

# Increase in mercury in Pacific yellowfin tuna

Paul E. Drevnick<sup>1,\*</sup>, Carl H. Lamborg<sup>2</sup>, and Martin J. Horgan<sup>3</sup>

<sup>1</sup>University of Michigan Biological Station and School of Natural Resources and Environment, Ann Arbor, MI 48109 USA

<sup>2</sup>Woods Hole Oceanographic Institution, Department of Marine Chemistry & Geochemistry, Woods Hole, MA 02543 USA

<sup>3</sup>230 Northridge Dr., Oxford, OH 45056

\*Corresponding Author: \*E-mail: [drevnick@umich.edu](mailto:drevnick@umich.edu). Phone (734) 763-6280.

## Abstract

Mercury is a toxic trace metal that can accumulate to levels that threaten human and environmental health. Models and empirical data suggest that humans are responsible for a great deal of the mercury actively cycling in the environment at present. Thus, we would predict that the concentration of mercury in fish should have increased dramatically since the Industrial Revolution. Evidence in support of this hypothesis has been hard to find, however, and some studies have suggested that analyses of fish show no change in mercury concentration. By compiling and re-analyzing published reports on yellowfin tuna (*Thunnus albacares*) caught near Hawai'i over the past half century, we find that the concentration of mercury in these fish is currently increasing at a rate  $\geq 3.8\%$  per year. This rate of increase is consistent with a model of anthropogenic forcing on the mercury cycle in the North Pacific, and suggests fish mercury concentrations are keeping pace with current loadings increases to the ocean. Future increases in mercury in yellowfin tuna and other fishes can be avoided by reductions in atmospheric mercury emissions from point sources.

## Introduction

Mercury is a potent toxin that can accumulate to high concentrations in fish, posing a health risk to humans who eat fish. Methylmercury, the predominant form of mercury in fish, is formed from mercuric ions (Hg[II]) by microbes and perhaps abiotically in waters and sediments, then enters the base of the food web and increases in concentration with each successive trophic level. Consumption of mercury-contaminated fish from gross pollution events, e.g., in Minamata, Japan [1] where fish with concentrations as high as 36 ppm could "easily be captured by hand", has resulted in severe neurologic damage in humans, most acutely in children exposed prenatally via maternal fish consumption. In waters not directly affected by local pollution, mercury concentrations in fish are typically  $< 1$  ppm. However, even at this lower level, prenatal exposure is associated with developmental deficits [2]. In the U.S. annually, Trasande et al. [3] found that *c.* 300,000-600,000 children are born with mercury concentrations in cord blood that exceed  $5.8 \mu\text{g/L}$ , a value associated with significant loss of IQ (intelligence quotient); the economic consequence of which is estimated to be US\$8.7 billion annually in lost income.

Humans are exposed to methylmercury primarily via consumption of ocean fish [4], and while models (see review in [5]) and empirical data [6] suggest an increase in mercury content of the global ocean since preindustrial times, a corresponding increase in

1  
2  
3 47 methylmercury has not been shown for ocean fish. Local pollution of nearshore  
4 48 environments occurs where there are point-source water discharges, but for open ocean  
5 49 the principal source of mercury is atmospheric deposition [7]. Atmospheric mercury is  
6 50 now dominated by human contributions (primarily from fossil fuel combustion and  
7 51 artisanal gold mining), and present-day rates of mercury deposition are 3-5x greater than  
8 52 natural (pre-anthropogenic) rates [8]. It had been thought that mercury pollution could  
9 53 only result in a negligible increase in mercury concentration in open ocean waters [9], but a  
10 54 recent synthesis of data from water column profiles of total mercury points to a 2.6x  
11 55 increase (since the 1500s) in waters shallower than 1000 m, globally [6].

12 56 Here we show for a commercially important species, the yellowfin tuna (*Thunnus*  
13 57 *albacares*), a temporal increase in mercury concentration. Mercury data for Pacific  
14 58 yellowfin tuna from waters near Hawai'i present a unique record in that the same  
15 59 population/location was sampled three different time periods over 37 years, muscle tissue  
16 60 samples were analyzed for Hg, and data were reported in peer-reviewed literature. A  
17 61 network of fish aggregation devices in Hawai'i have documented the high site fidelity of  
18 62 yellowfin tuna [10]. The following is a report of our compilation, re-analysis, and  
19 63 interpretation of the mercury data.  
20 64

## 21 65 **Methods**

22 66 We compiled published reports on mercury in yellowfin tuna caught from waters of the  
23 67 North Pacific Ocean near Hawai'i during 1971 [11, 12], 1998 [13], and 2008 [14]. From  
24 68 each specimen, muscle tissue was subsampled and measured for total mercury. Data are  
25 69 comparable across studies because of strict adherence to quality assurance/quality control  
26 70 (QA/QC) that ensured accuracy of data. Muscle tissue samples from 1971 [11, 12] and  
27 71 1998 [13] were acid digested and analyzed by cold-vapor atomic absorption  
28 72 spectrophotometry. Rivers et al. [11] reported the mean recovery of spiked samples was  
29 73 101% and that the result for each sample was validated by separate analysis of  
30 74 methylmercury, indeed confirming that nearly all of total mercury is methylmercury [15].  
31 75 Thieleke [12; see also 16] had each sample analyzed by two independent laboratories, with  
32 76 excellent agreement between laboratories, as the maximum deviation among samples was  
33 77 6%. Kraepiel et al. [13] reported detailed QA/QC procedures, including sample handling  
34 78 and use of duplicates (mean relative percent difference of 4.7%), spikes (mean recovery of  
35 79 96.7%), and a reference sample (mean concentration of 0.128 ppm with standard deviation  
36 80 of 0.0098 ppm on 6 samples) for analyses. Muscle tissue samples from 2008 [14] were  
37 81 analyzed by thermal decomposition, amalgamation, and atomic absorption  
38 82 spectrophotometry. Choy et al. [14; see also 17] also reported detailed QA/QC procedures,  
39 83 including analysis of duplicates (data only accepted if relative percent difference between  
40 84 duplicates was <5%) and certified reference materials (mean recoveries of DORM-3 and  
41 85 High Purity Standard Trace Metal Fish were 100.3% and 96.1%, respectively).

42 86 To compare Hg concentration among the three data sets (1971, 1998, and 2008), we  
43 87 used the analysis-of-covariance (ANCOVA) model, with body size as the covariate. ANCOVA  
44 88 is the standard parametric test for comparing a characteristic of groups of subjects while  
45 89 controlling for the effect of another variable on that characteristic. Controlling for the  
46 90 effect of body size when assessing mercury concentration among groups of fish is thus a  
47 91 classic use of ANCOVA. Fish from 22–76 kg were included in the analysis as this size range  
48 92 ( $\pm 5$  kg) was common to all three data sets. It was necessary to remove the fish < 22 kg,  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

93 because these fish did not adhere to the assumption of linearity. Mercury concentrations in  
94 young tuna tend to be low, but highly variable [18]. A diet shift occurs in young tuna when  
95 a critical body mass is developed that enables endothermic capability to allow access to  
96 prey in deeper, colder water [19]. At a certain size (depending on species), likely due to  
97 this ontogenetic diet shift, the mercury concentration versus size relationship conforms to  
98 expectations (i.e, a linear relationship). Outliers, identified with Tukey box plots and  
99 confirmed by one-sided Grubbs' tests, were also removed from the data sets.

## 101 Results

102 The ANCOVA revealed that slopes of the relationships between mercury concentration and  
103 fish size were not statistically different among the three time periods ( $F_{2,223} = 1.17, P =$   
104  $0.31$ ; Fig. 1), but mercury concentrations were higher in 2008 than either 1971 or 1998  
105 ( $F_{2,225} = 11.6, P < 0.0001$ ; Tukey's HSD; Fig. 1). Accordingly, the average mercury  
106 concentration (least square mean  $\pm$  std. error) was considerably higher in 2008 ( $0.336$   
107  $\pm 0.023$ ) than in 1971 ( $0.229 \pm 0.008$ ), or 1998 ( $0.218 \pm 0.008$ ). Sample size (for 1971,  
108 1998, and 2008,  $n = 111, 104,$  and  $14,$  respectively) is incorporated in the ANCOVA, and  
109 with the significant result, it can be ruled out – with 95% probability – that the effect of  
110 sample year was due to chance. The fourteen data points from 2008 are elevated relative  
111 to the two other data sets. Note that we conducted statistical diagnostics and found five  
112 points among the three datasets with potentially high leverage or high influence. We  
113 performed the ANCOVA without these points and found no change in the qualitative results  
114 of the ANCOVA (no difference in slopes, significant difference in intercepts).

## 116 Discussion

117 Our analysis, at least for the early part of the record, is in agreement with that of Kraepiel et  
118 al. [13], who found no change in tuna mercury between the same 1971 and 1998 datasets.  
119 This conclusion led Kraepiel et al. [13] to hypothesize that methylmercury forms from  
120 mercury naturally occurring in deep waters, sediments, or possibly hydrothermal vents  
121 and is therefore largely natural. Subsequently developed independent lines of evidence  
122 have suggested that vents are not strong enough sources of mercury to supply foodwebs  
123 [20], that fish do acquire methylated mercury from shallow depths in the ocean [21], and  
124 that seawater mercury concentrations are increasing ocean-wide [6], including near  
125 Hawai'i [22]. Thus, we should expect to see changes in the concentration of mercury in  
126 tuna now and in the foreseeable future.

127 Such changes are discernable in the recent part of the record, from 1998 to 2008,  
128 which shows an increase in mercury concentration in tuna at a rate of  $\geq 3.8\%$  per year, in  
129 agreement with recent and modeled changes in mercury cycling in the North Pacific (Fig. 2).  
130 Sunderland et al. [22] found a significant rise in mercury concentrations in seawater at all  
131 depths (0-1000 m) from 2002 to 2006. The largest increases, modeled at 3% per year  
132 between 1995-2006, are occurring in intermediate waters (150-1000 m), which in addition  
133 to receiving mercury from atmospheric deposition to surface waters above (0-150 m) also  
134 receive inputs associated with lateral flow of mercury-enriched waters from the coast of  
135 the Northwest Pacific [22, 23]. Yellowfin tuna near Hawai'i spend most of their time in  
136 surface waters or immediately below [24]. The agreement between the data/modeling by  
137 Sunderland et al. [22] and the updated tuna record compiled and re-analyzed here provides

1  
2  
3 138 support for the alternative hypothesis that mercury and methylmercury concentrations in  
4 139 the ocean are increasing due to human activity and that anthropogenic methylmercury  
5 140 accumulates in important commercial fish.

7 141 A criticism of the Kraepiel et al. [13] study, that effects of fishing on oceanic food  
8 142 webs may have affected methylmercury accumulation in tuna [25], also applies here. Data  
9 143 for age, growth, and trophic level are not available for the individual fish in our synthesis.  
10 144 In the Pacific Ocean, the largest tunas have become more rare, but no detectable change in  
11 145 trophic level has occurred in any population [26]. Growth statistics of yellowfin tuna  
12 146 appear unchanged over the past half century [27], but the data aren't ideal for determining  
13 147 temporal trends. One could expect an increase in growth rate, as a density-dependent  
14 148 response to over-exploitation of the population [28]. With increased growth, there would  
15 149 be "growth dilution" of mercury [29]. To sum, there are no data to suggest Pacific yellowfin  
16 150 tuna have different growth rates or trophic level for the study period, but if either of those  
17 151 two factors have changed, it would likely be in a direction that would tend to lower the  
18 152 concentration of mercury in tuna, masking the effect of increased mercury loading to the  
19 153 system.

22 154 Because fish mercury concentrations appear to be increasing in step with modeled  
23 155 loadings, these data indicate that mercury concentrations in open ocean fish are responsive  
24 156 to anthropogenic mercury releases. Lamborg et al. [6] suggested that if atmospheric  
25 157 mercury emissions continue to increase, the deepwater sink will become 'overwhelmed' in  
26 158 the coming decades. Therefore, it should be expected that the rate of increase in mercury  
27 159 in surface waters (0-1000 m) will be greater than the rate of increase in emissions.  
30 160 Mercury contamination of ocean fish is a serious global health issue, now being addressed  
31 161 by the UN Minamata Convention on Mercury. Current goals of the convention – that aim to  
32 162 reduce atmospheric mercury emissions from point sources – would result in avoided  
33 163 increases in rates of atmospheric mercury deposition [30]. However, even if current  
34 164 deposition rates are maintained, North Pacific intermediate waters are expected to double  
35 165 in mercury concentration by 2050 [22]. Thus, more stringent reductions in emissions are  
36 166 necessary.

37 166

38 167

39 168

40 168

41 169

42 170

43 171

44 172

45 173

46 174

47 175

48 176

49 177

50 178

51 179

52 180

53 181

54 180

55 181

56 181

57 181

58 181

59 181

60 181

### Supplemental Data

The data reported were previously published [11-14], but can also be found in Table S1.

### Acknowledgments

PED was supported by the University of Michigan and the Fonds de recherche du Québec – Nature et Technologies. CHL was supported by NSF OCE-1129339, 1232760 and the Woods Hole Oceanographic Institution. Bruce Monson, Ed Swain, and anonymous reviewers provided constructive comments on earlier drafts of the manuscript. The authors declare no competing financial interests or any other conflicts of interest.

### References

[1] Harada M. 1995. Minamata disease: methylmercury poisoning in Japan caused by environmental pollution. *Crit. Rev. Toxicol.* 25:1-24.

- 1  
2  
3 182 [2] McKelvey W, Oken E. 2012. Mercury and Public Health: An Assessment of Human  
4 183 Exposure. In Banks MS, Ed, *Mercury in the Environment: Pattern and Process*, University  
5 184 of California Press, Oakland, CA, pp 267-287.
- 7 185 [3] Trasande L, Landrigan PJ, Schechter C. 2005. Public health and economic consequences  
8 186 of methyl mercury toxicity to the developing brain. *Environ. Health Perspect.* 113:590-  
9 187 596.
- 11 188 [4] Sunderland EM. 2007. Mercury exposure from domestic and imported estuarine and  
12 189 marine fish in the U.S. seafood market. *Environ. Health Perspect.* 115:235-242.
- 13 190 [5] Black FJ, Conaway CH, Flegal AR. 2012. Mercury in the marine environment. In Banks  
14 191 MS, Ed, *Mercury in the Environment: Pattern and Process*, University of California Press,  
15 192 Oakland, CA, pp. 167-219.
- 17 193 [6] Lamborg CH, Hammerschmidt CR, Bowman KL, Swarr GJ, Munson KM, Ohnemus DC,  
18 194 Lam PJ, Heimbürger L-E, Rijkenberg MJA, Saito MA. 2014. A global ocean inventory of  
19 195 anthropogenic mercury based on water column measurements. *Nature* 512:65-68.
- 20 196 [7] Mason RP, Fitzgerald WF, Morel FMM. 1994. The biogeochemical cycling of elemental  
21 197 mercury: anthropogenic influences. *Geochim. Cosmochim. Acta* 58:3191-3198.
- 23 198 [8] Engstrom DR, Fitzgerald WF, Cooke CA, Lamborg CH, Drevnick PE, Swain EB, Balogh SJ,  
24 199 Balcom PH. 2014. Atmospheric Hg emissions from preindustrial gold and silver  
25 200 extraction in the Americas: a reevaluation from lake sediment archives. *Environ. Sci.*  
26 201 *Technol.* 48:6533-6543.
- 28 202 [9] Hammond AL. 1971. Mercury in the environment: natural and human factors. *Science*  
29 203 171:788-789.
- 30 204 [10] Itano DG, Holland KN. 2000. Movement and vulnerability of bigeye (*Thunnus obesus*)  
31 205 and yellowfin tuna (*Thunnus albacares*) in relation to FADs and natural aggregation  
32 206 points. *Aquat. Living Resour.* 13:213-223.
- 34 207 [11] Rivers JB, Pearson JE, Shultz CD. 1972. Total and organic mercury in marine fish. *Bull.*  
35 208 *Environ. Contam. Toxicol.* 8:257-266.
- 36 209 [12] Thieleke JR. 1973. Mercury Levels in Five Species of Commercially Important Pelagic  
37 210 Fish Taken from the Pacific Ocean Near Hawaii. Ph.D. Dissertation, University of  
38 211 Wisconsin-Madison.
- 40 212 [13] Kraepiel AML, Keller K, Chin HB, Malcolm EG, Morel FMM. 2003. Sources and  
41 213 variations of mercury in tuna. *Environ. Sci. Technol.* 37:5551-5558.
- 42 214 [14] Choy CA, Popp BN, Kaneko JJ, Drazen JC. 2009. The influence of depth on mercury  
43 215 levels in pelagic fishes and their prey. *Proc. Natl. Acad. Sci. U.S.A.* 106:13865-13869.
- 44 216 [15] Bloom NS. 1992. On the chemical form of mercury in edible fish and marine  
45 217 invertebrate tissue. *Can. J. Fish. Aquat. Sci.* 49:1010-1017.
- 47 218 [16] Boush GM, Thieleke JR. 1983. Total mercury content in yellowfin and bigeye tuna. *Bull.*  
48 219 *Environ. Contam. Toxicol.* 30:291-297.
- 49 220 [17] Choy CA. 2013. Pelagic Food Web Connectivity in the North Pacific Subtropical Gyre: A  
50 221 Combined Perspective from Multiple Biochemical Tracers and Diet. Ph.D. Dissertation,  
51 222 University of Hawai'i at Manoa.
- 53 223 [18] Cumont G, Viallex G, Lelièvre H, Bobenrieth P. 1975. Mercury Contamination in Sea  
54 224 Fish. Fisheries and Marine Service Canada, Translation Series No. 3373, Halifax, NS.
- 55 225 [19] Graham BS, Grubbs D, Holland K, Popp BN. 2006. A rapid ontogenetic shift in the diet  
56 226 of juvenile yellowfin tuna from Hawai'i. *Mar. Biol.* 150:647-658.
- 57  
58  
59  
60

- 1  
2  
3 227 [20] Lamborg CH, Von Damm KL, Fitzgerald WF, Hammerschmidt CR, Zierenberg R. 2006.  
4 228 Mercury and monomethylmercury in fluids from Sea Cliff submarine hydrothermal field,  
5 229 Gorda Ridge. *Geophys. Res. Lett.* 33:L17606, doi:10.1029/2006GL026321.
- 7 230 [21] Blum JD, Popp BN, Drazen JC, Choy CA, Johnson MW. 2013. Methylmercury production  
8 231 below the mixed layer in the North Pacific Ocean. *Nat. Geosci.* 6:879-884.
- 10 232 [22] Sunderland EM, Krabbenhoft DP, Moreau JW, Strode SA, Landing WM. 2009. Mercury  
11 233 sources, distribution, and bioavailability in the North Pacific Ocean: insights from data  
12 234 and models. *Glob. Biogeochem. Cycles* 23:GB2010, doi:10.1029/2008GB003425.
- 13 235 [23] Hammerschmidt CR, Bowman KL. 2012. Vertical methylmercury distribution in the  
14 236 subtropical North Pacific Ocean. *Mar. Chem.* 132-133:77-82.
- 16 237 [24] Brill RW, Block BA, Boggs CH, Bigelow KA, Freund EV, Marcinek DJ. 1999. Horizontal  
17 238 movements and depth distribution of large adult yellowfin tuna (*Thunnus albacares*)  
18 239 near the Hawaiian Islands, recorded using ultrasonic telemetry: Implications for the  
19 240 physiological ecology of pelagic fishes. *Mar. Biol.* 133:395-408.
- 20 241 [25] Renner R. 2004. Where is the mercury? *Environ. Sci. Technol.* 38:12A.
- 22 242 [26] Sibert J, Hampton J, Kleiber P, Maunder M. 2006. Biomass, size, and trophic status of  
23 243 top predators in the Pacific Ocean. *Science* 314:1773-1776.
- 24 244 [27] Zhu G, Xu L, Dai X, Liu W. 2011. Growth and mortality rates of yellowfin tuna, *Thunnus*  
25 245 *albacares* (Perciformes: Scombridae), in the eastern and central Pacific Ocean. *Zoologia*  
26 246 28:199-206.
- 27 247 [28] Polacheck T, Eveson JP, Laslett GF. 2004. Increase in growth rates of southern bluefin  
28 248 tuna (*Thunnus maccoyii*) over four decades: 1960 to 2000. *Can. J. Fish. Aquat. Sci.*  
29 249 61:307-322.
- 31 250 [29] Newman MC, Unger MA. 2003. *Fundamentals of Ecotoxicology*, 2<sup>nd</sup> ed., CRC Press, Boca  
32 251 Raton, FL.
- 33 252 [30] Selin NE. 2014. Global change and mercury cycling: challenges for implementing a  
34 253 global mercury treaty. *Environ. Toxicol. Chem.* 33:1202-1210.
- 35 254  
36 255

### 37 255 **Figure Captions**

38 256 **Fig. 1.** Linear regressions of fish size (kg) versus mercury concentration (parts-per-million)  
39 257 for three datasets for yellowfin tuna from North Pacific waters near Hawai'i; 1971 (ref. 11,  
40 258 12; black):  $Hg = -0.0799 + 0.0068 \times \text{mass}$ ,  $n = 111$ ,  $r^2 = 0.413$ ,  $P < 0.0001$ ; 1998 (ref. 13;  
41 259 red):  $Hg = -0.1619 + 0.0083 \times \text{mass}$ ,  $n = 104$ ,  $r^2 = 0.375$ ,  $P < 0.0001$ ; 2008 (ref. 14; green):  
42 259  $Hg = -0.0718 + 0.0093 \times \text{mass}$ ,  $n = 14$ ,  $r^2 = 0.656$ ,  $P = 0.0004$ .

44 261  
45 262 **Fig. 2.** Least square mean ( $\pm$  std err) mercury concentration in yellowfin tuna from 1971  
46 263 (black), 1998 (red), and 2008 (green) from waters near Hawai'i. Overlaid on the fish data  
47 264 are mercury concentrations in seawater; gray squares ( $\pm$  std dev) represent point  
48 265 estimates from integrated 1000-m profiles in the eastern North Pacific, and the solid gray  
49 266 line represents modeled trends for intermediate waters (150-1000 m) basin wide (dashed  
50 267 lines represent 95% confidence interval). Seawater data and model output are from  
51 268 Sunderland et al. [22].

52 267  
53 268  
54  
55  
56  
57  
58  
59  
60

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

Fig. 1.

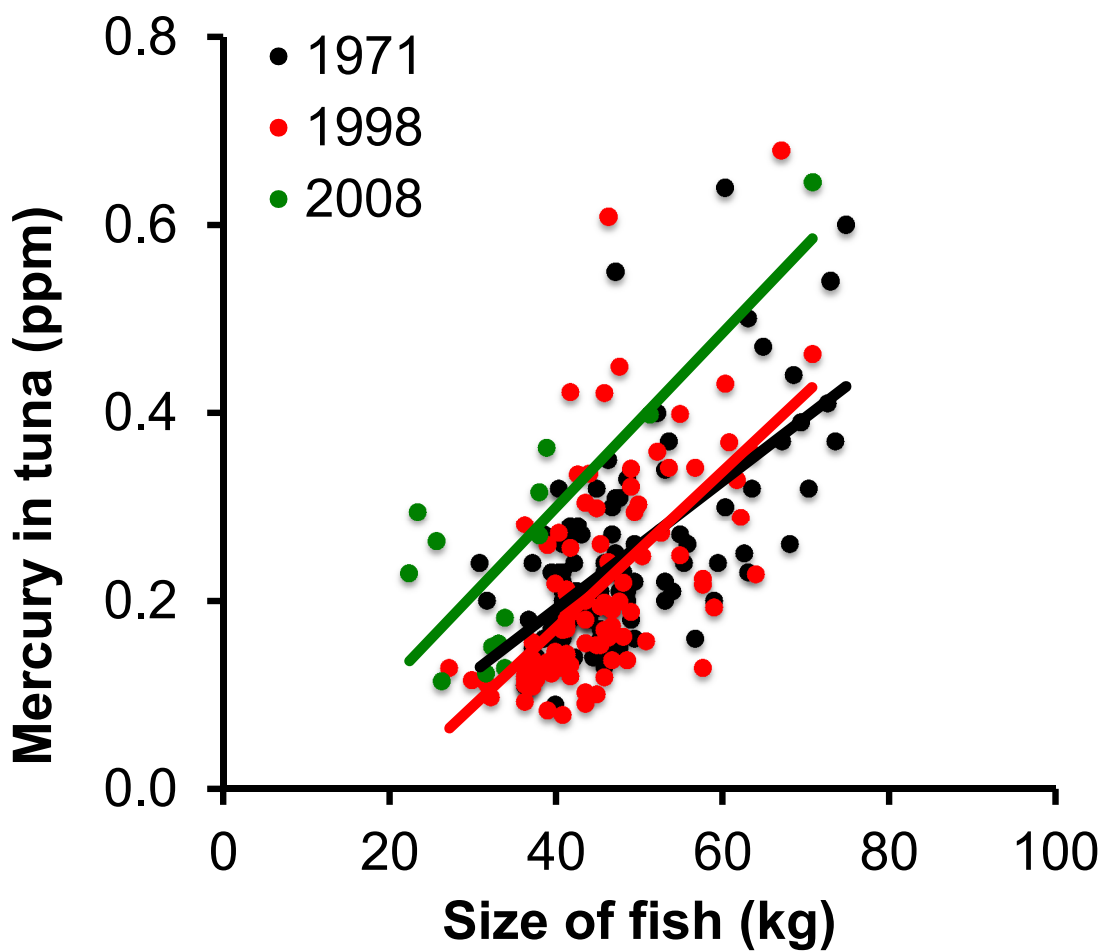
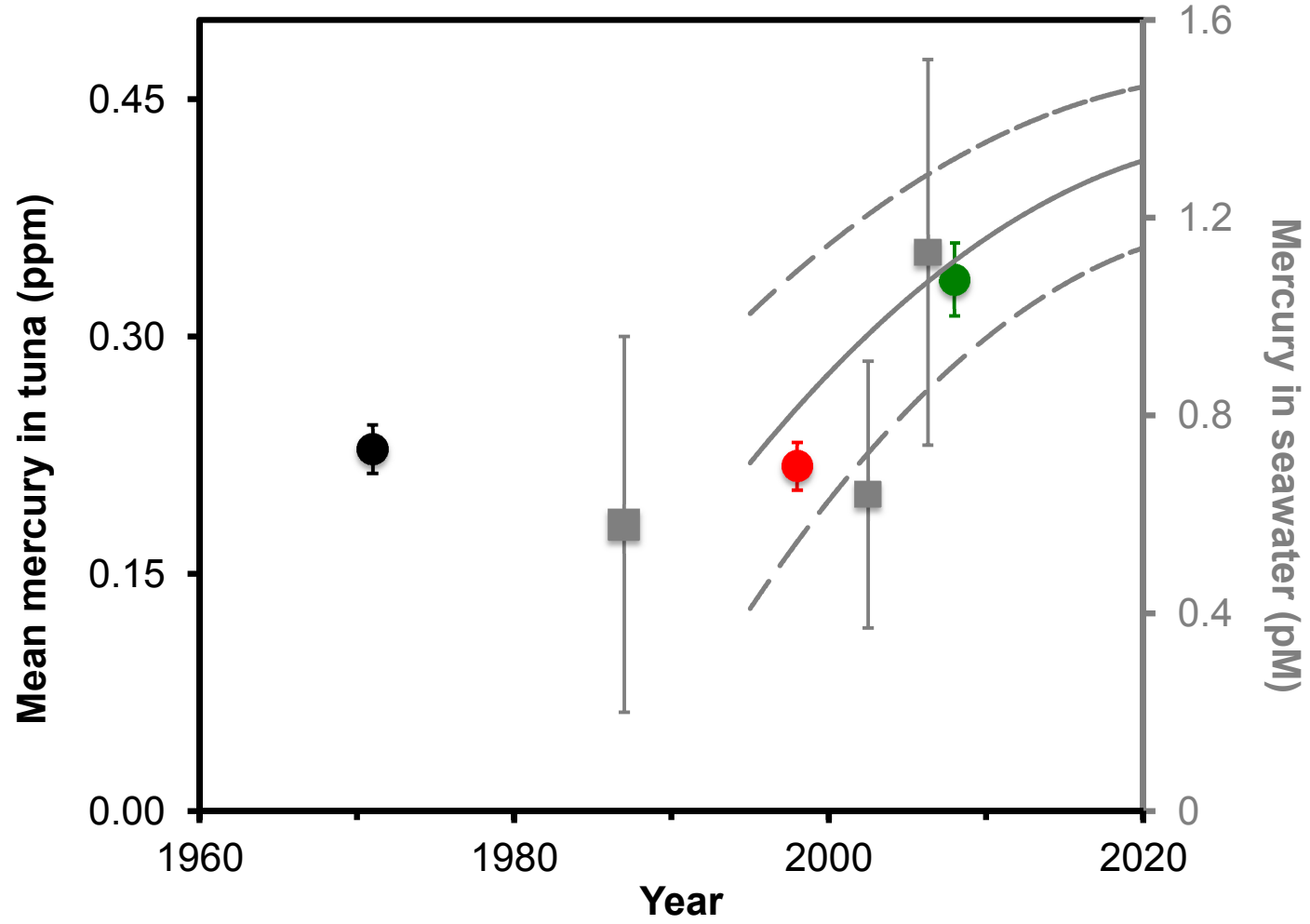


Fig. 2.



1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49