Javdan *et al. BMC Res Notes* (2019) 12:363 https://doi.org/10.1186/s13104-019-4396-8

BMC Research Notes

RESEARCH NOTE Open Access

Check for

Agr typing of *Staphylococcus aureus* species isolated from clinical samples in training hospitals of Isfahan and Shahrekord

Saeid Javdan¹, Tahmine Narimani², Milad Shahini Shams Abadi³ and Abolfazl Gholipour^{1,4*}

Abstract

Objective: As an opportunistic pathogen, *Staphylococcus aureus* is associated with serious nosocomial infections and growing antimicrobial resistance against beta-lactams among *S. aureus* strains has become a global challenge. The current study was designed to investigate the presence of *agr* genes among *S. aureus* strains recovered from clinical samples in university hospitals of Isfahan and Shahrekord.

Results: A total of 150 *S. aureus* isolates were screened by Disk diffusion method (DDM) and conventional PCR. The minimum (17.3%) and maximum (46%) antibiotic resistance rates were found in vancomycin and cefoxitin, respectively. The majority of our isolates were classified as agr type I followed by type II, type IV, and type III. The statistical analysis showed a significant correlation between agr type I and antibiotic resistance against cefoxitin and erythromycin (p=0.04 and p=0.03, respectively). Based on our findings, the agr typing could be considered an effective approach for molecular tracking of *S. aureus* infections.

Keywords: Staphylococcus aureus, Agr type, mecA gene, Antibiotic resistance, Methicillin

Introduction

As a part of microflora of skin and mucous membranes of healthy individuals, *Staphylococcus aureus* is also an opportunistic pathogen and associated with hospital acquired infections such as septicemia, pneumonia, septic arthritis, osteomyelitis, toxic shock syndrome after surgery, folliculitis, endocarditis, and urinary tract infections (UTIs) [1, 2]. Antibiotic resistance by affecting more than two million people annually is one of the biggest global challenges. The increasing antimicrobial resistance among *S. aureus* species against beta-lactam antibiotics has led to serious problems with the treatment of their related infections. Despite considerable efforts in controlling antibiotic resistance, methicillin-resistance *S. aureus* (MRSA) is raising worldwide, in addition geographical and local variations influence its dynamic and

crisis [1, 3]. The methicillin resistance development in S. aureus is related to the several Staphylococcal Cassette chromosome mec elements (SCCmec) encoding mecA gene for a penicillin binding protein (PBP2a) [4]. MRSA strains are usually multi-drug resistant (MDR) and show resistancy to other antibiotics such tetracyclines, aminoglycosides, lincosamides etc. [1, 5, 6]. Rapid and precise typing of S. aureus is really crucial to transmission identification of this pathogen. In this regard, Pulsed-Field gel electrophoresis and spa typing (Staphylococcal protein A) are common typing methods. The spa gene is one of the most distinctive factors related to this organism, and various patterns of it have been identified by several studies [7]. One of the major regulatory and control factors in the virulence gene expression of *S. aureus* is the accessory gene regulatory (agr) system. Indeed, agr operon including agrA, agrB, agrC, and agrD genes regulate over 70 genes in S. aureus 23 of which control its pathogenicity and invasive infections [8]. Moreover, S. aureus can be stratified into 4 different groups (agr I, agr II, agr III, and agr IV) according to the sequences of agrC (auto inducing peptide) and agrD (cyclic AIP) genes. It is stated that

Full list of author information is available at the end of the article



^{*}Correspondence: gholipour_abolfazl@yahoo.com

⁴ Department of Microbiology and Immunology, Cellular and Molecular Research Center, Shahrekord University of Medical Sciences, Shahrekord,

Javdan et al. BMC Res Notes (2019) 12:363 Page 2 of 6

agr types are different in their properties and prevalence in various geographical areas thus, identification of predominant types in each region may well be functional [9].

Given to the critical roles of *agr* genes, the current study was designed to detect and identify the *agr* groups of *S. aureus* strains isolated from clinical samples in training hospitals of Isfahan and Shahrekord cities.

Main text

Materials and methods

Samples and bacterial isolates

This cross sectional study was conducted in microbiology department of Shahrekord University of Medical Sciences. During May to November 2017, a total of 150 isolates of *S. aureus* were collected from clinical samples (wound, blood, urine, tissue etc.) of patients attending university hospitals in Isfahan (Alzahra and Kashani) and Shahrekord (Kashani and Hajar).

Characterization assays

The isolates were identified using Gram staining, catalase test, slide or tube coagulase test, DNase test, and growth on Mannitol Salt Agar (MSA) as a differential growth medium [10].

Antibiogram testing

Disk diffusion method (DDM) as described by CLSI 2016 guideline [11] was performed for following antibiotics: erythromycin (15 mg), tetracycline (30 mg), vancomycin (30 mg), gentamicin (10 mg), rifampin (5 mg), cefoxitin (15 mg), trimethoprim (5 mg), rifampicin (5 mg). In addition, all isolates were subjected to cefoxitin disc diffusion test to identify the methicillin sensitive *S. aureus* (MSSA) and MRSA.

DNA extraction

The nucleic acids of *S. aureus* isolates were extracted by phenol chloroform method followed by RNase treatment [12]. The purity of extraction was assessed using the A260/280 ratio and agarose gel electrophoresis.

PCR amplification of the mecA gene

molecular detection of mecA gene was carried out according to the following condition: initial denaturation at 95 °C for 3 min followed by 33 cycles of denaturation at 94 °C for 1 min, annealing at 53 °C for 30 s, and extension at 72 °C for 1 min and final extension step at 72 °C for 6 min.

PCR detection of agr genes

PCR assay for amplification of *agr* genes was set as follows: hot start at 95 °C/6 min, 32 cycles of 94 °C/45 s, 60 °C/1 min, 72 °C/70 s and a final extension step of

72 °C/8 min. All reactions performed in duplicate and along with the negative control (water) and positive (previously known positive-PCR products) control. The final products were detected by electrophoresis on 1% agarose gel containing DNA safe stain (Sinagene, Iran) and the sizes of the PCR products were estimated by the migration pattern of a 100-bp DNA ladder (Sinagene, Iran).

Statistical analysis

Statistical analysis was performed using SPSS version 22. The chi-square test was used to calculate statistical significance (p < 0.05).

Results

Study population

150 *S. aureus* isolates were collected from patients attending training hospitals in Isfahan (110 isolates from Alzahra hospital) and Shahrekord (25 cases from Kashani hospital and15 isolates from Hajar hospital). The mean age of the participants was 47.6 years (SD: 21.5) and male/female ratio was 90/60. However, there was not any significant difference in sex and age of patients with *S. aureus* infection. *S. aureus* isolates were obtained from several clinical samples and different hospital wards.

Antibiotic susceptibility

According to our results the lowest (17.3%) and the highest (46%) antibiotic resistance rates were found in vancomycin and cefoxitin, respectively. In addition, MRSA strains were verified by PCR amplification of mecA gene. The antibiotic resistance distribution among different agr groups is shown in Table 1. The results of this study showed a significant correlation between agr type and antibiotic resistance against cefoxitin and erythromycin (p=0.04 and p=0.03, respectively).

agr typing

Molecular detection of 150 *S. aureus* isolates has indicated that agr type I was the predominant one (82/150) followed by type II (37/150), type IV (21/150), and type III (10/150) (Fig. 1). Table 2 is shown the frequency distribution of different agr types among different clinical samples.

Discussion

There is a dramatic increase in *S. aureus* infections, both with community-associated and hospital-acquired types, and development of antibiotic-resistant species, especially MRSA and vancomycin-resistant strains, is the major cause of the infections and further treatment complications [13]. Identification and typing of the isolates may imply a common source of infection; therefore, accurate analysis of these patterns can help to break the

 Table 1 The antibiotic resistance profiles among 4 different agr types

AGR	FOX		ш		-	_			NA VA		RP			MT			GM		
type	S (%)	R (%)	S (%)	S (%) I (%) R (%)	R (%)	(%) S	(%) I	R (%)	S (%)	R (%)	(%) S	1 (%) 1	(%)	(%) S	I (%)	R (%)	(%) I (%) S		R (%)
_	37 (45.1%)	45 (54.9%)	38 (46.3%)	21 (25.6%)	23 (28) 2	47 (57.3%)	7 (8.5%)	28 (34.1%)	70 (85.4%)	12 (14.6%)	49 (59.8%)	3 (3.7%)	30 (36.6%)	52 (63.4%)	6 (7.3%)	24 (29.3%)	37 (45.1%) 45 (54.9%) 38 (46.3%) 21 (25.6%) 23 (28) 47 (57.3%) 7 (8.5%) 28 (34.1%) 70 (85.4%) 12 (14.6%) 49 (59.8%) 3 (3.7%) 30 (36.6%) 52 (63.4%) 6 (7.3%) 24 (29.3%) 58 (70.7%) 2 (2.4%) 2 (2.4%) 22 (26.8%)	(2.4%) 2	2 (26.8%)
=	22 (59.5%) 15 (40.5%) 12 (32.4%) 5 (13.5%) 20 20 (54.1%)	15 (40.5%)	12 (32.4%)	5 (13.5%)	20 (54.1%)	20 (54.1%)	1 (2.7%)	16 (43.2%)	28 (75.7%)	9 (24.3%)	27 (73.0%)	2 (5.4%)	8 (21.6%)	18 (48.6%)	5 (13.5%)	14 (37.8%)	20 (54.1%) 1 (2.7%) 16 (43.2%) 28 (75.7%) 9 (24.3%) 27 (73.0%) 2 (5.4%) 8 (21.6%) 18 (48.6%) 5 (13.5%) 14 (37.8%) 23 (62.2%) 0		14 (37.8%)
=	7 (70%)	3 (30%) 3	3 (30%)	1 (10%) 6 (60%)	(%09) 9	0 (%02) 2	0	30 (30%)	8 (80%)	2 (20%)	8 (80%) 1 (10%)	1 (10%)	1 (10%)	4 (40%)	1 (10%)	5 (50%)	7 (70%) 1 (10%)	(10%)	2 (2%)
≥	15 (71.4%)	6 (28.6%)	15 (71.4%) 6 (28.6%) 6 (28.6%) 7 (33.3%) 8 (38.1%)	7 (33.3%)		10 (47.6%) 1 (4.8%)	1 (4.8%)	10 (47.6%)	10 (47.6%) 18 (85.7%) 3 (14.3%) 14 (66.7%) 1 (4.8%)	3 (14.3%)	14 (66.7%)		6 (28.6%)	14 (66.7%)	1 (4.8%)	6 (28.6%)	6 (28.6%) 14 (66.7%) 1 (4.8%) 6 (28.6%) 13 (61.9%) 0		8 (38.1%)
Total		81 (54%) 69 (46%)	59 (39.3%)	59 (39.3%) 34 (22.7%) 57 (38%) 84 (56%)	57 (38%)	84 (56%)	(%9) 6	57 (38%)	124 (82.7%)	26 (17.3%%)	98 (65.3%)	7 (4.7%)	98 (65.3%) 7 (4.7%) 45 (30%)	88 (58.7%)	13 (8.7%)	88 (58.7%) 13 (8.7%) 49 (32.7%) 101 (67.3%)	· ·	3 (2%) 4	46 (30.7%)

Javdan *et al. BMC Res Notes* (2019) 12:363 Page 4 of 6

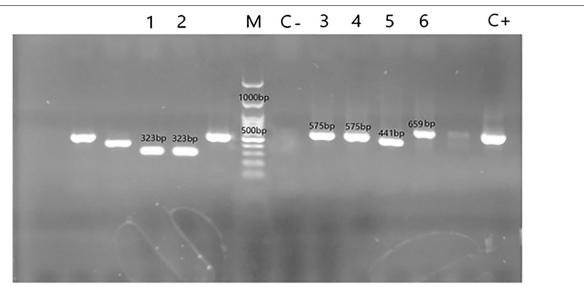


Fig. 1 Lanes 1 and 2: agr type III, lanes 3 and 4: agr type II, lane 5: agr type I, lane 6: agr type IV, M: 100 bp DNA ladder, C— and C+: negative and positive controls

chain of transmission. Accordingly, the present study was designed to identification of *agr* types among *S. aureus* isolates and possible association of these pathogens with some phenotypic characteristics such as antibiotic resistance and pathogenesis.

Dufour et al. [14] used agr typing method for the first time to stratify S. aureus isolates and affirmed that these bacteria can be divided into four groups I, II, III, IV by this system. Ever since, many researches have been applied the agr typing approach and in several studies such as those by Lee et al. and Shopsin et al. [15, 16], the agr group I was the most dominant S. aureus type. Our findings indicated that agr type I was the most predominant type among S. aureus isolated from Isfahan and Shahrekord cities. Similarly, in several previous studies such as those by Cheraghi, Bibalan, Peerayeh, Khoramrooz, Mohsenzadeh, and Goudarzi agr type I has been reported as the most dominant isolate of S. aureus in different regions of Iran [17-22]. It is declared that certain agr groups of S. aureus are involved in some particular disease and infections, for example agr type I isolates are associated with bacteremia and invasive infections [21]. In the present study, wound and tracheal aspirates, were sequentially the most frequent clinical samples and as it is summarized in Table 2, the agr group I was the major agr type among these sample. However, we couldn't find any significant difference or correlation between agr types and certain clinical specimen.

In the current study, the antimicrobial susceptibility testing revealed that the highest antibiotic resistance rate was against cefoxitin (46%) followed by erythromycin and tetracycline (both 38%). Several studies that have reported erythromycin and tetracycline as the antimicrobial agents with lowest susceptibility among S. aureusagr group I isolates [20, 23, 24]. As it is summarized in Table 1, the agr types III and I showed the maximum and minimum resistance rates against tetracycline, respectively. In the present study, agr type I isolates had the highest sensitivity to vancomycin; however, the smallest resistance rate against this agent was related to agr type IV (Table 1). The greatest susceptibility and resistance to rifampin were found among agr types IV and I of S. aureus strains, respectively (Table 1). As it could be seen in Table 1, agr types IV and III have shown the highest and the lowest susceptibility to trimethoprim, respectively. The maximum percentage of gentamicin susceptibility was related to agr type I, while type III isolates had the highest resistance against this antimicrobial agent (Table 1). We found a significant correlation between agr type and antibiotic resistance against cefoxitin and erythromycin (p = 0.04 and p = 0.03, respectively). Indeed, in this report, the agr types never implied the sensitivity or resistance to antibiotics, but in the case of cefoxitin and erythromycin the agr group I isolates showed the highest resistance against these agents.

The majority of *S. aureus* isolates in this study were classified as *agr* group I and our results suggest a probable correlation between this type and antibiotic resistance to cefoxitin and erythromycin. Here we can conclude that *agr* typing is a suitable and effective approach for molecular tracking of *S. aureus* infection.

Javdan et al. BMC Res Notes (2019) 12:363 Page 5 of 6

Table 2 Frequency distribution of agr types in different clinical samples

Sample	agr type				Tota
	I	II	III	IV	
Tracheal asp	oirate				
No .	13	6	2	2	23
%	56.5%	26.1%	8.7%	8.7%	1009
Wound					
No	30	11	4	5	50
%	60.0%	22.0%	8.0%	10.0%	1009
Tissue					
No	3	2	0	0	5
%	60.0%	40.0%	0.0%	0.0%	1009
Abscess					
No	5	5	1	1	12
%	41.7%	41.7%	8.3%	8.3%	1009
Conjunctiva		11.7 70	0.570	0.570	1007
No	0	1	0	1	2
%	0.0%	50.0%	0.0%	50.0%	1009
Blood cultu		50.070	0.070	50.070	1007
No	1	2	0	0	3
%	33.3%	66.7%	0.0%	0.0%	1009
Sputum	33.370	00.7 70	0.070	0.070	1007
•	2	0	0	า	4
No %	2	0		2	
	50.0%	0.0%	0.0%	50.0%	1009
Nasal aspira		1	1	0	4
No	2	1	1	0	4
%	50.0%	25.0%	25.0%	0.0%	
Synovial flui		0	0	2	2
No	0	0	0	3	3
%	0.0%	0.0%	0.0%	100%	1009
Discharge					_
No	3	3	1	0	7
%	42.9%	42.9%	14.3%	0.0%	1009
Bactace					
No	7	1	0	2	10
%	70.0%	10.0%	0.0%	20.0%	1009
Blood					
No	4	3	0	1	8
%	50.0%	37.5%	0.0%	12.5%	1009
Peritoneal fl	luid				
No	1	0	1	0	2
%	50.0%	0.0%	50.0%	0.0%	1009
Urine cultur	e				
No	9	2	0	3	14
%	64.3%	14.3%	0.0%	21.4%	1009
Pharyngeal	swab				
No	2	0	0	1	3
%	66.7%	0.0%	0.0%	33.3%	1009
Total					
No	82	37	10	21	150

Table 2 (continued)

Sample	agr type				Total
	I	II	III	IV	
%	54.7%	24.7%	6.7%	14.0%	100%

Limitations

The lack of investigation on others typing methods in *S. aureus* isolates can be mentioned as one of the main limitations of the present study.

Abbreviations

MRSA: methicillin-resistance *Staphylococcus aureus*; MDR: multi-drug resistant; SCCmec: Staphylococcal Cassette chromosome mec elements; PCR: polymerase chain reaction.

Acknowledgements

Not applicable.

Authors' contributions

SA, TN, MSSA, AG: design of study. MSSA, SA, AG: acquisition of data. TN, SJ, AG: evaluation of data, preparation of the manuscript. MSSA, AG: assessment of data. All authors read and approved the final manuscript.

Funding

This research was supported by the budget of research projects of the Shahre-kord University of medical sciences (Number of project: 2443). Funding body were used to purchase equipment and tools.

Availability of data and materials

All relevant data are included in the manuscript.

Ethics approval and consent to participate

This study was approved by the Ethics Committee of Shahrekord University of Medical Sciences. The informed consent was obtained from all the participants, and informed consent obtained was written.

Consent to publish

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

 Cellular and Molecular Research Center, Basic Health Sciences Institute, Shahrekord University of Medical Sciences, Shahrekord, Iran.
 Department of Microbiology, Isfahan University of Medical Sciences, Isfahan, Iran.
 Department of Bacteriology & Virology, School of Medicine, Shiraz University of Medical Sciences, Shiraz, Iran.
 Department of Microbiology and Immunology, Cellular and Molecular Research Center, Shahrekord University of Medical Sciences, Shahrekord, Iran.

Received: 19 May 2019 Accepted: 18 June 2019 Published online: 27 June 2019

References

- 1. Lowy FD. Antimicrobial resistance: the example of *Staphylococcus aureus*. J Clin Investig. 2003;111(9):1265–73.
- Zouhir A, Taieb M, Lamine MA, Cherif A, Jridi T, Mahjoubi B, et al. ANTISTAPHYBASE: database of antimicrobial peptides (AMPs) and essential oils (EOs) against methicillin-resistant Staphylococcus aureus (MRSA) and Staphylococcus aureus. Arch Microbiol. 2017;199(2):215–22.

Javdan *et al. BMC Res Notes* (2019) 12:363 Page 6 of 6

- Medina Cruz D, Mi G, Webster TJ. Synthesis and characterization of biogenic selenium nanoparticles with antimicrobial properties made by Staphylococcus aureus, methicillin-resistant Staphylococcus aureus (MRSA), Escherichia coli and Pseudomonas aeruginosa. J Biomed Mater Res Part A. 2018;106:1400–12.
- García-Álvarez L, Holden MT, Lindsay H, Webb CR, Brown DF, Curran MD, et al. Meticillin-resistant Staphylococcus aureus with a novel mecA homologue in human and bovine populations in the UK and Denmark: a descriptive study. Lancet Infect Dis. 2011;11(8):595–603.
- Gorak EJ, Yamada SM, Brown JD. Community-acquired methicillin-resistant Staphylococcus aureus in hospitalized adults and children without known risk factors. Clin Infect Dis. 1999;29(4):797–800.
- Herold BC, Immergluck LC, Maranan MC, Lauderdale DS, Gaskin RE, Boyle-Vavra S, et al. Community-acquired methicillin-resistant *Staphylococcus aureus* in children with no identified predisposing risk. JAMA. 1998;279(8):593–8.
- Afrough P, Pourmand MR, Zeinalinia N, Yousefi M, Abdossamadi Z, Bagherzadeh Yazdchi S. Molecular typing of clinical and nasal carriage isolates of Staphylococcus aureus by spa gene patterns. J Mazandaran Univ Med Sci. 2012;22(94):28–34.
- 8. Thompson TA, Brown PD. Association between the agr locus and the presence of virulence genes and pathogenesis in *Staphylococcus aureus* using a *Caenorhabditis elegans* model. Int J Infect Dis. 2017;54:72–6.
- Bibalan MH, Shakeri F, Javid N, Ghaemi A, Ghaemi EA. Accessory gene regulator types of Staphylococcus aureus isolated in Gorgan, North of Iran. J Clin Diagn Res JCDR. 2014;8(4):DC07.
- Mahon CR, Lehman DC, Manuselis G. Textbook of diagnostic microbiology-E-Book. Maryland Heights: Elsevier Health Sciences; 2014.
- P.W. Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing. CLSI guideline M100S.2016.
- Sambrook J, Fritsch E, Maniatis T. Molecular cloning: a laboratory manual+ cold Spring Harbor. New York: Cold spring harbor laboratory press; 1989.
- Trinh TD, Zasowski EJ, Claeys KC, Casapao AM, Compton M, Lagnf A, et al. Role of vancomycin minimum inhibitory concentrations by modified population analysis profile method and clinical outcomes in high inoculum methicillin-resistant Staphylococcus aureus infections. Infect Dis Ther. 2018:7:1–9
- Dufour P, Jarraud S, Vandenesch F, Greenland T, Novick RP, Bes M, et al. High genetic variability of the agr locus in *Staphylococcus species*. J Bacteriol. 2002;184(4):1180–6.
- van Leeuwen W, van Nieuwenhuizen W, Gijzen C, Verbrugh H, van Belkum A. Population studies of methicillin-resistant and-sensitive Staphylococcus

- aureus strains reveal a lack of variability in the agrD gene, encoding a staphylococcal autoinducer peptide. J Bacteriol. 2000;182(20):5721–9.
- Shopsin B, Mathema B, Alcabes P, Said-Salim B, Lina G, Matsuka A, et al. Prevalence of agr specificity groups among *Staphylococcus aureus* strains colonizing children and their guardians. J Clin Microbiol. 2003;41(1):456–9.
- Cheraghi S, Pourgholi L, Shafaati M, Fesharaki SH, Jalali A, Nosrati R, et al. Analysis of virulence genes and accessory gene regulator (agr) types among methicillin-resistant Staphylococcus aureus strains in Iran. J Glob Antimicrob Resist. 2017;10:315–20.
- 18. Hasannejad Bibalan M, Ghaemi E, Shakeri F, Javid N. The relation between accessory gene regulator (agr) types of *S. aureus* and some phenotypic criteria. Relation. 2014;17(87):1–8.
- Khoramrooz SS, Mansouri F, Marashifard M, Hosseini SAAM, Chenarestane-Olia FA, Ganavehei B, et al. Detection of biofilm related genes, classical enterotoxin genes and agr typing among *Staphylococcus aureus* isolated from bovine with subclinical mastitis in southwest of Iran. Microb Pathog. 2016;97:45–51.
- Goudarzi M, Bahramian M, Tabrizi MS, Udo EE, Figueiredo AMS, Fazeli M, et al. Genetic diversity of methicillin resistant *Staphylococcus aureus* strains isolated from burn patients in Iran: ST239-SCCmec III/t037 emerges as the major clone. Microb Pathog. 2017;105:1–7.
- 21. Peerayeh SN, Azimian A, Nejad QB, Kashi M. Prevalence of agr specificity groups among *Staphylococcus aureus* isolates from university hospitals in Tehran. Lab Med. 2015;40(1):27–9.
- Mohsenzadeh M, Ghazvini K, Azimian A, editors. Frequency of specific agr groups and antibiotic resistance in *Staphylococcus aureus* isolated from bovine mastitis in the northeast of Iran. Veterinary Research Forum; 2015. Urmia: Faculty of Veterinary Medicine, Urmia University.
- 23. Ghasemian A, Peerayeh SN, Bakhshi B, Mirzaee M. High frequency of icaAD, clumping factors A/B, fib and eno Genes in *Staphylococcus aureus* species isolated from wounds in Tehran, Iran during 2012–2013. Arch Clin Infect Dis. 2015;10(4):e23033.
- 24. Arabestani MR, Kazemian H, Tabar ZK, Hosseini SM. Prevalence of virulence genes, agr and antimicrobial resistance of *Staphylococcus aureus* isolated from food and dairy products in Hamadan Iran. Der Pharm Lett. 2016;8(8):62–7.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

