1	Comparison of CO₂ Dynamics and Air-Sea Exchange in Differing Tropical Boof Environments
2	Keel Environments
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11 12 13 14 15 16 17 18 19 20 21 22 23 24	Abstract An array of MAPCO ₂ buoys, CRIMP-2, Ala Wai, and Kilo Nalu, deployed in the coastal waters of Hawaii have produced multiyear high temporal resolution CO ₂ records in three different coral reef environments off the island of Oahu, Hawaii. This study, which includes data from June 2008-December 2011, is part of an integrated effort to understand the factors that influence the dynamics of CO ₂ -carbonic acid system parameters in waters surrounding Pacific high island coral reef ecosystems and subject to differing natural and anthropogenic stresses. The MAPCO ₂ buoys are located on the Kaneohe Bay backreef, and fringing reef sites on the south shore of O'ahu, Hawai'i. The buoys measure CO ₂ and O ₂ in seawater and in the atmosphere at 3-hour intervals, as well as other physical and biogeochemical parameters (CTD, chlorophyll-a, turbidity). The buoy records, combined with data from synoptic spatial sampling, have allowed us to examine the interplay between biological cycles of productivity/respiration and calcification/dissolution and biogeochemical and physical forcings on hourly to inter-annual time scales.
25 26 27 28 29 30 31 32 33 34 35 36	Air-sea CO ₂ gas exchange was also calculated to determine if the locations were sources or sinks of CO ₂ over seasonal, annual, and interannual time periods. Net annualized fluxes for CRIMP-2, Ala Wai, and Kilo Nalu over the entire study period were 1.15 mol C m ⁻² yr ⁻¹ , 0.045 mol C m ⁻² yr ⁻¹ , and -0.0056 mol C m ⁻² yr ⁻¹ , respectively, where positive values indicate a source or a CO ₂ flux from the water to the atmosphere, and negative values indicate a sink or flux of CO ₂ from the atmosphere into the water. These values are of similar magnitude to previous estimates in Kaneohe Bay as well as those reported from other tropical reef environments. Total alkalinity (A _T) was measured in conjunction with pCO2 and the carbonic acid system was calculated to compare with other reef systems and open ocean values around Hawaii. These findings emphasize the need for high-resolution data of multiple parameters when attempting to characterize the carbonic-acid system in locations of highly variable physical, chemical, and biological parameters (e.g. coastal systems, reefs).
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1 Introduction

Atmospheric concentrations of carbon dioxide (CO₂) have been on the rise for the past 200 years due to human activities releasing more than 400 petagrams of carbon into the atmosphere since 1750. Approximately half of these emissions occurred during the past 40 years (CDIAC cdiac.ornl.gov/). Atmospheric concentrations of ~395 µatm, as observed at Mauna Loa in 2012 (http://www.esrl.noaa.gov/gmd/ccgg/trends/), are significantly higher than those at any time in the past 800,000 years (Petit et al., 1990; Augustin et al., 2004; Siegenthaler et al., 2005). Although the atmospheric record is now well characterized, much work remains to be done in the oceanic realm especially on the strength of the CO₂ exchange flux and its future.

The vast majority of research on the effect of the rise of atmospheric CO_2 on the oceans has been focused on the open ocean. (Andersson et al., 2007; Feely et al., 2004; Feely et al., 2008; Orr et al., 2005; Takahashi et al., 2009; Wanninkhof et al., 2012). The open ocean, in general, is regarded as a sink for atmospheric CO₂, meaning that the net flux of CO₂ presently is into the ocean. However, the global coastal ocean is quite variable with respect to the net flux of CO_2 into or out of the atmosphere. Values of coastal net CO_2 flux (mol C m⁻² yr⁻¹) range from 438 and 3330 in some European estuaries (Frankignoulle et al., 1998) to -34.6 and -33.6 in both the Gulf of Biscay and the East China Sea (Frankignoulle and Borges, 2001; Wang et al., 2000) and as high as -87.6 (Hales et al., 2003) in a highly productive upwelling region off the Oregon coast. Massaro et al. (2012) reported a net flux of 1.80 mol C m⁻² yr⁻¹ in the southern sector of Kaneohe Bay. Positive values indicate a flux of CO₂ from the ocean into the atmosphere (source) and negative values indicate a flux into the ocean (sink). However, many of these flux estimates were obtained through periodic synoptic sampling of the surface waters of these systems and very few long-term, high resolution, *in-situ* studies on the CO₂-carbonic acid system and CO₂ exchange dynamics have been performed in coastal environments (e.g. De Carlo et al., this volume; Drupp et al., 2011; Massaro et al., 2012).

As atmospheric CO_2 levels continue to rise for (at least) the remainder of the 21^{st} century, the global shallow ocean environment, which has been a source of CO_2 to the atmosphere for at least the past 300 years, may reverse its net flux to that of a sink for atmospheric CO₂ just like
the open ocean (Andersson and Mackenzie, 2004; Andersson et al., 2006; Mackenzie et al.,
2011; Wanninkhof et al., 2012).

2 Study Rationale

This study expands upon a 2.5 year *in-situ* high-resolution time series study that was carried out in southern Kaneohe Bay, Oahu, Hawaii (Figure 1a and 1b) through the use of the Coral Reef Instrumented Monitoring and CO₂ Platform (CRIMP-CO₂) buoy (Drupp et al. 2011; Massaro et al., 2012). Kaneohe Bay, in particular its southern sector, has been extensively studied for the past 40 years (e.g., Roy, 1970; Cox et al., 1973; Hollett, 1977; Smith et al., 1981; Taguchi and Laws, 1987, 1989; Jokiel et al., 1993; Hunter and Evans, 1995; Laws and Allen, 1996; Atkinson, 2000; Hearn and Atkinson, 2000; Kinzie et al., 2001; Falter et al., 2004; Ringuet and Mackenzie, 2005; Tanaka and Mackenzie, 2005; Hoover et al., 2006; De Carlo et al., 2007; Fagan and Mackenzie, 2007; Hoover and Mackenzie, 2009; Drupp et al. 2011; Massaro et al., 2012; Shamberger et al., 2011). The bay is thought to be representative of other high-island tropical reef environments found throughout the Pacific Ocean.

The Coral Reef Instrumented and CO_2 Monitoring (CRIMP-CO₂) program, initiated in late 2005, was the first long-term, high resolution CO₂ monitoring program conducted in tropical coastal waters. The CRIMP-CO₂ buoy also served as a platform for deploying additional water quality sondes measuring a variety of standard physical and biogeochemical parameters. In June 2008, the CRIMP buoy was relocated to its current location (CRIMP-2, Figure 1b) at a depth of 3 m on the inside edge of the Kaneohe Bay barrier reef, in the central sector of Kaneohe Bay. Two additional buoys, Water Quality Buoys Ala Wai and Kilo Nalu (WQB-AW and WQB-KN, respectively, Figure 1c) were also deployed at that time, on the south shore of Oahu in ~ 12 m water depth on sand patches adjacent to fringing reefs in Mamala Bay, approximately 200 m offshore of urban Honolulu. These three buoys provide high frequency $(3 \text{ hr}) \text{ CO}_2$ and ancillary water quality parameters of conditions in differing coastal coral reef environments, and complementary data are obtained through periodic synoptic sampling.

Figure 1

Periodic synoptic sampling, often performed during the day under favorable weather conditions (e.g. De Carlo et al., this volume), misses diel cycles and, frequently, changes induced by local rainfall and storm events. The latter have been shown to induce rapid and brief changes in stream flow (e.g., Tomlinson and De Carlo, 2003; Hoover and Mackenzie, 2009) and have a significant effect on short-term CO₂ variability and general biogeochemistry (e.g., nutrient concentrations) in the nearshore semi-estuarine environment of Kaneohe Bay (Ringuet and Mackenzie, 2005; De Carlo et al., 2007; Fagan and Mackenzie, 2007; Hoover and Mackenzie, 2009; Drupp et al., 2011; Massaro et al., 2012). Strong diel signals, due to water column and reef productivity/respiration and calcification/dissolution processes, have been observed by both Drupp et al. (2011) and Shamberger et al. (2011) in Kaneohe Bay. Seasonal and annual trends in water column pCO₂ were clearly evident in previous work by Fagan and Mackenzie (2007), Drupp et al. (2011), and Massaro et al. (2012). Based on these studies, it is evident that only a monitoring program that is both long-term (multiple years) and high resolution (multiple samples per day) can adequately characterize CO_2 exchange fluxes and processes controlling them in dynamic biogeochemical environments such as Kaneohe Bay and other tropical reef environments.

3 Methods

3.1 Environmental Setting

Kaneohe Bay, the location of the CRIMP-2 buoy, is the largest semi-enclosed body of water on the island of Oahu, Hawaii (Figure 1) and is located on the northeast (windward) coast. It is host to numerous fringing and patch reefs as well as a large barrier reef. The bay has a mean depth of 9.5 meters, with the barrier reef averaging ~2 meters depth (Bathen, 1968). Northeasterly tradewinds blow the majority (~80%) of the year and their speed is a major factor in governing the magnitude of air-sea CO₂ gas exchange fluxes (Massaro et al., 2012). Water residence time in the bay ranges from hours on the barrier reef to 30+ days in the more hydrologically isolated southern sector (Lowe et al., 2009a), where the original CRIMP buoy was deployed.

128 CRIMP-2 was deployed in 3 m of water over sandy sediment on the inside edge of the
129 barrier reef (Figure 1b) in the central sector of Kaneohe Bay. The site was chosen due to its

proximity to the barrier reef (~20 m distance from the backreef edge) and because the flow of water at this site is nearly unidirectional, thereby allowing measurements of water chemistry and its CO₂ content after the water flows across the barrier reef from the open ocean. Effectively, the water chemistry at this site represents an end-member following the alteration of its chemistry due to biological and physical processes during its transit across the barrier reef. It is important to note that much of the reef may not have been exposed to the same water chemistry that is measured at this buoy, as the water has been undergoing biogeochemical alteration continuously during its transit across the reef. For example, data from Shamberger et al., 2011 (discussed later) collected from a mid-reef location falls in between the open ocean values and our CRIMP-2 end-member. The CRIMP-2 location is also generally isolated from direct terrestrial and riverine inputs from the Kaneohe Bay watershed.

Recent work has shown that the biogeochemistry of the CRIMP-2 location is driven largely by benthic productivity and calcification occurring on the barrier reef (Shamberger et al., 2011). This is in sharp contrast to the original CRIMP location, where land runoff combined with the physical characteristics of the site modified considerably the biogeochemistry of the water and CO₂ exchange dynamics on various time scales (Ringuet and Mackenzie, 2005; De Carlo et al. 2007; Drupp et al., 2011; Massaro et al., 2012). At the CRIMP-2 location, runoff has very little effect on the magnitude of the air-sea flux of CO₂ and the location also can be considered a relatively healthy coral reef environment in which to study the CO₂-carbonic acid system and CO₂ exchange.

Fringing reefs in Mamala Bay on the south shore of Oahu are rather unprotected. These reefs are generally subject to the direct physical effects of tides, winds, and open ocean wave action, and nearshore water exchanges readily with surrounding open ocean water. The two sites (Ala Wai and Kilo Nalu) differ from each other in the presence or absence, respectively, of riverine input from the Ala Wai Canal. The canal drains approximately 1/3 of urban Honolulu (population ~850,000) and contributes significant amounts of fluvial discharge containing high nutrient and suspended sediment loads to coastal waters. The effects of this discharge are especially evident at the Ala Wai buoy following (large) storm events (Tomlinson et al., 2011). Occasionally, depending on wind and swell conditions, this riverine signal can reach the Kilo Nalu buoy, about 1.6 km to the west, although the impacts of the fluvial discharge on water

quality at that site are typically highly attenuated. The substrate in Mamala Bay out to the 30m
isobaths consists mainly of sand, limestone pavement, and spur and groove coral reef.
Encrusting algae and coral dominates the reef with some *Porites lobata* and *Pocillopora meandrina*. Macroalgae have 10-90% local coverage of the pavement (Battista et al., 2007,
Tomlinson et al., 2011).

3.2 In-situ Measurements

The buoys are equipped to make a broad range of measurements. A LICOR-820 infrared sensor and a MaxtexMAXTM-250 Series sensor measuring CO₂ and O₂, respectively, are located within the buoy body, as well as a Sensirion humidity sensor. Data are collected every 3 hours and transmitted daily via Iridium satellite to NOAA/PMEL (Pacific Marine Environmental Laboratory). The accuracy of the LICOR-820 is conservatively estimated to be 2.5% of the measured value. Details regarding the analytical scheme of the buoy can be found in Massaro et al. (2012) and Shamberger et al. (2011). CO₂ and O₂ data from all three buoys can be viewed in "near real time" at http://www.pmel.noaa.gov/co2/coastal/HI/

Both WQB-AW and KN buoys are also equipped with Sea-Bird Electronics SeaCat16 v2plus CTD's (Conductivity, Temperature, Depth) and WET Labs FLNTUS ECO fluorometer/turbidity sensors. Temperature, conductivity (salinity), dissolved oxygen, chlorophyll, and turbidity are measured at 20 minute intervals. SBE16 data from WQB-AW and WQB-KN are transmitted every 20 minutes via a Satlantic STOR-X data logger and cell phone 40 178 modem. A Yellow Springs Instrument (YSI) 6600 Multi-Parameter sonde and a Sea-Bird Electronics MicroCat CT (37-SMP) are attached to the CRIMP-2 buoy and measure temperature, conductivity, dissolved oxygen, chlorophyll, turbidity, and pH. Water quality data from the SBE16's can be viewed in real time on the Pacific Islands Ocean Observing System (PacIOOS) website.

184 <u>http://oos.soest.hawaii.edu/pacioos/data_product/WQ/</u>

Wind and tidal data were obtained from the NOAA National Ocean Service (NOS)
Station OOUH1 in downtown Honolulu (for south shore buoys) and from the Hawaii Institute of
Marine Biology (HIMB) weather station on Coconut Island (for CRIMP-2). Rainfall data for the
windward coast were obtained from gauges at the Hawaii Institute of Marine Biology (HIMB) on

Coconut Island in southern Kaneohe Bay and the National Weather Service (NWS) HI15 Luluku gauge in the Kaneohe Stream watershed. South shore rainfall was measured at the NWS HI-18 Manoa Lyon Arboretum and NWS HI-20 Nuuanu Valley gauges (in the upper watersheds which drain much of Honolulu) and from NWS HI-26 Aloha Tower within Honolulu harbor.

3.3 Sample Analysis

Discrete water samples were collected at each location, during periods of routine equipment maintenance, usually every 4 weeks. Samples for total alkalinity (A_T) were collected from the surface waters in 300 mL borosilicate bottles. Each sample was fixed with 200 µL of saturated HgCl₂ to inhibit any biological activity. The bottles were sealed with Apiezon-greased ground glass stoppers. Total alkalinity was determined using the open cell potentiometric titration method (Dickson et al., 2007) on a Metrohm Titrando 905. The accuracy of the A_T determinations was evaluated using certified reference materials (CRM) from the Scripps Institution of Oceanography (Dickson et al., 2001; Dickson et al., 2003; Dickson et al., 2007) and found to be 1.37 μ mol kg⁻¹ (n=26). The precision between replicates was 1.52 μ mol kg⁻¹ (n=63).

3.4 Calculations

Concentrations of CO₂ measured by the LICOR-820 were corrected using the relative humidity in the equilibrator to calculate a dry xCO₂ (mole fraction at 0% humidity). Using the methods from Zeebe and Wolf-Gladrow (2001), originally developed by Weiss and Price (1980). pCO₂ was calculated utilizing ambient water vapor pressure, dependent on T and S, and assuming that the air directly above the sea surface was 100% saturated.

Calculations of air-sea fluxes were made using Equation (1) below:

$F = k\alpha \Delta p CO_2$, (1)

where k is the CO₂ gas transfer velocity, α is the solubility of CO₂ in seawater at the specified temperature and salinity, and ΔpCO_2 is the difference between atmospheric and seawater pCO₂ concentrations (Weiss, 1974; Liss, 1983; Wanninkhof, 1992). The gas transfer velocity, k, was calculated using the Ho et al. (2006) parameterization shown in equation (2):

$k_{(600)} = (0.266 \pm 0.019) (U_{10})^2$ (2)

where U_{10} is the wind speed measured at (or corrected to) ten meters above sea level. Error bars for fluxes were calculated by error propagation, using a 2.5% error for the CO₂ measurement and an uncertainty of 0.019 in the Ho et al. parameterization.

Normalization of pCO₂ to an average seawater temperature was performed according to Takahashi et al. (1993, 2002) assuming a 4.23% change in pCO₂ per 1°C change in temperature (equation 3). The effect of only temperature on pCO_2 at a given time was also calculated (equation 4) from Takahashi et al. (1993 and 2002) using the mean pCO_2 of the study period. This represents the pCO₂ changes induced by temperature fluctuations under isochemical conditions.

 pCO_2 at $T_{mean} = pCO_{2obs} * exp(0.0423*(T_{mean}-T_{-obs}))$ (3) $pCO_2 \text{ at } T_{obs} = pCO_{2mean} * exp(0.0423*(T_{obs}-T_{mean})),$ (4)

where T_{obs} and pCO_{2obs} are the measured *in-situ* temperature and pCO₂, and T_{mean} and pCO_{2mean} are the annual mean values of temperature and pCO_2 .

Dissolved inorganic carbon (C_T), pH (total scale), and calcium carbonate saturation state (Ω) were calculated using CO2SYS (Lewis and Wallace, 1998) with carbonic acid constants by Mehrbach et al. (1973) refit by Dickson and Millero (1987).

Nutrient analyses were performed on a Seal Analytical AA3 HR Nutrient Autoanalyzer. Nitrate (NO_3) and nitrite (NO_2) were analyzed according to the methods of Armstrong et al. (1967) and Grasshoff et al. (1983) with a detection limit of 0.006 μ mol L⁻¹ and a relative standard deviation (CV) of 0.3%. Silicate (Si(OH₄)) was analyzed based on the methods of Grasshoff et al. (1983) with a detection limit of 0.350 μ mol L⁻¹ and a CV of 0.5%. Soluble reactive phosphate (PO4³⁻) was analyzed according to the colorimetric method of Murphy and Riley (1962) with a detection limit of 0.018 μ mol L⁻¹ and a CV of 0.4%.

4 Results

Figure 2a shows the pCO₂ of seawater at all three buoy locations over the entire study period of June 2008-December 2011. A seasonal trend is evident in all three data sets; however,

CRIMP-2 (blue) shows much greater short term variations in pCO₂. The solid yellow line in this figure indicates 377 μ atm, which was the mean air pCO₂ during the study period. When seawater pCO_2 is above this line, the location is a source of CO_2 to the atmosphere and when seawater pCO_2 is below the line, the location is a sink for CO_2 from the atmosphere. However, source or sink activity is dependent upon the instantaneous flux, which was calculated from the *in-situ* measured atmospheric pCO_2 at the time of the seawater measurement. The mean seawater pCO₂ values at CRIMP-2, Kilo Nalu, and Ala Wai were $439 \pm 11 \mu atm$, $379 \pm 10 \mu atm$, and 389 ± 10 µatm, respectively. Minimum and maximum values are reported in Table 1. CRIMP-2 displayed the largest variation with a range of 780 µatm. The standard deviation of the mean was 83.4 µatm at CRIMP-2 compared to values of 18.2 and 25.3 µatm for KN and AW, respectively, further demonstrates the greater short-term variations which occur at the CRIMP-2 location.

Figure 2b displays the pCO₂ normalized to average T during the entire study period, as calculated by Equation 3. Even when normalized to the mean temperature of 25° C, pCO₂ still exhibits large short term fluctuations at all three buoys. The T-normalized pCO₂ also displays a seasonal trend (albeit much less than in the raw pCO₂ data shown in Figure 2a) which can be difficult to discern due to the short term variations.

260 Figure 2

Table 1

Monthly mean pCO₂ was calculated at each location and is shown in Figure 3a. The trend of higher pCO₂ in the summer and lower pCO₂ in the winter was also observed at the original CRIMP location (Drupp et al., 2011; Massaro et al., 2012). Figure 3b shows the weekly temperatures throughout the study period. All three locations follow the same trend of maximum temperatures (~27 °C) during the summer and minimum temperatures (~23-24 °C) in the winter. Water at CRIMP-2 does become slightly cooler than water at AW and KN during the winter months, but does not become warmer than water at AW and KN during the summer.

269 Figure 3

The daily range of pCO_2 was also calculated for each day throughout the study period. Table 2 shows statistics for the daily ranges at all three buoys, as well as the original CRIMP buoy. CRIMP-2 exhibited a mean daily range of 196 \pm 16 µatm, four times greater than observed at the Ala Wai location. The maximum daily ranges at AW and KN are considerably lower than the mean daily range at CRIMP-2. At each location, pCO_2 dropped throughout the day reaching a minimum value in the afternoon, around $\sim 14:00$ hrs. Maximum daily pCO₂ values occurred late at night to the early morning (\sim 4:00-8:00 hrs). The pCO₂ varied in the opposite direction of oxygen concentrations, which peaked in the mid-afternoon and reached minimum concentrations during the night. Pearson correlations between pCO_2 and pO_2 at each buoy are shown in Table 3. An anti-correlation of -0.75 was observed at CRIMP-2: negative, albeit weak, correlations were also observed at Ala Wai and Kilo Nalu. An example of the diel anti-correlation cycle is shown in Figure 4, which displays pCO₂ and pO₂ at CRIMP-2 from April 22 to May 12, 2010.

- *Table 2*
- *Table 3*
- 285 Figure 4

Autocorrelation was also evaluated for each buoy data set to determine the periodicity of pCO₂. This value indicates the correlation between all points separated by specific time intervals in the same data set. A strong autocorrelation was found at a period of 24 hours for each location. An autocorrelation plot for CRIMP-2 is shown in Figure 5. This correlation stayed relatively strong for 48 and 72 hours but dropped off (<0.5) over greater multiples of 24 hours. The correlation of 0.74 at 24 hrs observed at CRIMP-2 was the highest of the three locations during the study period (see Table 3). A correlation greater than 0.50 existed out to 72 hours (24 hr intervals), meaning that even points 72 hours (3 days) apart had a correlation value of greater than 0.50. A much lower (and sometimes negative) correlation occurred between points 12 hours apart.

Figure 5

 Instantaneous CO₂ fluxes at the three buoy sites are presented in Figure 6 in units of mmol C m⁻² hr⁻¹ during the entire study period of June 2008 to December 2011. Statistics for these fluxes are shown in Table 4. As expected from the larger range of pCO₂ values, CRIMP-2 displayed the largest range of flux and had a mean instantaneous flux of 0.13 \pm 0.030 mmol C m⁻² hr⁻¹. The mean flux at Kilo Nalu was 0 and 0.005 \pm 0.005 at Ala Wai. Percentage of time spent as a source and a sink was also computed for the three sites. The CRIMP-2 and Ala Wai water columns were CO₂ sources to the atmosphere 78% and 66% of the study period, respectively. The Kilo Nalu location was a source 51% of the time.

Net annualized fluxes for all three locations are also shown in Table 4 in units of mol $C/m^2/yr$. CRIMP-2 was a source of $1.15 \pm 0.27 \text{ mol C} \text{ m}^{-2} \text{ yr}^{-1}$, while Ala Wai and Kilo Nalu were essentially neutral at $0.045 \pm 0.048 \text{ mol C} \text{ m}^{-2} \text{ yr}^{-1}$ and $-0.0050 \pm 0.072 \text{ mol C} \text{ m}^{-2} \text{ yr}^{-1}$, respectively. For comparison, CRIMP displayed an annualized flux of $1.76 \pm 0.37 \text{ mol C} \text{ m}^{-2} \text{ yr}^{-1}$ for the time period of December 2005 to June 2008 (Massaro et al., 2012).

Table 5 shows net annualized fluxes (in mol C $m^{-2} yr^{-1}$) for each year (2008-2011) of the study period as well as for each season of the study period. Seasons are separated as summer dry periods from May through October and the winter wet periods from November through April. The water at CRIMP-2 was a source of CO_2 to the atmosphere every year of the study period $(1.39 \pm 0.25 \text{ mol C m}^{-2} \text{ yr}^{-1}, 0.82 \pm 0.20 \text{ mol C m}^{-2} \text{ yr}^{-1}, 1.10 \pm 0.28 \text{ mol C m}^{-2} \text{ yr}^{-1}, \text{ and } 1.31 \pm 0.21 \text{ mol C m}^{-2} \text{ yr}^{-1}$ $0.36 \text{ mol C} \text{m}^{-2} \text{yr}^{-1}$). The water at Ala Wai was initially a source of CO₂ in 2008 and 2009, switched to a sink of CO₂ (albeit a small sink) in 2010 of -0.003 \pm 0.013 mol C m⁻² yr⁻¹ and back to a source of 0.092 mol C m⁻² yr⁻¹ in 2011. Initially, in 2008, the Kilo Nalu environment was a source $(0.073 \pm 0.081 \text{ mol C m}^{-2} \text{ yr}^{-1})$ but from 2009-2011, the surface seawater most likely transitioned to a sink (-0.018 mol C $m^{-2} yr^{-1}$, 0.00 mol C $m^{-2} yr^{-1}$, and -0.032 mol C $m^{-2} yr^{-1}$, respectively. Because of the small value of ΔpCO_2 and resulting high relative uncertainty, it is not possible to classify at present the Kilo Nalu location as a CO₂ source or sink. Each location also displayed a seasonal trend in flux. At CRIMP-2, waters oscillate between strong sources in the summers (mean of 1.66 mol C m^{-2} yr⁻¹) and weaker sources in the winter (mean of 0.66 mol C m⁻² yr⁻¹). The same trend was observed at both the Ala Wai and Kilo Nalu locations, as the water column switched between source and sink behavior on a summer to winter time scale.

Data are not shown from the Kilo Nalu buoy for the winter of 2010-2011 or the summer of 2011due to problems with the equilibrator.

Table 4

Table 5

Total alkalinity at the buoy locations ranged from a mean of 2225 µmol kg⁻¹at CRIMP-2 to 2306 and 2298 at Kilo Nalu and Ala Wai, respectively. AT data from synoptic sampling are shown in Figure 7. C_T and pH (total scale), calculated from A_T and pCO₂, were lower at CRIMP-2 (1960 µmol kg⁻¹ and 7.99) than at Ala Wai and Kilo Nalu (~1993 µmol kg⁻¹ and 20 333 ~8.06). Carbon system parameters and statistics are shown in Table 6. The Kilo Nalu environment displayed the least amount of variability, while the water at CRIMP-2 and at Ala Wai had similar absolute variability, despite differences in the mean values. Data from surface waters at Station ALOHA, from the Hawaiian Ocean Time-series (HOT, http://hahana.soest.hawaii.edu/hot/hot-dogs/), during the same time period, are also shown and are most comparable with Kilo Nalu. Data from Shamberger et al. (2011) taken from 3 separate synoptic experiments at the CRIMP-2 site and a mid-reef site, during 6/08, 8/09, and 2/10, are consistent with the data from this study. Figure 8 shows the relationship between A_T and C_T at each location. Both A_T and C_T have been normalized to a salinity of 35 (nA_T and nC_T) to help reduce the effects of mixing of water masses with different properties. HOT data from 2008-2011 (<25m) are also shown as an end-member point included in each regression.

Table 6

- 346 Figure 7
- 347 Figure 8
- **5 Discussion**

349 5.1 Physical Controls of pCO₂

During the 3.5 year study period, all three buoy environments displayed temporal cycles of pCO₂ on time scales ranging from daily to seasonal. Data (in Figure 2a), collected every three hours, from all three buoys show the long term seasonal variability, due in part to temperature

changes, as well as the short term (hourly-weekly) variability due to both biological activity and physical forcing.

Monthly mean pCO_2 is shown in Figure 3a to clarify the seasonal trends of pCO_2 . The summer dry period, from May-October, is marked by warmer water and air temperatures and significantly less precipitation, resulting in less nutrient-rich land runoff (Ringuet and Mackenzie, 2005; De Carlo et al. 2007, Hoover and Mackenzie, 2009; Drupp et al. 2011). CRIMP-2 exhibited a seasonal amplitude of pCO₂ of ~100 µatm, while seasonal amplitudes at Kilo Nalu and Ala Wai were ~ 35 μ atm. These amplitudes are based on monthly mean data in order to eliminate large transient increases or decreases unrelated to seasonal variation. Because increasing temperature raises seawater pCO₂ (and vice versa) by 4.23% per degree Celsius (Takahashi et al. 1993), pCO₂ is elevated in the summer and depressed in the winter at all three locations. Temperature changes, however, are not the only cause of pCO₂ changes in the water column at any of these locations. If temperature was the only factor, the annual seasonal amplitudes (based on monthly means) should be nearly twice the observed ranges at Kilo Nalu and Ala Wai, and slightly less than the observed range at CRIMP-2. The other main factors driving long-term pCO₂ trends include biological processes, residence time (a function of mixing), and air to sea gas exchange (a function of wind speed and ΔpCO_2). These processes are discussed below.

In order to separate the effects of temperature and biological processes on pCO₂, two different normalizations were applied (Takahashi et al., 1993). Equation 4 was used to derive the temperature-dependent effects on pCO₂. The values are calculated based on a mean pCO₂ at each location for the entire study period, and are representative of temperature-induced changes under isochemical conditions (i.e. no biological/mixing effects on water chemistry). On short time scales, greater variation is observed at CRIMP-2 than at either Kilo Nalu or Ala Wai, due to larger daily temperature fluctuations, the shallow depth of the reef and the longer water residence time of water in Kaneohe Bay. On a seasonal to annual scale, greater variation (~1.5 times) is also observed at CRIMP-2 compared to the other locations. This enhanced variation in pCO₂ is caused by an annual temperature range at CRIMP-2 approximately 1.5 times larger than at either Kilo Nalu or Ala Wai (Figure 3b). Temperature is the strongest contributor to long-term pCO₂ variations, and it even has a daily effect (mean daily range of 21 μ atm), but it is not the only

physical factor affecting pCO_2 .

Residence time is another important physical factor impacting pCO₂, as it controls the amount of time the water column is exposed to biological processes. Longer residence times allow for an accumulation of CO_2 from continuous respiration and calcification by the biological community. The residence time of water in Kaneohe Bay is controlled by wave action and wind speed and less by tidal mixing. Tradewinds blow from the northeast approximately 80% of the year with a mean speed of 4.9 m s⁻¹, and wave action is considered to be the dominant control on current speed across the reef (Lowe et al., 2009b). Water takes only a few hours to travel across the reef flat, although during light winds (and the consequent low wave action), the water can take more than a day to flow across the reef (Shamberger et al., 2011). Lowe et al. (2009 a/b) also show some model runs with residence times exceeding 24 hours. An example of longer residence time leading to increased pCO_2 is shown in Figure 4. Beginning on May 1, 2010 and lasting until May 5, wind speeds were dramatically lower ($<2 \text{ m s}^{-1}$) than typical ($\sim4-6 \text{ m s}^{-1}$). The current speed across the reef, based on offshore significant wave height using the relationship developed for the Kaneohe Bay barrier reef by Lowe et al. (2009b) immediately prior to this period was approximately 30 cm s⁻¹ and dropped to a low of ~10 cm s⁻¹ on May 3. The reduction in wind/current speed significantly increased the water residence time on the reef, resulting in an accumulation of CO₂ derived from calcification and respiration. In addition, low wind speeds reduce air-sea gas exchange, thereby lowering gas evasion rates, and further enhancing the accumulation of CO_2 in the water column (Massaro et al., 2012). In a related study on the Kaneohe Bay barrier reef, Shamberger et al. (2011) found that daily cycles of A_T were enhanced and pCO₂ was elevated on days when wind speeds were low and residence time was long (13+ hours). The higher mean pCO₂ at CRIMP in southern Kaneohe Bay, despite considerably lower rates of calcification (Drupp et al., 2011; Massaro et al., 2012), was attributed primarily to a longer residence time of approximately 15-30 days (Ostrander et al., 2008; Lowe et. al, 2009a), which further supports our findings.

The chemistry of water at Kilo Nalu, in stark contrast to Kaneohe Bay, displays little short- or long-term variability. It is reasonably representative of the surrounding open ocean water, with a mean pCO_2 of 378 µatm for the study period and a standard deviation that is four times less than that at CRIMP-2. The Kilo Nalu location is very weakly stratified and

consistently flushed by tides, currents, and waves. Stratification of the water column at Kilo Nalu is mainly caused by daily heating and is relatively weak, if present at all, while convective mixing destabilizes the site at night (Hamilton et al., 1995; Sevadjian et al., 2010). Water column productivity is generally low and chlorophyll-a concentrations rarely exceed 0.5-1.0 µg L^{-1} (Tomlinson et al., 2011). Benthic productivity is also likely low as the bottom substrate is primarily sand over limestone pavement with very little coral or algal cover. This deeper (relative to CRIMP-2), fairly well mixed water column masks much of the CO₂ signal, which is low to begin with at this site. The Ala Wai buoy location experiences similar physical forcings as Kilo Nalu, which contribute to a similar mean pCO₂ of 389 µatm, although its proximity to the mouth of the Ala Wai Canal results in stronger land-ocean interactions at this site due to enhanced fluvial runoff and nutrient inputs (discussed below).

424 5.2 Biogeochemical Controls of pCO₂

The water column at CRIMP-2 (blue in Figure 2) exhibits the largest short-term variability of pCO_2 observed at all three locations owing to high rates of daily calcification and productivity on the reef flat (Shamberger et al., 2011). A similar trend of short-term variability was described by Drupp et al. (2011) at the CRIMP location in southern Kaneohe Bay.

Normalization of pCO_2 to both mean temperature and mean pCO_2 separates the effects of temperature-independent and temperature-dependent processes on pCO₂ (Takahashi et al. 1993, 2002). Temperature normalization (equation 3) alone removes the effect of temperature on pCO_2 and provides a value that Takahashi (1993) refers to as reflective of the "net biology effect." It is clearly evident in Figure 2b that T-normalized pCO₂ ("net biology effect") accounts for the vast majority of short-term variability of pCO_2 and even displays a slight seasonal trend. The standard deviation at all three study locations barely changes between the *in-situ* pCO₂ and the T-normalized value, indicating that the majority of the variance in the *in-situ* pCO₂ values cannot be attributed to temperature. The CRIMP-2, Kilo Nalu, and Ala Wai sites have T-normalized standard deviations of 84.5 µatm, 17.5 µatm, and 24.4 µatm, respectively, nearly identical to the standard deviations of the *in-situ* values of 83.4 µatm, 18.2 µatm, and 25.3 µatm, respectively.

Figure 9 shows the A_T at CRIMP-2 versus salinity. The conservative mixing line (black
dashed line) was derived by Massaro et al. (2012) for Kaneohe Bay, using offshore seawater and

a zero salinity intercept derived from water samples collected at the mouth of Kaneohe Stream, the major river flowing into Kaneohe Bay. All of the CRIMP-2 samples, except for 11/14/2009, fall below the mixing line supporting the hypothesis that calcification/productivity processes across the reef are the main drivers of temporal changes in pCO₂ and A_T. The only sample that does not fall below the mixing line was taken on a day with stronger than average trade winds (gusts up to 9 m s⁻¹) resulting in faster current speeds (short residence time), as well as below average solar radiation reducing photosynthetic and calcification rates. The point falls virtually on the mixing line (A_T=2297 μ mol kg⁻¹) suggesting that the seawater sampled at that time may have represented open ocean water.

Figure 9

Daily pCO₂ values at CRIMP-2 on the reef fluctuate widely with a mean range four times greater than the two other buoys. Chlorophyll measured at all three buoys is low (mean<0.6 µg L^{-1}), implying that most of the photosynthesis/respiration and calcification/dissolution signals observed at CRIMP-2 are driven by the benthic reef flat organisms, rather than organisms in the water column. Average values of total nitrogen $(NO_3^- + NO_2^- + NH_3)$ were 0.55 µmol at sites across the reef and 0.46 µmol at CRIMP-2, and inorganic phosphate was below detection levels $(0.29 \text{ }\mu\text{mol }L^{-1} \text{ for all samples.}$ Water column production was likely nutrient limited as previously shown by Ringuet and Mackenzie (2005) and Drupp et al., (2011). A study performed by Shamberger et al. (2011) found that net ecosystem production (NEP) on a daily basis ranged from -378 mmol $m^{-2} d^{-1}$ to 80 mmol $m^{-2} d^{-1}$ (negative values indicate net respiration, positive values indicate net production). The reef varied from net autotrophic during the day to net heterotrophic at night during this study, a characteristic of many reef systems.

464 Daily cycles of pCO₂, typical of what would be expected from daytime photosynthesis 465 and nighttime respiration, are characterized by a maximum pCO₂ occurring at night/early 466 morning, usually between 04:00-08:00, with a corresponding minimum pCO₂ occurring in the 467 mid-afternoon, ~14:00-17:00. This cycle mirrors that of pO₂, which peaks in the late-afternoon 468 and reaches a minimum in the early morning. This same cycle, caused by photosynthesis 469 consuming CO₂ and producing O₂ during the day while respiration at night consumes O₂ and 470 produces CO₂, was also observed at CRIMP (Drupp et al., 2011 and Massaro et al., 2012).

 Autocorrelations, highlighted in the results and Table 3 shows that points separated by multiples of 24h (same time each day) are highly correlated even up to 72h apart (all points separated by 72h have correlations of ~0.56). After 72h, no points have a correlation better than 0.50. Because of this, CO_2 levels at the buoy locations can be fairly accurately predicted approximately three days into the future under normal conditions. All points separated by 12h show weak or negative correlations. Because the points 12h apart are not strongly anticorrelated, daytime p CO_2 values are not an indication of what the nighttime values will be and vice versa. This is due to daytime photosynthesis and calcification being controlled primarily by solar radiation and nutrient availability, which change day to day. In addition, the water chemistry is affected by changes in residence time, which is primarily a function of wind speeds and forereef wave action. Because many of these factors are transient in nature, it is difficult to predict p CO_2 more than 2-3 days in advance. This makes long-term high resolution monitoring all the more valuable.

Although the data at CRIMP-2 are consistent with P/R cycles, Shamberger et al. (2011), in a complementary study on the barrier reef, showed that calcification increases during the day, with some dissolution occurring at night, and daily net ecosystem calcification ranged from 174 to 331 mmol CaCO₃ m⁻² d⁻¹ (positive values indicate net calcification, negative values indicate net dissolution). The release of CO₂ during calcification would counter its uptake during photosynthesis. Because of this, the observed pCO₂ is a **net concentration** that would be much lower were it not for calcification concurrent with photosynthesis. The opposite can be true at night when the uptake of CO₂ during dissolution counters the release of CO₂ from respiration.

The subdued seasonal trend remaining in the T-normalized pCO_2 data is different between CRIMP-2 and the south shore buoys, Kilo Nalu and Ala Wai. T-normalized pCO₂ at CRIMP-2 decreases during the winter (Nov.-Apr.) and increases in the summer (May-Oct.). This trend, however, is reversed in both Kilo Nalu and Ala Wai where T-normalized pCO_2 decreases during the summer months and increases in the winter. The trend on the south shore is consistent with summer/winter photosynthetic/respiration cycles (and a relative lack of calcification), as shown, for example, by De Carlo et al. (this volume) in the Bay of Villefranche in the NW Mediterranean. Decreasing SST lowers the rate of water column primary production during the winter, leading to a slight increase in pCO₂ at Kilo Nalu and Ala Wai. In addition, at

Ala Wai, increased organic rich runoff in the winter can cause periods of elevated pCO₂. However, at CRIMP-2 on the barrier reef, where calcification exerts a major biological control on pCO₂, decreased wintertime temperatures also favor lower rates of calcification (e.g. Jokiel and Coles, 1977; Coles and Jokiel, 1978; Marshall and Clode, 2004; Langdon and Atkinson, 2005), driving the T-normalized pCO₂ down. However, this effect may be somewhat mitigated by the simultaneous temperature-driven decrease in primary productivity (raising pCO₂ via increased respiration).

Although the mean pCO_2 and seasonal pCO_2 trends at the Ala Wai buoy are similar to those at the nearby Kilo Nalu site, seawater does experience greater short-term variability (evident as a slightly higher standard deviation), owing to its location near the mouth of the Ala Wai canal. This canal drains a considerable portion of the Honolulu urban watershed and its waters are enriched both in nutrients and organic matter as well as a number of pollutant chemical species (e.g., De Carlo et al., 2004, 2005; Tomlinson et al., 2011). This continuous and occasionally strong supply of nutrients and organic matter from the Ala Wai canal contributes to an enhanced daily photosynthesis/respiration cycle. During periods of heavy rainfall, the Ala Wai delivers large pulses of freshwater, sediment, and nutrients (Tomlinson et al., 2011). Tomlinson et al. (2011) described a storm in the Honolulu watershed that occurred on 13 March 2009 resulting in elevated chlorophyll at the Ala Wai buoy, peaking at 3-5 times the baseline values observed during the week following this intense 24-hour storm. In the two days following this storm, daily pCO_2 cycles, measured by WQB-AW, were significantly dampened due to an increase in photosynthesis. However, the large amount of terrestrial organic matter, introduced by runoff, also enhanced respiration, driving pCO₂ levels back up at night. On 17 March 2011, pCO_2 reached its highest value of the season, 505 µatm. The following two days were marked by a phytoplankton bloom with chlorophyll levels 4.5 times greater than baseline values (Tomlinson et al., 2011), most likely caused by an increased nutrient subsidy following the remineralization of runoff derived organic matter. This same trend was also observed at the Kilo Nalu location, albeit on a considerably smaller scale. Following this storm, Kilo Nalu experienced daily cycles of pCO₂ of ~65 μ atm, with maximum pCO₂ above 400 μ atm from March 15 to 19. In comparison, the Kilo Nalu location had not shown pCO₂ over 400 µatm in more than a month. The response of Kilo Nalu to runoff from the Ala Wai canal is not typically this strong but

particularly large amounts of rain in the Ala Wai watershed and overland flow near the buoy location undoubtedly contributed to additional non-point source runoff and an influx of organic material. Runoff-induced phytoplankton blooms and subsequent remineralization were also shown to affect similarly the pCO₂ levels in southern Kaneohe Bay at the original CRIMP location (Drupp et al., 2011).

Even under baseline conditions, the effect of remineralization of runoff derived organic matter can be observed at the Ala Wai location. Figure 8 shows nA_T versus nC_T at the three buoy locations. After normalization, the sample values, especially those from Kilo Nalu, cluster around values similar to the open ocean. Mean HOT surface (<20 m depth) samples have $nA_T = 2307 \mu mol kg^{-1}$ and $nC_T = 1986 \mu mol kg^{-1}$ (updated from Keeling et al., 2004; http://hahana.soest.hawaii.edu/hot/hot-dogs/) and have been added as an end-member point included in the regression. Some of the Ala Wai samples have elevated C_T due to the higher levels of pCO₂ derived from net respiration and remineralization of organic rich runoff from land. The elevated A_T concentrations at Ala Wai, relative to the HOT data may reflect dissolution within the water column as it is very unlikely that sufficient alkalinity is being added from the land runoff. In contrast, the nA_T vs nC_T plot for CRIMP-2 is scattered due to the variety and greater variability of physical and biological processes previously discussed. Residence time is an important factor affecting A_T, as current speed and, on the rare occasion, direction across the reef are not always constant. Future plans include the deployment of acoustic Doppler current profilers (ADCP) to understand more fully the effects of residence time on pCO₂ and A_T in these different reef settings.

5.3 Air-Sea Gas Exchange

Instantaneous CO_2 fluxes are shown in Table 4 and Figure 6a-c. CRIMP-2 shows the largest range from 3.08 mmol C m⁻² hr⁻¹ to -1.00 mmol C m⁻² hr⁻¹. Although this range was nearly two times greater than the CRIMP range reported by Massaro et al. (2012), the mean instantaneous flux was 0.128 mmol C m⁻² hr⁻¹, only about half that of the CRIMP mean. This can be attributed to the latter location having higher average pCO₂ concentrations despite having smaller variations in pCO₂ and lower levels of biological activity (including very little calcification). The lower variation, but higher mean concentration, is a result of the longer residence time of the south bay waters, which is on the order of days to weeks rather than a few

hours as observed over the barrier reef. The waters surrounding all three buoys during parts of the study period acted as CO₂ sources to the atmosphere (positive flux), as well as sinks from the atmosphere (negative flux) during other time periods due to seasonal and diel variations in water column pCO₂ and transient events (often storms), resulting in phytoplankton blooms (pCO₂) drawdown) or organic matter respiration (pCO_2 increase). The net annualized fluxes for CRIMP-2, Ala Wai, and Kilo Nalu were 1.15 mol C $m^{-2} y^{-1}$, 0.045 mol C $m^{-2} y^{-1}$, and -0.0056 mol C m⁻² y⁻¹, respectively. The flux at CRIMP-2, on the barrier reef, is consistent with, but lower than the flux obtained by Massaro et al. (2012) at CRIMP of 1.76 mol C $m^{-2} y^{-1}$ in southern Kaneohe Bay from 2006-2008, and Fagan and Mackenzie's (2007) value of 1.45 mol C $m^{-2} y^{-1}$ for the entire Kaneohe Bay lagoon. However, these three flux estimates are considerably greater than those calculated for the fringing reefs of Honolulu.

The Ala Wai study site is strongly influenced by mixing with open ocean seawater. However, it remains a net annual source of CO₂ to the atmosphere due to its proximity to frequent nutrient- and organic carbon-rich freshwater discharges and to respiration of labile organic matter in the water column (previously discussed). Waters near the Ala Wai buoy do, however, switch between source and sink seasonally. In the rainy winter months, enhanced nutrient-rich discharge from the watershed stimulates primary productivity during the day, although delivery of pre-existing organic matter also can increase respiration at night. Furthermore, the enhanced solubility of CO₂ at lower seawater temperature during the winter leads to a lower seawater pCO_2 relative to the atmosphere, resulting in air-sea gas exchange. This seasonality is reflected in the data in Table 5.

582 At all three locations, the seasonality in CO_2 flux (source in summer, sink in winter) is 583 further enhanced by the seasonality of atmospheric CO_2 . In the winter, CO_2 in the Hawaiian 584 atmosphere is significantly higher than during the summer

50 585 (http://www.esrl.noaa.gov/gmd/ccgg/trends/). The higher atmospheric winter concentrations are 51 586 part of the annual northern hemisphere P/R cycles. Because the seasonal variability of 53 587 temperature and pCO₂ in the water column and in the atmosphere are in opposite directions, the 55 588 ΔpCO_2 is affected throughout the entire year. For example, lower atmospheric CO₂, coupled 57 589 with higher temperature-driven water column pCO₂ during the summer, enhances gas exchange 59 590 by increasing the concentration gradient between the two reservoirs. In the winter, high

atmospheric CO_2 and lower water column p CO_2 reduce the size of the ΔpCO_2 term. These opposing effects contribute to the seasonality in the source/sink function. At CRIMP-2, the ΔpCO_2 during the summer is driven even higher by calcification. Despite the finding from a recent short-term Kaneohe Bay barrier reef study of Shamberger et al. (2011) that did not show a significant increase in summertime calcification, many other studies support this conclusion. It has been found that corals in Hawaii and at other reef sites reach a maximum calcification rate at approximately 26°C, the ambient temperature of Hawaiian summers (Jokiel and Coles, 1977; Coles and Jokiel, 1978; Marshall and Clode, 2004; Langdon and Atkinson, 2005). Thus in general, we would anticipate higher calcification during the warm summertime. In contrast, at both Kilo Nalu and Ala Wai, the biological processes (higher summertime productivity) counteract the temperature seasonality (see Figure 6a-c), causing a decrease in ΔpCO_2 . In addition to the changes in the CO₂-carbonic acid system chemistry of the water, Hawaiian summers typically have stronger and more persistent tradewinds, which increase the rate of the gas exchange. It is these additive effects that lead to CRIMP-2 being a significant net annual source, and opposing effects that lead to Ala Wai and Kilo Nalu being a very mild source and mild sink, respectively.

Flux values presented here compare well to previous studies. Above we already compared the fluxes reported in Kaneohe Bay by Fagan and Mackenzie (2007) and Massaro et al. (2012). The fluxes estimated in this study are also of similar magnitude to those reported for other tropical reef environments. Ohde and Van Woesik (1999) calculated a flux of 1.8 mol C m^{-2} yr⁻¹ for an Okinawan reef in Japan. Comparable reef environments in Bermuda (1.2 mol C m^{-2} yr⁻¹ at Hog Reef Flat, Bates et al., 2001) and the northern Great Barrier Reef (1.1 to 1.5 mol C m^{-2} yr⁻¹ on Yonge Reef, Frankignoulle et al., 1996) also exhibited comparable fluxes.

614 6 Conclusions

615 Coastal tropical reef surface waters at multiple buoy moorings around the island of Oahu, 616 Hawaii, exhibit highly variable CO_2 -carbonic acid system chemistry and magnitude and 617 direction of CO_2 exchange due to differences in land/ocean interactions, water column and 618 benthic (CRIMP-2) photosynthesis/respiration, and residence time. Data from this study and 619 previous work in Kaneohe Bay (De Carlo et al., 2007; Drupp et al., 2011; Ringuet and

Mackenzie, 2005; Massaro et al., 2012) demonstrate the utility of long-term, high-resolution
sampling to characterize accurately the carbon chemistry of the coastal surface waters of Hawaii
and other similar tropical reef environments where diel cycles, land-derived runoff, and seasonal
changes and physical characteristics can all combine to alter the CO₂-carbonic acid system
chemistry of the water column on multiple time scales

Seasonal biological and temperature effects on the pCO_2 of the water column were in phase at CRIMP-2, due to vigorous benthic calcification, but at the Ala Wai and Kilo Nalu, where there is much less of an overall calcification signal, these seasonal effects were out of phase, leading to a smaller range of pCO₂. Total alkalinity data showed that the carbon chemistry of surface waters at the Ala Wai and Kilo Nalu buoy sites is primarily controlled by water mass mixing that is driven by tides and waves. At these locations, A_T and C_T values are very similar to open ocean values, with slightly elevated pCO₂ due to increased remineralization of organic matter derived from land runoff and respiration of organic matter produced in situ during nutrient-rich pulses of land runoff. A_T and C_T at the CRIMP-2 barrier reef location are primarily controlled by biological activity and the residence time of water on the reef.

Air-sea CO₂ exchange also showed seasonality. Both the Ala Wai and Kilo Nalu buoy locations became a sink during the winter months as cooler temperatures and brief phytoplankton blooms drew down pCO₂. While waters at CRIMP-2 never became a net sink during the winter months, it did display periods of sink activity causing an overall reduction in source strength in winter. During the summer months, high temperatures coupled with lower atmospheric CO₂ and stronger winds, as well as enhanced calcification (at CRIMP-2), led to each location acting as a source of CO_2 to the atmosphere. However, as previously stated, the processes controlling p CO_2 at CRIMP-2 are in phase, likely amplifying the net air-sea gas exchange by increasing seasonal variability, whereas, at Ala Wai and Kilo Nalu these processes are competing (out of phase) and likely reduce the net air-sea gas exchange. Currently Kaneohe Bay, both within the inner lagoon (Fagan and Mackenzie, 2007; Massaro et al., 2012) and its barrier reef, is a net source of CO₂ to the atmosphere, while the Ala Wai and Kilo Nalu areas are a weak source and weak sink of this gas, respectively.

It is important to note that this study along with the previous studies in southern KaneoheBay detailed by Drupp et al. (2011) and Massaro et al. (2012) have found significant differences

in CO_2 -carbonic acid system chemistry and CO_2 exchange between four Hawaiian reef settings located in close proximity to each other. It is critical that we do not over-extrapolate results from one setting to a global perspective as more and more studies show that CO_2 dynamics in coral reef environments are highly variable and often quite different between various locations.

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Figure 4: pCO_2 and pO_2 at CRIMP-2 buoy are anti-correlated (see Table 3) since the reef is highly productive. Photosynthesis during the day decreases pCO_2 while increasing pO_2 with a max O_2 saturation resulting at ~14:00. Respiration at night drives pCO₂ up, with maximum values measured around 04:00-07:00. Elevated pCO₂ from 5/2-5/4 was caused by low wind period (0-2 m s⁻¹ rather than normal 6-8 m s⁻¹) increasing residence time and lowering the gas transfer velocity resulting in a buildup of CO₂ from respiration and calcification.

Figure 5: Autocorrelation of hourly pCO₂ at CRIMP-2 clearly shows a daily cycle which is highly correlated even up to 72 hours.

Figure 6: Instantaneous air-sea gas exchange of CO_2 at all 3 buoy locations is shown. Note that positive values represent a source of CO_2 to the atmosphere, while negative values represent a sink of CO_2 from the atmosphere. Each environment displays a seasonal trend of source activity in the summer and sink activity in the winter.

²² 911 **Figure 7:** Total alkalinity measured at each buoy location from 2009-2011 is displayed. A_T at the CRIMP-2 buoy is consistently lower, due to calcification as open ocean water flows across the barrier reef. Water at the Ala Wai and Kilo Nalu sites is more representative of open ocean **914** water, showing only a small change from typical offshore water, which far less variability over time. 27 915

Figure 8: A_T and C_T have been normalized to S=35 to remove mixing effects and show the relationship between the two parameters. Regression lines include the HOT mean data from 2008-2011 to serve as an end-member point. A system controlled solely by photosynthesis and 32 918 respiration would have a theoretical slope of 1, while a system controlled only by calcification and dissolution would have a slope of 2. The data for Ala Wai and Kilo Nalu cluster around **921** typical open ocean values, although a small addition of pCO₂ from organic matter remineralization results in higher C_T values at Ala Wai. CRIMP-2 is highly variable due to the combined effects of P/R, calcification/dissolution and residence time across the reef.

Figure 9: The A_T vs salinity plot can help identify whether alkalinity was added or removed as water flowed across the reef as water moved towards CRIMP-2. Because all but one of the samples fall below the conservative mixing line, alkalinity was biogeochemically removed 44 926 across the reef flat. The black square is the open ocean end member point used to calculate the 47 928 conservative mixing line.

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Figures and Tables Click here to download Colour Figure: Drupp et al. Comparison of CO2 Dynamics Figures.doc

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Figure 1a



Figure 1b



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Figure 8



- **Figure 9**

- 53 Tables

	Min	Max	Mean	
pCO _{2 sw}	(µatm)	(µatm)	(µatm)	St. Dev
CRIMP-2	196	976	439	83.4
Kilo Nalu	316	479	378	18.2
Ala Wai	302	514	389	25.3
CRIMP-1	225	671	448	50.8

- **Table 1:** pCO_2 statistics are shown for each of the three buoys from 6/10/08-12/31/11, as well as
- from the original CRIMP-1 buoy in southern Kaneohe Bay from 11/30/05-5/17/08.

pCO _{2 sw}	Daily Range Mean (μatm)	Daily Range Max (µatm)	Daily Range St. Dev
CRIMP-2	196	557	72.2
Ala Wai	52	146	21.3
Kilo Nalu	30	124	19.3
CRIMP-1	42	204	25.4

61 **Table 2:** Daily ranges of pCO_2 for each buoy over the study period were calculated and a mean,

62 min, max, and standard deviation of the ranges were calculated. Daily pCO₂ values at CRIMP-2 63 on the reef fluctuate widely with a range 4x greater than any other buoy due to elevated levels of

64 productivity, respiration, calcification, and dissolution.

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Buoy	pCO ₂ vs pO ₂	pCO ₂ A	utocorr	elation
вибу	Correlation	12hrs	24hrs	72 គ្រ ទ្រ
CRIMP-2	-0.75	-0.20	0.76	0.59
Ala Wai	-0.29	0.31	0.72	0.62
Kilo Nalu	-0.23	0.48	0.75	0.39
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78 **Table 3:** A negative (albeit weak for AW and KN) anti-correlation is exhibited between pCO₂

and pO₂. Autocorrelation shows a strong 24 hour periodicity indicative of P/R cycles.

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% Time Net June 2008 -% Time Max Max Annualized Mean As Dec 2011 Source Sink As Sink Source Flux mol C m⁻² mmol C $m^{-2} hr^{-1}$ yr⁻¹ CRIMP-2 3.25 -1.04 78.0% 21.3% 0.13 1.15 Ala Wai 1.72 -0.89 0.005 66.0% 33.9% 0.045 Kilo Nalu -1.56 0.00 51.4% 48.4% -0.0056 0.373 CRIMP-1* 1.34 -0.71 0.20 94.2% 5.5% 1.78*

82 **Table 4:** Instantaneous flux ranges are shown for each buoy as well as the percentage of time

83 spent as a source and a sink. Note that these values do not add up to 100% due to periods of no

- wind or of pCO_{2sw} equaling the pCO_{2air} . Net annualized flux measurements show that the
- CRIMP-2 buoy environments is a significant source of CO₂ to the atmosphere (positive values
- indicate sources), while the Ala Wai and Kilo Nalu environments are barely a net source and

sink, respectively.

- *CRIMP-1 data set from 11/05-6/08

	2008		2009		2010		2011
CRIMP-2	1.39		0.82		1.10		1.31
Ala Wai	0.064		0.027		-0.003		0.092
Kilo Nalu	0.073		-0.018		0.00		-0.032
	Summer	Winter	Summer	Winter	Summer	Winter	Summer
	' 08	'08-'09	' 09	'09-'10	'10	'10-'11	'11
CRIMP-2	1.60	0.31	1.63	0.56	1.71	1.12	1.71
Ala Wai	0.069	-0.035	0.12	-0.057	0.049	0.028	0.13
Kilo Nalu	0.074	-0.082	0.086	-0.040	0.055	N/A	N/A
Fluxes in mol C m ⁻² yr ⁻¹							

Table 5: Annualized fluxes for each year of the study period as well as broken into the summer

and winter (dry and wet) seasons. Note that positive values indicate source behavior of CO_2 to

the atmosphere while negative (and red) values indicate sink behavior of CO_2 from the

atmosphere. Because the buoys were deployed in June 2008, the annualized flux for 2008 is only

calculated from June 2008-December 2008. Each buoy oscillates between source behavior in the

summer and weak source or sink behavior in the winter. Summer (dry season) is defined as

May-October and winter (wet season) is November-April.

Location	nA _T (µmol kg ⁻¹)		nC _T (µmol kg ⁻¹)		pН		$\Omega_{ m Ar}$	
	Mean	Range	Mean	Range	Mean	Range	Mean	Range
CRIMP-2	2229.53	2181.63-	1966.79	1850.94-	7.99	7.90-8.13	2.96	2.40-3.77
(this study)		2278.55		2020.93				
Ala Wai	2305.53	2166.87-	2001.44	1879.57-	8.05	7.99-8.09	3.40	3.04-3.73
(this study)		2349.53		2050.10				
Kilo Nalu	2301.75	2262.85-	1988.35	1945.80-	8.06	8.03-8.08	3.50	3.26-3.63
(this study)		2335.35		2013.35				
HOT (2009-	2306.53	2298.03-	1984.74	1969.02-	8.07	8.06-8.08	3.63	3.56-3.71
2011)		2317.14		1998.93				
CRIMP-2							2.83	2.50-3.07
(Shamberger								
et al., 2011)								
K-Bay							3.18	3.08-3.25
Midreef								
(Shamberger								
et al., 2011)								

112 **Table 6**: Carbon system parameters C_T , pH, and Ω_{Ar} were calculated from A_T and pCO₂ samples

113 collected at each buoy. Both A_T and C_T have been normalized to salinity=35 (n A_T and n C_T). Ω_{A_T}

114 data from other studies in Kaneohe Bay compare well with the values calculated for CRIMP-2

during this study. Data from Station ALOHA and the HOT program also agree with our findings

that the water column at Ala Wai and Kilo Nalu represents open ocean water that has been

117 slightly altered by respiration and organic matter remineralization.