1	Molecular signature of organic nitrogen in septic-impacted groundwater
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## 26 Abstract

### 27

28 Dissolved inorganic and organic nitrogen levels are elevated in aquatic systems due to 29 anthropogenic activities. Dissolved organic nitrogen (DON) arises from various sources, and its 30 impact could be more clearly constrained if specific sources were identified and if the molecular-31 level composition of DON were better understood. In this work, the pharmaceutical 32 carbamazepine was used to identify septic-impacted groundwater in a coastal watershed. Using ultrahigh resolution mass spectrometry data, the nitrogen-containing features of the dissolved 33 34 organic matter in septic-impacted and non-impacted samples were compared. The septic-35 impacted groundwater samples have a larger abundance of nitrogen-containing formulas. 36 Impacted samples have additional DON features in the regions ascribed as 'protein-like' and 37 'lipid-like' in van Krevelen space and have more intense nitrogen-containing features in a 38 specific region of a carbon versus mass plot. These features are potential indicators of dissolved 39 organic nitrogen arising from septic effluents, and this work suggests that ultrahigh resolution 40 mass spectrometry is a valuable tool to identify and characterize sources of DON. 41

## 42 Introduction

43 Groundwater is an important source of freshwater to coastal systems.<sup>1, 2</sup> Thus,

44 groundwater is a source of nitrogen to coastal waters, especially if it has been impacted by

45 human activities. Dissolved nitrogen is comprised of both inorganic (nitrate, nitrite, and

46 ammonia) and organic forms. The quantitative balance between inorganic and organic nitrogen

47 will depend on various factors, including sources and biological activity, but evidence suggests

48 that dissolved organic nitrogen (DON) can be quantitatively as important as inorganic forms of

49 nitrogen in groundwater.<sup>3</sup> For example, Kroeger et al.<sup>3</sup> found that DON comprised 10-93% of

50 total dissolved nitrogen in a set of coastal watersheds. Both the source and environmental

51 conditions influence the bioavailability and processing of DON, with bioavailability ranging

52 from 1-60%.<sup>4, 5</sup> Sources of DON include precipitation, agriculture, natural runoff/infiltration,

53 artificial recharge of wastewater, and septic systems. In populated coastal areas, septic systems

54 may be particularly important sources of both inorganic and organic nitrogen.<sup>3, 6-8</sup> At present,

bowever, it is difficult to link the bulk DON found in impacted aquatic systems to specific

56 sources or processes due to a lack of information about the molecular-level composition of DON.

57 Information regarding the molecular-level composition of DON is important to 58 improving understanding the reactivity of the pool of nitrogen in aquatic systems. The amount 59 of DON present is changed (both increases and decreases have been observed) by human alteration of the landscape or introduction of runoff/effluents.<sup>9, 10</sup> For example, septic systems are 60 a major source of dissolved nitrogen in low-residential watersheds.<sup>11</sup> Compositional changes in 61 62 the DON pool in human altered systems have received limited attention, and such changes have 63 potentially important ramifications. For example, DON is more recalcitrant than inorganic 64 nitrogen, leading to longer residence times and a larger zone of impact. Additionally, DON 65 concentration and composition will affect water treatment and human exposure to nitrogenous disinfection byproducts.<sup>12</sup> 66

67 Ultrahigh resolution mass spectrometry techniques, such as Fourier transform ion 68 cyclotron resonance mass spectrometry (FT-ICR-MS), are used to gain insight into the molecular-level composition of dissolved organic matter (DOM).<sup>13-17</sup> Such instruments provide 69 70 the opportunity to see differences among samples that may not be apparent through bulk analyses (e.g., measurement of dissolved organic carbon (DOC)) or targeted analysis of specific 71 72 molecules. FT-ICR-MS has been used to analyze incorporation of nitrogen into organic matter and transformations and sources of the DON pool.<sup>18-20</sup> While recent studies have shown 73 compositional differences between effluent organic matter and natural organic matter,<sup>21, 22</sup> a 74 75 specific focus on the nitrogen-containing component has yet to be undertaken. That said, past 76 work on septic-impacted groundwater on Cape Cod (MA) noted that a large portion of assigned formulas contained nitrogen, potentially due to microbial activity and only partial mineralization 77 of septic contributions to the DON pool.<sup>17</sup> 78

79 Groundwater impacted by septic system effluent contains numerous human-use compounds, including antibiotics, over-the-counter pharmaceuticals, steroid estrogens, 80 prescription medications, artificial sweeteners, and personal care products.<sup>23-27</sup> On Cape Cod. 81 82 studies have found such compounds in impacted groundwater, which also serves as the drinking water source for the population.<sup>27, 28</sup> Some of these compounds, including carbamazepine (an 83 84 anti-convulsant) and artificial sweeteners, appear to be persistent and sorb to soils or sediments only to a limited extent. Thus, they have been proposed as markers for wastewater/septic impact 85 and for contaminant source tracking.<sup>26, 29-32</sup> In particular, carbamazepine is generally recognized 86 87 as being persistent in groundwater and an indicator of impact on natural waters by untreated or treated wastewater.<sup>29, 30</sup> Laboratory studies have shown carbamazepine to be stable in on-site 88 89 treatment scenarios,<sup>33</sup> and recent results demonstrated the utility of carbamazepine in discriminating nutrient sources in coastal groundwater.<sup>8</sup> 90

On Cape Cod, the vast majority of residents have on-site wastewater treatment,<sup>34</sup> and 91 92 nationwide, 20% of households rely on septic systems. Even when functioning properly, septic 93 systems are a source of nitrogen to groundwater. Because of the prevalence of septic systems on Cape Cod, past detection of pharmaceuticals in groundwater,<sup>27, 28</sup> known nitrogen pollution 94 issues,<sup>3, 35, 36</sup> and the fact that the aquifer to which septic systems discharge is also the source of 95 96 drinking water, this is a logical site to investigate the composition of DON in groundwater with 97 and without potential septic system impact. Our goals were to both discern septic impacted sites 98 and begin to characterize the DON arising from septic tanks. In this work, electrospray ionization 99 (ESI) FT-ICR-MS was used to obtain detailed information about the dissolved organic nitrogen 100 component of DOM, with carbamazepine used as a marker to identify impacted vs. non-101 impacted waters.

## 103 Experimental

All solvents were obtained from Fisher and were Optima<sup>®</sup> grade. Trace metal grade 104 105 hydrochloric acid was also from Fisher. Ultrapure water (18.2 M $\Omega$ ·cm) was produced by a 106 Millipore Elix 3 and Gradient system. Carbamazepine (Sigma-Aldrich) standards (0.1-1,000 ng/mL and a d<sub>10</sub>-carbamazepine (Cambridge Isotopes) spiking solution in methanol were 107 prepared previously.<sup>37</sup> All groundwater samples were collected via a peristaltic pump into 108 109 prewashed (overnight soak with soap and water, rinsed with ultrapure water, soaked in 10% HCl, 110 rinse with ultrapure water) and autoclaved 2.5 L polycarbonate bottles. The bottles were rinsed 111 three times with  $\sim 1$  L of collected groundwater and then filled to capacity, with 5 L being 112 collected for each sample. The samples were transported on ice to the laboratory, where they were filtered through 0.2 µm, 47 mm hydrophilic PTFE Omnipore<sup>TM</sup> membranes (EMD-113 Millipore) and acidified to pH 3 using concentrated HCl. If not extracted immediately, the 114 filtered and acidified samples were stored at 4 °C. 115

116 Sample sites

117 Four sites in Falmouth (Cape Cod), MA were sampled in September and October of 2013. The shallow groundwater at two of the sites, Head of Waquoit Bay and Sage Lot Pond, 118 with population densities of 190 and 0 people per km<sup>2</sup> respectively, was expected to have 119 minimal, if any, impact from septic systems.<sup>3</sup> The groundwater sample at Head of Waquoit Bay 120 121 was taken within the Waquoit Bay National Estuarine Research Reserve east of the boathouse at the north point of the transect described by Kroeger and Charette,<sup>36</sup> using a small diameter drive 122 point piezometer.<sup>38</sup> Groundwater at the Sage Lot Pond site was collected in a forested area from 123 124 previously installed wells. The other two sites, Green Pond and Site D, were near residences in

more densely populated areas  $(1,000-1,500 \text{ people per km}^2)^3$  and were expected to be impacted 125 126 by septic system effluent. Groundwater at Green Pond was collected on the west shore 127 approximately 0.4 kilometers north of the pond outlet into Vineyard Sound. A small diameter 128 drive point piezometer was used approximately 5 m from shore. At Site D, a densely populated 129 residential area west of Waquoit Bay, samples were collected at three depths (6.30, 9.97, and 130 16.11 m below ground surface) during ongoing USGS sampling efforts from a multilevel well. 131 These specific depths for characterization of the DOM and quantification of carbamazepine were 132 chosen based on nitrate and ammonia levels observed in previous sampling efforts to capture 133 potentially different redox zones.

#### 134 *DOC and water quality analysis*

For the six samples described above, DOC and total dissolved nitrogen (TDN) were measured with a Shimadzu TOC  $V_{CSH}$  with a total nitrogen attachment. Conductivity was measured with a YSI 556MPS handheld meter. Nutrients were measured using an AA3 fourchannel segmented flow autoanalyzer (SEAL Analytical). DON was calculated by subtracting the nitrate/nitrite and ammonia concentrations from the TDN. Water quality data from other sampling depths at Site D were provided by the USGS.

#### 141 *Carbamazepine*

142 Duplicate 1-L samples of collected groundwater were spiked with 5  $\mu$ L of 2  $\mu$ g/mL d<sub>10</sub>-143 carbmazepine for use as a surrogate and extracted following the protocols of ref.<sup>37</sup> using Empore 144 SDB-RPS disks (3M) that had been pre-conditioned with acetone, isopropyl alcohol, methanol, 145 and ultrapure water. The disks were stored at -20 °C until extraction. The disks were warmed to 146 room temperature and eluted with 10 mL of methanol and 10 mL of acetone. The combined 147 extract was blown down with nitrogen to a volume of ~5 mL and then reduced to near-dryness

148 via vacuum centrifugation. The extract was then reconstituted with 60 µL of methanol and 140

149  $\mu$ L of ultrapure water. Carbamzepine and d<sub>10</sub>-carbamzepine were quantified by ultrahigh

150 pressure liquid chromatography tandem mass spectrometry (Thermo TSQ Vantage) with the

151 electrospray ion source in positive mode according to previously described methods.<sup>37, 39</sup>

152 ESI-FT-ICR-MS

153 Samples (1-L) were extracted using Bond Elut PPL cartridges (Agilent) following the protocol of Dittmar et al.<sup>40</sup> The resulting methanol extracts were analyzed via direct infusion 154 with the electrospray ionization interface of a 7T LTQ FT-ICR-MS (LTQ FT-Ultra, Thermo 155 156 Scientific). The D2 sample was analyzed twice to test instrument reproducibility. The methanol 157 extracts were diluted with a solution of 70% methanol/30% water (2-fold dilution by volume for 158 Site D samples, 3-fold for Green, and 10-fold for Head of Waquoit and Sage Lot) to achieve a 159 stable spray and analyzed in positive mode, because nitrogen functional groups are generally 160 more amenable to detection under positive ionization. The resolving power was 400,000 at m/z161 400, and 200 transients were collected per sample. The transients were co-added and processed using the protocols described in Kido Soule et al.<sup>41</sup> 162

After correcting the m/z values of the features detected via ESI-FT-ICR-MS for the presence of sodium, a previously developed algorithm<sup>42, 43</sup> was used to assign elemental formulas with an allowed error of 1 ppm. The algorithm allows formulas including C, H, O, N, S, and P and allows inclusion of <sup>13</sup>C if the error is reduced by doing so. All data analysis and processing were performed in MATLAB R2013b (MathWorks, Inc.).

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#### 169 **Results and Discussion**

170 Dissolved organic carbon, dissolved nitrogen and carbamazepine

171 Sample collection depths, dissolved organic carbon, total dissolved nitrogen, and 172 carbamazepine data are shown in Table 1. The specific conductance values demonstrate that all 173 samples were collected from fresh water zones, rather than salt water intrusion where specific 174 conductance would be >  $30,000 \mu$ S/cm.

The DOC levels at Head of Waquoit and Sage Lot are higher than those at Green Pond and Site D. The lower DOC concentration for the D1 sample may be due to the deeper vadose zone at Site D, which would allow greater DOC mineralization during transport to the water table. At Site D, the DOC level decreases with depth, consistent with previous observations on Cape Cod,<sup>44</sup> and with a greater extent of degradation for organic matter that has traveled further or is older (see below).

181 The total dissolved nitrogen and nitrate+nitrite (Table 1) at Green Pond and Site D are 182 substantially higher than the levels at Head of Waquoit and Sage Lot, supporting the 183 expectations regarding the degree of anthropogenic impact at the two sites. The nitrate+nitrite is 184 likely derived from septic discharge and fertilizer application. The DON levels are lowest at the 185 two unimpacted sites (Head of Waquoit and Sage Lot), but at these sites, the DON comprises the 186 largest fraction of TDN (>65%). This indicates decreased processing of the organic nitrogen and less septic impact. At Site D, the DON levels are equal or higher than those at Head of Waquoit 187 188 or Sage Lot, but DON is only 4-8% of the total TDN. The elevated levels, however, clearly 189 indicate impact from septic systems. While the nitrate levels at Green Pond also indicate septic 190 influence, the calculated DON level is zero. This is in contrast to previous samples from 191 groundwater around Green Pond, which saw levels  $\sim 2 \text{ mg/L}$  as DON.<sup>3</sup> This does not mean, 192 however, that one would not expect to observe N-containing formulas via ESI-FT-ICR-MS in 193 our sample. The zero value for DON merely indicates that determining this value as the

difference between TDN and nutrients does not have the required precision and that additionalanalyses are needed to evaluate the role of septic systems as a source of organic nitrogen.

For the Head of Waquoit and Sage Lot groundwater samples, average carbamazepine concentrations were below the limit of quantification (1.6 ng/L). Carbamazepine was detected at concentrations ranging from 7.8 to 46.7 ng/L in the samples from Green Pond and Site D. These levels are consistent with those in various wastewater-impacted groundwaters<sup>25, 30, 32</sup> and a recent analysis of drinking waters drawn from the shallow aquifer on Cape Cod.<sup>28</sup>

201 It is clear that the samples collected from shallow depths (Green and D1) as well as the 202 deeper samples D2 and D3 are impacted by septic effluent based on the presence of 203 carbamazepine. Yet, the trend of values with depth at Site D merits discussion. Groundwater on Cape Cod flows in layers,<sup>45, 46</sup> with deeper waters travelling farther distances and being of 204 205 greater age. Thus, the water from the shallow depth near the water table (D1) is likely derived 206 from proximal sources and influenced by local septic systems. The water at the greatest depth (D3) is likely 10-20 years old<sup>44, 46, 47</sup> and has travelled from the recharge mound further north. 207 208 The presence of carbamazepine at this depth suggests that the water is from an area impacted by 209 septic systems or wastewater recharge and supports the notion that carbamazepine is persistent in 210 groundwater systems. The carbamazepine concentration at the intermediate depth is lowest. This 211 may indicate that the region in which this groundwater layer receives recharge is less 212 impacted/populated than the area providing recharge to the deeper groundwater.

213 Molecular characterization of the organic matter

For the seven samples analyzed via ESI-FT-ICR-MS (including the duplicate runs for sample D2), 12,992 features were detected and 11,434 formulas were assigned. Because of the small number of samples, cluster analysis was performed based on the features found in each

sample, and the dendrogram is shown in the Supplementary Information (Figure S1). The two
unimpacted sites, Head of Waquoit and Sage Lot, clustered together as did the replicate
injections for D2. Overall, the calculated differences were not large (all distances between 0.2
and 0.45), and the clustering was not used to guide further analyses.

221 The total formulas and the percentage of formulas in each sample containing only CHO 222 and CHON are given in Table 2. Also given are the percentages of all formulas found by the 223 algorithm that contained N in any allowed combination with other elements. In general, Head of 224 Waquoit and Sage Lot (the two unimpacted sites) had a higher percentage ( $\geq$ 56%) of CHO 225 formulas compared to the sites where carbamazepine was detected ( $\leq$ 56%). Using a 1-tail t-test 226 with equal variance (as determined by an F-test) to evaluate if the impacted samples have a lower 227 CHO level, gives a p-value of 0.115 (significant with 88% confidence). Similarly, the CHON 228 percentages ( $\leq 26\%$ ) and all N-containing formulas ( $\leq 40\%$ ) at the unimpacted sites are lower than 229 at the impacted sites (28-36% for CHON and 41-56% for all N). In this case, the t-test gives a p-230 value of 0.063 (significant with 93% confidence). Previous comparisons of effluent organic 231 matter from a wastewater treatment plant with Suwannee River natural organic matter analyzed 232 using negative mode ESI-FT-ICR-MS showed reduced abundance of CHO formulas in the wastewater samples.<sup>21</sup> The molar ratios of O:C and N:C (Table 2) also indicate that the Head of 233 234 Waquoit and Sage Lot samples are more oxidized (based on O:C ratio) and that the impacted 235 samples have higher nitrogen content. While these differences are subtle and more data would 236 be needed to derive more robust statistical certainty, these results suggest that septic-impacted 237 groundwater has a greater abundance of N-containing formulas.

The van Krevelen plots for all features and for only N-containing features are shown in
the Supplementary Information (Figures S2-S7). The four samples collected from shallow depths

have a larger number of points with O:C ratios > 0.5, with the Head of Waquoit and Sage Lot samples having more than the Green Pond and D1 samples. This is true of the plots for all of the detected features as well as those containing N, as quantified in Table 2. The exception is the Ncontaining compounds for the Head of Waquoit sample, but the count is biased by the fact that this sample has the fewest N-containing features. Overall, these results indicate a more reduced character for the septic-impacted samples.

Differences in the N-containing compounds are more readily assessed by identifying features that are common or unique among the samples. The samples were divided into three pairs for this analysis: Head of Waquoit + Sage Lot, Green + D1, and D2 + D3. In each case, the N-containing features present in both samples of one pair and not in both samples of the other pair (and vice versa) were determined. Also found were the features contained in all four of the compared samples.

252 In the van Krevelen diagrams shown in Figure 1a and 1b, the septic impacted samples are 253 compared to the unimpacted samples. It is apparent that the impacted samples have N-containing 254 features that are absent from the unimpacted samples. In comparing Green Pond and D1 with 255 Head of Waquoit and Sage Lot, the impacted samples contain features of H:C > 1.5 across the 256 O:C range that are not in the unimpacted samples (green points in Figure 1a). Similarly, D2 and 257 D3 contain features of H:C > 1.5 and O:C < 0.6 that are not in the unimpacted samples (red 258 points in Figure 1b). The features with H:C > 1.5 overlap with the regions of the diagram generally ascribed to be 'protein-like' and 'lipid-like',<sup>48</sup> and it may be expected that the lipid and 259 260 protein content of human waste would be different than that of natural systems. Nitrogen-261 containing compounds may also arise from household products/pharmaceuticals that are washed down the drain.<sup>49</sup> A previous study identified the sulfur-containing linear alkyl benzene 262

sulfonates in wastewater effluent using ESI-FT-ICR-MS, supporting the hypothesis that human use chemicals alter the composition of the dissolved organic matter.<sup>21</sup>

265 In Figure 1c, the differences between the shallow and deep samples impacted by septic 266 effluents are shown. The shallow samples have a cluster of features with higher O:C ratios (0.2-267 0.8, green points), and the deep samples (red points) have additional features with O:C < 0.3. 268 This could be consistent with reduction of the N-containing features in older, deeper samples in which dissolved oxygen has been consumed.<sup>48</sup> Also supporting the greater processing of N-269 270 containing compounds in the deep, impacted samples are the number of formulas in the 'protein-271 like' area of the van Krevelen diagrams for the individual samples (181 and 387 for D2 and D3, 272 respectively), compared to D1 (528). The reduced numbers in the deep samples may indicate 273 degradation of a portion of the septic-derived N-containing features in these samples.

274 Another means to visualize the differences in nitrogen content are carbon versus mass plots.<sup>50</sup> In such plots, the number of carbon atoms in a molecule is plotted versus the molecular 275 276 weight of the molecule, and the size of the point is proportional to the intensity of the ESI-FT-277 ICR-MS signal. Shown in Figure 2 are such plots for each sample. Note that the four impacted 278 samples have abundant (number of dots) and intense (larger size) features with N:C  $\ge$  0.5 from 279 carbon numbers 10 to 30 with m/z values from 400 to 700 (denoted by pink ovals). These 280 features are less intense and less numerous in the Head of Waquoit and Sage Lot samples, 281 indicating this pattern may be indicative of septic impacts. Interestingly, the four samples from 282 shallow depths all have a greater intensity of points in the upper right portion of the plot (carbon 283 number > 30, m/z > 600; black ovals) with low nitrogen content. The greatest intensity for these 284 points is in the two impacted samples (D1 and Green). While this is not an indicator of organic

nitrogen from the septic system effluent, the greater intensity in this region may be an indicatorof recent or less-processed septic input.

287 Both the van Krevelen diagrams and carbon versus mass plots show that it is possible to 288 identify features/molecular formulas of DON associated with septic-derived organic matter. 289 Elucidating molecular level composition is the first step to a predictive understanding of the 290 reactivity of the DON pool. Such features can be used to identify the relative contributions of 291 different sources of DON to the total nitrogen pool. This has the potential to allow the 292 assessment of the availability of DON in aquatic systems. With additional structural 293 characterization to identify the abundance of functional groups, it would also be possible to 294 evaluate impacts on water treatment (e.g., the formation of nitrogenous disinfection by products).

## 295 Conclusion

296 This work has provided molecular-level information regarding the DON in groundwaters 297 impacted by septic system effluent. Carbamazepine and nitrate/nitrite were found to be valid 298 indicators of septic influence, but depth, age, and recharge location of groundwater layers need 299 to be carefully considered when comparing these measurements. Overall, there appears to be a 300 greater abundance of N-containing features in DOM impacted by septic effluent, even if the 301 water is older and/or deeper. In van Krevelen space, the impacted samples had N-containing 302 features present at H:C > 1.5 that plotted in the 'lipid-like' and 'protein-like' areas of the 303 diagram. An abundance of features in this area of the diagram may be a useful marker of septic 304 impact in groundwater. The carbon versus mass plots also have abundant, intense features in a 305 specific region of the plot for the impacted samples. Thus, these plots may also be useful in 306 identifying the source of DON. Further work will be needed to identify features in van Krevelen 307 space and carbon versus mass plots that are indicators of other sources of DON in groundwater.

308 This work also demonstrates that unimpacted groundwater samples from the targeted watershed 309 are needed to allow comparisons to be performed to find features indicative of alteration of the 310 organic matter by human activities.

This work focused on abundance and intensity of N-containing features. Quantitative sample-to-sample comparison of intensities is limited using ESI-FT-ICR-MS due to issues associated with variations in ion suppression among samples. Thus, it is important to recognize that while the features detected in this work are a first step in using ESI-FT-ICR-MS to develop a molecular level understanding of DON, the overall impact of DON on aquatic systems will be a combination of abundance, concentration, and bioavailability of specific DON-moieties, and ESI-FT-ICR-MS analysis alone will not allow this assessment.

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327	Refer	ences
328	1.	W. S. Moore, Annu. Rev. Mar. Sci., 2010, 2, 59-88.
329	2.	A. E. Mulligan and M. A. Charette, J. Hydrol., 2006, 327, 411-425.
330	3.	K. D. Kroeger, M. L. Cole and I. Valiela, <i>Limnol. Oceanogr.</i> , 2006, <b>51</b> , 2248-2261.
331	4.	D. A. Bronk, Q. N. Roberts, M. P. Sanderson, E. A. Canuel, P. G. Hatcher, R. Mesfioui,
332		K. C. Filippino, M. R. Mulholland and N. G. Love, Environ. Sci. Technol., 2010, 44,
333		5830-5835.
334	5.	S. P. Seitzinger, R. W. Sanders and R. Styles, <i>Limnol. Oceanogr.</i> , 2002, 47, 353-366.
335	6.	M. Badruzzaman, J. Pinzon, J. Oppenheimer and J. G. Jacangelo, J. Environ. Manage.,
336		2012, <b>109</b> , 80-99.
337	7.	C. Meile, W. P. Porubsky, R. L. Walker and K. Payne, <i>Water Res.</i> , 2010, 44, 1399-1408.
338	8.	S. Zhao, P. Zhang, J. Crusius, K. D. Kroeger and J. F. Bratton, J. Environ. Monit., 2011,
339		<b>13</b> , 1337-1343.
340	9.	B. A. Pellerin, S. S. Kaushal and W. H. McDowell, <i>Ecosytems</i> , 2006, 9, 852-864.
341	10.	E. H. Stanley and J. T. Maxted, Ecol. Appl., 2008, 18, 1579-1590.
342	11.	S. S. Kaushal, P. M. Groffman, L. E. Band, E. M. Elliott, C. A. Shields and C. Kendall,
343		Environ. Sci. Technol., 2011, 45, 8225-8232.
344	12.	A. D. Shah and W. A. Mitch, Environ. Sci. Technol., 2011, 46, 119-131.
345	13.	E. B. Kujawinski, P. G. Hatcher and M. A. Freitas, Anal. Chem., 2002, 74, 413-419.
346	14.	C. E. Rostad and J. A. Leenheer, Anal. Chim. Acta, 2004, 523, 269-278.
347	15.	A. C. Stenson, A. G. Marshall and W. T. Cooper, Anal. Chem., 2003, 75, 1275-1284.
348	16.	B. P. Koch, M. Witt, R. Engbrodt, T. Dittmar and G. Kattner, Geochim. Cosmochim.
349		Acta, 2005, <b>69</b> , 3299-3308.
350	17.	K. Longnecker and E. B. Kujawinski, Geochim. Cosmochim. Acta, 2011, 75, 2752-2761.
351	18.	G. McKee and P. G. Hatcher, Geochim. Cosmochim. Acta, 2010, 74, 6436-6450.
352	19.	F. Schmidt, M. Elvert, B. P. Koch, M. Witt and KU. Hinrichs, Geochim. Cosmochim.
353		<i>Acta</i> , 2009, <b>73</b> , 3337-3358.
354	20.	F. Schmidt, B. P. Koch, M. Elvert, G. Schmidt, M. Witt and KU. Hinrichs, Environ. Sci.
355		<i>Technol.</i> , 2011, <b>45</b> , 5223-5229.
356	21.	M. Gonsior, M. Zwartjes, W. J. Cooper, W. Song, K. P. Ishida, L. Y. Tseng, M. K. Jeung,
357		D. Rosso, N. Hertkorn and P. Schmitt-Kopplin, Water Res., 2011, 45, 2943-2953.
358	22.	P. F. Greenwood, L. J. Berwick and JP. Croue, <i>Chemosphere</i> , 2012, <b>87</b> .
359	23.	J. Carrera, C. J. Ptacek, W. D. Robertson, D. W. Blowes, M. C. Moncur, E. Sverko and S.
360		Backus, <i>Environ. Sci. Technol.</i> , 2008, <b>42</b> , 2805-2811.
361	24.	E. Godfrey, W. W. Woesnner and M. J. Benoitti, <i>Ground Water</i> , 2007, <b>45</b> , 263-271.
362	25.	D. J. Lapworth, N. Baran, M. E. Stuart and R. S. Ward, <i>Environ. Poll.</i> , 2012, 163, 287-
363		303.
364	26.	W. D. Robertson, D. R. Van Stempvoort, D. K. Solomon, J. Homewood, S. J. Brown, J.
365		Spoelstra and S. L. Schiff, <i>J. Hydrol.</i> , 2013, <b>477</b> , 43-54.
366	27.	C. H. Swartz, S. Reddy, M. J. Benoitti, H. Yin, L. B. Barber, B. J. Brownawell and R. A.
367		Rudel, <i>Environ. Sci. Technol.</i> , 2006, <b>40</b> , 4894-4902.
368	28.	L. A. Schalder, R. A. Rudel, J. M. Ackerman, S. C. Dunagan and J. G. Brody, <i>Sci. Total</i>
369	• •	<i>Environ.</i> , 2014, <b>268-269</b> , 384-393.
370	29.	G. Gasser, M. Rona, A. Voloshenko, R. Shelkov, O. Lev, s. Elhanany, F. T. Lange, M.
371		Scheure and I. Pankratov, Desalination, 2011, 273, 398-404.

- 372 A. Jurado, R. Lopez-Serna, E. Vazquez-Sune, J. Carrera, E. Pujades, M. Petrovic and D. 30. 373 Barcelo, Chemosphere, 2014, in press. 374 http://dx.doi.org/10.1016/j.chemosphere.2014.01.014. 375 31. M. E. Stuart, D. J. Lapworth, J. Thomas and L. Edwards, Sci. Total Environ., 2014, 468-376 **469**, 564-577. 377 D. R. Van Stempvoort, J. W. Roy, J. Grabuski, S. J. Brown, G. Bickerton and E. Sverko, 32. Sci. Total Environ., 2013, 461-462, 348-359. 378 379 J. Teerlink, V. Martinez-Hernandez, C. P. Higgins and J. E. Drewes, Water Res., 2012, 33. 380 **46**, 5174-5184. 381 Massachusetts Executive Office of Environmental Affairs, Cape Cod Watershed 34. 382 Assessment and 5-Year Action Plan, Boston, MA, 2004. 383 J. R. Barbaro, D. A. Walter and D. R. LeBlanc, U.S. Geological Survey Scientific 35. 384 Investigations Report 2013–5061, 2013, 37 p. http://pubs.usgs.gov/sir/2013/5061/. K. D. Kroeger and M. A. Charette, Limnol. Oceanogr., 2008, 53, 1025-1039. 385 36. 386 37. D. R. Griffith, Joint Program in Oceanography/Applied Ocean Science and Engineering 387 and Department of Civil and Environmental Engineering, Massachusetts Institute of 388 Technology/Woods Hole Oceangraphic Institution, Ph.D. Thesis. Available at: 389 http://dspace.mit.edu/handle/1721.1/85824, 2013, [Accessed May 20, 2014]. 390 M. A. Charette and M. C. Allen, Ground Water Monit. Remed., 2006, 26, 87-93. 38. 391 39. X.-S. Miao and C. S. Metcalfe, Anal. Chem., 2003, 75, 3731-3738. 392 T. Dittmar, B. Koch, N. Hertkorn and G. Kattner, Limnol. Oceanogr.: Methods, 2008, 6, 40. 393 230-235. 394 M. C. Kido Soule, K. Longnecker, S. J. Giovannoni and E. B. Kujawinski, Org. 41. 395 Geochem., 2010, 41, 725-733. 396 E. B. Kujawinski and M. D. Behn, Anal. Chem., 2006, 78 4363-4373. 42. 397 43. E. B. Kujawinski, K. Longnecker, N. V. Blough, R. Del Vecchio, L. Finlay, J. B. Kitner and S. J. Giovannoni, Geochim. Cosmochim. Acta, 2009, 73, 4384–4399. 398 399 44. W. J. Pabich, I. Valiela and H. F. Hemond, *Biogeochemistry*, 2001, 55, 247-268. 400 T. C. Cambareri and E. M. Eichner, Ground Water, 1998, 36, 626-634. 45. 401 O. Portniaguine and D. K. Solomon, Water Resour. Res., 1998, 34, 637-645. 46. 402 D. K. Solomon, R. J. Poreda, P. G. Cook and A. Hunt, Ground Water, 1995, 33, 988-996. 47. 403 S. Kim, R. W. Kramer and P. G. Hatcher, Anal. Chem., 2003, 75, 5336-5344. 48. 404 49. X. Li and B. J. Brownawell, Environ. Sci. Technol., 2010, 44, 7561-7568. 405 50. T. Reemtsma, A. These, M. Linscheid, J. Leenheer and A. Spitzy, *Environ. Sci. Technol.*, 406 2008, 42, 1430-1437. 407 408
  - 409

412 Table 1. Specific conductance, nutrient, dissolved organic carbon, and carbamazepine levels in the sampled groundwaters.

Site <sup>a</sup>	Depth below surface (m)	SC (µS/cm)	DOC (mg C/L)	[NO <sub>3</sub> <sup>-+</sup> NO <sub>2</sub> <sup>-</sup> ] (mg/L as N)	$[NH_4^+]$ (mg/L as N)	TDN (mg/L as N)	DON (mg/L as N)	Carbamazepine (ng/L; average of n = 2)
Head of Waquoit	1.17	680	4.6	0.05	0.03	0.3	0.3	<loq<sup>e</loq<sup>
Sage Lot	$2 - 4^{b}$	-	8.6	0.01	0.03	0.2	0.2	<loq< td=""></loq<>
Green	0.86	531	1.7	3.68	0.02	3.7	0	12.1
Site D1	6.30 <sup>c</sup>	171	1.8	7.33	0.14	7.8	0.3	46.7
	7.53	181	1.1	1.67	0.57	2.4	0.11	
	8.75	203	1.2	2.34	0.69	3.2	0.17	
Site D2	9.97	236	0.4	12.09	0.51	13.7	1.1	7.8
	11.19	225	0.3	15.7	< 0.02	14.9	_ d	
	12.41	209	0.5	12.9	< 0.02	12.3	_ <sup>d</sup>	
	13.66	219	0.5	14.5	< 0.02	14.9	0.4	
	14.88	255	0.4	19.4	< 0.02	18.7	_ d	
Site D3	16.11	321	0.5	24.39	0.02	26.3	1.9	30.6
	17.33	1081	0.6	23.1	< 0.02	16.1	_ d	

<sup>a</sup>For unlabeled rows at Site D, only water quality data were collected by the USGS (provided courtesy of Denis LeBlanc and John Colman), and no quantification of

carbamazepine or analysis by ESI-FT-ICR-MS was performed.

<sup>b</sup>Exact depth between these depths. Water table at  $\sim 2$  m below ground surface.

<sup>c</sup>The interval at 5.1 m was dry, indicating the water table is between 5.1 and 6.3 m in depth

<sup>d</sup>Subtracting the inorganic components from total nitrogen results in a negative value.

413 414 415 416 417 418 <sup>e</sup>One replicate was above the LOQ of 1.6 ng/L and one below, with the average value below the LOQ.

0:4-	Total Formulas	%CHO	% CHON	% containing N <sup>b</sup>	(O:C) <sub>wa</sub>	(N:C) <sub>wa</sub>	O:C > 0.5	O:C > 0.5
Site							all formulas	N-containing formulas
Head of Waquoit	5,667	61	25	37	0.34	0.07	1193	278
Sage Lot	6,638	56	26	40	0.35	0.07	1337	522
Green	5,731	54	28	43	0.30	0.09	947	374
Site D1	5,803	56	29	41	0.30	0.07	661	375
Site D2 <sup>a</sup>	4,413	41	36	56	0.20	0.17	425	177
Site D3	5,368	53	30	44	0.27	0.09	505	262

**Table 2.** The total number of formulas and percentages (by number) of the types of formulas assigned to the groundwater samples, weighted averages of O:C and N:C molar ratios, and number of formulas with O:C ratio > 0.5. 

<sup>a</sup>Average of duplicate injections. <sup>b</sup>Includes all possible formulas found by the algorithm that contain N in any allowed combination with C,H,O,S, and/or P.



- 429 Figure 1. Comparisons of N-containing features in selected samples. (a) The green symbols are
- 430 features present in the shallow, impacted samples (Green and D1) but not in the unimpacted sites
- 431 (Head of Waquoit and Sage Lot). The blue are the features in Head of Waquoit and Sage Lot, but
- 432 not the other two samples. Black symbols are present in all four samples. (b) The blue symbols
- 433 are features present in the shallow, unimpacted samples (Head of Waquoit and Sage Lot) but not
- in D2 and D3. The red are the features in D2 and D3, but not the other two samples. Black
- 435 symbols are present in all four samples. (c) The green symbols are features present in the
- 436 shallow, impacted samples (Green and D1) but not in D2 and D3. The red are the features in D2
- 437 and D3, but not the other two samples. Black symbols are present in all four samples
- 438



Figure 2. Carbon versus mass plots for the nitrogen containing features detected via ESI-FT-ICR-MS in the groundwater samples.
 Larger diameter denotes greater intensity and color indicates the N:C ratio. The pink oval marks the region of the plot that is

442 comprised of high nitrogen ratio features that are more intense in the septic -impacted samples. The black oval indicates low nitrogen
 443 features that are more abundant/intense in shallow water samples.

# **Supplementary Information for**

# Molecular signature of organic nitrogen in septic-impacted groundwater

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Dendrogram from Cluster Analysis (Figure S1)	p. S2
Van Krevelen diagrams (Figures S2-S7)	p. S3



**Figure S1.** Dendrogram derived from cluster analysis determined from the FTICR-MS data from the six samples. The sample for Site D2 was analyzed twice, and the close association of the replicates indicates limited instrument variability.

# Van Krevelen diagrams

In the following van Krevelen diagrams, the left hand plot contains all of the identified features for which formulas were assigned. The plot on the right hand side contains only N-containing features. The rectangles indicate regions associated with condensed hydrocarbons (red), lignin (dark blue), lipids (green), proteins (pink), and carbohydrates (light blue), as shown below.





Figure S2. Van Krevelen diagrams for the Waquoit ground water samples.



Figure S3. Van Krevelen diagrams for the Sage Lot ground water samples.



Figure S4. Van Krevelen diagrams for the Green Pond ground water samples.



Figure S5. Van Krevelen diagrams for the Site D1 ground water samples.



Figure S6. Van Krevelen diagrams for the Site D2 ground water samples.



Figure S7. Van Krevelen diagrams for the Site D3 ground water samples.