| 1 | High energy or protein concentrations in food as possible offsets for cyanide |
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| 2 | consumption by specialized bamboo lemurs in Madagascar? |
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| 24 | Short title: Variation in food composition of bamboo lemurs |
| 25 | |
| 26 | Abstract |
| 27 | Plants producing toxic plant secondary metabolites (PSM) deter feeding of folivores. Animals |
| 28 | that are able to cope with noxious PSMs have a niche with a competitive advantage over other |
| 29 | species. However, the ability to cope with toxic PSMs incurs costs for detoxification. In order |
| 30 | to assess possible compensations for the ingestion of toxic PSMs, we compare the chemical |
| 31 | quality of plants consumed by bamboo lemurs (genera Hapalemur and Prolemur; |
| 32 | strepsirrhine primates of Madagascar) in areas with and without bamboo. Some bamboo |
| 33 | lemurs consume bamboo containing concentrations of cyanogenic substances $10 - 50$ times |
| 34 | above the average lethal dosage for mammals, and we postulate that animals consuming |

35 cyanogenic substances need supplementary protein or readily available energy for

36 detoxification. We compared the chemical composition of food consumed by three species of

- 37 bamboo lemurs that feed mainly (>80% of their time) on bamboo in the evergreen rainforest
- 38 of Ranomafana (Madagascar) with published data of the diets of bamboo lemurs at two sites
- 39 without highly cyanogenic plants (reed beds of Lac Alaotra and the evergreen littoral forest of
- 40 Mandena) and with food of sympatric folivorous lemur species that do not feed on bamboo.
- 41 Lemurs feeding on bamboo consumed up to twice as much protein than bamboo lemurs in
- 42 areas without bamboo and sympatric lemur species that feed on leaves of trees.
- 43 Concentrations of non-structural carbohydrates (a source of energy) showed the opposite
- 44 trend. This result supports the hypothesis that feeding on cyanogenic plants is linked to high
- 45 protein intake, either as a source of protein or for sulphur containing amino acids that can be
- 46 used for detoxification. Due to the high protein concentrations in bamboo, however, we

47 cannot distinguish between the hypothesis that lemurs that eat bamboo target additional food

48 items with higher protein, from the hypothesis that lemurs feeding on bamboo unavoidably

- 49 obtain higher concentrations of protein than animals feeding on leaves of trees, without an
- 50 added nutritional benefit.
- 51

52 Keywords Primates; Strepsirrhine; *Hapalemur* spp.; *Prolemur simus*; Detoxification;

- 53 Folivory; Plant secondary chemicals; Cyanogenic substances
- 54

55 Introduction

56 Generalist folivores feeding on toxic plants have to diversify their diet so that different 57 plant secondary metabolites (PSMs) can be detoxified by different metabolic pathways, 58 because single pathways can be limiting by their detoxification rates (Foley and Moore 2005; 59 Nersesian et al. 2012). In contrast, some folivorous specialists focus on only a few plant 60 species (Shipley et al. 2009), such as the giant panda (Ailuropoda melanoleuca; Schaller et al. 1985; Nie et al. 2015), red panda (Ailurus fulgens; Johnson et al. 1988), pygmy rabbit 61 62 (Brachylagus idahoensis; Shipley et al. 2006), some marsupials (Dearing et al. 2000; Marsh et al. 2003), and the bamboo lemurs of Madagascar (gentle and golden bamboo lemurs: 63 64 Hapalemur spp.; greater bamboo lemur: Prolemur simus; Mutschler 1999; Tan 1999, 2006). 65 Here, plants and folivorous animals are linked in an evolutionary arms race in which plants 66 produce PSM- to deter feeding, and folivores evolve morphological and/or physiological 67 mechanisms for detoxification of these components, or exhibit behavioural selection of higher quality food or reduced energy expenditure to cope with PSMs (Freeland and Janzen 1974; 68

69 Guglielmo et al. 1996; Provenza et al. 2003; Iason and Villalba 2006; Moore and DeGabriel 70 2012). From the plants' perspective, production of PSMs is costly and therefore some PSMs 71 are only produced when plants are threatened by folivores (Dolch and Tscharntke 2000) or 72 after they have actually been fed upon (induced defences; Schuman and Baldwin 2012). From 73 the animals' point of view, detoxification also requires additional energy expenditure that 74 sometimes needs to be compensated for by ingestion of more or higher quality food (Glander 75 1982; Provenza et al. 2003; Dearing et al. 2005; Foley and Moore 2005; Villalba and 76 Provenza 2005; Lee et al. 2006; Shaw et al. 2006). A dramatic cost of detoxification of a 77 single PSM (benzoate) has been described for common brushtail possums (Trichosurus 78 *vulpecula*) where the costs for detoxification account for about 30% of the dietary nitrogen 79 intake (Au et al. 2013). Common brushtail possums that fed on a low protein diet had a 80 negative protein balance (Au et al. 2013). Thus, animals that consume toxic food, possibly in 81 order to avoid competition, should incur higher energy and nutrient requirements. 82 The bamboo lemurs of Madagascar are among the very few mammals with a diet dominated by bamboo (Schaller et al. 1985; Tan 1999), a monocotyledonous flowering plant 83 84 belonging to the grass family Poaceae. In the altitudinal Malagasy rain forest at Ranomafana, 85 three species of bamboo lemurs (Hapalemur griseus ranomafanensis, H. aureus, and 86 Prolemur simus) spend approximately 80 - 95% of their feeding time consuming various 87 bamboo species, including cyanogenic parts of the giant bamboo (Cathariostachys 88 madagascariensis; Tan 1999, 2006). These bamboo lemurs consume cyanogenic items (Tan 89 2000, 2006) and excrete cyanide (or more likely thiocyanate) mainly in urine (Yamashita et al. 2010). The parts consumed contain up to 0.6% cyanide per gram dry weight which is about 90 91 10 - 50 times the lethal dosage for other mammals (Glander et al. 1989; Ballhorn et al. 2009a, 92 2016). Cyanide is detoxified by conversion to thiocyanate (Huang et al. 2016). This reaction 93 is catalysed by the enzyme rhodanese and requires a sulphur donor, most likely sulphur 94 containing amino acids (Conn 1979; Sousa et al. 2003). While the enzyme is widespread in 95 animal tissues, a 10- to 50-fold increase of cyanogenic substances beyond the lethal dosage 96 ought to require production of supplementary rhodanese and availability of sulphur containing 97 amino acids, making this a protein expensive detoxification mechanism. Detoxification is 98 likely also to be linked to increased energy requirements, which could be provided by non-99 structural carbohydrates that are easily digestible and metabolized to produce energy. 100 In contrast to the altitudinal rain forest at Ranomafana, some bamboo lemur species,

101 Hapalemur alaotrensis and H. meridionalis, occur in reed beds and littoral forests without

102 bamboo. Hapalemur alaotrensis feed predominately on Cyperaceae and Poaceae, mainly

103 reeds (*Phragmites*; Mutschler 1999), and *H. meridionalis* on a variety of plants, mainly 104 Poaceae, including terrestrial grass (Eppley et al. 2011, 2016). Cyanogenesis is frequent in 105 Poaceae and Juncaceae (Conn 1979; Naik 1984; Harborne 1988), but cyanogenic substances 106 are not characteristic for terrestrial grasses (Frohne and Jensen 1973). Neither bamboo nor 107 grasses consumed by mountain gorillas (Gorilla beringei beringei) contain cyanogenic 108 substances (Grueter et al., 2016; J. Rothman, pers. comm.). Additionally, a variety of grasses 109 taken from botanical gardens did not contain cyanogenic substances or only in concentrations 110 too low to poison folivorous mammals (Ballhorn, unpubl.).

111 Though the notion that protein is a limiting factor for animals feeding on plants is not 112 always supported (Oftedal 1991; Wasserman and Chapman 2003; Stolter 2008; Rothman et 113 al. 2011), protein is thought to play an important role in primate food selection because food 114 for primate folivores is often characterized by higher concentrations of different measures of 115 protein than items not eaten (protein measured as crude, soluble or digestible protein; e.g., Milton 1979; Ganzhorn 1988, 1992; Oates et al. 1990; Chapman et al. 2002; DeGabriel et al. 116 117 2008, 2009, 2014; Wallis et al. 2012). Refined approaches demonstrated that primates may not simply select high protein dietary items, but adjust their food composition to meet their 118 119 protein requirements by ingesting a certain amount of protein per day (Felton et al. 2009; 120 Johnson et al. 2013). While the emphasis on protein for folivores' food selection has a long 121 tradition, an increasing number of studies indicates that folivores do not need to optimize 122 protein intake because food items contain enough protein to satisfy their needs (Oftedal 1991; 123 Stolter 2008; Ganzhorn et al. 2017). These studies rather indicate an important role of easily 124 digestible energy for the maintenance of animals (Wasserman and Chapman 2003; Rothman 125 et al. 2011). There are not enough studies available yet to analyse which component is 126 favoured over the other at any given circumstances.

At Ranomafana and in the littoral forests of eastern Madagascar, bamboo lemurs 127 128 coexist with other folivorous lemurs that feed on leaves of trees. These sympatric species can 129 be used for comparisons of the chemical properties of food consumed by animals that ingest 130 cyanogenic substances versus species that do not, or ingest cyanogenic components in much lower concentrations. We tested the prediction that the cyanogenic plant parts fed upon by 131 132 bamboo lemurs contain higher concentrations of protein and/or easily extractable energy than 133 food items consumed by bamboo lemurs or other folivorous lemur species that feed on plants 134 without or with lower cyanide concentrations.

135 If the consumption of cyanogenic substances requires additional protein or energy, we136 expect that:

137 (1) bamboo eating bamboo lemurs consume items with higher protein or non-structural

138 carbohydrate content than bamboo lemurs that do not feed on bamboo,

139 (2) bamboo eating bamboo lemurs consume items with higher protein or non-structural

140 carbohydrate concentrations than sympatric folivorous lemur species that do not feed on

141 bamboo, and

142 (3) items consumed by bamboo lemurs that do not feed on bamboo should not differ in their

143 protein or non-structural carbohydrate concentrations from items consumed by other

144 sympatric folivorous lemur species.

145

146 Methods

147 Study species and sites

148 Bamboo lemurs (Hapalemur and Prolemur) are strepsirrhine primates of Madagascar 149 with body mass ranging from 0.8 - 1.7 kg (*Hapalemur* spp.) and 2.2 - 2.6 kg (*Prolemur* simus) (Tan 2006; Mittermeier et al. 2010). All species considered here inhabit the evergreen 150 151 rainforest or swamps of eastern Madagascar and data are based on observations in the wild (Table 1). Apart from *H. alaotrensis* from the marshes of Lac Alaotra, all other bamboo lemur 152 153 species occur in sympatry with other folivorous lemur species, such as Avahi spp. and 154 Propithecus spp., that do not consume bamboo or other grasses but rely on leaves from trees. 155 The species and study sites have been described previously: Hapalemur alaotrensis in the 156 reed beds of Lac Alaotra (Mutschler et al. 1998; Mutschler 1999), H. meridionalis in the 157 littoral forest of Mandena (Eppley et al. 2011, 2015a), Avahi meridionalis from the littoral 158 forest of Ste. Luce (Norscia et al. 2012), a forest fragment close to Mandena, described by 159 Donati et al. (2011), H. griseus ranomafanensis, H. aureus, Prolemur simus and Propithecus edwardsi in the evergreen rainforest of Ranomafana National Park (Tan 1999, 2006; Arrigo-160 161 Nelson 2006). H. alaotrensis and H. meridionalis occur at sites without bamboo and feed on 162 plants without or with very low cyanogenic potential (HCNp; the maximum amount of cyanide that can be released from the accumulated cyanogenic precursors; Ballhorn et al. 163 164 2009b). Hapalemur griseus ranomafanensis, H. aureus and P. simus feed mainly on bamboo, 165 some of which are characterized by very high HCNp. The two other lemur species (A. 166 meridionalis and P. edwardsi) do not eat bamboo and were included for comparison of 167 species that ingest other possible food items available at the same site. 168 169

| 1/1 I able I Life history traits of lemur species included in the present st |
|---|
|---|

| | | Body | Feeding | Resting | Observation | Group composition | # Food | |
|------------------------------|-------------|------|---------|---------|--------------|-------------------|---------|----------------|
| Species | Site | mass | % | % | time | M / F | species | Reference |
| Sites without cyanogenic bar | mboo | | | | | | | |
| Hapalemur alaotrensis | Lac Alaotra | 1.2 | < 22% | > 60% | 13 months: | 3 / 2 | 11 | Mutschler |
| | | | | | 5 days/month | 1 / 1 | | 1999 |
| H. meridionalis | Mandena | 1.1 | 42% | 44% | 12 months: | 6 / 8 | 72 | Eppley et al. |
| | | | | | 5 days/month | (3 groups) | | 2011, 2016 |
| Avahi meridionalis | Ste. Luce | 1.2 | 15% | 67% | 7 months: | 1 / 1 | 47 | Norscia et al. |
| | | | | | 256 hours | 1 / 1 | | 2012 |
| Sites with cyanogenic bambo | <u>00</u> | | | | | | | |
| H. griseus | Ranomafana | 0.9 | 48% | 41% | 12 months: | 1 / 2 | > 40 | Tan 1999, |
| ranamofanensis | | | | | 5 days/month | 1 / 2 | | 2006, C. Tan |
| | | | | | | | | unpubl. data |
| H. aureus | Ranomafana | 1.4 | 37% | 54% | 12 months: | 1 / 1 | > 21 | Tan 1999, |
| | | | | | 5 days/month | 1 / 1 | | 2006, C. Tan |
| | | | | | | | | unpubl. data |
| Prolemur simus | Ranomafana | 2.6 | 41% | 50% | 12 months: | 3 / 2 | 7 | Tan 1999, |
| | | | | | 5 days/month | 1 / 2 | | 2006 |
| Propithecus edwardsi | Ranomafana | 5.8 | 28% | 42% | 12 months: | 7 / 8 | > 75 | Arrigo-Nelson |
| | | | | | 5 days/month | (7 groups) | | 2006 |

172 Body mass (kg) from cited literature or Mittermeier et al. (2010)

174 Food composition: Comparison between species and sites

175 In our analyses, we distinguish between food types and food items. A food item 176 represents the item consumed by an animal in a given feeding event (= one sample). Food 177 types represent plant parts. The chemical composition of a food type can be represented by the chemical analysis of a single sample of that food type. In this case, the food type based on just 178 179 one or two samples is considered to be representative for the type throughout the study area. 180 This category of data is available for Hapalemur alaotrensis (Mutschler 1999) and Avahi 181 meridionalis (Norscia et al. 2012). It ignores temporal, spatial and individual variation of the 182 chemical composition between plant individuals (Ganzhorn and Wright 1994; Chapman et al. 183 2003; Rothman et al. 2012). For the other lemur species (H. meridionalis, H. griseus ranomafanensis, H. aureus, P. simus, P. edwardsi) we collected food items whenever an 184 185 animal was seen feeding and sampling was possible. We analysed the chemical composition 186 of food types for the pooled samples per food type (in case of *H. meridionalis*) or calculated

¹⁷³

187 as the mean of the concentration from all items (= samples) collected per food type. We used

188 food consumed by other folivorous lemur species, (Avahi meridionalis) in the littoral forest

189 (Norscia et al. 2012), and Propithecus edwardsi in Ranomafana (Arrigo-Nelson 2006; Arrigo-

190 Nelson et al. unpublished data) to compare the chemical composition of food consumed by

191 bamboo lemurs with other potential food items available at the same site. Behavioural

192 observations and sample collection for these species are based on systematic observations of

193 habituated animals (Table 1).

194

195 Food selection of bamboo lemurs at Ranomafana

In Ranomafana, when possible, we collected items equivalent to the item eaten for
chemical analyses at the moment of a feeding event. For this, we collected a similar item as
the item consumed from the same plant individual. This procedure allows us to consider the
chemical variability within plant species (Chapman et al. 2003; Moore and Foley 2005;
Rothman et al. 2012).

201 In order to assess possible selection criteria, we correlated the frequency of 202 consumption of any given food type with the mean concentrations of the various chemical 203 components of the food type. Weighing the consumption of food types by the frequency of 204 consumption seems a suitable procedure since each item was analysed chemically. Repeated 205 consumption of the same item might represent some kind of preference, though we did not 206 consider the availability of the item in question. Other procedures to assess selection criteria, 207 such as measuring the time of consumption or counting bites, calculating intake based on bites 208 or comparing items consumed with items not consumed have other advantages (Zinner 1999; 209 Rothman et al. 2012; DeGabriel et al. 2014; Aristizabal et al. 2017), but are simply not 210 available for comparisons.

211

212 *Chemical analyses*

Chemical analyses have been performed and published for food items consumed by *Hapalemur alaotrensis* (Pollock 1986; Mutschler 1999), and by *H. meridionalis* (Eppley et al.
2011, 2016). We analysed dried samples for all other species in the lab of Hamburg
University, following the methods described by Bollen et al. (2004).

217 Samples were dried in the sun or in a drying oven, ground with a Culatti MFC mill to 218 pass a 1 mm sieve, and dried again overnight at 50-60°C prior to analyses. Total nitrogen was 219 determined with a Gerhardt Kjeldatherm using the Kjeldahl procedure. Crude protein was 220 calculated as nitrogen concentration * 6.25. Samples were analysed for neutral detergent fibre

(NDF) (Goering and van Soest 1970; van Soest 1994) modified according to the instructions
for use in an ANKOM FIBER ANALYZER. All concentrations are expressed as percentages
of dry weight. Biochemical analyses were carried out at the Institute of Zoology, Department
of Ecology and Conservation at Hamburg University.

225 Originally, HCNp of the dried food items consumed by the different bamboo lemurs 226 from Ranomafana had been analysed enzymatically using the Spectroquant® cyanide test 227 (Merck KGaA, Darmstadt, Germany; Ballhorn et al. 2009a). None of these samples showed 228 any detectable cyanide. In contrast, plant material from Ranomafana stored in alcohol or 229 analysed in the field, reliably released up to 0.6% cyanide per dry plant material (Glander et 230 al. 1989; Ballhorn et al. 2009a) and showed positive reactions for cyanides in semi-231 quantitative tests (Tan, 1999, 2006; Yamashita et al. 2010). Therefore, we assume that the 232 dried samples contained cyanogenic substances but our analyses for HCNp did not produce 233 viable results likely because potentially specific enzymes (β -glucosidases) necessary to 234 release cyanide from the bamboo tissue had been destroyed during the drying process or 235 storage of samples. In order to have at least some qualitative estimates for the cyanogenic 236 potential of bamboo lemur food items, we performed the analyses of potential food items in 237 the field as listed below and compiled published information on the cyanide concentrations of 238 various parts and species of bamboo (Table 2).

239

240 **Table 2** Possible cyanide concentrations found in bamboo and grass from southeastern

241 Madagascar. All analyzed plant species are known food items of *Hapalemur griseus*

242 ranomafanensis (H.g.r.), H. aureus (H.a.) and/or Prolemur simus (P.s.) (Tan 2006); however,

only the species/parts consumed within Ranomafana are indicated. Published concentrations
of cyanide (in % per dry weight) are referenced at the end of the table.

| Species and Part | (| Consumed (+ | -) | Cyanide [%] |
|-------------------------|-----------|-------------|-------------|--|
| | H.g.r. | H.a. | <i>P.s.</i> | |
| Arundinaria sp. | | | | · |
| Leaf tip | | | | $0.40 \text{ (mean)} - 0.81 \text{ (max)}^{-1}$ |
| Culm pith | | | | $0.01 \text{ (mean)} - 0.02 \text{ (max)}^{1,2}$ |
| Ground shoot | | | | $0.40 \text{ (mean)} - 0.81 \text{ (max)}^{1,2}$ |
| Cathariostachys madagas | cariensis | | | · |
| Ground shoot | + | + | + | $0.40 \text{ (mean)} - 0.81 \text{ (max)}^3$ |
| Branch shoot | + | + | + | $0.61 \text{ (mean)} - 1.08 \text{ (max)}^3$ |
| Branch complement | + | + | + | 0.61 ^{3, 4} |
| Culm pith | | | + | $0.01 \text{ (mean)} - 0.02 \text{ (max)}^5$ |

| Leaves | | | + | 0 ^{3, 7} |
|---------------------------|---|---|---|-------------------|
| Mature leaf base | | | + | 0 5,7 |
| Mature leaf tip | | | + | 0 5,7 |
| Young leaf base | + | + | + | 0 5,7 |
| Young leaf tip | | | + | 0 5,7 |
| Sokinochloa chiataniae | | | | |
| Mature culm | | | | 0 6,7 |
| Growing tip | + | + | + | 0.02 6 |
| Leaf blade | | | + | 0 6, 7 |
| Sokinochloa brachyclada | | I | | |
| Branch shoot | + | + | + | 0 6,7 |
| Ground shoot | + | + | + | 0 6, 7 |
| Mature leaves | | | + | 0 6, 7 |
| Young leaf base | + | + | + | 0 6,7 |
| Young leaf tip | | | + | 0 6, 7 |
| Nastus elongatus | | I | | |
| Branch shoot | + | + | + | + 7 |
| Nastus sp. | | | | |
| Stem | | | | 0 7, 8 |
| Poecilostachys festucaceu | S | I | | |
| Stem | + | + | + | 0 9 |
| Leaves | | | + | 0 9 |
| Mature leaf tip | | | | 0 9 |

245

¹ Eisler (1991), ² assumed to be equivalent to *Cathariostachys madagascariensis* (Eisler

1991), ³ Ballhorn et al. (2009b), ⁴ assumed to be equivalent to "branch shoots", ⁵ Ballhorn 246 (unpubl. data), ⁶ Glander et al. (1989), ⁷ Tan (2006), 247

⁸http://tech.groups.yahoo.com/group/bamboo-plantations/message/2019, ⁹ Yamashita et al. 248

249 (2010)

250 For statistical analyses, we assigned a concentration of 0.01% HCN to items that showed a

251 positive response (+) in the Feigl-Anger Test, if only qualitative data are available.

252

253 Total non-structural carbohydrates (TNC) were used as a proxy for easily digestible

254 carbohydrates (for a discussion see Conklin-Brittain et al. 2006; Rothman et al. 2012). TNC

were calculated as: TNC = 100% - Crude protein – Neutral detergent fibre. Lipids and ash 255

ought to be subtracted from this measure, but these analyses were not available. Lipids are not an important component in leaves. Ash (= mineral) contents can amount to several percent of dry mass in leaves (National Research Council 2003) but has not measured consistently in the samples used for the present analysis. We assume that the error introduced by not subtracting lipids and ash is small compared to the large fibre fraction. Neutral detergent fibre (NDF) was only used to calculate TNC. We did not have any specific hypothesis concerning NDF itself. Therefore, NDF were not discussed specifically.

263 For analysis of plant HCNp, fresh items were selected from bamboo in Ranomafana several years after the behavioural observation of the bamboo lemurs had been completed 264 265 (Ballhorn et al. 2009b). These items were used as proxy for cyanogenic components in the 266 original food samples of bamboo lemurs because the analyses of the original food items failed 267 and could not be repeated. For each plant species, we analysed different plant tissues for 268 HCNp to obtain information on the quantitative variability of cyanogenic precursor content as there frequently is substantial within-plant variation with young and reproductive tissues 269 270 showing the highest amounts of cyanogenic precursors (Ballhorn et al. 2008, 2009b). Fresh plant samples were weighted to the nearest 0.001 g and ground with liquid nitrogen and 271 272 cooled mortar and pestle (4°C) under addition of the fourfold volume (v fresh weight⁻¹) of ice-273 cold Na₂HPO₄ solution (67 mmol L⁻¹). Samples were quantitatively analysed for their HCNp 274 by complete enzymatic hydrolysis of cyanogenic precursors under addition of β-glucosidase 275 from almonds (Emulsin®, Merck, NY, USA). We used closed glass vessels (Thunberg 276 vessels) for incubation (20 min at 30 °C in a water bath) of plant extracts together with 277 enzyme solution adjusted to an activity of 20 nkat. Quantification of released HCN was 278 conducted spectrophotometrically at 585 nm using the Spectroquant® cyanide test (Merck 279 KGaA, Darmstadt, Germany; Ballhorn et al. 2009b).

280

281 Statistical analysis

Data were tested for deviations from normality with Kolmogorov-Smirnov-one sample 282 283 tests. If residuals deviated from normality, we used non-parametric tests. Kruskal-Wallis 284 Analysis of Variance was used for comparisons of more than two groups. Post-hoc tests for 285 subsequent pairwise comparisons were based on Mann-Whitney-U-tests and significance 286 levels were Bonferroni corrected. For parametric analyses of more than two groups we used 287 Analysis of Variance with subsequent Scheffé's post-hoc test. All statistical analyses were 288 performed in IBM SPSS Statistics v. 22 (SPSS Inc., Chicago, IL, U.S.A.), with significance 289 set at p < 0.05.

290

291 Results

292 Basic information on life history traits and study design of the bamboo lemur species 293 considered are given in Table 1. Analyses were not separated by sex because no published 294 data were available for this comparison. Since sex ratios were balanced in all species, there 295 should not be any bias in the results due to different protein or energy requirements of females 296 and males. The number of food plant species listed in Table 1 represents the total number of 297 species known to be consumed by the different lemur species. Chemical analyses 298 (summarized in Table 3) are only available for subsets of plant species consumed most 299 frequently.

300

301 Unweighted samples

The unweighted data in Table 3 are a summation of the nutrients in all of the plant types that have been recorded as consumed by a given species and were analysed for their chemical composition; for example, chemical analyses for *Hapalemur aureus* are available for four different plant types that have been consumed by *H. aureus*. The unweighted data in Table 3 represent the total nutritional value of these plant types as if they were consumed in equivalent amounts.

308 Hapalemur g. ranomafanensis and P. simus consumed plants with much lower 309 cyanogenic potential (HCNp) than H. aureus. Due to small sample size of the unweighted 310 samples, this difference is not significant (Table 3). Though the median of plant HCNp is zero 311 or close to zero for H. g. ranomafanensis and P. simus, respectively, both species include 312 some plant parts with very high cyanide concentration (Table 3). According to the unweighted samples, food types consumed by Hapalemur aureus have higher concentrations of nitrogen 313 314 than food consumed all other lemur species, except for the sympatric H. g. ranomafanensis 315 (Table 3).

Food of all bamboo lemur species from the sites with non-cyanogenic plants and the non-bamboo-lemur species has lower nitrogen concentrations than the two *Hapalemur* species from Ranomafana (Table 3; Fig. 1a). *Prolemur simus*, with respect to nitrogen in its diet, is closer to the non-bamboo eating *Propithecus edwardsi* than to the other two bamboo lemur species of Ranomafana.

321 Concentrations of non-structural carbohydrates did not differ consistently between
 322 Ranomafana and Mandena. According to the unweighted samples, *Prolemur simus* ate the
 323 items with the lowest concentrations of total non-structural carbohydrates (TNC) while *Avahi*

- 324 meridionalis and Propithecus edwardsi had the highest concentrations of TNC (Fig. 1b). The
- 325 high concentrations in food of non-bamboo lemur species do not differ between Avahi
- *meridionalis* and *Propithecus edwardsi* (Table 3).



Fig. 1 Concentrations of nitrogen (Fig. 1a) and non-structural carbohydrates as a proxy of
energy (Fig. 1b) in food items consumed by bamboo lemurs and sympatric folivorous lemur
species at sites with cyanogenic bamboo (rain forest of Ranomafana) and at sites without
cyanogenic bamboo (marsh of Lac Alaotra, littoral forests of Mandena and Ste. Luce). Values
are means and 95% confidence intervals; items not weighted by frequency of consumption. *Hau: Hapalemur aureus; Hgr: H. griseus ranomafanensis; Ps: Prolemur simus; Hal: H. alaotrensis; Hm: H. meridionalis; Am: Avahi meridionalis; Pe: Propithecus edwardsi.*

337

338 Weighted samples

339 Frequency of consumption was considered in the weighted data. Weighted data are340 only available for *Hapalemur meridionalis* (from a non-bamboo site) and the lemur species341 that inhabit the sites with cyanogenic bamboo (i.e., Ranomafana). In this case, the diet342 constituents are weighted by the frequency of consumption, such that the nutritional contents343 reported in Table 3, are a representation of the lemur's average nutritional intake.

Based on the frequency of consumption, *Hapalemur aureus* ate the highest proportion of cyanogenic items (65.1% of collected food items), followed by *Prolemur simus* (64.1% of collected food items) and *H. griseus ranomafanensis* (18.2%). The frequency of consumption of cyanogenic plant items differs significantly between the three species ($\chi^2 = 14.11$, df = 2, p

348 < 0.01).

349 For most species and chemicals, the averages of the weighted samples were

350 remarkably similar to the means of the unweighted samples, but not for the nitrogen

351 concentration in the food of *Hapalemur g. ranomafanensis* (Table 3). While the

352 concentrations of cyanogenic potential in the weighted food item samples were lower for *H. g.*

353 ranomafanensis compared to P. simus and H. aureus, this difference was not statistically

354 significant (Table 3). The three bamboo lemur species from Ranomafana have significantly

355 higher concentrations of nitrogen than *H. meridionalis*. Food items of *H. aureus* have

356 somewhat higher concentrations of nitrogen than food consumed by *H. g. ranomafanensis and*

P. simus, though this difference is not significant. According to the weighted samples, food of

the three bamboo eating lemur species did not differ in their concentrations of NDF or TNC.

359 All bamboo lemur species from Ranomafana had significantly lower concentrations of NDF

and TNC than food items consumed by *H. meridionalis* (Table 3).

361

362 **Discussion**

Animals that are able to feed on plants with specific plant secondary metabolites have exclusive access to resources not used by competitors. While avoiding competition is advantageous, the need for detoxification is likely to be associated with costs, either in terms of increased energy or protein requirements (Dearing et al. 2005, Foley and Moore 2005,

367 Villalba and Provenza 2005; Lee et al. 2006, Shaw et al. 2006; Nersesian et al. 2012; Au et al. 368 2013). Bamboo lemurs of Madagascar are of special interest because some of the species 369 ingest cyanogenic bamboo up to about 50 times the lethal dose of cyanide, which range 370 between 74 and 370 mmol kg⁻¹ for other mammals, such as sheep, mice, cats or rats (Glander 371 et al. 1989; Ballhorn et al. 2009a). While all bamboo lemur species feed on bamboo where 372 bamboo is available and all of the Ranomafana bamboo lemur species have been found to 373 ingest and excrete cyanide (Yamashita et al. 2010), the proportion of cyanogenic food 374 ingested by the different species varies and can be negligible at sites without bamboo 375 (Glander et al. 1989; Mutschler et al. 1998; Mutschler 1999; Tan 1999; Eppley et al. 2011, 376 2016). This offers the possibility to investigate, whether or not species feeding on cyanogenic 377 bamboo compensate for the assumed costs of detoxification by increased consumption of 378 items that provide easily digestible energy or protein, either for supplying enzymes for 379 detoxification or for maintenance or to supply sulphur for the detoxification mechanism. As 380 tests of this hypothesis, we compared the chemical composition of food items consumed by 381 bamboo lemur species living in areas with cyanogenic bamboo with the food composition in 382 areas where bamboo is absent and where the animals feed on non-cyanogenic plants. In 383 addition, we compared the chemical composition of food items consumed by bamboo lemurs 384 with the diet of other folivorous lemurs from the same areas (Avahi meridionalis in the littoral 385 forest and Propithecus edwardsi in the rainforest of Ranomafana).

386 The inter-site comparison revealed that all bamboo lemur species feeding on 387 cyanogenic bamboo (Hapalemur aureus, H. griseus ranomafanensis, Prolemur simus) 388 ingested up to 1.7 times more nitrogen than the two bamboo lemur species (Hapalemur 389 alaotrensis, H. meridionalis) studied at sites without cyanogenic bamboo. The Hapalemur 390 spp. feeding on cyanogenic bamboo in Ranomafana also ingest more nitrogen than the 391 sympatric non-bamboo lemur species (Propithecus edwardsi) feeding on the leaves of trees. 392 Food items of Prolemur simus have higher nitrogen concentrations than items of the diet of 393 Propithecus edwardsi, though this difference is not significant. In contrast to the Hapalemur 394 species at Ranomafana, the Hapalemur spp. feeding on non-cyanogenic items do not ingest 395 more nitrogen than the non-bamboo lemur species (Avahi meridionalis) that is sympatric to H. 396 meridionalis. Cyanogenic glycosides contribute to the nitrogen concentrations in cyanogenic 397 plants. But their relative contribution is small (maximum 0.3% nitrogen, corresponding to 398 0.6% cyanide) compared to the nitrogen contained in protein. The nitrogen concentrations in 399 food plants of the lemur species feeding on cyanogenic food plants ranges between 2.4% and 400 4.4% (Table 3). Nitrogen concentrations in food consumed by bamboo lemur species that do

401 not rely on bamboo (in areas where bamboo is absent) ranges between 1.2% and 2.1%. Even 402 when the maximum nitrogen content of 0.3% bound in cyanogenic substances is subtracted 403 from the total nitrogen concentration of cyanogenic food parts, the average nitrogen 404 concentration in food items of bamboo consuming species would be about 1.1% higher than 405 in food plants of species that do not include bamboo in their diet. Nitrogen concentrations of 406 1.1% are equivalent to about 6.9% more crude protein. According to studies that linked 407 protein in food to reproductive success and growth, this difference can have a major impact 408 for folivorous mammal fitness (Moore and Foley 2005; DeGabriel et al. 2009).

409 The actual difference in the consumption of digestible nitrogen might be more 410 pronounced than indicated by the measure of nitrogen or crude protein. Grasses do not contain 411 condensed tannins and therefore these components do not interfere with protein digestibility 412 as may do tannin-rich leaves of trees (Waterman and Mole 1994; Wallis et al. 2012). Thus, 413 while the nitrogen concentrations reported for bamboo are closely linked to the concentrations 414 of digestible nitrogen, the concentrations of digestible nitrogen in leaves of trees are likely to 415 be much lower and do not show a tight relationship between digestible and crude protein 416 (Wallis et al. 2012). This would increase the difference in protein between a diet based on 417 bamboo and a diet based on the leaves of trees. This argument could be used to support the 418 finding (in favour of the original hypothesis that additional protein is needed for 419 detoxification) that the nitrogen concentrations in food of Prolemur simus do not differ from 420 the nitrogen concentration of the sympatric Propithecus edwardsi.

Based on our limited knowledge of the population dynamics of these species, the bamboo lemur species at the different sites do not seem to differ in their reproductive output or population dynamics (Nievergelt et al. 2002; Tan 2006; Eppley et al. 2015b, 2016b). Therefore, the surplus of protein ingested from bamboo in Ranomafana is not reflected in higher reproductive rates. In contrast to the nitrogen content, there is no indication that energy (measured as total non-structural carbohydrates; though proteins can certainly also be converted to energy) plays a role in detoxification or food selection.

Morphological traits could provide hints for special adaptations to deal with food peculiarities, such as a complex forestomach that could facilitate detoxification (Chivers and Hladik 1980; Alexander 1993; Waterman and Kool 1994). However, at least *Hapalemur* griseus does not have any morphological specialization that indicates any specific adaptation of the digestive tract. The only difference between *H. griseus* and a dietary generalist lemurid species such as *Lemur catta* is a shorter cecum and a shortened and sacculated colon of *H.* griseus (Campbell et al. 2004; Perrin 2013). However, these are unlikely candidates for

detoxification adaptations, as detoxification should occur at the beginning of the digestive
tract rather than at the end. However, *H. griseus* achieves higher digestion of fibre and protein
than other lemurs studied so far, indicating some kind of specialization that cannot be linked
to morphological characteristics as both features of the digestive tract hint towards lower
rather than higher digestion rates, just the opposite of what has been found in digestion studies
(Campbell et al. 2000). Thus, it remains unclear what the surplus protein in bamboo is used
for.

442 On the basis of the comparison between lemur species occurring at different sites, the 443 utilization of protein for detoxification is consistent with the original hypothesis. It is also 444 consistent with studies on sheep and brushtail possums, which describe that animals can 445 ingest more PSM if supplemented with protein (Villalba and Provenza 2005; Nersesian et al. 446 2012; Au et al. 2013). Our comparison of the three sympatric bamboo lemur species at 447 Ranomafana matches this scenario, providing evidence that cvanide tolerance may be related 448 to protein ingestion. In Ranomafana, Hapalemur g. ranomafanensis is the species that 449 consumes the lowest amounts of cyanogenic substances (Table 3; Yamashita et al. 2010), 450 while it is also the species ingesting the lowest portion of nitrogen in its diet.

451 Despite our results supporting the hypotheses, our sample size is small and we are 452 aware that the study design is not optimal as it is non-experimental and lacks direct 453 physiological measurements of energy and protein expenditure, but all the lemur species 454 involved are threatened, with most being classified as "Endangered" or "Critically 455 Endangered" according to the most recent IUCN classification (July 2012; Schwitzer et al. 456 2013). Therefore, it is impossible to use these animals in standard physiological experiments 457 and, in particular, it is out of the question that the animals could be fed cyanogenic food for an 458 experimental study. Thus, we had to rely on descriptive field studies. The results of our 459 comparisons are consistent with the hypothesis that lemurs consuming cyanogenic bamboo 460 can compensate for the toxins by consuming more protein because (1) bamboo eating bamboo 461 lemurs consume items with higher protein content than bamboo lemurs that do not feed on 462 bamboo, (2) two out of three bamboo eating lemur species consume items with higher protein 463 than sympatric folivorous lemur species that do not feed on bamboo, and (3) items consumed 464 by bamboo lemurs that do not feed on bamboo do not differ in their protein concentrations 465 from items consumed by other sympatric folivorous lemur species. The results for total non-466 structural carbohydrates show the reverse trend. Therefore, we conclude that protein serves a 467 greater function than simply providing a source of energy for these animals.

468 Yet, bamboo is rich in protein but low in total non-structural carbohydrates. Therefore, 469 it could well be that if lemur species feed on bamboo they unavoidably ingest high 470 concentrations of protein. Also, it would have to be expected that lemurs feeding on 471 cyanogenic substances would show some indication of protein selection. This was not found 472 in our analyses and it remains enigmatic what the bamboo lemurs do with the high 473 concentrations of protein in their diet. These issues cannot be further resolved without 474 experimental approaches or at least by applying new conceptual frameworks on how to study 475 food selection (Felton et al. 2009; Rothman et al. 2011; Johnson et al. 2013; DeGabriel et al. 476 2014).

477

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814 **Table captions**

- 815 **Table 1** Life history traits of lemur species included in the present study.
- 816
- 817 **Table 2** Possible cyanide concentrations found in bamboo and grass from southeastern
- 818 Madagascar. Plant species are known food items of Hapalemur griseus ranomafanensis
- 819 (H.g.r.), H. aureus (H.a.) and/or Prolemur simus (P.s.), with those parts consumed within
- 820 Ranomafana indicated. Published concentrations of cyanide are referenced at the end of the
- table.
- 822
- **Table 3** Chemical composition of food items eaten by bamboo lemurs *Hapalemur* spp.,
- 824 *Prolemur simus*; and sympatric folivorous lemur species.
- 825

826 Figure legend

- Fig. 1 Concentrations of nitrogen (Fig. 1a) and non-structural carbohydrates as a proxy of
- 828 energy (Fig. 1b) in food items consumed by bamboo lemurs and sympatric folivorous lemur
- 829 species at sites with cyanogenic bamboo (rain forest of Ranomafana) and at sites without
- 830 cyanogenic bamboo (marsh of Lac Alaotra, littoral forests of Mandena and Ste. Luce). Values
- are means and 95% confidence intervals; items not weighted by frequency of consumption.
- 832 Hau: Hapalemur aureus; Hgr: H. griseus ranomafanensis; Ps: Prolemur simus; Hal: H.
- 833 alaotrensis; Hm: H. meridionalis; Am: Avahi meridionalis; Pe: Propithecus edwardsi.