Title - Detecting Ovarian Cancer Using Extracellular Vesicles: Progress and

2 Possibilities

3

1

- 4 Authors
- 5 Emanuela Carollo¹, Bianca Paris¹, Priya Samuel¹, Paschalia Pantazi¹, Thais Fernanda Bartelli^{1,2},
- 6 Emmanuel Dias-Neto^{2,3}, Susan Ann Brooks¹, Ryan Charles Pink¹, David Raul Francisco
- 7 Carter¹†

8 9

Author Affiliations

- 10 1. Department of Biological and Medical Sciences. Faculty of Health and Life Sciences. Oxford
- Brookes University, Gipsy Lane, Headington, Oxford, UK, OX3 0BP.
- 12 2. Laboratory of Medical Genomics, A.C.Camargo Cancer Center, São Paulo, SP, Brazil.
- 13 3. Laboratory of Neurosciences Alzira Denise Hertzog Silva (LIM-27), Instituto de Psiquiatria,
- 14 Faculdade de Medicina da Universidade de São Paulo, São Paulo, SP, Brazil.
- † Corresponding author. <u>dcarter@brookes.ac.uk</u>

16 **Abstract**

- 17 Ovarian cancer (OC) is the deadliest gynecological malignancy. Most patients are diagnosed when
- they are already in the later stages of the disease. Earlier detection of OC dramatically improves
- 19 the overall survival, but this is rarely achieved as there is a lack of clinically implemented
- 20 biomarkers of early disease. Extracellular vesicles (EVs) are small cell-derived vesicles that have
- been extensively studied in recent years. They contribute to various aspects of cancer pathology,
- including tumour growth, angiogenesis and metastasis. EVs are released from all cell types and the
- 23 macromolecular cargo they carry reflects the content of the cells from which they were derived.
- 24 Cancer cells release EVs with altered cargo into biofluids, and so they represent an excellent
- 25 potential source of novel biomarkers for the disease. In this review we describe the latest
- developments in EVs as potential biomarkers for earlier detection of OC. The field is still relatively
- young, but a number of studies have shown that EVs and the cargo they carry, including miRNAs
- and proteins, can be used to detect OC. They could also give insight into the stage of the disease
- and predict the likely therapeutic outcome. There remain a number of challenges to the use of EVs
- 30 as biomarkers, but through ongoing research and innovation in this exciting field there is great
- 31 potential for the development of diagnostic assays in the clinic that could improve patient
- 32 outcome.

33 34

Keywords

35 Extracellular vesicles, exosomes, ovarian cancer, miRNAs, diagnosis, liquid biopsy.

3637

Introduction

38

Ovarian cancer (OC) is the 7th most commonly diagnosed cancer in women and the disease causes

40 more than 150,000 deaths around the world each year, making it the deadliest gynecological

malignancy^{1, 2}. The 5-year survival for OC patients is less than 50%³, and this is mostly due to the frequently late presentation of the disease (when the cancer has usually already spread to the peritoneal space) and the subsequent acquisition of therapeutic resistance during treatment^{3, 4}.

OC is a heterogeneous disease that is thought to originate from the epithelium of the ovaries⁵, though recent studies suggest that it may originate from cells shed from the fallopian tube⁶. It can be classified into two groups based on their distinct origin and histological characteristics⁵. The first group, originating from epithelial cells and known as epithelial ovarian cancer (EOC), accounts for almost 90% of all OC cases; this group can be further subdivided on the basis of histological morphology into serous (the majority of these are high-grade serous carcinoma [HGSC] and the rest are low-grade serous carcinoma [LGSC]), mucinous, endometrioid, and clear cell carcinomas⁷. The second group can originate from stromal and germ cells and include non-epithelial ovarian cancers that account for 10% of all OC cases⁸.

The treatment of most patients with EOC comprises surgical debulking of the tumour mass followed by chemotherapy with platinum-based compounds such as carboplatin, cisplatin and oxaliplatin⁹. Successful debulking is the best correlate for subsequent survival^{10, 11}. The treatment regime for EOC has changed little in the past few decades, with the addition of taxanes such as paclitaxel (which work by inhibition of microtubule function) being the only substantial change to the way in which EOCs are treated⁹. The report that the presence of tumor infiltrating lymphocytes is a good indicator of chemotherapy response and improved survival for OC implies a synergistic effect of immunotherapy and chemotherapy that may benefit some patients¹².Immunotherapy for OC is still limited to clinical trials and only a low proportion of patients appear to benefit clinically, unlike immunotherapy treatment in melanoma which appears to be particularly successful¹³. There is therefore an urgent need for improved therapeutics for OC, but most importantly, an urgent need for the identification of improved high-sensitivity biomarkers to facilitate earlier detection of the disease and to monitor treatment response to standard and new therapy.

Current biomarkers for ovarian cancer detection

Almost 70% of OC cases are diagnosed at an advanced stage (III or IV) of the disease^{7,14}. Early symptoms of OC, including back pain and digestive problems such as dyspepsia, are general and non-specific and are often related to benign conditions. It tends to be only later, once the less serious causes of symptoms have been eliminated and/or symptoms become more intense or severe, that the cancer is diagnosed¹⁵. Late diagnosis has a significant impact on prognosis; patients that are diagnosed in earlier stages have a five-year survival rate of >70% but this drops to <40% for those diagnosed in later stages^{4, 16}. Transvaginal ultrasonography is an imaging-based approach, which could be used to identify earlier-stage disease, but it is invasive and trials have shown that as a screening tool it only has limited ability to reduce overall survival¹⁷⁻¹⁹. The inability to detect OC at an earlier stage reveals one of the biggest challenges in biomarker research: the

need of finding a highly sensitive, non-invasive screening method applicable to an asymptomatic population subject to develop OC during their life.

> Due to the high incidence and low survival rates, much research effort has been put into finding potential biomarkers for screening the disease. One of the first OC biomarkers identified was Cancer Antigen 125 (CA125), also known as MUC16²⁰. CA125 is a high molecular weight transmembrane glycoprotein produced by coelomic epithelium, and thus far it is the most clinically utilised biomarker for monitoring the response to treatment and detecting disease recurrence²¹. When the treatment is successful the level of circulating CA125 can decrease, while increased levels are correlated with drug resistance and disease progression²². Although CA125 is used in the clinic, it has some limitations for the purpose of early detection, as it can be associated with both false positive and false negative results²¹. Non-epithelial ovarian tumours are not associated with increased levels of this mucin, and the various EOC subtypes produce different levels of CA125 (levels are higher in HGSC compared to mucinous carcinoma, for example) ²²⁻²⁴. Some studies reported a higher correlation between the FIGO (International Federation of Gynecology and Obstetrics) staging and preoperative CA125 levels, whilst other studies suggest the correlation is less clear-cut^{23, 24}. CA125 can also be detected in other physiological or pathological conditions leading to false positives; for example, it can be raised in the presence of other cancers including cervix, breast, colon and lung cancer²⁵, and it can be altered in inflammatory pathologies²⁶, during the menstrual cycle and in pregnancy^{27, 28}. Moreover, in some cases of EOC the level of CA125 is not raised, and this is particularly true in the early stages of the disease when the presence of a biomarker would be most clinically useful²⁹. CA125, therefore, does not represent the ideal biomarker for robust and diagnostic detection of early OC, and this is confirmed by clinical trials that reveal CA125 as a screening tool lead to only modest (if any) improvements in patient survival^{17, 18}.

Another biomarker that has been investigated is the Human Epididymis protein 4 (HE4)³⁰. This is a small protein encoded by the *WFDC2* (WAP Four-Disulfide Core Domain 2) gene and secreted by epithelial cells. HE4 is secreted by pulmonary epithelial cells, appears to be involved in sperm maturation, but is also highly expressed in the serum of OC patients when compared with healthy individuals^{30, 31} and it could be used for distinguishing OC from benign disease and to monitor treatment and recurrence³². Unfortunately, like CA125, HE4 has limitations as a biomarker; its expression has been associated with other conditions (especially with endometrial and lung cancer and cystic fibrosis) and different factors (including menstrual cycle, hormone treatment, age and smoking) can modify its expression, leading to false negatives and false positives^{28, 33}.

Greater predictive power could be achieved by combining different biomarkers. For example, the Risk for Ovarian Malignancy Algorithm combines CA125 and HE4 levels, leading to increases in both sensitivity and specificity and thus diagnostic accuracy³⁵. The Risk of Malignancy Index combines CA125 levels, ultrasound results and menopausal status³⁶. Another multivariate index assay known as OVA1 combines the results of measuring several different proteins alongside

CA125³⁷. However, even these multiparametric tests suffer from false positives and negatives and the only definitive way to diagnose patients is during surgery, making these tests unsuitable for routinely screening OC in the female population. Further research is therefore needed to identify other potential biomarkers for earlier detection of OC.

Extracellular Vesicles as cancer biomarkers

Extracellular vesicles (EVs) are a heterogeneous group of cell-derived submicron vesicles surrounded by a lipid bilayer³⁸. EVs can be broadly classified into three main types depending on the mechanisms of their biogenesis (Fig 1). Apoptotic bodies are considered to be larger EVs (>1000 nm in diameter, though smaller apoptotic vesicles can also be released) produced by apoptotic cells. Microvesicles (MVs) are a class of EVs produced by outward membrane budding of a cell with sizes ranging between 50 nm and 1 µm. Exosomes are small (30-150 nm in diameter) EVs produced when multivesicular bodies (MVBs) fuse with the plasma membrane leading to the release of the intraluminal vesicles (which, upon release, are then redefined as exosomes). Another class of EVs that is gaining attention in the tumour microenvironment is large oncosomes (LOs). LOs are a big class of EVs (3-4 um diameter), mostly originated from highly aggressive tumor cells and characterized by carrying a variety of oncogenic signals^{39, 40}. More recently, the group of David Lyden (Weill Cornell Medicine, New York, US) reported the identification of a new class of EVs, dubbed exomeres⁴¹. These particles, whose origin and mechanism of formation are still unknown, were found using asymmetric flow field-flow fractionation, are smaller than exosomes (<50nm) and were described as the most predominant particle secreted by cancer cells⁴¹.

EVs were, until recently, thought to primarily perform the role of facilitating the release of unwanted cellular material⁴². While this may be one of their roles, we now know that they can serve a variety of functions⁴³. They play significant roles in many biological processes, including angiogenesis and the immune response⁴³. EVs are released from all cell types and they carry various types of cargo such as long nucleic acids (including mRNAs, lncRNAs and DNA), short nucleic acids (including miRNAs, vault RNAs, tRNAs and YRNAs), proteins, glycoproteins, carbohydrates, metabolites and lipids^{44, 45}. Their biologically active cargo can be transferred into and used by recipient cells, leading to changes in the function of these cells^{46, 47}. EVs are therefore an important part of the signaling dialogue that occurs between cells. Many of the EVs produced by cells can make their way into biofluids, including saliva, urine, cerebrospinal fluid, blood, semen, sweat and tears⁴³. The molecular composition of EVs partially reflects the molecular landscape of the parental cell; given that this landscape can be altered in cancer, and that EVs can reach biofluids, these vesicles could serve as an easily accessible window into the state of a tumour⁴⁸. Indeed, the very presence in biofluids of vesicles carrying cancer-related cargo could be a diagnostic indicator that a tumour is present.

One approach that is often taken is to identify potential vesicular biomarkers in cancer patients is to initially use cultured cancer cell lines as a proxy. The cargo (miRNAs in particular) of EVs

released by cancer cell lines (compared to non-cancer cell lines) can be profiled using relatively unbiased techniques (such as microarrays or RNA-seq); transcripts that are identified as altered in cancer cell-derived EVs can then be measured in biofluids to see if they are also de-regulated in patients⁴⁸. However, one potential issue is that, even though the foetal bovine serum that the cultured cells grow in has been pre-cleared of vesicles, it still contains bovine-derived EVs carrying bovine miRNAs⁴⁹. Directly testing biofluids using unbiased techniques may, therefore, be preferable as a method for identifying suitable EV biomarkers; however, working with EVs presents several technical challenges and there are a number of pre-analytical variables that affect the interpretation of the results of such experiments^{50, 51}. Further work is also needed to establish the most reliable EV isolation strategy for the analysis of vesicular cargo in biofluids⁵².

Despite these barriers, there is great excitement for the potential use of EVs as biomarkers to inform clinicians not just on the *presence* of tumours but also on the *state* of the tumour and its microenvironment^{48, 53}. EVs can play an active role in the pathology of cancer, contributing to an increase in various undesirable phenotypes such as metastasis⁵⁴, angiogenesis⁵⁵ and drug resistance⁵⁶. EVs released by stressed cells (including stress induced by the chemotherapeutic agent cisplatin) are able to induce a range of effects in neighbouring tumour cells, including increased invasion, bystander DNA damage and an adaptive response⁵⁷⁻⁵⁹. miRNAs are short noncoding RNAs that can repress the expression of multiple genes, meaning that changes in their levels can lead to substantial phenotypic effects in a cell⁶⁰. They are known to be involved in stress response⁶¹ and in mediating drug resistance in ovarian cancer⁶²⁻⁶⁴. miRNAs can also contribute to other cancer phenotypes, including migration and angiogenesis⁶⁵. For these reasons, the miRNA content of EVs is of particular interest as a potential diagnostic in cancer⁴⁸. A growing body of literature describes the early attempts to capitalize on EV cargo as a biomarker in a variety of cancer types. In the following section we will review current work investigating EVs as biomarkers in OC (Table 1).

EVs as biomarkers in ovarian cancer

Early forays into EV biomarker research in OC focused on simply counting the number of vesicles in circulation. Cancer cells produce more EVs when compared with normal cells, and this could be related to specific conditions in the tumour microenvironment⁶⁶. The levels of circulating EVs were seen to be elevated in patients with EOC and these may correlate with disease stage^{67, 68}.

For the reasons described in the previous section , the detection of circulating vesicular RNA, particularly miRNA, could be used as a biomarker for OC. In addition, the RNA is protected from degradation whilst encapsulated in vesicles, and sensitive techniques can be used to amplify specific targets from relatively few copies of the nucleic acid, allowing a global view of the complex RNA landscape in these organelles^{69, 70}. Many studies have analysed changes of miRNA levels in biofluids such as plasma or serum, but it is not always clear whether these changes represent free circulating miRNAs or those encapsulated in EVs; a more comprehensive review of

non-vesicular biomarkers in OC has been previously published⁷¹. Here we will focus on studies where EVs have been specifically investigated.

207208209

210

211

212213

214

215

216

217

218219

220

221

222

206

In one early study it was shown that certain miRNAs (including miR-21, miR-141, miR-200a/b/c and miR-214) were more highly expressed in circulating EVs from OC compared to patients with benign disease⁶⁸. In another study the level of miR-200a/b/c and miR-373 were shown to be elevated in circulating EVs of EOC patients⁶⁷. Correlations with stage and lymph node involvement were observed for miR-200a and miR-373, while lower overall survival correlated with levels of miR-200b/c⁶⁷. EV miRNAs were measured in peritoneal or pleural effusions and the level of a combination of miRNAs was correlated with stage, progression free survival and overall survival⁷². An expression signature of eight miRNAs circulating in serum was also shown to distinguish healthy control from patients with OC⁷³. Most of these miRNAs were shown to be vesicular when released by cell lines and xenografts⁷³. In another study the levels of miR-21 were elevated in ovarian carcinoma EVs⁷⁴. Pan *et al* showed that the levels of a number of miRNAs are altered in the plasma EVs of ovarian cancer patients. Interestingly, the levels of miR-200b correlated with CA125 and overall survival⁷⁵. Urine could also be used as a source of diagnostic EVs. In one study the levels of miR-30a-5p were elevated in the urine of ovarian cancer patients, and this miRNA was shown to be found in EVs⁷⁶.

223224225

226

227

228229

230

231

232

233

234

235

236

237

238

239

240241

242

243

244

245

246

247

Proteins in EVs have also been investigated as potential OC biomarkers. Interestingly, CA125 has been identified in EVs and its vesicular levels were higher than freely circulating CA125 plasma levels at an earlier stage, suggesting that studying CA125 vesicular levels instead of freely circulating serum CA125 could be used to detect OC earlier⁷³. Another study demonstrated that the expression of Claudin 4 protein in EVs obtained from plasma of OC patients was positively correlated with tumor stage, with a sensitivity of 51% and specificity of 98%⁷⁸. Furthermore, the dual measurement of CA125 and Claudin4 inside EVs could be used as a new combination biomarker, although further validation experiments need to be performed⁷⁸. Patients with stage I EOC had a low level of circulating CA125 but high levels of Claudin 4, suggesting that relative levels of the two could be informative⁷⁸. It has been shown that EVs derived from OC patients' plasma contain increased levels of TGF\$1 and melanoma associated antigen 3 (MAGE3) compared with patients with benign disease, suggesting they could serve as potential biomarkers to distinguish between malignant and benign patients⁷⁹. The levels of EpCAM, ADAM10 and EMMPRIN have also been shown to increase in EOC^{68, 80}. Zhang et al developed a microfluidic device combined with ELISA to detect EVs; with this device they show that in a small cohort of patients the levels of EpCAM in plasma EVs from OC patients was higher compared to controls⁸¹. CD24 has been associated with poor prognosis in OC and it is highly enriched in EVs from ascitic fluid⁸², with most of the CD24 positive exosomes being secreted by tumour cells⁸². Combined measurement of vesicular EpCAM and CD24 can distinguish between patients that are responsive or nonresponsive to therapy⁸³. Zhao et al developed a microfluidic device named 'ExoSearch Chip' to isolate serum exosomes that contain CA125, EpCAM and CD24. When EVs were isolated using antibodies against CA125 it was noted that patient samples contained a greater amount of EVs compared to healthy controls⁸⁴. Similarly, another microfluidic-based platform was used to show

that the number of EpCAM+ EVs is correlated with disease progression in OC⁸⁵. In another study, soluble E-cadherin was found to be released with EVs into ascitic fluid and the levels were able to distinguish between OC and benign disease⁸⁶. Taken together, these results suggest that vesicular proteins have promising potential as biomarkers and that they could be potentially used as point of care testing (POCT) as quick, cheap and sensitive technique that could help to overcame the challenges related with early diagnosis.

253254255

256

257

258

259

260

248

249

250251

252

Other EV-associated molecules can also be used as biomarkers. A recent study published by Lea *et al* developed an ELISA assay that can detect and bind picogram-levels of phosphatidylserine (PS)-containing EVs from the plasma of OC patients, and, based on the difference in the number of PS-positive EVs, it can differentiate between malignant and benign disease⁸⁷. The use of single EV-methods that can distinguish between subpopulations of vesicles may also be useful in identifying cancer-specific EVs⁸⁸. In another study, microvesicle-associated tissue factor procoagulant activity was able to distinguish between plasma from OC patient and healthy controls⁸⁹.

261262263

264

265

266

Future perspective

EVs are not the only potential source of non-invasive circulating biomarkers for detecting and monitoring cancer. In particular, the use of cell-free DNA, circulating tumour DNA and circulating tumour cells have shown both diagnostic and prognostic utility in the diagnosis and monitoring of $OC^{90,\,91}$.

267268269

Although EVs have gained a lot of attention in recent years as a potential biomarker for OC and other cancer types, there is still much work required before their potential can be realised.

270271272

273

274

275

276

277

278279

280

281

282

283

284285

286

287

288

There remain many challenges associated with working with EVs^{48, 50, 52}. There are several methods for extracting EVs but no universal agreement on which technique is most appropriate for diagnostic purposes. The most commonly used techniques are ultracentrifugation (UC) and size exclusion chromatography (SEC), but both of these approaches lead to the enrichment of EVs with different biofluid impurities including soluble proteins and lipoproteins. Combining methodologies, such as SEC and density cushion centrifugation can help to remove lipoproteins and therefore improve EV purity^{52, 92}. Antibody-bound magnetic beads that are specific to EV markers such as CD63 can be used to isolate vesicles and enhance purity⁹². Methods that rely on polyethylene glycol-based precipitation result in much lower EV purity but give higher yields⁹³; miRNA/protein biomarkers discovered using such methods cannot be definitively attributed as vesicular (unless further validation is performed) but if the biomarker is reproducibly and robustly altered in OC then it has value. Microfluidic approaches for EV isolation are being developed that can be combined with novel sensor technologies^{81, 84, 85}; these approaches may be less useful for biomarker discovery (due to the smaller scale of material that they produce) but could be used effectively to detect differences in specific EV cargo in ovarian cancer patients. Further research is needed to develop isolation and detection technology, and to assess the effect of isolation methodology on EV purity.

289290

291

292

293

294

295

Purifying EVs from different biofluids presents various challenges⁹⁴, and there are many preanalytical variables that can affect the measurement of biomarkers such as vesicular miRNA and proteins⁵¹. These include the method of biofluid collection, time of day, whether the patient has fasted and the presence of other conditions. These factors could affect biomarker discovery and the testing of biomarkers in a clinical setting. More work is therefore required to establish the effect of pre-analytical variables on the detection of EV cargo and robust procedures must be put in place for diagnostic applications.

296297298

299

300

301

302

Another challenge to quantifying EV cargo is choosing the most appropriate 'reference gene'. It is not clear, at present, which proteins or RNAs are most appropriate as a reference for normalisation. The normalisation of expression of RNAs or proteins of interest can therefore be problematic, with outcomes depending on the choice of reference. More studies are needed to identify EV content whose levels are the most stable in different conditions and between different individuals.

303304305

306

307

308

309

310

311

312

313

314

315

316

317

318

319

320

EVs in any given biofluid are released from a variety of cells. A tumour may contribute EVs to this heterogeneous population, and the proportion of tumour-derived vesicles will increase as the disease progresses. The ability to detect a smaller number of cancer EVs in this sea of normal EVs depends on several factors, including the nature of extraction methodology, the sensitivity of the detection assay and the 'normal levels' of the EV-cargo being measured. In the ideal test, the EVcargo being detected would be absent in normal biofluid but present in high levels after the appearance of a tumour. The test could be run on a small amount of biofluid at the point-of-care and be sufficiently sensitive to pick up very small numbers of cancer-derived EVs. As the sensitivity of EV-based detection methods increase it may be possible to move beyond the use of a test to monitor treatment/relapse towards a true diagnostic test for early-stage OC in asymptomatic individuals. Ideally, the collection of this biofluid would be minimally invasive for the patients and part of their routine disease follow-up (such as blood collection, for example). Whilst improvements are being made in all these areas, this hypothetical test does not currently exist. Further work is necessary to identify novel potential biomarkers in EVs and develop the technology required to isolate and detect them. Exciting progress is being made in this area which we hope will allow us to unlock the potential of EVs for earlier diagnosis of ovarian cancer in the near future.

321322

323

324

325

326

327

328

329

Acknowledgements

We thank Cancer Research UK, The Cancer and Polio Research Fund and Oxford Brookes University for funding. DRFC is also supported by a BBSRC Project Grant. E-DN is a research fellow from Conselho Nacional de Desenvolvimento Científico e tecnológico (CNPq) and acknowledges the support received from Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP grants 11/18784-6; 14/26897-0 and 15/50257-3) and Associação Beneficente Alzira Denise Hertzog Silva (ABADHS). We also thank the Newton Fund and FAPESP for funding of the 'Extracellular Vesicles

and non-cellular RNA: roles in health and neglected tropical diseases' Workshop. We apologise to other authors whose excellent work we could not include in this review due to space constraints.

332333334 References

335

- 336 1. Siegel, R. L.; Miller, K. D.; Jemal, A., Cancer Statistics, 2017. *CA Cancer J Clin* **2017**, *67* 337 (1), 7-30.
- 2. Reid, B. M.; Permuth, J. B.; Sellers, T. A., Epidemiology of ovarian cancer: a review. 339 *Cancer Biol Med* **2017**, *14* (1), 9-32.
- 340 3. Aletti, G.; Gallenberg, M.; Cliby, W.; Jatoi, A.; Hartmann, L., Current management strategies for ovarian cancer. *Mayo Clin Proc* **2007**, *82* (6), 751-70.
- 342 4. Cannistra, S. A., Cancer of the ovary. *N Engl J Med* **2004**, *351* (24), 2519-29.
- 5. Kurman, R. J.; Shih, I. M., The origin and pathogenesis of epithelial ovarian cancer: a proposed unifying theory. *Am J Surg Pathol* **2010**, *34* (3), 433-43.
- 6. Kurman, R. J., Origin and molecular pathogenesis of ovarian high-grade serous carcinoma. *Ann Oncol* **2013**, *24 Suppl 10*, x16-21.
- 7. Berkenblit, A.; Cannistra, S., Advances in the management of epithelial ovarian cancer. *J Reprod Med* **2005**, *50* (6), 426-38.
- 8. Matulonis, U. A.; Sood, A. K.; Fallowfield, L.; Howitt, B. E.; Sehouli, J.; Karlan, B. Y., Ovarian cancer. *Nat Rev Dis Primers* **2016**, *2*, 16061.
- 9. Raja, F. A.; Chopra, N.; Ledermann, J. A., Optimal first-line treatment in ovarian cancer. *Ann Oncol* **2012**, *23 Suppl 10*, x118-27.
- 353 10. Schorge, J. O.; McCann, C.; Del Carmen, M. G., Surgical debulking of ovarian cancer: 354 what difference does it make? *Rev Obstet Gynecol* **2010**, *3* (3), 111-7.
- 355 11. Polterauer, S.; Vergote, I.; Concin, N.; Braicu, I.; Chekerov, R.; Mahner, S.; Woelber, L.;
- 356 Cadron, I.; Van Gorp, T.; Zeillinger, R.; Castillo-Tong, D. C.; Sehouli, J., Prognostic value of
- residual tumor size in patients with epithelial ovarian cancer FIGO stages IIA-IV: analysis of the OVCAD data. *Int J Gynecol Cancer* **2012**, *22* (3), 380-5.
- 359 12. Zhang, L.; Conejo-Garcia, J. R.; Katsaros, D.; Gimotty, P. A.; Massobrio, M.; Regnani, G.;
- 360 Makrigiannakis, A.; Gray, H.; Schlienger, K.; Liebman, M. N.; Rubin, S. C.; Coukos, G.,
- Intratumoral T cells, recurrence, and survival in epithelial ovarian cancer. *N Engl J Med* **2003**, 362 348 (3), 203-13.
- 363 13. Santoiemma, P. P.; Powell, D. J., Tumor infiltrating lymphocytes in ovarian cancer. 364 *Cancer Biol Ther* **2015**, *16* (6), 807-20.
- Rosen, D. G.; Yang, G.; Liu, G.; Mercado-Uribe, I.; Chang, B.; Xiao, X. S.; Zheng, J.; Xue,
- F. X.; Liu, J., Ovarian cancer: pathology, biology, and disease models. *Front Biosci (Landmark Ed)* **2009**, *14*, 2089-102.
- 368 15. Goff, B. A.; Mandel, L.; Muntz, H. G.; Melancon, C. H., Ovarian carcinoma diagnosis. 369 *Cancer* **2000**, *89* (10), 2068-75.
- Holschneider, C. H.; Berek, J. S., Ovarian cancer: epidemiology, biology, and prognostic factors. *Semin Surg Oncol* **2000**, *19* (1), 3-10.
- 372 17. Jacobs, I. J.; Menon, U.; Ryan, A.; Gentry-Maharaj, A.; Burnell, M.; Kalsi, J. K.; Amso, N.
- 373 N.; Apostolidou, S.; Benjamin, E.; Cruickshank, D.; Crump, D. N.; Davies, S. K.; Dawnay, A.;
- Dobbs, S.; Fletcher, G.; Ford, J.; Godfrey, K.; Gunu, R.; Habib, M.; Hallett, R.; Herod, J.;
- 375 Jenkins, H.; Karpinskyj, C.; Leeson, S.; Lewis, S. J.; Liston, W. R.; Lopes, A.; Mould, T.;
- Murdoch, J.; Oram, D.; Rabideau, D. J.; Reynolds, K.; Scott, I.; Seif, M. W.; Sharma, A.; Singh,
- N.; Taylor, J.; Warburton, F.; Widschwendter, M.; Williamson, K.; Woolas, R.; Fallowfield, L.;
- 378 McGuire, A. J.; Campbell, S.; Parmar, M.; Skates, S. J., Ovarian cancer screening and mortality

- in the UK Collaborative Trial of Ovarian Cancer Screening (UKCTOCS): a randomised
- 380 controlled trial. *Lancet* **2016**, *387* (10022), 945-956.
- 381 18. Buys, S. S.; Partridge, E.; Black, A.; Johnson, C. C.; Lamerato, L.; Isaacs, C.; Reding, D.
- 382 J.; Greenlee, R. T.; Yokochi, L. A.; Kessel, B.; Crawford, E. D.; Church, T. R.; Andriole, G. L.;
- Weissfeld, J. L.; Fouad, M. N.; Chia, D.; O'Brien, B.; Ragard, L. R.; Clapp, J. D.; Rathmell, J. M.;
- Riley, T. L.; Hartge, P.; Pinsky, P. F.; Zhu, C. S.; Izmirlian, G.; Kramer, B. S.; Miller, A. B.; Xu, J.
- 385 L.; Prorok, P. C.; Gohagan, J. K.; Berg, C. D.; Team, P. P., Effect of screening on ovarian cancer
- 386 mortality: the Prostate, Lung, Colorectal and Ovarian (PLCO) Cancer Screening Randomized
- 387 Controlled Trial. *JAMA* **2011**, *305* (22), 2295-303.
- 388 19. van Nagell, J. R.; Hoff, J. T., Transvaginal ultrasonography in ovarian cancer screening:
- current perspectives. *Int J Womens Health* **2013**, *6*, 25-33.
- 390 20. Bast, R. C.; Klug, T. L.; St John, E.; Jenison, E.; Niloff, J. M.; Lazarus, H.; Berkowitz, R. S.;
- Leavitt, T.; Griffiths, C. T.; Parker, L.; Zurawski, V. R.; Knapp, R. C., A radioimmunoassay using
- a monoclonal antibody to monitor the course of epithelial ovarian cancer. *N Engl J Med* **1983**,
- 393 *309* (15), 883-7.
- 394 21. Nolen, B. M.; Lokshin, A. E., Biomarker testing for ovarian cancer: clinical utility of
- 395 multiplex assays. *Mol Diagn Ther* **2013**, *17* (3), 139-46.
- 396 22. Høgdall, E. V.; Christensen, L.; Kjaer, S. K.; Blaakaer, J.; Kjaerbye-Thygesen, A.;
- 397 Gayther, S.; Jacobs, I. J.; Høgdall, C. K., CA125 expression pattern, prognosis and correlation
- 398 with serum CA125 in ovarian tumor patients. From The Danish "MALOVA" Ovarian Cancer
- 399 Study. *Gynecol Oncol* **2007**, *104* (3), 508-15.
- 400 23. But, I.; Gorisek, B., Preoperative value of CA 125 as a reflection of tumor grade in
- 401 epithelial ovarian cancer. *Gynecol Oncol* **1996**, *63* (2), 166-72.
- 402 24. Köbel, M.; Kalloger, S. E.; Boyd, N.; McKinney, S.; Mehl, E.; Palmer, C.; Leung, S.;
- Bowen, N. J.; Ionescu, D. N.; Rajput, A.; Prentice, L. M.; Miller, D.; Santos, J.; Swenerton, K.;
- 404 Gilks, C. B.; Huntsman, D., Ovarian carcinoma subtypes are different diseases: implications for
- 405 biomarker studies. *PLoS Med* **2008**, *5* (12), e232.
- 406 25. Johnson, C. C.; Kessel, B.; Riley, T. L.; Ragard, L. R.; Williams, C. R.; Xu, J. L.; Buys, S. S.;
- 407 Prostate, L., C.lorectal and Ovarian Cancer Project Team, The epidemiology of CA-125 in
- 408 women without evidence of ovarian cancer in the Prostate, Lung, Colorectal and Ovarian
- 409 Cancer (PLCO) Screening Trial. *Gynecol Oncol* **2008**, *110* (3), 383-9.
- 410 26. Szekanecz, E.; Sándor, Z.; Antal-Szalmás, P.; Soós, L.; Lakos, G.; Besenyei, T.;
- 411 Szentpétery, A.; Simkovics, E.; Szántó, J.; Kiss, E.; Koch, A. E.; Szekanecz, Z., Increased
- 412 production of the soluble tumor-associated antigens CA19-9, CA125, and CA15-3 in
- rheumatoid arthritis: potential adhesion molecules in synovial inflammation? *Ann N Y Acad*
- 414 *Sci* **2007**, *1108*, 359-71.
- 415 27. Badgwell, D.; Bast, R. C., Early detection of ovarian cancer. Dis Markers 2007, 23 (5-6),
- 416 397-410.
- 417 28. Anastasi, E.; Granato, T.; Marchei, G. G.; Viggiani, V.; Colaprisca, B.; Comploj, S.; Reale,
- 418 M. G.; Frati, L.; Midulla, C., Ovarian tumor marker HE4 is differently expressed during the
- phases of the menstrual cycle in healthy young women. *Tumour Biol* **2010**, *31* (5), 411-5.
- 420 29. Meinhold-Heerlein, I.; Hauptmann, S., The heterogeneity of ovarian cancer. Arch
- 421 *Gynecol Obstet* **2014**, *289* (2), 237-9.
- 422 30. Schummer, M.; Ng, W. V.; Bumgarner, R. E.; Nelson, P. S.; Schummer, B.; Bednarski, D.
- 423 W.; Hassell, L.; Baldwin, R. L.; Karlan, B. Y.; Hood, L., Comparative hybridization of an array of
- 424 21,500 ovarian cDNAs for the discovery of genes overexpressed in ovarian carcinomas. *Gene*
- 425 **1999,** *238* (2), 375-85.
- 426 31. Hellström, I.; Raycraft, J.; Hayden-Ledbetter, M.; Ledbetter, J. A.; Schummer, M.;
- 427 McIntosh, M.; Drescher, C.; Urban, N.; Hellström, K. E., The HE4 (WFDC2) protein is a
- 428 biomarker for ovarian carcinoma. *Cancer Res* **2003**, *63* (13), 3695-700.

- 429 Moore, R. G.; Brown, A. K.; Miller, M. C.; Skates, S.; Allard, W. J.; Verch, T.; Steinhoff, 32.
- 430 M.; Messerlian, G.; DiSilvestro, P.; Granai, C. O.; Bast, R. C., The use of multiple novel tumor
- 431 biomarkers for the detection of ovarian carcinoma in patients with a pelvic mass. *Gynecol*
- 432 Oncol 2008, 108 (2), 402-8.
- 433 Ferraro, S.; Schiumarini, D.; Panteghini, M., Human epididymis protein 4: factors of
- 434 variation. Clin Chim Acta 2015, 438, 171-7.
- 34. Ferraro, S.; Robbiano, C.; Tosca, N.; Panzeri, A.; Paganoni, A. M.; Panteghini, M., Serum 435
- 436 human epididymis protein 4 vs. carbohydrate antigen 125 in ovarian cancer follow-up. Clin
- 437 Biochem 2018, 60, 84-90.
- 438 Moore, R. G.; Miller, M. C.; Disilvestro, P.; Landrum, L. M.; Gajewski, W.; Ball, J. J.;
- 439 Skates, S. J., Evaluation of the diagnostic accuracy of the risk of ovarian malignancy algorithm
- 440 in women with a pelvic mass. *Obstet Gynecol* **2011**, *118* (2 Pt 1), 280-8.
- 441 Jacobs, I.; Oram, D.; Fairbanks, J.; Turner, J.; Frost, C.; Grudzinskas, J. G., A risk of
- 442 malignancy index incorporating CA 125, ultrasound and menopausal status for the accurate
- 443 preoperative diagnosis of ovarian cancer. Br J Obstet Gynaecol 1990, 97 (10), 922-9.
- 444 Ueland, F. R.; Desimone, C. P.; Seamon, L. G.; Miller, R. A.; Goodrich, S.; Podzielinski, I.;
- 445 Sokoll, L.; Smith, A.; van Nagell, J. R.; Zhang, Z., Effectiveness of a multivariate index assay in
- 446 the preoperative assessment of ovarian tumors. *Obstet Gynecol* **2011**, *117* (6), 1289-97.
- Hessvik, N. P.; Llorente, A., Current knowledge on exosome biogenesis and release. Cell 447
- 448 Mol Life Sci **2018**, 75 (2), 193-208.
- Di Vizio, D.; Morello, M.; Dudley, A. C.; Schow, P. W.; Adam, R. M.; Morley, S.; 449
- 450 Mulholland, D.; Rotinen, M.; Hager, M. H.; Insabato, L.; Moses, M. A.; Demichelis, F.; Lisanti,
- 451 M. P.; Wu, H.; Klagsbrun, M.; Bhowmick, N. A.; Rubin, M. A.; D'Souza-Schorey, C.; Freeman,
- 452 M. R., Large oncosomes in human prostate cancer tissues and in the circulation of mice with
- metastatic disease. Am J Pathol 2012, 181 (5), 1573-84. 453
- Minciacchi, V. R.; You, S.; Spinelli, C.; Morley, S.; Zandian, M.; Aspuria, P. J.; Cavallini, 454 40.
- L.; Ciardiello, C.; Reis Sobreiro, M.; Morello, M.; Kharmate, G.; Jang, S. C.; Kim, D. K.; 455
- 456 Hosseini-Beheshti, E.; Tomlinson Guns, E.; Gleave, M.; Gho, Y. S.; Mathivanan, S.; Yang, W.;
- 457 Freeman, M. R.; Di Vizio, D., Large oncosomes contain distinct protein cargo and represent a
- 458 separate functional class of tumor-derived extracellular vesicles. Oncotarget 2015, 6 (13),
- 459 11327-41.
- 460 Zhang, H.; Freitas, D.; Kim, H. S.; Fabijanic, K.; Li, Z.; Chen, H.; Mark, M. T.; Molina, H.;
- 461 Martin, A. B.; Bojmar, L.; Fang, J.; Rampersaud, S.; Hoshino, A.; Matei, I.; Kenific, C. M.;
- 462 Nakajima, M.; Mutvei, A. P.; Sansone, P.; Buehring, W.; Wang, H.; Jimenez, J. P.; Cohen-Gould,
- 463 L.; Paknejad, N.; Brendel, M.; Manova-Todorova, K.; Magalhães, A.; Ferreira, J. A.; Osório, H.;
- 464 Silva, A. M.; Massey, A.; Cubillos-Ruiz, J. R.; Galletti, G.; Giannakakou, P.; Cuervo, A. M.;
- 465
- Blenis, J.; Schwartz, R.; Brady, M. S.; Peinado, H.; Bromberg, J.; Matsui, H.; Reis, C. A.; Lyden,
- 466 D., Identification of distinct nanoparticles and subsets of extracellular vesicles by asymmetric
- 467 flow field-flow fractionation. Nat Cell Biol 2018, 20 (3), 332-343.
- Johnstone, R. M.; Adam, M.; Hammond, J. R.; Orr, L.; Turbide, C., Vesicle formation 468
- 469 during reticulocyte maturation. Association of plasma membrane activities with released
- 470 vesicles (exosomes). J Biol Chem 1987, 262 (19), 9412-20.
- 471 Yáñez-Mó, M.; Siljander, P. R.; Andreu, Z.; Zavec, A. B.; Borràs, F. E.; Buzas, E. I.; 43.
- 472 Buzas, K.; Casal, E.; Cappello, F.; Carvalho, J.; Colás, E.; Cordeiro-da Silva, A.; Fais, S.; Falcon-
- Perez, J. M.; Ghobrial, I. M.; Giebel, B.; Gimona, M.; Graner, M.; Gursel, I.; Gursel, M.; 473 474
- Heegaard, N. H.; Hendrix, A.; Kierulf, P.; Kokubun, K.; Kosanovic, M.; Kralj-Iglic, V.; Krämer-
- 475 Albers, E. M.; Laitinen, S.; Lässer, C.; Lener, T.; Ligeti, E.; Linē, A.; Lipps, G.; Llorente, A.;
- 476 Lötvall, J.; Manček-Keber, M.; Marcilla, A.; Mittelbrunn, M.; Nazarenko, I.; Nolte-'t Hoen, E.
- N.; Nyman, T. A.; O'Driscoll, L.; Olivan, M.; Oliveira, C.; Pállinger, É.; Del Portillo, H. A.; 477
- Reventós, J.; Rigau, M.; Rohde, E.; Sammar, M.; Sánchez-Madrid, F.; Santarém, N.; 478

- 479 Schallmoser, K.; Ostenfeld, M. S.; Stoorvogel, W.; Stukelj, R.; Van der Grein, S. G.;
- Vasconcelos, M. H.; Wauben, M. H.; De Wever, O., Biological properties of extracellular vesicles
- and their physiological functions. *J Extracell Vesicles* **2015**, *4*, 27066.
- 482 44. Kim, D. K.; Lee, J.; Simpson, R. J.; Lötvall, J.; Gho, Y. S., EVpedia: A community web
- resource for prokaryotic and eukaryotic extracellular vesicles research. Semin Cell Dev Biol
- 484 **2015,** *40*, 4-7.
- 485 45. Février, B.; Raposo, G., Exosomes: endosomal-derived vesicles shipping extracellular
- 486 messages. *Curr Opin Cell Biol* **2004,** *16* (4), 415-21.
- 487 46. Skog, J.; Würdinger, T.; van Rijn, S.; Meijer, D. H.; Gainche, L.; Sena-Esteves, M.; Curry,
- 488 W. T.; Carter, B. S.; Krichevsky, A. M.; Breakefield, X. O., Glioblastoma microvesicles transport
- 489 RNA and proteins that promote tumour growth and provide diagnostic biomarkers. *Nat Cell*
- 490 *Biol* **2008**, *10* (12), 1470-6.
- 491 47. Mulcahy, L. A.; Pink, R. C.; Carter, D. R., Routes and mechanisms of extracellular vesicle
- 492 uptake. J Extracell Vesicles 2014, 3.
- 493 48. Lane, R. E.; Korbie, D.; Hill, M. M.; Trau, M., Extracellular vesicles as circulating cancer
- 494 biomarkers: opportunities and challenges. *Clin Transl Med* **2018,** *7* (1), 14.
- 495 49. Wei, Z.; Batagov, A. O.; Carter, D. R.; Krichevsky, A. M., Fetal Bovine Serum RNA
- Interferes with the Cell Culture derived Extracellular RNA. *Sci Rep* **2016**, *6*, 31175.
- 497 50. Ramirez, M. I.; Amorim, M. G.; Gadelha, C.; Milic, I.; Welsh, J. A.; Freitas, V. M.; Nawaz,
- 498 M.; Akbar, N.; Couch, Y.; Makin, L.; Cooke, F.; Vettore, A. L.; Batista, P. X.; Freezor, R.; Pezuk,
- 499 J. A.; Rosa-Fernandes, L.; Carreira, A. C. O.; Devitt, A.; Jacobs, L.; Silva, I. T.; Coakley, G.;
- Nunes, D. N.; Carter, D.; Palmisano, G.; Dias-Neto, E., Technical challenges of working with
- 501 extracellular vesicles. *Nanoscale* **2018**, *10* (3), 881-906.
- 502 51. Witwer, K. W.; Buzás, E. I.; Bemis, L. T.; Bora, A.; Lässer, C.; Lötvall, J.; Nolte-'t Hoen,
- 503 E. N.; Piper, M. G.; Sivaraman, S.; Skog, J.; Théry, C.; Wauben, M. H.; Hochberg, F.,
- 504 Standardization of sample collection, isolation and analysis methods in extracellular vesicle
- research. *J Extracell Vesicles* **2013**, *2*.
- 506 52. Mateescu, B.; Kowal, E. J.; van Balkom, B. W.; Bartel, S.; Bhattacharyya, S. N.; Buzás, E.
- 507 I.; Buck, A. H.; de Candia, P.; Chow, F. W.; Das, S.; Driedonks, T. A.; Fernández-Messina, L.;
- Haderk, F.; Hill, A. F.; Jones, J. C.; Van Keuren-Jensen, K. R.; Lai, C. P.; Lässer, C.; Liegro, I. D.;
- 509 Lunavat, T. R.; Lorenowicz, M. J.; Maas, S. L.; Mäger, I.; Mittelbrunn, M.; Momma, S.;
- Mukherjee, K.; Nawaz, M.; Pegtel, D. M.; Pfaffl, M. W.; Schiffelers, R. M.; Tahara, H.; Théry, C.;
- Tosar, J. P.; Wauben, M. H.; Witwer, K. W.; Nolte-'t Hoen, E. N., Obstacles and opportunities in
- 512 the functional analysis of extracellular vesicle RNA an ISEV position paper. *J Extracell Vesicles*
- **2017**, *6* (1), 1286095.
- 514 53. Pink, R. C.; Elmusrati, A. A.; Lambert, D.; Carter, D. R. F., Royal Society Scientific
- Meeting: Extracellular vesicles in the tumour microenvironment. Philos Trans R Soc Lond B
- 516 *Biol Sci* **2018**, *373* (1737).
- 517 54. Weidle, U. H.; Birzele, F.; Kollmorgen, G.; Rüger, R., The Multiple Roles of Exosomes in
- Metastasis. *Cancer Genomics Proteomics* **2017**, *14* (1), 1-15.
- 519 55. Song, W.; Yan, D.; Wei, T.; Liu, Q.; Zhou, X.; Liu, J., Tumor-derived extracellular
- vesicles in angiogenesis. *Biomed Pharmacother* **2018**, *102*, 1203-1208.
- 521 56. Samuel, P.; Fabbri, M.; Carter, D. R. F., Mechanisms of Drug Resistance in Cancer: The
- Role of Extracellular Vesicles. *Proteomics* **2017**, *17* (23-24).
- 523 57. Samuel, P.; Mulcahy, L. A.; Furlong, F.; McCarthy, H. O.; Brooks, S. A.; Fabbri, M.; Pink,
- R. C.; Carter, D. R. F., Cisplatin induces the release of extracellular vesicles from ovarian cancer
- 525 cells that can induce invasiveness and drug resistance in bystander cells. *Philos Trans R Soc*
- 526 Lond B Biol Sci **2018**, 373 (1737).

- 527 58. Bewicke-Copley, F.; Mulcahy, L. A.; Jacobs, L. A.; Samuel, P.; Akbar, N.; Pink, R. C.;
- 528 Carter, D. R. F., Extracellular vesicles released following heat stress induce bystander effect in
- unstressed populations. J Extracell Vesicles 2017, 6 (1), 1340746.
- 530 59. Al-Mayah, A. H.; Irons, S. L.; Pink, R. C.; Carter, D. R.; Kadhim, M. A., Possible Role of
- 531 Exosomes Containing RNA in Mediating Nontargeted Effect of Ionizing Radiation. *Radiat Res*
- 532 **2012**.
- 533 60. Bartel, D. P., MicroRNAs: target recognition and regulatory functions. *Cell* **2009**, *136*
- 534 (2), 215-33.
- 535 61. Jacobs, L. A.; Bewicke-Copley, F.; Poolman, M. G.; Pink, R. C.; Mulcahy, L. A.; Baker, I.;
- Beaman, E. M.; Brooks, T.; Caley, D. P.; Cowling, W.; Currie, J. M.; Horsburgh, J.; Kenehan, L.;
- Keyes, E.; Leite, D.; Massa, D.; McDermott-Rouse, A.; Samuel, P.; Wood, H.; Kadhim, M.;
- 538 Carter, D. R., Meta-analysis using a novel database, miRStress, reveals miRNAs that are
- frequently associated with the radiation and hypoxia stress-responses. *PLoS One* **2013**, *8* (11),
- 540 e80844.
- 541 62. Pink, R. C.; Samuel, P.; Massa, D.; Caley, D. P.; Brooks, S. A.; Carter, D. R., The
- passenger strand, miR-21-3p, plays a role in mediating cisplatin resistance in ovarian cancer
- 543 cells. *Gynecol Oncol* **2015**, *137* (1), 143-51.
- 544 63. Samuel, P.; Pink, R. C.; Caley, D. P.; Currie, J. M.; Brooks, S. A.; Carter, D. R., Over-
- expression of miR-31 or loss of KCNMA1 leads to increased cisplatin resistance in ovarian
- cancer cells. *Tumour Biol* **2015**.
- 547 64. Samuel, P.; Pink, R. C.; Brooks, S. A.; Carter, D. R., miRNAs and ovarian cancer: a miRiad
- of mechanisms to induce cisplatin drug resistance. Expert Rev Anticancer Ther **2016**, 16 (1),
- 549 57-70.
- 550 65. Zaman, M. S.; Maher, D. M.; Khan, S.; Jaggi, M.; Chauhan, S. C., Current status and
- implications of microRNAs in ovarian cancer diagnosis and therapy. J Ovarian Res 2012, 5 (1),
- 552 44.
- 553 66. King, H. W.; Michael, M. Z.; Gleadle, J. M., Hypoxic enhancement of exosome release by
- breast cancer cells. *BMC Cancer* **2012**, *12*, 421.
- 555 67. Meng, X.; Müller, V.; Milde-Langosch, K.; Trillsch, F.; Pantel, K.; Schwarzenbach, H.,
- 556 Diagnostic and prognostic relevance of circulating exosomal miR-373, miR-200a, miR-200b
- and miR-200c in patients with epithelial ovarian cancer. *Oncotarget* **2016**, *7* (13), 16923-35.
- 558 68. Taylor, D. D.; Gercel-Taylor, C., MicroRNA signatures of tumor-derived exosomes as
- diagnostic biomarkers of ovarian cancer. *Gynecol Oncol* **2008,** *110* (1), 13-21.
- 560 69. Mitchell, P. S.; Parkin, R. K.; Kroh, E. M.; Fritz, B. R.; Wyman, S. K.; Pogosova-
- Agadjanyan, E. L.; Peterson, A.; Noteboom, J.; O'Briant, K. C.; Allen, A.; Lin, D. W.; Urban, N.;
- Drescher, C. W.; Knudsen, B. S.; Stirewalt, D. L.; Gentleman, R.; Vessella, R. L.; Nelson, P. S.;
- Martin, D. B.; Tewari, M., Circulating microRNAs as stable blood-based markers for cancer
- detection. *Proc Natl Acad Sci U S A* **2008,** *105* (30), 10513-8.
- 565 70. Amorim, M. G.; Valieris, R.; Drummond, R. D.; Pizzi, M. P.; Freitas, V. M.; Sinigaglia-
- Coimbra, R.; Calin, G. A.; Pasqualini, R.; Arap, W.; Silva, I. T.; Dias-Neto, E.; Nunes, D. N., A
- total transcriptome profiling method for plasma-derived extracellular vesicles: applications
- 568 for liquid biopsies. *Sci Rep* **2017**, *7* (1), 14395.
- 569 71. Samuel, P.; Carter, D. R., The Diagnostic and Prognostic Potential of microRNAs in
- 570 Epithelial Ovarian Carcinoma. *Mol Diagn Ther* **2017**, *21* (1), 59-73.
- 72. Vaksman, O.; Tropé, C.; Davidson, B.; Reich, R., Exosome-derived miRNAs and ovarian
- carcinoma progression. *Carcinogenesis* **2014**, *35* (9), 2113-20.
- 573 73. Yokoi, A.; Yoshioka, Y.; Hirakawa, A.; Yamamoto, Y.; Ishikawa, M.; Ikeda, S. I.; Kato,
- 574 T.; Niimi, K.; Kajiyama, H.; Kikkawa, F.; Ochiya, T., A combination of circulating miRNAs for
- 575 the early detection of ovarian cancer. *Oncotarget* **2017**, *8* (52), 89811-89823.

- 576 74. Cappellesso, R.; Tinazzi, A.; Giurici, T.; Simonato, F.; Guzzardo, V.; Ventura, L.;
- 577 Crescenzi, M.; Chiarelli, S.; Fassina, A., Programmed cell death 4 and microRNA 21 inverse
- 578 expression is maintained in cells and exosomes from ovarian serous carcinoma effusions.
- 579 *Cancer Cytopathol* **2014,** *122* (9), 685-93.
- 580 75. Pan, C.; Stevic, I.; Müller, V.; Ni, Q.; Ferrer, L. O.; Pantel, K.; Schwarzenbach, H.,
- Exosomal microRNAs as tumor markers in epithelial ovarian cancer. *Mol Oncol* **2018**.
- 582 76. Zhou, J.; Gong, G.; Tan, H.; Dai, F.; Zhu, X.; Chen, Y.; Wang, J.; Liu, Y.; Chen, P.; Wu, X.;
- 583 Wen, J., Urinary microRNA-30a-5p is a potential biomarker for ovarian serous
- adenocarcinoma. *Oncol Rep* **2015**, *33* (6), 2915-23.
- 77. Peng, P.; Yan, Y.; Keng, S., Exosomes in the ascites of ovarian cancer patients: origin
- and effects on anti-tumor immunity. *Oncol Rep* **2011**, *25* (3), 749-62.
- 587 78. Li, J.; Sherman-Baust, C. A.; Tsai-Turton, M.; Bristow, R. E.; Roden, R. B.; Morin, P. J.,
- 588 Claudin-containing exosomes in the peripheral circulation of women with ovarian cancer.
- 589 *BMC Cancer* **2009**, *9*, 244.
- 590 79. Szajnik, M.; Derbis, M.; Lach, M.; Patalas, P.; Michalak, M.; Drzewiecka, H.; Szpurek,
- D.; Nowakowski, A.; Spaczynski, M.; Baranowski, W.; Whiteside, T. L., Exosomes in Plasma of
- Patients with Ovarian Carcinoma: Potential Biomarkers of Tumor Progression and Response
- to Therapy. *Gynecol Obstet (Sunnyvale)* **2013**, *Suppl* 4, 3.
- 594 80. Keller, S.; König, A. K.; Marmé, F.; Runz, S.; Wolterink, S.; Koensgen, D.; Mustea, A.;
- 595 Sehouli, J.; Altevogt, P., Systemic presence and tumor-growth promoting effect of ovarian
- 596 carcinoma released exosomes. *Cancer Lett* **2009**, *278* (1), 73-81.
- 597 81. Zhang, P.; He, M.; Zeng, Y., Ultrasensitive microfluidic analysis of circulating exosomes
- using a nanostructured graphene oxide/polydopamine coating. Lab Chip 2016, 16 (16), 3033-
- 599 42.
- 82. Runz, S.; Keller, S.; Rupp, C.; Stoeck, A.; Issa, Y.; Koensgen, D.; Mustea, A.; Sehouli, J.;
- 601 Kristiansen, G.; Altevogt, P., Malignant ascites-derived exosomes of ovarian carcinoma
- 602 patients contain CD24 and EpCAM. *Gynecol Oncol* **2007**, *107* (3), 563-71.
- 603 83. Im, H.; Shao, H.; Park, Y. I.; Peterson, V. M.; Castro, C. M.; Weissleder, R.; Lee, H.,
- 604 Label-free detection and molecular profiling of exosomes with a nano-plasmonic sensor. *Nat*
- 605 Biotechnol **2014**, 32 (5), 490-5.
- 84. Zhao, Z.; Yang, Y.; Zeng, Y.; He, M., A microfluidic ExoSearch chip for multiplexed
- exosome detection towards blood-based ovarian cancer diagnosis. Lab Chip 2016, 16 (3),
- 608 489-96.
- 609 85. Hisey, C. L.; Dorayappan, K. D. P.; Cohn, D. E.; Selvendiran, K.; Hansford, D. J.,
- 610 Microfluidic affinity separation chip for selective capture and release of label-free ovarian
- 611 cancer exosomes. *Lab Chip* **2018**.
- 612 86. Tang, M. K. S.; Yue, P. Y. K.; Ip, P. P.; Huang, R. L.; Lai, H. C.; Cheung, A. N. Y.; Tse, K. Y.;
- Ngan, H. Y. S.; Wong, A. S. T., Soluble E-cadherin promotes tumor angiogenesis and localizes to
- 614 exosome surface. *Nat Commun* **2018,** 9 (1), 2270.
- 615 87. Lea, J.; Sharma, R.; Yang, F.; Zhu, H.; Ward, E. S.; Schroit, A. J., Detection of
- phosphatidylserine-positive exosomes as a diagnostic marker for ovarian malignancies: a
- 617 proof of concept study. *Oncotarget* **2017**, *8* (9), 14395-14407.
- 618 88. Carney, R. P.; Hazari, S.; Colquhoun, M.; Tran, D.; Hwang, B.; Mulligan, M. S.; Bryers, J.
- 619 D.; Girda, E.; Leiserowitz, G. S.; Smith, Z. J.; Lam, K. S., Multispectral Optical Tweezers for
- 620 Biochemical Fingerprinting of CD9-Positive Exosome Subpopulations. Anal Chem 2017, 89
- 621 (10), 5357-5363.
- 622 89. Claussen, C.; Rausch, A. V.; Lezius, S.; Amirkhosravi, A.; Davila, M.; Francis, J. L.;
- 623 Hisada, Y. M.; Mackman, N.; Bokemeyer, C.; Schmalfeldt, B.; Mahner, S.; Langer, F.,
- Microvesicle-associated tissue factor procoagulant activity for the preoperative diagnosis of
- ovarian cancer. *Thromb Res* **2016**, *141*, 39-48.

- 90. Zhou, Q.; Li, W.; Leng, B.; Zheng, W.; He, Z.; Zuo, M.; Chen, A., Circulating Cell Free DNA as the Diagnostic Marker for Ovarian Cancer: A Systematic Review and Meta-Analysis. *PLoS One* **2016**, *11* (6), e0155495.
- 629 91. Kamat, A. A.; Baldwin, M.; Urbauer, D.; Dang, D.; Han, L. Y.; Godwin, A.; Karlan, B. Y.; 630 Simpson, J. L.; Gershenson, D. M.; Coleman, R. L.; Bischoff, F. Z.; Sood, A. K., Plasma cell-free
- DNA in ovarian cancer: an independent prognostic biomarker. *Cancer* **2010**, *116* (8), 1918-25. G2. Karimi, N.; Cvjetkovic, A.; Jang, S. C.; Crescitelli, R.; Hosseinpour Feizi, M. A.;
- Nieuwland, R.; Lötvall, J.; Lässer, C., Detailed analysis of the plasma extracellular vesicle proteome after separation from lipoproteins. *Cell Mol Life Sci* **2018**, *75* (15), 2873-2886.
- 93. Van Deun, J.; Mestdagh, P.; Sormunen, R.; Cocquyt, V.; Vermaelen, K.; Vandesompele, J.; Bracke, M.; De Wever, O.; Hendrix, A., The impact of disparate isolation methods for extracellular vesicles on downstream RNA profiling. *J Extracell Vesicles* **2014**, *3*.
 - 94. Soares Martins, T.; Catita, J.; Martins Rosa, I.; A B da Cruz E Silva, O.; Henriques, A. G., Exosome isolation from distinct biofluids using precipitation and column-based approaches. *PLoS One* **2018**, *13* (6), e0198820.

Figures/Tables

638

639

640641642

643644645

646 647

648

649

650

651

655

656

657

Figure 1 – Illustration of biogenesis pathways for EVs. Microvesicles (MVs) are directly released by outward budding of the plasma membrane. The precursors for exosomes are formed inside multivesicular bodies (MVBs) as intraluminal vesicles (ILVs). MVBs can fuse with the lysosomes leading to the degradation of their content or fuse with the PM leading to the release of exosomes into the extracellular space. Exosomes can then be taken up by recipient cells through different pathways leading to the transfer of their cargo and potentially modifying the behavior of recipient cells.

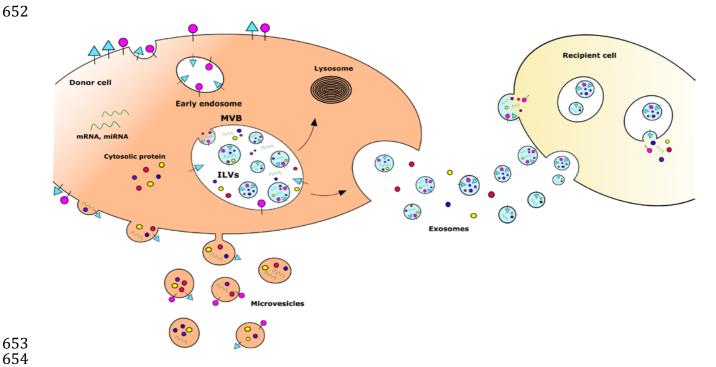


Table 1 – EV components identified in EOCs patients that could be used as potential biomarkers for diagnosis or prognosis of ovarian cancer patients.

Biomarker type	Candidate	Sample source	Comment	Reference
Proteins	CA125	Ascites	Vesicular CA125 higher than circulating CA125 in early stage	Peng P <i>et al</i> 2011 ⁷⁷
	Claudin 4	Plasma and cell lines	Abundant in EVs from plasma of OC patients in late stages (potential use in combination with CA125)	Li J <i>et al</i> 2009 ⁷⁸
	TGFβ1 and MAGE3	Plasma	Higher in plasma from OCs patients. Total EV-protein concentration decreased in responsive patients after treatment	Szajnik M <i>et al</i> 2013 ⁷⁹
	ADAM10, EpCAM and EMMPRIN	Ascites and serum	Ascites EVs correlates with serum EV levels and carry late stage disease markers	Keller S <i>et al</i> 2009 ⁸⁰ , Taylor D and Gercel-Taylor 2008 ⁶⁸
	CD24	Ascites and cell lines	High CD24 is associated with poor prognosis in OC and is released in tumour-derived EVs	Runz S <i>et al</i> 2007 ⁸²
	CD24 and EpCAM	Ascites	Expression is associated with chemotherapy response	Im H et al 2014 83
	EpCAM	Serum	EpCAM positive EVs increase in plasma of stage IV patients	,
	E-cadherin	Ascites and non- cancer ovarian cell line	Can help distinguish between healthy and benign disease	Tang MKS <i>et al</i> 2018 ⁸⁶
miRNA	miR-21, miR-141, miR-200, miR-214	Serum	Levels of these miRNAs were similar in the EVs and tumour cells and were predictive of disease stage	Taylor D and Gercel-Taylor 2008 ⁶⁸

	miR-200 a/b/c and miR-373	Serum	Higher levels in EOC patients. miR200 specific for malignant disease. miR200b/c higher in patients with stage III-IV disease and are associated with CA125 level	Meng X <i>et al</i> 2016 ⁶⁷
	miR- 21, miR- 23b and miR- 29a	Ascites and pleural effusion	Associated with poor survival	Vaksman O et al 2014 ⁷²
	miR-142-3p, miR- 26a-5p, let-7d-5p, miR-374a-5p, miR- 766-3p, miR-200a- 3p, miR-328-3p, miR-130b-3p	Serum	Specific for early-stage. Most of these found in EVs	Yokoi A <i>et al</i> 2017 ⁷³
	miR-21	Ascites	Up-regulated in malignant cells and tumour-derived EVs	Cappellesso R et al 2014 ⁷⁴
	miR-200b and miR-320	Plasma	EV levels higher in stage IV patients. Both miRNA are higher in patients compared with healthy controls and they are positively correlated with CA125	Pan C <i>et al</i> 2018 ⁷⁵
	miR-30a-5p	Urine	Higher in urine of ovarian cancer patients, particularly in stage I and II	Zhou J <i>et al</i> 2015 ⁷⁶
Other	Phosphatidylserine (PS)	Plasma and cell lines	Higher in cancer-derived EVs	Lea J <i>et al</i> 2017 ⁸⁷
	Microvesicle- associated TF procoagulant activity	Plasma	Improved diagnostic benefit when combined with CA-125 levels	Claussen C <i>et al</i> 2016 ⁸⁹