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## HORMONE SIGNALLING

# A novel gibberellin promotes seedling establishment

A previously unknown biologically active gibberellin present in seeds of *Arabidopsis thaliana* is formed by hydration of the gibberellin precursor GA<sub>12</sub> through the action of GAS2, a 2-oxoglutarate-dependent dioxygenase that decreases sensitivity to abscisic acid and promotes seed germination and seedling establishment.

### Peter Hedden

he gibberellin (GA) group of plant hormones promote growth and other developmental processes, including seed germination. While there are currently 136 naturally occurring compounds that are classified as GAs on the basis of their structure, only a relatively small number of these possess biological activity and are involved in the regulation of plant development. For a GA to be biologically active it was thought to require certain structural characteristics<sup>1-3</sup>, as exemplified by GA<sub>4</sub>, which is the main biologically active GA in Arabidopsis (Fig. 1). All bioactive GAs possess a C<sub>19</sub> skeleton (containing 19 carbon atoms), which is formed biosynthetically from C<sub>20</sub> precursors, including GA<sub>12</sub> (Fig. 1), that were thought to possess no intrinsic biological activity. Now in a paper published in Nature Communications, Liu et al.<sup>4</sup> show that a simple derivative of GA<sub>12</sub>, formed by hydration of its double bond and with seemingly little structural similarity to known biologically active GAs, has GA-like activity. The compound, which was detected in Arabidopsis and maize seeds, was chemically characterized as 16,17-dihydroGA<sub>12</sub> 16α-ol (DHGA<sub>12</sub>) (Fig. 1). It stimulated hypocotyl elongation when applied to Arabidopsis seedlings grown in far-red light and promoted cotyledon greening in the presence of the hormone abscisic acid (ABA), although in both cases with lower activity than GA<sub>4</sub>. Furthermore, the authors used microscale thermophoresis to show that DHGA<sub>12</sub> binds to a GA receptor (GA INSENSITIVE DWARF1C) and molecular docking with a receptor model to suggest how this interaction occurs. The authors propose a role for DHGA<sub>12</sub> in promoting seed germination and early seedling growth.

Seed germination is regulated by a balance between GA and ABA, which act antagonistically<sup>5</sup>. The authors used a chemical activation screen for genes that overcome the inhibitory effect of ABA on *Arabidopsis* 

seed germination. The screen for promotion of germination in the presence of ABA utilized transgenic Arabidopsis plants with transfer DNA (T-DNA) insertions containing an estradiol-inducible promoter, such that expression of genes with insertion of the T-DNA in their promoters is activated by application of the chemical6. Expression of one of the induced genes, namely GAIN-**OF-FUNCTION IN ABA-MODULATED** SEED GERMINATION 2 (GAS2), resulted in reduced sensitivity to ABA. The gene is expressed in roots and leaves, induced by light and ABA, and suppressed by treatment with GA4. Overexpression of GAS2 promoted seed germination and early seedling development in the presence of ABA, while germination was delayed in non-chemically induced plants - in which GAS2 is not expressed — or plants containing mutations in GAS2 produced by genome editing. GAS2 was found to encode a 2-oxoglutaratedependent dioxygenase (20DD) of previously unknown function. Due to the antagonistic action of GAs with respect to ABA and the known involvement of 20DDs in GA biosynthesis<sup>7</sup>, the authors suspected that GAS2 may have a role in GA metabolism, which they tested using enzyme assays with GAS2 protein produced in Escherichia coli. They found that GAS2 acted on GA<sub>12</sub>, but surprisingly, it did not catalyse the expected oxidation but instead hydrated the C-16,17 double bond, a reaction not previously associated with 20DD activity. Consistent with this activity, plants in which GAS2 was overexpressed contained higher amounts of DHGA<sub>12</sub> in imbibed seeds compared with wild-type seeds, whereas it was not detectable in seeds from non-induced T-DNA plants. Compared with wild-type seeds, those from the overexpression line contained lower amounts of GA12 and also of GA4, a later product of the biosynthetic pathway. While the reduction of GA<sub>4</sub> can be partly explained by diversion of its precursor from the main biosynthetic pathway, another cause appears

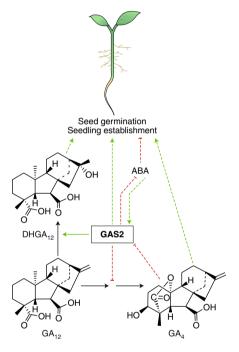


Fig. 1 | GAS2 reduces sensitivity to ABA, promoting seed germination and seedling establishment. GAS2 encodes a 2-oxoglutaratedependent dioxygenase which was shown in vitro to hydrate the double bond of the GA precursor GA<sub>12</sub> to form the previously unidentified DHGA<sub>12</sub>. Application of DHGA<sub>12</sub> promoted seed germination and seedling establishment in the presence of ABA, although less strongly than GA<sub>4</sub>. Imbibed seeds of plants overexpressing GAS2 accumulated DHGA<sub>12</sub>, but contained reduced levels of GA<sub>12</sub>, GA<sub>4</sub> and ABA. Expression of GAS2, which was strongest in the roots and leaves, was promoted by exogenous ABA and repressed by GA<sub>4</sub>. Green arrows indicate promotive activities, while red barheaded lines indicate repression. Continuous lines indicate enzymatically catalysed reactions, while dashed lines indicate regulatory activities.

to be reduced expression of a GA 20-oxidase gene, which is required for  $GA_4$  synthesis, in the *GAS2* overexpression line.

These are interesting and novel findings, showing a clear involvement of GAS2 in promoting seed germination. However, they may not tell the whole story. Of particular note is the strong reduction in the content of the highly active GA<sub>4</sub> in seeds of the overexpression line, while there is a smaller increase in the level of the less active DHGA<sub>12</sub>. This is inconsistent with the ability of GAS2 to promote seed germination, for which the observed reduction in ABA levels in the GAS2 overexpression line must be relevant. Furthermore, cotyledon greening and hypocotyl elongation could be only partially recovered in the gas2 mutant lines by application of DHGA<sub>12</sub> or GA<sub>4</sub>, indicating the involvement of other factors. The altered expression of genes involved in ABA and GA metabolism in the GAS2 overexpression and genesilenced lines suggests that GAS2 may have a regulatory role, perhaps through

formation of an unidentified product. The mechanism by which GAS2 catalyses the hydration of GA<sub>12</sub> is unclear and this may not be its only activity. Could it also have bona fide oxygenase activity against an unknown substrate? Detailed comparison of metabolites in the lines with altered GAS2 expression could prove helpful in identifying potential substrates and products. Nevertheless, the demonstration that DHGA12 has GA-like activity, despite its apparent dissimilarity from the canonical structure thought to be essential for activity, is a surprising and unexpected finding that will be of considerable interest to researchers in the plant hormone field. 

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#### **Competing interests**

The author declares no competing interests.

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