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Associations of the Top 20 Alzheimer Disease Risk Variants With Brain Amyloidosis

Liana G. Apostolova, MD, MSc, ^{[1,2,3} Shannon L. Risacher, PhD,² Tugce Duran, BS,¹ Eddie C. Stage, BS,¹ Naira Goukasian, BS,⁴ John D. West, MS,² Triet M. Do, BS,⁴ Jonathan Grotts, MA,⁵ Holly Wilhalme, MS,⁵ Kwangsik Nho, PhD,² Meredith Phillips, BA,¹ David Elashoff, PhD,⁵ and Andrew J. Saykin, PsyD^{2,3}, for the Alzheimer's Disease Neuroimaging Initiative

¹Department of Neurology, School of Medicine, Indiana University, Indianapolis

²Department of Radiology and Imaging Sciences, Center for Neuroimaging, School of Medicine, Indiana University, Indianapolis

³Department of Medical and Molecular Genetics, School of Medicine, Indiana University, Indianapolis

⁴Department of Neurology, David Geffen School of Medicine at UCLA, Los Angeles, California

⁵Department of Medicine Statistics Core, David Geffen School of Medicine at UCLA, Los Angeles, California

Article Information

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Group Information: The members of the Alzheimer's Disease Neuroimaging Initiative are listed at the end of this article.

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Corresponding Author: Liana G. Apostolova, MD, MSc, Department of Neurology, School of Medicine, Indiana University, 355 W 16th St, Ste 4700, Indianapolis, IN 46202 (<u>lapostol@iu.edu</u>).

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Study concept and design: Apostolova, Saykin.

Acquisition, analysis, or interpretation of data: All authors.

Drafting of the manuscript: Apostolova, Stage, Goukasian, Grotts, Nho.

Critical revision of the manuscript for important intellectual content: Apostolova, Risacher, Duran, Stage, Goukasian, West, Do, Wilhalme, Nho, Phillips, Elashoff, Saykin.

Statistical analysis: Apostolova, Risacher, Duran, Stage, Goukasian, West, Do, Grotts, Wilhalme, Nho, Phillips, Elashoff.

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Group Information: A complete listing of Alzheimer's Disease Neuroimaging Initiative (ADNI) investigators is as follows: ADNI I, GO, II, and III Leadership and Infrastructure: Principal Investigator: Michael W. Weiner, MD, University of California, San Francisco; ATRI PI and Director of Coordinating Center Clinical Core: Paul Aisen, MD, University of Southern California; Executive Committee: Michael Weiner, MD, University of California, San Francisco; Paul Aisen, MD, University of Southern California; Ronald Petersen, MD, PhD, Mayo Clinic, Rochester; Clifford R. Jack, Jr, MD, Mayo Clinic, Rochester; William Jagust, MD, University of California, Berkeley; John Q. Trojanowki, MD, PhD, University of Pennsylvania; Arthur W. Toga, PhD, University of Southern California; Laurel Beckett, PhD, University of California, Davis; Robert C. Green, MD, MPH, Brigham and Women's Hospital/Harvard Medical School; Andrew J. Saykin, PsyD, Indiana University; John Morris, MD, Washington University, St Louis; Leslie M. Shaw, University of Pennsylvania; ADNI External Advisory Board: Zaven Khachaturian, PhD, Prevent Alzheimer's Disease 2020 (chair); Greg Sorensen, MD, Siemens; Maria Carrillo, PhD, Alzheimer's Association; Lew Kuller, MD, University of Pittsburgh; Marc Raichle, MD, Washington University; St Louis; Steven Paul, MD, Cornell University; Peter Davies, MD, Albert Einstein College of Medicine of Yeshiva University; Howard Fillit, MD, AD Drug Discovery Foundation; Franz Hefti, PhD, Acumen Pharmaceuticals; David Holtzman, MD, Washington University, St Louis; M. Marcel Mesulam, MD, Northwestern University; William Potter, MD, National Institute of Mental Health; Peter Snyder, PhD, Brown University; ADNI 3 Private

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Jack, Jr, MD, Mayo Clinic, Rochester (core principal investigator); Matthew Bernstein, PhD, Mayo Clinic, Rochester; Nick Fox, MD, University of London; Paul Thompson, PhD, UCLA School of Medicine; Norbert Schuff, PhD, University of California, San Francisco, MRI; Charles DeCarli, MD, University of California, Davis; Bret Borowski, RT, Mayo Clinic; Jeff Gunter, PhD, Mayo Clinic; Matt Senjem, MS, Mayo Clinic; Prashanthi Vemuri, PhD, Mayo Clinic; David Jones, MD, Mayo Clinic; Kejal Kantarci, Mayo Clinic; Chad Ward Mayo Clinic; PET Core Leaders and Key Personnel: William Jagust, MD, University of California, Berkeley (core principal investigator); Robert A. Koeppe, PhD, University of Michigan; Norm Foster, MD, University of Utah; Eric M. Reiman, MD, Banner Alzheimer's Institute; Kewei Chen, PhD, Banner Alzheimer's Institute; Chet Mathis, MD, University of Pittsburgh; Susan Landau, PhD, University of California, Berkeley; Neuropathology Core Leaders: John C. Morris, MD, Washington University, St Louis; Nigel J. Cairns, PhD, FRCPath, Washington University, St Louis; Erin Franklin, MS, CCRP Washington University, St Louis; Lisa Taylor-Reinwald, BA, HTL, Washington University, St Louis (past investigator); Biomarkers Core Leaders and Key Personnel: Leslie M. Shaw, PhD, University of Pennsylvania School of Medicine; John Q. Trojanowki, MD, PhD, University of Pennsylvania School of Medicine; Virginia Lee, PhD, MBA, University of Pennsylvania School of Medicine; Magdalena Korecka, PhD, University of Pennsylvania School of Medicine; Michal Figurski, PhD, University of Pennsylvania School of Medicine; Informatics Core Leaders and Key Personnel: Arthur W. Toga, PhD, University of Southern California (core principal investigator); Karen Crawford, University of Southern California; Scott Neu, PhD, University of Southern California; Genetics Core Leaders and Key Personnel: Andrew J. Saykin, PsyD, Indiana University; Tatiana M. Foroud, PhD, Indiana University; Steven Potkin, MD, University of California, Irvine; Li Shen, PhD, Indiana University; Kelley Faber, MS, CCRC, Indiana University; Sungeun Kim, PhD, Indiana University; Kwangsik Nho, PhD, Indiana University; Initial Concept Planning & Development: Michael W. Weiner, MD, University of California, San Francisco; Lean Thal, MD, University of California, San Diego; Zaven Khachaturian, PhD, Prevent Alzheimer's Disease 2020; Early Project Proposal Development: Leon Thal, MD, University of California, San Diego; Neil Buckholtz, National Institute on Aging; Michael W. Weiner, MD, University of California, San Francisco; Peter J. Snyder, PhD, Brown University; William Potter, MD, National Institute of Mental Health; Steven Paul, MD, Cornell University; Marilyn Albert, PhD, Johns Hopkins University; Richard Frank, MD, PhD, Richard Frank Consulting; Zaven Khachaturian, PhD, Prevent Alzheimer's Disease 2020; National Institute on Aging: John Hsiao, MD, National Institute on Aging; Investigators by Site Oregon Health & Science University: Joseph Quinn, MD; Lisa C. Silbert, MD; Betty Lind, BS; Jeffrey A. Kaye, MD (past investigator); Raina Carter, BA (past investigator); Sara Dolen, BS (past investigator); University of Southern California: Lon S. Schneider, MD; Sonia Pawluczyk, MD; Mauricio Becerra, BS; Liberty Teodoro, RN; Bryan M. Spann, DO, PhD (past investigator); University of California, San Diego: James Brewer, MD, PhD; Helen Vanderswag, RN; Adam Fleisher, MD (past investigator); University of Michigan: Jaimie Ziolkowski, MA, BS, TLL; Judith L. Heidebrink, MD, M; Joanne L. 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Green, MD, MPH, Brigham and Women's Hospital/Harvard Medical School (chair); Resource Allocation Review Committee: Tom Montine, MD, PhD, University of Washington (chair); Clinical Core Leaders: Michael Weiner MD (core principal investigator); Ronald Petersen, MD, PhD, Mayo Clinic, Rochester (core principal investigator); Paul Aisen, MD, University of Southern California; Clinical Informatics and Operations: Gustavo Jimenez, MBS, University of Southern California; Michael Donohue, PhD, University of Southern California; Devon Gessert, BS. University of Southern California; Kelly Harless, BA, University of Southern California; Jennifer Salazar, MBS, University of Southern California; Yuliana Cabrera, BS, University of Southern California; Sarah Walter, MSc, University of Southern California; Lindsey Hergesheimen, BS, University of Southern California; San Francisco Veterans Affairs Medical Center: Thomas

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Neylan, MD, University of California, San Francisco; Jacqueline Hayes, University of California, San Francisco; Shannon Finley, University of California, San Francisco; Biostatistics Core Leaders and Key Personnel: Danielle Harvey, PhD, University of California, Davis (core principal investigator); Michael Donohue, PhD, University of California, San Diego; MRI Core Leaders and Key Personnel: Clifford R. Jack, Jr, MD, Mayo Clinic, Rochester (core principal investigator); Matthew Bernstein, PhD, Mayo Clinic, Rochester; Bret Borowski, RT, Mayo Clinic; Jeff Gunter, PhD, Mayo Clinic; Matt Senjem, MS, Mayo Clinic; Kejal Kantarci, Mayo Clinic; Chad Ward, Mayo Clinic; PET Core Leaders and Key Personnel: William Jagust, MD, University of California, Berkeley (core principal investigator); Robert A. Koeppe, PhD, University of Michigan; Norm Foster, MD, University of Utah; Eric M. 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Potkin, MD; Adrian Preda, MD; Dana Nguyen, PhD; Medical University South Carolina: Jacobo Mintzer, MD, MBA; Dino Massoglia, MD, PhD, Olga Brawman-Mintzer, MD; Premiere Research Inst (Palm Beach Neurology): Carl Sadowsky, MD; Walter Martinez, MD; Teresa Villena, MD; University of California, San Francisco: William Jagust MD; Susan Landau, PhD; Howard Rosen, MD; David Perry; Georgetown University Medical Center: Raymond Scott Turner, MD, PhD; Kelly Behan Brigid Reynolds, NP; Brigham and Women's Hospital: Reisa A. Sperling, MD; Keith A. Johnson, MD; Gad Marshall, MD; Banner Sun Health Research Institute: Marwan N. Sabbagh, MD; Sandra A. Jacobson, MD; Sherye A. Sirrel, MS, CCRC; Howard University: Thomas O. Obisesan, MD, MPH; Saba Wolday, MSc; Joanne Allard, PhD; University of Wisconsin: Sterling C. Johnson, Ph.D. J. Jay Fruehling, MA; Sandra Harding, MS; University of Washington: Elaine R. Peskind, MD; Eric C. Petrie, MD, MS; Gail Li, MD, PhD; Stanford University: Jerome A. Yesavage, MD; Joy L. Taylor, PhD; Ansgar J. Furst, PhD; Steven Chao, MD; Cornell University: Norman Relkin, MD, PhD, Gloria Chiang, MD; Lisa Ravdin, PhD; ADNI Depression Part A: Leadership and Infrastructure Principal Investigator: Scott Mackin, PhD, University of California, San Francisco; ATRI PI and Director of Coordinating Center Clinical Core: Paul Aisen, MD, University of Southern California; Rema Raman, PhD, University of Southern California; Executive Committee: Scott Mackin, PhD, University of California, San Francisco; Michael Weiner, MD, University of California, San Francisco; Paul Aisen, MD, University of Southern California; Rema Raman, PhD, University of Southern California; Clifford R. Jack, Jr, MD, Mayo Clinic, Rochester; Susan Landau, PhD, University of California, Berkeley; Andrew J. Saykin, PsyD, Indiana

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University; Arthur W. Toga, PhD, University of Southern California; Charles DeCarli, MD, University of California, Davis; Robert A. Koeppe, PhD, University of Michigan; Data and Publication Committee (DPC): Robert C. Green, MD, MPH, Brigham and Women's Hospital/Harvard Medical School (chair); Erin Drake, MA, Brigham and Women's Hospital/Harvard Medical School (director); Clinical Core Leaders: Michael Weiner, MD (core principal investigator); Paul Aisen, MD, University of Southern California; Rema Raman, PhD, University of Southern California; Mike Donohue, PhD, University of Southern California; Clinical Informatics, Operations and Regulatory Affairs: Gustavo Jimenez, MBS, University of Southern California; Devon Gessert, BS, University of Southern California; Kelly Harless, BA, University of Southern California; Jennifer Salazar, MBS, University of Southern California; Yuliana Cabrera, BS, University of Southern California; Sarah Walter, MSc, University of Southern California; Lindsey Hergesheimer, BS, University of Southern California; Elizabeth Shaffer, BS; Psychiatry Site Leaders and Key Personnel: Scott Mackin, PhD, University of California, San Francisco; Craig Nelson, MD, University of California, San Francisco; David Bickford, BA, University of California, San Francisco; Meryl Butters, PhD, University of Pittsburgh; Michelle Zmuda, MA, University of Pittsburgh; MRI Core Leaders and Key Personnel: Clifford R. Jack, Jr, MD, Mayo Clinic, Rochester (core principal investigator); Matthew Bernstein, PhD, Mayo Clinic, Rochester; Bret Borowski, RT, Mayo Clinic, Rochester; Jeff Gunter, PhD, Mayo Clinic, Rochester; Matt Senjem, MS, Mayo Clinic, Rochester; Kejal Kantarci, MD, Mayo Clinic, Rochester; Chad Ward, BA, Mayo Clinic, Rochester; Denise Reyes, BS, Mayo Clinic, Rochester; PET Core Leaders and Key Personnel: Robert A. Koeppe, PhD, University of Michigan Susan Landau, PhD, University of California, Berkeley; Informatics Core Leaders and Key Personnel: Arthur W. Toga, PhD, University of Southern California (core principal investigator); Karen Crawford, University of Southern California; S Neu, PhD, University of Southern California; Genetics Core Leaders and Key Personnel: Andrew J. Saykin, PsyD, Indiana University; Tatiana M. Foroud, PhD, Indiana University; Kelley M. Faber, MS, CCRC, Indiana University; Kwangsik Nho, PhD, Indiana University; Kelly N. Nudelman, Indiana University; Part B: Investigators by Site: University of California, San Francisco: Scott Mackin, PhD; Howard Rosen, MD; Craig Nelson, MD; David Bickford, BA; Yiu Ho Au, BA; Kelly Scherer, BS; Daniel Catalinotto, BA; Samuel Stark, BA; Elise Ong, BA; Dariella Fernandez, BA; University of Pittsburgh: Meryl Butters, PhD; Michelle Zmuda, MA; Oscar L. Lopez, MD; MaryAnn Oakley, MA; Donna M. Simpson, CRNP, MPH.

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Key Points

Question

Which of the recently validated Alzheimer disease genetic risk variants are associated with brain amyloidosis?

Findings

In this study of 977 individuals from the Alzheimer's Disease Neuroimaging Initiative, the adenosine triphosphate–binding cassette subfamily A member 7 gene had the strongest association with brain

amyloidosis after apolipoprotein E ε 4. The fermitin family homologue 2 gene had a stage-dependent association with brain amyloidosis, which was most pronounced in the mild cognitive impairment stage.

Conclusions

This study found an association of AD risk variants with brain amyloidosis.

Abstract

Importance

Late-onset Alzheimer disease (AD) is highly heritable. Genome-wide association studies have identified more than 20 AD risk genes. The precise mechanism through which many of these genes are associated with AD remains unknown.

Objective

To investigate the association of the top 20 AD risk variants with brain amyloidosis.

Design, Setting, and Participants

This study analyzed the genetic and florbetapir F 18 data from 322 cognitively normal control individuals, 496 individuals with mild cognitive impairment, and 159 individuals with AD dementia who had genomewide association studies and ¹⁸F-florbetapir positron emission tomographic data from the Alzheimer's Disease Neuroimaging Initiative (ADNI), a prospective, observational, multisite tertiary center clinical and biomarker study. This ongoing study began in 2005.

Main Outcomes and Measures

The study tested the association of AD risk allele carrier status (exposure) with florbetapir mean standard uptake value ratio (outcome) using stepwise multivariable linear regression while controlling for age, sex, and apolipoprotein E ε 4 genotype. The study also reports on an exploratory 3-dimensional stepwise regression model using an unbiased voxelwise approach in Statistical Parametric Mapping 8 with cluster and significance thresholds at 50 voxels and uncorrected *P* < .01.

Results

This study included 977 participants (mean [SD] age, 74 [7.5] years; 535 [54.8%] male and 442 [45.2%] female) from the ADNI-1, ADNI-2, and ADNI–Grand Opportunity. The adenosine triphosphate–binding cassette subfamily A member 7 (*ABCA7*) gene had the strongest association with amyloid deposition ($\chi^2 = 8.38$, false discovery rate–corrected *P* < .001), after apolioprotein E ε 4. Significant associations were found between *ABCA7* in the asymptomatic and early symptomatic disease stages, suggesting an association with rapid amyloid accumulation. The fermitin family homolog 2 (*FERMT2*) gene had a stage-dependent association with brain amyloidosis (*FERMT2* × diagnosis $\chi^2 = 3.53$, false discovery rate–corrected *P* = .05), which was most pronounced in the mild cognitive impairment stage.

Conclusions and Relevance

This study found an association of several AD risk variants with brain amyloidosis. The data also suggest

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Associations of the Top 20 Alzheimer Disease Risk Variants With Brain Amyloidosis

that AD genes might differentially regulate AD pathologic findings across the disease stages.

Introduction

Sporadic Alzheimer disease (AD) is 70% to 80% heritable. The strongest genetic risk factor for AD is the apolipoprotein E (*APOE*) gene (OMIM <u>107741</u>). The *APOE* ϵ 4 allele carries the greatest risk through the reduction of β -amyloid (A β) clearance. *APOE* ϵ 4 carriers have a significantly higher prevalence of Pittsburgh compound B uptake than noncarriers across all disease stages, including presymptomatic amyloidosis in cognitively normal control individuals. Peripheral blood apoE protein levels correlate with amyloid positron emission tomography (PET) binding. These data indicate that imaging phenotypes can provide meaningful information related to gene function and pathophysiologic findings.

Previous large-scale genome-wide association studies (GWASs) have identified and validated 20 novel AD genetic risk loci. Few of these loci are in or near genes associated with A β aggregation and clearance and are thought to influence amyloid deposition. For the remainder, the precise disease-associated mechanism remains unknown.

Several imaging genetics studies have reported associations of some of the AD risk genes with brain amyloidosis or neurodegeneration. Phosphatidylinositol-binding clathrin assembly protein (PICALM) (OMIM <u>603025</u>) <u>rs3851179</u>, bridging integrator 1 (BIN1) (OMIM <u>601248</u>) <u>rs7561528</u>, complement component receptor 1 (CR1) rs1408077 (OMIM 120620), adenosine triphosphate-binding cassette subfamily A member 7 (ABCA7) (OMIM 605414) rs3764650, and membrane-spanning 4-domains, subfamily A, member 6a (MS4A6A) (OMIM 606548) rs610932 are associated with cortical and hippocampal atrophy. ABCA7 rs3764650 and rs3752246; BIN1 rs744373; CR1 rs6701713, rs3818361, and rs6656401; and clusterin (CLU) rs3818361 (OMIM 185430) are associated with amyloid deposition. Although these studies enrich the imaging genetics field, they also have significant shortcomings. Many of these research studies have focused on a single variant or a few variants while ignoring the complex polygenic disease background. In addition, all analyses of gene-endophenotype associations to date have largely used averaged phenotypic records across all disease stages. Such an approach is justified if the risk variant has a static or conserved effect during the disease course. However, considering the complicated and constantly evolving disease pathophysiologic process with early amyloid deposition, later onset of neuronal degeneration, and variable degree of inflammation, we considered stage-dependent genetic associations. Furthermore, improved understanding of the polygenetic risk factors for AD could enable personalized risk assessment, whereas an in-depth characterization of disease-associated mechanism could lead to new therapeutic avenues.

We report a comprehensive analysis of the associations of all well-validated AD risk variants with brain amyloidosis. Our goal was to establish their relative contribution to the amyloid burden. We hypothesized that our multivariable analytic approach would help us more accurately model the probability distribution of our imaging outcome measure and that we would detect several genetic variants in addition to *APOE* ϵ 4 that are associated with brain amyloidosis. In addition, we hypothesized that we might also find stage-dependent associations with amyloid accumulation.

Methods

Participants

Data used in this study were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI)

database (<u>http://adni.loni.usc.edu</u>). The ADNI is a longitudinal study with approximately 50 sites across the United States and Canada that was launched in 2003 (<u>http://adni.loni.usc.edu</u>). The goal of the ADNI is to track the progression of AD by using clinical and cognitive tests, magnetic resonance imaging (MRI), fludeoxyglucose PET, amyloid PET, cerebrospinal fluid, and blood biomarkers. The institutional review boards of all sites participating in the ADNI provided review and approval of the ADNI data collection protocol.

The clinical description of the ADNI cohort has been previously published. Diagnosis of AD was based on the National Institute of Neurological and Communicative Disorders and Stroke and the Alzheimer's Disease and Related Disorders Association criteria. Individuals with AD dementia were required to have Mini-Mental State Examination (MMSE) scores between 20 and 26 and a Clinical Dementia Rating (CDR) score of 0.5 to 1 at baseline. Qualifying individuals with mild cognitive impairment (MCI) had memory concerns but no significant functional impairment, scored between 24 and 30 on the MMSE, had a global CDR score of 0.5, had a CDR memory score of 0.5 or greater, and had objective memory impairment on the Wechsler Memory Scale–Logical Memory II test. The controls had MMSE scores between 24 and 30, had a global CDR score of 0, and did not meet criteria for MCI and AD. Individuals were excluded if they refused or were unable to undergo MRI; had other neurologic disorders, active depression, a history of psychiatric diagnosis, a history of alcohol or other substance dependence within the past 2 years; had less than 6 years of education; or were not fluent in English or Spanish. The full list of inclusion and exclusion criteria can be accessed on pages 23 to 29 of the online ADNI protocol (http://adni.loni.usc.edu/wp_content/uploads/2010/09/ADNI_GeneralProceduresManual.pdf). Written informed consent was obtained from all participants, and all data were deidentified.

Gene Variant Selection and Imputation

The ADNI-1 participants were genotyped using the Illumina Human610-Quad BeadChip array (Illumina Inc), whereas the ADNI-2 and the ADNI–Grand Opportunity (GO) participants were genotyped using the Illumina HumanOmniExpress BeadChip (Illumina Inc) according to the manufacturer's protocol. We focused on the 20 well-established AD risk genes identified and validated in the largest AD GWASs to date. In addition to the variants reported in these articles, we included all other variants that were previously associated with brain amyloidosis (eTable 1 in the <u>Supplement</u>), which yielded a total of 36 variants.

Missing genotypes (eTable 2 in the <u>Supplement</u>) were imputed using MACH and minimac in a 2-stage procedure using the 1000 Genomes project pilot data as a reference panel. Minimac yielded the posterior probabilities of the imputed genotypes at ungenotyped marker loci for each individual. The threshold to accept each imputed genotype was set at $r^2 = 0.30$.

Nine genes were represented by more than 1 single-nucleotide polymorphism (SNP). Because linkage disequilibrium (LD) introduces colinearity bias, we performed LD analyses followed by Cohen κ statistics (eFigure 1 and eTable 3 in the <u>Supplement</u>). When choosing between 2 variants with significant overlap (high LD and high κ), we retained the variant with least data missingness. Our final number of variants was thus reduced to 27. *ABCA7, BIN1, CLU, CR1,* ephrin receptor EphA1 (*EPHA1*) (OMIM <u>179610</u>), and sortilin-related receptor (*SORL1*) (OMIM <u>602005</u>) were represented with more than 1 variant in the analyses (eTable 3 in the <u>Supplement</u>).

Allele frequencies for each gene variant were assessed. Genotypes were collapsed when the minor allele homozygote frequency was less than 2% as follows: *ABCA7* <u>rs3764650</u> GG/GT vs TT, Cass scaffolding

protein family member 4 (*CASS4*) (HGNC <u>15878</u>) <u>rs7274581</u> CC/TC vs TT, *CLU* <u>rs9331949</u> AG/GG vs AA, desmoglein 2 (*DSG2*) (OMIM <u>125671</u>) <u>rs8093731</u> TT/TC vs CC, fermitin family homologue 2 *FERMT2* (OMIM <u>607746</u>) <u>rs17125944</u> CC/TC vs TT, and *SORL1* <u>rs112183431</u> CC/TC vs TT. The remaining variants were coded by minor allele dosage.

Florbetapir F 18 PET Data Acquisition Protocol and Analyses

The florbetapir F 18 PET acquisition and preprocessing protocols are available at <u>http://www.adni-info.org</u>. In our main analyses, we used the mean whole-`brain standard uptake volume ratios (SUVRs) from University of California, Berkeley downloaded from the ADNI database (<u>http://adni.loni.usc.edu</u>). This variable was obtained by averaging the SUVRs obtained using whole cerebellum as the reference region across the frontal, anterior-posterior cingulate, lateral-parietal, and lateral-temporal gray matter regions. The University of California, Berkeley, protocols for ¹⁸F-florbetapir preprocessing, coregistration, and normalization have been previously described.

To visualize the regional pattern of associations in 3 dimensions, we downloaded all preprocessed ¹⁸Fflorbetapir data from the Laboratory of Neuroimaging Image Data Archive (<u>https://ida.loni.usc.edu</u>). We aligned the images to the corresponding MRI from the same visit, normalized to MNI space using measures obtained from the MRI spatial transformation and intensity normalized to the intensity of the whole cerebellum reference region to create SUVR images, as previously described.

Statistical Analysis

R Statistical Analyses Clinical and demographic characteristics (age, sex, educational level, MMSE, *APOE* $\varepsilon 4$ genotype, and diagnosis) for each variant were compared using *t* tests or χ^2 tests with 2-sided *P* values as appropriate. Stepwise multivariable linear regression models with all 27 AD risk variants were performed first in the pooled sample and second in each diagnostic category using amyloid PET mean SUVR as the outcome measure. An additional model in the pooled sample using only amyloid-positive individuals (SUVR>1.17) is available in the eResults in the <u>Supplement</u>. All regression models included age, sex, and *APOE* $\varepsilon 4$ genotype as covariates. The regression model for the pooled sample was also corrected for diagnosis. The decision to exclude variables was based on the Akaike information criterion critical *P* value threshold of .16. Because we included only previously validated candidate genes, our significance threshold was set at *P* < .05. Correction for false discovery rate (FDR) was applied.

Analyses in Imaging Space All imaging analyses were performed in an exploratory fashion. To explore the spatial distribution of the associations, we reproduced the final stepwise regression models using voxelwise regression in Statistical Parametric Mapping 8 (SPM8; Wellcome Department of Cognitive Neuroscience). The SPM8 models included all variants retained in the R statistical models (including those that were retained based on the Akaike information criterion) covaried for age, sex, and *APOE* ϵ 4 genotype. The pooled model also included diagnosis as a covariate. Because of the exploratory nature of our secondary results, we allowed a less stringent visualization threshold: voxelwise threshold of *P* < .01 (uncorrected) with a minimum cluster size (k) of 50 voxels. We also computed familywise error (FWE) and FDR-corrected cluster and peak statistics as appropriate.

Results

The study population was composed of participants from the ADNI-1, ADNI-2, and ADNI-GO stages and

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consisted of 322 controls, 496 individuals with MCI, and 159 individuals with AD who had available GWAS and ¹⁸F-florbetapir PET data (mean [SD] age, 74 [7.5] years; 535 [54.8%] male and 442 [45.2%] female). Group comparisons of demographic characteristics and distribution of the genotypes that were retained in the regression models are given in <u>Table 1</u>. *APOE* ϵ 4 had significant associations with brain amyloidosis (eFigure 2 in the <u>Supplement</u>). There were no significant differences in age, sex, educational level, MMSE score, and *APOE* ϵ 4 distribution between carriers and noncarriers or by allele dosage for any of the genotypes except for zinc finger CW-type and PWWP domain containing 1 (*ZCWPW1*) (HGNC 23486) for which risk allele homozygotes were less educated (*P* = .02).

Pooled Sample

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In the pooled sample, the stepwise linear regression model achieved an R^2 of 0.35 (95% CI, 0.33-0.37; P < .001). *ABCA7* rs3752246 ($\chi^2 = 8.38$, FDR-corrected P < .001), *EPHA1* rs11771145 ($\chi^2 = 4.08$, FDR-corrected P = .03), and *PICALM* rs3851179 ($\chi^2 = 3.67$, FDR-corrected P = .04) were significantly associated with mean SUVR in the pooled sample. Other associations were as follows: *ZCPWPW1* rs1476679 ($\chi^2 = 2.74$, FDR-corrected P = .08), *FERMT2* rs17125944 ($\chi^2 = 3.63$, FDR-corrected P = .08), and protein tyrosine-kinase 2 β (*PTK2B*) rs28834970 (OMIM 601212) ($\chi^2 = 2.52$, FDR-corrected P = .01). *ABCA7* rs3764650 and *CLU* rs11136000 were included in the model based on the Akaike selection criterion. A reduced model that included only age, sex, educational level, and *APOE* $\varepsilon 4$ achieved a reduced R^2 of 0.31 (95% CI, 0.29-0.33). The between-model difference in R^2 and reduced R^2 was 0.038 (95% CI, 0.029-0.047). Figure 1 and Figure 2 show these associations and Table 2 gives FWE- and FDR-corrected cluster-level results and within-cluster peak associations for genetic variants identified in our models.

Interaction Analyses

To further test for the presence of a stage-specific association, we conducted a linear regression analysis in the pooled sample including interaction terms. *FERMT2* was the only variant that had a significant interaction (*FERMT2* × diagnosis $\chi^2 = 3.53$, FDR-corrected P = .05). The effect sizes for the remaining genes remained unchanged. Figure 3 shows the β -coefficient maps of the main effect size of *FERMT2* and its interaction with diagnosis as well as the *FERMT2* effect size within each diagnostic group.

Exploratory Analyses Within Diagnostic Groups

In the control group, the model achieved an R^2 of 0.17 (95% CI, 0.14-0.21; P < .001; reduced $R^2 = 0.14$; 95% CI, 0.11-0.17; R^2 -reduced R^2 difference = 0.032; 95% CI, 0.015-0.05). Significant associations were seen for *PICALM* <u>rs3851179</u> ($\chi^2 = 3.56$, FDR-corrected P = .04). The association for *ABCA7* <u>rs3764650</u> was $\chi^2 = 3.16$ (FDR-corrected P = .09). *ABCA7* <u>rs3752246</u> was included in the model based on the Akaike selection criterion.

In the MCI group, the model achieved an R^2 of 0.3 (95% CI, 0.27-0.32; P < .001; reduced $R^2 = 0.24$; 95% CI, 0.21-0.27; R^2 -reduced R^2 difference = 0.058; 95% CI, 0.042-0.074). *ABCA7* rs3752246 ($\chi^2 = 7.22$, FDR-corrected P = .002), *EPHA1* rs11771145 ($\chi^2 = 3.74$, FDR-corrected P = .03), *FERMT2* rs17125944 ($\chi^2 = 10.38$, FDR-corrected P = .002), and *SORL1* rs1131497 ($\chi^2 = 3.66$, FDR-corrected P = .03) were significantly associated with mean SUVR. The association for *ABCA7* rs3764650 was $\chi^2 = 2.9$ (FDR-corrected P = .09).

In the dementia group, the model achieved an R^2 of 0.35 (95% CI, 0.29-0.41; P < .0001; reduced $R^2 = 0.22$;

JAWA Network

95% CI, 0.16-0.28; R^2 -reduced R^2 difference = 0.13; 95% CI, 0.09-0.17). Other associations were as follows: *EPHA1* rs11771145 (χ^2 = 5.05, FDR-corrected P = .01), *ZCWPW1* rs1476679 (χ^2 = 3.79, FDR-corrected P = .04), *DSG2* rs8093731 (χ^2 = 3.27, FDR-corrected P = .08), *CLU* rs9331949 (χ^2 = 4.09, FDR-corrected P = .058), and *SORL1* rs1131497 (χ^2 = 2.51, FDR-corrected P = .08).

Figure 1 and Figure 2 present exploratory visualization of these associations, and <u>Table 2</u> presents the FWEand FDR-corrected cluster-level results and within-cluster peak associations for genetic variants identified in our models.

Discussion

Improved understanding of the polygenetic risk factors that are associated with AD could enable personalized risk assessment. To our knowledge, this is the first comprehensive analysis of the association of the top 20 AD risk variants with brain amyloidosis. We were able to confirm the previously reported association between *ABCA7* and brain amyloidosis as described by Shulman et al and Hughes et al. Our study found that after *APOE* ε 4, *ABCA7* has the strongest association with amyloid deposition. We were unable to confirm the reported associations of *CR1* likely because the associations previously reported were determined using a univariable approach. It is plausible that the previously reported *CR1* association is better accounted for by other AD-related genes, which were not part of the original analysis. We also found evidence of a stage-dependent gene association of *FERMT2* with brain amyloidosis. This is, to our knowledge, the first report of such an association.

Several genes had associations with brain amyloidosis. *ABCA7* encodes a 2146–amino acid ABC family transporter protein. The ABC protein family is responsible for the transport of a variety of molecules across cellular membranes, primarily lipids. *ABCA7* is expressed in nervous tissue, with the highest expression in microglia. Loss of function of *ABCA7* was associated with increased β -secretase cleavage of amyloid precursor protein (APP), leading to higher levels of A β in vitro and in vivo. A previous ADNI study analyzed the associations of 15 *ABCA7* loci with cerebrospinal fluid A β and florbetapir SUVR. Three variants (rs3752242, rs3752240, and rs4147912) were significantly associated with brain amyloidosis but not with brain atrophy. One of these 3 SNPs (rs3752242) is in LD with *ABCA7* rs3752246, lending support to our findings. Further evidence of the role of *ABCA7* in AD comes from a study that reported one rare missense variant (rs72973581; minor allele frequency of 4.3%) to confer a significant protection against AD. In a previous publication, a late but profound effect of *ABCA7* was found on neurodegeneration. Individuals with AD dementia had significant associations of *ABCA7* rs3752246 with gray matter density throughout the brain. Individuals with MCI and controls did not have such an association.

CLU encodes for clusterin, an extracellular chaperone protein that consists of 427 amino acids. CLU is highly expressed in neurons and ependymal cells. It seems to be involved in a variety of processes throughout the body, including synaptic maintenance and programmed cell death. Under physiologic conditions, clusterin reduces aggregation and promotes clearance of A β . CLU is highly expressed in the hippocampi in patients with AD and Pick disease. Clusterin protein levels are also elevated in AD, and its pattern of distribution correlates positively with that of A β 42 and A β 40 in postmortem tissue.

DSG2 encodes a cell adhesion desmosome cadherin protein. *DSG2* binds plaque proteins and intermediate filaments and seems to play a role in inflammation. Although this gene was reported to be associated with AD risk, a mechanistic explanation of this association has not yet been elucidated. *DSG2* is expressed in epithelial-derived tissues, such as epithelial cell lines, epithelial malignant tumors, and the brain, especially

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the corpus callosum region. We found an association with amyloid deposition later in the disease course, indicating a late modulatory effect on amyloid deposition.

EPHA1 encodes a 976–amino acid protein that belongs to the EPH family of receptor tyrosine kinases. *EPHA1* plays a role in contact-dependent signaling and nervous system development. *EPHA1* is highly expressed in the cerebral cortex and hippocampus. A previous analysis of ADNI-1 data reported that *EPHA1* <u>rs11771145</u> is associated with less brain atrophy and higher cerebral metabolic rate in MCI. Analyses of the cognitively normal imaging subcohort of the Ginkgo Evaluation of Memory study implicated another *EPHA1* allele (<u>rs11767557</u>), which is in LD with ours, to have a negative effect on brain amyloidosis.

FERMT2 encodes for a 680–amino acid scaffolding extracellular matrix protein that plays a role in cell adhesions. *FERMT2* is expressed in the brain (http://www.proteinatlas.org/ENSG00000073712-<u>FERMT2/tissue</u>). *FERMT2* is upregulated in atherosclerotic plaques, suggesting a possible role in inflammation and leukocyte extravasation. *FERMT2* is a coactivator of β 3-integrin—a microglial and reactive astrocyte marker that plays a role in poststroke brain tissue recovery. *FERMT2* has also been associated with a cognitive decline in AD and modifies tau neurotoxicity in a *Drosophila* model.

PICALM encodes a 652–amino acid protein that binds to clathrin's heavy chain and assists in vesicle assembly and endocytosis. *PICALM* was recently identified as a risk gene for late-onset AD. PICALM colocalizes with APP. *PICALM* knockdown resulted in a reduction in the amount of APP internalized and a reduction in A β generation. In a previous study, *PICALM* was found to modulate the clearance of tau and thus autophagy. *PICALM* has been associated with brain changes in AD. Morgen et al reported a negative association with prefrontal brain volume and working memory, whereas Biffi et al found associations with hippocampal amygdalar and white matter lesion volume, as well as with entorhinal, parahippocampal, and temporal pole cortical thickness.

SORL1 encodes a 2186–amino acid protein from the low-density lipoprotein receptor family. SORL1 readily binds APOE and lipoprotein lipase and localizes to both the Golgi apparatus and the plasma membrane, where it likely mediates endocytosis. *SORL1* plays a role in APP trafficking and recycling. *SORL1* is downregulated in lymphoblasts and cortical pyramidal neurons of patients with AD. The neuronal SORL1 protein level determines cognitive decline and conversion from MCI to AD. The protein level also correlates with the levels of the APP soluble products that result from β -secretase cleavage. An SNP in LD with our variant (rs1133174) has also been linked to brain atrophy in AD.

The *ZCWPW1* gene codes for a 648–amino acid protein. *ZCWPW1* is considered to be a risk gene for lateonset AD. Its proposed mechanism of action is through epigenetic regulation of gene expression.

Strengths and Limitations

Several strengths and limitations of our study warrant discussion. One of the major strengths lies in the careful clinical, biomarker, and genetic characterization of all individuals enrolled in the ADNI. The ADNI protocol uses unified subject assessment, standardization of all imaging, biofluid and DNA and RNA data collection and processing, and meticulous data quality control across all study sites. Another strength of the study is the fairly large sample size that allowed us to achieve enough statistical power to test the associations of 27 AD-associated risk variants using a polygenic model.

A major limitation of our study is that we only report cross-sectional analyses; thus, we cannot make

definitive conclusions regarding genetic effects on amyloid deposition over time. From our cross-sectional observations across the disease continuum, we drew conclusions about early vs late genetic influences on brain amyloidosis that will need to be further tested using a longitudinal design, which is what we plan to do next. Another limitation of our work is that the sample size was not big enough to allow us to test for gene-gene and gene-environment interactions. Last but not least, the ADNI uses rigorous exclusion criteria typical of clinical trials, rendering the ADNI cohort not representative of the general population, which may negatively affect the generalizability of our results. Thus, our next steps will be to validate our findings in a large, independent, longitudinal cohort.

Conclusions

We found an association of genetic variants with brain amyloidosis, the salient pathognomonic feature of AD. Four of the genetic variants reported here, *ABCA7*, *CLU*, *EPHA1*, and *SORL1*, have been previously linked to the amyloidogenic AD pathways. To our knowledge, we are the first to report a stage-specific association for a genetic variant (ie, *FERMT2*).

Notes

Supplement.

eTable 1. All Variants of the Top 20 AD Risk Genes That Were Considered for Inclusion in Our Analyses

eTable 2. List of Variants That Needed Partial or Full Imputation

eTable 3. Linkage Disequilibrium and κ Statistics Results for the Variants Retained in Our Analyses

eFigure 1. Linkage Disequilibrium Results

eFigure 2. Association of Apolipoprotein E (APOE) E4 With Brain Amyloidosis

eResults. Amyloid-Positive Pooled Sample

References

1. Wingo TS, Lah JJ, Levey AI, Cutler DJ. Autosomal recessive causes likely in early-onset Alzheimer disease. Arch Neurol. 2012;69(1):59-64. [PMCID: PMC3332307] [PubMed: 21911656]

2. Sleegers K, Lambert JC, Bertram L, Cruts M, Amouyel P, Van Broeckhoven C. The pursuit of susceptibility genes for Alzheimer's disease: progress and prospects. Trends Genet. 2010;26(2):84-93. [PubMed: 20080314]

3. Riddell DR, Zhou H, Atchison K, et al. Impact of apolipoprotein E (ApoE) polymorphism on brain ApoE levels. J Neurosci. 2008;28(45):11445-11453. [PubMed: 18987181]

4. Deane R, Sagare A, Hamm K, et al. apoE isoform-specific disruption of amyloid beta peptide clearance from mouse brain. J Clin Invest. 2008;118(12):4002-4013. [PMCID: PMC2582453] [PubMed: 19033669]

5. Kim J, Basak JM, Holtzman DM. The role of apolipoprotein E in Alzheimer's disease. Neuron. 2009;63(3):287-303. [PMCID: PMC3044446] [PubMed: 19679070]

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6. Fleisher AS, Chen K, Liu X, et al. Using positron emission tomography and florbetapir F18 to image cortical amyloid in patients with mild cognitive impairment or dementia due to Alzheimer disease. Arch Neurol. 2011;68(11):1404-1411. [PubMed: 21747008]

7. Reiman EM, Chen K, Liu X, et al. Fibrillar amyloid-beta burden in cognitively normal people at 3 levels of genetic risk for Alzheimer's disease. Proc Natl Acad Sci U S A. 2009;106(16):6820-6825. [PMCID: PMC2665196] [PubMed: 19346482]

 Lazaris A, Hwang KS, Goukasian N, et al. ; Alzheimer's Disease Neuroimaging Initiative . Alzheimer risk genes modulate the relationship between plasma apoE and cortical PiB binding. Neurol Genet. 2015;1(3):e22. [PMCID: PMC4809461] [PubMed: 27066559]

9. Gupta VB, Laws SM, Villemagne VL, et al. ; AIBL Research Group . Plasma apolipoprotein E and Alzheimer disease risk: the AIBL study of aging. Neurology. 2011;76(12):1091-1098. [PubMed: 21422459]

Harold D, Abraham R, Hollingworth P, et al. Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease [published correction appears in *Nat Genet*. 2009;45(6):712]. Nat Genet. 2009;41(10):1088-1093. [PMCID: PMC2845877] [PubMed: 19734902]

11. Hollingworth P, Harold D, Sims R, et al. ; Alzheimer's Disease Neuroimaging Initiative; CHARGE consortium; EADI1 consortium . Common variants at ABCA7, MS4A6A/MS4A4E, EPHA1, CD33 and CD2AP are associated with Alzheimer's disease. Nat Genet. 2011;43(5):429-435. [PMCID: PMC3084173] [PubMed: 21460840]

12. Lambert JC, Heath S, Even G, et al. ; European Alzheimer's Disease Initiative Investigators . Genomewide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. Nat Genet. 2009;41(10):1094-1099. [PubMed: 19734903]

 Lambert JC, Ibrahim-Verbaas CA, Harold D, et al. Meta-analysis of 74,538 individuals identifies 11 new susceptibility loci for Alzheimer's disease. *Nat Genet.* 2013;45(12):1452-1458.
[PMCID: PMC3896259] [PubMed: 24162737]

14. Naj AC, Jun G, Beecham GW, et al. Common variants at MS4A4/MS4A6E, CD2AP, CD33 and EPHA1 are associated with late-onset Alzheimer's disease. Nat Genet. 2011;43(5):436-441. [PMCID: PMC3090745] [PubMed: 21460841]

 Seshadri S, Fitzpatrick AL, Ikram MA, et al. ; CHARGE Consortium; GERAD1 Consortium; EADI1 Consortium . Genome-wide analysis of genetic loci associated with Alzheimer disease. JAMA.
2010;303(18):1832-1840. [PMCID: PMC2989531] [PubMed: 20460622]

16. Shulman JM, Chen K, Keenan BT, et al. Genetic susceptibility for Alzheimer disease neuritic plaque pathology. JAMA Neurol. 2013;70(9):1150-1157. [PMCID: PMC3773291] [PubMed: 23836404]

17. Hohman TJ, Koran ME, Thornton-Wells T; Alzheimer's Neuroimaging Initiative . Epistatic genetic effects among Alzheimer's candidate genes. PLoS One. 2013;8(11):e80839. [PMCID: PMC3832488] [PubMed: 24260488]

Hughes TM, Lopez OL, Evans RW, et al. Markers of cholesterol transport are associated with amyloid deposition in the brain. Neurobiol Aging. 2014;35(4):802-807. [PMCID: PMC3896052] [PubMed: 24199960]

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19. Thambisetty M, An Y, Nalls M, et al. ; Baltimore Longitudinal Study of Aging and the Alzheimer's Disease Neuroimaging Initiative . Effect of complement CR1 on brain amyloid burden during aging and its modification by APOE genotype. Biol Psychiatry. 2013;73(5):422-428. [PMCID: PMC3535537] [PubMed: 23022416]

20. Chibnik LB, Shulman JM, Leurgans SE, et al. CR1 is associated with amyloid plaque burden and agerelated cognitive decline. Ann Neurol. 2011;69(3):560-569. [PMCID: PMC3066288] [PubMed: 21391232]

21. Braskie MN, Ringman JM, Thompson PM. Neuroimaging measures as endophenotypes in Alzheimer's disease. *Int J Alzheimers Dis* 2011;2011:490140. [PMCID: PMC3087508] [PubMed: 21547229]

22. Ramirez LM, Goukasian N, Porat S, et al. Common variants in ABCA7 and MS4A6A are associated with cortical and hippocampal atrophy. Neurobiol Aging. 2016;39:82-89. [PubMed: 26923404]

23. Saykin AJ, Shen L, Foroud TM, et al. ; Alzheimer's Disease Neuroimaging Initiative . Alzheimer's Disease Neuroimaging Initiative biomarkers as quantitative phenotypes: genetics core aims, progress, and plans. Alzheimers Dement. 2010;6(3):265-273. [PMCID: PMC2868595] [PubMed: 20451875]

24. Potkin SG, Guffanti G, Lakatos A, et al. ; Alzheimer's Disease Neuroimaging Initiative . Hippocampal atrophy as a quantitative trait in a genome-wide association study identifying novel susceptibility genes for Alzheimer's disease. PLoS One. 2009;4(8):e6501. [PMCID: PMC2719581] [PubMed: 19668339]

25. Morris JC, Roe CM, Xiong C, et al. APOE predicts amyloid-beta but not tau Alzheimer pathology in cognitively normal aging. Ann Neurol. 2010;67(1):122-131. [PMCID: PMC2830375] [PubMed: 20186853]

26. Petersen RC, Aisen PS, Beckett LA, et al. Alzheimer's Disease Neuroimaging Initiative (ADNI): clinical characterization. Neurology. 2010;74(3):201-209. [PMCID: PMC2809036] [PubMed: 20042704]

27. Petersen RC, Negash S. Mild cognitive impairment: an overview. CNS Spectr. 2008;13(1):45-53. [PubMed: 18204414]

28. Petersen RCS, Smith GE, Waring SC, Ivnik RJ, Tangalos EG, Kokmen E. Mild cognitive impairment: clinical characterization and outcome. Arch Neurol. 1999;56(3):303-308. [PubMed: 10190820]

29. McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlan EM. Clinical diagnosis of Alzheimer's disease: report of the NINCDS-ADRDA Work Group under the auspices of Department of Health and Human Services Task Force on Alzheimer's Disease. Neurology. 1984;34(7):939-944. [PubMed: 6610841]

30. Dubois B, Feldman HH, Jacova C, et al. Research criteria for the diagnosis of Alzheimer's disease: revising the NINCDS-ADRDA criteria. Lancet Neurol. 2007;6(8):734-746. [PubMed: 17616482]

31. Jack CRJA Jr, Albert MS, Knopman DS, et al. Introduction to the recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimers Dement. 2011;7(3):257-262. [PMCID: PMC3096735] [PubMed: 21514247]

32. Folstein MFF, Folstein SE, McHugh PR. "Mini-mental state": a practical method for grading the cognitive state of patients for the clinician. J Psychiatr Res. 1975;12(3):189-198. [PubMed: 1202204]

33. Morris JC. The Clinical Dementia Rating (CDR): current version and scoring rules. Neurology. 1993;43(11):2412-2414. [PubMed: 8232972]

Associations of the Top 20 Alzheimer Disease Risk Variants With Brain Amyloidosis

JAWA Network:

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34. D. W. Wechsler Memory Scale-Revised. San Antonio, TX: Psychological Corp; 1987.

35. Nho K, Corneveaux JJ, Kim S, et al. ; Multi-Institutional Research on Alzheimer Genetic Epidemiology (MIRAGE) Study; AddNeuroMed Consortium; Indiana Memory and Aging Study; Alzheimer's Disease Neuroimaging Initiative (ADNI) Whole-exome sequencing and imaging genetics identify functional variants for rate of change in hippocampal volume in mild cognitive impairment. Mol Psychiatry. 2013;18(7):781-787. [PMCID: PMC3777294] [PubMed: 23608917]

36. Jagust WJ, Bandy D, Chen K, et al. ; Alzheimer's Disease Neuroimaging Initiative . The Alzheimer's Disease Neuroimaging Initiative positron emission tomography core. Alzheimers Dement. 2010;6(3):221-229. [PMCID: PMC2920531] [PubMed: 20451870]

37. Risacher SL, Kim S, Shen L, et al. ; Alzheimer's Disease Neuroimaging Initiative (ADNI) The role of apolipoprotein E (APOE) genotype in early mild cognitive impairment (E-MCI). Front Aging Neurosci. 2013;5:11. [PMCID: PMC3612590] [PubMed: 23554593]

38. Akaike H. Information theory and an extension of the maximum likelihood principle In: Parzen E, Tanabe K, Kitagawa G., eds. Selected Papers of Hirotugu Akaike. Springer Series in Statistics (Perspectives in Statistics). New York, NY: Springer; 1998.

39. Weiner MW, Veitch DP, Aisen PS, et al. ; Alzheimer's Disease Neuroimaging Initiative . The Alzheimer's Disease Neuroimaging Initiative: a review of papers published since its inception. Alzheimers Dement. 2012;8(1)(suppl):S1-S68. [PMCID: PMC3329969] [PubMed: 22047634]

40. Nuchnoi P, Nantakomol D, Chumchua V, Plabplueng C, Isarankura-Na-Ayudhya C, Prachayasittikul V. The Identification of functional non-synonymous SNP in human ATP-binding cassette (ABC), subfamily member 7 gene: application of bioinformatics tools in biomedicine. J Bioanal Biomed. 2011;3(1):26-31.

41. Li H, Karl T, Garner B. Understanding the function of ABCA7 in Alzheimer's disease. Biochem Soc Trans. 2015;43(5):920-923. [PubMed: 26517904]

42. Satoh K, Abe-Dohmae S, Yokoyama S, St George-Hyslop P, Fraser PE. ATP-binding cassette transporter A7 (ABCA7) loss of function alters Alzheimer amyloid processing. J Biol Chem. 2015;290(40):24152-24165. [PMCID: PMC4591804] [PubMed: 26260791]

43. Zhao QF, Wan Y, Wang HF, et al. ; Alzheimer's Disease Neuroimaging Initiative . ABCA7 genotypes confer Alzheimer's disease risk by modulating amyloid-β pathology. J Alzheimers Dis. 2016;52(2):693-703. [PubMed: 27003212]

44. Sassi C, Nalls MA, Ridge PG, et al. ; ARUK Consortium . ABCA7 p.G215S as potential protective factor for Alzheimer's disease. Neurobiol Aging. 2016;46:235.e1-235.e9. [PMCID: PMC5024078] [PubMed: 27289440]

45. Stage E, Duran T, Risacher SL, et al. The effect of the top 20 Alzheimer disease risk genes on graymatter density and FDG PET brain metabolism. Alzheimers Dement (Amst). 2016;5:53-66. [PMCID: PMC5198883] [PubMed: 28054028]

46. Murphy BF, Kirszbaum L, Walker ID, d'Apice AJ. SP-40,40, a newly identified normal human serum protein found in the SC5b-9 complex of complement and in the immune deposits in glomerulonephritis. J Clin Invest. 1988;81(6):1858-1864. [PMCID: PMC442636] [PubMed: 2454950]

47. Danik M, Chabot JG, Hassan-Gonzalez D, Suh M, Quirion R. Localization of sulfated glycoprotein-2/clusterin mRNA in the rat brain by in situ hybridization. J Comp Neurol. 1993;334(2):209-227. [PubMed: 8366194]

48. Wong P, Pineault J, Lakins J, et al. Genomic organization and expression of the rat TRPM-2 (clusterin) gene, a gene implicated in apoptosis. J Biol Chem. 1993;268(7):5021-5031. [PubMed: 7680346]

49. Beeg M, Stravalaci M, Romeo M, et al. Clusterin binds to Aβ1-42 oligomers with high affinity and interferes with peptide aggregation by inhibiting primary and secondary nucleation. J Biol Chem. 2016;291(13):6958-6966. [PMCID: PMC4807280] [PubMed: 26884339]

50. Duguid JR, Bohmont CW, Liu NG, Tourtellotte WW. Changes in brain gene expression shared by scrapie and Alzheimer disease. Proc Natl Acad Sci U S A. 1989;86(18):7260-7264. [PMCID: PMC298037] [PubMed: 2780570]

51. Miners JS, Clarke P, Love S. Clusterin levels are increased in Alzheimer's disease and influence the regional distribution of Aβ. Brain Pathol. 2017;27(3):305-313. [PubMed: 27248362]

52. Kamekura R, Nava P, Feng M, et al. Inflammation-induced desmoglein-2 ectodomain shedding compromises the mucosal barrier. Mol Biol Cell. 2015;26(18):3165-3177. [PMCID: PMC4569309] [PubMed: 26224314]

53. Koch PJ, Goldschmidt MD, Walsh MJ, Zimbelmann R, Franke WW. Complete amino acid sequence of the epidermal desmoglein precursor polypeptide and identification of a second type of desmoglein gene. Eur J Cell Biol. 1991;55(2):200-208. [PubMed: 1935985]

54. Wang H, Li ZY, Liu Y, et al. Desmoglein 2 is a receptor for adenovirus serotypes 3, 7, 11 and 14. Nat Med. 2011;17(1):96-104. [PMCID: PMC3074512] [PubMed: 21151137]

55. Besco JA, Hooft van Huijsduijnen R, Frostholm A, Rotter A. Intracellular substrates of brain-enriched receptor protein tyrosine phosphatase rho (RPTPrho/PTPRT). Brain Res. 2006;1116(1):50-57. [PubMed: 16973135]

56. Kljuic A, Christiano AM. A novel mouse desmosomal cadherin family member, desmoglein 1 gamma. Exp Dermatol. 2003;12(1):20-29. [PubMed: 12631243]

57. Miyata S, Yoshikawa K, Taniguchi M, et al. Sgk1 regulates desmoglein 1 expression levels in oligodendrocytes in the mouse corpus callosum after chronic stress exposure. Biochem Biophys Res Commun. 2015;464(1):76-82. [PubMed: 26043694]

58. Maru Y, Hirai H, Yoshida MC, Takaku F. Evolution, expression, and chromosomal location of a novel receptor tyrosine kinase gene, eph. Mol Cell Biol. 1988;8(9):3770-3776. [PMCID: PMC365435] [PubMed: 3221865]

59. Yamazaki T, Masuda J, Omori T, Usui R, Akiyama H, Maru Y. EphA1 interacts with integrin-linked kinase and regulates cell morphology and motility. J Cell Sci. 2009;122(Pt 2):243-255. [PubMed: 19118217]

60. Chen G, Wang Y, Zhou M, et al. EphA1 receptor silencing by small interfering RNA has antiangiogenic and antitumor efficacy in hepatocellular carcinoma. Oncol Rep. 2010;23(2):563-570. [PubMed: 20043122]

61. Torii M, Hashimoto-Torii K, Levitt P, Rakic P. Integration of neuronal clones in the radial cortical

JAWA Network:

columns by EphA and ephrin-A signalling. Nature. 2009;461(7263):524-528. [PMCID: PMC2874978] [PubMed: 19759535]

62. Hattori M, Osterfield M, Flanagan JG. Regulated cleavage of a contact-mediated axon repellent. Science. 2000;289(5483):1360-1365. [PubMed: 10958785]

63. Noberini R, Rubio de la Torre E, Pasquale EB. Profiling Eph receptor expression in cells and tissues: a targeted mass spectrometry approach. Cell Adh Migr. 2012;6(2):102-112. [PMCID: PMC3499309] [PubMed: 22568954]

64. Wang HF, Tan L, Hao XK, et al. ; Alzheimer's Disease Neuroimaging Initiative . Effect of EPHA1 genetic variation on cerebrospinal fluid and neuroimaging biomarkers in healthy, mild cognitive impairment and Alzheimer's disease cohorts. J Alzheimers Dis. 2015;44(1):115-123. [PubMed: 25182741]

65. Weinstein EJ, Bourner M, Head R, Zakeri H, Bauer C, Mazzarella R. URP1: a member of a novel family of PH and FERM domain-containing membrane-associated proteins is significantly over-expressed in lung and colon carcinomas. Biochim Biophys Acta. 2003;1637(3):207-216. [PubMed: 12697302]

66. Tu Y, Wu S, Shi X, Chen K, Wu C. Migfilin and Mig-2 link focal adhesions to filamin and the actin cytoskeleton and function in cell shape modulation. Cell. 2003;113(1):37-47. [PubMed: 12679033]

67. Oksala N, Pärssinen J, Seppälä I, et al. Kindlin 3 (FERMT3) is associated with unstable atherosclerotic plaques, anti-inflammatory type II macrophages and upregulation of beta-2 integrins in all major arterial beds. Atherosclerosis. 2015;242(1):145-154. [PubMed: 26188538]

68. Ma YQ, Qin J, Wu C, Plow EF. Kindlin-2 (Mig-2): a co-activator of beta3 integrins. J Cell Biol. 2008;181(3):439-446. [PMCID: PMC2364684] [PubMed: 18458155]

69. Ellison JA, Velier JJ, Spera P, et al. Osteopontin and its integrin receptor $\alpha(v)\beta 3$ are upregulated during formation of the glial scar after focal stroke. Stroke. 1998;29(8):1698-1706. [PubMed: 9707214]

70. Kang WS, Choi JS, Shin YJ, et al. Differential regulation of osteopontin receptors, CD44 and the $\alpha(v)$ and $\beta(3)$ integrin subunits, in the rat hippocampus following transient forebrain ischemia. Brain Res. 2008;1228:208-216. [PubMed: 18638458]

71. Wang X, Lopez OL, Sweet RA, et al. Genetic determinants of disease progression in Alzheimer's disease. J Alzheimers Dis. 2015;43(2):649-655. [PMCID: PMC4245313] [PubMed: 25114068]

72. Shulman JM, Imboywa S, Giagtzoglou N, et al. Functional screening in *Drosophila* identifies Alzheimer's disease susceptibility genes and implicates tau-mediated mechanisms. Hum Mol Genet. 2014;23(4):870-877. [PMCID: PMC3900103] [PubMed: 24067533]

73. Tebar F, Bohlander SK, Sorkin A. Clathrin assembly lymphoid myeloid leukemia (CALM) protein: localization in endocytic-coated pits, interactions with clathrin, and the impact of overexpression on clathrin-mediated traffic. Mol Biol Cell. 1999;10(8):2687-2702. [PMCID: PMC25500] [PubMed: 10436022]

74. Gharesouran J, Rezazadeh M, Khorrami A, Ghojazadeh M, Talebi M. Genetic evidence for the involvement of variants at APOE, BIN1, CR1, and PICALM loci in risk of late-onset Alzheimer's disease and evaluation for interactions with APOE genotypes. J Mol Neurosci. 2014;54(4):780-786. [PubMed: 25022885]

75. Xiao Q, Gil SC, Yan P, et al. Role of phosphatidylinositol clathrin assembly lymphoid-myeloid leukemia (PICALM) in intracellular amyloid precursor protein (APP) processing and amyloid plaque pathogenesis. J Biol Chem. 2012;287(25):21279-21289. [PMCID: PMC3375549] [PubMed: 22539346]

76. Moreau K, Fleming A, Imarisio S, et al. PICALM modulates autophagy activity and tau accumulation. Nat Commun. 2014;5:4998. [PMCID: PMC4199285] [PubMed: 25241929]

77. Morgen K, Ramirez A, Frölich L, et al. Genetic interaction of PICALM and APOE is associated with brain atrophy and cognitive impairment in Alzheimer's disease. Alzheimers Dement. 2014;10(5) (suppl):S269-S276. [PubMed: 24613704]

78. Biffi A, Anderson CD, Desikan RS, et al. ; Alzheimer's Disease Neuroimaging Initiative (ADNI) Genetic variation and neuroimaging measures in Alzheimer disease. Arch Neurol. 2010;67(6):677-685. [PMCID: PMC2956757] [PubMed: 20558387]

79. Jacobsen L, Madsen P, Moestrup SK, et al. Molecular characterization of a novel human hybrid-type receptor that binds the alpha2-macroglobulin receptor-associated protein. J Biol Chem. 1996;271(49):31379-31383. [PubMed: 8940146]

80. Jacobsen L, Madsen P, Jacobsen C, Nielsen MS, Gliemann J, Petersen CM. Activation and functional characterization of the mosaic receptor SorLA/LR11. J Biol Chem. 2001;276(25):22788-22796. [PubMed: 11294867]

81. Rogaeva E, Meng Y, Lee JH, et al. The neuronal sortilin-related receptor SORL1 is genetically associated with Alzheimer disease. Nat Genet. 2007;39(2):168-177. [PMCID: PMC2657343] [PubMed: 17220890]

82. Scherzer CR, Offe K, Gearing M, et al. Loss of apolipoprotein E receptor LR11 in Alzheimer disease. Arch Neurol. 2004;61(8):1200-1205. [PubMed: 15313836]

83. Sager KL, Wuu J, Leurgans SE, et al. Neuronal LR11/sorLA expression is reduced in mild cognitive impairment. Ann Neurol. 2007;62(6):640-647. [PMCID: PMC2669325] [PubMed: 17721864]

84. Alexopoulos P, Guo LH, Tsolakidou A, et al. Interrelations between CSF soluble AβPPβ, amyloid-β 142, SORL1, and tau levels in Alzheimer's disease. J Alzheimers Dis. 2012;28(3):543-552. [PubMed: 22045485]

85. Assareh AA, Piguet O, Lye TC, et al. Association of SORL1 gene variants with hippocampal and cerebral atrophy and Alzheimer's disease. Curr Alzheimer Res. 2014;11(6):558-563. [PubMed: 24938503]

86. Beecham GW, Hamilton K, Naj AC, et al. ; Alzheimer's Disease Genetics Consortium (ADGC) Genome-wide association meta-analysis of neuropathologic features of Alzheimer's disease and related dementias. PLoS Genet. 2014;10(9):e1004606. [PMCID: PMC4154667] [PubMed: 25188341]

87. He F, Umehara T, Saito K, et al. Structural insight into the zinc finger CW domain as a histone modification reader. Structure. 2010;18(9):1127-1139. [PubMed: 20826339]

88. Allen M, Kachadoorian M, Carrasquillo MM, et al. Late-onset Alzheimer disease risk variants mark brain regulatory loci. Neurol Genet. 2015;1(2):e15. [PMCID: PMC4807909] [PubMed: 27066552]

89. Karch CM, Jeng AT, Nowotny P, Cady J, Cruchaga C, Goate AM. Expression of novel Alzheimer's

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disease risk genes in control and Alzheimer's disease brains. PLoS One. 2012;7(11):e50976. [PMCID: PMC3511432] [PubMed: 23226438]

Figures and Tables

Table 1.

Demographic Characteristics and Distribution of Genotypes

Variable	Control Group (n = 322)	MCI Group (n = 496)	AD Dementia Group (n = 159)	<i>P</i> Value
Age, mean (SD), y	75 (6.5)	73 (7.8)	75 (7.8)	<.001
Male sex, No. (%)	156 (48.4)	284 (57.3)	95 (59.7)	.02
Educational level, mean (SD), y	16.6 (2.6)	16.2 (2.7)	15.9 (2.7)	.03
MMSE score, mean (SD)	28.9 (2.1)	27.8 (2.6)	22.8 (2.9)	<.001
<i>APOE</i> ε4, 0/1/2, %	72.4/25.8/1.9	53.4/37.3/9.3	32.7/48.4/18.9	<.001
Amyloid positive, No. (%)	85 (26.4)	252 (50.8)	133 (83.6)	<.001
<i>ABCA7</i> rs3752246, % 0/1/2 alleles	69.3/28.3/2.5	67.7/28.4/3.8	64.8/30.8/4.4	.47
<i>ABCA7</i> rs3764650, % 0/1 or 2 alleles	82.9/17.1	81.3/18.8	83.6/16.4	.72
<i>CLU</i> rs11136000, % 0/1/2 alleles	35.4/50.6/14.0	35.9/49.6/14.5	39.6/44.7/15.7	.91
<i>CLU</i> rs9331949, % 0/1 or 2 alleles	94.7/5.3	96.6/3.4	94.3/5.7	.32
<i>DSG2</i> rs8093731, % 0/1 or 2 alleles	97.8/2.2	98.0/2.0	98.1/1.9	.98
<i>EPHA1</i> rs11771145, % 0/1/2 alleles	44.7/43.8/11.5	44.8/42.3/12.9	33.3/49.7/17.0	.02
<i>FERMT2</i> rs17125944, % 0/1 or 2 alleles	82.9/17.1	85.1/14.9	81.8/18.2	.53
<i>PICALM</i> rs3851179, % 0/1/2 alleles	40.4/46.6/13.0	42.3/45.2/12.5	42.8/48.4/8.8	.59
<i>PTK2B</i> rs28834970, % 0/1/2 alleles	42.2/41.9/15.8	43.1/42.7/14.1	39.0/46.5/14.5	.74
<i>SORL1</i> rs1131497, % 0/1/2 alleles	33.5/47.8/18.6	31.9/52.0/16.1	38.4/48.4/13.2	.26
<i>ZCWPW1</i> rs1476679, % 0/1/2 alleles	50.6/40.1/9.3	52.4/39.5/8.1	54.7/37.7/7.5	.62

Abbreviation: MMSE, Mini-Mental State Examination.

Figure 1.

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Association of Alzheimer Disease Risk Genes With Brain Amyloidosis in the Pooled Sample

Images were visualized using $P \le .01$ (uncorrected) and cluster size (k) of 50 voxels. Scale indicates T values.

Figure 2.



Association of Alzheimer Disease Risk Genes With Brain Amyloidosis in the Normal Control, Mild Cognitive Impairment, and Dementia Groups

Images were visualized using $P \le .01$ (uncorrected) and cluster size (k) of 50 voxels. Scale indicates T values.

Table 2.

FWE- and FDR-Corrected Cluster Analyses and Within-Cluster Peak Effects^a

Gene	Cluster Level				Peak Level			
Variant	FWE- Corrected	FDR- Corrected	Cluster Size,	Uncorrected <i>P</i> Value	Τ	Uncorrected <i>P</i> Value	Talairach Coordinates,	Brain Region
	P Value	q Value	Voxels				x/y/z	
Pooled Sam	P Value	q Value	Voxels				x/y/z	

JAWA Network-

Associations of the	Top 20	Alzheimer	Disease	Risk '	Variants	With	Brain	Amyloidosi	s
								-	

rs3752246							inferior temporal gyrus (BA20)
<i>CLU</i> rs11136000	.07	0.101	1246	.002	3.64 <.001	4/48/0	Right cerebellum
<i>EPHA1</i> rs11771145	.03	0.033	1484	.001	4.18 <.001	-22/-72/48	Left precuneus (BA7)
	.03	0.033	1520	.001	3.57 <.001	44/-82/0	Right middle occipital gyrus (BA18)
FERMT2 rs17125944	.01	0.020	1871	<.001	4.48 <.001	-28/6/-44	Left superior temporal gyrus (BA38)
ZCWPW1 rs1476679	.047	0.082	1380	.001	3.57 <.001	10/18/-26	Right rectal gyrus (BA11)
Normal Con	trol Group						
<i>ABCA7</i> rs3752246	.006	0.008	1914	<.001	4.14 <.001	-22/66/8	Left middle

Open in a separate window

Abbreviations: BA, Brodmann area; FDR, false discovery rate; FWE, familywise error.

^aIn the pooled sample, *ABCA7* rs3764650, *PICALM* rs3851179, and *PTK2B* rs28834970 had no significant clusters; in the control group, *ABCA7* rs3764650 and *PICALM* rs3851179 had no significant clusters; and in the dementia group, *EPHA1* rs11771145 and *SORL1* rs1131497 had no significant clusters.

Figure 3.

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LANA Network



β-Coefficient Maps of the Main Association of *FERMT2* and Its Interaction With Diagnosis and the Association of *FERMT2* Within Each Diagnostic Group

Main association of *FERMT2* with brain amyloidosis (A), its interaction with diagnosis (B), and the association of *FERMT2* with brain amyloidosis in each diagnostic group (C) displayed using Statistical Parametric Mapping 8.

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