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3	Planktonic Larval Duration, Age and Growth of Ostorhinchus doederleini (Pisces: Apogonidae) on
4	the Southern Great Barrier Reef, Australia
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26 Abstract

27 Cardinalfishes (Apogonidae) are abundant on corals reefs, but there are few data on demography to 28 understand trophodynamics and population dynamics. Ostorhinchus doederleini is a small and abundant 29 apogonid on the Great Barrier Reef (GBR) and throughout the Western Pacific Ocean. We present key 30 demographic parameters for the entire life history from the southern GBR. Daily deposition of 31 increments in otoliths was validated. Fish had a Planktonic Larval Duration (PLD) of 16 to 26 days. 32 PLD was established from fish collected immediately prior to settlement as no settlement mark was found. Fish grew at about 0.35 mm d^{-1} for the first 20 d after settlement. Fish reached a maximum 33 34 standard length at about 200 d and no fish lived longer than 368 d at four reefs separated by kilometers 35 to tens of kilometres. There was no evidence for differences in size at age between sexes. Mortality 36 was very high, for fish greater than 60 days old mortality rates ranged from 2.9 to 4.6% per day. Short 37 lives and high mortality rates makes O. doederleini, and potentially other apogonids, vulnerable to 38 recruitment failure. Here we review data on the demographic characteristics of other reef fishes. 39 Although some taxa live to over 50 years, the short lives of apogonids are most aligned with the 40 Gobiidae and Blenniidae (i.e. typically < 1.5 years). Descriptions of fish size, age, longevity, growth 41 and mortality; from hatching to age maxima are very rare for most taxa, even at the level of family.

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43 Introduction

Nocturnal fishes are an important trophic group on both coral and temperate reefs (e.g. Marnane and
Bellwood 2002; Annesse and Kingsford 2005). Fishes belonging to the Apogonidae, Holocentridae,
Nemipteridae and Pempheridae are abundant on reefs and species richness is high (Hobson and Chess
1978; Sale and Douglas 1984; Bellwood 1996; Greenfield 2003; Mabuchi et al. 2006; Thacker and Roje

48 2009; Gardiner and Jones 2010; Boaden and Kingsford 2012). These taxa feed at night on a range of 49 invertebrates and fishes (Hiatt and Strasberg 1960; Marnane and Bellwood 2002; Annesse and 50 Kingsford 2005; Barnett et al. 2006) and are the prey of many piscivorous taxa (e.g. Kingsford 1992). 51 Apogonids, and other nocturnal fishes, generally occupy caves and other habitats that provide 52 shelter (e.g. coral Gardiner and Jones 2005) during the day and undergo feeding migrations at night. 53 Movements range from spatial scales of metres to hundreds of metres. In general these fishes feed in 54 habitats that are different from those they occupy during the day (Gladfelter 1979; Marnane 2000; 55 Annesse and Kingsford 2005; Boaden and Kingsford 2012). The trophic importance of these 56 movements to reef ecosystems are is potentially substantial in that their feeding behavior and availability 57 to predators are likely to be significant carbon pathways. Furthermore, feeding behavior often varies 58 with size of fish. Many apogonids and other young nocturnal fishes (Annesse and Kingsford 2005) feed 59 in different habitats than large juveniles and adults (Finn and Kingsford 1996). 60 Most apogonids have paternal mouth brooding and a larval phase (Brothers et al. 1983). An 61 exception, the apogonid *Pterapogon kauderni* has paternal mouth brooding and direct development of 62 juveniles with no larval phase (Vagelli and Volpedo 2004). Great attention has been given to filial 63 cannibalism of eggs brooded in the buccal cavity of males O. doederleini (Okuda and Yanagisawa 1996) 64 and other species (Okuda et al. 1997; 2003; Kume et al. 2000). Despite their novel reproductive 65 biology, trophic importance and the recognition that there is high local population turnover (e.g. on 66 patch reefs Sale and Douglas 1984), there are few data on demography (but see Okuda et al. 1998). An 67 exception is Longenecker and Langston (2006) who found that Ostorhinchus rubrimacula (Apogonidae) 68 lived to a maximum of 274 days at a reef in Fiji. There are some estimates of Planktonic Larval Duration 69 (PLD) for 14 species of apogonids in Japan (mean PLDs ranged from 14 to 31 days, Ishihara and 70 Tachihara 2011), but the generality of these values is unknown.

71 A thorough understanding of population dynamics of any species requires information on growth 72 and mortality at all stages of life history (Hilborn and Walters 1992). The potential for high volatility in 73 population size would be greatest in species that have low age-maxima as there would be little in the 74 way of 'stored' year classes from seasons of high recruitment. Great variation in coral reef fish 75 recruitment among seasons is common (Doherty and Williams 1988), but populations persist, often 76 supported by a small number of large peaks in recruitment (Doherty and Fowler 1994; Kingsford 2009). 77 Apogonids are abundant on tropical reefs, but predation on these fishes is known to be very high (e.g. 78 Sale and Ferrell 1984). We have not found any other studies on the growth of apogonids prior to 79 settlement, immediately after settlement and to their age maxima. We hypothesised that age maxima in 80 Ostorhinchus doederleini would be low and this would be consistent among reefs. If this were the case 81 it would greatly influence the outcomes of connectivity among reefs and the persistence of population 82 genotypes. Recent evidence suggests that reef-based local populations of O. doederleini can be 83 genetically distinct even when the reefs are only separated by kilometers to tens of kilometers. Is has 84 been proposed that despite the relatively poor swimming abilities of larval apogonids (Stobutzki and 85 Bellwood 1997), they are capable of detecting olfactory cues from their natal reefs and this combined 86 with larval behavior facilitates reduced dispersal that can create genetically distinct populations (Gerlach 87 et al. 2007), but a knowledge of fish demography is essential for a full understanding of connectivity. 88 The objective of the present study was to describe the growth and mortality of Ostorhinchus 89 doederleini (Jordan and Snyder 1901) at four sites within the southern Great Barrier Reef. Wee 90 examined otolith microstructures to (1) validate the aging technique, and (2) determine the Planktonic 91 Larval Duration (PLD), the duration of the presettlement phase. We also, (3) describe the early growth 92 characteristics in presettlement and newly settled fish and (4) potential differences in the growth rates 93 and demographics of fish at multiple reefs. We also provide information on (5) mortality rates based 94 upon length-at-age estimates of post-settlement fishes. Finally, we (6) review and compare the
95 demographic characteristics of apogonids with other reef fishes.

96

97

98 Materials and methods

99 Study sites and fish sampling

100 Ostorhinchus doederleini were sampled at multiple sites on four reefs of the Capricorn Bunker group,

101 southern Great Barrier Reef: Heron Island, One Tree Island, Lamont and Fitzroy reefs (Fig. 1).

102 Presettlement fish were only collected at One Tree Island. Channel nets were set to sample fish on the

103 flood tides only at night in January 1994, February 1995 and February 2009 (Table 1). The net was

square mouthed (0.75 x 0.75 m), the mesh was 500 µm, organized as a box/pyramid with an efficiency

105 of 1:12 (mouth area: open sifting surface; full procedures see Kingsford and Finn 1997). To preserve

106 the calcium carbonate of otoliths, subsamples of collections from nets were preserved in ethanol.

107 Samples were then rinsed in alcohol to reduce the seawater content, the final percentage ethanol was

108 80%. Ostorhinchus doederleini entering the lagoon to settle were easily identified by morphology and

109 pigmentation (e.g. a black dot on the caudal peduncle).

Newly settled *O. doederleini* (10 to 18 mm SL) settle on sand and often away from continuous
reef (Finn and Kingsford 1996). We constructed six peaked patch reefs on sand (diameter ~ 1.5 m by 0.
5 m maximum height) 20 to 60 m from continuous reef. Patch reefs were monitored on a daily basis and
all newly settled fish were collected with fine-meshed hand nets and a wide size range was preserved in
80% ethanol for ageing. Some fish that colonized patch reefs were larger than new settlers (18 to 27
mm SL), but were collected for a description of early post-settlement growth (Table 1).

116 Juvenile and adult O. doederleini were collected at four reefs during January and February of 117 2009, 2010 and two large individuals in 2013 (Table 1). At each reef we sampled at multiple sites 118 separated by hundreds of metres to tens of kilometers to obtain a representative sample, and fish were 119 pooled for analyses. Juveniles and adults were collected from caves or branching coral, (especially 120 *Porites cylindrica* and branching *Acropora* spp.) and preserved in 80% ethanol. A wide size-range of 121 fish were collected using a mix of 5:1 ethanol: clove oil. At Heron Island and Lamont reef, we sampled 122 all fish from some large patch reefs and this allowed us to estimate mortality rates (see below). Great 123 care was taken to obtain all size classes of fish, especially large individuals that can be missed because 124 they are the most secretive in the backs of caves. Because O. doederleini were potentially short-lived 125 (e.g. Longenecker and Langston 2006) we also sampled fish in October 2010 in case the time of 126 sampling altered our estimates of age maxima; these samples did not allow estimates of mortality as 127 only large fish were selected. All fish collected in October 2010 were sexed macroscopically (ie the 128 presence of oocytes in females or fine-grain spermatogenic tissue in males) to determine size and age 129 versus sex relationships.

130

131 Otolith preparation

Increments in otoliths were used to obtain estimates of the age and growth of *O. doederleini*. All fish were weighed (\pm 0.01 g) and then measured (standard length [SL] \pm 1 mm). Sagittal otolith pairs were dissected, rinsed in water, and dried overnight. One otolith from each pair was randomly selected, embedded to the edge of a glass microscope slide using Crystal Bond thermoplastic glue. The sulcal groove of the otolith was orientated perpendicular to the surface of the slide and outer edge of the primordium was flush with the inside edge of the slide. Slides were then held vertically and ground to the transverse mid-plane using a Gemmasta GF4 Faceting Machine with a 3000µ diamond encrusted 139 steel disc. The glue was then reheated and otoliths were remounted with the newly ground surface

140 placed face down in the centre of the glass slide. Otoliths were ground until a thin section (20 µm thick)

141 was obtained which included the primordium.

142

143 Analysis of growth increments

144 The opaque zones visible in the internal structure of the otolith were counted along a radius from the 145 primordium to the outer edge of the otolith from an image taken with a compound microscope (Leica 146 DMLB using a Leica DC300 digital camera); a white incident light source was used. Alternating 147 translucent and opaque increments were interpreted as daily rings; no annuli were observed, even in the 148 largest fish. The counting of increments began from the first clear increment closest to the primordium. 149 Sections were coded and examined in random order and the opaque increments counted on two 150 occasions by the same observer separated by at least seven days. Counts of daily rings were compared 151 between these two occasions in order to assess the confidence that could be placed in the interpretation 152 of otolith structure. If increment counts differed by more than 10% between counting occasions, then the 153 otoliths were re-examined. If, following a third reading, agreement between the third and one of the two 154 other counts was not reached (only closely matching counts were used in analyses), then the otolith was 155 not included in the analysis. To calculate the age, an average of the two closest age estimates was used. 156 Rejection rate over the entire sample was 2.8%, leaving a total sample of n=581 for aging.

157

158 Validation of aging technique

159 To determine the periodicity of increment deposition in *O. doederleini* otoliths, a validation experiment

160 (using tetracycline) was completed at One Tree Island Research Station in January 1994. Fish were

161 treated with tetracycline, it is transported to the endolymph of the inner ear via the blood and deposited

in the otoliths and other bony tissues as a mark (Odense and Logan 1974). The tetracycline mark can be
viewed as a fluorescent line under ultraviolet light. The otoliths of 20 untreated fish were examined for
autofluorescence.

165 Newly settled and other juvenile O. doederleini between 16.2 and 32.0 mm SL were collected 166 from isolated patch reefs. Fish were collected with a small hand net and transferred to buckets filled with 167 seawater for transport back to the laboratory. They were immersed in tetracycline-treated seawater (0.25 168 $g L^{-1}$) for 18 hours in the laboratory in order to mark their otoliths (higher concentrations killed small 169 fish). Fish were transferred to 25 L buckets with one mm mesh windows in the lagoon after removal 170 from the tetracycline solution. Treated fish, therefore, were exposed to the natural cycle of day and night 171 and to normal levels of salinity and water temperature. Fish were fed on wild zooplankton and Artemia 172 nauplii that were reared in the laboratory. Fish of a range of sizes were euthanased after five, 10 and 15 173 d; fish were then preserved in 98% ethanol. Sagittal otoliths were removed, processed and viewed under 174 transmitted and fluorescent light using a compound microscope. Under fluorescent light an ocular 175 marker was aligned with the fluorescent band if present. The otolith was then viewed under natural light 176 and increments counted between the ocular marker and the otolith margin. Processed otoliths from a 177 similar size range of untreated O. doederleini were also examined for the presence of auto fluorescence. 178 All otoliths were examined without knowledge of size or treatment.

179

180 Spacing of otolith increments

181 Most reef fish have a mark in otoliths that corresponds with the time of settlement (Brothers et al. 1983).

182 Settlement marks are an abrupt change in increment spacing, usually from wide to narrow (Victor 1986;

183 Kingsford et al. 2011). Our initial observation of otoliths in recently settled fish failed to detect a

184 settlement mark. We tested this more critically by measuring the spacing of increments in eight

randomly selected juvenile *O. doederleini* to determine how the increment pattern changed with distance from the primordium. Transverse sections of the sagittal otolith were prepared. In the first group of fish increment widths were measured from the primordium to the otolith margin (n = 4 fish) and in the second group of fish, from sulcal groove along the longest axis of the otolith to the ventral edge (n = 4fish) were measured. Increment widths were measured using a calibrated video-analysis system (Leica IM50) linked to a compound microscope (Leica DMLB).

191

192 Age and Growth

Growth was described using the von Bertalanffy growth function (VBGF) which provided the best fit to size-at-age data when compared with estimates of the Schnute growth function (Schnute 1981). The von Bertalanffy expression for length at age t (L_t), as a function of time was calculated using equation (1):

196
$$L_t = L_{\infty} [1 - e^{-K(t-t_0)}]$$
(1)

where L_t = length at age *t* (*days*), L_{∞} = the mean asymptotic standard length, *K* = the rate at which the growth curve approaches L_{∞} , and t_0 = the age at which the fish have a theoretical length of zero. The growth function was run by solver (Microsoft Excel) which minimizes the sum of squares by changing L_{∞} , *K* and t_0 . Some of our demographic comparisons use 'Age max' = maximum age of an individual within a sample; and adapted from the convention of Choat and Robertson (2002), max 5% refers to the average age of fish from the 95th to 100th percentile, max 10% the average age from the 90th to the 100th percentile of the sample.

206 Mortality

207 The instantaneous rate of mortality (Z) was calculated using log-linear regression analyses of age-

208 frequency data sets for *O. doederleini* populations from two reefs where all fish from new recruits to

209 large adults were collected. This method assumes that recruitment is consistent over time at each reef

210 (Haddon 2001). Year classes to the left of the age-frequency mode were excluded from the analysis

211 because our sampling technique was biased against small fish. Mortality was only estimated where fish

212 (over 45 mm SL) were collected representatively (i.e. Heron Island and Lamont reef). The slope of the

213 regression line for each reef provided an estimate of daily instantaneous mortality (*Z*); equation (2),

when expressed as a percentage = loss of individuals per day.

where *F* is fishing mortality and *M* is natural mortality (Gust et al. 2002). Since there is not a fishery for *O. doederleini* on the GBR, *F* equals zero and therefore *Z* estimates natural mortality only. Daily
survival rate estimates (S) were then calculated (i.e. percentage surviving per day, Ricker 1975)
according to the equation (3).

220

221 $S = e^{-Z}$ (3)

222

Data from each site were pooled for each reef. Similarities in mortality rates among replicate reefsallowed comparisons of mortality between reefs.

225 Results

226 Validation of aging

227 The otoliths of most *Ostorhinchus doederleini* (75%) immersed in tetracycline showed a bright

228 fluorescent band close to the otolith edge (Fig. 2b). Twenty untreated O. doederleini otoliths were

229 checked for autofluorescence and showed no fluorescent band (n = 20). There was close agreement

230 between the number of increments laid down from the time of immersion in tetracycline and known

number of days since treatment; this was true of fish sampled at 5, 10 and 15 days (Table 2).

232 Settlement marks were not found in O. doederleini regardless of the orientation of counts with 233 respect to the otolith primordium (Fig. 3a & c). There were no rapid changes in increment width that 234 could be attributed to settlement. The approximate age at settlement could be estimated from the average 235 age that potential settlers entered the lagoon to settle (Fig. 3a & b, Table 3). Although increment width 236 varied across the otolith, they did not relate to settlement. Increments close to the primordium ranged 237 from 2 to 4 μ m in width and at approximately the 5 to 8 day mark increments rapidly became wider (5 to 238 7 μ m); Fig. 3a. The rapid increase in increment widths at the 5 to 8 day mark was not related to 239 settlement as the mean number of increments in O. doederleini caught just prior to settlement was 19.5 240 (SE = 0.38, n = 34; Table 3). Similar patterns were found in separate profiles, but increments were

241 generally wider (5 to $18 \mu m$) across the profile (Fig.3b).

242

Age and size of presettlement and recently settled Ostorhinchus doederleini

244 The age of presettlement *O. doederleini* caught in the channel nets ranged from 15.7 to 26.0 days (mean

245 = 19.9; n= 34) and SL ranged from 7.5 to 13.8 mm (mean 10.7; n=34); Table 3. The age of recently

246 settled *O. doederleini* collected from experimental patch reefs ranged from 16 to 42 days (mean= 25<u>.0</u>;

n = 232) and SL ranged from 8.6 - 18.0 mm (mean 12.7; n = 677); Table 3.

There was great overlap in the age and size of presettlement fish entering the lagoon and that of recently settled fish. Youngest fish on patch reefs were 16 to 17 days old and were as small as 8.6 mm SL. Older and larger fish were found on patch reefs, but many of them would have been associated with the reef for days as some recently settled fish move between patch reefs (Finn and Kingsford 1996). There was a strong linear relationship between age and standard length in presettlement fish over the first 20 days and recently settled fish and growth was rapid at about 0.<u>35-26</u> mm per day (Fig. 4).

255 Variation in growth and sizes among reefs

256 The maximum age of O. doederleini among all reefs was 368 days old and at most reefs few fish were 257 older than 200 days old (Fig. 5; Table 4). Time of sampling did not affect estimates of age maxima. The 258 oldest fish caught in Januaryto February at One Tree Island was 284 days and 59 mm long (SL), while in 259 October it was 263 days and 62 mm long (SL). When the average age of the top 5 and 10% of fish was 260 considered at each reef, fish were 150250 days old (Table 4). All von Bertalanffy relationships were 261 well anchored with small fish around 15to 20 mm SL. Best estimates of asymptotic length were 262 obtained from One Tree Island, where fish grew little in length from 150to 200 days old. Although there 263 were no significant differences in the size of fish among reefs in the age range of 90to 110 days 264 (ANOVA, $F_{(3, 60)} = 0.568$, P = 0.63), the maximum size of fish did vary. By reef the average size for the 265 top 5% of fish and the maximum SL was (average, max): One Tree Island (61.0, 66 mm), Heron (57.0, 266 65 mm); Lamont (62.6, 74 mm) and Fitzroy (65.5, 68 mm). The largest fish caught at Heron Island, 267 One Tree Island and Fitzroy reef were 65to 70 mm SL (= ~80to 85 Total Length). The two largest and 268 oldest fish ever collected (days/SL, mm: 246/69, 368/74) were from Lamont reef, January 2013. The 269 mean and range of lengths (SL) and ages differed little between males and females (Table 5).

271 Mortality

272 Instantanous mortality rates calculated from catch-curves indicated that mortality rates were high for

273 Lamont and Fitzroy reefs(2.9 to 4.6 % per day, Fig.6). Further, mortality rates were highest at Lamont

where large fish were rare. Survival (S) rates ranged from 0.971 to 0.955 d¹.

275 **Discussion**

276 Ostorhinchus doederleini had a short PLD and once they settled, grew fast and died young. High

277 mortality rates within days of settlement are common for reef fishes (Sale and Ferrell 1988) and low age

278 maxima have been found in other apogonids (Longenecker and Langston 2006). Our study has

demonstrated that O. doederleini has a high mortality (2.9 to 4.6% per day) and a low age-max. Ages of

280 one year or less clearly put these tropical apogonids at the lower end of the longevity spectrum for

tropical fishes along with the Gobiidae and Blenniidae (Table 6). Although some taxa such as gobies

may live for less than a year and have high mortality rates (e.g. *Eviota silligata* has an age max of 59

283 days, mortality rate 7.8% per day; Depczynski and Bellwood 2005), species from families of common

reef fishes such as the Pomacentridae (e.g. Doherty and Fowler 1994; Kingsford and Hughes 2005),

Labridae (Choat et al. 2006), Scaridae (Choat et al. 1996; Gust et al. 2002), Serranidae, Lutjanidae

286 (Kingsford 2009) and Acanthuridae (Choat and Axe 1996) live for years and sometimes decades

287 (Munday and Jones 1998; Fowler 2009) Table 6. Fishes with the lowest age-max were generally small

and represented three different trophic groups: nocturnal planktivores, micro/meso carnivores and

289 herbivores/detrital feeders. Although short-lived reef fish appear to be small, not all small fish are short

290 lived (e.g. *Pomacentrus mollucensis* SL max 90 mm (TL) and age max 18 years; Doherty and Fowler

291 1994).

To the best of our knowledge, this is the first study to report information on the growth rates of *O*. *doederleini* prior to settlement and immediately after settlement. Further, it is the first to provide data

on longevity (age maxima) of the species in the tropics.. Some families had data on multiple stages of life 294 295 history when multiple species were pooled (e.g. omnivorous/herbivorous damselfishes), but given the 296 differences in demographic characteristics within a family general patterns would be crude or simply 297 inaccurate. There was an abundance of data on age, growth and mortality of representatives of families 298 that are the focus of fisheries, especially macrocarnivores and piscivores such as lutianids, serranids and 299 carangids. However, data on presettlement growth, PLDs and post-settlement growth was 300 comparatively rare. In other families the focus has only, or primarily, been on early life history stages 301 (e.g. Holocentridae, Pomacentridae). For many important families, we found no data at all on age and 302 growth (e.g. Holocentridae, Pempheridae, Caesionidae and Pomacanthidae). The paucity of data is due 303 in part to a relatively recent understanding that the otoliths of tropical species can provide reliable 304 estimates of age (Meekan et al. 2001). Despite this there is a clear need to increase knowledge in this 305 area for the management reef assemblages and tropical fisheries

306

We concluded that accurate estimates of age could be obtained from daily increments. Evidence that counts were accurate included the following: the technique was validated, daily increments were easily observed (Fig. 2), the asymptote for age/length relationship for all reefs was beyond 200 days and, adult fish were collected from a size of 49 mm and 132 days or older (based on brooding fish and macroscopic examination of gonads).

312

The oldest fish we found was 368 days (n=581); at 23°35" S) and, unpublished data by Michael Marnane at One Tree Island also indicated that some *O. doederleini* can reach an age of over one year. Similar to our study, Longenecker and Langston (2006) found that *Ostorhinchus rubrimacula* (Apogonidae) lived to a maximum of 280 days in Fiji (17°15" S). However, there are great differences

317 between our results for O. doederleini, and other data from temperate waters in Japan. Okuda et al. 318 (1998) concluded age estimates of up to seven years in at Shikoku Island, Japan (33° 40" N). The fish 319 sampled in Japan grew to a maximum SL of 86 mm SL whereas we only found fish to a size of 74 mm 320 (93 mm TL). The maximum size of O. doederleini recorded for the GBR is 90 mm TL ~ 74 mm SL, but 321 no location is given by Randall et al. (1990) for this specimen. Okuda et al. (1998) had few data on fish 322 less than one year for us to make comparisons of age versus a fish of known size. There are three 323 possibilities to explain latitudinal variation in O. doederleini age and growth: (1) there are great 324 differences in age over 10 degrees of latitude, which is partly reflected by differences in size max; (2) a 325 methodological issue where the accuracy of aging techniques varies; Okuda et al. (1998) used scales, 326 whereas we used daily increments in otoliths; (3) mortality rates vary greatly between latitudes. With 327 respect to options one and two, we favour option one as latitudinal variation in growth and age maxima 328 are common in fishes (growth, Conover et al. 1997; age-max, Boehlert and Kappenman, 1980) 329 Furthermore, scales tend to underestimate not overestimate ages. In Japan Okuda et al. (1998) fin 330 clipped some fish that were monitored for up to two years, so they clearly get older than their tropical 331 conspecifics. The apogonid Apogon ruepillii found in estuaries near Perth (~ 32°S) were over a year old 332 (Chrystal et al. 1985). All of the evidence from tropical waters indicates that small O. doederleini have 333 age maxima of a year or less. Estimates of L_{∞} would be most reliable for all reefs combined (Table 4) 334 as very large fish were rare. Our comparisons of the size/age of fish among reefs in the 80to 90 mm SL 335 range would be robust, but beyond 200 days would be weak. Latitudinal gradients in growth and life 336 span would be interesting to examine. Conover et al. (1997), for example, demonstrated a strong 337 positive correlation between the latitude of origin of *Morone saxatilis* and growth. Further, it is 338 common for there to be positive relationships between life-span and latitude in ectotherms, including 339 fishes (Boehlert and Kappenman, 1980; review, Munch and Salinas 2009). At present there are no

340 comparative data on growth by latitude for O. doederleini. The presettlement phase of O. doederleini 341 was 16 to 26 days and fish settled at 9.5 to 13 mm SL. Settlement marks were not observed and the age 342 of settling fish was determined from potential settlers collected in nets at night and new arrivals to patch 343 reefs. Ishihara and Tachihara (2011) estimated the PLDs of six O. doederleini in Japan at ~ 16to 24 344 days and 10to 11.5 mm SL, which fell within the ranges of values we had from the southern GBR; these 345 authors also collected fish in channel nets. The range of ages over which fish settled was 38% of the 346 maximum age at settlement, a little less than that recorded for the damselfish Pomacentrus coelestis 347 (45%; Kingsford et al. 2011). Although many damselfishes spend a similar time in the plankton to 348 O. doederleini, there are many other taxa that have longer PLDs. For example, scarids have PLDs of 34 349 to 58 days; chaetodonts 25 to 40 and acanthurids 80+ days (Brothers et al. 1983).

Ostorhinchus doederleini have strong olfactory senses and can respond to the presence of
lagoonal waters (Atema et al. 2002). Presettlement fish swim and/or are carried over the reef crest at
night (Kingsford 2001) into One Tree lagoon where they find suitable settlement habitat to settle.
Ostorhinchus doederleini commonly settle away from continuous reef on sections of substratum,
potentially to avoid predatory parents (Finn and Kingsford 1996). We demonstrated that during this
phase fish had linear growth, this is typical of other fishes during the early juvenile phase (Kingsford &

356 Milicich).

Although reef fishes often have a mark in their otoliths related to settlement from pelagic to benthic environments (Brothers et al. 1983; Kingsford et al. 2011), 'settlement marks' were absent in the otoliths of *O. doederleini*. Brothers et al. (1983) also concluded that *O. doederleini* settle at 16to 27 days (n=5) and did not mention a settlement mark (='transition'). Settlement marks are not found in all fishes and, it has been speculated that the eco-behavioral transitions are less traumatic in these

362 circumstances (e.g. monacanthids settling from drift algae to algae attached to the substratum, Kingsford

and Milicich 1987). Ostorhinchus doederleini may experience less post-settlement trauma by not
settling on continuous reef, where the risk of predation (and stress) is high. Further, they usually settle
with conspecifics (pers. obs.) and they already have an established ability to feed in low light (Job and
Bellwood 2000).

367 There is strong evidence that population structure within a metapopulation of O. doederleini can 368 be complex at spatial scales of kilometers to tens of kilometers. Despite having a PLD of 16 to 26 days, 369 a combination of favorable oceanography and larval behavior means that a disproportionately high 370 number of fish return to natal reefs and this can result in significant genetic differences among reefs 371 (Gerlach et al. 2007). The robustness of population structures through time is likely to depend on larval 372 survival, physical forcing (Sissenwine 1984), post-settlement processes (Jones 1991; Almany and 373 Webster 2006) and demographic characteristics. It is critical to consider age for numerical modeling of 374 ecological variation in population size and genetic robustness. For short-lived species such as 375 O. doederleini, there is a great vulnerability to recruitment failure, especially if they do not have a 376 prolonged spawning season. This in turn could compromise the resistance and resilience of a local 377 genotype to change. Information on the timing and duration of spawning of O. doederleini is largely 378 anecdotal, but does suggest a prolonged spawning season. Mouth brooding fish have been observed in 379 October and January-February and peak recruitment is in January to February (i.e. from spawning in 380 December and January), Further, the collection of O. doederleini larvae, in June by Kingsford (2001) 381 indicates that at least some spawning occurs in winter and backdating the age of fish collected in 382 January and February puts many birthdates in the Austral winter to spring.

In conclusion, *O. doederleini* have a short PLD, they settle onto sand and rubble away from continuous reefs and grow quickly. They reach asymptotic size beyond 200 days, but few fish reach 200 days old and the age maximum we found at four reefs was 368 days. These small fish were

- 386 characterized by high mortality rates and presumably have a high population turn over; this should be
- 387 considered in models relating to the trophodynamics of reefs and the population biology of
- 388 *O. doederleini.* A review of the literature indicated that other fishes with similar demographic
- 389 characteristics are small gobies and blennies.
- 390

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- 396

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Table 1 Collection details for *Ostorhinchus doederleini* specimens, 1994 to 2013. OTI- One Tree Island;

616 H- Heron Reef; F- Fitzroy Reef; and L= Lamont.

Dates Sampled	Methods used	Reefs	Life stage
Jan 1994	Channel nets; Patch Reefs	OTI	Presettlement and
			early post-settlement
Feb 1995	Channel nets; Patch Reefs	OTI	Presettlement and
			early post-settlement
Feb 2009	Channel nets, Hand nets;	OTI, H, F, L	Juveniles and adults
	clove oil		
Jan-Feb. 2010	Hand nets; clove oil	OTI, H, F, L	Juveniles and adults
Oct 2010	Hand nets; clove oil	OTI	Juveniles and adults
Jan 2013	Hand nets; clove oil	L	Adults

Table 2 Results of validation marking experiments for *Ostorhinchus doederleini*. No. of days = number
of days since fish were immersed in tetracycline.

No. of days	п	SL range (mm)	Mean of counts (SE)
5	42	17.0 - 25.2	4.95 (0.07)
10	16	17.4 - 34.0	10.06 (0.11)
15	11	16.2 - 25.0	14.82 (0.23)

Table 3 Age (days) and size (mm; SL) of presettlement and recently settled *Ostorhinchus doederleini* in
two recruitment seasons. Data shown are the mean (range) and total n is given by row. For recently
settled fish n varied for size and age, more fish were collected for measurements of size and were not
preserved in ethanol (Kingsford and Finn 1997).

	1994	1995	п
Presettlement fish			
SL	10.7 (7.5 - 13.0) 4	10.6 (9.0 - 13.8) 30	34
Age	18 (15.7 - 19.7) 4	19.7 (16.0 - 26.0) 30	34
Recently settled fish			
SL	14.1 (9.8 - 18.0) 299	11.23 (8.6 - 18.0) 378	677
Age	28.2 (17.0 - 41.5) 113	22.5 (16.0 - 40.0) 119	232

Table 4 von Bertalanffy growth parameters for four reefs and estimates of age maxima for individual
fish from samples at each location and average age of the oldest five and 10 percent of fish (i.e. Max %).

Reef	n	Γ∞	Κ	r^2	Age Max	Max 5% (n) SE	Max10% (n) SE
Heron	122	76.24	0.008	0.86	215	173 (6) 13	144 (12) 11
One Tree Island	257	60.76	0.012	0.88	284	255 (13) 4	240 (26) 4
Lamont	91	83.84	0.006	0.82	368	213 (5) 6	176 (9) 27
Fitzroy	123	70.48	0.009	0.77	282	224 (6) 12	207 (12) 8
All reefs	593	65.04	0.010	0.89	368	249 (30) 5	226 (59) 4
635							

Table 5 Size, age and sex differences for *Ostorhinus doederleini* collected at One Tree Island, October
2010.

	Female	Male
n	55	70
Mean (SL)	53.27 mm	52.09 mm
SE	0.535	0.596
Min (SL)	43 mm	41 mm
Max (SL)	60 mm	66 mm
Mean (Age)	191 days	190 days
Min (Age)	139 days	132 days
Max (Age)	258 days	263 days

Table 6 A comparison of with examples of demographic characteristics among multiple families of tropical reef fishes, by trophic groups (adapted from Randall et al. 1990); in some cases we found more references than listed, but the age range was within the other publication listed. A Yes (Y) indicates that some information is available; ND- no data, y- years, d- days. Brothers et al. (1983) provided estimates of PLD for many of the groups listed, but they relied on the presence of settlement marks which are not found in some families such as apogonids. Some families appear in more than one trophic group (e.g. Pomacentridae). Data from temperate regions are not included.

Family	Species	Presettlement growth	PLD (range)	Settlement mark	Early Post-settlement growth	Life time growth	Age maxima	Estimates of mortality	Country	Source
Nocturnal plank	tivores/piscivores									
Apogonidae	Apogon cyanosoma	Ν	18-19d	ND	ND	ND	ND	ND	GBR, Australia	Brothers et al. 1983
	Cheilodipterus quinquelineatus	Ν	17-29d	ND	ND	ND	ND	ND	GBR, Australia	Brothers et al. 1983
	Ostorhinchus doederleini	Y	16-26d	ND	Y	Y	< 1 year	Y	GBR, Australia	This study
	Ostorhinchus rubrimacula	Ν	Ν	ND	Y	Y	< 1 year	ND	Fiji	Longenecker and Langston 2006
Holocentridae	Multiple (7)	Y	32-56d	ND	ND	ND	ND	ND	Western Atlantic	Tyler et al. 1993
Pempheridae		ND	ND	ND	ND	ND	ND	ND		
Diurnal plankti	vores									
Caesionidae		ND	ND	ND	ND	ND	ND	ND		
Pomacentridae	Multiple (8)	ND	8-32d	Y	Y	ND	ND	ND	GBR, Australia	Bay et al. 2006; Green and
Acanthuridae	Naso sp (6)	ND	ND	ND	ND	Y	9-30y	ND	GBR, Australia	Choat and Robertson 2002

Serranidae	e.g. Anthias spp. and Pseudoanthias spp	ND	ND	ND	ND	ND	ND	ND		
Micro-Meso car	nivores									
Balistidae	Multiple (2)	ND	ND	ND	ND	Y	11-14y	Y	Brazil	de Albuquerque et al. 2011;
Chaetodontidae	Multiple (8)	Y	23-45d	Y	Y	Y	6-20y	Y	GBR, Australia	Bernardes 2002 Fowler 1989; Berumen 2005; Berumen et al.
Gobiidae	Multiple (3)	ND	24-26d	Y	Y	Y	59-100d	Y	GBR, Australia	2012 Depczynski and Bellwood 2006
	Coryphopterus kuna	ND	45-74d	Y	Y	Y	120d	Y	Caribbean	Victor et al. 2010
	Eviota sigillata	ND	23-27d	Y	Y	Y	59d	Y	GBR, Australia	Depczynski and Pollwood 2005
Labridae	Multiple (9)	ND	21-50d	Y	ND	Y	15-36y	Y	GBR, Australia	Brothers et al. 1983;
	Thalassoma bifasciatum	Y	40-72d	Y	Y	ND	ND	ND	Caribbean	Victor 1982
Nemipteridae	Scolopsis bilineatus	ND	ND	ND	ND	Y	16	Y	GBR, Australia	Boaden and Kingsford 2012
Pomacanthidae	Pomacanthus imperator	ND	ND	ND	ND	Y	16	ND	China	Chung and Woo 1999
Macrocarnivore/ J	piscivore									
Carangidae	Multiple (2)	ND	ND	ND	ND	Y	6-10y	ND	Hawaii	Sudekum et al. 1991
	Caranx bucculentus	ND	ND	ND	ND	Y	ND	Y	Northern Australia	Brewer et al.
Lethrinidae	Multiple (7)	ND	ND	ND	ND	Y	15-36y	Y	GBR, Australia	Currey et al. 2013; Brown and Sumpton 1998
Lutjanidae	Multiple (11)	ND	ND	ND	ND	Y	12-55y	Y	Northern Australia, Indonesia	Marriott et al. 2007; Newman et al. 1996; Newman et al. 2000; Heupel et al. 2010
	Lutjanus carponotatus	ND	ND	ND	ND	Y	18y	Y	GBR, Australia	Kingsford 2009
Scorpaenidae	Pterois volitans	ND	20-35d	ND	ND	ND	ND	ND	Bahamas	Ahrenholz and Morris 2010
Serranidae	Cephalopholis cvanostisma	ND	ND	ND	ND	Y	45y	Y	GBR, Australia	Mosse et al. 2002
	C 11 11 11	ND		ND	ND	37	10	37	CDD A / I'	*****

	Epinephelus fuscoauttatus	ND	ND	ND	ND	Y	42y	Y	GBR, Australia	Pears et al. 2006
	Plectropomus laevis	ND	ND	ND	ND	Y	16y	Y	GBR, Australia	Heupel et al. 2010
	Plectropomus leopardus	ND	ND	ND	ND	Y	15y	Y	GBR, Australia	Kingsford 2009
Omnivores/herb	oivores/ scrapers and detrital	feeder								
Acanthuridae	Multiple (5)	ND	84d	Y	ND	Y	21-70y	ND	GBR, Australia	Choat and Axe 1996
Blenniidae	Multiple (3)	ND	12-33d	Y	ND	ND	ND	ND	GBR, Australia	Beldade et al. 2007; Brothers et
	Salarias patzneri	ND	ND	Y	Y	ND	350d	ND	GBR, Australia	Wilson 2004
Pomacentridae	Multiple (100)	Y	12-39d	Y	ND	ND	ND	ND	GBR, Australia; 'acific, Atlantic	Bay et al. 2006; Wellington and Victor 1989
	Acanthochromis	ND	ND	Y	Y	Ν	9-11y	Y	GBR, Australia	Kingsford and Hughes 2005
	Pomacentrus	Y	15-23d	ND	ND	ND	ND	ND	GBR, Australia	Kerrigan 1996
	amboinensis									
	Pomacentrus coelestis	Y	15-27d	Y	Y	ND	ND	ND	GBR, Australia	Kingsford et al. 2011
	Pomacentrus	ND	ND	ND	Y	Y	18y	Y	GBR, Australia	Doherty and Fowler 1994
	moluccensis									
	Stegastes acapulcoensis	ND	ND	ND	ND	Y	12-32y	Y	Panama, Galapagos	Meekan et al. 2001
Scaridae	Multiple (7)	Y	30-50d	Y	ND	Y	5-20y	Y	GBR, Australia	Brothers et al. 1983; Gust et al. 2002; Choat et al. 1996
	Sparisoma viride	ND	ND	ND	ND	Y	7-9y	Y	Caribbean	Choat et al. 2003

646 Figure Captions

647

- Fig. 1 Map showing the location of the four reefs in the Capricorn-Bunker Group, southern Great
 Barrier Reef, Australia where *Ostorhinchus doederleini* was collected.
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- **Fig. 2** (a) Sagittal otolith section of a 21 day old *Ostorhinchus doederleini*. Width of otolith section is
- 652 1500 μm. Initial measurement made at first visible increment (Insert showing enlargement of
- 653 300 μm long); (b) Otolith of a fish euthanised 10 days after immersion in tetracycline viewed under UV
- light, scale bar = $100 \mu m$; (c) A section of an 90 day old fish (from OTI; 34 mm SL) showing track used
- for aging, which extended from the primordium to the outer edge of the otolith along the longest axis
- 656 (Section is $2100 \,\mu m \log$).
- 657

Fig. 3 Otolith increment widths plotted as a function of increment number for a subset of four post-

- 659 settlement Ostorhinchus doederleini, (a) 1994 and 1995; (b) 2010 from One Tree Island, Great Barrier
- 660 Reef, Australia. The dotted vertical line indicates the mean no. of increments deposited by presettlement
- apogonids caught just prior to settlement measured from the primordium to along the sulcal groove;
- 662 from the primordium to the ventral edge of the otolith. Initial measurement made at first visible
- 663 increment.
- 664

Fig. 4 Relationship between standard length and age (days) of presettlement versus newly settled
 recruits *Ostorhinchus doederleini* in 1994 and 1995 together with fitted regression lines and summary
 statistics.

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Fig. 5 von Bertalanffy growth curves for *Ostorhinchus doederleini* for the four reefs in the Capricorn
Bunker group, Great Barrier Reef, Australia.

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672 **Fig 6** Age-based 'catch-curves' that provide estimates of mortality for *Ostorhinchus doederleini* for

Heron and Lamont reefs in the Capricorn Bunker group, Great Barrier Reef, Australia; each data point
indicates Log_e frequency of each age class.







Increment Width (µm)







Loge Frequency