

1 Prophylactic allergen immunotherapy with Der p 2 prevents murine 2 asthma by regulating lung GM-CSF

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38 Capsule summary

39 This mouse study demonstrates that repetitive inhalation of a single major house dust mite (HDM)
40 allergen prevents HDM-induced allergic asthma development through suppressing the function of
41 lung dendritic cells, thus providing an alternative to classical allergen-specific immunotherapy.

42 Key words

43 Allergic asthma, house dust mite/HDM, Der p 2/Derp2, allergen-specific immunotherapy/AIT,
44 inhalation, pulmonary, dendritic cell, type 2 conventional dendritic cell/cDC2, GM-CSF

45 To the Editor,

46 The prevalence of allergic diseases is increasing, urging new ways of prevention. Allergen
47 immunotherapy (AIT) is currently the only clinical intervention that can alter the natural course of
48 allergy and can offer long-term clinical benefit. Although AIT is traditionally used as treatment in
49 patients with established disease, the National Institute of Health Immune Tolerance Network (ITN)
50 has proposed that prophylactic AIT could also be used as primary prevention for new sensitizations
51 and allergic disease, in high risk children born to atopic parents and with a personal history of atopic
52 dermatitis and/or food allergen sensitization in early life¹. AIT involves the repeated administration of
53 allergen extracts, leading to the induction of an ill-defined state of systemic allergen-specific
54 immunological tolerance that is associated with decreased symptom scores, particularly in allergic
55 rhinitis patients, but less so in asthmatics². While current AIT involves the subcutaneous or sublingual
56 administration of crude allergen extracts, regimens based on natural routes of mucosal allergen
57 administration and defined single allergens might improve success rate. Several studies have
58 suggested that the natural route of allergen exposure (e.g. ingestion of peanut allergen in infants,
59 bee stings in bee keepers, inhalation of high doses of cat allergens in pet owners) can be very
60 successful in preventing the onset of clinical allergies³⁻⁵. The exposure to immunodominant allergens
61 in these studies, such as Arah6 peanut allergen in breast milk, phospholipase A₂ in bee venom, and
62 Feld1 cat allergen in house dust, suggests that the same allergens that can cause disease are also
63 best at inducing tolerance via the natural route of exposure.

64 To improve the success rate of prophylactic AIT in asthma, we hypothesized that inhalation of Derp2,
65 a major immunodominant house dust mite (HDM) allergen, would prevent the onset of HDM
66 sensitization and HDM-induced asthma. Recombinant Derp2 was produced in the yeast *Pichia*
67 *pastoris*, and was given as prophylactic AIT intranasally to naive C57Bl/6J mice every other day for a
68 period of 15 days, prior to intratracheal (i.t.) sensitization to the full HDM extract, and HDM extract
69 airway challenges (Fig 1a). In mice treated with sham PBS AIT, HDM sensitization and challenges
70 induced robust asthma features, including airway and tissue eosinophilia (Fig 1b-c), goblet cell
71 metaplasia (Fig 1b), T and B cell influx in the bronchoalveolar space (BAL) (Fig 1c), T helper 2 (Th2)
72 cytokine production by lung-draining lymph node (LDLN) cells (Fig 1d), immunoglobulin (Ig)E
73 synthesis (Fig 1e) and bronchial hyperreactivity (BHR) to methacholine (Fig 1f). These allergic asthma
74 features were almost completely abolished in mice that received active Derp2 AIT (Fig 1b-f). To
75 address if intranasal Derp2 AIT would also be effective in a more therapeutic secondary prevention
76 setting, mice were first sensitized to HDM extract, and after one week treated for 17 days with Derp2
77 AIT, followed by HDM airway challenges. In this setting too, Derp2 AIT prevented development of
78 the salient features of asthma, among which eosinophilia, Th2 cytokine production in LDLNs, and BHR
79 ($p < 0.09$) (Fig 1g-i).

80 We next sought for the immunological mechanism(s) of prophylactic AIT. In the active Derp2 AIT
81 group, we observed increased concentrations of HDM-specific IgG2c and IgG1 in serum, and of HDM-
82 specific IgA in BAL fluid (Fig 1e). In humans, successful AIT is often accompanied by increased titers of
83 IgG1, IgG4 and/or IgA⁶. However, despite the increase in several Igs, our prophylactic Derp2 AIT did
84 not require B cells, since the effects of AIT on lung eosinophilia and Th2 cytokine production were
85 preserved in *Mb1^{Cre}* x *Rosa26-Lox-Stop-Lox^{DTA}* mice genetically lacking B cells (Fig E1).

86 The generation of adaptive type 2 immunity to HDM depends on allergen presentation by type 2
87 conventional dendritic cells (cDC2s), which bridge innate and adaptive immunity. The capacity of
88 lungs DCs to take up HDM allergen in the lungs and transport it to the LDLNs was not reduced by
89 prophylactic Derp2 AIT (Fig E2a), and there were only small effects of Derp2 AIT on the expression of
90 the co-stimulatory molecules CD80 and CD86 on cDC1s and cDC2s (Fig E2b). To study the impact of
91 Derp2 AIT on functional allergen presentation by DCs, we used TCR-transgenic 1-DER mice in which
92 all CD4⁺ T cells react to an immunodominant peptide of Derp1⁷. CD4⁺ 1-DER T cells were transferred
93 to mice previously treated with Derp2 or sham PBS AIT, and their active division induced by HDM
94 extract inhalation. 4, 7 and 10 days after HDM inhalation, the LDLNs of sham treated mice contained
95 highly proliferating 1-DER T cells, yet proliferation was strongly reduced in mice receiving Derp2 AIT
96 (Fig. 2a-b). Derp2 AIT also suppressed IL-5, IL-13, and IL-10 secretion (Fig. 2c), and *Gata3* mRNA
97 expression (Fig E2c) by LDLN cells, and reduced the total and effector CD44⁺CD62L⁻ 1-DER T cell
98 numbers in lung tissue (Fig 2d-e). Similar effects of Derp2 AIT on 1-DER T cell activity were observed
99 when i.t. Derp1 was administered instead of HDM to trigger 1-DER T cell responses (data not shown).
100 In support of a role for Derp2 AIT-induced suppression of cDC2 functions, we found that the adoptive
101 transfer of *in vivo* HDM-primed cDC2, obtained from LDLNs of mice that had never been exposed to
102 Derp2 AIT, was sufficient to break the tolerant state induced by prophylactic Derp2 AIT (Fig. 2f).
103 Furthermore, vice versa, LDLN cDC2s from mice undergoing prophylactic Derp2 AIT and then exposed
104 to a single HDM extract inhalation, were less efficient in priming type 2 immunity upon adoptive
105 transfer to naive hosts (Fig E2d). In another set of experiments, cDCs were sorted from the LDLNs of
106 sham or Derp2 AIT mice that were primed with a single Derp1 protein inhalation, and were co-
107 cultured with 1-DER T cells *ex vivo*. cDC2s from the Derp2 AIT group induced less 1-DER T cell
108 proliferation than cDC2s from the sham AIT group (Fig 2g). T cell proliferation induced by cDC1s was
109 not influenced by Derp2 AIT (data not shown).

110 We finally addressed the mechanism of the reduced cDC2-mediated antigen presentation in mice
111 treated with prophylactic Derp2 AIT. In response to HDM inhalation, cDC2s are activated to perform
112 this function through the epithelial release of DC-instructing cytokines⁸. We therefore measured
113 whether prophylaxis with Derp2 affected the release of pro-allergic cytokines and chemokines
114 following a first inhalation of whole HDM allergen extract. In the sham AIT group, a single dose of i.t.
115 instilled HDM extract led to an increased production of MCP-1, KC, IL-1 α , and GM-CSF, in lungs,
116 compared to i.t. instilled PBS (Fig 2h). Prophylactic Derp2 AIT only significantly reduced the
117 production of GM-CSF, a cytokine that has been shown to break inhalation tolerance by activating
118 DCs⁹. To study the functional importance of reduced GM-CSF release in the tolerance mediated by
119 Derp2, mice treated with sham or Derp2 AIT and sensitized and challenged with HDM extract, were
120 supplemented i.t with recombinant GM-CSF at the time of sensitization. Strikingly, Derp2 AIT mice
121 that received GM-CSF at the time of HDM sensitization were no longer protected from allergy
122 development, as assessed by their development of a robust BAL eosinophilia (Fig 2i). We found
123 comparable results in a secondary prevention setting (Fig E3a). Similarly, the effect of prophylactic
124 Derp2 AIT on 1-DER T cell division was also rescued by GM-CSF supplementation (Fig 2j and Fig E3b).
125 In conclusion, our findings in mice show that prophylactic exposure to the single immunodominant
126 HDM allergen Derp2 via the airways offers an alternative way to prevent respiratory allergy to the
127 complex allergen HDM, by suppressing GM-CSF-driven activation of lung cDC2s.

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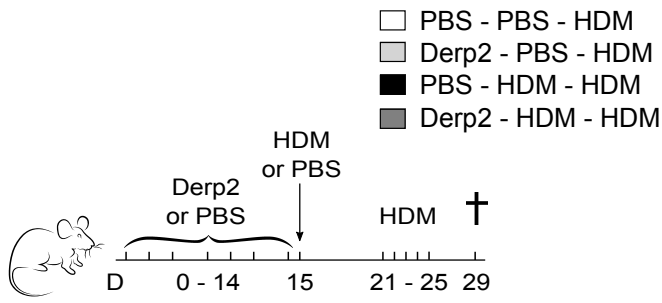
180 Figure legends

181 **Figure 1: Prophylactic Derp2 inhalation prevents HDM-induced allergic asthma development.** (a)
182 Experimental setup for Fig 1b-f. (b) Mucus production (purple, upper panel), and eosinophils (yellow,
183 lower panel) in lungs. (c) Immune cells in bronchoalveolar lavage (BAL) fluids. (d) Cytokine production
184 by HDM-restimulated lung-draining lymph node (LDLN) cells. (e) Immunoglobulins (Igs) in serum (IgE,
185 IgG1, IgG2c), or BAL (IgA). (f) Airway resistance in response to methacholine (Mch). (g-i) Secondary
186 prevention setting. (g) Immune cells in BAL. (h) Cytokine production by HDM-restimulated LDLN cells.
187 (i) Airway resistance in response to Mch. Results are representative of 1 (b), 2 (e), 4 (d) or 6 (c), or
188 pooled from 2 (f, i) or 3 (g, h) independent experiments, with $n = 4-7$ (b-f) or $3-6$ (g-i) mice/group for
189 single experiments. Data are shown as means \pm SEM, or in (f) and (i) as predictions of means \pm SEM
190 obtained from repeated measurement analysis using residual maximum likelihood (REML). * $p < 0.05$,
191 ** $p < 0.01$, *** $p < 0.001$. OD: optical density at 650 nm/850 nm.

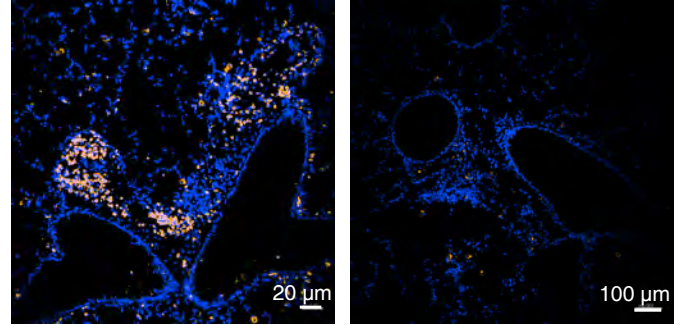
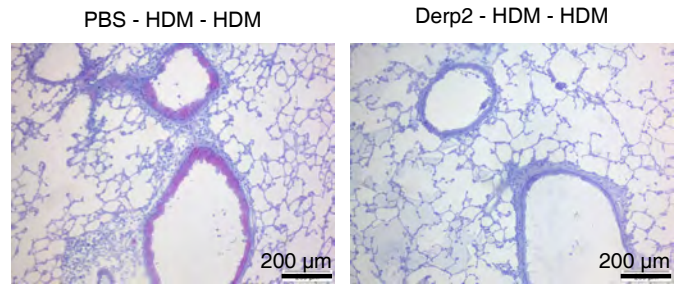
192 **Figure 2: Prophylactic Derp2 inhalation suppresses cDC2-mediated Th2 responses by blocking GM-**
193 **CSF release.** (a-e) Derp2 or sham PBS mice received CFSE-labeled 1-DER T cells intravenously (iv) and
194 HDM intratracheally (i.t.), and were analyzed 4, 7 or 10 d later. (a) Cell division profiles, and (b)
195 proliferation parameters, of tissue-resident ($CD45^{iv\ injected (iv)}$) 1-DER T cells in lung-draining lymph
196 nodes (LDLNs). (c) Cytokine production by HDM-restimulated LDLN cells. (d) Number and (e)
197 phenotype of tissue-resident lung 1-DER T cells. (f) Treatment as in Fig 1a, with 2 groups of mice i.t.
198 sensitized with donor HDM-primed cDC2s instead of HDM. Immune cells in bronchoalveolar lavage
199 (BAL) fluids. (g) V450-labeled 1-DER T cells co-cultured with LDLN cDC2s of Derp2 or sham PBS mice
200 i.t. instilled with Derp1. Cell division profiles, expansion indexes, and counts of 1-DER T cells. (h)
201 Cytokines and chemokines in lung tissue of Derp2 or sham PBS mice 2 h after HDM instillation. (i)
202 Treatment as in Fig 1a, with 1 group of mice i.t. sensitized with HDM + GM-CSF. BAL immune cells. (j)
203 Derp2 or sham PBS mice received CFSE-labeled 1-DER T cells iv and Derp1 with or without GM-CSF
204 i.t.. Proliferation parameters of LDLN 1-DER T cells. Results are representative of 2 (f-h, j) or 3 (a-e), or
205 pooled from 2 (i) independent experiments, with $n = 4-8$ (a-f, h-j) mice/group or 2-3 (g)
206 replicates/group for single experiments. Shown as means \pm SEM (or \pm SD in (g)). * $p < 0.05$, ** $p < 0.01$,
207 *** $p < 0.001$, **** $p < 0.0001$.

Figure 1

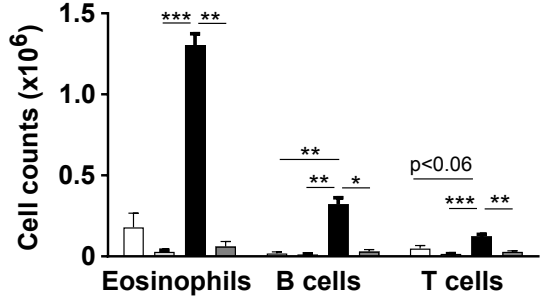
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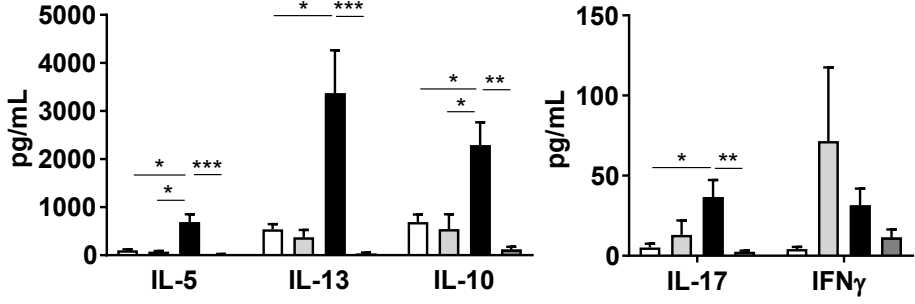
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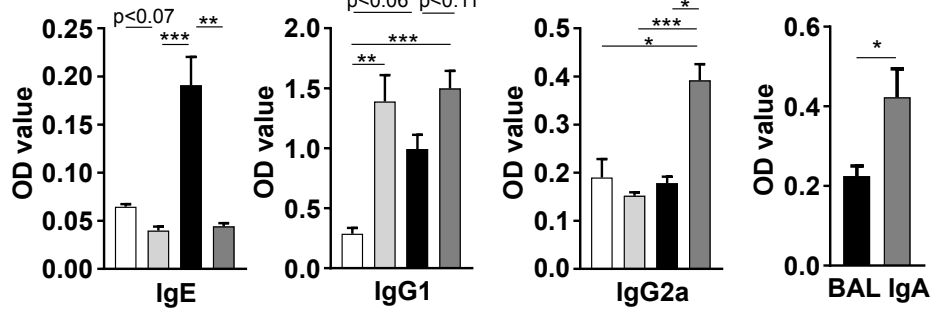
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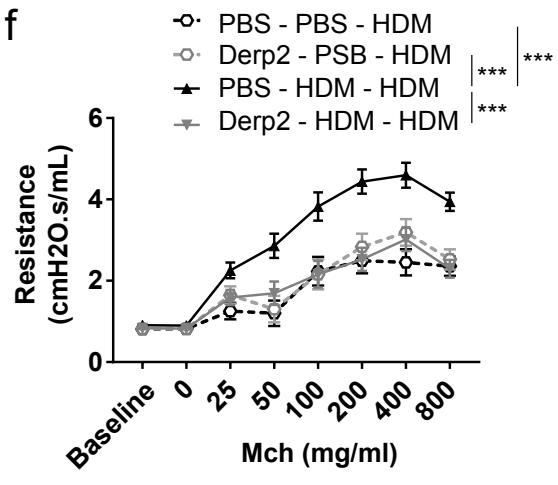
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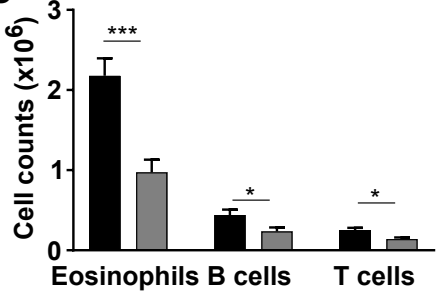
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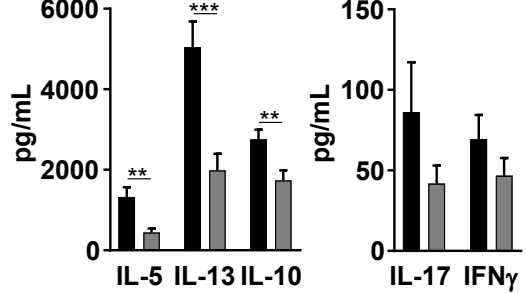
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