hot. The Quarterly Review of Biology, 73, 3-49.
- [61] Sharifi-Rad J, Mnayer D, Tabanelli G, Stojanović-Radić Z, Sharifi-Rad M, Yousaf Z, Vallone L, Setzer WN, Iriti M. (2016) Plants of the genus *Allium* as antibacterial agents: From tradition to pharmacy. *Cellular and Molecular Biology (Noisy-le-Grand, France)*, 62, 57-68.
- [62] Sharifi-Rad J, Sureda A, Tenore GC, Daglia M, Sharifi-Rad M, Valussi M, Tundis R, Sharifi-Rad M, Loizzo MR, Ademiluyi AO. (2017) Biological activities of essential oils: From plant chemoecology to traditional healing systems. *Molecules*, 22, 70.
- [63] Sharifi-Rad M, Varoni EM, Salehi B, Sharifi-Rad J, Matthews KR, Ayatollahi SA, Kobarfard F, Ibrahim SA, Mnayer D, Zakaria ZA. (2017) Plants of the genus *Zingiber* as a source of bioactive phytochemicals: From tradition to pharmacy. *Molecules*, 22, 2145.
- [64] Sharifi-Rad J, Ayatollahi SA, Varoni EM, Salehi B, Kobarfard F, Sharifi-Rad M, Iriti M, Sharifi-Rad M. (2017) Chemical composition and functional properties of essential oils from *Nepeta schiraziana* Boiss. *Farmacia*, 65, 802-812.
- [65] Salehi B, Ayatollahi S, Segura-Carretero A, Kobarfard F, Contreras M, Faizi M, Sharifi-Rad M, Tabatabai S, Sharifi-Rad J. (2017) Bioactive chemical compounds in *Eremurus persicus* (Joub. & Spach) Boiss. essential oil and their health implications. *Cellular and Molecular Biology* (Noisy-le-Grand), 63, 1-7.
- [66] Sharifi-Rad J, Salehi B, Varoni EM, Sharopov F, Yousaf Z, Ayatollahi SA, Kobarfard F, Sharifi-Rad M, Afdjei MH, Sharifi-Rad M. (2017) Plants of the *Melaleuca* genus as antimicrobial agents: From farm to pharmacy. *Phytotherapy Research*, 31, 1475-1494.
- [67] De La Torre Torres, JE, Gassara F, Kouassi AP, Brar SK, Belkacemi, K. (2017) Spice use in food: Properties and benefits. Critical Reviews in Food Science and Nutrition, 57, 1078-1088.
- [68] Sharifi-Rad M, Nazaruk J, Polito L, Morais-Braga MFB, Rocha JE, Coutinho HDM, Salehi B, Tabanelli G, Montanari C, del Mar Contreras M, Yousaf Z, Setzer WN, Verma DR, Martorell M, Sureda A, Sharifi-Rad J. (2018) *Matricaria* genus as a source of antimicrobial agents: From farm to pharmacy and food applications. *Microbiological Research*, 215, 76-88.
- [69] Sharifi-Rad M, Mnayer D, Morais-Braga MFB, Carneiro JNP, Bezerra CF, Coutinho HDM, Salehi B, Martorell M, del Mar Contreras M, Soltani-Nejad A, Uribe YAH, Yousaf Z, Iriti M, Sharifi-Rad J. (2018) *Echinacea* plants as antioxidant and antibacterial agents: From traditional medicine to biotechnological applications. *Phytotherapy Research*, early view (https://doi.org/10.1002/ptr.6101).
- [70] Sharifi-Rad J, Salehi B, Schnitzler P, Ayatollahi S, Kobarfard F, Fathi M, Eisazadeh M, Sharifi-Rad M. (2017) Susceptibility of herpes simplex virus type 1 to monoterpenes thymol, carvacrol, *p*-cymene and essential oils of *Sinapis arvensis* L., *Lallemantia royleana* Benth. and *Pulicaria vulgaris* Gaertn. *Cellular and Molecular Biology (Noisy-le-Grand)*, 63, 42-47.
- [71] Gottardi D, Bukvicki D, Prasad S and Tyagi AK. (2016) Beneficial Effects of Spices in Food Preservation and Safety. Frontiers in microbiology, 7.
- [72] Dorantes L, Colmenero R, Hernandez H, Mota L, Jaramillo ME, Fernandez E, Solano C. (2000) Inhibition of growth of some foodborne pathogenic bacteria by *Capsicum annum* extracts. *International Journal of Food Microbiology*, 57, 125-128.
- [73] Nascimento PL, Nascimento TC, Ramos NS, Silva GR, Gomes JEG, Falcão RE, Moreira KA, Porto AL, Silva T. (2014) Quantification, antioxidant and antimicrobial activity of phenolics isolated from different extracts of *Capsicum frutescens* (Pimenta Malagueta). *Molecules*, 19, 5434-5447.
- [74] Bacon K, Boyer R, Denbow C, O'Keefe S, Neilson A, Williams R. (2017) Antibacterial activity of jalapeño pepper (*Capsicum annuum* var. *annuum*) extract fractions against select foodborne pathogens. *Food Science & Nutrition*, 5, 730-738.

- [75] Hleba L. (2015) Antibacterial activity of habanero chili sauces against selected pathogenic bacteria. Scientific Papers Animal Science and Biotechnologies, 48, 132-136.
- [76] Sokołowska B, Skapska S, Fonberg-Broczek M, Niezgoda J, Chotkiewicz M, Dekowska A, Rzoska S. (2012) The combined effect of high pressure and nisin or lysozyme on the inactivation of *Alicyclobacillus acidoterrestris* spores in apple juice. *High Pressure Research*, 32, 119-127.
- [77] Omolo MA, Wong Z-Z, Mergen K, Hastings JC, Le NC, Reil HA, Case KA, Baumler DJ. (2014) Antimicrobial properties of chili peppers. *Journal of Infectious Diseases and Therapy*, 2(4), 1000145
- [78] Careaga M, Fernández E, Dorantes L, Mota L, Jaramillo ME, Hernandez-Sanchez H. (2003) Antibacterial activity of *Capsicum* extract against *Salmonella typhimurium* and *Pseudomonas aeruginosa* inoculated in raw beef meat. *International Journal of Food Microbiology*, 83, 331-335.
- [79] Sulieman HA, Allaahmed AA. (2012) Effect of antimicrobial properties of pepper fruits on some spoilage organism of Sudanese wet-salted fermented fish (fassiekh) product. *World's Veterinary Journal*, 2(1), 5-10.
- [80] Xing F, Cheng G, Yi K. (2006) Study on the antimicrobial activities of the capsaicin microcapsules. *Journal of Applied Polymer Science*, 102, 1318-1321.
- [81] Howard L, Talcott S, Brenes C, Villalon B. (2000) Changes in phytochemical and antioxidant activity of selected pepper cultivars (*Capsicum* species) as influenced by maturity. *Journal of Agricultural and Food Chemistry*, 48, 1713-1720.
- [82] Naidu KA, Thippeswamy NB. (2002) Inhibition of human low density lipoprotein oxidation by active principles from spices. *Molecular and Cellular Biochemistry*, 229, 19-23.
- [83] Salimath B, Sundaresh C, Srinivas L. (1986) Dietary components inhibit lipid peroxidation in erythrocyte membrane. *Nutrition Research*, 6, 1171-1178.
- [84] Materska M, Konopacka M, Rogolinski J, Slosarek K. (2015) Antioxidant activity and protective effects against oxidative damage of human cells induced by X-radiation of phenolic glycosides isolated from pepper fruits *Capsicum annuum* L. *Food Chemistry*, 168, 546-553.
- [85] Chen CY, Yen CY, Shen GM, Yu TJ, Liao YS, Jian RI, Wang SC, Tang JY, Chang HW. (2017) Antioxidant properties of fractions for unripe fruits of Capsicum annuum L. var. conoides. Anti-Cancer Agents in Medicial Chemistry, 17, 1971-1977.
- [86] Ito K, Nakazato T, Yamato K, Miyakawa Y, Yamada T, Hozumi N, Segawa K, Ikeda Y, Kizaki M. (2004) Induction of apoptosis in leukemic cells by homovanillic acid derivative, capsaicin, through oxidative stress: implication of phosphorylation of p53 at Ser-15 residue by reactive oxygen species. *Cancer Research*, 64, 1071-1078.
- [87] Kim HG, Bae JH, Jastrzebski Z, Cherkas A, Heo BG, Gorinstein S, Ku YG. (2016) Binding, antioxidant and anti-proliferative properties of bioactive compounds of sweet paprika (Capsicum annuum L.). Plant Foods for Human Nutrition, 71, 129-136.
- [88] Park JH, Jeon GI, Kim JM, Park E. (2012) Antioxidant activity and antiproliferative action of methanol extracts of 4 different colored bell peppers (Capsicum annuum L.). Food Science and Biotechnology, 21, 543-550.
- [89] Morittu V, Musco N, Vassalotti G, Gaito S, Badia AD, Spina A, Pero M, Squillacioti C, Tundis R, Mastellone V. (2017) Effects of total extracts from *Capsicum annuum* L. fruits at two different stages of ripening on oxidative status and metabolic profile in mice. *Journal of Nutritional Ecology and Food Research*, 4, 93-95.
- [90] Hassan MH, Edfawy M, Mansour A, Hamed A-A. (2012) Antioxidant and antiapoptotic effects of capsaicin against carbon tetrachloride-induced hepatotoxicity in rats. *Toxicology and Industrial Health*, 28, 428-438.
- [91] Clark R, Lee S-H. (2016) Anticancer properties of capsaicin against human cancer. Anticancer Research, 36, 837-843.
- [92] Liu NC, Hsieh PF, Hsieh MK, Zeng ZM, Cheng HL, Liao JW, Chueh PJ. (2012) Capsaicin-mediated tNOX (ENOX2) up-regulation enhances cell proliferation and migration in vitro and in vivo. *Journal of Agricultural and Food Chemistry*, 60, 2758-2765.
- [93] Pabalan N, Jarjanazi H, Ozcelik H. (2014) The impact of capsaicin intake on risk of developing gastric cancers: a meta-analysis. *Journal of Gastrointestinal Cancer*, 45, 334-341.
- [94] Chapa-Oliver AM, Mejía-Teniente L. (2016) Capsaicin: From plants to a cancer-suppressing agent. Molecules, 21, 931.
- [95] Lin S, Zhang J, Chen H, Chen K, Lai F, Luo J, Wang Z, Bu H, Zhang R, Li H, Tong H. (2013) Involvement of endoplasmic reticulum stress in capsaicin-induced apoptosis of human pancreatic cancer cells. *Evidence-Based Complementary and Alternative Medicine*, 2013, 629750.
- [96] Mori A, Lehmann S, O'Kelly J, Kumagai T, Desmond JC, Pervan M, McBride WH, Kizaki M, Koeffler HP. (2006) Capsaicin, a component of red peppers, inhibits the growth of androgen-independent, p53 mutant prostate cancer cells. *Cancer Research*, 66, 3222-3229.
- [97] Pramanik KC, Srivastava SK. (2012) Apoptosis signal-regulating kinase 1-thioredoxin complex dissociation by capsaicin causes pancreatic tumor growth suppression by inducing apoptosis. *Antioxidants & Redox Signaling*, 17, 1417-1432.
- [98] Skrzypski M, Sassek M, Abdelmessih S, Mergler S, Grötzinger C, Metzke D, Wojciechowicz T, Nowak K, Strowski M. (2014) Capsaicin induces cytotoxicity in pancreatic neuroendocrine tumor cells via mitochondrial action. *Cellular Signalling*, 26, 41-48.
- [99] Lee SH, Krisanapun C, Baek SJ. (2010) NSAID-activated gene-1 as a molecular target for capsaicin-induced apoptosis through a novel molecular mechanism involving GSK3beta, C/EBPbeta and ATF3. *Carcinogenesis*, 31, 719-728.
- [100] Amantini C, Morelli MB, Nabissi M, Cardinali C, Santoni M, Gismondi A, Santoni G. (2016) Capsaicin triggers autophagic cell survival which drives epithelial mesenchymal transition and chemoresistance in bladder cancer cells in an Hedgehog-dependent manner. *Oncotarget*, 7, 50180.
- [101] Buiatti E, Palli D, Decarli A, Amadori D, Avellini C, Bianchi S, Biserni R, Cipriani F, Cocco P, Giacosa A. (1989) A case-control study of gastric cancer and diet in Italy. *International Journal of Cancer*, 44, 611-616.
- [102] Alonso-Castro AJ, Villarreal ML, Salazar-Olivo LA, Gomez-Sanchez M, Dominguez F, Garcia-Carranca A. (2011) Mexican medicinal plants used for cancer treatment: Pharmacological, phytochemical and ethnobotanical studies. *Journal of Ethnopharmacology*, 133, 945-972.
- [103] Rezanka T, Sigler K. (2008) Biologically active compounds of semi-metals. *Phytochemistry*, 69, 585-606.
- [104] Lo YC, Yang YC, Wu IC, Kuo FC, Liu CM, Wang HW, Kuo CH, Wu JY, Wu DC. (2005) Capsaicin-induced cell death in a human gastric adenocarcinoma cell line. World Journal of Gastroenterology, 11, 6254-6257.
- [105] Meral O, Alpay M, Kismali G, Kosova F, Cakir DU, Pekcan M, Yigit S, Sel T. (2014) Capsaicin inhibits cell proliferation by cytochrome c release in gastric cancer cells. *Tumour Biology*, 35, 6485-6492.
- [106] Lee SH, Richardson RL, Dashwood RH, Baek SJ. (2012) Capsaicin represses transcriptional activity of β-catenin in human colorectal cancer cells. *Journal of Nutritional Biochemistry*, 23, 646-655.
- [107] Kim JD, Kim JM, Pyo JO, Kim SY, Kim BS, Yu R, Han IS. (1997) Capsaicin can alter the expression of tumor forming-related genes which might be followed by induction of apoptosis of a Korean stomach cancer cell line, SNU-1. *Cancer Letters*, 120, 235-241.
- [108] Park SY, Kim JY, Lee SM, Jun CH, Cho SB, Park CH, Joo YE, Kim HS, Choi SK, Rew JS. (2014) Capsaicin induces apoptosis and modulates MAPK signaling in human gastric cancer cells. *Molecular Medicine Reports*, *9*, 499-502.
- [109] Huh HC, Lee SY, Lee SK, Park NH, Han IS. (2011) Capsaicin induces apoptosis of cisplatin-resistant stomach cancer cells by causing degradation of cisplatin-inducible Aurora-A protein. *Nutrition and Cancer*, 63, 1095-1103.
- [110] Lu HF, Chen YL, Yang JS, Yang YY, Liu JY, Hsu SC, Lai KC, Chung JG. (2010) Antitumor activity of capsaicin on human colon cancer cells in vitro and colo 205 tumor xenografts in vivo. *Journal of Agricultural and Food Chemistry*, 58, 12999-13005.
- [111] Jin J, Lin G, Huang H, Xu D, Yu H, Ma X, Zhu L, Ma D, Jiang H. (2014) Capsaicin mediates cell cycle arrest and apoptosis in human colon cancer cells via stabilizing and activating p53. *International Journal of Biolical Sciences*, 10, 285-295.

- [112] Maoka T, Mochida K, Kozuka M, Ito Y, Fujiwara Y, Hashimoto K, Enjo F, Ogata M, Nobukuni Y, Tokuda H, Nishino H. (2001) Cancer chemopreventive activity of carotenoids in the fruits of red paprika *Capsicum annuum* L. *Cancer Letters*, 172, 103-109.
- [113] Brito-Argáez L, Tamayo-Sansores JA, Madera-Piña D, García-Villalobos FJ, Moo-Puc RE, Kú-González Á, Villanueva MA, Islas-Flores I. (2016)
 Biochemical characterization and immunolocalization studies of a *Capsicum chinense* Jacq. protein fraction containing DING proteins and antimicrobial activity. *Plant Physiology and Biochemistry*, 109, 502-514.
- [114] Fernando MS, Ligia BA, Mayra DB, Ignacio IF. (2011) A review of a promising therapeutic and agronomical alternative: Antimicrobial peptides from *Capsicum* sp. *African Journal of Biotechnology*, 10, 19918-19928.
- [115] Kwon YI, Apostolidis E, Shetty K. (2007) Evaluation of pepper (Capsicum annuum) for management of diabetes and hypertension. Journal of Food Biochemistry, 31, 370-385.
- [116] Oboh G, Ademiluyi AO, Akinyemi AJ, Henle T, Saliu JA, Schwarzenbolz U. (2012) Inhibitory effect of polyphenol-rich extracts of jute leaf (*Corchorus olitorius*) on key enzyme linked to type 2 diabetes (α-amylase and α-glucosidase) and hypertension (angiotensin I converting) in vitro.

 Journal of Functional Foods, 4, 450-458.
- [117] Ahuja KD, Robertson IK, Geraghty DP, Ball MJ. (2006) Effects of chili consumption on postprandial glucose, insulin, and energy metabolism. American Journal of Clinical Nutrition, 84, 63-69.
- [118] Tundis R, Loizzo MR, Menichini F, Bonesi M, Conforti F, Statti G, De Luca D, de Cindio B, Menichini F. (2011) Comparative study on the chemical composition, antioxidant properties and hypoglycaemic activities of two Capsicum annuum L. cultivars (acuminatum small and cerasiferum). Plant Foods for Human Nutrition, 66, 261.
- [119] Sricharoen P, Lamaiphan N, Patthawaro P, Limchoowong N, Techawongstien S, Chanthai S. (2017) Phytochemicals in *Capsicum* oleoresin from different varieties of hot chilli peppers with their antidiabetic and antioxidant activities due to some phenolic compounds. *Ultrasonics Sonochemistry*, 38, 629-639.
- [120] Vieira Bard GC, Nascimento VV, Oliveira AE, Rodrigues R, Da Cunha M, Dias GB, Vasconcelos IM, Carvalho AO, Gomes VM. (2014) Vicilin-like peptides from *Capsicum baccatum* L. seeds are α-amylase inhibitors and exhibit antifungal activity against important yeasts in medical mycology. *Biopolymers*, 102, 335-343.
- [121] Khan I, Ahmad H, Ahmad B. (2014) Anti-glycation Anti-oxidation properties of *Capsicum frutescens* and *Curcuma longa* fruits: Possible role in prevention of diabetic complication. *Pakistan Journal of Pharmaceutical Sciences*, 27, 1359-1362.
- [122] Chaiyasit K, Khovidhunkit W, Wittayalertpanya S. (2009) Pharmacokinetic and the effect of capsaicin in *Capsicum frutescens* on decreasing plasma glucose level. *Journal of the Medical Association of Thailand*, 92, 108-113.
- [123] Chaiyata P, Puttadechakum S, Komindr S. (2003) Effect of chili pepper (*Capsicum frutescens*) ingestion on plasma glucose response and metabolic rate in Thai women. *Journal of the Medical Association of Thailand*, 86, 854-860.
- [124] Nagasukeerthi P, Mooventhan A, Manjunath NK. (2017) Short-term effect of add on bell pepper (*Capsicum annuum* var. *grossum*) juice with integrated approach of yoga therapy on blood glucose levels and cardiovascular functions in patients with type 2 diabetes mellitus: A randomized controlled study. *Complementary Therapies in Medicine*, 34, 42-45.
- [125] Li J, Wang R, Xiao C. (2014) Association between chilli food habits with iron status and insulin resistance in a Chinese population. *Journal of Medicinal Food*, 17, 472-478.
- [126] Yuan LJ, Qin Y, Wang L, Zeng Y, Chang H, Wang J, Wang B, Wan J, Chen SH, Zhang QY, Zhu JD, Zhou Y, Mi MT. (2016) Capsaicin-containing chili improved postprandial hyperglycemia, hyperinsulinemia, and fasting lipid disorders in women with gestational diabetes mellitus and lowered the incidence of large-for-gestational-age newborns. *Clinical Nutrition*, 35, 388-393.
- [127] Varghese S, Kubatka P, Rodrigo L, Gazdikova K, Caprnda M, Fedotova J, Zulli A, Kruzliak P, Büsselberg D. (2017) Chili pepper as a body weight-loss food. *International Journal of Food Sciences and Nutrition*, 68, 392-401.
- [128] Srinivasan K. (2016) Biological activities of red pepper (*Capsicum annuum*) and its pungent principle capsaicin: A review. *Critical Reviews in Food Science and Nutrition*, 56, 1488-1500.
- [129] Platel K, Srinivasan K. (2004) Digestive stimulant action of spices: A myth or reality? Indian Journal of Medical Research, 119, 167.
- [130] Jensen-Jarolim E, Gajdzik L, Haberl I, Kraft D, Scheiner O, Graf J. (1998) Hot spices influence permeability of human intestinal epithelial monolayers. *Journal of Nutrition*, 128, 577-581.
- [131] Debreceni A, Abdel-Salam OM, Figler M, Juricskay I, Szolcsányi J, Mózsik G. (1999) Capsaicin increases gastric emptying rate in healthy human subjects measured by ¹³C-labeled octanoic acid breath test. *Journal of Physiology-Paris*, 93, 455-460.
- [132] van Wyk CW, Olivier A, de Miranda C, van der Bijl P, Grobler-Rabie AF, Chalton DO. (1995) Effect of chilli (*Capsicum frutescens*) extract on proliferation of oral mucosal fibroblasts. *Indian Journal of Experimental Biology*, 33, 244-248.
- [133] Maji AK, Banerji P. (2016) Phytochemistry and gastrointestinal benefits of the medicinal spice, Capsicum annuum L. (chilli): A review. Journal of Complementary and Integral Medicine, 13, 97-122.
- [134] Satyanarayana MN. (2006) Capsaicin and gastric ulcers. Critical Reviews in Food Science and Nutrition, 46, 275-328.
- [135] Mózsik G, Szolcsányi J, Dömötör A. (2007) Capsaicin research as a new tool to approach of the human gastrointestinal physiology, pathology and pharmacology. *Inflammopharmacology*, 15, 232-245.
- [136] Mózsik G. (2014) Capsaicin as new orally applicable gastroprotective and therapeutic drug alone or in combination with nonsteroidal antiinflammatory drugs in healthy human subjects and in patients. *Progress in Drug Research*, 68, 209-258.
- [137] Bortolotti M, Coccia G, Grossi G, Miglioli M. (2002) The treatment of functional dyspepsia with red pepper. *Alimentary Pharmacology and Therapeutics*, 16, 1075-1082.
- [138] Chen CL, Liu TT, Yi CH, Orr WC. (2010) Effects of capsaicin-containing red pepper sauce suspension on esophageal secondary peristalsis in humans. *Neurogastroenterol Motil*, 22, 1177-1182, e1312-1173.
- [139] Liu TT, Yi CH, Lei WY, Hung XS, Yu HC, Chen CL. (2014) Influence of repeated infusion of capsaicin-contained red pepper sauce on esophageal secondary peristalsis in humans. *Neurogastroenterology & Motility*, 26, 1487-1493.
- [140] Herrera-López JA, Mejía-Rivas MA, Vargas-Vorackova F, Valdovinos-Díaz MA. (2010) Capsaicin induction of esophageal symptoms in different phenotypes of gastroesophageal reflux disease. *Revista de Gastroenterología de México*, 75, 396-404.
- [141] Kang JY, Yeoh KG, Chia HP, Lee HP, Chia YW, Guan R, Yap I. (1995) Chili--protective factor against peptic ulcer? *Digestive Diseases and Sciences*, 40, 576-579.
- [142] Mozsik G, Szolcsanyi J, Racz I. (2005) Gastroprotection induced by capsaicin in healthy human subjects. World Journal of Gastroenterology, 11, 5180-5184.
- [143] Yeoh KG, Kang JY, Yap I, Guan R, Tan CC, Wee A, Teng CH. (1995) Chili protects against aspirin-induced gastroduodenal mucosal injury in humans. *Digestive Diseases and Sciences*, 40, 580-583.
- [144] Bortolotti M, Coccia G, Grossi G, Miglioli M. (2002) The treatment of functional dyspepsia with red pepper. *Alimentary Pharmacology & Therapeutics*, 16, 1075-1082.
- [145] Khan FA, Mahmood T, Ali M, Saeed A, Maalik A. (2014) Pharmacological importance of an ethnobotanical plant: Capsicum annuum L. Natural Product Research, 28, 1267-1274.

- [146] De Marino S, Borbone N, Gala F, Zollo F, Fico G, Pagiotti R, Iorizzi M. (2006) New constituents of sweet *Capsicum annuum* L. fruits and evaluation of their biological activity. *Journal of Agricultural and Food Chemistry*, 54, 7508-7516.
- [147] Mokhtar M, Ginestra G, Youcefi F, Filocamo A, Bisignano C, Riazi A. (2017) Antimicrobial activity of selected polyphenols and capsaicinoids identified in pepper (*Capsicum annuum* L.) and their possible mode of interaction. *Current Microbiology*, 74, 1253-1260.
- [148] Careaga M, Fernandez E, Dorantes L, Mota L, Jaramillo ME, Hernandez-Sanchez H. (2003) Antibacterial activity of *Capsicum* extract against *Salmonella typhimurium* and *Pseudomonas aeruginosa* inoculated in raw beef meat. *International Journal of Food Microbiology*, 83, 331-335.
- [149] Lampe JW. (2003) Spicing up a vegetarian diet: Chemopreventive effects of phytochemicals. *American Journal of Clinical Nutrition*, 78, 579S-583S.
- [150] Adamu HM, Abayeh OJ, Agho MO, Abdullahi AL, Uba A, Dukku HU, Wufem BM. (2005) An ethnobotanical survey of Bauchi State herbal plants and their antimicrobial activity. *Journal of Ethnopharmacology*, 99, 1-4.
- [151] Taveira GB, Mathias LS, da Motta OV, Machado OL, Rodrigues R, Carvalho AO, Teixeira-Ferreira A, Perales J, Vasconcelos IM, Gomes VM. (2014) Thionin-like peptides from *Capsicum annuum* fruits with high activity against human pathogenic bacteria and yeasts. *Biopolymers*, 102, 30-39.
- [152] Chatterjee S, Asakura M, Chowdhury N, Neogi SB, Sugimoto N, Haldar S, Awasthi SP, Hinenoya A, Aoki S, Yamasaki S. (2010) Capsaicin, a potential inhibitor of cholera toxin production in *Vibrio cholerae*. *FEMS Microbiology Letters*, 306, 54-60.
- [153] Yamasaki S, Asakura M, Neogi SB, Hinenoya A, Iwaoka E, Aoki S. (2011) Inhibition of virulence potential of *Vibrio cholerae* by natural compounds. *Indian Journal of Medical Research*, 133, 232-239.
- [154] Ribeiro SF, Carvalho AO, Da Cunha M, Rodrigues R, Cruz LP, Melo VM, Vasconcelos IM, Melo EJ, Gomes VM. (2007) Isolation and characterization of novel peptides from chilli pepper seeds: antimicrobial activities against pathogenic yeasts. *Toxicon*, 50, 600-611.
- [155] Cruz LP, Ribeiro SF, Carvalho AO, Vasconcelos IM, Rodrigues R, Da Cunha M, Gomes VM. (2010) Isolation and partial characterization of a novel lipid transfer protein (LTP) and antifungal activity of peptides from chilli pepper seeds. *Protein and Peptide Letters*, 17, 311-318.
- [156] Anaya-López JL, López-Meza JE, Baizabal-Aguirre VM, Cano-Camacho H, Ochoa-Zarzosa A. (2006) Fungicidal and cytotoxic activity of a Capsicum chinense defensin expressed by endothelial cells. Biotechnology Letters, 28, 1101-1108.
- [157] Jones NL, Shabib S, Sherman PM. (1997) Capsaicin as an inhibitor of the growth of the gastric pathogen *Helicobacter pylori*. *FEMS Microbiology Letters*, 146, 223-227.
- [158] Luo XJ, Peng J, Li YJ. (2011) Recent advances in the study on capsaicinoids and capsinoids. European Journal of Pharmacology, 650, 1-7.
- [159] Derry S, Sven-Rice A, Cole P, Tan T, Moore RA. (2013) Topical capsaicin (high concentration) for chronic neuropathic pain in adults. *Cochrane Database of Systematic Reviews*, 2, CD007393.
- [160] McCarthy GM, McCarty DJ. (1992) Effect of topical capsaicin in the therapy of painful osteoarthritis of the hands. *Journal of Rheumatology*, 19, 604-607.
- [161] Deal CL, Schnitzer TJ, Lipstein E, Seibold JR, Stevens RM, Levy MD, Albert D, Renold F. (1991) Treatment of arthritis with topical capsaicin: A double-blind trial. *Clinical Therapeutics*, 13, 383-395.
- [162] Group CS. (1992) Effect of treatment with capsaicin on daily activities of patients with painful diabetic neuropathy. Diabetes Care, 15, 159-165.
- [163] Derry S, Lloyd R, Moore RA, McQuay HJ. (2009) Topical capsaicin for chronic neuropathic pain in adults. *Cochrane Database of Systematic Reviews*, 2009, CD007393.
- [164] Anand P, Bley K. (2011) Topical capsaicin for pain management: Therapeutic potential and mechanisms of action of the new high-concentration capsaicin 8% patch. *British Journal of Anaesthesiology*, 107, 490-502.
- [165] Glinski W, Glinska-Ferenz M, Pierozynska-Dubowska M. (1991) Neurogenic inflammation induced by capsaicin in patients with psoriasis. *Acta Dermato-venereologica*, 71, 51-54.
- [166] Arnold WP, van de Kerkhof PC. (1994) Topical capsaicin in pruritic psoriasis. Journal of the American Academy of Dermatology, 31, 135.
- [167] Mankowski C, Poole CD, Ernault E, Thomas R, Berni E, Currie CJ, Treadwell C, Calvo JI, Plastira C, Zafeiropoulou E. (2017) Effectiveness of the capsaicin 8% patch in the management of peripheral neuropathic pain in European clinical practice: The ASCEND study. *BMC Neurology*, 17, 80.
- [168] Yong YL, Tan LT-H, Ming LC, Chan K-G, Lee L-H, Goh B-H, Khan TM. (2017) The effectiveness and safety of topical capsaicin in postherpetic neuralgia: A systematic review and meta-analysis. *Frontiers in Pharmacology*, 7, 538.
- [169] Chung M-K, Campbell JN. (2016) Use of capsaicin to treat pain: Mechanistic and therapeutic considerations. Pharmaceuticals, 9, 66.
- [170] Knotkova H, Pappagallo M, Szallasi A. (2008) Capsaicin (TRPV1 agonist) therapy for pain relief: Farewell or revival? *Clinical Journal of Pain*, 24, 142-154.
- [171] Whiting S, Derbyshire E, Tiwari B. (2012) Is there potential to use bio-active compounds (capsaicinoids) as innovative weight management aids? A meta-analysis of evidence. *The Proceedings of the Nutrition Society*, 71(OCE2), E169.
- [172] Lejeune MP, Kovacs EM, Westerterp-Plantenga MS. (2003) Effect of capsaicin on substrate oxidation and weight maintenance after modest bodyweight loss in human subjects. *British Journal of Nutrition*, 90, 651-659.
- [173] Ahuja KD, Ball MJ. (2006) Effects of daily ingestion of chilli on serum lipoprotein oxidation in adult men and women. *British Journal of Nutrition*, 96, 239-242.
- [174] Lim K, Yoshioka M, Kikuzato S, Kiyonaga A, Tanaka H, Shindo M, Suzuki M. (1997) Dietary red pepper ingestion increases carbohydrate oxidation at rest and during exercise in runners. *Medicine and Science in Sports and Exercise*, 29, 355-361.
- [175] Henry CJ, Emery B. (1986) Effect of spiced food on metabolic rate. Human Nutrition. Clinical Nutrition, 40, 165-168.
- Yoshioka M, Doucet E, Drapeau V, Dionne I, Tremblay A. (2001) Combined effects of red pepper and caffeine consumption on 24 h energy balance in subjects given free access to foods. *British Journal of Nutrition*, 85, 203-211.
- [177] Yoshioka M, Lim K, Kikuzato S, Kiyonaga A, Tanaka H, Shindo M, Suzuki M. (1995) Effects of red-pepper diet on the energy metabolism in men. Journal of Nutritional Science and Vitaminology (Tokyo), 41, 647-656.
- [178] Yoshioka M, St-Pierre S, Suzuki M, Tremblay A. (1998) Effects of red pepper added to high-fat and high-carbohydrate meals on energy metabolism and substrate utilization in Japanese women. *British Journal of Nutrition*, 80, 503-510.
- [179] Yoshioka M, Imanaga M, Ueyama H, Yamane M, Kubo Y, Boivin A, St-Amand J, Tanaka H, Kiyonaga A. (2004) Maximum tolerable dose of red pepper decreases fat intake independently of spicy sensation in the mouth. *British Journal of Nutrition*, 91, 991-995.
- [180] Smeets AJ, Janssens PL, Westerterp-Plantenga MS. (2013) Addition of capsaicin and exchange of carbohydrate with protein counteract energy intake restriction effects on fullness and energy expenditure. *Journal of Nutrition*, 143, 442-447.
- [181] Ludy MJ, Mattes RD. (2011) The effects of hedonically acceptable red pepper doses on thermogenesis and appetite. *Physiology & Behavior*, 102, 251-258.
- [182] Westerterp-Plantenga MS, Smeets A, Lejeune MP. (2005) Sensory and gastrointestinal satiety effects of capsaicin on food intake. *International Journal of Obesity (London)*, 29, 682-688.
- Diepvens K, Westerterp KR, Westerterp-Plantenga MS. (2007) Obesity and thermogenesis related to the consumption of caffeine, ephedrine, capsaicin, and green tea. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 292, R77-R85.

- [184] Yoshioka M, St-Pierre S, Suzuki M, Tremblay A. (1998) Effects of red pepper added to high-fat and high-carbohydrate meals on energy metabolism and substrate utilization in Japanese women. *British Journal of Nutrition*, 80, 503-510.
- [185] Tsi D, Nah AKH, Kiso Y, Moritani T, Ono H. (2003) Clinical study on the combined effect of capsaicin, green tea extract and essence of chicken on body fat content in human subjects. *Journal of Nutritional Science and Vitaminology*, 49, 437-441.
- [186] Ohnuki K, Niwa S, Maeda S, Inoue N, Yazawa S, Fushiki T. (2001) CH-19 Sweet, a non-pungent cultivar of red pepper, increased body temperature and oxygen consumption in humans. *Bioscience Biotechnology and Biochemistry*, 65, 2033-2036.
- [187] Yoneshiro T, Aita S, Kawai Y, Iwanaga T, Saito M. (2012) Nonpungent capsaicin analogs (capsinoids) increase energy expenditure through the activation of brown adipose tissue in humans. *American Journal of Clinical Nutrition*, 95, 845-850.
- [188] Lejeune MP, Kovacs EM, Westerterp-Plantenga MS. (2003) Effect of capsaicin on substrate oxidation and weight maintenance after modest bodyweight loss in human subjects. *British Journal of Nutrition*, 90, 651-659.
- [189] Tetens I. (2011) EFSA Panel on Dietetic Products, Nutrition and Allergies; Scientific opinion on the substantiation of health claims related to creatine and increased attention (ID 1524) and improvement of memory (ID 1528) pursuant to Article 13 (1) of Regulation (EC) No 1924/2006. EFSA Journal, 9(6), 2216.
- [190] Saito M, Yoneshiro T. (2013) Capsinoids and related food ingredients activating brown fat thermogenesis and reducing body fat in humans. Current Opinion in Lipidology, 24, 71-77.
- [191] Ibrahim M, Jang M, Park M, Gobianand K, You S, Yeon S-H, Park S, Kim MJ, Lee H-J. (2015) Capsaicin inhibits the adipogenic differentiation of bone marrow mesenchymal stem cells by regulating cell proliferation, apoptosis, oxidative and nitrosative stress. *Food & Function*, 6, 2165-2178.
- [192] Song J-X, Ren H, Gao Y-F, Lee C-Y, Li S-F, Zhang F, Li L, Chen H. (2017) Dietary capsaicin improves glucose homeostasis and alters the gut microbiota in obese diabetic ob/ob mice. *Frontiers in Physiology*, 8, 602.
- [193] Shen W, Shen M, Zhao X, Zhu H, Yang Y, Lu S, Tan Y, Li G, Li M, Wang J. (2017) Anti-obesity effect of capsaicin in mice fed with high-fat diet is associated with an increase in population of the gut bacterium *Akkermansia muciniphila*. Frontiers in Microbiology, 8, 272.
- [194] Hu Y-W, Ma X, Huang J-L, Mao X-R, Yang J-Y, Zhao J-Y, Li S-F, Qiu Y-R, Yang J, Zheng L. (2013) Dihydrocapsaicin attenuates plaque formation through a PPARγ/LXRα pathway in apoe-/- mice fed a high-fat/high-cholesterol diet. *PLoS One*, 8, e66876.
- [195] Singletary K. (2011) Red pepper: overview of potential health benefits. *Nutrition Today*, 46, 33-47.
- [196] da Silva Tremarin C, Casali KR, Meurer L, Schaan BDA. (2014) Capsaicin-induced metabolic and cardiovascular autonomic improvement in an animal model of the metabolic syndrome. *British Journal of Nutrition*, 111, 207-214.
- [197] Hsu Y-J, Huang W-C, Chiu C-C, Liu Y-L, Chiu W-C, Chiu C-H, Chiu Y-S, Huang C-C. (2016) Capsaicin supplementation reduces physical fatigue and improves exercise performance in mice. *Nutrients*, 8, 648.
- [198] Li J, Wang R, Xiao C. (2014) Association between chilli food habits with iron status and insulin resistance in a Chinese population. *Journal of Medicinal Food*, 17, 472-478.
- [199] Huang W, Cheang WS, Wang X, Lei L, Liu Y, Ma KY, Zheng F, Huang Y, Chen Z-Y. (2014) Capsaicinoids but not their analogue capsinoids lower plasma cholesterol and possess beneficial vascular activity. *Journal of Agricultural and Food Chemistry*, 62, 8415-8420.
- [200] Fraenkel L, Bogardus ST, Concato J, Wittink DR. (2004) Treatment options in knee osteoarthritis: The patient's perspective. *Archives of Internal Medicine*, 164, 1299-1304.
- [201] Prasad NS, Raghavendra R, Lokesh BR, Naidu KA. (2004) Spice phenolics inhibit human PMNL 5-lipoxygenase. *Prostaglandins, Leukotrienes and Essential Fatty Acids*, 70, 521-528.

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