Chapter 2 Harmful Elements in Estuarine and Coastal Systems

Md. Badiuzzaman Khan, Mauro Masiol, Angelika Hofer, and Bruno Pavoni

Abstract Estuaries and coastal zones are dynamic transitional systems which provide many economic and ecological benefits to humans, but also are an ideal habitat for other organisms as well. These areas are becoming contaminated by various anthropogenic activities due to a quick economic growth and urbanization. This chapter explores the sources, chemical speciation, sediment accumulation and removal mechanisms of the harmful elements in estuarine and coastal seawaters. It also describes the effects of toxic elements on aquatic flora and fauna. Finally, the toxic element pollution of the Venice Lagoon, a transitional water body located in the northeastern part of Italy, is discussed as a case study, by presenting the procedures adopted to measure the extent of the pollution, the impacts on organisms and the restoration activities.

Keywords Harmful elements • Coastal areas • Aquatic organisms • Health effects • Venice Lagoon

1 Introduction

Estuarine and coastal areas being the interaction zone between fresh and marine waters are highly complex due to tidal currents and waves (Morris et al. 1995). According to Ridgway and Shimmield (2002), they are dynamic systems where various ecological and physicochemical functions occur. In many ways, estuaries are very important ecosystems (Legorburu et al. 2013) being a vital site for primary

M.B. Khan • A. Hofer • B. Pavoni (🖂)

Dipartimento di Scienze Ambientali, Informatica e Statistica, Università Ca' Foscari Venezia, Dorsoduro 2137, 30123 Venice, Italy

e-mail: brown@unive.it

M. Masiol

Division of Environmental Health and Risk Management, School of Geography, Earth and Environmental Sciences, University of Birmingham, Edgbaston, Birmingham B15 2TT, UK

production (Bricker et al. 2008) and a suitable habitat for many species (Lotze 2010). In addition, they control exchange processes of elements between river and coastal areas (Das et al. 2010) and supply goods and services (Pinto et al. 2010). Recently, researchers throughout Europe have given priority to estuaries and coastal areas especially after the European Commission, in the European Water Framework Directive (WFD, 2000/60/EC), has emphasized the objective of achieving "Good Ecological and Chemical Status" in all European water bodies by 2015 (Borja et al. 2009; Tueros et al. 2009).

However, river banks and estuaries have always been a focus for human settlement and industrial development, recreation, tourism, development of port and boating facilities, agriculture, housing, mining and forestry. Estuaries are becoming polluted by various human activities due to a quick economic growth and urbanization (Xu et al. 2013). Estuarine and coastal areas act as a sink for trace metals and other pollutants incoming from adjacent catchments, up-watershed activities and nearby terrestrial areas, but also they are a source for the same materials to the adjoining coastal marine areas (Kennish and Fertig 2012). Moreover, estuaries have been frequently used as dumping grounds for sewages, rubbish and industrial wastes. All these activities are responsible for the pollution of estuaries and coastal marine areas. Waste discharges, accidental spills, urban and agricultural runoff and ground water flow carry a wide range of pollutants to the coastal marine areas (Water and River Commission 1997). Therefore, substantial amount of nutrients and trace metals are transported through riverine input and submarine groundwater discharges to coastal sea waters due to progressive anthropization in coastal areas (Moore 2006). Trace elements are taken by river and ultimately mixed into coastal marine areas after transportation and alteration in estuarine areas (Swarzenski et al. 2006).

The estuarine and coastal seas play a vital role in the biogeochemical cycles of various essential and non-essential elements, by favouring physico-chemical and biological processes such as variations in the chemical speciation, inorganic scavenging, sedimentation, bio-alteration, bio-accumulation and bio-magnification (Costa et al. 2012). Trace metals are natural elements of crustal materials which enter the aquatic environment via erosion process (Di Leonardo et al. 2009; Rocha et al. 2011; Weber et al. 2009). Although some metals are vital to biota, all are potentially toxic when concentrations exceed a certain limit. Metals are received in the estuaries through rivers, many of them are persistent, toxic and bioaccumulative in nature (Liang 2011). Trace metals enter the aquatic systems mostly from anthropogenic inputs such as leaching of impervious urban surfaces (roads, parking lots and roofs) and soil surfaces by runoff and to sewage overflow from sanitary sewer systems during high rainfall events, locally treated wastewater discharge and industrial effluents (Bay et al. 2003; Bothner et al. 2002; Nicolau et al. 2012). The entrance of trace elements from river to open ocean via estuaries depend on the partitioning of trace metals between dissolved and particulate phase, riverine and anthropogenic inputs, coastal and seafloor erosion, biological activities and also physical - chemical transport and biological interaction within these areas. Trace elements may be accumulated in sediments after recycling due to chemical and biological processes and also may return back to the water column after resuspension (Förstner 1984; Tessier and Campbell 1987).

There are two processes controlling the partitioning of metallic species between the solution and suspended particulate matter (SPM) phase during estuarine mixing. Firstly, the removal of metals by flocculation of humic and fulvic acid-metal complexes and desorption of metals from river particles or from resuspended sediments (Roux et al. 1998). The features of a river or estuary's watershed such as climate (e.g., rain frequency and intensity), morphology, hydrology, geology and land use drive estuarine dynamics, particularly flushing times of waters and particle concentrations and residence times, which are among the most important factors controlling metal reactivity in estuaries (Elbaz-Poulichet 2005). Redox conditions (Zwolsman et al. 1997) and the presences of dissolved organic matter (DOM) also control the fate of metals in estuaries.

Keeping the above points in mind, the sources, chemistry, sediment accumulation and removal mechanisms of the harmful elements in estuarine and coastal seawater are described below. Elements are discussed in the following chapter in an order that reproduces a decreasing impact on the organisms and ecosystems. However, the chapter of the book first discussed metal, then non-metal (As) followed by organotin compounds (Tributyltin and Triphenyltin).

2 Harmful Elements in the Estuary and Coastal Areas

2.1 Mercury

Mercury is a widely distributed hazardous pollutant and has received great attention globally because of its persistence in environments, high toxicity to organisms, reactivity and tendency of forming more toxic organic mercury compounds and also biomagnifications capability along the food web (Jiang et al. 2006; Craig 1986; Beckvar et al. 1996). Although the use of Hg has declined considerably during the past several years, still now the metal has to be considered important in several sectors such as energy generation, incineration and mobilization from mining activities (Hines et al. 2012; Fitzgerald et al. 2007). Mining, industrial activities and urban expansion in estuarine and coastal zone are the main anthropogenic sources of Hg (Luo et al. 2012).

The coastal zone plays a vital role in the global mercury cycle (Mason et al. 1994) both as sink for terrestrial Hg and as source of methylated Hg to the ocean (Mason and Benoit 2003). Approximately 50–80 % of the total Hg coming from rivers is deposited within the estuarine zone (Cossa et al. 1996; Benoit et al. 1998). Input of Hg in the coastal zone is influenced by various natural and anthropogenic activities such as coastal and seafloor erosion, river and sewage/mining discharge, and atmospheric Hg deposition (Ci et al. 2011). Hg can exist in different forms which control its availability, complex distribution, and toxicity.

The most important forms of Hg are elemental Hg (Hg⁰), inorganic Hg (Hg²⁺), monomethylmercury (CH₃Hg⁺) and dimethylmercury (CH₃HgCH₃) (Leermakers et al. 2001). All of these are influenced by various factors such as redox and pH conditions, and concentrations of inorganic and organic complexing agents (Horvat et al. 2003).

Approximately 10–30 % of the Hg exists as elemental Hg in ocean water (Mason and Fitzgerald 1993), where Hg^0 is mainly formed from the reduction of Hg (II) by aquatic microorganisms (Mason et al. 1995) and from photoreduction of Hg (II) (Amvot et al. 1997; Costa and Liss 2000) followed by geotectonic activity (Ferrara et al. 2003; Horvat et al. 2003). Abiotic methylation may take place in an environment loaded with humic organic matter (Weber 1993). Methylmercury (MeHg) is produced from inorganic mercury due to the activity of sulfate reducing bacteria (Mason et al. 1993; Morel et al. 1998; King et al. 1999). Although methylmercury is present in estuarine water at very low concentrations (Cossa and Coquery 2005), it is the most toxic form of Hg and may be accumulated in human body through the consumption of sea foods (Baldi 1997; Morel et al. 1998). Methyl mercury is a unique example of metal biomagnification in food chains due to its special characteristics such as (i) high lipid solubility and easy transfer through membranes and (ii) longer biological half-life. There are several factors controlling the production of MeHg such as temperature (35 °C is the optimum temperature in river), salinity (methylation rate decreases at increasing salinity due to the formation of chloride complexes), redox/sulphide levels (MeHg is higher in moderately anoxic sediments and increased proportionately with sulphide concentrations up to 1.8 mg S g^{-1}) (Callister and Winfrey 1986; Compeau and Bartha 1984; Breteler et al. 1981; Craig and Moreton 1983).

Dimethylmercury is an organic form of Hg found in seawater and mainly produced in oxygenated environments (Mason et al. 1995). There is little evidence of the production of dimethylmercury in surface water. However, it comes to the surface from deep water through deep water upwelling and deep thermocline mixing and degrades quickly due to its low stability with light and higher surface water temperature (Fitzgerald and Mason 1997). There is also evidence of the demethylation of the methylmercury (Marvin-DiPasquale et al. 2000; Barkay and Wagner-Dobler 2005) mediated by the bacteria through both reductive process (products: Hg⁰ and CH₄) and also oxidative process (products: Hg²⁺ and CO₂) (Barkay et al. 2003).

The concentration of dissolved mercury ranged from 0.5–3 ng L⁻¹ to 2– 15 ng L⁻¹ in ocean and coastal sea-water, respectively (WHO 1989), whereas its concentrations varied from 20 to 100 μ g kg⁻¹ in ocean sediments. The concentration of methylmercury in UK estuarine sediments is less than 0.02 μ g g⁻¹ (Langston 1982). Dissolved mercury has strong attraction for organic matter and suspended sediment and becomes accumulated in sediment after adsorption onto these particles. After deposited in sediment, inorganic Hg is transformed into methylmercury, which is bioavailable and toxic to aquatic organisms (Campbell et al. 1986). Methyl mercury can be accumulated at concentrations 10 (*Fucus vesiculosus*) and 100 (*Mytilus edulis*) times higher than sediment concentrations in estuarine algae and invertebrates (Langston et al. 1996). The concentration of total mercury in the Isonzo River (northern Adriatic Sea) ranged from 1.72 to 31.4 ng L⁻¹, whereas the average value was 9.2 ± 7.9 ng L⁻¹. On the other hand, the concentration of methyl mercury ranged from 1.65 to 17.2 ng g⁻¹ in sediments. However, the concentration of methylmercury varied from 0.21 to 6.28 ng g⁻¹ which is 0.121 % of total Hg (Bratkič et al. 2013).

2.1.1 Effects of Mercury on Aquatic Organisms

The European Water Framework Directive – 2000/60/EC (2000) indicates mercury as a priority harmful substance. In addition, mercury is included in the first 20 compounds of the priority list of dangerous substances published by the Agency for Toxic Substances and Disease Registry (ATSDR 2007). This metal behaves as a resistant cation, which forms complexes and can interact with proteins, enzymes and cell membranes in biological systems. It is present under different oxidation states such as Hg^{2+} , Hg^+ or Hg^0 in the methylated forms dimethylmercury (CH₃)₂Hg and methylmercury CH₃Hg⁺. This two derivates are persistent and bioaccumulable. The methylmercury with its ionic form is hydrophile and quite mobile in the environment, while the dimethylmercury is lipophylic and volatile. In mammals, depression and irritability indicate a low intoxication of mercury, whereas damages to the nervous system such as blindness, madness, paralysis, as well as effects on the DNA (Renzoni et al. 1998) can indicate a heavy intoxication of mercury. Elemental mercury is less dangerous because it is easily eliminated with the urine. As described by (Grieb et al. 1990), the methylmercury accumulates in muscle tissues, liver and kidney of fishes (Mackey et al. 1996) with a biological half life varying from 2 to 3 years (USEPA 1984 and references therein). Considering the trophic chain, the methylmercury contained in the benthos or the smaller fish tissues will deposit in different tissues of the predator (e.g. kidney, bile, muscle). As the level of intoxication in fish depends on its diet (Adams et al. 2010), age, weight and length, as well as on the physical-chemical parameters of the water body, no definitive intoxication parameters can be established, but a stress condition can be proved (Das et al. 1980), because a significant impairment of metabolitic functions, and a modified morphology and behaviour can be detected (Alam and Manghan 1992; Scott and Sloman 2004). Adams et al. (2010) reported that at levels higher than 5 μ g g⁻¹ of wet weight in fish muscles impairment in the reproduction can be observed. In the benthos, some difficulties were found in determining the toxicity of Hg, rather than of a mixture of metals (Suchanek et al. 2008). Brown and Ahsanullah (1971 and Arizzi Novelli et al. 2002b) demonstrated that Hg is the most toxic element compared to others such as Cu, Cd, Zn, Pb for larvae of Ophryotrocha and Artemia as well as for sperm cells and embryos of Paracentrotus lividus, respectively.

2.2 Cadmium

Great attention has been given to cadmium due to its toxic behavior in the environment (Steinnes 1989; Flyhammer 1995; Porter et al. 2004). The main dissolved forms of Cd are chloride complexes whereas the most available species is the free ion (Cd²⁺), the proportion of which increases with decreasing salinity (Engel and Fowler 1979; Cross and Sunda 1985). Many scientists reported that there was a relationship between the Cd concentration in sediment and biota. A linear relationship was found between the concentration in the polychate *Nereis diversicolor* and in surface sediment from estuaries in Devon and Cornwall (Bryan and Hummerstone 1973a).

The concentration of Cd in Ocean was found to range from 0.02 to 0.12 μ g L⁻¹ (Bruland et al. 1979), whereas the concentration was 0.01–0.17 μ g L⁻¹ in UK coastal waters. However, the concentration of Cd in estuary fluctuated from 1 to 50 μ g L⁻¹ (Morris 1984; Bryan and Gibbs 1983). Although chlorinity is a vital parameter controlling Cd behavior in estuaries, other factors such as ionic strength (increase of Ca²⁺ and Mg²⁺ concentrations competing with Cd²⁺ for adsorption onto particles), pH, redox potential, microbial activity, organic matter degradation and the residence time of the particles in the mixing zone (type of estuary) can be involved.

Cd has higher affinity for the chloride ion rather than to organic ligands (Bilinski et al. 1991; Tipping et al. 1998). During the mixing of river water with sea water, mobilization processes occur from particulate matter by the formation of highly stable and soluble Cd-chloride complexes. The formation of chloride complexes increase with the increase of salinity which regulates the activity of the free ion (Paalman et al. 1994; Thouvenin et al. 1997; Lemaire et al. 2006). An organic complex is occurred at low salinity condition. There was also evidence of Cd desorption in the estuary (Boyle et al. 1982).

2.2.1 Effects of Cadmium on Organisms

This metal can be complexed by the seawater and form soluble forms such as CdCl⁺, CdCl₂(aq) and CdCl₃⁻. Under these forms, Cd can be taken up from different organisms living in the aquatic environment, stored in liver, kidney and gills, and cause problems in the enzymatic regulation of the carbohydrate metabolism. Cd is one of the elements next to Mn and Ni which can disrupt the storage or mobilization of metabolic substrates like glucose, glycogen, lactate, lipids and proteins (Scott and Sloman 2004). As Cd and Zn have the same positive loading, it can substitute Zn in some enzymes causing an alteration of their structure, which compromises their activities. Moreover, Cd (in addition to Cu, As, Hg, Pb) influences reproduction processes of fishes (Das et al. 1980). Cd is accumulated in small amounts in the fishes' muscle but in higher amounts in other tissues and organs (USEPA 2001). In mammals, Cd can cause caught, nausea, thoracic diseases, but in

small quantities it is bound by proteins and eliminated easily with the urine. When exposed to high quantities, red blood cells can be destroyed and the kidney can be damaged. Therefore, Cd is considered a potentially toxic element (Mackey et al. 1996). In benthic organisms, Cd was linked to lysosomal damage and mortality of the organisms (Thompson et al. 2007).

2.3 Lead

Lead is one of the major anthropogenic pollutants and is now considered as ubiquitous (Chakraborty et al. 2012; Snape and Townsend 2008; Lobinski and Marczenko 1996). Nutritionally or physiologically lead is not an essential nutrient for either humans or other organisms. It is toxic, bioaccumulative and persistent. The contamination of lead from gasoline burning has declined in different environmental matrices, including aerosols (Migon et al. 1993; Grousset et al. 1994), seawater (Boutron et al. 1991; Wu and Boyle 1997) and rivers (Singh and Singh 2006).

Various uses of lead such as in storage batteries and as organic antiknocking additives (tetraalkyllead) to petrol, cables, solders, steel products, ammunition, shielding systems from radiation and x-rays, circuit boards in computers and electronics equipments, superconductor and optical technology, insecticides, pigments, paints, ceramics, enamels, glass, plastics and rubber products, coal-fired power stations, wastes from runoff and incineration and other industrial effluents have contributed significantly for the wide distribution of lead in the environment (Ritson et al. 1999; Hansmann and Köppel 2000). Although legislations have been implemented to enforce the use of alternative petroleum additives and recover lead from used batteries, the uses of lead are continuing in other areas of application.

Various forms of lead are available in the environment. Organometallic forms of lead are more lipophilic and can easily penetrate biological membranes. As a result, alkyllead species may be bioaccumulated in food chains. The main dissolved forms of Pb are PbCO₃ and PbOH (Nurnberg and Valenta 1983), whereas the most available inorganic form is the free ion Pb²⁺ (Freedman et al. 1980). Pb can be accumulated in the sediments by the deposit-feeding species. Pb levels in the polychaete *Nereis diversicolor* were correlated linearly to the Pb/organic matter ratio in the sediment (Bryan 1985). The availability of lead decreases with the increases of sediment organic matter concentration. This is due to the complexation of the free ion Pb²⁺ in the interstitial water, or possibly in the gut following sediment ingestion (Windom et al. 1982). Fe oxides have also effect on Pb concentration and its availability decrease with the increase of Fe oxides (Luoma and Bryan 1982). Inorganic lead may be a source for the contamination of birds. Tetramethyllead and tetraethyllead are the two main lead compounds introduced into the environmental. Both of these compounds are used as gasoline antiknock additives. However, tetramethyllead and derivatives of methylatedlead are the two main compounds found in the environment as organometallic compounds of lead.

Methyllead compounds are only found in the immediate vicinity of anthropogenic sources which are normally present at nanograms per gram level and decay occurs via R_3Pb^+ and R_2Pb^{2+} , with inorganic lead as the final products (Craig 1986). The principal factors governing the biological activity of lead in aquatic environment depend on its geochemical behavior. The physical and chemical properties of surface water and sediments such as pH, total hardness, porosity, Eh and organic matter affect the concentration of lead. Lead carbonates, lead sulfates and lead sulfides are predominant in sediments, whereas lower amount of lead were detected in surface water in dissolved form. Lower pH helps for the mobilization of inorganic lead.

Flegal and Patterson (1983) conducted an experiment and found the concentration of Pb in Ocean to be in the range 0.001–0.014 μ g L⁻¹. The dissolved concentration of Pb ranged from 0.015 to 0.135 μ g L⁻¹ along the east coast basin of Britain, whereas in the Humber estuary the concentration fluctuated from 0.010 to 0.055 μ g L⁻¹ (Balls 1985). However, the highest portion of lead in coastal waters and estuaries is connected with particles. The concentration of inorganic Pb in UK estuarine sediments ranged from 25 μ g g⁻¹ to more than 2,700 μ g g⁻¹ of sediment (Bryan et al. 1985). Organolead compounds have also been detected in sediments such as the concentrations of tetraethyl, triethyl and diethyl Pb in Canada were 1.15, 0.19 and 0.022 μ g g⁻¹, respectively (Chau et al. 1984). Riley and Towner (1984) conducted an experiment and found higher concentrations of tri and dialkyl Pb than those of dissolved inorganic Pb.

2.3.1 Effects of Lead on Organisms

When present in its ionic form Pb^{2+} , it becomes hazardous for the environment as it is easily taken up by organisms. Therefore, it is also included in the priority list of ATSDR (2007). In mammals, Pb is distributed firstly in the soft tissues, then in the bones, where it takes the place of Ca²⁺. It may cause damage to liver and the reproductive system and to the DNA as well. In fish, the lead poisoning can cause permanent and severe damages in the reproductive system (Das et al. 1980) after long term exposure, depending on the species, size and age of the fish (Alam and Manghan 1992). Temperature, depth and movement of the water body influence the possibility for marine organisms to take up Pb, stored in liver, hepatopancreas and hard tissues and finally influence the metabolism of the organism (Prosi 1989). However, lead (Pb²⁺) also influences the cytoskeleton, mitochondria, the nucleus in liver cells, causing the cell death, whereas in the kidney it inhibits only some enzymes (Rabitto et al. 2005).

2.4 Zinc

Zinc acts as metal cofactor for many enzymatic processes of both eukaryotic and prokaryotic organisms (Vallee and Auld 1993) and is used in nucleic acid transcription and repair protein (Anton et al. 2007). It also participates in nutrient uptake mechanisms especially for the uptake of CO_2 through the enzyme Carbonic Anhydrase (CA) (Morel et al. 1994) and also for the uptake of phosphate through the enzyme alkaline phosphatase (Shaked et al. 2006).

There are several forms of available Zn in the aquatic system. However, the most bioavailable and dissolved one is the free ion Zn^{2+} (O'Brien et al. 1990). The concentration of zinc in coastal areas and estuaries are much higher than in the ocean (less than 1 µg L⁻¹, Bruland et al. 1979). The concentration of zinc in a North Sea transect was 0.3–70 µg L⁻¹ (Duinker and Nolting 1982), whereas the concentration varied from 11 to 23 µg L⁻¹ in Bristol Channel-Seven Estuary (Morris 1984). The Zn concentration of sediment in Restronguet Creek, UK, ranged from less than 100 to around 3,000 µg g⁻¹. In interstitial waters, the concentration of Zn in Creek sediments veried from 262 to 396 µg g⁻¹, whereas it fluctuated from 67 to 216 µg g⁻¹ at a depth of 10 cm (Bryan and Gibbs 1983). The average Zn concentration in the western coast of Mauritius was 107 mg kg⁻¹ (Ramessur 2004).

Generally dissolved Zn is predominant in river water, whereas in estuaries a larger portion of the zinc is adsorbed to suspended particles where concentrations of suspended particles are higher (CCREM 1987). Zinc may be mobilized from sediments in low saline estuarine areas, due to the microbial degradation of organic matter and also displacement by calcium and magnesium. Zinc also can be deposited with flocculated particles during high tubidity and may accumulate in anaerobic sediments.

2.4.1 Effects of Zinc on Organisms

Zinc is a micronutrient for marine organisms, important in the enzymatic and metabolic regulation, but at high concentrations it becomes toxic and can cause different permanent and severe damages, e.g. in the reproduction processes (Das et al. 1980). Low concentrations of Zn were observed to stimulate the growth of the organisms in the marine environment. The water hardness (hardness <1 mg L⁻¹ as CaCO₃), when Zn is present only at low concentrations can influence the egg production and the fertilization process in fish (Sprague 1971 and references therein). An irregular development of some marine organisms has been detected when the concentration of Zn increases (Volpi Ghirardini et al. 2005). Further, it seems that diet borne Zn is not toxic, but it can influence the concentrations of Fe and Cu in the fish. However this feature is still depending on species, age, length and weight of the organism. However, in fish Zn is principally accumulated in the digestive tract after short-term and long-term exposure. Then, gills, liver, kidney,

spleen and skeletal tissues divide the second position in Zn accumulation (Clearwater et al. 2002).

2.5 Chromium

Chromium is an important component in industrial sectors and used for the production of ferrochrome, electroplating, pigment production and tanning. The major sources of Cr in the marine environment are the wastes of these industries, dumping of solid wastes and municipal wastes. Different forms of chromium are available in the aquatic environment, but the main biologically important forms are Cr (III) and Cr (VI). Trivalent species participates in glucose, lipid and protein metabolism (Eisler 1986), whereas hexavalent Cr is comparatively more toxic than the trivalent form due to its oxidizing potential, solubility and capability to cross cell membrane (Levy and Venitt 1986). The main forms of Cr (III) are CrOH^{2+} , Cr(OH)_{3}^{0} and organic and inorganic complexes, whereas HCrO_{4}^{-} and CrO_{4}^{2-} are the main forms of Cr(VI) in aquatic system (Rai et al. 1987, 1986).

In offshore waters, the average concentration of Cr ranged from 0 to 234 μ g L⁻¹, while the concentration of Cr varied between 30 to over 200 μ g g⁻¹ in UK estuarine sediments (Murray et al. 1983). The total concentration of Cr in the upper 500 m water of the Sargasso Sea (Near to the Bermuda in the North Atlantic Ocean) ranged from 2.5 to 4.5 nM where the concentrations of Cr (III) and Cr (VI) were 1.2 and 2.4 nM, respectively (Connelly et al. 2006). The main way of the removal of Cr (III) from the sea water is adsorption to suspended particles such as oxides of Fe or by surfaces (Peternac and Legovic 1986; Mance et al. 1984). The adsorption of trivalent Cr increases with pH. However, adsorption process may decrease with the increases of competing cations. On the other hand, the adsorption of hexavalent Cr decreases with pH and the presence of dissolved anions. Chromium migrations are controlled by several processes such as competition between complexation, dissolution/precipitation, redox process and adsorption/desorption mechanisms. The migration of trivalent chromium occurs under acidic conditions and it occurs as dissolved organic complexes. On the contrary, Cr (VI) migrates rapidly but the process may slow down due to the presence of Fe (II) and high organic matter concentration (Richard and Bourg 1991). With the presence of dissolved oxygen, oxidation of Cr (III) to Cr (VI) is very low but increases with the presence of MnO₂. On the other hand, the reduction of Cr (VI) to Cr (III) increases with the presences of organic matter-rich environment. Finally, after settling of the Cr in sediment, remobilization of Cr occurs either as chromate or as organic Cr (III) complexes (Fig. 2.1).



Fig. 2.1 Chromium cycling in the aquatic environment (Adapted from Richard and Bourg 1991)

2.5.1 Effects of Chromium on Organisms

Cr is present in the environment mainly in its inorganic forms Cr(III) and Cr(VI). The latter is a highly toxic compound as it is soluble at each value of pH. Therefore, it is also a quite mobile species. Cr(III) is important in the metabolism of fish, whereas Cr(VI) has increased the maximum lifespan of females in the study of Perez – Benito (2006). As reported by USEPA (1980), *Daphnia magna* is highly sensitive to Cr(VI) in the water body. Further, the photosynthesis in algae and the growth of some fish species varies with the concentration of Cr(VI). However, the uptake of Cr(VI) in a marine organism depends on the conditions of the water body in terms of temperature, salinity and pH.

2.6 Nickel

Nickel is a ubiquitous and nutritionally essential nutrient for plants, animals and micro-organisms. Nickel is used for the production of stainless steel and nickel alloys which are ultimately used in vehicles, processing machinery, armaments, tools, electrical equipment, household appliances and coinage. Moreover, it is also used in catalysts, pigments and batteries. The main sources of nickel to the environment are the uses of coal and oil for power generation, incineration of wastes, nickel mining, steel manufacturing, electroplating, cement manufacturing. However, nickel from all of these activities finally reaches the marine environment due to dumping of wastes, surface-runoff of wastes and incineration (WHO 1991).

Nickel may exist in several forms in the aquatic system. However, Ni (II) is the prevalent oxidation state under normal environmental conditions. The other forms exist in oxidation of 1–, 1+, 3+ and 4+ (Young 1995; Clayton and Clayton 1994; Coogan et al. 1989). The concentration of Ni in fresh water ranged from 2 to 10 μ g L⁻¹, whereas it ranged from 0.2 to 0.7 μ g L⁻¹ (WHO 1991) in Ocean. In coastal, bay and estuary waters it varied from 0.20 to 5.3 μ g L⁻¹ (Domínguez-Lledó et al. 2007). The concentration of nickel in sediment of Ria Ferrol (NW Spain) were in the range 11.3–67.2 μ g g⁻¹, whereas the average concentration was 32 μ g g⁻¹ (Barciela-Alonso et al. 2003).

Nickel exists in aquatic systems as soluble salts, absorbed on clay particles or together with organic materials such as humic and fulvic acids and proteins. However, sediments also act as a source of nickel through desorption (WHO 1991). The fate of nickel in sea water is controlled by several factors including pH, pE, ionic strength, type and concentration of organic and inorganic ligands, and the presence of solid surfaces for adsorption (Barciela-Alonso et al. 2003).

2.6.1 Effects of Nickel on Organisms

Ni is an essential element for all organisms, aquatic or not, which becomes toxic at high concentrations. In addition, it has been indicated as possible carcinogenic (Coogan et al. 1989; Denkhaus and Salnikow 2002). A long term exposure of marine organisms can modify their morphology and behavior (Alam and Manghan 1992) and disrupt the storage and/or mobilization of essential metabolic substrates (e.g. glucose, proteins) (Scott and Sloman 2004). However, there is no strong evidence about the relationship between the concentration of nickel and its toxicity in the organism (Griffitt et al. 2008).

2.7 Cobalt

Cobalt is an essential component for human and other organisms since it is an important component of Vitamin B-12. Cobalt has both natural and anthropogenic sources. The main human sources are coal and oil burning, industrial activities, vehicular exhausts and sewage sludge. Cobalt is used in steel and alloys, metal-lurgy, electroplating, nuclear technology, fertilizers, medicine, drier for paint, foam stabilizer in beer brewing (Tsalev and Zaprianov 1985; ATSDR 1999).

The main forms of cobalt in seawater are Co^{2+} and its sulfate-, chloro- and carbonate-complexes (Ahrland 1975) and is removed quickly from seawater with MnO₂ (Knauer et al. 1982). About 90 % of the cobalt can be found in the sediments and in suspended particulate matter in sea waters (Robertson et al. 1973). Sometimes cobalt acts as a limiting nutrient for marine phytoplankton when present at low concentration in the ocean waters (Knauer et al. 1982).

The mean concentration of cobalt detected in seawater was $0.3 \ \mu g \ L^{-1}$, whereas it varied from 0.1 to 10 $\ \mu g \ L^{-1}$ in fresh water. The concentrations of cobalt in estuarine sediments ranged from 15 to 60 nM (Huynh-Ngoc et al. 1989), whereas Duchart et al. (1973) found the cobalt concentration 34–650 nM in estuarine sediments. The cobalt concentration in surface marine water is 18–300 pM, whereas it ranged from 20 to 50 pM at depth (Donat and Bruland 1995). The concentration of cobalt is comparatively higher in estuarine and coastal waters than in ocean. Achterberg et al. (1999) measured 140–310 pM cobalt in coastal water near Washed Humber Estuary.

The mobilization of cobalt in estuarine and coastal areas depends on the partitioning of the metal between dissolved and particulate phases (Martino et al. 2002). This is influenced by various factors such as types of metal ion, metal concentration, pH, salinity and dissolved oxygen (Stumm 1992). The primary ways of removal process of cobalt are particle scavenging and biological uptake. However, the metal is also released from the sediment to the dissolved phase due to desorption as a consequence of salinity changes (Kraepiel et al. 1997)

2.7.1 Effects of Cobalt on Organisms

Cobalt seems to be an essential element for certain blue-green algae, nitrogenfixing bacteria and symbiotic systems as well as for animals. It can be found in two oxidation states, Co^{2+} and Co^{3+} . Co^{3+} is found rarely in organisms as it is thermodynamically unstable in natural waters and cannot be taken up easily. Co can also substitute other trace metals (e.g. Cu, Pb, Zn, Cd) in a wide variety of minerals due to its similar geochemical properties, to form complexes (EnvGovCa 2003). In addition, it can influence negatively the growth rate of, and the Ca-uptake in aquatic herbivores (De Schamphelaere et al. 2008)

2.8 Vanadium

Vanadium is a vital component for aquatic organisms such as marine phytoplankton (Moore et al. 1996), macroalgae (Patrick 1978; Nalewajko et al. 1995) and other organisms (Taylor et al. 1997). Vanadium is present in many enzymes such as haloperoxidases (Butler and Carter-Franklin 2004), nitrate reductases (Antipov et al. 1999) and nitrogenases (Rehder 2000) and also participates in metabolic activity of many components such as chlorophyll synthesis (Meisch et al. 1977; Wilhelm and Wild 1984), cell division (Meisch and Benzschawel 1978), phosphate uptake kinetics (Lee 1982), sulfoxidation (Andersson et al. 1997). Vanadium, on the other hand, is a very important metal for technology. Approximately 75–85 % of the vanadium is used in ferrous metallurgy (alloy additives in steel). The main inputs of vanadium into environment are the burning of fossil fuels, crude oil spillages and industrial activities (atomic energy industry, air craft construction and space technology, as catalyst for the production of sulfuric acid and plastics) (Kalogeropoulos et al. 1989; WHO 1988).

Vanadium in the aquatic environment exist in the form of many species, but the predominant stable oxidized forms are V(IV) and V(V). However, V(V) is more toxic than V(IV) (Pinto et al. 2013). El-Mosehy (2006), who collected sediment samples from the coastal area of the Egyptian seas (Mediterranean and Red Seas) found a mean concentration of 52.61 μ g g⁻¹ in the Red Sea sediments, whereas the mean value was 40.58 μ g g⁻¹ in the Mediterranean Sea. The concentrations of dissolved and particulate vanadium in the seawater of English Channel were in the ranges 15–28 nmol L⁻¹ and 2–32 nmol L⁻¹, respectively (Auger et al. 1999).

The speciation of vanadium is influenced by redox condition of the aquatic environment (Taylor and van Staden 1994; Pettersson et al. 2003). Various redox pairs such as NO_3^{-}/NH_4^{+} , Fe^{3+}/Fe^{2+} , MnO_2/Mn^{2+} , SO_4^{-2-}/H_2S play a significant role in vanadium speciation and cycling in coastal seawaters. The solubility of vanadium decreases with decreasing oxidation number. Most of the vanadium present in ocean waters has a conservative behavior, which turns into a non-conservative one in coastal water (Paulsen and List 1997). As most of the vanadium is accumulated in the sediment and only 10 % is present in a soluble form, it is evident that a continuous removal from the sea water occurs (WHO 1988).

2.8.1 Effects of Vanadium on Organisms

Vanadium is an essential element for mammals depending on its concentration and it is bioavailable in the environment. In marine mammals Vanadium was detected at concentrations around 0.2–1.2 μ g g⁻¹, wet weight, in liver tissues, where the concentration is proportional to the animal size and age. Furthermore, V shows a good correlation with the concentration of Hg in tissues of marine organisms and it forms complexes with phosphorus. These complexes cause the inhibition of

ATPase. It accumulates in bones, liver, kidney, muscles (Mackey et al. 1996). When V is present at low quantities, it stimulates the biosynthesis of chlorophyll in green algae in the presence of light (Meisch and Bielig 1975). Further, V^{3+} and V^{4+} show antimicrobial activities at low pH values. Therefore, a Vanadium accumulation in the tissues of an organism can be a defense against predators, bacterial infections and biofouling competitors (Odate and Pawlik 2007). At low concentrations (0.041 mg L⁻¹), Vanadium stimulates the growth and reproductive performance in fish, whereas high concentrations (11.2 mg L⁻¹) are toxic (Perez-Benito 2006).

2.9 Iron

Iron is one of the most important essential plant nutrients and plays an important role in various metabolic processes such as photosynthesis, respiration, nitrogen fixation and detoxification of reactive oxygen species (Sunda 2001; Crichton 2001). Iron enters the aquatic environment from weathering as well as from human activities such as burning of coke and coal, acid mine drainage, mineral processing, sewage, iron related industries and the corrosion of iron and steel (CCREM 1987).

Coastal waters receive iron from river water, groundwater inputs and terrestrial runoff. Generally iron exists in sea water in two oxidation states such as iron (II) and iron (III). These oxidation states play a significant role in the formation of soluble organic and inorganic complexes, colloids, insoluble ferric oxyhydroxides and particle phases (Bruland and Rue 2002; Achterberg et al. 2001). The iron (III) oxidation form is predominating in oxygenated waters, but it becomes insoluble after formation of oxyhydroxides.

The largest portion of dissolved iron (III) (80–99 %) occurs in the form of complexes with organic ligands probably produced by iron limited phytoplankton (Rue and Bruland 1995) or bacteria (Granger and Price 1999), whereas only a small portion is found in free hydrated (Fe³⁺) or inorganically complexed form (Gledhill and Van den Berg 1994; Rue and Bruland 1995; Nolting et al. 1998). On the other hand, iron (II) is thermodynamically unstable in oxygenated water and is quickly oxidized to iron (III). The main sources of iron (II) in sea water are photoreduction of iron in surface water, atmospheric deposition and diffusion from sediments (Miller et al. 1995; Zhuang et al. 1992). This iron (II) contributes for about 50 % of the total dissolved iron in sea water (Gledhill and Van den Berg 1994; Hong and Kester 1986).

The concentration of iron in unpolluted oceanic seawater was found to range from $1.9-2.8 \text{ ng L}^{-1}$ to $224-1,228 \text{ ng L}^{-1}$ and higher concentrations could be found in estuarine water (Whitehouse et al. 1998). The mean concentration of iron was $20.8 \mu \text{g g}^{-1}$, which was found in coastal salt marshes. Ergul et al. (2010) found an iron concentration of 0.033 ng g⁻¹ and 25.1 $\mu \text{g g}^{-1}$ in sea water and sediment samples, respectively.

The greater part of the dissolved iron in river waters exists as small colloidal particles, whereas the estimated value of dissolved iron (Fe) input of rivers in the



Fig. 2.2 Schematic diagram of biogeochemical iron cycling in the ocean (Adapted from Achterberg et al. 2001)

world oceans is 26×10^{19} mol year⁻¹ (de Baar and de Jong 2001). Removal of particulate Fe occurs through flocculation during the estuarine mixing of the river water with seawater (Sholkovitz 1978). The biogeochemical cycle of Fe in oceanic environment is given in Fig. 2.2. Organic complexation is one of the key factors of biogeochemistry of Fe in estuarine waters and keeps iron in dissolved phase at high salinities beyond the flocculation zone. This dissolved phase is being flushed from the estuary, whereas the non-organically complexed fraction becomes aggregate

and adsorb to particles, thus it remains within the internal cycle of estuary for a longer period of time (Morris et al. 1986). In anoxic marine waters, ferrous iron is mobilized from sediments and diffuses into the water column.

2.9.1 Effects of Iron on Organism

The bioessential element iron is not toxic itself, but it enhances the uptake of As(III) in some marine organisms, which causes adverse health effects for the organism, while it inhibits those of As(V) (Khang et al. 2012). Further, zero-valent iron has been tested by Gheju (2011) as a promoter of the reduction of the heavily toxic and mobile Cr(VI) into the less toxic Cr(III), a process which depends straightly of the environmental conditions of the aquatic system. The same effect was detected when marine organisms were exposed to Cd bound to ferric hydroxide which makes Cd less bioavailable for the organisms (Wu et al. 2012). Nevertheless, Fe is a fundamental element for many biological processes occurring in living organisms. One of them is the interference to the thyroid metabolism and to different enzymes in mammals and fish (Rejitha and Subhash Peter 2013).

2.10 Copper

Copper is a ubiquitous element existing in all natural waters and sediments. Copper is an essential plant nutrient and plays a vital role in the metabolic activities of the organisms. However, it may become toxic to aquatic organisms at concentrations 10–50 times higher than their normal recommended limit (Hall et al. 1997). Recently, the use of Cu as antifouling coating in ships has increased due to the ban of tributyltin and triphenyltin in antifouling coatings. Therefore, concerns are growing that copper has a detrimental effect on the aquatic environment and now it has become an issue in the European saltwater environment because of ecological risk from copper exposure (IMO 1997). Copper exists in various chemical forms such as free copper ions, or complexed with inorganic anions or organic ligands or as suspended particles when present as a precipitate or adsorbed onto organic matter (Mance et al. 1984). The free Cu^+ and Cu^{2+} ions are among the most toxic forms to marine life and toxicity decreases according to the order of Cu⁺ and Cu²⁺> inorganic copper and organic copper (Jones and Bolam 2007; Zamuda et al. 1985). Actually dissolved copper exists in aquatic environment in different forms such as free hydrated cations, organic or inorganic complexes (Achterberg and Braungardt 1999), whereas particulate copper is bound to various mineral phases (clays, quartz, carbonates), iron/manganese oxides and organics (Millward 1995). There are many factors that control the bioavailability and concentration of dissolved Cu such as the cupric ion concentration. These include salinity, the presence of organic ligands and pH (Bryan and Langston 1992).

Hall and Anderson (1999) conducted a research on seawater and found that the concentration of Cu ranged from 0.26 to 16 μ g L⁻¹, 0.28 to 5.48 μ g L⁻¹ and 0.069 to 5.56 μ g L⁻¹ in marina/harbor, estuary and coastal/open sea, respectively. The average highest concentration was found in a harbor (geometric mean 1.53 μ g L⁻¹), the slightly lower concentration was observed in an estuary (geometric mean value 1.49 μ g L⁻¹) and the lowest concentration was found in the open sea (0.68 μ g L⁻¹).

In estuarine systems copper may be removed from solution by adsorption onto suspended particles which are deposited and accumulate in sediments. The bio-availability of copper increases with decreasing salinity due to the reduced competition from Ca and Mg for uptake sites (Wright and Zamuda 1987). The sea weed *Fucus vesiculosus* has the capability of desorbing and accumulating Cu adsorbed onto particles of suspended sediment (Luoma and Bryan 1982). Estuarine sediments are thought to be the most important depositional sites for particulate copper transported from rivers, although re-mobilization may occur when sediment is disturbed. Copper is removed from oceans' over time through the formation of sediments by the help of several biological and physiological processes (Martin and Windom 1991) or by particulate settling.

2.10.1 Effects of Copper on Organisms

Cu at low concentrations is essential for living organisms as it maintains cellular functions and it plays an important role as a cofactor for different metabolic enzymes (Monteiro et al. 2009). However at higher concentrations free cupric ions have been indicated as the most toxic forms of Cu present in the aquatic environment for marine organisms (Nor 1987), as they are supposed to damage intracellular proteins which lead towards apoptotic cell death (Monteiro et al. 2009). Other studies (USEPA 1985; James et al. 2008; Jorge et al. 2013) revealed a strong correlation between the concentration of Cu in the water body and physiological changes such as a lag in growth in the organisms they studied. Further, Cu can influence negatively the reproduction process of different marine organisms (USEPA 1985; James et al. 2008) as well as the community structure of phyto – and zooplankton (Moore and Winner 1989). However, Jorge et al. (2013) have verified adaptive or compensatory mechanisms when organisms are exposed even to higher concentrations of Cu for longer terms. Furthermore, the toxicity of Cu can be mitigated by the presence of humic matter in the aqueous environment as it complexes Cu (Nor 1987).

2.11 Manganese

Manganese is a vital micronutrient for marine organisms and plays a significant role in photosynthesis (Horsburgh et al. 2002; Kernen et al. 2002). It is also associated with some proteins and enzymes which regulate physiological functions (Frausto da

Silva and Williams 1991). However, a higher concentration of dissolved Mn is helpful to reduce the toxicity of copper (Sunda and Huntsman 1983; Stauber and Florence 1985), cadmium and zinc (Sunda and Huntsman 1996).

The uses of manganese show an increasing trend and recent yearly data account for more than 1,5,000,000 ton (Zhang 2007). Various forms of manganese are present in water such as Mn(IV) oxides, Mn(III) oxyhydroxide and Mn(II) (Post 1999). In marine oxygenated environments, manganese (IV, III) oxides exist as insoluble forms, whereas Mn(II) is available in soluble or dissolved condition. Marine sediments have an aerobic surface enriched with Mn (III, IV) phases. When these oxides enter into deeper layers become reduced to soluble Mn (II) (Burdige 1993). Dissolved Mn (II) may move towards the oxidized zone and precipitate again as Mn (III, IV) (Sundby and Silverberg 1985). In anoxic sediments, Mn is primarily available as Mn(II) carbonates and in solid solution with CaCO₃.

The sediment concentration of Mn in Galician rias (north-west Spain) was reported to vary from 93 to 500 mg kg⁻¹ (Barreiro and Carballeira Real 1994; Belzunce-Segarra et al. 1997). Hartwell et al. (2000) conducted a research in Chesapeake Bay, USA and found that the total manganese level in sediment was from 940 to 2,400 mg kg⁻¹, dry weight. The total manganese level in the northern Adriatic Sea (Italy) ranged from 200 to 800 mg kg⁻¹ dry weight, with an average value of 370 mg kg⁻¹ dry weight (Fabbri et al. 2001). The average manganese concentrations in surface sediments of the Baltic Sea were 3,550 (Bothnian Sea), 5,070 (Gulf of Finland) and 8,960 (Bothnian Bay) mg kg⁻¹ dry weight. The concentrations of dissolved Mn in sediment pore waters were in the range 0.2-24 mg L⁻¹ (Eriksson and Baden 1998), whereas it fluctuated from 0.2 to 17 μ g L⁻¹ (Hall et al. 1996) in bottom water. Bryan and Hummerstone (Bryan and Hummerstone 1973b) measured manganese concentrations in seaweeds from south-west England varying from 128 to 393 mg kg^{-1} (geometric mean), whereas manganese concentrations were 350 and 29 mg kg⁻¹ in a seaweed of North Sea (Fucus vesiculosus) and mussels (Mytilus edulis), respectively (Struck et al. 1997). The concentration of manganese in lobsters living on sediments was 92 mg kg⁻¹ of body weight (Eriksson 2000).

In anoxic conditions the concentration of manganese in Chesapeake Bay, USA, was 237 μ g L⁻¹ (Eaton 1979), whereas it was approximately 700–800 μ g L⁻¹ in anoxic bottom water in the Baltic Sea (Kremling 1983). Roitz et al. (2002) conducted a research on the distribution of dissolved manganese within the San Francisco Bay and found manganese concentrations in the range 12–210 nM at the freshwater interface, from 35 to 756 nM at intermediate salinities and from 86 to 435 nM at the oceanic interface. Tueros et al. (2008) determined concentration of dissolved Mn according to the ranges in salinity in estuary and coastal waters within the Basque Country (northern Spain). The median value of Mn within the polyhaline range (18–50 PSU) was 5.8 μ g L⁻¹ while in the euhaline ranges (salinity \leq 0.5 PSU) the value of Mn was 1 μ g L⁻¹.

Dissolved manganese is particle reactive and may be removed from the water column by sorption onto inorganic and organic particles (Hunt 1983), bacteria

(Sunda and Huntsman 1990) and phytoplankton (Brand et al. 1983; Sunda and Huntsman 1996). However, there is also evidence of desorption of dissolved manganese from riverine suspended particles at the freshwater/seawater interface (Li et al. 1984; Yang and Sañudo-Wilhelmy 1998). The main sources of dissolved Mn in estuary are fluvial and atmospheric inputs, point and non-point source inputs and benthic fluxes from sediments.

Dissolved Mn shows a complex behavior in the estuarine mixing water zone where redox conditions play a vital role in the sedimentation, solubilization and dissolved-particulate partitioning processes. One of the main sources of dissolved Mn in the water column is the re-dissolution of Mn (II) ions (Laslett and Balls 1995). Desorption from suspended particulate matter (SPM) is favored in an environment with low dissolved oxygen and low medium salinity which also contributes for the formation of stably-dissolved Mn (III) chloride and sulphate complexes (Owens and Balls 1997). Mn (II) can be removed from the coastal areas through oxidative processes and the precipitation of Mn (IV) hydroxides (Laslett 1995; Nolting et al. 1999). However, the removal of Mn (II) has also occurred in estuarine reaches where dissolved oxygen increased. Changing of solubility of Mn ions due to the Fe (II)/Fe (III) solubilisation-precipitation processes affect the dissolution and co-precipitation of the other metals such as nickel and zinc (Laslett 1995; Audry et al. 2007). Generally the highest concentration was found in mesohaline-polyhaline ranges, whereas the concentration decreased gradually in the outer estuary and the coastal zone (Kraepiel et al. 1997; Nolting et al. 1999; Audry et al. 2007).

2.11.1 Effects of Manganese on Organisms

Manganese is an essential trace element which is involved in many metabolic processes in plants and animals, with the problem that an excess of the bioavailable ionic form Mn²⁺ can cause neurotoxic effects on marine organisms (Martin et al. 2008). Oweson and Hernoth (2009) detected immunotoxic effects of Mn in crustaceans. Mn was accumulated in the glands and the blood of the animals. This reduced the bactericidal capacity of the organisms, making them more susceptible to different infections. However, Mn was eliminated from the organism back into the water body when the Mn concentration of the latter decreased. Different aquatic organisms are widely used as bioindicators for the Mn concentrations prevailing in an aqueous environment (Steenkamp et al. 1994; Baden et al. 1999; Norwood et al. 2006). Mn-oxidizing bacteria also exist (Tyler 1970) in the aquatic environment. The resulting compounds can be faster taken up by organisms which use the oxidized Mn as energy source in their growth process (Kepkay and Nealson 1987).

2.12 Arsenic

Arsenic has received extensive attention during the last decades due to its relative mobility over a wide range of redox conditions, carcinogenic properties and also its occurrence in the aquatic environment (Wang et al. 2012). Arsenic is a ubiquitous element on earth with metalloid property and complex chemistry. In aquatic systems, arsenic can be originated from both natural and anthropogenic sources such as volcanism, weathering, as by-product of mining, metal refining process, burning of fossil fuels, wood preservation, herbicides, pesticides, fungicides, cement works, electronics industries, ammunition factories, chemistry (dyes and colors, wood preservatives, pesticides, pyrotechnics, drying agent for cotton, oil and dissolvent recycling), pharmaceutical works (medication) (Savory and Wills 1984; Pacyna 1987; Winter 1998; Cullen and Reimer 1989).

Arsenic is found in the atmosphere mainly in four oxidation states such as arsenate (As^V), arsenite (As^{III}), arsenic (As^0), and arsine (As^{III-}) (Sharma and Sohn 2009). In Earth, arsenic is commonly disseminated as the arsenides of copper, nickel, and iron, or as arsenic sulfide or oxide. Estuaries are zones between terrestrial and marine ecosystems and play important role in arsenic cycling (Maher 1984; Bone et al. 2006). The features of the estuaries change with the changes in salinity and redox potential (Wang et al. 2012).

However, arsenic (III) oxide is generated due to copper smelting, whereas methylated arsenic compounds are produced due to biological activity. The noxiousness of As in the aquatic system depends on its speciation and concentration. In water, arsenate and arsenite are more dominant than other forms of arsenic. Arsenate is predominant in oxygenated water, whereas arsenite is predominant in a reduced condition. Arsenic is transported from its sources to estuaries and marine system via atmospheric deposition and riverine input. Marine microorganisms are responsible for the transformation of inorganic arsenic into more complex organic compounds such as arsenobetaine, arsenocholine and arsoniumphospholipids.

Aquatic micro-organisms such as phytoplankton and bacteria keep a significant role in arsenic speciation, distribution and cycling in aquatic ecosystems (Howard et al. 1995; Hasegawa et al. 2001; Hellweger and Lall 2004; Sharma and Sohn 2009). Phytoplankton species are responsible for the conversion of inorganic arsenic into methylarsenicals and high order organic arsenic compounds such as arsenosugars, whereas bacteria play an important role for the mineralization of organic arsenic to methylarsenicals (Francesconi and Edmonds 1996; Hanaoka et al. 1995). Aquatic organisms keep and transform arsenic species inside their body after being exposed to arsenic from food or other sources such as water and sediments (Edmonds et al. 1997; Hasegawa et al. 2001). The ability of microorganisms to accumulate, transform, detoxify, and modify the arsenic speciation depends on several parameters such as the phosphate concentration, pH, redox potential, salinity, temperature, and light intensity (Karadjova et al. 2008).

The average concentration of arsenic in marine water is less fluctuating than in freshwaters (Neff 1997; Smedley and Kinniburgh 2002). The mean concentration

of arsenic in marine water is 1.5 mg L^{-1} . The average concentrations of As in Pacific coast near Nakaminato (Ibaraki, Japan) and Onagawa (Miyagi, Japan) were 3.1 µg L^{-1} and 0.6 µg L^{-1} , respectively (Ishikawa et al. 1987). In deep Pacific and Atlantic waters it ranges from 1.0 to 1.8 mg L^{-1} (Cullen and Reimer 1989). However, the concentrations of arsenic in estuaries are much more uniform than in open waters. Arsenic concentration in estuaries might be influenced by effluents from industry and mining activities, bioactivities of aquatic organisms and geothermic water (Smedley and Kinniburgh 2002). The distribution of arsenic is influenced by thermal stratification (Hasegawa 1996). Arsenic was detected uniformly distributed in the water column in Vestfjord estuary in Norway, where the value ranged from 0.75 to 1.05 mg L^{-1} (Abdullah et al. 1995).

Physical mixing of the fresh and seawater masses and salinity may influence the concentration of dissolved arsenic in estuaries and continental shelves. There is a linear relationship between arsenic concentration and salinity. For instance, Seyler and Martin (1991) reported a linear increase in total arsenic concentrations varying from 0.13 mg L⁻¹ in freshwater to 1.8 mg L⁻¹ in offshore waters, with the increase in salinity in Krka Estuary, Yugoslavia. Redox potential (Eh) and pH are the most important factors controlling arsenic speciation. Under oxidizing conditions, $H_2AsO_4^-$ is dominant at low pH (less than about pH 6.9), whilst at higher pH, $HAsO_4^{2-}$ becomes dominant. H_3AsO_4 and AsO_4^{3-} may be present in extremely acidic and alkaline conditions respectively. Under reducing conditions at pH less than about pH 9.2, the uncharged arsenite species $H_3AsO_3^0$ will predominate (Brookins 1988).

Arsenic can be removed from the estuary in several ways. Although As can be scavenged from the water column by the particulate matter, adsorption of As onto iron (and to a lesser extent Mn) oxyhydroxides may be a dominant way of removal (Crecelius et al. 1975; Langston 1983). There is a decreasing trend of arsenic from estuary to ocean due to flocculation of iron oxides with increased pH and salinity (Cullen and Reimer 1989).

2.12.1 Effects of Arsenic on Organisms

Arsenic is included in the priority list of dangerous substances established by ATSDR (2007) as it shows a high ability to accumulate in lipid tissues of organisms and can be transported easily upwards in the trophic chain. It can be found in water bodies as As(V), As(III), As(0), As(III-). Among these ionic forms, As(III) and As (V) are the most toxic ones. Those compounds derive from contaminated water and/or sediments and can be possibly biomagnified within the aquatic food web (Rahman et al. 2012 and references therein). In mammals, the poisoning can be acute (e.g. gastroenteritis) and chronic (e.g. effects on the reproductive – and the immune system) (Norwood et al. 2007). Also in fish, chronic poisoning of As can cause problems in the reproduction process as well as in the development of the young fish at concentrations as low as 1 ppb (Das et al. 1980). Major amounts (85–90 %) of arsenic compounds were found in edible portions of fish, whereas a

smaller amount (5–44 %) was detected in lipids of macroalgal species. The concentrations of arsenic differ significantly in phytoplankton and microalgae. Generally, brown algae contain higher amount of total arsenic (around 230 mg g⁻¹ d. w.) than green (approximately 23.3 mg g⁻¹ d. w.) and red algae (approximately 38 mg g⁻¹ d. w.) (Thomson et al. 2007). There are two different assumptions about the degradation of arsenic species: the first one is that As (V) is taken up mainly by phytoplankton and reduced to As (III), while the second one suggests that As(III), taken up by phytoplanktonic organisms, is converted to As(V) and to methylated forms of As which are further eliminated by excretion (Phillips 1990). Those methylated forms are in turn degraded by the action of bacteria, forming again the more bioavailable form As(V) (Rahman et al. 2012).

2.13 Tin and Organotin Compounds (Tributyltin and Triphenyltin)

The environmental significance of tin is mostly related to its organometallic derivatives, namely Organotin compounds (OTs) which are extensively used in the industrial sectors because of their special properties such as PVC stabilizers and catalysts, as biocides in agriculture and as antifouling agents in marine paints to limit the growth of aquatic organisms on ship hulls, fish cages, oil rig supports etc. Even though some natural methylated forms of tin are known, the impact of organotin compounds in marine ecosystems derives mainly from anthropogenic activities (Amouroux et al. 2011). Scientists are globally concerned about organotin due to their ubiquitous presence (at nanomolar concentration in water) and accumulation in aquatic organisms (Atanasov et al. 2005; Hu et al. 2006). Guruge et al. (1996) reported an annual use in 1992 of 50,000 tons, whereas in 1986 it was 35,000 tons. During the last 60 years, significant amounts of organotins have been added into various ecosystems (Hoch 2001). Thirty six organotin compounds are harmful to the natural resources out of 260 known compounds (Eisler 1989). Various factors control the solubility of organotin compounds such as oxidation/ reduction potential, pH, temperature, ionic strength, and concentration and composition of the dissolved organic matter (USEPA 2003).

Organotin compounds are persistent in the aquatic environment because of their slow degradation rate and consistent flux (Michel and Averty 1999). Among the organotin compounds, tributyltin (TBT) and triphenyltin (TPT) are the most hazardous and largest discharged compounds into the marine environment. Although triphenyltin (TPT) is mainly used in agriculture such as fungicide and insecticide, it is also used in marine paint formulations. On the contrary, tributyltin (TBT) is widely used in antifouling agent in boat paints, in wood preservation, as an antifungal agent for textile and in industrial water systems (Donard et al. 2001). TBT is more hazardous than TPT and causes chronic and acute toxic effect to aquatic organisms such as zooplankton, algae, molluscs and fishes at larval stage (Gibbs and Bryan 1996). However, OT compounds are recognized as priority substances in the EU directive on environmental quality standards in the area of the water framework policy (EU 2008). In 2008, the total ban on using harmful organotins in antifouling paints was introduced (EU 2003; IMO 2001).

Tributyltin and triphenyltin are ionizable and make complexes by reacting with different ligands present in the environment (Arnold et al. 1997; Buck et al. 2003). The fate and accumulation of the organotion compounds are determined by the speciation of organotin compounds. In water, tributyltin and triphenyltin exist as various states such as neutral, non-dissociated molecules (triorganotin hydroxide and triorganotin chloride) and/or as organotin cations. Speciation is normally regulated by the pH and salinity of the water. In seawater the hydroxyl-complexes (>93 %) are the dominant species, whereas under acidic medium (pH 6), the TBT-cation exists in addition to the neutral TBT-Cl and TBT-OH complexes (Arnold et al. 1997; Burton et al. 2005)

2.13.1 The Behavior and Distribution in the Environment

The biological effects of the TBT and TPT are mainly determined by the distribution of the compounds in the environment, timescale and environmental conditions where they exist. However, the environmental distributions depend on the physical, chemical and biological properties of these compounds and the myriad of the localized conditions.

TBT and TPT are hydrophobic in nature and quickly partition into water-borne particles and their aqueous solubility depend on salinity, pH and temperature (Hoch 2001). After partitioning in water, OTs are trapped in the sediments. These may undergo resuspension into the water column and OTs are returned back into the water. This accumulation acts as a temporary sink and sediments are also a long-term source of dissolved-phase contamination (Hoch 2001; Donard et al. 2001). Organotin compounds can be accumulated at the surface layer of air-water interface due to their hydrophobic characters and may be quickly lost due to volatilization and UV degradation (Adelman et al. 1990).

2.13.2 Bioaccumulation and Biotransformation

Organotin compounds are accumulated significantly by aquatic species while some species degrade or eliminate the effects of bioaccumulation because of having an efficient enzymatic mechanism (Laughlin 1996). Biomethylation is the main process of biotransformation of the organotin compounds, whereas chemical agents such as methylcobalamin (it transfers a methyl group to metals), methyl iodide (it is produced by microalgae and reacts with metals and metal sulfides), humic and fulvic acid (produced due to decomposition of vegetation) and other methyl metals can transmethylate OTs (Hamasaki et al. 1995). Although methylation or biomethylation can engage sediment-or estuary –containing microorganisms,

Table 2.1 Estimated half-lives of TBT in various environmental matrixes	Environmental matrixes	Half-life
	Freshwater Estuarine water	6–26 days (light), 4 months (dark) 1–2 weeks
	Sea water	6–127 days
	Water/sediment mixture	5 months–5 years
	Estuarine sediment	3.8 years
	Soil	4–5 months (TPhT)
	Adapted from Cima et al. (2003)	

sulfate-reducing bacteria and yeasts are predominant in methylating organotins. Organotin compounds are degraded through different processes such as photolysis, biological degradation and chemical cleavage. The degradation rates of organotin compounds can be evaluated from its half-life. Table 2.1 shows the half-life values of TBT at different environmental matrixes.

2.13.3 Effects of Tributyltin (TBT) and Triphenyltin (TPT) on Organisms

Tributyltin and Triphenyltin show adverse effects on the survival, growth and reproduction processes of different aquatic organisms (USEPA 2003; Arizzi Novelli et al. 2002a). Further, (Duft et al. 2003) tested the carcinogenicity of TBT on some organisms. They influence the reproduction process acting as endocrine disrupters causing imposex phenomena in many adult aquatic organisms, e.g. gastropods and daphnids (USEPA 2003 and references therein), sea- and mudsnails and mollusks (Duft et al. 2003; Arizzi Novelli et al. 2002b), Nassarius nitidus (Pavoni et al. 2007). The action of tin-compounds alone can introduce those adverse effects in aquatic organisms, but the presence of some other organic or inorganic compounds can improve the effects (Pavoni et al. 2007 and references therein). Further, Arizzi Novelli et al. (2002a, b) tested the toxicity of TBT and TPT on embryos of sea urchin and their sperm cells, with the results of high poisoning symptoms to phenyltin and butyltin, respectively. In fish, TBT inhibits the gonad development and it reduces the thymus as well as the number of lymphocytes. In mammals, the immune system is suffering from a poisoning in TBT and TPT as well as the reproductive one (Sekizawa et al. 2001 and references therein). As organisms of the higher trophic chain have appropriate detoxifying enzymes, TBT and TPT seem to be toxic only for lower organisms (Arizzi Novelli et al. 2002b). Nevertheless, it can have lethal effects on several aquatic organisms, such as crustaceans, algae, echinoderma and fish (Dimitriou et al. 2003 and references therein).

3 The Lagoon of Venice as a Case of Study of Metal Contamination

The international scientific literature offers a large number of studies focusing on the presence, dynamics and cycling of various polluting and harmful elements in both estuarine and coastal environments. Among these, the pollution of the Lagoon of Venice (Northeastern Italy) has received increasing attention from various scientific and socio-economical authorities in the past two decades. The reasons are principally due to its geomorphological complexity and the anthropogenic pressure. This includes the industrial impact and the intensive lagoon use for tourism and fishing purposes. Due to the international significance, Venice and its lagoon were included in the World Cultural Heritage list (UNESCO 1987) and, at present, it hosts 2.5 million tourists per year staving more than six million nights (Comune di Venezia 2012). Scientific studies, have focused on the physical, chemical, biological and ecological aspects of the Lagoon of Venice. Most studies evidenced that it suffers from numerous and worrying environmental and social problems arising from geological features, climate change, economical causes and many kinds of anthropogenic pressures. Among those, the increased frequency and intensity of flooding events, caused by the climate change related sea level rise and land subsidence, certainly represent the main threat for the future of Venice. The debate on this is still far from being conclusive (Fontini et al. 2010; Carbognin et al. 2010). Another controversial issue is related to the high density of anthropogenic pressures, which impacts seriously all the environmental compartments of the lagoon and its ecosystem and cause sediment, water and air pollution, eutrophication phenomena, exploitation of biological resources, degradation of biota, sediment erosion and the consequent salt marsh losses. Since the pollution of the Lagoon of Venice was largely studied in the last decades, it represents an ideal case of study for transitional environments. Various pollutants from different pathways and from both diffuse and point sources have polluted the lagoon. Solidoro et al. (2010) reported a comprehensive overview of the responses of the lagoon ecosystem to both natural and anthropogenic pressures over the last 50 years. Thereafter, some key findings of recent researches are reported and subdivided by pollution sources and environmental issues. This section provides a quick overview of the studies conducted on the lagoon and highlights the major issues addressed in the last 20 years.

3.1 Source of Contamination: Industrial Zone of Porto Marghera

The direct discharges from the industrial zone of Porto Marghera are largely recognized as the main source of the pollution for the Lagoon (e.g., (Pavoni et al. 1988; Frignani et al. 1997; Bernardello et al. 2006; Carrer and Leardi 2006;

Zonta et al. 2007)). Porto Marghera is one of the most important industrial districts of Italy and includes chemical and metallurgical activities, oil-refining and storage, shipbuilding, incineration and thermoelectric power plants burning coal, gas and refuse derived fuel, wastewater treatments, hazardous waste incineration and other factories. It was built on the border of the lagoon starting from 1917 with activities related to oil and coal and largely grew in the following 50 years, reaching a maximum of 31,000 employees and a surface area up to 20 km² in the early 1970s. Today, following the reduction of the industrial activity, the employees have decreased to ~12,000, but the area of Porto Marghera probably is still the main source of contamination for the lagoon. In addition to the main industrial zone, the island of Murano hosts small and mid-size glass-making factories having an impact on the lagoon and the atmosphere (Giusti and Polo 2002; Rampazzo et al. 2008). All these inputs resulted in this sediment pollution near the main industrial areas.

A large number of studies, including the older ones, indicated that the industrial zone of Porto Marghera was the main responsible of elevated concentrations of heavy metals in the Lagoon of Venice. For example, Donazzolo et al. (1984) studied the heavy metals pollution of sediments and attributed it to the industrial area. These findings were followed by a large number of more specific studies. For example, Bellucci et al. (2002) reported maximum values of As, Cd, Hg, Pb and Zn in channels of the oldest part of the industrial area. Bernardello et al. (2006) studied the distribution of various heavy metals in the superficial sediments and pointed out the industrial zone of Porto Marghera as the main source of contamination. They also reported maps of pollutant concentrations for the inner part of the lagoon. Coccioni et al. (2009) indicated that the lagoon is affected by trace element pollution (Cd, Cu, Ni, Pb, Zn and Hg) with the highest concentrations in its inner part, which corresponds to the Porto Marghera industrial area, and also revealed a strong relationship between trace elements (in particular Mn, Pb and Hg) and the occurrence of abnormalities in tests with foraminifera. In addition, a study by (Sommerfreund et al. 2010) showed that the current industrial discharges are the main cause of the pollution nearby the industrial area, but they also pointed out the role of tributary loadings as a source of contaminants to the lagoon. Apitz et al. (2009) used statistical methods to separate natural background and anthropogenic concentrations of trace elements in radio-chronologically selected surface sediments of the Venice Lagoon. Han et al. (2011) studied the relocation effects of dredged marine sediments on mercury geochemistry and found that organic matter play a key role in the control of methylation rates via the control of Hg solubility.

Also the role of glass-making factories on the Island of was assessed, but the extent of studies was largely lower. For example, Giusti and Zhang (2002) described the trace element distribution in sediments, marine water and mussels of the Venetian Lagoon around Murano. They found that sediments are contaminated with many elements (Zn, Cu, Ag, As and Pb) with the highest levels in the fine fraction (pelite, <63 μ m), which was the main carrier of heavy metals in most sites.

Some studies have also reported on the cycling, transport and the accumulation of many elements in the finest sediments. For example, Martin et al. (1995)

highlighted the significance of colloids in the biogeochemical cycling of some trace metals, whereas Zonta et al. (1994) showed a strong correlation between heavy metals and finer particles content in the sediment (d < 7.8 μ m) in a study conducted in a salt marsh. Perin et al. (1997) measured the bio-available and non-bio-available phases of some heavy metals in a channel near the industrial zone. They found that the inner canals were very polluted with heavy metals in the form of highly insoluble sulphides.

3.2 Sources: Urban Settlements

The urban agglomerations include the historical part of Venice on the main lagoon island, and the mainland urban settlement of Mestre (a total of 270,000 inhabitants) across the causeway, with the surrounding inland areas. The population dynamics of the historical center had a population peak during the 1950s, before declining to $\sim 112 \cdot 10^3$ in the early 1970s and to $66 \cdot 10^3$ in the 2000s. The sewer system is normally organized for the mainland, whereas for the islands it is not conventionally structured with some leaks of untreated domestic sewage into the lagoon. There are no more major industries and factories in Venice historical centre and the direct role of sewer systems discharges in heavy metals pollution of the lagoon can be of minor importance. However, a significant indirect effect on the bioavailability of many elements can be supposed.

3.3 Source: Drainage Basin

Rivers, streams and land runoff (Collavini et al. 2005) reach the lagoon. The lagoon has a drainage basin of ~1,850 km², which provides a mean yearly freshwater input of 35.5 m³ s⁻¹ peaking up to 350–600 m³ s⁻¹ during significant adverse meteorological conditions. The main freshwater tributaries, contributing with more than 50 % of the total annual load (Zonta et al. 2001; Zuliani et al. 2005, 2001), are few small rivers mainly located in the northern lagoon; some of them are partially canalized. The sediment input associated with these river inflows was estimated to be ~33 × 10³ tons year⁻¹ in the period 1999–2000. This hydrological pattern creates a typical brackish environment with a salinity gradient that ranges from 10 psu near the mainland border to 32–37 psu at the inlets (Molinaroli et al. 2007) with summer peaks over 40 psu in the most confined areas of the northern lagoon (Facca et al. 2011). Some studies were carried out to assess the role of drainage basin in the contamination of the lagoon. Among others, the DRAIN project (Collavini et al. 2005) studied the pollutant loads from the 12 major tributaries of the Venice Lagoon during 1998–2000.

3.4 Source: Air Pollution

The atmospheric wet and dry polluting depositions are principally linked to the industrial emission processes (e.g., Guerzoni et al. 2005; Rampazzo et al. 2008; Rossini et al. 2010). Rossini et al. (2010) and Bettiol et al. (2005) analyzed the atmospheric bulk deposition of Venice and estimated the metal fluxes to the lagoon. They indicate that industrial sources have a quasi-permanent effect in the composition of atmospheric depositions.

3.5 Source: Engineering Works

In addition, the continuous intervention of man with hydraulic works and the excavation of the lagoon canals resulted in continuous changes of geomorphology. Among those, the re-construction of stone jetties at the lagoon inlets (1860–1934), the dredging of deep and large canals for navigation purposes (1926 and 1970) and the mechanical and often illegal clam catching (1990s) had wide impacts on many lagoon areas (Guerzoni and Tagliapietra 2006). Starting from the second half of 2000s, the construction of mobile barriers to safeguard the lagoon from exceptionally high tides (MoSE project) has further altered the structure of the lagoon inlets with likely consequences in hydrodynamics, sediment exchanges with the open sea and the whole ecosystem (Ghezzo et al. 2010). A comprehensive chronology of the most relevant events that affected the lagoon morphology in the last century is summarized in Sarretta et al. (2010).

3.6 Effects of Pollution on Organisms

A number of studies were carried out in both animal and plant species to detect the bioaccumulation and the effects of many pollutants in the Lagoon of Venice. Some studies found high levels of metals in both sediments and target gastropods (Berto et al. 2012). Nesto et al. (2007) evaluate levels and effects of trace metals (Cd, Cr, Cu, Fe, Mn, Pb, Zn), PAHs and PCBs in mussels and fish native of the Lagoon of Venice.

Berto et al. (2006) analyzed the levels of mercury in both sediments and Gastropoda *Nassarius reticulates* in the southern Venice Lagoon and stated that a main cause for mercury pollution might be attributed to the antifouling paints used in great quantity in the recent past near the town of Chioggia. Berto et al. (2007) analyzed the recent contamination of TBT and found high content of organotin compounds in the gastropods sampled near the dockyards, harbors, and marinas.

Among other species, the mollusc *Mytilus galloprovincialis* was used in most studies, probably for being edible and largely available in the lagoon. Zatta

et al. (1992) measured the content of heavy metals (Cd, Co, Cu, Cr, Hg, Fe, Mn, Ni, Pb, Zn) and As accumulated in the mollusc Mytilus galloprovincialis from the Venetian lagoon. Widdows et al. (1997) studied the effects of pollution on *Mytilus* galloprovincialis and demonstrated that the inhibitory effects of toxicants on the growth of organisms living in the more polluted inner sites. Da Ros et al. (2000) studied the cellular and biochemical response of mussels transplanted from a relatively pristine site to a polluted one and vice versa and evidenced a direct influence in the thinning of the digestive cells and in the increasing number of lysosomes in mussels from the more polluted site. Giusti and Zhang (2002) described the trace element distribution of the Mytilus galloprovincialis around the Island of Murano. Their results indicated that the highest trace element concentrations were in the soft tissue of mussels. However, the levels detected were within the recommended Italian and international guidelines for human shellfish consumption, except for arsenic. Irato et al. (2003) sampled a series of different bivalve mollusks: Mytilus galloprovincialis, ark clam (Scapharca inaequivalvis), and Japanese littleneck (Tapes philippinarum) in two sites and analyzed the concentrations of essential and nonessential metals in digestive gland and gills of each bivalve. Results indicated that all the three species accumulated metals to a considerable extent, and also evidenced the *M. galloprovincialis* and S. inaequivalvis as sentinel organisms to be used specifically for Zn pollution. Sfriso et al. (2008) investigated the concentration changes of elements (As, Hg, Cd, Cr, Co, Cu, Pb, Mn, Ni, V, Zn, Fe, and Hg) in the soft tissues of Tapes philippinarum during its growth. Arsenic was reported higher in clam tissues than in surface sediments and suspended particles, whereas other elements (Cr, Pb, Mn, Ni, and Fe) were found higher in suspended particles and sediments than in Tapes.

A large number of studies was also conducted in plant organisms, mainly algal species. Bragato et al. (2006) monitored growth dynamics, nutrient and heavy metal accumulation in the two dominating macrophytes *Phragmites australis* and *Bolboschoenus maritimus*, but they found that the heavy metal concentration in water and soil was not correlated to the plant content. Favero et al. (1996) investigated the metal accumulation of *Ulva rigida* and reported that this specie specifically accumulates Fe. Caliceti et al. (2002) measured the concentrations of heavy metals (Fe, Zn, Cu, Cd, Ni, Pb, Cr, As) in seven seaweeds of environmental and commercial relevance. They found high Pb levels in *Ulva* species and to a lesser extent in *Gracilaria*, whereas Brown seaweeds (*Cystoseira*) were highly contaminated by arsenic. Sfriso et al. (1995) reported that the dominant macroalgal species in the central lagoon, *Ulva rigida*, shows a preferential enrichment of Zn and Mn compared to other metals.

4 A Concluding Remark

After the period of the worst industrial impact and eutrophication, which caused anoxic events till the eighties, the lagoon appeared to enter a phase of progressive improvement of environmental conditions. In this process, in addition to a significant reduction of industrial activity and stricter limits to emissions enforced by law, an important role was played by the water exchange with the sea that ensured a water renewal and enabled a removal of the finest most polluted sediments from inside the lagoon out to the sea. However, as a consequence of the building of the MoSE barriers at the lagoon-sea channels, the water exchange is expected to be significantly limited. More research is in progress.

References

- Abdullah MI, Shiyu Z, Mosgren K (1995) Arsenic and selenium species in the oxic and anoxic waters of the Oslofjord, Norway. Mar Pollut Bull 31:116–126
- Achterberg EP, Braungardt C (1999) Stripping voltammetry for the determination of trace metal speciation and in-situ measurements of trace metal distribution in marine waters. Anal Chim Acta 400:381–397
- Achterberg EP, Colombo C, van den Berg CMG (1999) The distribution of dissolved C, Zn, Ni, Co, and Cr in English coastal surface waters. Cont Shelf Res 19:537–558
- Achterberg EP, Holland TW, Bowie AR, Fauzi R, Mantoura C, Wosfold PJ (2001) Review: determination of iron in seawater. Anal Chim Acta 442:1–14
- Adams DH, Sonne C, Basu N, Dietz R, Nam DH, Leifsson PS, Jensen AL (2010) Mercury contamination in spotted seatrout, Cynoscion nebulosus: an assessment of liver, kidney, blood, and nervous system health. Sci Total Environ 408:5808–5816
- Adelman D, Hinga KR, Pilson MEQ (1990) Biogeochemistry of butyltins in an enclosed marine ecosystem. Environ Sci Technol 24:1027–1032
- Ahrland S (1975) Metal complexes in seawater. In: Gold-berg ED (ed) Dahlem workshop on the nature of sea-water Dahlem Konferenzen. Abakon Verlagsgesellschaft, Berlin, pp 219–244
- Alam MK, Manghan OE (1992) The effect of malathion, diazinon and various concentrations of zinc, copper, nickel, lead, iron and mercury on fish. Biol Trace Elem Res 34:225–236
- Amouroux D, Seby F, Monperrus M, Pannier F, Mendiguchia C, Benoit- Bonnemason C, Donard OFX (2011) Chemical species. In: Quevauviller P, Roose P, Verreet G (eds) Chemical marine monitoring: policy framework and analytical trends, 1st edn. Wiley, Chichester, pp 101–160
- Amyot M, Mierle G, Lean D, Mc Queen DJ (1997) Effect of solar radiation on the formation of dissolved gaseous mercury in temperate lakes. Geochim Cosmochim Acta 61(5):975–987
- Andersson MA, Willets A, Allenmark SG (1997) Asymmetric sulfoxidation catalyzed by a vanadium-containing bromoperoxidase. J Org Chem 62:8455–8458
- Antipov AN, Lyalikova NN, Khiznjak TV, L'vov NP (1999) Some properties of dissimilatory nitrate reductases lacking molybdenum and molybdenum factors. Biochemistry (Moscow) 64 (5):483–487
- Anton M, Andrew EA, Sacha C, Alessandra DeM Angela F, Manuela M, Magali S, Marc H, Kamel J, Uma M, Edda R, Assaf V, Kirk EA, John AB, Anthony C, Aubrey KD, Kimberlee T, Massod ZH, Todd WL, Lippmeier JC, Diego M, Micaela SP, Gregory JP, Mak AS, Dan SR, Armbrust EV, Chris B (2007) Identification and comparative genomic analysis of signaling and regulatory components in the diatom Thalassiosira psuedonana. J Phycol 43(3):585–604

- Apitz SE, Degetto S, Cantaluppi C (2009) The use of statistical methods to separate natural background and anthropogenic concentrations of trace elements in radio-chronologically selected surface sediments of the Venice Lagoon. Mar Pollut Bull 58(3):402–414
- Arizzi Novelli A, Argese E, Tagliapietra D, Bettiol C, Volpi Ghirardini A (2002a) Toxicity of tributyltin and triphenyltin to early life – stages of Paracentrotus lividus (Echinodermata: Echinoidea). Environ Toxicol Chem 21(4):859–864
- Arizzi Novelli A, Losso C, Ghetti PF, Volpi Ghirardini A (2002b) Toxicity of heavy metals using sperm cell and embryo toxicity bioassays with Paracentrotus lividus (Echinodermata: Echinodea): comparison with exposure concentrations in the lagoon of Venice, Italy. Environ Toxicol Chem 22(6):1295–1301
- Arnold CG, Weidenhaupt A, David MM, Mu⁻Iler SR, Haderlein SB, Schwarzenbach RP (1997) Aqueous speciation and 1-octanol–water partitioning of tributyl- and triphenyltin: effect of pH and ion composition. Environ Sci Technol 31:2596–2602
- Atanasov AG, Nashev LG, Tam S, Baker ME, Odermatt A (2005) Organotins disrupt the 11(beta)hydroxysteroid dehydrogenase type 2-dependent local inactivation of glucocorticoids. Environ Health Perspect 113:1600–1606
- ATSDR (Agency for Toxic Substances and Disease Registry) (1999) Toxicological profile for lead. US Department of Health and Human Services, Public Health Service, Atlanta, p 465
- ATSDR (Agency for Toxic Substances and Disease Registry) (2007) Agency of toxic substances and disease registry. http://www.atsdr.cdc.gov/. Accessed May 2013
- Audry S, Blanc G, Schafer J, Guerin F, Masson M, Robert S (2007) Budgets of Mn, Cd and Cu in the macrotidal Gironde estuary (SW France). Mar Chem 107:433–448
- Auger Y, Bodineau L, Leclercq S, Wartel M (1999) Some aspects of vanadium and chromium chemistry in the English Channel. Cont Shelf Res 19:2003–2018
- Baden SP, Eriksson SP, Gerhardt L (1999) Accumulation and elimination kinetics of manganese from different tissues of the Norway Lobster *Nephrops norvegicus* (L.). Aquat Toxicol 46(2): 127–137
- Baldi F (1997) Microbial transformation of mercury species and their importance in the biogeochemical cycle of mercury. In: Sigel A, Sigel H (eds) Metal ions in biological systems – mercury and its effects on environment and biology, vol 34. Marcel Dekker Inc., New York, pp 213–248
- Balls PW (1985) Copper, lead and cadmium in coastal waters of the western North Sea. Mar Chem 15(4):363–378
- Barciela-Alonso MC, Tubio-Franco MC, Prego R (2003) Nickel and cobalt determination in marine sediments by electrothermal atomic absorption spectrometry and their distribution in the Ria of Ferrol (NW Spain). Mar Pollut Bull 46:1504–1515
- Barkay T, Wagner-Dobler I (2005) Microbial transformations of mercury: potentials, challenges, and achievements in controlling mercury toxicity in the environment. Adv Appl Microbiol 57:1–52
- Barkay T, Miller SM, Summers AO (2003) Bacterial mercury resistance from atoms to ecosystems. FEMS Microbiol Rev 27:355–384
- Barreiro RA, Carballeira Real C (1994) Heavy metals in sediment cores from NW Spain estuary. Bull Environ Contam Toxicol 53:368–373
- Bay SM, Zeng EY, Lorenson TD, Tran K, Alexander C (2003) Temporal and spatial distributions of contaminants in sediments of Santa Monica Bay, California. Mar Environ Res 56:255–276
- Beckvar N, Field J, Salazar S, Hoff R (1996) Contaminants in aquatic habitats at hazardous waste sites: mercury. NOAA technical memorandum, NOS ORCA 100, Washington
- Bellucci LG, Frignani M, Paolucci RM (2002) Distribution of heavy metals in sediments of the Venice Lagoon: the role of the industrial area. Sci Total Environ 295(1):35–49
- Belzunce-Segarra MJ, Bacon JR, Prego R, Wilson MJ (1997) Chemical forms of heavy metals in surface sediments of the San Simón inlet, Ria de Vigo, Galicia. J Environ Sci Health A 32: 1271–1292

- Benoit JM, Gilmour CC, Mason RP, Riedel GF, Riedel GS, Sullivan K (1998) Behavior of mercury in the Patuxent River Estuary, MD, USA. Biogeochem 40:249–265
- Bernardello M, Secco T, Pellizzato F, Chinellato M, Sfriso A, Pavoni B (2006) The changing state of contamination in the Lagoon of Venice. Part 2: Heavy metals. Chemosphere 64:1334–1345
- Berto D, Giani M, Covelli S, Boscolo R, Cornello M, Macchia S, Massironi M (2006) Mercury in sediments and *Nassarius reticulates (Gastropoda Prosobranchia)* in the southern Venice Lagoon. Sci Total Environ 368(1):298–305
- Berto D, Giani M, Boscolo R, Covedis S, Giovanardi O, Massironi M, Grassia L (2007) Organotins (TBT and DBT) in water, sediments, and gastropods of the southern Venice lagoon (Italy). Mar Pollut Bull 55(10–12):425–435
- Berto D, Brusa RB, Cacciatore F, Covelli S, Rampazzo F, Giovanardi O, Giani M (2012) Tin free antifouling paints as potential contamination source of metals in sediments and gastropods of the southern Venice lagoon. Cont Shelf Res 15:34–41
- Bettiol C, Collavini F, Guerzoni S, Molinaroli E, Rossini P, Zaggia L, Zonta R (2005) Atmospheric and riverine inputs of metals, nutrients and persistent organic pollutants into the lagoon of Venice. Hydrobiology 550(1):151–165
- Bilinski H, Kozar S, Plavsic M, Kwokal Z, Branica M (1991) Trace metal adsorption on inorganic solid phases under estuarine conditions. Mar Chem 32:225–233
- Bone SE, Gonneea ME, Charette MA (2006) Geochemical cycling of arsenic in a coastal aquifer. Environ Sci Technol 40(10):3273–3278
- Borja A, Bald J, Franco J, Larreta J, Muxika I, Revilla M, Rodriguez JG, Solaun O, Uriarte A, Valencia V (2009) Using multiple ecosystem components, in assessing ecological status in Spanish (Basque Country) Atlantic marine waters. Mar Pollut Bull 59(1–3):54–64
- Bothner MH, Casso MA, Rendigs RR, Lamothe PJ (2002) The effect of the new Massachusetts Bay sewage outfall on the concentrations of metals and bacterial spores in nearby bottom and suspended sediments. Mar Pollut Bull 44:1063–1070
- Boutron CF, Go"rlach U, Candelone JP, Bolshov MA, Delmas RJ (1991) Decrease in anthropogenic lead, cadmium and zinc in Greenland snows since the late 1960s. Nature 353:53–156
- Boyle EA, Huested SS, Grant B (1982) The chemical mass balance of the Amazon plume, II. Copper, nickel and cadmium. Deep-Sea Res 29(1355):1364
- Bragato C, Brix H, Malagoli M (2006) Accumulation of nutrients and heavy metals in Phragmites australis (Cav.) Trin. Ex steudel and Bolboschoenus maritimus (L.) Palla in a constructed wetland of the Venice lagoon watershed. Environ Pollut 144:967–975
- Brand LE, Sunda WG, Guillard RRL (1983) Limitation of marine phytoplankton reproductive rates by zinc, manganese, and iron. Limnol Oceanogr 28:1182–1198
- Bratkič A, Ogrinc N, Kotnik J, Faganeli J, Zagar D, Yano S, Tada A, Horvat M (2013) Mercury speciation driven by seasonal changes in a contaminated estuarine environment. Environ Res 125:171–178
- Breteler RJ, Valiela I, Teal JM (1981) Bioavailability of mercury in several north-eastern U.S. Spartina ecosystems. Estuar Coast Shelf Sci 12:155–166
- Bricker SB, Longstaff B, Dennison W, Jones A, Boicourt K, Wicks C, Woerner J (2008) Effects of nutrient enrichment in the nation's estuaries: a decade of change. Harmful Algae 8(1):21–32
- Brookins DG (1988) Eh-pH diagrams for geochemistry. University of Michigan, Michigan
- Brown B, Ahsanullah M (1971) Effect of heavy metals on mortality and growth. Mar Pollut Bull 2 (12):182–187
- Bruland KW, Rue EL (2002) Analytical methods for the determination of concentrations and speciations of iron. In: Turner DR, Hunter KA (eds) The biogeochemistry of iron in seawater. Wiley, New York, pp 255–289
- Bruland KW, Franks RD, Knauer GA, Martin JH (1979) Sampling and analytical methods for the determination of copper, cadmium, zinc and nickel at the nanogram per liter level in sea water. Anal Chim Acta 105:233–245

- Bryan GW (1985) Bioavailability and effects of heavy metals in marine deposits. In: Ketchum BH, Capuzzo JM, Burt WV, Duedall IW, Park PK, Kester DR (eds) Wastes in the ocean, vol 6: Nearshore waste disposal. Wiley, New York, pp 42–79
- Bryan GW, Gibbs PE (1983) Heavy metals in the Fal estuary, Cornwall: a study of long-term contamination by mining waste and its effects on estuarine organisms, Marine Biological Association of the United Kingdom, Occasional publication, No. 2. Marine Biological Association of the United Kingdom, Plymouth, 112 pp
- Bryan GW, Hummerstone LG (1973a) Adaptation of the polychaete *Nereis diversicolor* to estuarine sediments containing high concentrations of zinc and cadmium. J Mar Biol Ass UK 53(4):839–857
- Bryan GW, Hummerstone LG (1973b) Brown seaweed as an indicator of heavy metals in estuaries in south-west England. J Mar Biol Ass UK 53:705–720
- Bryan GW, Langston WJ (1992) Bioavailability, accumulation and effects of heavy metals in sediments with special reference to United Kingdom estuaries: a review. Environ Pollut 76:89–131
- Bryan GW, Langston WJ, Hummerstone LC, Burt GR (1985) A guide to the assessment of heavymetal contamination in estuaries using biological indicators, Journal of Marine Biological Association of UK, Occasional publication, No. 4. Marine Biological Association of the UK, Plymouth, 92 pp
- Buck B, Mascioni A, Que JL, Veglia G (2003) Dealkylation of organotin compounds by biological dithiols: toward the chemistry of organotin toxicity. J Am Chem Soc 125:13316–13317
- Burdige DJ (1993) The biogeochemistry of manganese and iron reduction in marine sediment. Earth Sci Rev 35:249–284
- Burton ED, Philips IR, Hawker DW (2005) In situ partitioning of butyltin compounds in estuarine sediments. Chemosphere 59:585–592
- Butler A, Carter-Franklin JN (2004) The role of vanadium bromoperoxidase in the biosynthesis of halogenated marine natural products. Nat Prod Rep 21:180–188
- Caliceti M, Argese E, Sfriso A, Pavoni B (2002) Heavy metal contamination in the seaweeds of the Venice lagoon. Chemosphere 47(4):443–454
- Callister SM, Winfrey MR (1986) Microbial methylation of mercury in upper Wisconsin River sediments. Wat Air Soil Pollut 29:453–465
- Campbell JA, Chan EYL, Riley JP, Head PC, Jones PD (1986) The distribution of mercury in the Mersey estuary. Mar Pollut Bull 17(1):36–40
- Carbognin L, Teatini P, Tomasin A, Tosi L (2010) Global change and relative sea level rise at Venice: what impact in term of flooding. Clim Dyn 35(6):1039–1047
- Carrer S, Leardi R (2006) Characterizing the pollution produced by an industrial area. Chemometric methods applied to the Lagoon of Venice. Sci Total Environ 370:99–116
- CCREM (Canadian Council of Resource and Environmental Ministers) (1987) Canadian water quality guidelines. Inland Waters Directorate, Environmental Canada, Ottawa
- Chakraborty P, Babu PVR, Sarma VV (2012) A study of lead and cadmium speciation in some estuarine and coastal sediments. Chem Geol 294(295):217–225
- Chau, YK, Wong PTS, Bengert GA, Dunn JL (1984) Determination of dialkyllead, trialkyllead, tetralkyllead and lead (II) compounds in sediment and biological samples. Anal Chem 56:271–274. dberg, Plenum Press, New York/London, pp 671–697
- Ci Z, Zhang X, Wang Z, Niu Z (2011) Phase speciation of mercury (Hg) in coastal water of the Yellow Sea, China. Mar Chem 126:250–255
- Cima F, Craig PJ, Harrington C (2003) In: Craig PJ (ed) Organometallic compounds in the environment, 2nd edn. Wiley, Chichester, pp 101–150
- Clayton GD, Clayton FE (1994) Patty's industrial hygiene toxicology, 4th edn. A Wiley-Interscience Publication, New York, pp 2157–2173
- Clearwater SJ, Fara AM, Meyer JS (2002) Review bioavailability and toxicity of dietborne copper and zinc to fish. Comp Biochem Physiol Part C 132:269–313

- Coccioni R, Frontalini F, Marsili A, Mana D (2009) Benthic foraminifera and trace element distribution: a case-study from the heavily polluted lagoon of Venice (Italy). Mar Pollut Bull 59:257–267
- Collavini F, Bettiol C, Zaggia L, Zonta R (2005) Pollutant loads from the drainage basin to the Venice Lagoon (Italy). Environ Int 31:939–947
- Compeau G, Bartha R (1984) Methylation and demethylation of mercury under controlled redox, pH, and salinity conditions. Appl Environ Microbiol 50:498–502
- Connelly DP, Statham PJ, Knap AH (2006) Seasonal changes in speciation of dissolved chromium in the surface Sargasso Sea. Deep-Sea Res I 53:1975–1988
- Coogan TP, Latta DM, Snow ET, Costa M (1989) Toxicity and carcinogenicity of nickel compounds. Crit Rev Toxicol 19(4):341–384
- Cossa D, Coquery M (2005) The Mediterranean mercury anomaly, a geochemical or a biological issue. Handbook environmental chemistry, vol5. Springer-Verlag, Heidelberg
- Cossa D, Coquery M, Gobeil C, Martin JM et al (1996) Mercury fluxes at the ocean margins. In: Baeyens W (ed) Global and regional mercury cycles: sources, fluxes, and mass balances., pp 229–247
- Costa M, Liss P (2000) Photoreduction and evaluation of mercury from sea water. Sci Total Environ 261:125–135
- Costa MF, Landing WM, Kehrig HA, Barletta M, Holmes CD, Barrocas PRG, Evers DC, Buck DG, Vasconcellos AC, Hacon SS, Moreira JC, Malm O (2012) Mercury in tropical and subtropical coastal environments. Environ Res 119:88–100
- Craig PJ (1986) Organomercury compounds in the environment. In: Craig PJ (ed) Organometallic compounds in the environment. Longman, Harlow, pp 55–110
- Craig PJ, Moreton PA (1983) Total mercury, methyl mercury and sulphide in River Carton sediments. Mar Pollut Bull 14:408–411
- Crecelius EA, Bothner MH, Carpenter R (1975) Geochemistries of arsenic, antimony, mercury and related elements in sediments of Puget sound. Environ Sci Technol 9(4):325–333
- Crichton R (2001) Inorganic biochemistry of iron metabolism: from molecular mechanisms to clinical consequences, 2nd edn. Wiley, Chichester
- Cross FA, Sunda, WG (1985) The relationship between chemical speciation and bioavailability of trace metals to marine organisms–a review. In: Chao NL, Kirby-Smith W (eds) Proceedings of the international symposium on utilization of coastal ecosystems, vol 1. Fundaqao Universidade do Rio Grande and Duke University Marine Laboratory, pp 169–182
- Cullen WR, Reimer KJ (1989) Arsenic speciation in the environment. Chem Rev 89:713-764
- Da Ros L, Nasci C, Marigomez I, Soto M (2000) Biomarkers and trace metals in the digestive gland of indigenous and transplanted mussels, *Mytilus galloprovincialis*, in Venice Lagoon, Italy. Mar Environ Res 50(1):417–423
- Das KK, Dastidar SG, Chakrabarty S, Banerjee SK (1980) Toxicity of mercury: a comparative study in air breathing and non air breathing fish. Hydrobiology 68(3):225–229
- Das A, Justic⁷ D, Swenson E (2010) Modeling estuarine-shelf exchanges in a deltaic estuary: implications for coastal carbon budgets and hypoxia. Ecol Mode 221(7):978–985
- de Baar HJW, de Jong JTM (2001) Distributions, sources and sinks of iron in seawater. In: Hunter KA, Turner DR (eds) The biogeochemistry of iron in seawater, IUPAC book series on analytical and physical chemistry of environmental systems. Wiley, Chichester, pp 123–254
- De Schamphelaere KAC, Koene JM, Heijerick DG, Janssen CR (2008) Reduction of growth and haemolymph Ca levels in the freshwater snail Lymnaea stagnalis chronically exposed to cobalt. Ecotoxicol Environ Saf 71(1):65–70
- Denkhaus E, Salnikow K (2002) Nickel essentiality, toxicity, and carcinogenicity. Crit Rev Oncol Hematol 42(1):35–56
- Di Leonardo R, Vizzini S, Bellanca A, Mazzola A (2009) Sedimentary record of anthropogenic contaminants (trace metals and PAHs) and organic matter in a Mediterranean coastal area (Gulf of Palermo, Italy). J Mar Syst 78:136–145

- Comune di Venezia (2012) Yearbook of tourism in Venice. Available at: http://www.comune. venezia.it/flex/cm/pages/ServeBLOB.php/L/IT/IDPagina/53175
- Dimitriou P, Castritsi-Catharios J, Miliou H (2003) Acute toxicity effects of tributyltin chloride and triphenyltin chloride on gilthead seabream, Sparus aurata L., embryos. Ecotoxicol Environ Saf 54:30–35
- Domínguez-Lledó FC, Galindo-Riaño MD, Díaz-López IC, García-Vargas M, Granado-Castro MD (2007) Applicability of a liquid membrane in enrichment and determination of nickel traces from natural waters. Anal Bioanal Chem 389:653–659
- Donard OFX, Lespes G, Amouroux D, Morabito R (2001) Organotin compounds in the environment: still a critical issue. In: Ebdon L, Pitts L, Cornelis R, Crews H, Donard OFX, Quevauviller P (eds) Trace element speciation for environment. Food & Health. Royal Society of Chemistry, Cambridge, pp 142–175
- Donat JR, Bruland K (1995) Win trace elements in natural waters. In: Salbu B, Steinnes E (eds) CRC Press, Boca Raton, pp 247–281
- Donazzolo R, Orio AA, Pavoni B, Perin G (1984) Heavy-metals in sediments of the Venice Lagoon. Oceanologica Acta 7(1):25–32
- Duchart P, Calvert SE, Price NB (1973) Distribution of trace metals in the porewaters of shallow water marine sediments. Limnol Oceanogr 18:605–610
- Duft M, Schulte-Oehlmann U, Tillmann M, Markert B, Oehlmann J (2003) Toxicity of triphenyltin AND tributyltin to the freshwater mudsnail *Potamopyrgus antipodarum* in a new sediment biotest. Environ Toxicol Chem 22(1):145–152
- Duinker JC, Nolting RF (1982) Dissolved copper, zinc and cadmium in the Southern Bight of the North Sea. Mar Pollut Bull 13:93–96
- Eaton A (1979) The impact of anoxia on Mn fluxes in the Chesapeake Bay. Geochim Cosmochim Acta 43:429–432
- EC (2000) Directive 2000/60/EC of the European Parliament and of the Council of 23 October 2000. Establishing a framework for Community action in the field of water policy. Off J Eur Commun. http://eur-lex.europa.eu/LexUriServ/LexUriServ.do?uri=OJ:L:2000:327:0001: 0072:EN:PDF
- Edmonds JS, Shibata Y, Francesconi KA, Rippingale RJ (1997) Arsenic transformations in short marine food chains studied by HPLC-ICP MS. Appl Organometal Chem 11(4):281–287
- Eisler R (1986) Chromium hazards to fish, wildlife, and invertebrates: a synoptic review. U.S. Fish and Wildlife Service Biological report 85 (1.6). U.S. Department of the Interior, Fish and Wildlife Service, Laurel, Maryland, p 60
- Eisler R (1989) Tin hazards to fish, wildlife, and invertebrates: a synoptic review, Contaminant hazard reviews. Report No. 15. Biological report 85 (1.15). U.S. Department of the Interior, Fish and Wildlife Service, Laurel, Maryland, p 60
- Elbaz-Poulichet F (2005) River inputs of metals and Arsenic. In: Part K, Saliot A (eds) The Mediterranean Sea, vol 5, The handbook of environmental chemistry. Springer Verlag, Berlin/ Heidelberg, pp 211–235
- El-Moselhy KM (2006) Distribution of vanadium in bottom sediments from the marine coastal area of the Egyptian seas. Egypt J Aquat Res 32(1):12–21
- Engel DW, Fowler BA (1979) Factors influencing cadmium accumulation and its toxicity to marine organisms. Environ Health Perspect 28:81–88
- EnvGovCa (2003) Technical report water quality guidelines for cobalt. http://www.env.gov.bc. ca/wat/wq/BCguidelines/cobalt/cobalt_tech.pdf. Accessed June 2013
- Ergul HA, Ay U, Karademir A, Cayir B, Topcuoglu S, Telli B, Terzi M (2010) Heavy metal concentrations in biota, sediments and sea water samples from di iskelesi region. Rapp Commun Int Mer Médit 39:246
- Eriksson SP (2000) Temporal variations of manganese in the haemolymph and tissues of the Norway lobster, *Nephrops norvegicus* (L.). Aquat Toxicol 48(2–3):297–307
- Eriksson SP, Baden SP (1998) Manganese in the haemolymph and tissues of the Norway lobster, *Nephrops norvegicus*, along the Swedish west -coast, 1993–1995. Hydrobiology 375–376: 255–295

- EU (2003) Regulation (EC) No 782/2003 of the European Parliament and of the Council of 14 April 2003 on the prohibition of organotin compounds on ships. Off J Eur Union, L 115/1, 09.05.2003
- EU (2008) Directive 2008/105/EC of the European Parliament and of the Council on environmental quality standards in the field of water policy, amending and subsequently repealing Council Directives 82/176/EEC, 83/513/EEC, 84/156/EEC, 84/491/EEC, 86/280/EEC and amending Directive 2000/60/EC of the European Parliament and of the Council. Off J Eur Union, L 348, 24.12.2008
- Fabbri D, Gabbianelli G, Locatelli C, Lubrano D, Trombini C, Vassura I (2001) Distribution of mercury and other heavy metals in core sediments of the northern Adriatic Sea. Water Air Soil Pollut 129(1–4):143–153
- Facca C, Pellegrino N, Ceoldo S, Tibaldo M, Sfriso A (2011) Trophic conditions in the waters of the Venice lagoon (Northern Adriatic Sea, Italy). Open Oceanogr J 5:1–13
- Favero N, Cattalini F, Bertaggia D, Albergoni V (1996) Metal accumulation in a biological indicator (Ulva rigida) from the Lagoon of Venice (Italy). Arch Environ Contam Toxicol 31 (1):9–18
- Ferrara R, Ceccarini C, Lanzillotta E, Gardfeldt K, Sommar J, Horvat M, Logar M, Fajon V, Kotnik J (2003) Profiles of dissolved gaseous mercury concentration in the Mediterranean seawater. Atmos Environ 37(1):85–92
- Fitzgerald WF, Mason RP (1997) Biogeochemical cycling of mercury in the marine environment. In: Sigel A, Sigel H (eds) Mercury and its effects on environment and biology. Marcel Dekker, New York, pp 53–111
- Fitzgerald WF, Lamborg CH, Hammerschmidt CR (2007) Marine biogeochemical cycling of mercury. Chem Rev 107:641–662
- Flegal AR, Patterson CC (1983) Vertical concentration profiles of lead in the Central Pacific at 15° and 20°S. Earth Planet Sci Lett 64:19–32
- Flyhammer P (1995) Heavy-metals in the environment—analysis of the cadmium flux in Sweden with special emphasis on landfill leachate. J Environ Qual 24:612–621
- Fontini F, Umgiesser G, Vergano L (2010) The role of ambiguity in the evaluation of the net benefits of the MOSE system in the Venice Lagoon. Ecol Econ 69(10):1964–1972
- Förstner U (1984) Chemical forms and reactivities of metals in sediments. In: Leschber R, Davis RD, Hermite PL (eds) Chemical methods for assessing bio-available metals in sludges and soils. Elsevier Applied Science Publishers, New York
- Francesconi KA, Edmonds JS (1996) Arsenic and marine organisms. In: Sykes AG (ed) Advances in inorganic chemistry. Academic Press, San Diego, pp 147–189
- Frausto da Silva JJR, Williams RJP (1991) Manganese: dioxygen evolution and glycosylation. In: The biological chemistry of the elements/the inorganic chemistry of life. Clarendon, Oxford, pp 370–387
- Freedman ML, Cunningham PM, Schindler JE, Zimmeman MJ (1980) Effect of lead speciation on toxicity. Bull Environ Contam Toxicol 25:389–393
- Frignani M, Bellucci LG, Langone L, Muntau H (1997) Metal fluxes to the sediments of the northern Venice Lagoon. Mar Chem 58(3):275–292
- Gheju M (2011) Hexavalent chromium reduction with zero-valent iron (ZVI) in aquatic systems. Water Air Soil Pollut 222:103–148
- Ghezzo M, Guerzoni S, Cucco A, Umgiesser G (2010) Changes in Venice Lagoon dynamics due to construction of mobile barriers. Coast Eng 57:694–708
- Gibbs PE, Bryan GW (1996) Reproductive failure in the gastropod Nucella lapillus associated with imposex caused by tributyltin pollution: a review. In: Champs MA, Seligman PF (eds) Organotin – environmental fate and effects. Chapman & Hall, London, pp 259–281
- Giusti L, Polo MC (2002) A review of the glass-making industry of Murano (Venice, Italy) under an environmental perspective. Environ Manage Health 13:40–54
- Giusti L, Zhang H (2002) Heavy metals and arsenic in sediments, mussels and marine water from Murano (Venice, Italy). Environ Geochem Health 24(1):47–65

- Gledhill M, Van den Berg CMG (1994) Determination of complexation of iron (III) with natural organic complexing ligands in seawater using cathodic stripping voltammetry. Mar Chem 47(1):41–54
- Granger J, Price NM (1999) The importance of siderophores in iron nutrition of heterotrophic marine bacteria. Limnol Oceanogr 44:541–555
- Grieb TM, Bowie GL, Driscoll CT, Gloss SP, Schofield CL, Porcella DB (1990) Factors affecting mercury accumulation in fish in the upper Michigan peninsula. Environ Toxicol Chem 9(7): 919–930
- Griffitt RJ, Luo J, Gao J, Bonzongo JC, Barber DS (2008) Nanomaterials in the environment effects of particle composition and species on toxicity of metallic nanoparticles in aquatic organisms. Environ Toxicol Chem 27(9):1972–1978
- Grousset FE, Quetel CR, Thomas B, Buat-Me´nard P, Donard OF, Bucher A (1994) Transient Pb isotopic signatures in the Western European atmosphere. Environ Sci Technol 28:1605–1608
- Guerzoni S, Tagliapietra D (2006) Atlante della laguna. Venezia tra terra e mare (Atlas of the lagoon: Venice between land and sea). Marsilio Editore, Venice, 242 pp
- Guerzoni S, Rancazzo G, Molinaroli E, Rossini P (2005) Atmospheric bulk deposition to the Lagoon of Venice Part II. Source apportionment analysis near the industrial zone of Porto Marghera, Italy. Environ Int 31:975–982
- Guruge KS, Tanabe S, Iwata H (1996) Distribution, biomagnifications and elimination of butyltin compound residues in common cormorants (*Phalacrocorax carbo*) from Lake Biwa, Japan. Arch Environ Contam Toxicol 31:210–217
- Hall JLW, Anderson RD (1999) A deterministic ecological risk assessment for copper in European saltwater environment. Mar Pollut Bull 38:207–218
- Hall IR, Hydes DJ, Statham PJ, Overnell J (1996) Dissolved and particulate trace metals in a Scottish sea loch: an example of a pristine environment. Mar Pollut Bull 32:846–854
- Hall LW Jr, Scott MC, Killen WD (1997) A screening level probabilistic ecological risk assessment of copper and cadmium in the Chesapeake Bay watershed. Report. Maryland University, Queenstown. Wye Research and Education Center; Environmental Protection Agency, Annapolis, MD. Chesapeake Bay Program
- Hamasaki T, Nagase H, Yoshioka Y, Sato T (1995) Formation, distribution, and ecotoxicity of methymetals of tin, mercury, and arsenic in the environment. Crit Rev Environ Sci Technol 25:45–91
- Han S, Gieskes J, Obraztsova A, Deheyn DD, Tebo BM (2011) Relocation effects of dredged marine sediments on mercury geochemistry: Venice lagoon, Italy. Estuar Coast Shelf Sci 93:7–13
- Hanaoka K, Nakamura O, Ohno H, Tagawa S, Kaise T (1995) Degradation of arsenobetaine to inorganic arsenic by bacteria in seawater. Hydrobiologia 316:75–80
- Hansmann W, Köppel V (2000) Lead-isotopes as tracers of pollutants in soils. Chem Geol 171:123-144
- Hartwell SI, Alden RW, Wright DA, Ailstock S, Kerhin R (2000) Correlation of measures of ambient toxicity and fish community diversity in a Chesapeake Bay tributary, Maryland, USA: a biological, chemical, and geological assessment. Environ Toxicol Chem 19(7):1753–1763
- Hasegawa H (1996) Seasonal changes in methylarsenic distribution in Tosa Bay and Uranouchi Inlet. Appl Organometal Chem 10:733–740
- Hasegawa H, Sohrin Y, Seki K, Sato M, Norisuye K, Naito K, Matsui M (2001) Biosynthesis and release of methylarsenic compounds during the growth of freshwater algae. Chemosphere 43:265–272
- Hellweger FL, Lall U (2004) Modeling the effect of algal dynamics on arsenic speciation in Lake Biwa. Environ Sci Technol 38:6716–6723
- Hines ME, Poitras EN, Covelli S, Faganelli J, Emili A, Zizek S, Horvat M (2012) Mercury methylation and demethylation in Hg-contaminated lagoon sediments (Marano and Grado Lagoon, Italy). Estuar Coast Shelf Sci 113:85–95
- Hoch M (2001) Organotin compounds in the environment—an overview. Appl Geochem 16(7–8): 719–743

- Hong HS, Kester DR (1986) Redox state of iron in the offshore waters of Peru. Limnol Oceanogr 31:512–524
- Horsburgh MJ, Wharton SJ, Karavolos M, Foster SJ (2002) Manganese: elemental defense for a life with oxygen. Trends Microbiol 10(11):496–501
- Horvat M, Kotnik J, Logar M, Fajon V, Zvonarić T, Pirrone N (2003) Speciation of mercury in surface and deep-sea waters in the Mediterranean Sea. Atmos Environ 37(1):S93–S108
- Howard AG, Comber SDW, Kifle D, Antai EE, Purdie DA (1995) Arsenic speciation and seasonal changes in nutrient availability and micro-plankton abundance in Southampton water, UK. Estuar Coast Shelf Sci 40:435–450
- Hu J, Zhen H, Wan Y (2006) Trophic magnification of triphenyltin in a marine food web of Bohai Bay, North China: comparison to tributyltin. Environ Sci Technol 40:3142–3147
- Hunt CD (1983) Incorporation and deposition of Mn and other trace metals by flocculent organic matter in a controlled marine ecosystem. Limnol Oceanogr 28:302–308
- Huynh-Ngoc L, Whitehead NE, Boussemart M, Calmet D (1989) Dissolved nickel and cobalt in the aquatic environment around Monaco. Mar Chem 26:119–132
- IMO (International Maritime Organization) (1997) Marine environmental protection Committee. Harmful effects of the use of antifouling paints for ships. 40th session, Agenda Item 11, Annex 2
- IMO (International Maritime Organization) (2001) International convention on the control of harmful anti-fouling systems on ships. International Maritime Organization. http://www.imo.org
- Irato P, Santovito G, Cassini A, Piccinni E, Albergoni V (2003) Metal accumulation and binding protein induction in Mytilus galloprovincialis, Scapharca inaequivalvis, and Tapes philippinarum from the Lagoon of Venice. Arch Environ Contam Toxicol 44(4):0476–0484
- Ishikawa M, Okoshi K, Kurosawa M, Kitao K (1987) Trace element analysis of seawater by PIXE. In: Sebe T, Yamamoto Y (eds) Proceedings of the 12th international symposium on application of ion beams in material science. Hosei University Press, Tokyo, pp 445–456
- James R, Sampath K, Jothilakshmi S, Vasudhevan I, Thangarathinam R (2008) Effects of copper toxicity on growth, reproduction and metal accumulation in chosen ornamental fishes. Ecohydrol Hydrobiol 8(1):89–97
- Jiang GB, Shi JB, Feng XB (2006) Mercury pollution in China. Environ Sci Technol 40: 3672–3678
- Jones B, Bolam T (2007) Copper speciation survey from UK marinas, harbours and estuaries. Mar Pollut Bull 54:1127–1138
- Jorge MB, Loro VL, Bianchini A, Wood CM, Gillis PL (2013) Mortality, bioaccumulation and physiological responses in juvenile freshwater mussels (Lampsilis siliquoidea) chronically exposed to copper. Aquat Toxicol 126:137–147
- Kalogeropoulos N, Scoullos M, Vassilaki-Grimant M, Grimanis P (1989) Vanadium in particles and sediments of the northern Saronikos gulf, Greece. Sci Total Environ 79:241–252
- Karadjova IB, Slaveykova VI, Tsalev DL (2008) The biouptake and toxicity of arsenic species on the green microalga Chlorella salina in seawater. Aquat Toxicol 87:264–271
- Kennish MJ, Fertig B (2012) Application and assessment of a nutrient pollution indicator using eelgrass (Zostera marina L.) in Barnegat Bay-Little Egg Harbor estuary, New Jersey. Aquat Bot 96:23–30
- Kepkay PE, Nealson KH (1987) Growth of a manganese oxidizing *Pseudomonas sp.* in continuous culture. Arch Microbiol 148:63–67
- Kernen N, Kidd MJ, Penner-Hahn JE, Pakrasi HB (2002) A light-dependent mechanism for massive accumulation of manganese in the photosynthetic bacterium Synechocytis sp. PCC 6803. Biochemistry 41(50):15085–15092
- Khang HV, Hatayama M, Inoue C (2012) Arsenic accumulation by aquatic macrophyte coontail (Ceratophyllum demersum L.) exposed to arsenite, and the effect of iron on the uptake of arsenite and arsenate. Environ Exp Bot 83:47–52
- King JK, Saunders M, Lee RF, Jahnke RA (1999) Coupling mercury methylation rates to sulfate reduction rates in marine sediments. Environ Toxicol Chem 18:1362–1369
- Knauer GA, Martin JH, Gordon RM (1982) Cobalt in north-east Pacific waters. Nature 297:49-51

- Kraepiel AML, Chiffoleau JF, Martin JM, Morel FMM (1997) Geochemistry of trace metals in the Gironde estuary. Geochim Cosmochim Acta 61:1421–1436
- Kremling K (1983) The behaviour of Zn, Cd, Cu, Ni, Co, Fe, and Mn in anoxic Baltic waters. Mar Chem 13:87–108
- Langston WJ (1982) The distribution of mercury in British estuarine sediments and its availability to deposit feeding bivalves. J Mar Biol Ass UK 62(667):84
- Langston WJ (1983) The behaviour of arsenic in selected United Kingdom estuaries. Can J Fish Aquat Sci 40(Suppl 2):143–150
- Langston WJ, Burt GR, Pope N, Mcevoy J (1996) Bioaccumulation of methylmercury (Mersey estuary 1995). Report to NRA North West Region
- Laslett RE (1995) Concentrations of dissolved and suspended particulate Cd, Cu, Mn, Ni, Pb and Zn in surface waters around the coasts of England and Wales and in adjacent seas. Estuar Coast Shelf Sci 40:67–85
- Laslett RE, Balls PW (1995) The behaviour of Mn, Ni and Zn in the Forth, an industrialised partially mixed estuary. Mar Chem 48:311–328
- Laughlin RB (1996) Bioaccumulation of TBT by aquatic organisms. In: Champ M, Seligman PF (eds) Organotin: environmental fate and effects. Chapman & Hall, London, pp 331–355
- Lee K (1982) The effects of vanadium on phytoplankton: field and laboratory studies. Ph.D. thesis, University of Toronto, 431 p. National Library Canada, Canadian Theses Microfilm No. 58302
- Leermakers M, Galletti S, Galan SD, Brion N, Baeyens W (2001) Mercury in the Southern North Sea and Scheldt estuary. Mar Chem 75(3):229–248
- Legorburu I, Rodriguez JG, Borja A, Menchaca I, Solaun O, Valencia V, Galparsoro I, Larreta J (2013) Source characterization and spatio-temporal evolution of the metal pollution in the sediments of the Basque estuaries (Bay of Biscay). Mar Pollut Bull 66:25–38
- Lemaire E, Blanc G, Schafer J, Coynel A, Etcheber H (2006) Dissolved trace metal-organic complexes in the Lot–Garonne river system determined using the C18 Sep–Pak system. Aquat Geochem 12:21–38
- Levy LS, Venitt S (1986) Carcinogenicity and mutagenicity of chromium compounds: the association between bronchial metaplasia and neoplasia. Carcinogenesis 7(831):835
- Li YH, Burkhardt H, Teraoka H (1984) Desorption and coagulation of trace elements during estuarine mixing. Geochim Cosmochim Acta 48:1879–1884
- Liang GD (2011) Redox metal processes and controls in estuaries. Elsevier 115:141
- Lobinski R, Marczenko Z (1996) In: Weber SG (ed) Spectrochemical trace analysis for metals and metalloids, Wilson and Wilson's comprehensive analytical chemistry. Elsevier, Dordrecht, p 481
- Lotze HK (2010) Historical reconstruction of human-induced changes in US estuaries. Oceanogr Mar Biol Ann Rev 48:267–338
- Luo W, Wang T, Jiao W, Hu W, Naile JE, Khim JS, Giesy JP, Lu Y (2012) Mercury in coastal watersheds along the Chinese Northern Bohai and Yellow Seas. J Hazard Mater 215(216):199–207
- Luoma SN, Bryan GW (1982) A statistical study of environmental factors controlling concentrations of heavy metals in the burrowing bivalve *Scrobicularia plana* and the polychaete *Nereis diversicolor*. Estuar Coast Shelf Sci 15:95–108
- Mackey EA, Becker PR, Demiralp R, Greenberg RR, Koster BJ, Wise SA (1996) Bioaccumulation of vanadium and other trace metals in livers of Alaskan cetaceans and pinnipeds. Arch Environ Contam Toxicol 30(4):503–512
- Maher WA (1984) Mode of occurrence and speciation of arsenic in some pelagic and estuarine sediments. Chem Geol 47(3-4):333-345
- Mance G, Brown VM, Gardiner J, Yates J (1984) Proposed environmental quality standards for list II substances in water – chromium, Technical report TR 207. WRc, Medmenham
- Martin JM, Windom HL (1991) Present and future roles of ocean margins in regulating marine biogeochemical cycles of trace elements. In: Mantoura RFC, Martin J-M, Wollast R (eds)

Ocean margin processes in global change. Dahlem workshop. Wiley, Chichester, pp 45–67; 469 pp

- Martin JM, Dai MH, Cauwet G (1995) Significance of colloids in the biogeochemical cycling of organic carbon and trace metals in the Venice Lagoon (Italy). Limnol Oceanogr 40(1):119–131
- Martin K, Huggins T, King C, Carrol MA, Catapane EJ (2008) The neurotoxic effects of manganese on the dopaminergic innervation of the gill of the bivalve mollusc, Crassostrea virginica. Comp Biochem Physiol C 148:152–159
- Martino M, Turner A, Nimmo M, Millward GE (2002) Resuspension, reactivity and recycling of trace metals in the Mersey Estuary, UK. Mar Chem 77:171–186
- Marvin-DiPasquale MC, Agee J, McGowan C, Oremland RS, Thomas M, Krabbenhoft D, Gilmour CC (2000) Methyl-mercury degradation pathways: a comparison among three mercury-impacted ecosystems. Environ Sci Technol 34:4908–4917
- Mason RP, Benoit JM (2003) Organomercury compounds in the environment. In: Craig P (ed) Organometallics in the environment. Wiley, New York, pp 57–99
- Mason RP, Fitzgerald WF (1993) The distribution and biogeochemical cycling of mercury in the equatorial Pacific Ocean. Deep Sea Res 40:1897–1907
- Mason RP, Fitzgerald WF, Hurley J, Hanson AK, Donaghay PL, Sieburth JM (1993) Mercury biogeochemical cycling in a stratified estuary. Limnol Oceanogr 38(6):1227–1241
- Mason RP, Fitzgerald WF, Morel FMM (1994) The biogeochemical cycling of elemental mercury, anthropogenic influences. Geochim Cosmochim Acta 58:3191–3198
- Mason RP, Rolfhus KR, Fitzgerald WF (1995) Methylated and elemental mercury cycling in the surface and deep ocean waters of the North Atlantic. Water Air Soil Pollut 80:665–677
- Meisch HU, Benzschawel H (1978) The role of vanadium in green plants. III. Influence on cell division of Chlorella. Arch Microbiol 116:91–95
- Meisch HU, Bielig HJ (1975) Effect of Vanadium on growth, chlorophyll formation and iron metabolism in unicellular green algae. Arch Microbiol 105:77–82
- Meisch HU, Benzschawel H, Bielig HJ (1977) The role of vanadium in green plants. II. Vanadium in green algae—two sites of action. Arch Microbiol 114:67–70
- Michel P, Averty B (1999) Distribution and fate of tributyltin in surface and deep waters of the northwestern Mediterranean. Environ Sci Technol 33:2524–2528
- Migon C, Alleman L, Leblond N, Nicolas E (1993) Evolution of the atmospheric lead in northwestern Mediterranean between 1986 and 1992. Atmos Environ 27A:2161–2167
- Miller WL, King DW, Lin J, Kester DR (1995) Photochemical redox cycling of iron in coastal seawater. Mar Chem 50:63–77
- Millward GE (1995) Processes affecting trace element speciation in estuaries, a review. Analyst 120:609–614. Mining waste and its effects on estuarine organisms. Mar Biol Ass UK, Occasional Publication, No. 2, 112 pp
- Molinaroli E, Guerzoni S, Sarretta A, Cucco A, Umgiesser G (2007) Link between hydrology and sedimentology in the Lagoon of Venice, Italy. J Mar Syst 68:303–317
- Monteiro SM, dos Santos NMS, Calejo M, Fontainhas-Fernandes A, Sousa M (2009) Copper toxicity in gills of the teleost fish, *Oreochromis niloticus*: effects in apoptosis induction and cell proliferation. Aquat Toxicol 94(3):219–228
- Moore WS (2006) The role of submarine groundwater discharge in coastal biogeochemistry. J Geochem Explor 88:389–393
- Moore MV, Winner RW (1989) Relative sensitivity of *Cerodaphnia dubia* laboratory tests and pond communities of zooplankton and benthos to chronic copper stress. Aquat Toxicol 15:311–330
- Moore RM, Webb R, Tokarczyk R, Wever R (1996) Bromoperoxidase and iodoperoxidase enzymes and production of halogenated methanes in marine diatom cultures. J Geophys Res 101:20899–20908
- Morel FMM, Reinfelder JR, Roberts SB, Chamberlain CP, Lee JG, Yee D (1994) Zinc and carbon co-limitation of marine phytoplankton. Nature 369:740–742

- Morel FMM, Kraepeil AML, Amyot M (1998) The chemical cycle and bioaccumulation of mercury. Annu Rev Ecol Syst 29:543–566
- Morris AW (1984) The chemistry of the Severn Estuary and Bristol Channel. Mar Pollut Bull 15:57-61
- Morris AW, Bale AJ, Howland RJM, Millward GE, Ackroyd DR, Loring DH, Rantala RTT (1986) Sediment mobility and its contribution to trace-metal cycling and retention in a macrotidal estuary. Water Sci Technol 18(4–5):111–119
- Morris AW, Allen JI, Howland RJM, Wood RG (1995) The estuary plume zone: source or sink for land-derived nutrient discharges? Estuar Coast Shelf Sci 40:387–402
- Murray JW, Spell B, Paul B (1983) The contrasting geochemistry of manganese and chromium in the eastern tropical Pacific Ocean. In: Wong CS, Boyle E, Bruland KW, Burton JD, Goldberg ED (eds) Trace metals in sea water. Plenum Press, New York/London, pp 643–669
- Nalewajko G, Lee K, Jack TR (1995) Effects of vanadium on freshwater phytoplankton photosynthesis. Water Air Soil Pollut 81:93–105
- Neff JM (1997) Ecotoxicology of arsenic in the marine environment. Environ Toxicol Chem 16:917–927
- Nesto N, Romano S, Moschino V, Mauri M, Da Ros L (2007) Bioaccumulation and biomarker responses of trace metals and microorganic pollutants in mussels and fish from the Lagoon of Venice, Italy. Mar Pollut Bull 55(10–12):469–484
- Nicolau R, Lucas Y, Merdy P, Raynaud M (2012) Base flow and storm water net fluxes of carbon and trace metals to the Mediterranean Sea by an urbanized small river. Water Res 46:6625–6637
- Nolting RF, Gerringa LA, Swagerman MW, Timmermans KR, de Baar HJW (1998) Fe (III) speciation in the high nutrient, low chlorophyll Pacific region of the Southern Ocean. Mar Chem 62:335–352
- Nolting RF, Helder W, de Baar HJW, Gerringa LJA (1999) Contrasting behaviour of trace metals in the Scheldt estuary in 1978 compared to recent years. J Sea Res 42:275–290
- Nor YM (1987) Ecotoxicity of copper to aquatic biota a review. Environ Res 43:274-282
- Norwood WP, Borgmann U, Dixon DG (2006) Saturation models of arsenic, cobalt, chromium and manganese bioaccumulation by *Hyalella azteca*. Environ Pollut 143:519–528
- Norwood WP, Borgmann U, Dixon DG (2007) Chronic toxicity of arsenic, cobalt, chromium and manganese to Hyalella azteca in relation to exposure and bioaccumulation. Environ Pollut 147 (1):262–272
- Nurnberg HW, Valenta P (1983) Potentialities and applications of voltammetry in chemical speciation of trace metals in the sea. In: Wong CS, Boyle E, Bruland KW, Burton JD, Goldberg ED (eds) Trace metals in sea water. Springer, New York
- O'Brien P, Rainbow PS, Nugegoda D (1990) The effect of the chelating agent EDTA on the rate of uptake of zinc by *Palaemon elegans* (Crustacea: Decapoda). Mar Environ Res 30:155–159
- Odate S, Pawlik JR (2007) The role of Vanadium in the Chemical Defense of the Solitary Tunicate, Phallusia nigra. J Chem Ecol 33(3):643–654
- Owens RE, Balls PW (1997) Dissolved trace metals in the Tay Estuary: implications for monitoring and modelling. Estuar Coast Shelf Sci 44:421–434
- Oweson C, Hernroth B (2009) A comparative study on the influence of manganese on the bactericidal response of marine invertebrates. Fish Shellfish Immunol 27:500–507
- Paalman PAA, van der Weijden CH, Loch JPG (1994) Sorption of cadmium on suspended matter under estuarine conditions: competition and complexation with major sea-water ions. Water Air Soil Pollut 73:49–60
- Pacyna JM (1987) Atmospheric emissions of arsenic, cadmium, lead and mercury from high temperature processes in power generation and industry. In: Hutchinson TC, Meema KM (eds) Lead, mercury, cadmium and arsenic in the environment. Wiley, Lillestrom
- Patrick R (1978) Effects of trace metals in the aquatic ecosystem. Am Sci 66:185-191
- Paulsen SC, List EJ (1997) A study of transport and mixing in natural waters using ICPMS: water-particle interactions. Water Air Soil Pollut 99:149–156

- Pavoni B, Marcomini A, Sfriso A, Orio AA (1988) Multivariate analysis of heavy metal concentrations in sediments of the lagoon of Venice. Sci Total Environ 77(2):189–202
- Pavoni B, Centanni E, Valcanover S, Fasolato M, Ceccato S, Tagliapietra D (2007) Imposex levels and concentrations of organotin compounds (TBT and its metabolites) in Nassarius nitidus from the Lagoon of Venice. Mar Pollut Bull 55:505–511
- Perez-Benito JF (2006) Effects of Cr(VI) and V(V) on the lifespan of fish. J Trace Elem Med Biol 20(3):161–170
- Perin G, Fabris R, Manente S, Wagener AR, Hamacher C, Scotto S (1997) A five-year study on the heavy metal pollution of Guanabara bay sediments (Rio De Janeiro, Brazil) and evaluation of the metal bioavailability by means of geochemical speciation. Water Res 31:3017–3028
- Peternac B, Legovic T (1986) Uptake, distribution and loss of Cr in the crab *Xantho hydrophilus*. Mar Biol 91:467–471
- Pettersson L, Andersson I, Gorzsas A (2003) Speciation in peroxovanadate systems. Coord Chem Rev 237:77–87
- Phillips DJH (1990) Arsenic in aquatic organisms: a review, emphasizing chemical speciation. Aquat Toxicol 16:151–186
- Pinto R, Patricio J, Neto JM, Salas F, Marques JC (2010) Assessing estuarine quality under the ecosystem services scope: ecological and socioeconomic aspects. Ecol Complex 7(3):389–402
- Pinto JJ, Garcia-Vargas M, Moreno C (2013) A bulk liquid membrane-flow injection (BLM-F1) coupled system for the preconcentration and determination of vanadium in saline waters. Talanta 103:161–165
- Porter SK, Scheckel KG, Impellitteri CA, Ryan JA (2004) Toxic metals in the environment: thermodynamic considerations for possible immobilization strategies for Pb, Cd, As, and Hg. Crit. Rev. Environ Sci Technol 34:495–604
- Post JE (1999) Manganese oxide minerals: crystal structures and economic and environmental significance. Proc Natl Acad Sci U S A 96:3447–3454
- Prosi F (1989) Factors controlling biological availability and toxic effects of lead in aquatic organisms. Sci Total Environ 79:157–169
- Rabitto IS, Alves Costa JRM, Silva de Assis HC, Pelletier E, Akaishi FM, Anjos A, Randi MAF, Oliveira Ribeiro CA (2005) Effects of dietary Pb (II) and tributyltin on neotropical fish, *Hoplias malabaricus*: histopathological and biochemical findings. Ecotoxicol Environ Saf 60 (2):147–156
- Rahman MA, Hasegawa H, Lim RP (2012) Bioaccumulation, biotransformation and trophic transfer of arsenic in the aquatic food chain. Environ Res 116:118–135
- Rai D, Zachara JM, Eary LE, Girvin DC, Moore DA, Resch CT, Sass BM, Schmidt RL (1986) Geochemical behavior of chromium species. Interim report EPRI EA-4544, E.P.R.I., Pale Alto, CA
- Rai D, Sass BM, Moore DA (1987) Chromium(III) hydrolysis constants and solubility of chromium(III) hydroxide. Inorg Chem 26:345–349
- Ramessur RT (2004) Statistical comparison and correlation of zinc and lead in estuarine sediments along the western coast of Mauritius. Environ Int 30:1039–1044
- Rampazzo G, Masiol M, Visin F, Rampado E, Pavoni B (2008) Geochemical characterization of PM10 emitted by glass factories in Murano, Venice (Italy). Chemosphere 71:2068–2075
- Rehder D (2000) Vanadium nitrogenase. J Inorg Biochem 80:133-136
- Rejitha V, Subhash Peter MC (2013) Adrenaline and triiodothyronine modify the iron handling in the freshwater air-breathing fish Anabas testudineus Bloch: role of ferric reductase in iron acquisition. Gen Comp Endocrinol 181:130–138
- Renzoni A, Zino F, Franchi E (1998) Mercury levels along the food chain and risk for exposed populations. Environ Res 77(2):68–72
- Richard FC, Bourg CM (1991) Aqueous geochemistry of chromium: a review. Water Res 25(7): 807–816
- Ridgway J, Shimmield G (2002) Estuaries as repositories of historical contamination and their impact on shelf seas. Estuar Coast Shelf Sci 6:903–928

- Riley JD, Towner JV (1984) The distribution of alkyl lead species in the Mersey estuary. Mar Pollut Bull 15:153–158
- Ritson PI, Bouse RM, Flegal AR, Luoma SN (1999) Stable lead isotopic analyses of historic and contemporary lead contamination of San Francisco Bay estuary. Mar Chem 64:71–83
- Robertson DE, Silker WB, Langford JC, Petersen MR, Perkins RW (1973) Transport and depletion of radionuclides in the Columbia river. In: Radioactive contamination of the marine environment. IAEA, Vienna, pp 141–158
- Rocha L, Rodrigues SM, Lopes I, Soares AMVM, Duarte AC, Pereira E (2011) The water-soluble fraction of potentially toxic elements in contaminated soils: relationships between ecotoxicity, solubility and geochemical reactivity. Chemosphere 84:1495–1505
- Roitz JS, Flega AR, Bruland KW (2002) The biogeochemical cycling of manganese in San Francisco Bay: temporal and spatial variations in surface water concentrations. Estuar Coast Shelf Sci 54:227–239
- Rossini P, Matteucci G, Guerzoni S (2010) Atmospheric fall-out of metals around the Murano glass-making district (Venice, Italy). Environ Sci Pollut Res 17:40–48
- Roux LL, Roux SL, Appriou P (1998) Behaviour and speciation of metallic species Cu, Cd, Mn and Fe during estuarine mixing. Mar Pollut Bull 36:56–64
- Rue EL, Bruland KW (1995) Complexation of iron(II1) by natural organic ligands in the Central North Pacific as determined by a new competitive ligand equilibration/adsorptive cathodic stripping voltammetric method. Mar Chem 50:117–138
- Sarretta A, Pillon S, Molinaroli E (2010) Sediment budget in the Lagoon of Venice, Italy. Cont Shelf Res 30(8):934–949
- Savory J, Wills MR (1984) Arsenic. In: Merian E (ed) Metalle in der Ummell. Verlag Chemie, Weinheim-Deerfirid Beach/Florida Basel, pp 315–334
- Scott GR, Sloman KA (2004) The effects of environmental pollutants on complex fish behaviour: integrating behavioural and physiological indicators of toxicity. Aquat Toxicol 68(4):369–392
- Sekizawa J, Suter G II, Birnbaum L (2001) C. Tributyltin and Triphenyltin compounds. http:// whqlibdoc.who.int/hq/2001/a76785_tributyltin.pdf. Accessed June 2013
- Seyler P, Martin JM (1991) Arsenic and selenium in a pristine river-estuarine system: the Krka (Yugoslavia). Mar Chem 34:137–151
- Sfriso A, Marcomini A, Zanette M (1995) Heavy metals in sediments, SPM and phytozoobenthos of the Lagoon of Venice. Mar Pollut Bull 30(2):116
- Sfriso A, Argese E, Bettiol C, Facca C (2008) *Tapes philippinarum* seed exposure to metals in polluted areas of the Venice lagoon. Estuar Coast Shelf Sci 79(4):581–590.124
- Shaked Y, Xu Y, Leblanc K, Morel FMM (2006) Zinc availability and alkaline phosphate activity in Emiliania huxleyi: implications for Zn-P co-limitation in the ocean. Limnol Oceanogr 51(1): 299–309
- Sharma VK, Sohn M (2009) Aquatic arsenic: toxicity, speciation, transformations, and remediation. Environ Int 35:743–759
- Sholkovitz ER (1978) Flocculation of dissolved Fe, Mn, Al, Cu, Ni, Co and Cd during estuarine mixing. Earth Planet Sci Lett 41(1):77–86
- Singh AK, Singh M (2006) Lead decline in the Indian environment resulting from the petrol-lead phase-out programme. Sci Total Environ 368(2–3):686–694
- Smedley PL, Kinniburgh DG (2002) A review of the source, behaviour and distribution of arsenic in natural waters. Appl Geochem 17:517–568
- Snape L, Townsend AT (2008) Multiple Pb sources in marine sediments near the Australian Antarctic Station, Casey. Sci Total Environ 389:466–474
- Solidoro C, Bandelj V, Bernardi FA, Camatti E, Ciavatta S, Cossarini G, Facca C, Franzoi P, Libralato S, Canu DM, Pastres R, Pranovi F, Raicevich S, Socal G, Sfriso A, Sigovini M, Tagliapietra D, Torricelli P (2010) Response of Venice Lagoon ecosystem to natural and anthropogenic pressures over the last 50 years, In: Kennish M, Paerl H (eds) Coastal Lagoons: critical habitats and environmental change. CRC Press/Taylor & Francis, Boca Raton, pp 483–511. http://dx.doi.org/10.1201/EBK1420088304-c19. doi:10.1201/EBK1420088304-c19#_new

2 Harmful Elements in Estuarine and Coastal Systems

- Sommerfreund JK, Grandhi N et al (2010) Contaminant fate and transport in the Venice Lagoon: result from a multi-segment multimedia model. Ecotoxicol Environ Saf 73(3):222–230
- Sprague JB (1971) Measurement of pollutant toxicity to fish III. Water Res 5:245-266
- Stauber JL, Florence TM (1985) Interactions of copper and manganese: a mechanism by which manganese alleviates copper toxicity to the marine diatom, *Nitzschia closterium* (Ehrenberg) W. Smith. Aquat Toxicol 7:241–254
- Steenkamp VE, du Preez HH, Schonbee HJ, van Eeden PH (1994) Bioaccumulation of manganese in selected tissues of the freshwater crab, *Potamonautes warreni* (Calman), from industrial and mine-polluted freshwater ecosystems. Hydrobiologia 288:137–150
- Steinnes E (1989) Cadmium in the terrestrial environment- impact of long-range atmospheric transport. Toxicol Environ Chem 19:139–145
- Struck BD, Pelzer R, Ostapczuk P, Emons H, Mohl C (1997) Statistical evaluation of ecosystem properties influencing the uptake of As, Cd, Co, Cu, Hg, Mn, Ni, Pb and Zn in seaweed (*Fucus vesiculosus*) and common mussel (*Mytilus edulis*). Sci Total Environ 207:29–42
- Stumm W (1992) Chemistry of the solid water interface. Wiley, New York, 428 pp
- Suchanek TH, Eagles-Smith CA, Slotton DG, Harner EJ, Adam DP, Colwell AE, Anderson NL, Wooderward DL (2008) Mine – derived mercury: effects on lower trophic species in Clear Lake, California. Ecol Appl 18(Supplement 8):A158–A176
- Sunda WG (2001) Chapter 3: Bioavailability and bioaccumulation of iron in the sea. In: Turner DR, Hunter KA (eds) The biogeochemistry of iron in seawater. Wiley, New York, pp 41–84
- Sunda WG, Huntsman SA (1983) Effect of competitive interactions between manganese and copper on cellular growth in estuarine and oceanic species of the diatom *Thalassiosira*. Limnol Oceanogr 28:924–934
- Sunda WG, Huntsman SA (1990) Diel cycles in microbial manganese oxidation and manganese redox speciation in coastal waters of the Bahama Islands. Limnol Oceanogr 35:325–338
- Sunda WG, Huntsman SA (1996) Antagonisms between cadmium and zinc toxicity and manganese limitation in a coastal diatom. Limnol Oceanogr 41:373–387
- Sundby B, Silverberg N (1985) Manganese fluxes in the benthic boundary layer. Limnol Oceanogr 30:374–382
- Swarzenski PW, Orem WH, McPherson BF, Baskaran M, Wan Y (2006) Biogeochemical transformation in the Loxaharchee River estuary, Florida: the role of submarine groundwater discharge. Mar Chem 201:248–265
- Taylor MJC, van Staden JF (1994) Spectrophotometric determination of vanadium(IV) and vanadium(V) in each other's presence. Rev Anal 119:1263–1276
- Taylor SW, Kammerer B, Bayer E (1997) New perspectives in the chemistry and biochemistry of the tunichromes and related compounds. Chem Rev 97:333–346
- Tessier A, Campbell PGC (1987) Partitioning of trace metals in sediments: relationships with bioavailability. Hydrobiology 149:43–52
- Thompson B, Adelsbach T, Brown C, Hunt J, Kuwabara J, Neale J, Ohlendorf H, Schwarzbach S, Spies R, Taberski K (2007) Biological effects of anthropogenic contaminants in the San Francisco Estuary. Environ Res 105:156–174
- Thomson D, Maher W, Foster S (2007) Arsenic and selected elements in inter-tidal and estuarine marine algae, south-east coast, NSW, Australia. Appl Organometal Chem 21:396–411
- Thouvenin B, Gonzalez JL, Boutier B (1997) Modelling of pollutant behaviour in estuaries: application to cadmium in the Loire estuary. Mar Chem 58:147–161
- Tipping E, Lofts S, Lawlor AJ (1998) Modelling the chemical speciation of trace metals in the surface waters of the Humber system. Sci Total Environ 210(211):63–77
- Tsalev DL, Zaprianov ZK (1985) Atomic absorption spectrometry in occupational and environmental health practice, vol 1. CRC Press, Boca Raton, p 137
- Tueros I, Rodriguez JG, Borja A, Solaun O, Valencia V, Millan E (2008) Dissolved metal background levels in marine waters, for the assessment of the physico-chemical status, within the European Water Framework Directive. Sci Total Environ 407:40–52

- Tueros I, Borja A, Larreta J, Rodriguez JG, Valencia V, Millan E (2009) Integrating long-term water and sediment pollution data, in assessing chemical status within the European Water Framework Directive. Mar Pollut Bull 58(9):1389–1400
- Tyler PA (1970) Hyphomicrobium and the oxidation of manganese in aquatic ecosystems. Antonie Van Leeuwenhoek 36:567–578
- UNESCO (1987) Convention concerning the protection of the world cultural and natural heritage. Report of the World Heritage Committee, Eleventh session. UNESCO Headquarters, 7–11 Dec 1987
- USEPA (U.S. Environmental Protection Agency) (1980) Ambient water quality criteria for chromium. EPA 440/5-80-035
- USEPA (U.S. Environmental Protection Agency) (1984) Ambient water quality criteria for mercury. EPA 440/5-84-026
- USEPA (U.S. Environmental Protection Agency) (1985) Water quality criteria for copper. EPA 440/5-84-031
- USEPA (U.S. Environmental Protection Agency) (2001) 2001 Update of ambient water quality criteria for cadmium. EPA-822-R-01-001
- USEPA (U.S. Environmental Protection Agency) (2003) Ambient aquatic life water quality criteria for tributyltin. USEPA Office of Research and Development, Environmental. Office of Water 4304 T. US EPA 822-R-03-031
- Vallee BL, Auld DS (1993) Zinc: biological functions and coordination motifs. Acc Chem Res 26 (10):543–551
- Volpi Ghirardini A, Arizzi Novelli A, Losso C, Ghetti PF (2005) Sperm cell and embryo toxicity tests using the sea urchin Paracentrotus lividus (LmK). Techniques in aquatic toxicology, vol 2. CRC Press, Boca Raton, pp 147–168
- Wang S, Xu L, Zhao Z, Wang S, Jia Y, Wang H, Wang X (2012) Arsenic retention and remobilization in muddy sediments with high iron and sulfur contents from a heavily contaminated estuary in China. Chem Geol 314–317:57–65
- Water and River Commission (1997) River and estuarine pollution. Government of Western Australia. Water facts
- Weber HJ (1993) Review of possible paths for abiotic methylation of Mercury (II) in the aquatic environment. Chemosphere 26(11):2063–2077
- Weber FA, Voegelin A, Kretzschmar R (2009) Multi-metal contaminant dynamics in temporarily flooded soil under sulfate limitation. Geochim Cosmochim Acta 73:5513–5527
- Whitehouse P, Young W, Fawell J, Sutton A, Wilkinson M (1998) Proposed environmental quality standards for octylphenol in water (Report for the Environment Agency R&D technical report P59)
- WHO (1988) Environmental health criteria No. 81, Vanadium. IPCS, World Health Organisation, Geneva
- WHO (1989) Environmental health criteria No. 86 Mercury environmental aspects. World Health Organisation, Geneva
- WHO (World Health Organisation) (1991) Environmental Health Criteria 124, Lindane. Published under the joint sponsorship of the United Nations Environment Programme. The International Labour Organisation and the World Health Organisation
- Widdows J, Nasci C, Fossato VU (1997) Effects of pollution on the scope for growth of mussels (Mytilus galloprovincialis) from Venice Iagoon, Italy. Mar Environ Res 43:69–79
- Wilhelm C, Wild A (1984) The variability of the photosynthetic unit in Chlorella. I. The effect of vanadium on photosynthesis productivity P-700 and cytochrome F in undiluted and homocontinuous cultures of Chlorella fusca. J Plant Physiol 115:115–124
- Windom HL, Tenore KT, Rice DL (1982) Metal accumulation by the polychaete *Capitella capitata:* influences of metal content and nutritional quality of detritus. Can J Fish Aquat Sci 39:191–196
- Winter M (1998) Web elements 2.0. http:rrwww.shef.ac.ukrchemistryrweb-elements. University of Sheffield

- Wright DA, Zamuda CD (1987) Copper accumulation by two bivalve molluscs: salinity effect is independent of cupric ion activity. Mar Environ Res 23(1):1–14
- Wu J, Boyle EA (1997) Lead in the western North Atlantic Ocean: completed response to leaded gasoline phaseout. Geochim Cosmochim Acta 61:3279–3283
- Wu X, Jia Y, Zhu H (2012) Bioaccumulation of cadmium bound to ferric hydroxide and particulate organic matter by the bivalve *M. meretrix*. Environ Pollut 165:133–139
- Xu B, Burnett W, Dimova N, Diao S, Mi T, Jiang X, Zhigang Y (2013) Hydrodynamics in the Yellow River estuary via radium isotopes: ecological perspectives. Cont Shelf Res 66:19–28
- Yang M, Sañudo-Wilhelmy SA (1998) Cadmium and manganese distributions in the Hudson River estuary: interannual and seasonal variability. Earth Planet Sci Lett 160:403–418
- Young RA (1995) Toxicity profiles. Toxicity summary for nickel and nickel compounds. Web site: http://risk.lsd.ornl.gov/tox/profiles/nickel. Accessed 3 Mar 2005
- Zamuda CD, Wright DA, Smucker RA (1985) The importance of dissolved organic compounds in the accumulation of copper by American oyster, *Crassostrea virginica*. Mar Environ Res 16:1–12
- Zatta P, Gobbo S, Rocco P, Perazzolo M, Favarato M (1992) Evaluation of heavy metal pollution in the Venetian lagoon by using *Mytilus galloprovincialis* as biological indicator. Sci Total Environ 119:29–41
- Zhang JS (2007) Current challenge and chance in China's Mn-industry. China Manganese Ind 1:6-9
- Zhuang GS, Yi Z, Duce RA, Brown PR (1992) Link between iron and sulphur cycles suggested by detection of Fe(II) in remote marine aerosols. Nature 355:537–539
- Zonta R, Zaggia L, Argese E (1994) Heavy metal and grain-size distributions in estuarine shallow water sediments of the Cona Marsh (Venice Lagoon, Italy). Sci Total Environ 151(1):19–28
- Zonta R, Bettiol C, Collavini F, Fagarazzi OE, Zaggia L, Zuliani A (2001) DRAIN project fresh water and pollutant transfer from the drainage basin to the Venice Lagoon. Project report, vol 15B
- Zonta R, Botter M, Cassin D, Pini R, Scattoli M, Zaggia L (2007) Sediment chemical contamination of a shallow water area close to the industrial zone of Porto Marghera (Venice Lagoon, Italy). Mar Pollut Bull 55:529–542
- Zuliani A, Zonta R, Zaggia L, Fagarazzi OE (2001) Fresh water transfer from the drainage basin to the Venice lagoon. In: LLPP, MAV, CVN (eds) Determination of the pollutant load discharged into the Venice lagoon by the drainage basin. DRAIN project workshop. Arti Grafiche Venete, Venezia, pp 31–36
- Zuliani A, Zaggia L, Collavini F, Zonta R (2005) Freshwater discharge from the drainage basin to the Venice Lagoon (Italy). Environ Int 31:929–938
- Zwolsman JJG, Van Eck BTM, Van Der Weijden CH (1997) Geochemistry of dissolved trace metals (cadmium, copper, zinc) in the Scheldt estuary, southwestern Netherlands: Impact of seasonal variability. Geochim Cosmochim Acta 61:1635–1652