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Auther(s)	Nagasawa, Kazuya; Katahira, Hirotaka; Nitta, Masato
Citation	Biogeography , 15 : 11 - 20
Issue Date	2013-08-20
DOI	
Self DOI	
URL	http://ir.lib.hiroshima-u.ac.jp/00035284
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Relation	



Isoparorchis hypselobagri (Trematoda: Isoparorchiidae) from freshwater fishes in western Japan, with a review of its host-parasite relationships in Japan (1915–2013)

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Abstract. Specimens of *Isoparorchis hypselobagri* (Billet, 1898) were collected from the following freshwater fishes in western Japan: *Anguilla japonica* (Anguillidae) from Shimane and Ehime prefectures; *Silurus asotus* (Siluridae) from Hiroshima and Yamaguchi prefectures; *Acanthogobius flavimanus* (Gobiidae) from Shimane Prefecture; *Candidia temminckii* (Cyprinidae), *Pungtungia herzi* (Cyprinidae), *Rhinogobius fluviatilis* (Gobiidae), and *Rhinogobius* sp. (Gobiidae) from Hiroshima Prefecture. The collection of *I. hypselobagri* from *A. japonica*, *A. flavimanus*, *R. fluviatilis*, and *Rhinogobius* sp. represents new host records, and the parasite is reported for the first time from Hiroshima, Yamaguchi, and Ehime prefectures. Host-parasite relationships of *I. hypselobagri* infecting Japanese freshwater fishes are reviewed based on the literature published in 1915–2013.

Key words: Isoparorchis hypselobagri, Trematoda, fish parasite, new host records

Introduction

Trematodes of the digenean family Isoparorchiidae are endoparasites of freshwater fishes in Asia and Australsia (Gibson, 2002), where *Isoparorchis hypselobagri* (Billet, 1898) has been reported from the Russian Far East (Bykhovskaya-Pavlovskaya & Kulakova, 1987), Japan (Shimazu *et al.*, 2011), China (Chen *et al.*, 1973), Vietnam (Moravec & Sey, 1989), Indonesia (Bovien, 1927), Bangladesh (Chandra, 2006), India (Bhalerao, 1936), Pakistan (Bilquees & Khatoon, 1972), and Australia (Cribb, 1988). Because of its large body size of adult worms (up to over 40 mm) and impacts on fish hosts, much attention has been paid to various aspects of the biology of the species, such as the morphology (e.g., Tewari & Pandey, 1989), physiology (e.g., Adak & Manna, 2011), body composition (e.g., Srivastava & Gupta, 1976), occurrence in fishes (e.g., Devaraji & Ranganathan, 1967; Bashirullah, 1972), and pathology (e.g., Mahajan *et al.*, 1979; Li *et al.*, 2002).

During a study of the parasite fauna of freshwater fishes of western Japan, we collected specimens of *I. hypselobagri* in Shimane, Hiroshima, Yamaguchi, and Ehime prefectures. This collection includes new host and prefecture records for the parasite. In Japan, the species has been studied for its morphology and ecology since 1915 when Kobayashi (1915a) first described it. However, many of those studies have been published in Japanese, thus it is difficult for scientists of other countries to fully understand the knowledge of the species reported in Japan. The present paper reports on our findings and, using the literature published between 1915 and 2013, reviews

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the host-parasite relationships of the species infecting Japanese freshwater fishes.

Materials and Methods

Freshwater fishes were collected at various localities in Shimane, Hiroshima, Yamaguchi, and Ehime prefectures, western Japan, in 2008-2013. Fishes were transported alive to the laboratory at Hiroshima University, where they were identified using Nakabo (2002) and examined for parasites. Trematodes were flattened between slide glass and cover-slip, fixed in 70% ethanol, stained in Heidenhain's iron hematoxyline, dehydrated in a graded ethanol series, cleared in xylene, and mounted in Canada balsam. They were measured for body length (BL, mm) and examined for maturity: worms with and without eggs in the uterus were recorded as mature and immature, respectively. Drawings were made with the aid of a drawing tube. Representative voucher specimens are deposited in the Platyhelminthes (Pl) collection housed at the National Museum of Nature and Science (NSMT-Pl) in Tsukuba, Ibraki Prefecture, Japan, and the remaining specimens are retained in the senior author's collection. The scientific and Japanese names of fishes used in this paper are adopted from Nakabo (2013) and the English names of fishes are from Froese & Pauly (2013).

Results and Discussion

Specimens of *I. hypselobagri* were collected from the following fishes in western Japan:

1) Anguilla japonica Temminck & Schlegel, 1847 (Anguilliformes: Anguillidae): one immature specimen (6.4 mm BL, site: stomach wall tissue) from the Sozu River, Ainan Town, Ehime Prefecture, on 5 July 2008; four immature specimens (2.3–5.8 mm BL, sites: mesentery and outer surface of air bladder wall, NSMT-PI 6022 [N=1, Fig. 1A]) from Lake Shinji, Izumo City, Shimane Prefecture, on 3 August 2011; and one immature specimen (9.6 mm BL, site: mesentery) from Lake Nakaumi, Matsue City, Shimane Prefecture, on 8 November 2011. 2) Candidia temminckii (Temminck & Schlegel, 1846) (Cypriniformes: Cyprinidae): one immature (6.8 mm BL, site: mesentery) and one mature (9.6 mm BL, site: mesentery, NSMT-PI 6023 [N=1]) specimens from the Nukui River, a tributary of the Kurose River, Higashi-Hiroshima City, Hiroshima Prefecture, on 26 and 28 March 2012, respectively.

3) *Pungtungia herzi* Herzenstein, 1892 (Cypriniformes: Cyprinidae): one mature (26.1 mm BL, site: body muscle, NSMT-Pl 6024 [N=1, Fig. 2]) and one immature (11.2 mm BL, site: mesentery) specimens from the Nukui River, a tributary of the Kurose River, Higashi-Hiroshima City, Hiroshima Prefecture, on 24 July 2012.

4) *Silurus asotus* Linnaues, 1758 (Siluriformes: Siluridae): three mature (12.5–20.3 mm BL, site: air bladder) and one immature (3.2 mm BL, body cavity) specimens from the Awano River, Shimonoseki City, Yamaguchi Prefecture, on 22 February 2011; four mature (22.9–45.2 mm BL, site: air bladder, NSMT-Pl 6025 [N=1]) and three mature (35.9–40.1 mm BL, site: air bladder) specimens from the Furuko River, a tributary of the Kurose River, Higashi-Hiroshima City, Hiroshima Prefecture, on 21 February and 10 July 2012, respectively.



Fig. 1. Isoparorchis hypselobagri: A, immature specimen, NSMT-PI 6022, from Anguilla japonica in Lake Shinji, Shimane Prefecture; B, immature specimen, NSMT-PI 6026, from Acanthogobius flavimanus in Lake Shinji, Shimane Prefecture. Scale bars: 1 mm.

5) Acanthogobius flavimanus (Temminck & Schlegel, 1845) (Perciformes: Gobiidae): six immature specimens (2.4–5.3 mm BL, site: body cavity, NSMT-Pl 6026 [N=1, Fig. 1B]) from Lake Shinji, Matsue City, Shimane Prefecture, on 8 January 2013.

6) *Rhinogobius fluviatilis* Tanaka, 1925 (*=Rhinogobius* sp. LD of Nakabo [2002]; LD= large-dark type [Anonymous, 1989]) (Perciformes: Gobiidae): one immature (4.2 mm BL, site: liver tissue, NSMT-Pl 6027 [N=1]) and three immature (3.3–4.4 mm BL, site: liver tissue, NSMT-Pl 6028 [N=1]) specimens from the Furuko River, a tributary of the Kurose River, Higashi-Hiroshima City, Hiroshima Prefecture, on 10 January 2012 and 13 September 2012, respectively.

7) *Rhinogobius* sp. (*=Rhinogobius* sp. OR of Nakabo [2002]; OR=orange type [Anonymous, 1989]) (Perciformes: Gobiidae): one immature



Fig. 2. *Isoparorchis hypselobagri*: mature specimen, NSMT-Pl 6024, from *Pungtungia herzi* in the Nukui River, a tributary of the Kurose River, Hiroshima Prefecture. Scale bar: 4 mm.

specimen (2.5 mm BL, site: liver tissue, NSMT-Pl 6029 [N=1]) from the Furuko River, a tributary of the Kurose River, Higashi-Hiroshima, Hiroshima Prefecture, on 13 September 2012.

Since immature and mature worms of *I.* hypselobagri were well described by Yamashita & Nishida (1955) and Shimazu *et al.* (2011), respectively, and our observation did not add new information on its morphology, no description of the specimens collected is given in this paper. Four species of fishes, *A. japonica*, *A. flavimanus*, *R. fluviatilis*, and *Rhinogobius* sp. (*=Rhinogobius* sp. OR of Nakabo [2002]), are new host records for *I. hypselobagri*. This parasite is herein reported for the first time from Hiroshima, Yamaguchi, and Ehime prefectures in Japan. The fish hosts of *I. hypselobagri* in Japan is listed in Table 1.

There was a taxonomic confusion in *R. fluviatilis* and *Rhinogobius* sp. in Japan. The former species was earlier reported as "*Rhinogobius* sp. LD" (Nakabo, 2002), but Suzuki & Chen (2011) conducted a detailed taxonomic study of gobiids of the genus and concluded that both are identical with each other: thus, *R. fluviatilis* is a current valid name. Although *Rhinogobius* sp. reported in this paper was formerly regarded as "*Rhinogobius* sp. OR" (Nakabo, 2002), the latter has been demonstrated to include several species (Suzuki *et al.*, 2010) and, following Nakabo (2013), *Rhinogobius* sp. is herein used. Future identification of *Rhinogobius* sp. examined in this study, a specimen preserved in 70% ethanol is kept in the junior author's (M.N.) collection.

Review of the host-parasite relationships of Isoparorchis hypselobagri infecting Japanese freshwater fishes

In Japan, this parasite was originally described by Kobayashi (1915a) as *Leptolecithum eurytremum* using mature specimens from the air bladder of *Silurus asotus* (as *Parasilurus asotus*) and *Tachysurus nudiceps* (as *Pseudobagrus aurantiacus*) (see below for the scientific name of this species) and immature specimens from the body cavity of these fishes,

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Table 1. Fish hosts of Isoparorchis hypeselobagri in Japan, based on the previous and present studies. The classification scheme and scientific and Japanese names of fishes are adopted from Nakabo (2013). If present, English names of fishes are also adopted from Froese & Pauly (2013).

Order	Family	Scientific name	Japanese name	English name	Prefecture	Reference
Anguilliformes	Anguillidae	Anguilla japonica	nihon-unagi	Japanese eel	Shimane, Ehime	present study
Cypriniformes	Cyprinidae	Candidia sieboldii (as Nipponocypris sieboldii)	numamutsu	*	Shiga	Shimazu et al. (2011)
		Candidia temminckii (as Zacco temminckii)	kawamutsu	dark chub	Nara, Oita, Hiroshima	Sawada & Osako (1969); Kugi & Shiote (1979); present study
		Gnathopogon elongatus elongatus	ta-moroko	—	Nagano, Shiga	Yamaguti (1938); Shimazu et al. (2011)
		Hemibarbus barbus	nigoi	_	Tokyo	Suzuki et al. (1967); Kamegai et al. (1972)
		Opsariichthys platypus (as Zacco platypus)	oikawa	freshwater minnow	Oita	Okabe (1940); Kugi & Shiote (1979)
		Pseudogobio esocinus esocinus (as Pseudogobio esocinus)	kamatsuka		Gunma, Tokyo, Shiga	Nihei <i>et al.</i> (1964); Suzuki <i>et al.</i> (1967); Shimazu <i>et al.</i> (2011)
		Pungtungia herzi	mugitsuku	_	Shiga, Hiroshima, Fukuoka	Kifune (1978); Shimazu <i>et al.</i> (2011); present study
		Sarcocheilichthys vari- egatus microoculus (as Sar- cocheilichthys variegatus)	biwa-higai	_	Ibaraki	Yamaguti (1934)
		Tanakia lanceolata	yari-tanago	_	Shiga	Shimazu et al. (2011)
		Tribolodon hakonensis (as Richardosonius hakuen- sis, Leuciscus hakonensis)	ugui	big-scalled redfin	Ibaraki	Kobayashi (1915a, 1921); Yamaguti (1934)
Siluriformes	Bagridae	Tachysurus nudiceps (as Pseudobagrus aurantiacus)	gigi	_	Ibaraki, Chiba, Shiga, Okayama	Kobayashi (1915a, 1915b, 1921)
	Siluridae	Silurus biwaensis	biwako-ō-namazu	Eurasian catfish	Shiga	Shimazu et al. (2011)
		Silurus asotus (as Parasilurus asotus)	namazu	Amur catfish	Ibaraki, Gunma, Chiba, Tokyo, Nagano, Shiga, Kyoto, Okayama, Hiroshima, Yamaguchi, Miyazaki	Kobayashi (1915a, 1915b, 1921); Yamaguti (1934); Nihei et al. (1964); Suzuki et al. (1967); Iwata et al. (2007); Shimazu (2007); Shimazu et al. (2011); present study
Salmoniformes	Osmeridae	Hypomesus nipponensis (as Hypomesus olidus)	wakasagi	Japanese smelt	Not clear**	Kobayashi (1915a, 1921)
Perciformes	Centrarchidae	Lepomis macrochirus	burūgiru	bluegill	Shiga	Shimazu et al. (2011)
		Micropterus salmoides	burakku-basu	largemouth black bass	Shiga	Grygier (2004); Shimazu <i>et al.</i> (2011)
	Cottidae	Cottus reinii	utsusemi-kajika	_	Shiga	Shimazu et al. (2011)
	Odontobutidae	Odontobutis obscura (as Mogrunda obscura)	donko	—	Not reported	Yamaguti (1934)
	Gobiidae	Acanthogobius flavimanus	mahaze	yellowfin goby	Shimane	present study
		Gymnogobius urotaenia (as Chaenogobius macro- gnathos)***	ukigori	_	Nagano, Shiga	Yamaguti (1938); Shimazu et al. (2011)
		Rhinogobius fluviatilis	ō-yoshinobori	_	Hiroshima	present study
		Rhinogobius sp. BW	biwa-yoshinobori	_	Shiga	Shimazu et al. (2011)
		Rhinogobius sp.****		_	Hiroshima	present study
		Tridentiger brevispinis	numa-chichibu	_	Shiga, Kyoto	Shimazu & Urabe (2005); Shimazu <i>et al.</i> (2011)
	Channidae	Channa argus (as Ophicephalus argus)	kamuruchī	snakehead	Ibaraki, Chiba, Shiga, Shimane, Kochi, Kumamoto	Yamashita & Nishida (1955); Komatsu & Matsumura (1963); Suzuki et al. (1967); Uehara (1972); Shimazu et al. (2011)

*: No English name. **: No English name. **: Kobayashi (1915a, 1921) collected the infected fishes of four species, including *H. nipponensis*, in Okayama, Chiba, Ibaraki, and Shiga prefectures but did not indicate the sampling locality of the parasite. ***: See Shimazu (2007) for the scientific name of this gobiid. ****: This is identical with "*Rhinogobius* sp. OR" (Nakabo, 2002).

Hypomesus nipponensis (as H. olidus) and Tribolodon hakonensis (as Richardosonius hakuensis) collected in Okayama Chiba, Ibaraki, and Shiga prefectures. Kobayashi (1915a) did not designate a type locality nor a type host for this parasite. As the original description was published in Japanese, its English version was later published (Kobayashi, 1921). In a series of papers on the endoparasitic worms from Japan, Kobayashi (1915b, 1915c, 1915d) reported anatomical features of the species (as L. eurytremum) using adult specimens from S. asotus and T. nudiceps. The same scientific name of the parasite was also used by Odhner (1927) for the Japanese specimens provided by Kobayashi. Subsequently, the species was reported as Isoparorchis trisimiltubis Southwell, 1913 (Yamaguti, 1934, 1938; Yamashita & Nishida, 1955; Uehara, 1972), but both L. eurytremum and I. trisimiltubis have been regarded as a junior synonym of I. hypselobagri in Japan (see Shimazu, 1999, 2002; Shimazu et al., 2011). Recently, Shimazu et al. (2011) redescribed the species based on specimens from fishes in the Lake Biwa basin, central Honshu, Japan, and made a detailed historical review of the synonymy of the species. No molecular work using materials from Japan and other countries has so far been conducted.

Geographical distribution

The species has been found in the following 18 prefectures in Japan (Table 1): Gunma, Ibaraki, Chiba, Tokyo, Nagano, Shiga, Kyoto, Nara, Okayama, Shimane, Hiroshima, Yamaguchi, Ehime, Kochi, Fukuoka, Oita, Miyazaki, and Kumamoto. These prefectures are located in central and western Honshu, Shikoku, and Kyushu islands, all of which belong to the temperate portion of Japan. So far, the species has not been reported from the subarctic (e.g., northern Honshu and Hokkaido) and subtropical (e.g., Okinawa) portions of Japan.

Fish hosts

The species parasitizes various freshwater fishes in Japan. Based on the previous and present studies, 24 nominal and 2 unnamed species in 10 families and 5 orders are known as its hosts (Table 1), which indicates that the host specificity of the parasite is not strict for fishes. Kobayashi (1915a) used "gigi" as the Japanese name for P. aurantiacus, but these names were a problematic use, because P. aurantiacus indicates a different species of bagrid called "gibachi" in Japanese. Thus, Kifune (1978) suggested the fish reported as "gigi" by Kobayashi (1915a) might have been "Pelteobagrus nudiceps." Shimazu et al. (2011) also made a similar suggestion. As no Kobayashi's (1915a) specimen of "gigi" or P. aurantiacus remains, it is impossible at present to clarify what species was examined by him. Therefore, following the original Japanese name of the fish, we adopt "gigi" and its most current scientific name, T. nudiceps, is used in this paper instead of P. aurantiacus. Okabe (1940) collected "Isoparorchis sp." from an unspecified host but, according to Kifune (1978), it was I. hypselobagri from Opsariichthys platypus (as Zacco platypus). Also, Iwata et al. (2007) reported "Isoparorchis sp." from S. astotus, and their specimens are herein regarded as I. hypselobagri based on the pictures (figs. 1-2 in Iwata et al., 2007). On the other hand, Shimazu (2007) found a specimen of Isoparorchis from Tandanus tandanus in Satyu Yamaguti's Collection deposited at the Meguro Parasitological Museum, Tokyo, but this host record is not included in Table 1, because he suggests that the specimen was collected in Australia.

Prevalence and intensity of infection

Infection levels of *I. hypselobagri* vary between fish species, fish sizes, or sampling localities. For example, 70.0% of 10 *Channa argus* (as *Ophicephalus argus*) from Lake Shinji, Shimane Prefecture, were infected with a mean intensity of 3.1 worms (intensity range: 0–5) (Yamashita & Nishida, 1955), while only 3.2% of 2,061 *Candidia temminckii* (as *Zacco temmickii*) from the Shirazuna River, Nara Prefecture, harbored up to 3 worms (mean intensity: 1.1 worms) (Sawada & Osako, 1969). In the latter fish sample, no worm was found in small fish (\leq 44 mm BL) but prevalence of infection steadily increased with an increase in fish size: 2.3, 8.1, and 14.5% for fish of 45–70, 71–110, and \geq 111 mm BL, respectively (Sawada & Osako, 1969).

Prevalence of infection by I. hypselobagri fluctuated seasonally in C. temminckii from the Shirazuna River (Sawada & Osako, 1969). Although it was high in late winter and spring (February to May), it decreased in early and mid-summer (June and July) and reached the lowest value in late summer (August). It, however, increased in fall (September to November) and showed high values in early and midwinter (December and January). For the observed high prevalence in winter, Sawada & Osako (1969) suggested that cercariae of this parasite may invade the fish in fall after they develop in and emerge from snail intermediate hosts during summer, but since the parasite is known to use three hosts (i.e., snail first intermediate hosts, arthropod second intermediate hosts, and fish final hosts) to complete its life cycle (Besprozvannykh & Ermolenko, 1989), it may be reasonable to consider that C. temminckii gets infected by feeding on the second intermediate hosts in fall. On the other hand, Sawada & Osako (1969) gave no explanation for the decline in prevalence in summer, but it is likely that this summer decline in prevalence was caused by mortality of infected hosts after their spawning, because C. temminckii spawns in May to August and reduces its adult density in the population during summer and fall (Katano, 1989).

Distribution and maturation in fish

When worms parasitize the body muscles of *C.* temminckii, they occur in the tunnel-like tubes built by themselves (Sawada & Osako, 1969). The coloration of the tissues surrounding the tubes changes to blackish brown, and the skin near the infection site is swollen (see fig. 1 in Sawada & Osako, 1969). Similar tube formation and blackish pigmentation also have been reported from other Japanese fishes, such as *C. argus* (Yamashita & Nishida, 1955), *Hemibarbus barbus* (Kamegai *et al.*, 1972), and *Pungtungia herzi* (Kifune, 1978). It is not yet known how much the parasite affects the physiology and behavior of those infected fishes in Japan.

Mature worms (with eggs in the uterus) are found

in the air bladder of S. asotus (Kobayashi, 1915a, 1921; Yamaguti, 1934; Sawada & Osako, 1969; Shimazu, 2007; Iwata et al., 2007; Shimazu et al., 2011; this paper) but also occur in various body parts of some other fishes, i.e., the body cavity of Silurus biwaensis (Shimazu et al., 2011), the air bladder of T. nudiceps (Kobayashi, 1915a, 1921), the body muscles of C. temminckii (Sawada & Osako, 1969; Kugi & Shiote, 1979; this paper), O. platypus (Kugi & Shiote, 1979), Lepomis macrochirus (Shimazu et al., 2011) and Micropterus salmoides (Shimazu et al., 2011), the hypodermal tissues and body muscles of P. herzi (Kifune, 1978: this paper), and around the air bladder and in the body cavity of C. argus (Suzuki et al., 1967; Shimazu et al., 2011). Mature worms from the air bladder of S. asotus were constantly large (up to 45.2 mm BL, see below), which suggests that this fish and the air bladder are quite suitable for the parasite as its final host and infection site.

Immature worms (with no eggs in the uterus) also occur in various body parts of various fish species: the body cavity of S. asotus, Odontobutis obscura (as Mogrunda obscura), C. temminckii, Gnathopogon elongatus elongates, H. barbus, O. platypus, Acanthogobius flavimanus, Gymogobius urotaenia (as Chaenogobius macrognathus), and C. argus (Kobayashi, 1915a, 1921; Yamaguti, 1934, 1938; Okabe, 1940; Yamashita & Nishida, 1955; Sawada & Osako, 1969; Kamegai et al., 1972; this paper), hypodermal (as subcutaneous) tissues of Tribolodon hakonensis (as Leuciscus hakonensis) and Sarcocheilichthys variegatus microoculus (as S. variegatus) (Yamaguti, 1934), body muscles and flesh of T. hakonensis, S. variegatus microoculus, H. barbus and C. argus (Yamaguti, 1934; Yamashita & Nishida, 1955; Suzuki et al., 1967; Kamegai et al., 1972), air bladder and air bladder wall of Anguilla japonica and O. obscura (Yamaguti, 1934; this study), gut of Rhinogobius sp. BW (Shimazu et al., 2011), stomach wall of A. japonica and Cottus reini (Shimazu et al., 2011; this paper), intestine of S. asotus (Shimazu et al., 2011), intestinal wall of C. argus (Suzuki et al., 1967), liver and liver wall of C. reini, R. fluviatilis, Rhinogobius sp. and C. argus (Yamashita &

Nishida, 1955; Suzuki *et al.*, 1967; Shimazu *et al.*, 2011; this paper), mesentery of *A. japonica*, *C. temminckii* and *P. herzi* (this paper), gonad wall of *C. argus* (Suzuki *et al.*, 1967), and gills of *T. nudiceps* (Kobayashi, 1915a, 1921). It is interesting to note that the percentage of occurrence of mature worms differs between body parts in the same host species: in *C. temminckii*, about 50% of the worms from the body muscles were mature, while the worms from the body cavity were all immature (Sawada & Osako, 1969).

Worms of various sizes are found in fish hosts. Mature and immature worms from S. asotus measured 20-40 mm BL and 8 mm BL, respectively (Shimazu et al., 2011). A similarly large mature specimen (45.2 mm BL) was also collected from S. asotus in this study. However, mature worms from other fish species do not reach such a large size (e.g., 7.8-9.3 mm BL and 26.1 mm BL from P. herzi [Kifune, 1978; this study]; 9.6 mm BL from C. temminckii [this study]: 11 mm BL from C. temminckii or O. platypus [Kugi & Shiote, 1979]), suggesting that, even if those fishes harbor mature worms, they are less suitable for the parasite than S. asotus. Sawada & Osako (1969) reported a close positive relationship between body sizes of worms and their hosts (C. temminckii): larger worms occurred in larger fish, which implies that the worms have no short-life and grow with fish growth and age.

Life cycle

The life cycle of *I. hypselobagri* is poorly known in Japan. Kobayashi (1915a, 1922) suggested that the cercaria infecting freshwater snails *Semisulcospira libertina* (as *Melania libertina*) was identifiable as the species, but his suggestion was denied by Shimazu (1999, 2003). According to Shimazu (1999, 2003) and Shimazu *et al.* (2011), the morphology of *Cercaria introverta* Faust, 1924 from *Semisulcospira* spp. in Japan (Ito, 1964; Urabe, 2003) is similar to that of the cercaria of *I. hypselobagri* which was described in the Russian Far East (Besprozvannykh & Ermolenko, 1989). On the other hand, we consider that wild fishes become infected with the parasite in two pathways through prey-predator relationships: one is feeding by fish final hosts on arthropod second intermediate hosts and the other is predation by large piscivorus fishes on small fishes. In these pathways, metacercariae and immature worms may be transferred, respectively, from the second intermediate hosts to the final hosts and from the prey fishes to the predators. Yamaguti (1934) experimentally succeeded in transferring immature worms (as larvae) from O. obscura to S. asotus and regarded O. obscura as a reservoir host. The previous works in Japan on I. hypselobagri focused on its morphology and occurrence in freshwater fishes, and our knowledge on the life cycle of the species is still limited: nothing is known, for example, about its second intermediate hosts in Japan. We need more work on various aspects of the life cycle of the parasite.

Acknowledgments

We thank Kouki Mizuno (Ehime Prefectural Uwajima Fishery High School), Tsunehiro Miura (Shimane Prefectural Fisheries Technology Center), Toshihiro Hatama (Yamaguchi Prefectural Fisheries Research Center), and Hidekatsu Yamaguchi (Hoshizaki Green Foundation) for their assistance in fish collection. We are also grateful to Yuzo Ota (Hiroshima University) for his help in fish examination. This study was supported by Grants-in-Aid for JSPS Fellows (No. 22-08025 to H.K.) and Scientific Research (C) (No. 24580267 to K.N.), both of which were from the Japan Society for the Promotion of Science (JSPS), and the Hoshizaki Green Foundation to K.N.

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(Received May 30, 2013; Accepted June 26, 2013)