

1 **Dissolved organic carbon compounds in deep-sea hydrothermal vent fluids**
2 **from the East Pacific Rise at 9°50'N**

3

4 Krista Longnecker^{§*}, Stefan M. Sievert[†], Sean P. Sylva[§], Jeffrey S. Seewald[§], and Elizabeth B.

5 Kujawinski[§]

6 [§]Woods Hole Oceanographic Institution, Marine Chemistry and Geochemistry, Woods Hole,

7 MA 02543, U.S.A.

8 [†]Woods Hole Oceanographic Institution, Biology Department, Woods Hole, MA 02543, USA

9 *Corresponding author: klongnecker@whoi.edu

10

11 Declarations of interest: none

12 Revised for submission to: *Organic Geochemistry*

13

14 **Abstract**

15 Deep-sea hydrothermal vents are unique ecosystems that may release chemically distinct
16 dissolved organic matter to the deep ocean. Here, we describe the composition and
17 concentrations of polar dissolved organic compounds observed in low and high temperature
18 hydrothermal vent fluids at 9°50'N on the East Pacific Rise. The concentration of dissolved
19 organic carbon was 46 μM in the low temperature hydrothermal fluids and 14 μM in the high
20 temperature hydrothermal fluids. In the low temperature vent fluids, quantifiable dissolved
21 organic compounds were dominated by water-soluble vitamins and amino acids. Derivatives of
22 benzoic acid and the organic sulfur compound 2,3-dihydroxypropane-1-sulfonate (DHPS) were
23 also present in low and high temperature hydrothermal fluids. The low temperature vent fluids
24 contain organic compounds that are central to biological processes, suggesting that they are a by-
25 product of biological activity in the seafloor. These compounds may fuel heterotrophic and
26 other metabolic processes at deep-sea hydrothermal vents and beyond.

27

28 *Keywords:* metabolomics; hydrothermal vents; deep-sea; dissolved organic matter; vitamins

29 **1 Introduction**

30 Dissolved organic matter in seawater is a heterogeneous mixture of compounds, each
31 with different physical and chemical properties. While cycling of seawater through the
32 seafloor during hydrothermal circulation can be a source of dissolved organic carbon (DOC)
33 to the deep ocean (Druffel and Griffin, 2015; Hedges, 1992; McCarthy et al., 2011), the chemical
34 composition and lability of this organic matter may be distinct from organic matter produced in
35 the non-vent deep ocean. These potential differences in chemical composition can have a
36 substantial impact on carbon cycling. Within hydrothermal systems, microbial communities may
37 play a fundamental role in regulating the composition of dissolved organic matter released to the
38 water column. For example, metabolites formed as by-products of chemoautotrophic activity
39 (Brault et al., 1988) represent a source of dissolved organic matter in hydrothermal fluids
40 <121°C. Hydrothermal fluids also contain hydrocarbons generated during thermal alteration of
41 microbial biomass (Dittmar, 2008; Konn et al., 2011; Reeves et al., 2014). In addition to
42 biogenic sources, dissolved organic compounds may be derived from thermal alteration of
43 dissolved organic matter initially present in circulating seawater (Hawkes et al., 2016; Rossel et
44 al., 2017) and abiotic processes that occur during hydrothermal circulation (Lang et al., 2010;
45 Lin et al., 2017; McCollom and Seewald, 2007; McDermott et al., 2015a; Proskurowski et al.,
46 2008). We posit that information on the composition of organic compounds delivered to the
47 water column by hydrothermal fluids can help identify the source of these compounds, and the
48 roles that they play in deep-sea carbon cycling.

49 Here we use a combination of direct infusion and liquid chromatography (LC)-based
50 mass spectrometry methods to analyze polar organic compounds found in hydrothermal vent

51 fluids. Direct infusion coupled to ultrahigh resolution mass spectrometers has been used in a
52 variety of ecosystems to consider large-scale differences in the composition of dissolved organic
53 matter (e.g., Kujawinski et al., 2009; Medeiros et al., 2015; Ohno et al., 2010). To complement
54 these data, we rely on targeted analytical methods that allow us to identify and quantify known
55 organic compounds (Kido Soule et al., 2015). LC-based mass spectrometry methods have proven
56 valuable in characterizing the composition of metabolites produced by marine microorganisms
57 (Fiore et al., 2015; Johnson et al., 2016). These methods can characterize biologically relevant
58 organic molecules found within seafloor hydrothermal vent ecosystems. Our results allow us
59 to compare the composition and concentration of these compounds in hydrothermal fluids
60 compared to conditions in non-vent deep seawater.

61 **2 Methods**

62 **2.1 Sample collection and processing**

63 Hydrothermal vent fluids were collected using either the ROV *Jason* or HOV *Alvin*
64 deployed from the R/V *Atlantis* during two cruises to the basalt-hosted deep-sea hydrothermal
65 vent field at 9°50'North on the East Pacific Rise in January and November 2014 (AT26-10 and
66 AT26-23). During each cruise, vent fluid samples for chemical analysis were collected from the
67 Crab Spa and Bio 9 vents, which are both located within the axial summit caldera at 9°North (see
68 map in Fornari et al., 2012). Crab Spa is a low temperature (25°C) diffuse-flow hydrothermal
69 vent (McNichol et al., 2018; McNichol et al., 2016) and Bio 9 is a high temperature (366°C)
70 focused-flow hydrothermal vent. Both sites are dominated by basalt. During the January 2014
71 cruise, a background seawater sample was collected 20 m above the seafloor, away from the
72 hydrothermal vent sites, for comparison with the vent fluids. All water samples were collected

73 using a titanium inlet snorkel consisting of a coil of narrow titanium tubing (1/8" O.D. x 0.085"
74 I.D.) connected with Teflon tubing to a FlexFoil Plus sample bag (SKC, Pennsylvania, US). The
75 titanium coil allowed high temperature fluids to cool before entering the Teflon tubing. Prior to
76 deployment, the 86 ml of dead volume in the titanium and Teflon tubing was pre-filled with
77 Milli-Q water. The FlexFoil Plus bags have been previously tested to confirm the absence of
78 contaminants that would interfere with our analyses. Water samples were pumped into the bag
79 using a peristaltic pump with PharMed BPT tubing (Masterflex) at the pump head at
80 approximately 25-40 mL minute⁻¹. Two liters of fluid were collected for all samples except for
81 the November 2014 Bio 9 sample that had 1.4 L of fluid. Vent fluid temperature was monitored
82 continuously during sample collection using a type J thermocouple attached to the inlet of the
83 sampling snorkel.

84 The fluid samples were returned to the ship-board laboratory for processing within 5 – 10
85 h of collection with the longer 10-hour delay being typical for the January 2014 samples due to
86 the use of ROV *Jason* which operates for longer periods of time at the seafloor. Water samples
87 were filtered with 0.2-µm Omnipore filters (Millipore) mounted in perfluoroalkoxy (PFA) filter
88 holders (Advantec). A 40 ml aliquot of 0.2 µm-filtered water was acidified with concentrated
89 hydrochloric acid and stored in combusted glass vials at 4°C for measurement of DOC and
90 dissolved total nitrogen (TN) using a Shimadzu TOC-V_{CSH} total organic carbon analyzer
91 equipped with a TNM-1 nitrogen analyzer in a shore-based laboratory. Blanks (Milli-Q water
92 from both the ship and the shore-based laboratory) and known concentrations of potassium
93 hydrogen phthalate and potassium nitrate were interspersed with sample runs during the analysis.
94 Comparisons to standards provided by Prof. D. Hansell (University of Miami) were made daily.

95 The coefficient of variability between replicate injections averaged <1%. Dissolved Mg
96 concentrations were analyzed by ion chromatography with suppressed conductivity detection
97 using a DIONEX DX500 system. pH measurements (25°C) were done using an Accumet
98 Ag/AgCl combination reference electrode.

99 The remaining filtrate was acidified and dissolved organic compounds were extracted
100 using Bond Elut PPL cartridges (1 g/6 ml sized cartridges, Agilent) following the protocol of
101 Dittmar et al. (2008) as modified by Longnecker (2015). DOM was eluted from the cartridges
102 using 100% methanol and stored at -20°C. The amount of DOC extracted was determined by
103 evaporating the methanol solution to dryness using a Vacufuge (Eppendorf), re-dissolving the
104 residue in Milli-Q water, and analyzing for DOC as described above. The DOC concentration in
105 the filtrate was also measured, allowing for calculation of the PPL cartridge extraction
106 efficiency.

107 In addition to the seawater and vent fluids, we processed and analyzed a Milli-Q water
108 sample in the same manner as the fluid samples. The Milli-Q water came from the Milli-Q water
109 system on board the R/V *Atlantis*, and is the same water that was used to pre-fill the sampling
110 apparatus before each deployment. Data on the concentration of compounds in the Milli-Q water
111 are provided in Table S1. These data were used to correct the concentration of each organic
112 compound present in the samples that originated from the Milli-Q water.

113 **2.2 Ultrahigh resolution mass spectrometry – direct infusion**

114 The January 2014 samples were analyzed using direct infusion with a syringe pump in
115 negative ion mode on a 7 Tesla Fourier-transform ion cyclotron resonance mass spectrometer
116 (FT-ICR-MS, Thermo Fisher Scientific, Waltham, MA) using electrospray ionization (ESI). The

117 dried organic matter extracts were reconstituted in 50:50 methanol:water and infused into the
118 ESI interface at 4 $\mu\text{L min}^{-1}$. The capillary temperature was set to 250 °C and the spray voltage
119 was between 3.7 and 4 kV. At least 200 scans were collected for each sample which is a
120 sufficient number of scans for good peak reproducibility (Kido Soule et al., 2010). The resulting
121 data are measured mass-to-charge (m/z) values and peak heights for organic compounds within
122 the extracted organic matter. Elemental formulas were assigned using the algorithm developed
123 by Kujawinski and Behn (2006). Magnitude-averaged elemental ratios were calculated following
124 the formulas provided in Sleighter and Hatcher (2008). Values for the number of condensed
125 aromatic compounds were calculated using the corrected version of the aromaticity index (Koch
126 and Dittmar, 2016).

127 **2.3 Targeted metabolomics**

128 The concentrations of 92 organic compounds were determined using targeted
129 metabolomics methods outlined by Kido Soule et al. (2015). Dried sample extracts were re-
130 dissolved in 95:5 (v/v) water:acetonitrile with deuterated biotin (final concentration 0.05 $\mu\text{g ml}^{-1}$)
131 as an internal standard. Organic compounds were chromatographically separated using a Synergi
132 4u Fusion – RP 80A 150 \times 2.00 mm column (Phenomenex, Torrance, CA) before being
133 introduced to a Thermo Scientific TSQ Vantage Triple Stage Quadrupole Mass Spectrometer via
134 a heated electrospray ionization source for mass spectrometric analysis. The chromatographic
135 separation used a binary gradient with eluent A being water with 0.1% formic acid and eluent B
136 being acetonitrile with 0.1% formic acid. Samples run at 250 $\mu\text{L min}^{-1}$ with 5% B for 0–2
137 minutes, ramp to 65% B from 2 to 20 minutes, ramp to 100% B from 20 to 25 min, and hold
138 until 32.5 minutes. The column was re-equilibrated for 7 min between samples with 95% A. The

139 mass spectrometer was operated in selected reaction monitoring (SRM) mode; optimal SRM
140 parameters (s-lens, collision energy) for each target compound were optimized individually using
141 an authentic standard. Two SRM transitions per compound were monitored for quantification
142 and confirmation. Eight-point external calibration curves based on peak area were generated for
143 each compound. The resulting data were converted to mzML files using the msConvert tool
144 (Chambers et al., 2012) and processed with MAVEN (Melamud et al., 2010). The targeted
145 metabolomics data are available from MetaboLights under study accession number MTBLS428.

146 **3 Results**

147 **3.1 Bulk chemical parameters in hydrothermal fluids**

148 The Crab Spa fluid samples contained 49 mmol/kg Mg in both January and November
149 2014. Based on this concentration, Crab Spa vent fluid contained 90% seawater and is consistent
150 with extensive seafloor mixing of cold seawater and a higher temperature zero-Mg
151 hydrothermal fluid (McNichol et al., 2016). In contrast, the Bio 9 fluids samples contained 6.7
152 and 9.4 mmol/kg Mg in January 2014 and November 2014, respectively. Because high
153 temperature vent fluids in basalt-hosted hydrothermal systems are typically characterized by
154 near-zero dissolved Mg concentrations (German and Von Damm, 2003), the measured levels of
155 dissolved Mg in the Bio 9 samples suggest entrainment of 13% and 18% ambient bottom
156 seawater during sample collection in January and November, respectively.

157 The DOC concentration in the low temperature Crab Spa fluids was slightly elevated
158 compared to the background seawater, while the high temperature Bio 9 fluids were
159 characterized by DOC concentrations substantially below seawater values (Table 1).
160 Extrapolating the DOC concentrations at Bio 9 to a fluid with zero Mg results in an endmember

161 fluid with 8.5 μM DOC. The ship-board Milli-Q system contained 16 μM of DOC. Between 22
162 and 38% of the organic carbon was extracted from the hydrothermal fluids using solid phase
163 extraction (Table 1) and thus our assessment of the composition of organic matter from the vent
164 fluids is restricted to approximately one-third of the DOM present in these fluids. The high
165 temperature Bio 9 fluids and the low temperature Crab Spa fluids both had lower TN
166 concentrations compared to background seawater. Extrapolating the Bio 9 fluids to zero Mg
167 results in an endmember fluid with 1.1 μM TN. The concentration of TN and DOC in
168 background seawater is within the range of concentrations previously measured in the deep sea
169 (Hansell et al., 2009; Ogawa et al., 1999). The pH of the fluids from Crab Spa and Bio 9 at 25°C
170 is 5.7 and 3.3, respectively.

171 **3.2 Direct infusion mass spectrometry**

172 The negative ion mode spectra from the direct infusion mass spectrometry analysis of the
173 January 2014 samples did not reveal large differences across the three samples (Supplemental
174 Figure S1). The measured m/z values provide an overview of the similarities and differences in
175 the chemical composition of organic compounds within a sample. The Crab Spa sample had
176 slightly more m/z values and a higher mean molecular weight compared to the seawater sample,
177 while the extractable organic matter in the Bio 9 fluids had a lower number of m/z values and
178 lower mean molecular weight (Table 2).

179 Elemental formulas were calculated from the m/z values. The resulting formulas can be
180 grouped based on the elements present to provide an overview of the types of organic matter
181 within the samples. For the three samples analyzed by direct infusion in this project, the
182 elemental formulas primarily contained CHO and CHON, regardless of the source of the organic

183 matter extract (Figure 1). The elemental formulas can also be considered as molar ratios of
184 hydrogen:carbon or oxygen:carbon. The weighted H:C and O:C molar ratios were highest for the
185 Crab Spa sample and lowest for the Bio 9 fluids (Table 2). The elemental ratios can also be used
186 to assess the number of condensed aromatic compounds found in hydrothermal vent fluids
187 compared to background seawater. The total number of condensed aromatic compounds was
188 highest in background seawater which had 1228 features that would correspond to individual
189 aromatic compounds based on the corrected aromaticity index (Koch and Dittmar, 2016). In
190 contrast, 894 and 1155 condensed aromatic compounds were found in the high and low
191 temperature vent fluids, respectively.

192 **3.3 Concentrations of selected metabolites in hydrothermal fluids**

193 The extraction of dissolved organic carbon from fluids by solid phase extraction is a
194 widely-used technique in aquatic sciences. By extracting our samples using the solid phase Bond
195 Elut PPL resin, we can measure a range of organic compounds and we can compare our results to
196 existing data on the organic matter found at hydrothermal vents and in seawater. However, there
197 are known issues with this method and some compounds are not well-retained by PPL (Johnson
198 et al., 2017). To account for varying levels of retention on the extraction cartridges, we assumed
199 that extraction efficiencies are not influenced by minor differences between the composition of
200 seawater and the vent fluids and corrected the measured aqueous concentrations using extraction
201 efficiencies determined by Johnson et al. (2017). We are actively working on methods to
202 improve the extraction of dissolved organic matter from aqueous solutions; in the interim,
203 caution is warranted when considering the absolute values for those compounds with low
204 extraction efficiencies. To limit errors, the correction for extraction efficiency was applied only

205 to compounds with extraction efficiencies greater than 1% due to larger errors associated with
206 correcting concentrations for compounds with lower extraction efficiencies.

207 In general, the concentrations of quantifiable organic compounds were highest in the low
208 temperature vent fluids from Crab Spa (Table S1). Furthermore, the concentrations of
209 quantifiable organic compounds are in the picomolar (10^{-12}) range, which is well below the
210 micromolar (10^{-6}) concentrations obtained for the bulk organic carbon concentrations. The
211 difference between the low temperature and high temperature vent fluids was greatest for the
212 vitamins (Figure 2). Riboflavin and pantothenic acid were present at relatively high
213 concentrations in the Crab Spa fluids in both January and November 2014. Biotin and its
214 precursor, desthiobiotin, were also prevalent in January 2014, but were present at low levels in
215 November 2014. By contrast, in the background seawater and high temperature vent fluid, the
216 total amount of water-soluble vitamins was less than 20 pM.

217 Within the hydrothermal fluid samples, amino acids formed a substantial fraction of the
218 quantifiable organic compounds. The data have been corrected for the amino acids present in the
219 Milli-Q water used to pre-fill the dead volume in the tubing prior to deploying the sampling
220 apparatus. This adjustment represents a minor fraction of the total amino acid concentrations in
221 the fluid samples with the measured concentrations in the vent fluids ranging from 4 pM to 300
222 pM while the values in MilliQ water contributed less than 6 pM. Phenylalanine concentrations in
223 seawater were intermediate to the values obtained for the high temperature Bio 9 and low
224 temperature Crab Spa fluids (Figure 3). Tryptophan was also present in most of the samples
225 with a mean value of 41 pM (range = 0 to 113 pM, n=5). In the January 2014 samples,

226 leucine/isoleucine was present at low concentrations and showed no differences across the three
227 types of samples; no leucine/isoleucine was detected in the samples collected in November 2014.

228 Dissolved organic sulfur compounds and benzoic acid derivatives were present in notable
229 amounts in the vent fluids. In particular, the organic sulfur compound 2,3-dihydroxypropane-1-
230 sulfonate (DHPS) was the most abundant organic sulfur compound present. Three additional
231 organic sulfur compounds were measured (Supplemental Table S1), but represented a total of
232 less than 5 pM of organic matter. Measureable amounts of DHPS were found in the background
233 seawater sample and in both hydrothermal fluids, reaching the highest values in the high
234 temperature Bio 9 vent fluid (Table 3). DHPS was absent from the MilliQ water. The DHPS
235 concentrations presented here substantially underestimate the actual concentration because
236 DHPS has an extraction efficiency below 1% with our methods (Johnson et al., 2017). DHPS
237 concentrations in these samples are likely more than 100x higher than presented, thus
238 approaching nM levels. However, we opted not to extrapolate to concentrations in the fluids for
239 compounds with the lowest extraction efficiencies to limit errors in quantification. Three benzoic
240 acid derivatives (2,3-dihydroxybenzoic acid, 4-aminobenzoic acid, and 4-hydroxybenzoic acid)
241 were also present in vent fluids, and were not detected in MilliQ water or ambient seawater
242 (Table 3).

243 **4 Discussion**

244 **4.1 Dissolved organic carbon in hydrothermal vent fluids**

245 Hydrothermal activity at oceanic spreading centers can be generally characterized as
246 either high-temperature (250–400 °C) focused flow or lower temperature diffuse-flow. Low
247 temperature diffuse venting reflects subsurface mixing of high-temperature hydrothermal fluids

248 with cool geochemically unmodified seawater. In this study, we measured DOC concentrations
249 as non-purgeable organic carbon and thus volatile organic compounds within hydrothermal vent
250 fluids will not be included in our DOC measurements. The DOC concentration in the high
251 temperature Bio 9 fluids was substantially less than the DOC concentration in bottom seawater
252 that represents the source of the hydrothermal fluids. Lower DOC concentrations compared to
253 ambient deep-sea water have previously been measured in high-temperature vent fluids (Hawkes
254 et al., 2015; Lang et al., 2006) and ridge flank fluids (Lin et al., 2012), indicating that these
255 systems act as sinks for DOC in the ocean. In contrast, the DOC levels in the low temperature
256 vent fluid from Crab Spa at 9° North EPR were elevated by ~2 μM above ambient seawater,
257 consistent with observations at other hydrothermal systems (Brault et al., 1988; Lang et al., 2006;
258 Lang et al., 2010). Processes known to occur in hydrothermal ecosystems that could result in
259 elevated DOC levels include the metabolic activity of the seafloor microbial community
260 (Butterfield et al., 2004; McNichol et al., 2016), the degradation products generated during
261 microbial biomass heating in seafloor environments (Reeves et al., 2014), or abiotic
262 processes such as reduction of dissolved inorganic carbon to form aqueous organic compounds
263 (Lang et al., 2010; McDermott et al., 2015b). Regardless of its source, DOC can be consumed by
264 the resident microbial community (Rossel et al., 2015) thereby fueling heterotrophic processes in
265 the deep ocean (Meier et al., 2016). Consumption of hydrothermally-derived organic carbon can
266 also be traced to higher trophic levels (Pearson et al., 2005), and we posit that the lability of
267 DOC released at the seafloor is directly linked to its chemical composition.

268 **4.2 Overall composition of dissolved organic matter**

269 To obtain compositional information about organic compounds in vent fluids, we used
270 solid phase extraction to concentrate the organic compounds and remove salt that interferes with
271 the mass spectrometry-based methods. The extraction process biases measured concentrations
272 due to variable extraction efficiencies for individual compounds. Using the Bond Elut PPL resin,
273 we were able to extract 22-38% of the organic carbon in the samples from the vents we studied, a
274 value that is generally higher than the extraction efficiencies obtained in oceanic basement fluids
275 (LaRowe et al., 2017), but is lower than seawater extracts from the Arctic (Longnecker, 2015)
276 and temperate coastal regions (Dittmar et al., 2008). The extraction of organic compounds from
277 aqueous fluids sampled in hydrothermal environments, and elsewhere, is an on-going challenge
278 in marine science. In the sections that follow we consider our organic matter compositional data
279 obtained using solid phase extraction in the context of previous research.

280 The m/z values generated by a mass spectrometry allow consideration of the diversity of
281 organic matter across samples and they can be converted into elemental formulas that provide
282 compositional information about the organic matter within a sample. Within our data, the low
283 temperature vent fluid had the highest number of m/z values while the lowest number of m/z
284 values were measured within the high temperature vent fluid. As dissolved organic matter is
285 exposed to increasingly higher temperatures (up to 380°C), decreasing numbers of m/z values are
286 measured and the remaining dissolved organic matter has smaller oxygen:carbon molar ratios
287 (Hawkes et al., 2016). Our field data are consistent with the experimental results of Hawkes et al.
288 (2016) because we observed the lowest oxygen:carbon molar ratio in the sample from the high
289 temperature vent fluids at Bio 9. Yet, the observations average the set of m/z values for bulk

290 organic matter within our samples and obscure many of the details about the composition of
291 organic compounds from hydrothermal ecosystems. In the sections that follow, we focus on
292 specific groups of organic compounds that we characterized using direct infusion mass
293 spectrometry and targeted mass spectrometry.

294 **4.3 Aromatic compounds at hydrothermal vents**

295 Mass spectrometry assessments of aromatic organic compounds provide an opportunity
296 to compare the results of direct infusion mass spectrometry with the results of targeted mass
297 spectrometry. Aromatic organic matter ranges from compounds containing a single benzene ring
298 to condensed compounds with multiple fused benzene rings. The direct infusion data presents an
299 overview of the number of compounds classified as aromatics or condensed aromatics, but does
300 not provide concentration data. On the other hand, the targeted mass spectrometry approach
301 provides concentration information for known compounds, but requires advance decisions as to
302 which compounds will be analyzed. These are fundamentally different data sets. Here, we
303 present an analysis of both datasets in order to emphasize the value of considering a combination
304 of data streams rather than relying on a single assessment of the composition of dissolved
305 organic matter.

306 For the direct infusion mass spectrometry data, we used the aromaticity index (Koch and
307 Dittmar, 2006) to estimate how many different condensed aromatic compounds were in ambient
308 seawater compared to the low and high temperature vent fluids. Based on this index, we found
309 the highest number of condensed aromatic compounds within the dissolved, solid-phase
310 extractable organic matter in ambient seawater. In contrast, while Dittmar and Koch (2006)
311 suggested that deep-sea hydrothermal vents could be a source of increased numbers of

312 condensed aromatic compounds to the deep sea, temperatures as low as 100 °C can result in
313 decreases in the number of m/z values (Hawkes et al., 2016), consistent with our observations of
314 fewer types of aromatic compounds in both the high and low temperature hydrothermal fluids
315 relative to seawater. Thus, thermal heating of organic matter in the subseafloor reduces both the
316 concentration of organic matter (Hawkes et al., 2015) and its complexity as observed by the
317 reduced number of aromatic compounds measured by ultrahigh resolution mass spectrometry.

318 Using the targeted mass spectrometry approach, we measured picomolar concentrations
319 of three substituted benzoic acid compounds in the low and high temperature hydrothermal fluids
320 while concentrations in the background seawater sample were below our detection limits (Table
321 3). Bulk concentrations of organic carbon in our samples were in the micromolar range.
322 Therefore, the aromatic compounds we quantify are only a small fraction of the organic
323 compounds in vent fluids. Monocyclic aromatic hydrocarbons, including benzoic acid and its
324 derivatives, have been previously documented in hydrothermal vent fluids (Konn et al., 2009;
325 Simoneit et al., 1988). Although monocyclic aromatic hydrocarbons are reactive under
326 hydrothermal conditions, the aromatic ring remains intact (McCollom et al., 2001). Rossel et al.
327 (2015) observed a relative increase in aromatic molecular formulas during incubations with low
328 temperature hydrothermal vent fluids, and their interpretation is that the biological community is
329 not consuming aromatic compounds which then accumulate in the incubation. We did not
330 observe consistent differences in the benzoic acid derivatives between the low- and high-
331 temperature hydrothermal vent fluids, which is surprising considering the differences in how
332 each fluid is formed. Additional research will be necessary to distinguish between microbial

333 activity and thermogenic production of benzoic acid derivatives and to constrain the short-term
334 variability of these compounds in hydrothermal vent fluids.

335 **4.4 Sulfur-containing organic compounds**

336 Hydrogen sulfide represents a substantial source of reduced sulfur within basalt-hosted
337 hydrothermal systems. Vent fluids also contain organic sulfur compounds than can serve as an
338 energy source in hydrothermal systems (Reeves et al., 2014; Rogers and Schulte, 2012). The
339 organosulfur compound 2,3-dihydroxypropane-1-sulfonate (DHPS) has been observed in the
340 surface ocean (Durham et al., 2015), but it has not previously been identified within
341 hydrothermal fluids nor in the deep sea. In the surface ocean, the presumptive source of DHPS is
342 biological degradation of sulfolipids followed by excretion of DHPS (Denger et al., 2014; Roy et
343 al., 2003), although direct release of DHPS by diatoms has also been implicated (Durham et al.,
344 2015). At present, the source of DHPS in the diffuse-flow vent fluids and seawater is not clear.
345 However, *Sulfurimonas denitrificans*, a chemolithoautotrophic campylobacterium, has been
346 found to produce elevated levels of DHPS in response to increased salt concentrations in its
347 growth medium (Götz et al., 2018). *Campylobacteria*, previously classified as
348 *Epsilonproteobacteria* (Waite et al., 2017), dominate the microbial community at Crab Spa and
349 other diffuse-flow deep-sea vents (Huber et al., 2007; Longnecker and Reysenbach, 2001;
350 McNichol et al., 2016), suggesting that *Campylobacteria* may be a source of DHPS at Crab Spa
351 and possibly hydrothermal systems elsewhere. At the same time, thermal alteration of microbial
352 biomass may release DHPS during a process that is analogous to that postulated for the
353 production of methanethiol at hydrothermal vents (Reeves et al., 2014). However, there are
354 substantial differences in DHPS concentrations between the January and November samples, and

355 measureable levels of DHPS within ambient seawater. The future development of a more
356 efficient extraction protocol for DHPS will be critical because sulfur-based organic compounds
357 such as DHPS may constitute an important carbon and energy source for the heterotrophic
358 microorganisms found within and around hydrothermal ecosystems.

359 **4.5 Vitamins and amino acids in hydrothermal fluids**

360 Low temperature diffuse flow fluids are a potential source of vitamins to the deep ocean.
361 In general, water-soluble vitamins are thermally unstable and would not persist under extended
362 periods of exposure to high temperatures. We suggest that the measureable quantities of vitamins
363 in the Crab Spa fluids reflects production by microbial communities in the seafloor at levels
364 in excess of their metabolic needs. While we are not aware of research that quantifies the
365 production of vitamins in the deep sea, the process might be similar to the accumulation of
366 vitamins such as B₂ in the surface ocean (Heal et al., 2014). In the surface ocean, many
367 phytoplankton depend on an external supply of vitamins for growth (Croft et al., 2006), and
368 changes in the concentration of vitamins in the surface ocean have been attributed to the
369 extracellular release of B vitamins by picocyanobacteria (Bonnet et al., 2010). Less is known
370 about the vitamin requirements of chemoautotrophic microorganisms, although cultured
371 chemoautotrophs for which vitamin dependency has been tested are able to grow without
372 externally supplied vitamins (e.g., Sievert et al., 2000; Takai et al., 2004a; Takai et al., 2004b).
373 The B vitamins are required for enzymatic reactions and all known carbon fixation pathways
374 (Monteverde et al., 2017), including pathways that have been described for deep-sea
375 hydrothermal vent microorganisms (Hügler and Sievert, 2011). Given these examples, it is likely
376 that chemoautotrophic microorganisms at deep-sea hydrothermal vents are a source of vitamins

377 to the deep sea. These vitamins could be consumed by organisms that cannot synthesize requisite
378 vitamins and emphasizes the role that chemoautotrophic microorganisms may play in sustaining
379 life in the deep-sea.

380 The processes responsible for the production and alteration of amino acids has long been
381 a topic of interest at deep-sea hydrothermal vents (see review by Colín-García et al., 2016).
382 Amino acid concentrations can vary in hydrothermal systems depending on the type of samples
383 collected (Haberstroh and Karl, 1989) and the timing of sample collection (Klevenz et al., 2010).
384 Our data are most comparable to measurements of dissolved free amino acids that show low
385 concentrations or concentrations below detection in vent fluids and ridge flank basement fluids
386 (Fuchida et al., 2014; Lin et al., 2015; McCollom et al., 2015). However, there are
387 methodological differences that complicate direct comparisons between the data presented in this
388 project and previous studies. While our extraction method is well suited to the analysis of
389 aromatic amino acids including tryptophan and phenylalanine, it has lower extraction efficiencies
390 for other amino acids (Johnson et al., 2017).

391 Amino acids represent a significant component of the organic compounds measured in
392 vent fluids at 9° 50' N EPR. The amino acid concentrations are temporally variable at Bio 9 and
393 Crab Spa and did not show the clear division between low temperature fluids and high
394 temperatures fluids that was apparent in the vitamin data. This result contrasts with previous
395 results for Mid-Atlantic Ridge fluids where Klevenz et al. (2010) measured higher dissolved free
396 amino acid concentrations in the low temperature diffuse hydrothermal fluids compared to the
397 high temperature fluids. The average dissolved free amino acid concentration in the samples
398 from multiple sites on the Mid-Atlantic Ridge, including Logatchev and other sites between 4°S

399 and 9°S, was 143 nM (Klevenz et al., 2010), which is comparable to the amino acid levels we
400 measured. In contrast, further north on the Mid-Atlantic Ridge (30°N to 37°N), amino acid
401 concentrations were below detection in vent fluids that spanned a range of temperatures (51°C to
402 > 360°C) and hydrothermal environments (McCollom et al., 2015). Furthermore, fluids from
403 high temperature hydrothermal fluids from the sediment-covered Guaymas Basin also had amino
404 acid levels below detection (Haberstroh and Karl, 1989). The challenges associated with
405 sampling fluids at deep-sea hydrothermal vents did not allow us to collect replicate samples from
406 each vent site and therefore we are hesitant to extrapolate past these data to consider what our
407 data may reveal about factors controlling the composition of amino acids within vent fluids.
408 Additional sampling campaigns will be necessary in order to fully quantify the temporal and
409 spatial variability in the concentration of these organic compounds.

410 **5 Conclusions**

411 Dissolved organic carbon from deep-sea hydrothermal vents can be a source of
412 chemically distinct compounds to the deep ocean. Here, we used a combination of direct infusion
413 mass spectrometry and targeted mass spectrometry to expand our view into the composition of
414 polar organic compounds that are present in deep-sea hydrothermal vent fluids. Water-soluble
415 vitamins, amino acids, benzoic acid derivatives, and organic sulfur compounds were found to
416 varying degrees within high and low temperature vent fluids. Yet, identifying the source of these
417 compounds in hydrothermal systems is complicated by the interplay between biological
418 processes and thermal heating of microbial biomass within the subsurface. The prevalence of
419 vitamins in the low temperature vent fluid is strong evidence for an active seafloor biosphere

420 that is releasing compounds that can be used as energy, carbon, and organic nutrient sources for
421 the natural microbial community in the vent fluids and the surrounding deep ocean.

422 **Acknowledgements**

423 We thank the captain and crew of the R/V *Atlantis*, the *JASON* group, and the *Alvin*
424 group for assistance with sample collection. Winn Johnson, Bill Arnold, Cara Fiore, and Florian
425 Götz provided assistance in the lab. Eugene Melamud and Jill McDermott provided useful
426 computational tools and ideas. We thank Melissa C. Kido Soule for processing the mass
427 spectrometry samples at the WHOI FT-MS Users' Facility funded by the National Science
428 Foundation (grant OCE-0619608) and the Gordon and Betty Moore Foundation (GBMF1214).
429 We appreciate editorial comments from the Kujawinski laboratory members on this manuscript.
430 This project was funded by a grant from WHOI's Deep Ocean Exploration Institute and WHOI's
431 Ocean Ridge Initiative (to EBK and SMS) and by NSF OCE-1154320 (to EBK and KL), OCE-
432 1136727 (to SMS and JSS), and OCE 1131095 (to SMS).

433 **References**

- 434 Bonnet, S., Webb, E.A., Panzeca, C., Karl, D.M., Capone, D.G., Sañudo-Wilhelmy, S.A., 2010.
435 Vitamin B₁₂ excretion by cultures of the marine cyanobacteria *Crocospaera* and
436 *Synechococcus*. *Limnology and Oceanography* 55, 1959-1964.
- 437 Brault, M., Simoneit, B.R.T., Marty, J.C., Saliot, A., 1988. Hydrocarbons in waters and
438 particulate material from hydrothermal environments at the East Pacific Rise, 13°N. *Organic*
439 *Geochemistry* 12, 209-219.
- 440 Butterfield, D.A., Roe, K.K., Lilley, M.D., Huber, J.A., Baross, J.A., Embley, R.W., Massoth,
441 G.J., 2004. Mixing, reaction and microbial activity in the sub-seafloor revealed by temporal and
442 spatial variation in diffuse flow vents at Axial Volcano, in: Wilcock, W.S.D., DeLong, E.F.,
443 Kelley, D.S., Baross, J.A., Cary, S.C. (Eds.), *The subseafloor biosphere at mid-ocean ridges*.
444 American Geophysical Union, Washington, DC, pp. 269-290.
- 445 Chambers, M.C., Maclean, B., Burke, R., Amodei, D., Ruderman, D.L., Neumann, S., Gatto, L.,
446 Fischer, B., Pratt, B., Egertson, J., Hoff, K., Kessner, D., Tasman, N., Shulman, N., Frewen, B.,
447 Baker, T.A., Brusniak, M.-Y., Paulse, C., Creasy, D., Flashner, L., Kani, K., Moulding, C.,
448 Seymour, S.L., Nuwaysir, L.M., Lefebvre, B., Kuhlmann, F., Roark, J., Rainer, P., Detlev, S.,
449 Hemenway, T., Huhmer, A., Langridge, J., Connolly, B., Chadick, T., Holly, K., Eckels, J.,
450 Deutsch, E.W., Moritz, R.L., Katz, J.E., Agus, D.B., MacCoss, M., Tabb, D.L., Mallick, P.,
451 2012. A cross-platform toolkit for mass spectrometry and proteomics. *Nature Biotechnology* 30,
452 918-920.
- 453 Colín-García, M., Heredia, A., Cordero, G., Camprubí, A., Negrón-Mendoza, A., Ortega-
454 Gutiérrez, F., Beraldi, H., Ramos-Bernal, S., 2016. Hydrothermal vents and prebiotic chemistry:
455 a review. *Boletín De La Sociedad Geológica Mexicana* 68, 599-620.
- 456 Croft, M.T., Warren, M.J., Smith, A.G., 2006. Algae need their vitamins. *Eukaryotic Cell* 5,
457 1175-1183.
- 458 Denger, K., Weiss, M., Felux, A.-K., Schneider, A., Mayer, C., Spiteller, D., Huhn, T., Cook,
459 A.M., Schleheck, D., 2014. Sulphoglycolysis in *Escherichia coli* K-12 closes a gap in the
460 biogeochemical sulphur cycle. *Nature* 507, 114-117.
- 461 Dittmar, T., 2008. The molecular-level determination of black carbon in marine dissolved
462 organic matter. *Organic Geochemistry* 39, 396-407.
- 463 Dittmar, T., Koch, B., Hertkorn, N., Kattner, G., 2008. A simple and efficient method for the
464 solid-phase extraction of dissolved organic matter (SPE-DOM) from seawater. *Limnology and*
465 *Oceanography Methods* 6, 230-235.
- 466 Dittmar, T., Koch, B.P., 2006. Thermogenic organic matter dissolved in the abyssal ocean.
467 *Marine Chemistry* 102, 208-217.

- 468 Druffel, E.R.M., Griffin, S., 2015. Radiocarbon in dissolved organic carbon of the South Pacific
469 Ocean. *Geophysical Research Letters* 42, 4096-4101.
- 470 Durham, B.P., Sharma, S., Luo, H., Smith, C.B., Amin, S.A., Bender, S.J., Dearth, S.P., Van
471 Mooy, B.A.S., Campagna, S.R., Kujawinski, E.B., Armbrust, E.V., Moran, M.A., 2015. Cryptic
472 carbon and sulfur cycling between surface ocean plankton. *Proceedings of the National Academy
473 of Sciences* 112, 453-457.
- 474 Fiore, C.L., Longnecker, K., Kido Soule, M.C., Kujawinski, E.B., 2015. Release of ecologically
475 relevant metabolites by the cyanobacterium, *Synechococcus elongatus* CCMP 1631.
476 *Environmental Microbiology* 17, 3949-3963.
- 477 Fornari, D.J., Von Damm, K.L., Bryce, J.G., Cowen, J.P., Ferrini, V., Fundis, A., Lilley, M.D.,
478 Luther, G.W., Mullineaux, L.S., Perfit, M.R., Meana-Prado, M.F., Rubin, K.H., Seyfried, W.E.,
479 Shank, T.M., Soule, S.A., Tolstoy, M., White, S.M., 2012. The East Pacific Rise Between 9
480 degrees N and 10 degrees N: Twenty-Five Years of Integrated, Multidisciplinary Oceanic
481 Spreading Center Studies. *Oceanography* 25, 18-+.
- 482 Fuchida, S., Mizuno, Y., Masuda, H., Toki, T., Makita, H., 2014. Concentrations and
483 distributions of amino acids in black and white smoker fluids at temperatures over 200 degrees
484 C. *Organic Geochemistry* 66, 98-106.
- 485 German, C.R., Von Damm, K.L., 2003. Hydrothermal Processes, in: Turekian, K.K. (Ed.),
486 *Treatise on Geochemistry*. Pergamon, Oxford, pp. 181-222.
- 487 Götz, F., Longnecker, K., Kido Soule, M.C., Becker, K.W., McNichol, J., Kujawinski, E.B.,
488 Sievert, S.M., 2018. Targeted metabolomics reveals proline as a major osmolyte in the
489 chemolithoautotroph *Sulfurimonas denitrificans*. *MicrobiologyOpen*, e00586.
- 490 Haberstroh, P.R., Karl, D.M., 1989. Dissolved free amino acids in hydrothermal vent habitats of
491 the Guaymas Basin. *Geochimica et Cosmochimica Acta* 53, 2937-2945.
- 492 Hansell, D.A., Carlson, C.A., Repeta, D.J., Schlitzer, R., 2009. Dissolved organic matter in the
493 ocean: a controversy stimulates new insights. *Oceanography* 22, 202-211.
- 494 Hawkes, J.A., Hansen, C.T., Goldhammer, T., Bach, W., Dittmar, T., 2016. Molecular alteration
495 of marine dissolved organic matter under experimental hydrothermal conditions. *Geochimica et
496 Cosmochimica Acta* 175, 68-85.
- 497 Hawkes, J.A., Rossel, P.E., Stubbins, A., Butterfield, D., Connelly, D.P., Achterberg, E.P.,
498 Koschinsky, A., Chavagnac, V., Hansen, C.T., Bach, W., Dittmar, T., 2015. Efficient removal of
499 recalcitrant deep-ocean dissolved organic matter during hydrothermal circulation. *Nature
500 Geoscience* 8, 856-860.
- 501 Heal, K.R., Carlson, L.T., Devol, A.H., Armbrust, E.V., Moffett, J.W., Stahl, D.A., Ingalls, A.E.,
502 2014. Determination of four forms of vitamin B₁₂ and other B vitamins in seawater by liquid

503 chromatography/tandem mass spectrometry. *Rapid Communications in Mass Spectrometry* 28,
504 2398-2404.

505 Hedges, J.I., 1992. Global biogeochemical cycles: progress and problems. *Marine Chemistry* 39,
506 67-93.

507 Huber, J.A., Welch, D.B.M., Morrison, H.G., Huse, S.M., Neal, P.R., Butterfield, D.A., Sogin,
508 M.L., 2007. Microbial population structures in the deep marine biosphere. *Science* 318, 97-100.

509 Hügler, M., Sievert, S.M., 2011. Beyond the Calvin Cycle: autotrophic carbon fixation in the
510 ocean. *Annual Review of Marine Science* 3, 261-289.

511 Johnson, W.M., Kido Soule, M.C., Kujawinski, E.B., 2016. Evidence for quorum sensing and
512 differential metabolite production by a marine bacterium in response to DMSP. *ISME Journal*
513 10, 2304-2316.

514 Johnson, W.M., Kido Soule, M.C., Kujawinski, E.B., 2017. Interpreting the impact of matrix on
515 extraction efficiency and instrument response in a targeted metabolomics method. *Limnology*
516 *and Oceanography Methods* 15, 417-428.

517 Kido Soule, M.C., Longnecker, K., Giovannoni, S.J., Kujawinski, E.B., 2010. Impact of
518 instrument and experiment parameters on reproducibility of ultrahigh resolution ESI FT-ICR
519 mass spectra of natural organic matter. *Organic Geochemistry* 41, 725-733.

520 Kido Soule, M.C., Longnecker, K., Johnson, W.M., Kujawinski, E.B., 2015. Environmental
521 metabolomics: analytical strategies. *Marine Chemistry* 177, Part 2, 374-387.

522 Klevenz, V., Sumoondur, A., Ostertag-Henning, C., Koschinsky, A., 2010. Concentrations and
523 distributions of dissolved amino acids in fluids from Mid-Atlantic Ridge hydrothermal vents.
524 *Geochemical Journal* 44, 387-397.

525 Koch, B.P., Dittmar, T., 2006. From mass to structure: An aromaticity index for high-resolution
526 mass data of natural organic matter. *Rapid Communications in Mass Spectrometry* 20, 926-932.

527 Koch, B.P., Dittmar, T., 2016. From mass to structure: an aromaticity index for high-resolution
528 mass data of natural organic matter. *Rapid Communications in Mass Spectrometry* 30, 250-250.

529 Konn, C., Charlou, J.L., Donval, J.P., Holm, N.G., Dehairs, F., Bouillon, S., 2009. Hydrocarbons
530 and oxidized organic compounds in hydrothermal fluids from Rainbow and Lost City ultramafic-
531 hosted vents. *Chemical Geology* 258, 299-314.

532 Konn, C., Testemale, D., Querellou, J., Holm, N.G., Charlou, J.L., 2011. New insight into the
533 contributions of thermogenic processes and biogenic sources to the generation of organic
534 compounds in hydrothermal fluids. *Geobiology* 9, 79-93.

- 535 Kujawinski, E.B., Behn, M.D., 2006. Automated analysis of electrospray ionization Fourier-
536 transform ion cyclotron resonance mass spectra of natural organic matter. *Analytical Chemistry*
537 78, 4363-4373.
- 538 Kujawinski, E.B., Longnecker, K., Blough, N.V., Del Vecchio, R., Finlay, L., Kitner, J.B.,
539 Giovannoni, S.J., 2009. Identification of possible source markers in marine dissolved organic
540 matter using ultrahigh resolution mass spectrometry. *Geochimica et Cosmochimica Acta* 73,
541 4384-4399.
- 542 Lang, S.Q., Butterfield, D.A., Lilley, M.D., Paul Johnson, H., Hedges, J.I., 2006. Dissolved
543 organic carbon in ridge-axis and ridge-flank hydrothermal systems. *Geochimica et*
544 *Cosmochimica Acta* 70, 3830-3842.
- 545 Lang, S.Q., Butterfield, D.A., Schulte, M., Kelley, D.S., Lilley, M.D., 2010. Elevated
546 concentrations of formate, acetate and dissolved organic carbon found at the Lost City
547 hydrothermal field. *Geochimica et Cosmochimica Acta* 74, 941-952.
- 548 LaRowe, D.E., Koch, B.P., Robador, A., Witt, M., Ksionzek, K., Amend, J.P., 2017.
549 Identification of organic compounds in ocean basement fluids. *Organic Geochemistry* 113, 124-
550 127.
- 551 Lin, H.-T., Amend, J.P., LaRowe, D.E., Bingham, J.-P., Cowen, J.P., 2015. Dissolved amino
552 acids in oceanic basaltic basement fluids. *Geochimica et Cosmochimica Acta* 164, 175-190.
- 553 Lin, H.-T., Cowen, J.P., Olson, E.J., Amend, J.P., Lilley, M.D., 2012. Inorganic chemistry, gas
554 compositions and dissolved organic carbon in fluids from sedimented young basaltic crust on the
555 Juan de Fuca Ridge flanks. *Geochimica et Cosmochimica Acta* 85, 213-227.
- 556 Lin, Y.-S., Koch, B.P., Feseker, T., Ziervogel, K., Goldhammer, T., Schmidt, F., Witt, M.,
557 Kellermann, M.Y., Zabel, M., Teske, A., Hinrichs, K.-U., 2017. Near-surface heating of young
558 rift sediment causes mass production and discharge of reactive dissolved organic matter.
559 *Scientific Reports* 7, 44864.
- 560 Longnecker, K., 2015. Dissolved organic matter in newly formed sea ice and surface seawater.
561 *Geochimica et Cosmochimica Acta* 171, 39-49.
- 562 Longnecker, K., Reysenbach, A.-L., 2001. Expansion of the geographic distribution of a novel
563 lineage of ϵ -Proteobacteria to a hydrothermal vent site on the Southern East Pacific Rise. *FEMS*
564 *Microbiology Ecology* 35, 287-293.
- 565 McCarthy, M.D., Beaupre, S.R., Walker, B.D., Voparil, I., Guilderson, T.P., Druffel, E.R.M.,
566 2011. Chemosynthetic origin of ^{14}C -depleted dissolved organic matter in a ridge-flank
567 hydrothermal system. *Nature Geoscience* 4, 32-36.
- 568 McCollom, T.M., Seewald, J.S., 2007. Abiotic synthesis of organic compounds in deep-sea
569 hydrothermal environments. *Chemical Reviews* 107, 382-401.

570 McCollom, T.M., Seewald, J.S., German, C.R., 2015. Investigation of extractable organic
571 compounds in deep-sea hydrothermal vent fluids along the Mid-Atlantic Ridge. *Geochimica et*
572 *Cosmochimica Acta* 156, 122-144.

573 McCollom, T.M., Seewald, J.S., Simoneit, B.R.T., 2001. Reactivity of monocyclic aromatic
574 compounds under hydrothermal conditions. *Geochimica et Cosmochimica Acta* 65, 455-468.

575 McDermott, J.M., Ono, S., Tivey, M.K., Seewald, J.S., Shanks, W.C., Solow, A.R., 2015a.
576 Identification of sulfur sources and isotopic equilibria in submarine hot-springs using multiple
577 sulfur isotopes. *Geochimica et Cosmochimica Acta* 160, 169-187.

578 McDermott, J.M., Seewald, J.S., German, C.R., Sylva, S.P., 2015b. Pathways for abiotic organic
579 synthesis at submarine hydrothermal fields. *Proceedings of the National Academy of Sciences*
580 112, 7668-7672.

581 McNichol, J., Stryhanyuk, H., Sylva, S.P., Thomas, F., Musat, N., Seewald, J.S., Sievert, S.M.,
582 2018. Primary productivity below the seafloor at deep-sea hot springs. *Proceedings of the*
583 *National Academy of Sciences* 115, 6756-6761.

584 McNichol, J., Sylva, S.P., Thomas, F., Taylor, C.D., Sievert, S.M., Seewald, J.S., 2016.
585 Assessing microbial processes in deep-sea hydrothermal systems by incubation at in situ
586 temperature and pressure. *Deep Sea Research Part I: Oceanographic Research Papers* 115, 221-
587 232.

588 Medeiros, P.M., Seidel, M., Powers, L.C., Dittmar, T., Hansell, D.A., Miller, W.L., 2015.
589 Dissolved organic matter composition and photochemical transformations in the northern North
590 Pacific Ocean. *Geophysical Research Letters* 42, 863-870.

591 Meier, D.V., Bach, W., Girguis, P.R., Gruber-Vodicka, H.R., Reeves, E.P., Richter, M.,
592 Vidoudez, C., Amann, R., Meyerdierks, A., 2016. Heterotrophic *Proteobacteria* in the vicinity of
593 diffuse hydrothermal venting. *Environmental Microbiology*, n/a-n/a.

594 Melamud, E., Vastag, L., Rabinowitz, J.D., 2010. Metabolomic analysis and visualization engine
595 for LC-MS data. *Analytical Chemistry* 82, 9818-9826.

596 Monteverde, D.R., Gómez-Consarnau, L., Suffridge, C., Sañudo-Wilhelmy, S.A., 2017. Life's
597 utilization of B vitamins on early Earth. *Geobiology* 15, 3-18.

598 Ogawa, H., Fukuda, R., Koike, I., 1999. Vertical distributions of dissolved organic carbon and
599 nitrogen in the Southern Ocean. *Deep Sea Research Part I: Oceanographic Research Papers* 46,
600 1809-1826.

601 Ohno, T., He, Z., Sleighter, R.L., Honeycutt, C.W., Hatcher, P.G., 2010. Ultrahigh resolution
602 mass spectrometry and indicator species analysis to identify marker components of soil- and
603 plant biomass-derived organic matter fractions. *Environmental Science & Technology* 44, 8594-
604 8600.

- 605 Pearson, A., Seewald, J.S., Eglinton, T.I., 2005. Bacterial incorporation of relict carbon in the
606 hydrothermal environment of Guaymas Basin. *Geochimica et Cosmochimica Acta* 69, 5477-
607 5486.
- 608 Proskurowski, G., Lilley, M.D., Seewald, J.S., Früh-Green, G.L., Olson, E.J., Lupton, J.E.,
609 Sylva, S.P., Kelley, D.S., 2008. Abiogenic hydrocarbon production at Lost City hydrothermal
610 field. *Science* 319, 604-607.
- 611 Reeves, E.P., McDermott, J.M., Seewald, J.S., 2014. The origin of methanethiol in midocean
612 ridge hydrothermal fluids. *Proceedings of the National Academy of Sciences* 111, 5474-5479.
- 613 Rogers, K.L., Schulte, M.D., 2012. Organic sulfur metabolisms in hydrothermal environments.
614 *Geobiology* 10, 320-332.
- 615 Rossel, P.E., Stubbins, A., Hach, P.F., Dittmar, T., 2015. Bioavailability and molecular
616 composition of dissolved organic matter from a diffuse hydrothermal system. *Marine Chemistry*
617 177, Part 2, 257-266.
- 618 Rossel, P.E., Stubbins, A., Rebling, T., Koschinsky, A., Hawkes, J.A., Dittmar, T., 2017.
619 Thermally altered marine dissolved organic matter in hydrothermal fluids. *Organic*
620 *Geochemistry* 110, 73-86.
- 621 Roy, A.B., Hewlins, M.J.E., Ellis, A.J., Harwood, J.L., White, G.F., 2003. Glycolytic breakdown
622 of sulfoquinovose in bacteria: a missing link in the sulfur cycle. *Applied and Environmental*
623 *Microbiology* 69, 6434-6441.
- 624 Sievert, S.M., Heidorn, T., Kuever, J., 2000. *Halothiobacillus kellyi* sp. nov., a mesophilic,
625 obligately chemolithoautotrophic, sulfur-oxidizing bacterium isolated from a shallow-water
626 hydrothermal vent in the Aegean Sea, and emended description of the genus *Halothiobacillus*.
627 *International Journal of Systematic and Evolutionary Microbiology* 50 Pt 3, 1229-1237.
- 628 Simoneit, B.R.T., Kawka, O.E., Brault, M., 1988. Origin of gases and condensates in the
629 Guaymas Basin hydrothermal system (Gulf of California). *Chemical Geology* 71, 169-182.
- 630 Sleighter, R.L., Hatcher, P.G., 2008. Molecular characterization of dissolved organic matter
631 (DOM) along a river to ocean transect of the lower Chesapeake Bay by ultrahigh resolution
632 electrospray ionization Fourier transform ion cyclotron resonance mass spectrometry. *Marine*
633 *Chemistry* 110, 140-152.
- 634 Takai, K., Hirayama, H., Nakagawa, T., Suzuki, Y., Nealson, K.H., Horikoshi, K., 2004a.
635 *Thiomicrospira thermophila* sp. nov., a novel microaerobic, thermotolerant, sulfur-oxidizing
636 chemolithomixotroph isolated from a deep-sea hydrothermal fumarole in the TOTO caldera,
637 Mariana Arc, Western Pacific. *International Journal of Systematic and Evolutionary*
638 *Microbiology* 54, 2325-2333.

639 Takai, K., Nealson, K.H., Horikoshi, K., 2004b. *Hydrogenimonas thermophila* gen. nov., sp.
640 nov., a novel thermophilic, hydrogen-oxidizing chemolithoautotroph within the ϵ -
641 *Proteobacteria*, isolated from a black smoker in a Central Indian Ridge hydrothermal field.
642 International Journal of Systematic and Evolutionary Microbiology 54, 25-32.

643 Waite, D.W., Vanwonterghem, I., Rinke, C., Parks, D.H., Zhang, Y., Takai, K., Sievert, S.M.,
644 Simon, J., Campbell, B.J., Hanson, T.E., Woyke, T., Klotz, M.G., Hugenholtz, P., 2017.
645 Comparative genomic analysis of the class *Epsilonproteobacteria* and proposed reclassification
646 to Epsilonbacteraeota (phyl. nov.). Frontiers in Microbiology 8.

647

648

649

650 Table 1. Measured temperature, dissolved magnesium (Mg), dissolved organic carbon (DOC),
651 and dissolved total nitrogen (TN) concentrations for water samples collected from 9°50'N, East
652 Pacific Rise. Solid phase extraction was used to process fluids for mass spectrometry analysis
653 and the extraction efficiency is the percent of organic carbon captured by the solid phase
654 extraction method. 'n.a.' in the table indicates the value was not determined.

	Date	Temperature (°C)	Mg (mmol/kg)	DOC (µM)	TN (µM)	Extraction efficiency
Seawater	Jan. 2014	2 °C	53.5	44.3	43.2	31%
Crab Spa	Jan. 2014	25 °C	48.9	46.5	12.7	22 %
	Nov. 2014	24 °C	49.1	46.2	15.6	n.a.
Bio 9	Jan. 2014	366 °C	6.7	13.6	5.6	38%
	Nov. 2014	364 °C	9.4	14.2	9.4	n.a.

655 Table 2. Parameters for negative ion mode, direct infusion data collected from the January 2014
 656 samples. Data are number of m/z values, the number of elemental ratios for all m/z values with
 657 elemental formulas, mean molecular weight, and magnitude-averaged molar ratios calculated
 658 following Sleighter and Hatcher (2008).

Station	# of m/z values	# of elemental formulas	Mean molecular weight	H:C_w	O:C_w
Seawater	6475	6226	458.61	1.13	0.42
Crab Spa	7363	7059	467.13	1.18	0.45
Bio 9	4644	4314	425.83	1.01	0.35

661
 662 Table 3. Concentration of dissolved 2,3-dihydroxypropane-1-sulfonate (DHPS) and benzoic acid
 663 derivatives in seawater, and high and low temperature hydrothermal vent fluids at 9°50'N East
 664 Pacific Rise. The concentration data in the table have been corrected to consider the extraction
 665 efficiency of each compound (Johnson et al., 2017), except for DHPS where uncorrected (*)
 666 measured data are provided. 'b.d.' are values below detection given the analytical approach.

667

Station	Date	DHPS (pM)	2,3-dihydroxybenzoic acid (pM)	4-aminobenzoic acid (pM)	4-hydroxybenzoic acid (pM)
Seawater	Jan. 2014	9.5 (*)	b.d.	b.d.	b.d.
Crab Spa	Jan. 2014	4.3 (*)	1.1	57.7	17.7
Crab Spa	Nov. 2014	17.3 (*)	2.5	4.0	52.9
Bio 9	Jan. 2014	8.7 (*)	1.3	b.d.	10.3
Bio 9	Nov. 2014	30.1 (*)	8.8	1.2	52.4

668

669

670 **Figure legends**

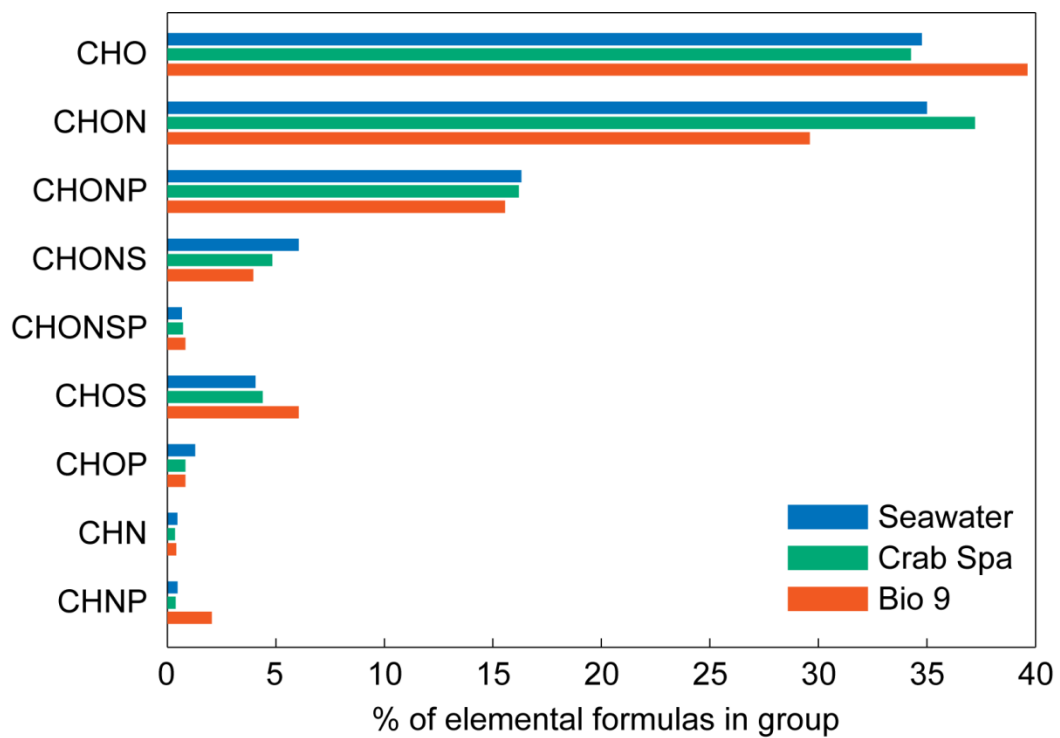
671 Figure 1. Percentage of elemental formulas assigned to each compositional group for seawater,
672 Crab Spa, and Bio 9 organic matter extracts from the January 2014 samples. Only elemental
673 formulas that represented more than 5% of the total elemental formulas are plotted.

674 Figure 2. Concentrations of vitamin B₂, B₅, B₇, and vitamin B₇'s precursor, desthiobiotin in
675 seawater and hydrothermal fluids from Crab Spa and Bio 9 vents.

676 Figure 3. Concentrations of tryptophan, phenylalanine, and leucine/isoleucine in seawater and
677 hydrothermal fluids from Crab Spa and Bio 9 vents.. Using the targeted metabolomics method
678 described in section 2.3, leucine and isoleucine cannot be quantified separately.

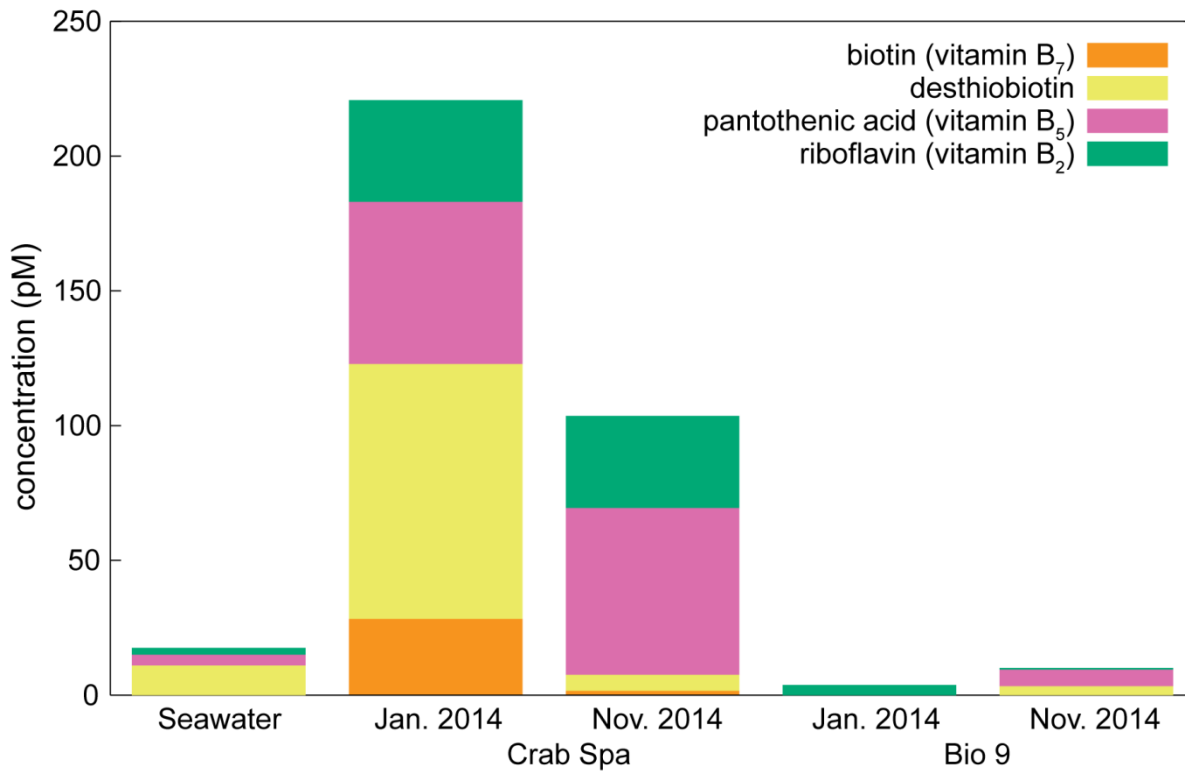
679

681 Figure 1

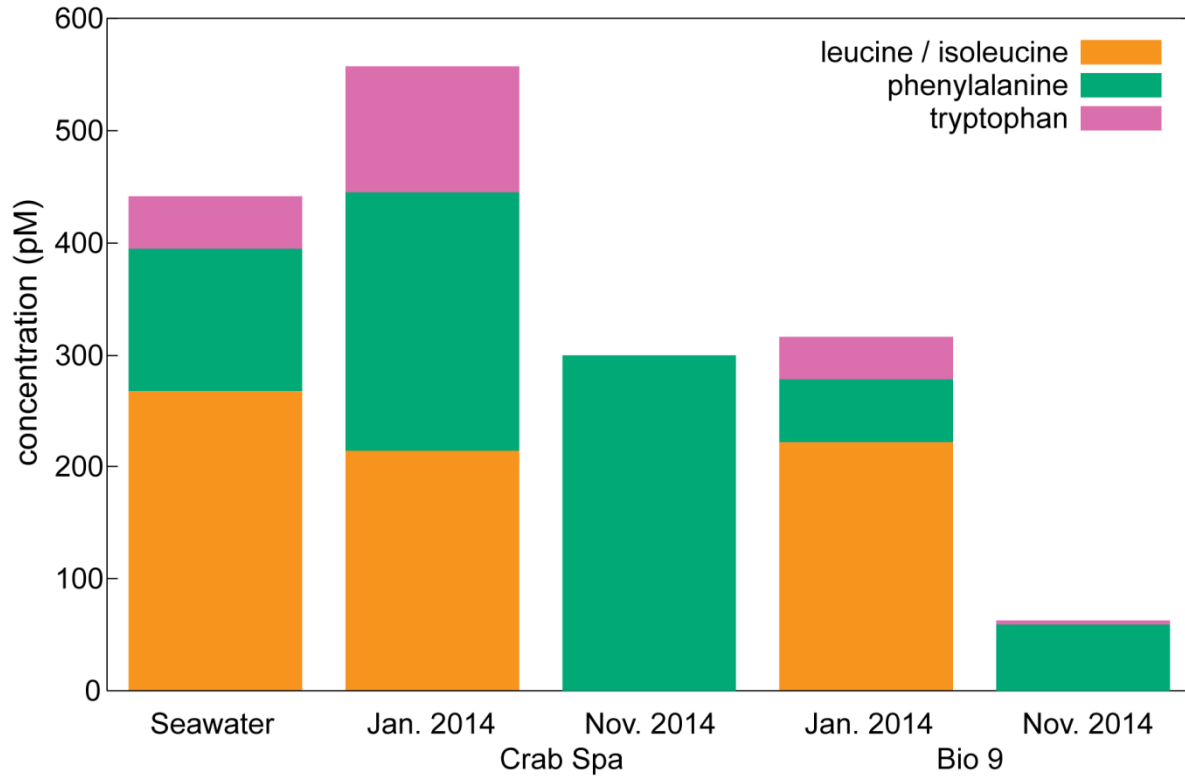


682

684 Figure 2



686 Figure 3



Dissolved organic carbon compounds in deep-sea hydrothermal vent fluids from the East Pacific Rise at 9°50'N

Krista Longnecker^{§*}, Stefan M. Sievert[†], Sean P. Sylva[§], Jeffrey S. Seewald[§], and Elizabeth B. Kujawinski[§]

[§]Woods Hole Oceanographic Institution, Marine Chemistry and Geochemistry, Woods Hole, MA 02543, U.S.A.

[†]Woods Hole Oceanographic Institution, Biology Department, Woods Hole, MA 02543, USA

*Corresponding author: klongnecker@whoi.edu

Supplemental information

Supplemental Table S1

List of metabolites detected in the extracellular fractions of dissolved organic matter. Four types of samples are given in the table: Milli-Q water, seawater, low temperature (low T) vent fluid from Crab Spa, and high temperature (high T) vent fluid from Bio 9. In each case, the fluids were filtered and the dissolved organic compounds extracted as described in the text.

Concentrations are given in picomolar, and are corrected for the extraction efficiency of each compound based on the values in Johnson et al. (2017). Compounds marked with [*] have extraction efficiencies less than 1% and the data in the table are the measured values that have not been corrected for the extraction efficiency of the compound. If no value is given in the table, the sample was below the detection limit for the metabolite; limit of detection and limit of quantification for these compounds are provided in Johnson et al. (2017). ‡The following amino acids were below detection in this project: arginine, glutamine, proline, glutamate, cysteine, serine, and homoserine/threonine.

The fluid samples were 2 liters except for the November 2014 Bio 9 sample that was 1.4 L of fluid. Each sample included 86 ml of Milli-Q water because the titanium and Teflon tubing was filled with Milli-Q water before each deployment. We have corrected the seawater and vent fluid samples to account for the concentration of each metabolite in the Milli-Q water.

References:

Johnson, W.M., Kido Soule, M.C., Kujawinski, E.B., 2017. Interpreting the impact of matrix on extraction efficiency and instrument response in a targeted metabolomics method. *Limnology and Oceanography Methods* 15, 417-428.

			Low T vent fluid		High T vent fluid	
	Milli-Q water	Seawater	Crab Spa		Bio 9	
	Jan. 2014	Jan. 2014	Jan. 2014	Nov. 2014	Jan. 2014	Nov. 2014
<i>Vitamins</i>						
biotin (vitamin B ₇)	1		28	2		
cyanocobalamin (vitamin B ₁₂)			0	0		
desthiobiotin (precursor to biotin)		11	95	6		3
folic acid (vitamin B ₉)		1	1	0		
pantothenic acid (vitamin B ₅)	1	4	60	62		6
riboflavin (vitamin B ₂)		2	38	34	4	1
<i>Amino acids</i> ‡						
leucine / isoleucine	45	268	214		222	
phenylalanine	92	127	231	300	57	60
tryptophan	21	47	112		38	4
<i>Dissolved organic sulfur compounds</i>						
2,3-dihydroxypropane-1-sulfonate [*]		10	4	17	9	30
3-mercapto proprionate					4	
5'-deoxy-5'(methylthio)adenosine	0		1	0	1	
dimethylsulfoniopropionate (DMSP) [*]					1	
<i>Dissolved organic phosphorus compounds</i>						
6-phosphogluconic acid [*]						3
D-glucosamine 6-phosphate [*]		4				
nicotinamide adenine dinucleotide (NAD)					1	
nicotinamide adenine dinucleotide phosphate (NADP)		9				
D-glucose 6-phosphate [*]		0	2			
glyphosate			8			

			Low T vent fluid		High T vent fluid	
	Milli-Q water	Seawater	Crab Spa		Bio 9	
	Jan. 2014	Jan. 2014	Jan. 2014	Nov. 2014	Jan. 2014	Nov. 2014
<i>Nucleic acid precursors</i>						
adenosine		14	45	30	11	9
guanosine						10
inosine 5'-monophosphate [*]			1			
inosine				122		
uridine 5-monophosphate [*]			5			
xanthine [*]				1		1
xanthosine				3		3
<i>Benzoic acid derivatives</i>						
2,3-dihydroxybenzoic acid			1	3	1	9
4-aminobenzoic acid			58	4		1
4-hydroxybenzoic acid	1		18	53	10	52
<i>Other metabolites</i>						
chitotriose						16
choline [*]				7		7
indole 3-acetic acid	1		50	1		1
kynurenine						6
n-acetyl glutamic acid			237		96	
n-acetyl muramic acid				383		
sodium taurocholate			3	5		
tryptamine				5		5

Supplemental Figure S1. Negative ion mode spectra from DOM extracted from seawater, low temperature vent fluids from Crab Spa, and high temperature vent fluids from Bio 9.

