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Zero-dimensional chemical kinetic simulation of ROS/RNS in pulsed-discharge exposed water

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The concentration variations of reactive oxygen/nitrogen species in water, such as H₂O₂, NO₂⁻, and NO₃⁻ generated by pulsed-discharge plasma exposure, are calculated using reaction rates of chemical reactions and acid-base equilibrium in water. The calculated concentrations and pH values are in good agreement with measured data within the range where the significant changes of the measured data are observed. The rate constant for ONOOH generation is estimated to be $7.8 \times 10^3 \text{ M}^{-2} \cdot \text{s}^{-1}$, and this value is in good agreement with previously reported values. The generation rates of H₂O₂, NO₂⁻, and NO₃⁻ are estimated to be 7.70×10^{-7} , 4.10×10^{-7} , and $1.10 \times 10^{-7} \text{ M} \cdot \text{s}^{-1}$, respectively.

1. Introduction

Discharge plasma in contact with water and in water vapor has been recently applied to a wide range of applications, such as biomolecule decontamination¹⁾, nanoparticle synthesis²⁾, plant growth promoting³⁾, the degradation of organic compounds^{4,5)}, disinfection⁶⁻¹⁵⁾, and the treatment of cancer cells.^{16,17)} In general, active species, such as electrons, ions, radicals, and reactive neutral species, are generated in plasma, and these species dissolve in water when the plasma is generated in contact with the water. Then, reactive oxygen species (ROS), such as H₂O₂ and HO₂, are generated in the water, and reactive nitrogen species (RNS), such as HNO₂ and ONOOH, are also generated when nitrogen gas is contained in the ambient and/or dissolved gases. The water containing ROS/RNS is well known as plasma-treated water⁸⁻¹²⁾, which is also called plasma-activated water¹³⁻¹⁵⁾ and plasma-activated medium.^{16,17)} Since ROS/RNS have high oxidation potential, these potentially contribute to bactericidal applications, and the bactericidal mechanisms induced by ROS/RNS in plasma-treated water has been partially elucidated. Ikawa *et al.*¹¹⁾ exposed distilled water to low-temperature atmospheric pressure helium plasma, demonstrated the bactericidal activity of ROS/RNS in the water by mixing with *Escherichia coli* suspension, and reported that the bactericidal activity is enhanced under acidic conditions. It was suggested that HO₂ released from O₂NOOH plays an important role in imparting the bactericidal activity to the water. It was also suggested that O₂NOOH is generated by the reaction between ONOOH and H₂O₂, and that ONOOH is generated from HNO₂ and H₂O₂. Some of these species are in equilibrium with those conjugate base in water; therefore, the pH value of the water significantly affect the concentrations of acid and its conjugated base.

Several groups reported ROS/RNS concentrations in water exposed to plasma with the biocidal effects and oxidative strength of those species. Gils *et al.*¹²⁾ produced plasma-treated water using an atmospheric pressure argon plasma jet for inactivation of *Pseudomonas aeruginosa*, and reported the significant influence of the acidity of the plasma-treated water on bactericidal effect. They also calculated ROS/RNS concentrations in the liquid phase using zero-dimensional solution kinetics simulations, estimated the flux of O₃, NO, and OH, and compared the calculated concentrations of H₂O₂, NO₂⁻, and NO₃⁻ with those measured concentrations; however, the chemical equilibrium of species and the reactions of long-lived species (*e.g.* the reaction between HNO₂ and H₂O₂) are not considered. Lukes *et al.*¹³⁾ reported the concentrations of H₂O₂, NO₂⁻, and NO₃⁻ in plasma-exposed water as functions of treatment time and post-discharge time, and that the bactericidal effects of the water is enhanced under acidic conditions. They also reported the rate constant for ONOOH

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3 generation, estimated by a kinetic study of post-discharge processes in pH-buffered aqueous
4 solution. Anderson *et al.*⁵⁾ exposed a pH-buffered aqueous solution containing indigo
5 carmine to non-equilibrium atmospheric-pressure plasma, and estimated the branching ratios
6 of OH/NO₂ and NO₃⁻/H⁺ generation through ONOOH decomposition. Hence, the
7 bactericidal effects and oxidative strength of ROS/RNS and the reactions of ROS/RNS have
8 been well investigated; however, ROS/RNS concentration variations in plasma-exposed
9 water accompanied with the pH variation of the water, caused by dissolving ROS/RNS in
10 the water, have not yet been fully reproduced by solution kinetics simulation, and it
11 contributes to predicting the reactions of ROS/RNS and estimating the generation rate of
12 ROS/RNS in the liquid phase.

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20 In this work, the variations of ROS/RNS concentrations and pH value in water exposed
21 to plasma were calculated by using reaction rates based on acid-base equilibrium and
22 chemical reactions. Our previous work¹⁸⁾ suggested that various discharge plasmas above
23 water, which are a pulsed discharge, a DC corona discharge, and a plasma jet, generate H₂O₂,
24 NO₂⁻, and NO₃⁻ in the water, and discussed the generation process of those species in the
25 gas and liquid phase; however, measurement conditions in the previous work, such as post-
26 discharge time to start analysing plasma-exposed water, were not necessarily the same. Thus,
27 the ROS/RNS concentrations and pH value in water exposed to the pulsed-discharge plasma,
28 which is effective in ROS/RNS generation using the plasmas, were remeasured under the
29 unified conditions in this work. Then, the variations of ROS/RNS concentrations and pH
30 value in the water were calculated by using reaction rates of chemical reactions in water.
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40 2. Experimental apparatus and conditions

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42 The pulsed discharge was generated in the same manner as in our previous work.¹⁸⁾ A
43 cylindrical discharge chamber to generate the pulsed discharge consisted of a needle
44 electrode and a water bath electrode. The needle electrode was a stainless-steel needle with
45 4.0 mm in diameter and 35 mm in length, and the water bath electrode was made of stainless
46 steel with 119 mm in inner diameter, 12 mm in depth, and a capacity of 0.13 L. Deionized
47 water of 100 mL was poured into the water bath electrode, and the distance between the tip
48 of the needle electrode and the water surface was fixed at 4 mm. Nitrogen gas was fed into
49 the chamber at a constant flow rate of 5 L·min⁻¹. A pulsed high voltage with a pulse width
50 of 500 ns generated by a Blumlein generator, which has two coaxial transmission lines, was
51 applied to the needle electrode to generate the pulsed discharge above the water surface. The
52 coaxial transmission lines were charged to a negative voltage of 14.14 kV, and a pulse
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3 repetition rate was 20 pulses per second. The pH value of the water and the ROS/RNS
4 concentrations in the water were measured 4 min after plasma exposure. The pH value and
5 temperature of the water and the ROS/RNS concentrations in the water were measured 4
6 min after plasma exposure. The pH value and temperature were measured using a pH meter
7 (CyberScan PCWP10), and 1.2 mL of water sample, taken from the plasma-exposed water,
8 were analyzed using a high-performance liquid chromatograph (HPLC; Shimadzu
9 Prominence) equipped with an ion chromatography column (Shodex IC NI-424) in
10 combination with an absorbance detector. The wavelength of the absorbance detector was
11 fixed at 220 nm. The eluent of the HPLC was a mixed aqueous solution of 3 mM (mmol·L⁻¹)
12 acetic acid and 1.9 mM potassium hydroxide, the pH value of the solution was 5.1, and the
13 column bath temperature of the HPLC was set to 40°C.
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23 3. Results and discussion

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25 Figures 1(a), 1(b), and 1(c) show photographs of the pulsed-discharge plasma immediately,
26 5 min, and 30 min after plasma exposure, respectively. The discharge reaches the water
27 surface, and then splits into several branches. A single thick discharge reaches the rim of the
28 water bath electrode immediately after plasma exposure, and several thin discharges that do
29 not reach the rim of the water bath electrode spread over the water surface after 5 min of the
30 plasma exposure. The extension of the discharge along the water surface tends to decrease;
31 the discharge area is about 100 mm in diameter after 5 min of plasma exposure, and about
32 60 mm in diameter after 30 min of plasma exposure. Figure 2 shows the variations of water
33 temperature as a function of the exposure time. The water temperature shows a tendency to
34 increase and then become constant with the plasma exposure.
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43 Figure 3 shows the chromatogram of the sample after 20 min of plasma exposure. H₂O₂,
44 NO₂⁻, and NO₃⁻ were detected at the retention time of 1.7, 11.6, and 18.0 min, respectively.
45 According to Refs. 11-13, ONOOH and O₂NOOH, which are key species in bactericidal
46 applications, can be produced in plasma-exposed water; however, the peaks corresponding
47 to those species were not detected as shown in Fig. 3. This is due to the lifetime of those
48 species. Since the half-life of ONOOH is typically less than 1 s, ONOOH is decomposed to
49 long-lived species, such as NO₃⁻. The lifetime of O₂NOOH depends on temperature¹⁹⁾ and
50 pH²⁰⁾, becoming shorter at higher temperature and higher pH. Nakashima *et al.*²¹⁾ reported
51 that O₂NOOH was detected at a column bath temperature of below 25°C but not at that of
52 40°C using an eluent of pH 2 in ion-exchange chromatography, because O₂NOOH
53 decomposed to nitrous acid and nitrate between the injector and detector. In this work, the
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column bath temperature and the pH value of eluent were set to 40°C and 5.1, respectively; therefore, O₂NOOH in water is completely decomposed between the injector and detector, and this results in no peak of O₂NOOH. Figure 4 shows the concentrations of H₂O₂, NO₂⁻, and NO₃⁻ in the sampled water and the pH value of the water as functions of the exposure time. H₂O₂ and NO₃⁻ concentrations increase monotonously with the exposure time, while NO₂⁻ concentration increases, reaches its peak, and then decreases to zero. This result shows a similar tendency, observed in the previous work¹⁸⁾, and these species were generated by dissolving H₂O₂, HNO₂, and HNO₃ in water.^{13,22-29)} Therefore, chemical reactions shown in Table I are deduced. Since the pH value decreases with the increase of the exposure time, the concentration of HNO₂, which is in equilibrium with NO₂⁻ (pK_a = 3.3)¹³⁾, increases and reacts with H₂O₂ to form ONOOH. Therefore, NO₂⁻ concentration drops off in the presence of H₂O₂ and the decrease of pH value. Furthermore, ONOOH is an unstable species and changes into NO₂/OH or NO₃⁻/H⁺. OH and NO₂ may change into H₂O₂ and NO₂⁻/NO₃⁻ via the reactions shown by the reactions shown by Eqs. (3) and (4), respectively.

According to the chemical reactions in water shown in Table I, the concentration variations of ROS/RNS in water exposed to pulsed-discharge plasma were calculated by using the reaction rates of the reactions. Since H₂O₂, NO₂⁻, and NO₃⁻ were generated as ROS/RNS in water by the plasma exposure, it was assumed that H₂O₂, NO₂⁻, and NO₃⁻ were generated in water with the plasma exposure. Accordingly, the reaction rates are represented as follows:

$$[\text{NO}_2^-] = f([\text{NO}_2^-] + [\text{HNO}_2]), \quad (5)$$

$$[\text{HNO}_2] = (1 - f)([\text{NO}_2^-] + [\text{HNO}_2]), \quad (6)$$

$$\frac{d[\text{ONOOH}]}{dt} = k_1[\text{H}_2\text{O}_2][\text{HNO}_2][\text{H}^+] - k_2[\text{ONOOH}], \quad (7)$$

$$\frac{d[\text{HNO}_2]}{dt} = -k_1[\text{H}_2\text{O}_2][\text{HNO}_2][\text{H}^+], \quad (8)$$

$$\frac{d[\text{H}_2\text{O}_2]}{dt} = G_{\text{H}_2\text{O}_2} - k_1[\text{H}_2\text{O}_2][\text{HNO}_2][\text{H}^+] + k_3[\text{OH}], \quad (9)$$

$$\frac{d[\text{NO}_2^-]}{dt} = G_{\text{NO}_2^-} + k_4[\text{NO}_2], \quad (10)$$

$$\frac{d[\text{NO}_3^-]}{dt} = G_{\text{NO}_3^-} + 0.76k_2[\text{ONOOH}] + k_4[\text{NO}_2], \quad (11)$$

$$\frac{d[\text{OH}]}{dt} = 0.24k_2[\text{ONOOH}] - 2k_3[\text{OH}]^2, \quad (12)$$

$$\frac{d[\text{NO}_2]}{dt} = 0.24k_2[\text{ONOOH}] - 2k_4[\text{NO}_2]^2, \quad (13)$$

where $[X]$ is the concentration of the species X at time t , f is the ionization degree of HNO_2 , G_X is the generation rate of the species X , and k_1 , k_2 , k_3 , and k_4 are the rate constants for the reactions shown by Eqs. (1), (2), (3), and (4), respectively. The coefficients 0.76 and 0.24 in Eqs. (11)-(13) are originated in the branching ratios of the reaction shown by Eq. (2). The value of f is determined by $\text{p}K_a$ for HNO_2 and the pH value of the water. The values of the rate constants k_2 , k_3 , and k_4 were reported as $0.13 + 0.87[\text{H}^+] \text{ s}^{-1}$ in Ref. 5, $4.2 \times 10^9 \text{ M}^{-1} \cdot \text{s}^{-1}$ in Ref. 31, and $1.0 \times 10^8 \text{ M}^{-1} \cdot \text{s}^{-1}$ in Ref. 32, respectively. The concentrations of H_2O_2 , NO_2^- , NO_3^- , HNO_2 , ONOOH , OH , and NO_2 and the pH value in the water, determined by NO_2^- and NO_3^- concentrations, were calculated by using the above reaction rates, and those were fitted to the measured data, as shown in Fig. 2, by varying the rate constant k_1 and the generation rates of H_2O_2 , NO_2^- , and NO_3^- . The concentration distribution was assumed to be uniform, and the time variation of concentrations during post-discharge period to start analyzing the plasma-exposed water and the pH variation in the liquid chromatographic analysis were considered. The above equations were calculated by the Runge-Kutta fourth-order method.

Figure 5 shows the calculated concentrations of ROS/RNS and pH value, together with the measured data as functions of time. The calculated concentrations of H_2O_2 , NO_2^- , and NO_3^- and pH value were in approximate agreement with the measured data below 3000 s, when k_1 was estimated to be $7.8 \times 10^3 \text{ M}^{-2} \cdot \text{s}^{-1}$ and the generation rates of H_2O_2 , NO_2^- , and NO_3^- were estimated to be 7.70×10^{-7} , 4.10×10^{-7} , and $1.10 \times 10^{-7} \text{ M} \cdot \text{s}^{-1}$, respectively. The estimated value of the rate constant k_1 is in good agreement with previously reported values, $8.3(\pm 0.6) \times 10^3 \text{ M}^{-2} \cdot \text{s}^{-1}$ in Ref. 33, $4.6 \times 10^3 (\pm 20\%) \text{ M}^{-2} \cdot \text{s}^{-1}$ in Ref. 34, and $(6.3 \pm 1.5) \times 10^3 \text{ M}^{-2} \cdot \text{s}^{-1}$ in Ref. 35. The calculated concentrations and pH were overestimated comparing to the measured data above 3000 s. This may be due to the change of the generation rate. Although the generation rates of H_2O_2 , HNO_2 , and HNO_3 were assumed to be constant in the calculation, the generation rates may decrease since the discharge area was contracted with time as shown in Fig. 1. The water temperature affects the reaction constant of chemical reactions in water, and the temperature change was observed as shown in Fig. 2. The calculated concentrations of ROS/RNS were in good agreement with measured data within the range where the temperature change was observed; therefore, the ROS/RNS generation rates may be significantly affected not by the temperature change but by the contraction of the discharge area.

This work suggests that the simple model considering only four reactions shown in Eqs.

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3 (1)-(4) with the assumption of complete mixing in the liquid phase can roughly reproduce
4 the measured data, so that it may allow to roughly estimate the generation rates of H₂O₂,
5 NO₂⁻, and NO₃⁻ and evaluate the generation efficiencies of those species. In order to
6 simulate the generation of ROS/RNS in liquid more accurately, it is desirable to consider the
7 variations of the generation rate depending on the discharge area and the temperature
8 dependence of the rate constants of chemical reactions in water.
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13 14 15 **4. Conclusions**

16 The concentrations of ROS/RNS and pH value in pulsed-discharge exposed water were
17 remeasured under the unified conditions and calculated using reaction rates based on acid-
18 base equilibrium and chemical reactions in the water. H₂O₂, NO₂⁻, and NO₃⁻ were generated
19 as a similar tendency in our previous work. The calculated concentrations of H₂O₂, NO₂⁻,
20 and NO₃⁻ and pH value in the water exposed to the pulsed-discharge plasma were in
21 approximate agreement with the measured data below 3000 s, when the rate constant for
22 ONOOH generation was set to $7.8 \times 10^3 \text{ M}^{-2} \cdot \text{s}^{-1}$, which is in good agreement with previously
23 reported values. The generation rates of H₂O₂, NO₂⁻, and NO₃⁻ were estimated to be
24 7.70×10^{-7} , 4.10×10^{-7} , and $1.10 \times 10^{-7} \text{ M} \cdot \text{s}^{-1}$, respectively.
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References

- 1) F. Fumagalli, O. Kylián, L. Amato, J. Hanus, and F. Rossi, *J. Phys. D* **45**, 135203 (2012).
- 2) N. Shirai, S. Uchida, and F. Tochikubo, *Jpn. J. Appl. Phys.* **53**, 046202 (2014).
- 3) D. P. Park, K. Davis, S. Gilani, C.-A. Alonzo, D. Dobrynin, G. Friedman, A. Fridman, A. Rabinovich, and G. Fridman, *Curr. Appl. Phys.* **13**, S19 (2013).
- 4) G. Tang, A. Komuro, K. Takahashi, and A. Ando, *J. Plasma Fusion Res.* **11**, 2406025 (2016).
- 5) C. E. Anderson, N. R. Cha, A. D. Lindsay, D. S. Clark, and D. B. Graves, *Plasma Chem. Plasma Process.* **36**, 1393 (2016).
- 6) D. Dobrynin, G. Friedman, A. Fridman, and A. Starikovskiy, *New J. Phys.* **13**, 103033 (2011).
- 7) Q. Zhang, P. Sun, H. Feng, R. Wang, Y. Liang, W. Zhu, K. H. Becker, J. Zhang, and J. Fang, *J. Appl. Phys.* **111**, 123305 (2012).
- 8) A. Kojtari, U. K. Ercan, J. Smith, G. Friedman, R. B. Sensenig, S. Tyagi, S. G. Joshi, H.-F. Ji, and A. D. Brooks, *J. Nanomedicine Biother. Discov.* **4**, 1000120 (2013).
- 9) H.-S. Kim, K. C. Wright, I.-W. Hwang, D.-H. Lee, A. Rabinovich, A. Fridman, and Y. I. Cho, *Int. Commun. Heat Mass Transf.* **42**, 5 (2013).
- 10) M. Naïtali, G. Kamgang-Youbi, J.-M. Herry, M.-N. Bellon-Fontaine, and J.-L. Brisset, *Appl. Environ. Microbiol.* **76**, 7662 (2010).
- 11) S. Ikawa, A. Tani, Y. Nakashima, and K. Kitano, *J. Phys. D.* **49**, 425401 (2016).
- 12) C. A. J. van Gils, S. Hofmann, B. K. H. L. Boekema, R. Brandenburg, and P. J. Bruggeman, *J. Phys. D* **46**, 175203 (2013).
- 13) P. Lukes, E. Dolezalova, I. Sisrova, and M. Clupek, *Plasma Sources Sci. Technol.* **23**, 015019 (2014).
- 14) M. J. Traylor, M. J. Pavlovich, S. Karim, P. Hait, Y. Sakiyama, D. S. Clark, and D. B. Graves, *J. Phys. D.* **44**, 472001 (2011).
- 15) Q. Zhang, Y. Liang, H. Feng, R. Ma, Y. Tian, J. Zhang, and J. Fang, *Appl. Phys. Lett.* **102**, 203701 (2013).
- 16) S. Mohades, M. Laroussi, J. Sears, N. Barekzi, and H. Razavi, *Phys. Plasmas* **22**, 122001 (2015).
- 17) J. Ikeda, H. Tanaka, K. Ishikawa, H. Sakakita, Y. Ikehara, and M. Hori, *Pathol. Int.* **68**, 23 (2018).
- 18) K. Takahashi, K. Satoh, H. Itoh, H. Kawaguchi, I. Timoshkin, M. Given, and S. MacGregor, *Jpn. J. Appl. Phys.* **55**, 07LF01 (2016).

- 19) J.-M. Régimbal and M. Mozurkewich, *J. Phys. Chem. A*, **101**, 8822 (1997).
- 20) R. A. Kenley, P. L. Trevor, and B. Y. Lan, *J. Am. Chem. Soc.* **103**, 2203 (1981).
- 21) Y. Nakashima, S. Ikawa, A. Tani, and K. Kitano, *J. Chromatogr. A*. **1431**, 89 (2016).
- 22) Y. Itikawa and N. Mason, *J. Phys. Chem. Ref. Data* **34**, 1 (2005).
- 23) W. Lindinger, *Phys. Rev. A* **7**, 328 (1973).
- 24) R. P. Joshi and S. M. Thagard, *Plasma Chem. Plasma Process.* **33**, 17 (2013).
- 25) R. Atkinson, D. L. Baulch, R. A. Cox, J. N. Crowley, R. F. Hampson, R. G. Hynes, M. E. Jenkin, M. J. Rossi, and J. Troe, *Atmos. Chem. Phys.* **4**, 1461 (2004).
- 26) S. Mededovic and B.R. Locke, *J. Phys. D* **40**, 7734 (2007).
- 27) R. Atkinson, D. L. Baulch, R. A. Cox, R. F. Hampson, Jr., J. A. Kerr, and J. Troe, *J. Phys. Chem. Ref. Data* **18**, 881 (1989).
- 28) S. Goldstein, J. Lind, and G. Marényi, *Chem. Rev.* **105**, 2457 (2005).
- 29) L. D'Ottone, P. Campuzano-Jost, D. Bauer, and A. J. Hynes, *J. Phys. Chem. A* **105**, 10538 (2001).
- 30) G. Merényi, J. Lind, G. Czapski, and S. Goldstein, *Inorg. Chem.* **42**, 3796 (2003).
- 31) A. J. Elliot and D. R. McCracken, *J. Chem. Soc. Faraday Trans.* **86**, 1539 (1990).
- 32) Y.-N. Lee and S. E. Schwartz, *J. Phys. Chem.* **85**, 840 (1981).
- 33) E. Halfpenny and P. L. Robinson, *J. Chem. Soc.* 928 (1952).
- 34) D. E. Damschen and L. R. Martin, *Atmos. Environ.* **17**, 2005 (1983).
- 35) Y.-N. Lee and J. A. Lind, *J. Geophys. Res. Atmos.* **91**, 2793 (1986).

Figure Captions

Fig. 1. (Color online) Photographs of pulsed-discharge plasma: (a) Immediately after plasma exposure, (b) After 5 min of plasma exposure, and (c) After 30 min of plasma exposure.

Fig. 2. (Black and white) The variations of water temperature.

Fig. 3. (Color online) Chromatogram of a sample taken after 20 min of plasma exposure.

Fig. 4. (Color online) The variations of concentrations and pH value as functions of exposure time: (a) concentrations and (b) pH.

Fig. 5. (Color online) Comparison of calculated and measured concentrations and pH value: (a) H_2O_2 , (b) NO_2^- , (c) NO_3^- , and (d) pH.

Table I. ROS/RNS reactions in water.

Reaction	Eq.	Ref.
$\text{HNO}_2 + \text{H}_2\text{O}_2 + \text{H}^+ \rightarrow \text{ONOOH} + \text{H}_2\text{O} + \text{H}^+$	(1)	27)
$\text{ONOOH} \rightarrow \text{NO}_2 + \text{OH}$ (24%) or $\text{NO}_3^- + \text{H}^+$ (76%)	(2)	13)
$\text{OH} + \text{OH} \rightarrow \text{H}_2\text{O}_2$	(3)	28)
$2\text{NO}_2 + \text{H}_2\text{O} \rightarrow \text{NO}_2^- + \text{NO}_3^- + 2\text{H}^+$	(4)	29)

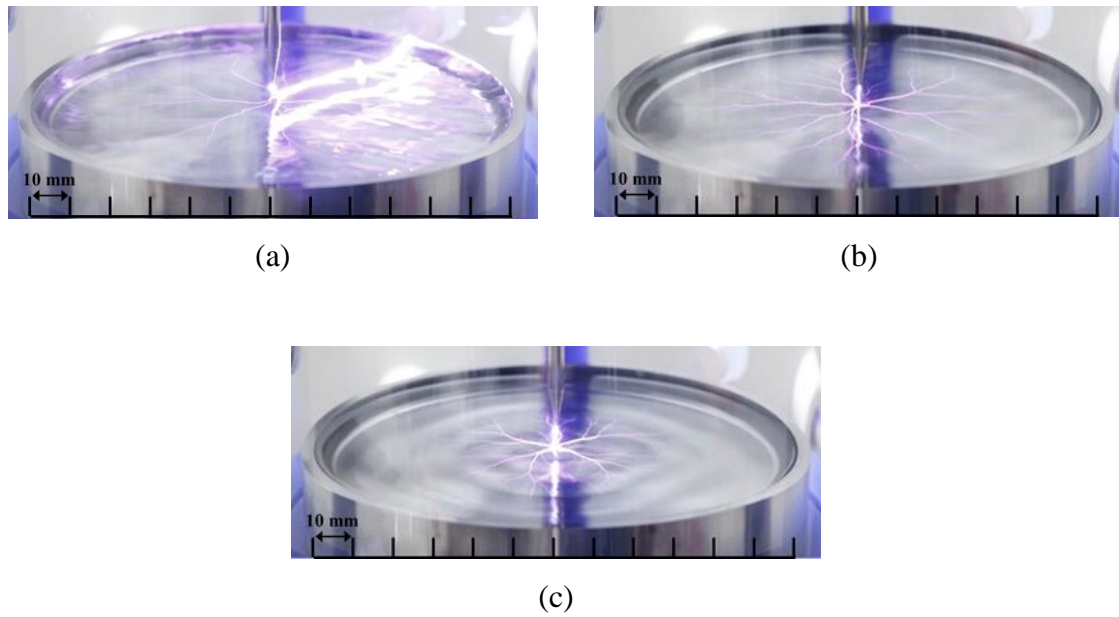


Fig. 1. (Color Online)

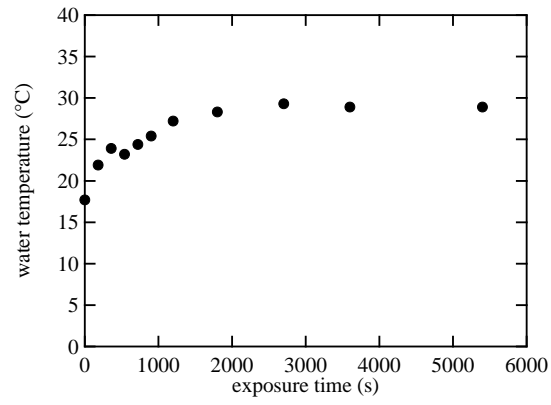


Fig. 2. (Black and white)

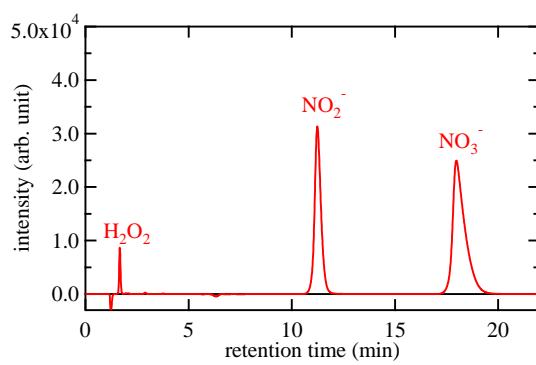


Fig. 3. (Color Online)

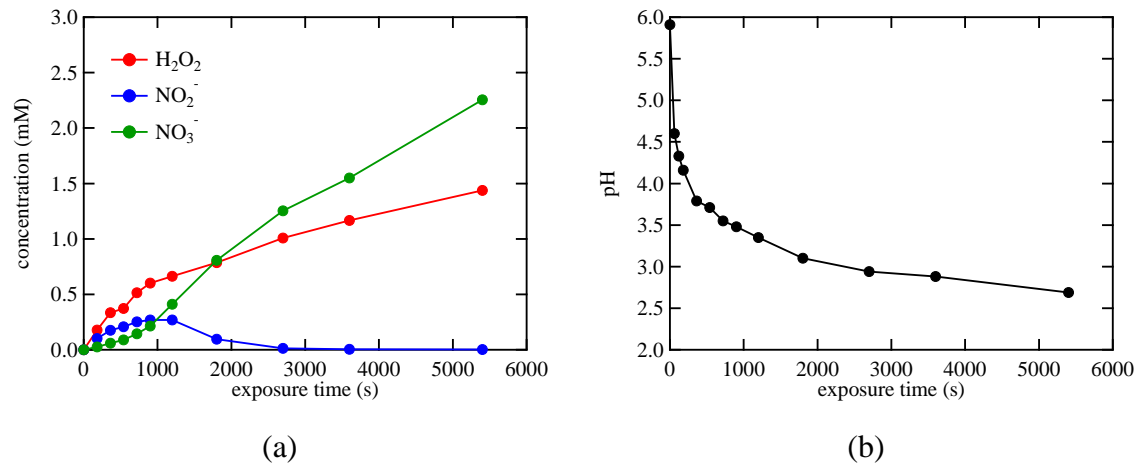


Fig. 4. (Color Online)

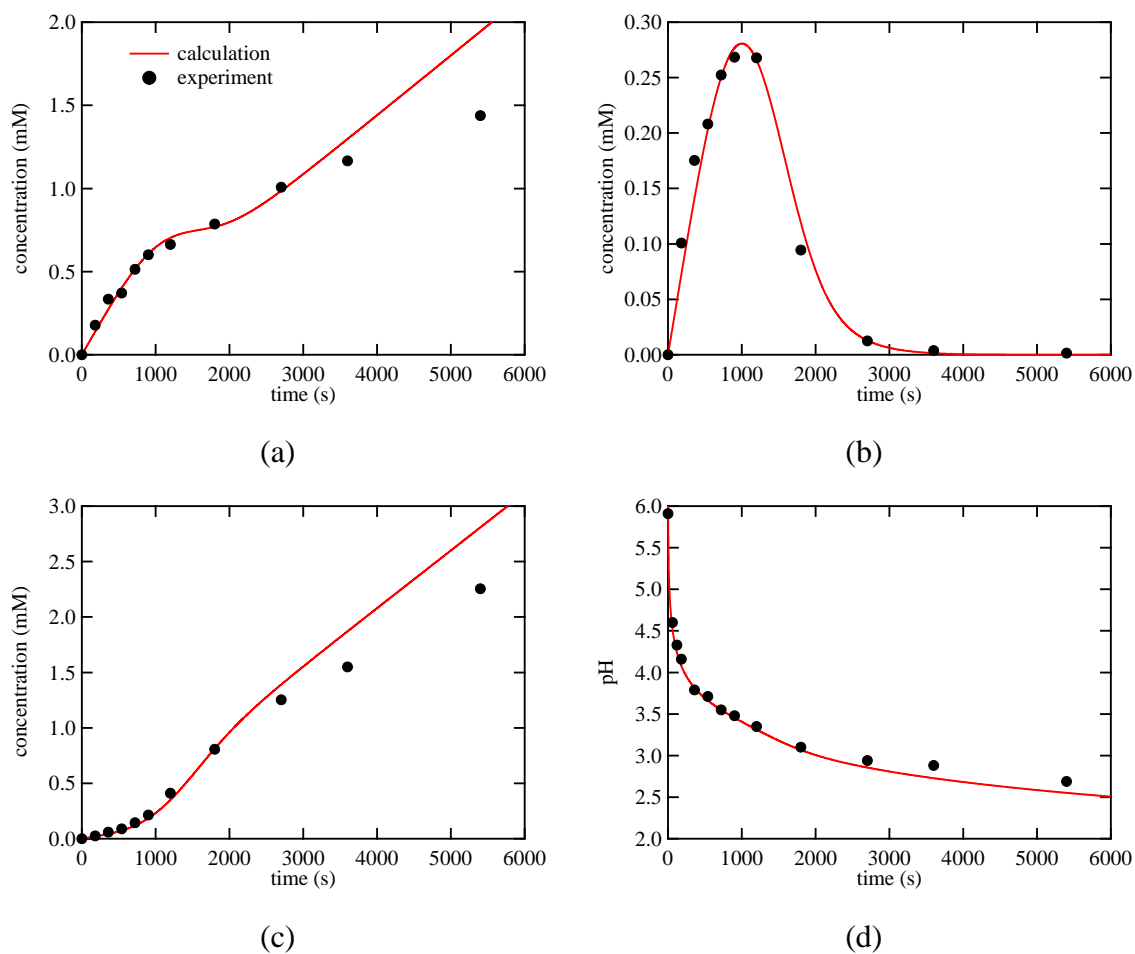


Fig. 5. (Color Online)