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The ecological niche of *Echinococcus multilocularis* in North America: understanding biotic and abiotic determinants of parasite distribution with new records in New Mexico and Maryland, United States

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Understanding the factors shaping the niche of parasites and its expression over geographical space and through time continues to be a modern scientific challenge with the results of research in this area directly influencing both theoretical and applied biology. This is especially important for proactive management of zoonotic parasites such as Echinococcus multilocularis, the etiologic agent of alveolar echinococcosis. Echinococcus multilocularis has a Holarctic distribution; with its geographic range and prevalence increasing recently in areas of the western Palearctic, while its distribution dynamics are poorly understood in the Nearctic. In this paper, we use an ecological niche modeling (ENM) approach to: i) estimate the current spatial distribution of suitable conditions for the parasite in the Nearctic. ii) Evaluate the abiotic and biotic factors influencing the species distribution. iii) Assess the potential impact of climatic change on the distribution of this species in the Nearctic. Additionally, we report two new occurrence records of this parasite that significantly expands its known geographic range. We reviewed the occurrence records of E. multilocularis for the Nearctic. This was complemented by two new records of the species from Maryland and New Mexico identified using morphology and multivariate morphometrics of the rostellar hooks. From these data we created two ENMs using the software Maxent. The first ENM included climatic variables, while the second included the same abiotic data plus biotic information consisting of four host community-related data sets. We evaluated model performance and variable importance to explore the relation of these variables to the parasite niche. Finally, we projected the resulting niche model onto future climate change scenarios. We found that an important portion of the Nearctic has suitable conditions for E. multilocularis with adequate habitat in the West and East of the continent where the parasite has not been detected. We also found that the proposed biotic variables improve the model performance and provide unique information, while the most critical abiotic variable was related to the amount of solar radiation. Finally, under the future emission scenarios explored, the distribution of suitable habitat for the parasite is predicted to increase by 56 % to 76 %. We obtained a robust model that provides detail on the distribution of suitable areas for E. multilocularis, including areas that have not been explored for the presence of the parasite. The host community variables included in this study seem a promising way to include biotic data for ecological parasite niche modeling.

El estudio de los factores que moldean el nicho de los parásitos y como este se expresa en la distribución espacial y temporal de estos organismos es un reto de importancia para la biología aplicada y teórica. Esta información puede ser de especial importancia para parásitos zoonóticos tales como Echinococcus multilocularis, el cestodo causante de echinococcosis alveolar. Este parasito presenta una distribución Holártica, con un incremento reciente en rango geográfico y prevalencia documentados en Asia y Europa, mientras que en el Neártico, se desconoce la dinámica de distribución de la especie. En este estudio usamos modelos de nicho ecológico para: i) estimar la distribución actual de hábitat para la especie en Norteamérica. ii) Evaluar el efecto de factores bióticos y abióticos sobre la distribución de este parasito. iii) Evaluar el impacto potencial del cambio climático sobre su distribución. Adicionalmente, reportamos dos nuevos registros para la especie. En este trabajo revisamos los registros de ocurrencia de E. multilocularis en el Neártico. Esta información es complementada con dos nuevos registros provenientes de Maryland y Nuevo México identificados a partir de análisis morfológicos y morfométricos. Empleando el software Maxent, creamos dos modelos de nicho a partir de estos registros. El primer modelo se basó únicamente en variables abióticas, mientras que el segundo además de incluir las variables abióticas incluyó variables bióticas relacionadas con la comunidad de hospederos potenciales. Evaluamos el desempeno de cada modelo y la contribución de cada variable para explorar la relación de estas con el nicho del parásito. Finalmente, proyectamos los modelos al futuro bajo dos escenarios de emisiones de CO₃. Encontramos que existen condiciones adecuadas para la especie en una porción importante del área de estudio, con áreas predichas al Este y Occidente del continente donde no se ha registrado el parásito. La inclusión de las variables bióticas resulta en modelos con mejor desempeño, así mismo, se evidencio que estas variables presentan información única no contenida en otras capas. La radiación solar fue la variable abiótica de mayor importancia. Finalmente, bajo los escenarios de cambio climático explorados, el área de hábitat adecuado para el parasito presenta un importante aumento de entre el 56 % y 76 %. En este trabajo obtuvimos un modelo robusto y detallado de la distribución de las condiciones ambientales adecuadas para E. multilocularis, el cual incluye zonas donde no ha sido reportada la presencia del parásito. Las variables relacionadas con la comunidad de hospederos incluidas en este trabajo parecen ser una manera prometedora de incluir información biótica en modelos de nicho de simbiontes.

Key words: Carnivora; disease geography; Echinococcosis; *Echinococcus multilocularis*; ecological niche model; maxent; Nearctic; parasite; Rodentia; multivariate statistics.

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Introduction

Emerging infectious diseases represent a major threat derived from accelerating climate change. We need to take proactive measures with respect to coping with the emerging disease crisis. One impediment in that task is a lack of proper documentation of the underlying biological factors influencing geographic distributions of pathogens and their hosts and vectors (Brooks et al. 2014; Sehgal et al. 2011; Escobar and Craft 2016). This is because the geographic distributions of pathogens, actual and potential hosts or vectors, and of active foci of disease associated with a given pathogen are often complex. Ecological niche modeling is a powerful tool for summarizing data about species biotic and abiotic requirements and their expression on geographical distributions, mapping known sites of occurrence and anticipating potential sites or areas of successful invasion. Accumulating and then monitoring these data provides information about changes in current distributions and interactions of parasites with their hosts that may aid in projecting suitable future pathogen distributions and potential disease outbreaks (Pickles et al. 2013; Ostfeld and Brunner 2015; Nylin et al. 2018). In a complementary manner, combining information about actual and projected environmental changes may allow us to prioritize monitoring activities to maximize the possibility of anticipating the arrival of pathogens, especially agents of endemic, emergent, and re-emergent zoonotic diseases, in novel areas. Knowing the transmission dynamics and potential hosts and vectors of any given pathogen in an anticipated site of invasion allows us to mitigate that pathogen's impact on wildlife communities and possible cascading zoonotic impacts on humans and domestic animals (Brooks and Boeger 2019; Brooks et al. 2014; Estrada-Peña et al. 2014). The cestode Echinococcus multilocularis Leuckart 1863, the etiological agent of alveolar echinococcosis in humans, is an excellent model system for applying these principles.

Echinococcus multilocularis is a small tapeworm with an adult form that occurs naturally in the small intestines of wild canids and less frequently in other carnivores throughout the northern regions of the Holarctic (Leiby and Kritsky 1972). The normal life-cycle includes a metastatic-proliferative larval stage in the visceral organs of muroid rodents (Rausch and Richards 1971). Alveolar echinococcosis occurs when humans accidentally ingest the tapeworm eggs through direct contact with the feces of infected dogs or wild canids or through environmental contamination (Rausch and Schiller 1954; Lass et al. 2015), and the larval cyst proliferates in the liver, lungs, or other tissues. In humans, this cyst slowly develops, eventually causing a disease that clinically show tumor-like symptoms with infiltrative growth and the potential for metastasis (Kern et al. 2017). Alveolar echinococcosis has been noted as an important neglected zoonotic disease (Da Silva 2010; Budke et al. 2017) and when the human infection data are combined and summarized with the cases of cystic echinococcosis, produced by infection with E. granulosus, around 18,000 new human cases of echinococcosis per year appear globally (Torgerson <u>et al. 2010</u>). If left untreated, 90 % of alveolar echinococcosis cases are fatal with echinococcosis, in general, having an impact of 666,433 human disability-adjusted years per annum worldwide (Romig et al. 2006; Torgerson et al. 2010).

As noted, *E. multilocularis* has a Holarctic distribution (Davidson et al. 2016) and recent studies have shown that the extent of the geographic distribution of this parasite in Asia and Europe is increasing, and is increasing in prevalence while infecting both normal sylvatic hosts as well as humans (Schweiger et al. 2007; Takumi et al. 2008; Staubach et al. 2011; Torgerson 2013).

Echinococcus multilocularis was first detected in the Nearctic region in the 1950's in people living on St. Lawrence Island, Alaska and in the 1960's from foxes and other mammals in the northern great-plains (Rausch and Schiller 1954; Leiby and Olsen 1964). Since then, the species has been reported from 14 of the United States and four provinces in Canada, with records from new areas being reported constantly (Massolo et al. 2014). Nonetheless, this trend of expanding geographic distribution is difficult to interpret given the lack of systematic sampling and poor representation of sampled specimens in georeferenced collections in recognized museums. This lack of data results in uncertainty as to whether the geographic range of E. multilocularis is expanding, or detection of the parasite by parasitologists is improving because of more intense sampling efforts. However, despite the paucity of verified museumbased georeferenced records of this species, it appears that E. multilocularis does have a more extensive distribution than initially thought and new records are recorded herein. Lack of extensive and systematic specimen-based and georeferenced museum collections of E. multilocularis in the Nearctic can partly be offset using correlative ecological niche models (ENM's) which provide information covering factors influencing its distribution in ecological (environmental) space and the resulting geographical distribution of suitable areas for the species (Estrada-Peña et al. 2014). The aim of this paper is to provide a robust estimate of both the current and predicted future distributions of this extreme human pathogen in the Nearctic.

ENM's provide robust tools that can improve the understanding of the potential and actual distribution of species in question, as well as the way in which environmental factors may influence a species under study over space and time (Phillips *et al.* 2006; Searcy and Shaffer 2016). ENM's use occurrence records and Geographic Information Systems (GIS) layers of environmental information, to infer multifarious relationships between occurrence and the environment (*i. e.*, the environmental niche) and then project this relation over the study area. These models have been widely used in evolutionary, ecological, public health and conservation applications (Chetan *et al.* 2014; Ostfeld and Brunner 2015; Almasieh *et al.* 2016; Haverkost *et al.* 2010). Recent studies have shown the potential of this tool to model the distribution of suitable conditions for pathogenic parasites, their vectors and potential reservoirs (<u>Samy et al. 2014</u>; <u>Ragha-van et al. 2016</u>). Furthermore, these models can be used to predict the effect of drivers of global environmental change such as climatic or landcover alterations on the distribution of parasites or pathogens (<u>Stensgaard et al. 2013</u>). Despite their potential and widespread availability, ENM's have many caveats that researchers using these tools should take into account to produce robust inferences (<u>Kramer-Schadt et al. 2013</u>; <u>Radosavljevic and Anderson 2014</u>).

Traditionally, ENM's have employed abiotic variables as predictors of distribution of species with the assumption that biotic interactions influence species distributions at much finer spatial scales than the coarse-level geographic distributions evaluated and predicted with ENMs (Soberon 2007). Nonetheless, other authors have proposed and successfully used ways to include biotic interactions in niche and distribution models (Giannini et al. 2013; Anderson 2017). Given the dependence of parasites on their hosts, biotic information is expected to be particularly relevant for modeling the niche and resulting distribution of this group. The successful inclusion of biotic data might help clarify the realized niche of the species, providing more precision in the projected distribution over environmental and geographical space. For this reason, some authors have included information on host distributions for parasite ENM (Pickles et al. 2013; Stensgaard et al. 2013; Kuhn et al. 2016). As E. multilocularis depends entirely on direct trophic interactions among carnivores and rodents and can infect a range of species, we assume that host communitylevel variables might influence the distribution of this parasite. By including these kinds of variables in our models we will increase the multivariate nature of the ENM and this will result in a model that includes both abiotic and biotic dimensions, probably providing a more accurate representation of the species realized niche.

Our goal in this study is to contribute to the understanding of the environmental space occupied by *E. multilocularis* in the Nearctic and project the derived geographical distribution of suitable conditions for the species, especially in the central part of the continent. Here we review and georeference all records of the species in North America available to us to accomplish the following objectives: i) create a robust ENM with a best estimate of the current spatial distribution of suitable conditions. ii) Evaluate the abiotic and biotic factors influencing the species niche space and their relative importance. iii) Assess the potential impact of climatic change on the distribution of suitable conditions for *E. multilocularis* in the future (2070). As part of our first objective, we include two unpublished records of the parasite for the USA.

Materials and Methods

Parasite data. To create a database on the *E. multilocularis* sampling and detection localities, we conducted a systematic search in the Web of Science using the search terms *"Echinococcus multilocularis"* and "North America", "Canada"

and "USA" or other variations of the country names. We complemented these sources with other references cited in the recovered papers. We further filtered out the publications that did not include new records (i. e., reviews) and the ones without enough information to positively georeference the sampling locality with a precision of less than 20 km. This threshold was selected as a compromise between precision and quantity of occurrence records. From each article, we collected the sampling localities (including those where the species was not detected), geo-coordinates, georeferencing precision, host species, infection status, number of sampled individuals, number of infected individuals, parasite prevalence and year of collection. We also included in our database, the records from the parasite collection in the Harold W. Manter Laboratory of Parasitology (HWML), data from the former US National Parasite Collection, the Museum of Southwestern Biology collection and sampling localities from the notes and records of R. L. Rausch's work on *E. multilocularis* in Alaska. Because most literature sources documenting E. multilocularis did not include study-site or collection locality geo-coordinates, we used gazetteers and Google Earth to georeference the sampling sites and estimated the assigned coordinate precision following the protocol of Chapman and Wieczorek (2006). When the paper presented a map (i. e., Rausch and Richards 1971), the map was georeferenced using the best matching coordinate system, and the coordinates were extracted using point size and georeferencing mismatch as precision.

New records. To increase the robustness of the current work, we included two previously unpublished records that were provided by HWML collaborators. The first record came from Maryland and represents a specimen of larval E. multilocularis that was collected by Gregory E. Glass who is now in the Department of Geography and the Emerging Pathogens Institute at the University of Florida. This specimen was determined to be definitely E. multilocularis and was identified to the level of the species by one of the experts in the field at the time, Everett L. Schiller (unfortunately the specimens were lost when Schiller died). The second specimen was collected by John E. Ubelaker, identified by John E. Ubelaker and Scott L. Gardner and deposited in the HWML Parasite Collection. To identify this specimen, protoscolexes recovered from the abdominal cavity of a single individual of Peromyscus maniculatus were prepared following the methods of Gardner et al. (2013). Both large and small hooks of the parasite recovered from stained protoscolexes were digitally imaged and measured. The species was identified based on hook morphology (shape and structure of the hooks: compare Figure 1, A to C). We also include a multivariate discriminant analysis of morphometric data to explore the affinities of the collected parasites to other populations of E. multilocularis. To do this, measurements of both large and small hooks were taken from specimens of E. multilocularis extracted from the alveolar cysts recovered from the liver of Microtus oeconomus col-



Figure 1. Images of rostellar hooks taken from alveolar cysts removed from the liver of rodents from three separate collection localities in Mongolia and North America. A) Large and small hooks from protoscolexes taken from *Peromysus maniculatus* collected near Taos, New Mexico, USA in 2009. B) Hooks taken from *Microtus limnophilus* collected near Har Us Lake, Khovd, Mongolia, 2012. C) Hooks from *Microtus oeconomus* from St. Lawrence Island Alaska, USA in 1950. All images and measurements made with a Zeiss Axiophot microscope and Zeiss Axiovision Software (SE64 v. 4.8). All hooks given at same ten micrometer scale as shown in Figure 1B.

lected in 1958 from St. Lawrence Island (HWML14934) and from *Microtus limnophilus* collected from near Har Us Lake in western Mongolia (HWML68052; <u>Gardner et al. 2013</u>). Canonical discriminant function analysis was performed using SAS 9.4 following the general methods of <u>Gardner</u> and <u>Schmidt (1988)</u> and <u>Gardner et al. (2013)</u>.

Environmental and biotic variables. We used two sets of predictor variables to model the niche of the parasite and project habitat suitability over North America. The first set (abiotic variables) includes average daily solar radiation and 15 bioclimatic variables which summarize several aspects of temperature and precipitation intra annual tendencies and variability acquired from the WorldClim V.2. dataset (Fick et al. 2017; bios 8, 9, 18 and 19 were excluded as they might show artefacts resulting from their dependence on both precipitation and temperature - Campbell et al. 2015). The second set consists of four variables related to host species assemblage, including definitive and intermediate host species richness and the proportion of carnivore assemblage represented by potential definitive host-species as well as the proportion of rodent assemblage represented by the potential intermediate hosts. The hosts were selected based on the species with more than two records of E. multilocularis infection in our database (definitive hosts: Canis latrans, Canis lupus, Vulpes vulpes, and V. lagopus; intermediate hosts: Lemmus trimucronatus, Microtus oeconomus, M. pennsylvanicus, Mus musculus, Myodes gapperi, Ondatra zibethicus, and Peromyscus maniculatus). Rasters displaying

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these variables were created by counting the number of species from each group whose range overlap at each pixel using the IUCN distribution maps (<u>IUCN 2016</u>).

To reduce both the number of variables and complexity of the model, we performed a Pearson product moment correlation test among all pairs of variables extracted using 10,000 random points from the study area (Warren et al. 2014). To attempt to decrease the effect of correlated variables on the analysis, one variable from each pair with correlation coefficients higher than 0.8 was removed. Variables that have been shown to be more biologically important or meaningful were retained in the analysis. This approach resulted in six climate-related, and four host-related variables (Table 3). These variables were masked using the presumed accessible area for the parasite. The selection of the modeling area is a key part of the EN modeling process as it should represent the sites accessible to the species of interest over relevant time periods (Barve et al. 2011). In this study, given the high dispersal potential of definitive hosts of E. multilocularis (Trewhella et al. 1988; Harrison 1992). We defined this area as all regions encompassing the geographic range of at least one definitive and intermediate host, resulting in an inclusive area of North America from above the arctic circle to the northern portion of Mexico. All variables were resampled to a 2.5' resolution.

Distribution modeling. We used the Maximum Entropy algorithm as implemented in the software Maxent (Version 3.4.1; <u>Phillips et al. 2017</u>) to model the niche of *E. multilocularis*, assess the effect of abiotic and biotic variables over its distribution and project habitat suitability over current and future environmental conditions. We selected Maxent as it has been shown repeatedly to perform well when only presence data is available (<u>Elith et al. 2006; Phillips</u> and <u>Dudík 2008</u>). Maxent compares environmental values at species presence sites against a background sample of

Table 3. Importance metrics for each of the variables included in the biotic+abiotic model. Jackknife values are reported in performance decrease percentage assessed using test data. The three most important variables according to each metric appear in bold.

Variable	Permutation importance	Jackknife without variable (test gain)	Jackknife with only variable (test gain)
Average daily solar radiation (kJ m ⁻² day ⁻¹)	7.2	1.70	1.310
Precipitation of Wettest Month	0.6	1.73	0.430
Precipitation of Driest Month	0.2	1.74	0.256
Max Temperature of Warmest Month	9.4	1.67	0.876
Temperature Annual Range	1.0	1.70	0.360
Precipitation Seasonality	3.7	1.72	0.153
Proportion definitive host in carnivore community	22.0	1.57	0.797
Definitive host species richness	34.3	1.61	0.678
Intermediate host species richness	17.4	1.69	0.298
Proportion intermediate host in rodent community	4.3	1.69	0.467

the environment and estimates the species distributions by finding the distribution of maximum entropy subject to the constraint that the expected value for each environmental variable under this estimated distribution matches its empirical average (Elith *et al.* 2011).

Despite the good performance of Maxent, several aspects should be taken in to account for maximizing model reliance and robustness. Probably, the most important is the fact that presence records usually do not come from a random sampling of the study area and sometimes might be spatially clumped which might result in biased models, which in the worst cases are predicting the sampling conditions rather than suitability for the species that is being studied (Fourcade et al. 2014). To tackle this issue and given the fact that the data were not evenly distributed through geographic space, we took the following two steps: i) We created a sampling bias raster from which Maxent would extract the background points in the proportion of sample intensity. For this, we pulled all the georeferenced records for E. multilocularis sampling (positive and negatives) as well as georeferenced records of other parasite species collected from potential hosts and available in the parasite collections accessible through the Arctos biodiversity data repository. With these records, we created a kernel density raster using a sigma = 0.8 representing parasite sampling intensity. ii) We further proceed to randomly remove one record from the pairs that were closer than 30 km (distance based on the data precision and visual inspection of occurrence clusters) to reduce the clustering of the data points. We did this using the R package SPTHIN (Aiello-Lammens et al. 2015) to maximize the number of usable points of occurrence. Another issue with Maxent models is the probability of overfitting due to exceedingly complex models. To alleviate this issue, we used the R package ENMeval (Muscarella et al. 2014) to assess the performance of several combinations of regularization parameters (RP; 1, 2, 3, 5, and 6) and feature classes (linear, quadratic, product and hinge - threshold was not included to avoid overly complex models; Phillips et al. 2017), both factors related to the complexity included in model fitting. From these, we selected the best combination based on the partial ROC significance (see below), difference between training and test omission threshold (a measure of overfitting) and best AICc.

To assess the effect of including biotic variables on model performance, we created two separate niche models, one using the dataset of all variables described above and another model employing only the abiotic variables. For each dataset, we created 50 bootstrapped models using a randomly chosen 20 % of the records for its assessment in each replicate. The models were fitted using the regularization parameter and features selected in the abovementioned step. We compared both models employing three criteria: AICc; partial ROC AUC ratio and the average tests data omission rate of the 50 replicates using the 10 % training omission threshold. The AICc was estimated for each model using the ENMeval package. The partial ROC AUC ratio is proposed as a superior way of comparing models and calculates the ratio among the null expectation and observed area under the Receiving Operator Curve starting from a defined omission threshold, which in this case given the time span of the records was set to 5 % (Peterson *et al.* 2008). The significance of the ratios was assessed with 500 bootstrapped replicates using 50 % of the records employing the NicheToolBox (http://shiny.conabio.gob.mx:3838/ nichetoolb2/). The mean tests data omission rate is considered a measure of model overfitting. The differences in pAUC and omission test between the two models were evaluated using a t-test.

We used two methods to assess the importance of each variable in determining environmental suitability for E. multilocularis. First, we used the average percent premutation importance of each variable from each replicate. This metric assesses the decrease or increase in model performance when each variable is permuted. Second, we used the Jackknife test in which the model is run without each variable and with only each variable, and the differences in performance are calculated as the percentage decrease in AUC. This jackknife analysis indicates the variables that contain the most influential and the most unique information respectively. To describe the relationship between the important variables and the parasite distribution, we employ response curves that show how suitability change when the variable of interest is varied while keeping all other environmental variables at their average sample value. All of these analyses were carried on the MaxEnt software (Phillips et al. 2017).

Maxent main output is a continuous raster of the estimated suitability. For predicting the potential distribution of a species, it is desirable to find a threshold of suitability above which the species is predicted to persist given the species can disperse, which would produce binary maps. Nonetheless, there are several ways of defining the threshold, and overall predictions are highly dependent on them (Norris 2014). In this paper, we report predicted distribution using two thresholds over the median suitability of the 50 model replicas, the first is the one resulting in a 5 % omission rate of all records, a threshold that has been commonly used for models at this spatial/temporal scale and coming from diverse sources (Raes et al. 2009; Campbell et al. 2015; Sage et al. 2017), and which can be interpreted as including the most suitable habitat. The second predicted distribution is more inclusive and represents the threshold that equates entropy of distributions at the threshold and original areas. This threshold has been proposed for organisms with high dispersal potential as they can reach distant areas in a relatively short time period (Filz and Schmitt 2015: Hill et al. 2016). We interpret the resulting model as describing the subset of the realized niche represented in the study area (Peterson and Soberón 2012). In turn, this model relates the environmental and biotic variables to suitability for the species occurrence. When this suitability value is projected over geographic space and reclassified,

we get an estimation of the suitable area.

Finally, given the small difference in performance and prediction ability (see results), we used the abiotic variables model to project the distribution of suitable conditions for E. multilocularis in the year 2070. For this, we used the climatic projections from the 17 general circulation models (GCM) available at the WorldClim database under the contrasting 4.5 and 8.5 emission scenarios (RCPs) from the 5th IPCC. The first scenario corresponds with a peak in emissions around 2040 and further emissions decline with stabilization around 2100; the second, more pessimistic scenario presents emissions rising throughout the 21st century (http://www.ipcc.ch/). We estimated suitability for each GCM RCP combination and predicted gain and loss of suitable habitat for the median suitability using thresholds that maintain the predicted suitable area found by the best model. As a way of assessing uncertainty arising from different GCM, we report the amount of consensus over suitable habitat gain and loss for each scenario.

Results

New records of Echinococcus multilocularis *in the Nearctic*. We include here a new record of a specimen of *E. multilocularis* found in the liver of an individual of *M. pennsylvanicus* collected by Gregory E. Glass in a grassy area in Mount Pleasant Park, Baltimore, Maryland in August, 1985. The parasite cyst in the liver was identified as *E. multilocularis* by Everett L. Schiller (see Methods, above). This represents the first record of *E. multilocularis* for Maryland and the most eastern record for the species in North America.

Our second unpublished record included herein was from a georeferenced, museum specimen of *E. multilocularis* recently deposited in the Manter Laboratory Parasite Collection (Catalog no. HWML68737). The host, an individual of *P. maniculatus*, was collected by John E. Ubelaker 10 km SW of Taos, New Mexico on 28 June, 2009 in willows along Pot Creek. The parasite was identified by J. E. Ubelaker and Scott L. Gardner.

To help confirm the identification of the sample from New Mexico, we examined the extent of morphological variation among three populations of E. multilocularis. For this, we measured seven characters of the larval/rostellar hooks including (large hooks) total-length, handle-length, guard-width, and blade-length and the same characters for the small hooks. We ran the analysis using both the CANDISC and STEPDISC commands in SAS 9.4. The analysis showed that the characters in order of importance for discrimination among the three populations of E. multilocularis were: large hook guard width, large hook total length, small hook guard width, large hook handle length, and small hook total length (see Table 1). All eight characters served to assist in discrimination among the three populations of these cestodes but those mentioned above were more important. Finally, in this analysis, the canonical ordination in Figure 2 shows that CANI separates the Alaskan form of E. multilocularis from the other two populations (Taos, New Mexico and Har Us Lake) fairly well while CANII shows a fairly good separation of *E. multilocularis* from the specimens collected from near Taos and Har Us Lake, Mongolia (Figures 1, 2; Table 1).

Performance and comparisons of the Models. For the final models, 178 presence records of *E. multilocularis*, collected through the last 60 years were used (Figure 3). An RP = 5 and all assessed features were selected for the biotic model, while a RP = 2 and the linear, quadratic and prod-



Figure 2. Plot of discriminant scores derived from measurements of rostellar hooks taken from specimens of *Echinococcus multilocularis* collected from three different collecting localities. The solid red dots represent the specimens of *E. multilocularis* collected from *Peromyscus maniculatus* near Taos New Mexico, USA. The Blue boxes represent specimens of *E. multilocularis* collected from *Microtus limnophilus* near Har Us Lake, Khovd, Mongolia, and the solid black dots represent specimens measured of *E. multilocularis* collected from *Microtus consulta*. Alaska, USA.

uct features were used for the only abiotic variables model. The models performed very well, with average pROC AUC above 0.9. The model including the biotic data showed the lower AICc and the higher ROC AUC ratio (p-value = 2.2^{-16}), while there was no statistically significant difference in omission rate between the two models (Table 2). This suggests that the model including host community informa-

Table 1. Character loadings and variation in canonical structure for 3 populations of *E. multilocularis* from *Peromyscus maniculatus* from Taos, New Mexico, USA; *Microtus oeconomus* from St. Lawrence Island, Alaska, USA; and *Microtus limnophilus* from Har Us Lake, Mongolia. Numbers in bold represent loading coefficients with greatest discriminatory power in the canonical analysis. For example, the character "Guard width" for the large hooks is most important with a loading on CAN I of 0.91.

Total Canonical Structure					
Variable		CANI	CAN II		
Large Hooks	Total length	0.438	0.802		
	Handle length	0.196	0.693		
	Guard width	0.914	0.049		
	Blade length	0.382	0.402		
	Total length	0.036	0.619		
Small Hooks	Handel length	-0.262	0.486		
	Guard width	0.762	0.270		
	Blade length	0.341	0.355		
Percentage of variation of each canonical axis		70.52 %	29.48 %		
Canonical Correlation		87.36 %	75.76 %		



Figure 3. Results of the Maxent distribution model of *Echinococcus multilocularis*. A) Median continuous environmental suitability index on a log scale resulting from the 50 replicas; warmer colors indicating higher suitability. B) Binary prediction of suitable environmental conditions using two different reclassification thresholds. The dark gray distribution represents the threshold that keeps 95 % of the records, while the light gray area represents the suitable area using a threshold that equates entropy of threshold and original distributions. Occurrence location data used in the model are displayed by date of collection.

tion is significantly more robust than the one including only abiotic variables. As the omission did not show differences between the models, the gain in performance of the biotic models appears to come from more restricted predictions, thus reducing overprediction error.

Distribution model. For the current distribution, we present the results from the abiotic+biotic model. Applying the more stringent threshold of a 5 % omission, we find that approximately 12 % of North American land mass appears as highly suitable for *E. multilocularis*. This results in a predicted suitable area of 2.6 million km² distributed mainly in a band from the east coast of North America to the east of the Rocky Mountains (Figure 3). The predicted suitable area encompasses the great lakes, the central great-plains from Minnesota to Alberta, the Appalachian range and a large portion of Quebec in Canada. Besides this continuous area, some smaller suitable sites are predicted through the continent, including St. Lawrence and St. George islands in Alaska as well as large areas of the state of Washington and isolated patches in British Columbia (Figure 3).

With the lower threshold, 36 % (7.4 million km²) of the study area is predicted as suitable for the parasite. This includes a large portion of NA, including the east coast from South Carolina to Newfoundland and Labrador and towards the west including Alberta, and a large area of the great plains as well as eastern and boreal forest and isolated areas of the Rocky Mountains south to New Mexico (Fig-

Table 2.Summary of performance metrics for models including both biotic andabiotic variables versus a model including only abiotic variables. Mean omission test rateis under a 10 % training omission threshold.

** indicates significant differences among the two models.

Model	AICc	pROC AUC Ratio**	Mean Omission Test Rate
Abiotic Only	4,808	1.81	10.46 %
Abiotic + Host variables	4,689	1.94	12.05 %

ure 3). Also, with this threshold, a strip of patches of suitable habitat is predicted to the northwest of Alberta up to Alaska. Some other disjunct areas in the north also appear suitable, with the largest being Baffin and Southampton islands in the Arctic. Under this threshold, the model also predicts two smaller isolated, suitable areas in Louisiana and the border between Texas and Oklahoma.

Variable importance. According to the permutation importance test, definitive host species richness is the most significant variable (34 % importance), followed by proportion of potential hosts in carnivore community (22 %) and intermediate host species richness (17 %). The jackknife test, using model gain on test data shows that the most influential variables are average daily solar radiation, maximum temperature of the warmest month, and proportion of potential hosts in carnivore community. This analysis also indicates that the variables containing unique information include again definitive host species richness, followed by proportion definitive host in carnivore community and maximum temperature of the warmest month (Table 2). When individual response curves are analyzed leaving the other variables at their average, definitive host species richness and proportion of potential intermediate hosts in rodent assemblages show a negative relationship with suitability, while intermediate host richness, proportion of potential hosts in carnivore communities, and maximum temperature of the warmest month show a positive relationship. This contrasts to solar radiation that shows an intermediate peak in suitability. In line with the permutation importance and jackknife results, the other variables don't show very strong effects on suitability (Figure 4).

Projected distribution. Projected change in median suitability values for all GCMs up to the year 2070 under 4.5 and 8.5 scenarios show a large expected increase in suitable habitat area. Using the lower threshold, the areas of predicted distribution are shown to increase by 56 % and 76 % under 4.5 and 8.5 scenarios respectively (Figure 5). Using this threshold, the distribution shift pattern is very similar for both scenarios with suitable area predicted to increase mainly north of the current suitable area in North America. Some areas are predicted to become unsuitable for the species in northwestern Canada and Alaska (accounting for less than 5 % of the original area in both scenarios). The uncertainty analysis shows that projections under different climate circulation models agree to a large extent in that suitable conditions for the parasite presence will appear up to Hudson Bay, Labrador as well



Figure 4. Response curves of *Echinococcus multilocularis* suitability relative to most influential variables used in this study. Curves show how predicted suitability changes as each variable is changed, keeping all other environmental variables at their average sample value. BIO5 = Max Temperature of Warmest Month; BIO7 = Temperature Annual Range; BIO13 = Precipitation of Wettest Month; BIO14 = Precipitation of Driest Month; BIO15 = Precipitation Seasonality (Coefficient of Variation); SRAD= average daily solar radiation; S_int_host = Intermediate host species richness; S_def_host = Definitive host species that are potential intermediate hosts; P_def_host = Proportion of carnivore species that are potential definitive hosts.

as west of the Rocky Mountains and in Alaska (Figure 5). The models show less consensus about the expected change in suitability for the taiga ecosystem in the north of the continent.

Discussion

To our knowledge, this is the first ecological niche model exercise that has been fully developed and implemented for *E. multilocularis* in the Nearctic region. The model presented here can be instrumental in efforts to both predict and to monitor the parasite as it continues to expand and perhaps make the jump into humans, domestic animals, and wildlife as it shows geographic areas of both suitable and highly suitable habitats for the species. The partial AUC as well as the average test omission rate indicates the model appears to have very good performance. This model also highlights areas where the parasite is predicted to be pres-

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ent, but so far there is no known record of it. These include eastern North America, especially the northern Appalachians, where highly suitable conditions are expected to occur. Labrador and Anticosti Island also appear as highly suitable for the parasite as well as the isolated patches in the state of Washington. Under the smaller threshold, an important area of North America appears as suitable, which deserves further investigation including a portion of Texas, and Louisiana, areas at this time that are far from the known distribution of the parasite.

This work also provides insights into the variables influencing E. multilocularis distribution, with a novel approach that included biotic variables from host community macroecological data. We found that by including host species richness and proportion of rodent and carnivore communities composed by potential host results in a significantly better model with reduced over-prediction at the time that average omission error does not change. The importance of including biotic variables in these models is further supported by the permutation variable importance analysis as well as the jackknife test that show these variables contribute significant and unique data to the species distribution model. This community data might be a proxy of potential trophic interactions among hosts and might even contain information on dilution effects as host species richness was negatively related to suitability whereas the proportion of host composing the carnivore community was positively related (Civitello et al. 2015). Nonetheless, more field and experimental data should be gathered to fully understand the mechanistic aspects behind this correlative model results.

Besides biotic variables, abiotic factors also had a relatively great impact in predicting the distribution of the parasite over geographic space. The primary variable with the greatest effect on the model was solar radiation, with an intermediate peak in suitability over all replicate models (Figure 4). This might be related to the effect of solar ultraviolet radiation on egg viability of taeniid cestodes (Veit *et al.* 1995); in addition, maximum temperature of the warmest month also appears as important, probably reflecting the effect of temperature relative to egg viability. Nonetheless, further research is needed to understand the mechanisms behind these correlations.

Despite excellent performance metrics, the model fails at identifying suitable areas in the Arctic where the parasite has been recorded such as Brooks Range, Banks Island, and Point Barrow. However, despite this, St. Lawrence and St. George islands, where the species has been recorded in very high prevalence, are predicted as highly suitable. We speculate that this under-prediction might come from two nonexclusive factors. The first is that as the distribution of the species is negatively influenced by radiation below a certain threshold and positively by maximum temperature, it might be that as suggested by <u>Haverkost *et al.* (2010)</u> for the tapeworm *Paranoplocephala macrocephala*, most of the life cycle of the parasite in the Arctic is subnivian and occurs



Figure 5. Projected changes in geographic distribution of suitable environmental conditions for *Echinococcus multilocularis* under the 4.5 (left column) and 8.5 (right column) global emission scenarios (RCP) for 2070. A) Changes in suitable habitat under the lower threshold for the median suitability of all GCM predictions under the 4.5 RCP. B) Changes in suitable habitat under the lower threshold for the median suitability of all GCM predictions under the 4.5 RCP. C) Consensus of predicted suitable habitat gain under the 4.5 RCP for all 17 GCM. Colors indicate the number of models that predict gain in each pixel. D) The same as C for the 8.5 RCP scenario. E) Consensus of predicted loss in suitable habitat under the 4.5 RCP for all 17 GCM. F) The same as E for the 8.5 RCP scenario.

in burrows under the snow were microclimatic conditions are not possible to model at continental scales. The other factor might be the fact that Arctic and central Nearctic populations of *E. multilocularis* represent at least two different genotypes (Nakao *et al.* 2009) or as some evidence suggests, the central US forms of *E. multilocularis* might actually represent another species, as is suggested by the discovery of several more genotypes (Gesy *et al.* 2014). Some support of this is shown in Figure 2 with a cluster of EM from near Taos, NM, showing distinct separation from other populations in our canonical plots. Thus, it is possible that the Arctic and central North American genotypes of these cestodes possess different niches and given the data quantity differences contrasting both areas, the present model is a representation of the central Nearctic genotypes. Under the two global carbon emission scenarios evaluated here, the extension of suitable habitat for *E. multilocularis* is projected to increase. Under the higher threshold, the projected change follows that of other pathogens in gaining suitable areas in the north, a pattern shown by other Nearctic parasites (Brownstein *et al.* 2005). Nonetheless, this pattern is not general and different parasite species are predicted to show unique responses including range contractions (Sage *et al.* 2017). For example, in the case of *E. multilocularis*, some northern areas are predicted to become unsuitable. Although we tried to account for the uncertainties involved in climate model predictions, these projections should be interpreted with care as there are certain untested assumption as well as uncertainties in them. One assumption is that the parasite niche will not evolve, an assumption that is difficult to support, given the potential niche divergence between strains shown here as well as the host-switching potential via ecological fitting of the parasite (Brooks *et al.* 2014; Brooks and Hoberg 2007). Another potential problem of the projections is that they do not include a deep set of host-community data, which will almost certainly show rearrangements due to climate change (Pandey and Papes 2018). Despite this, we think these models show the general trend that probably will exhibit some variation resulting from host and evolution complexities.

Here we show that by combining well-documented museum records, morphometric data, and ENM's we can improve model predictivity and provide extra data that use distribution models for enhancing knowledge on the distribution and ecology of zoonotic diseases. With our approach, we offer a robust hypothesis of the distribution of suitable habitats for E. multilocularis in the Nearctic. This approach to modeling the distribution of zoonotic parasites can be used by biodiversitists, parasitologists, and healthcare practitioners to assess both infections of zoonotic parasites in wildlife as well as in potentially vulnerable human communities in parasite-suitable or highly suitable areas. Besides this, we identified the factors shaping the parasite distribution and suggest a new set of variables coming from host geographic distribution that could be used in modeling of other parasites that have complex life cycles and that can only survive by infecting multiple hosts as Manter (1966) so eloquently stated:

These same parasites also hold promise of telling us something about host and geographical connections of long ago. They are simultaneously the product of an immediate environment and of a long ancestry reflecting associations of millions of years. The messages they carry are thus always bilingual and usually garbled. Today we know only a few selected pieces of the code. As our knowledge grows, studies based on adequate collections, correctly classified and correlated with knowledge of the hosts and life cycles involved should lead to a deciphering of the messages now so obscure. Eventually there may be enough pieces to form a meaningful language which could be called **parascript** - the language of parasites which tells of themselves and their hosts both of today and yesteryear.

With this contribution, we show that application of **both** abiotic and biotic variables in ecological niche models (of *E. multilocularis*) can expand on Manter's idea of Parascript in historical terms and predict where the parasites might be in the future.

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Literature cited

- AIELLO-LAMMENS, M. E., R. A. BORIA, A. RADOSAVLJEVIC, B. VILELA, AND R. P. ANDERSON. 2015. spThin: an R package for spatial thinning of species occurrence records for use in ecological niche models. Ecography 38:541–545.
- ALMASIEH, K., M. KABOLI, AND P. BEIER. 2016. Identifying habitat cores and corridors for the Iranian black bear in Iran. Ursus 27:18–30.
- ANDERSON, R. P. 2017. When and how should biotic interactions be considered in models of species niches and distributions? Journal of Biogeography 44:8–17.
- Barve, N., V. Barve, A. JIMÉNEZ-VALVERDE, A. LIRA-NORIEGA, S. P. MAHER, A. T. PETERSON, J. SOBERÓN, AND F. VILLALOBOS. 2011. The crucial role of the accessible area in ecological niche modeling and species distribution modeling. Ecological Modelling 222:1810–1819.
- BROOKS, D. R., AND D. A. MCLENNAN. 1993. Parascript: Parasites and the language of evolution. Smithsonian Institution Press, Washington D.C.
- BROOKS, D. R., AND W. A. BOEGER. 2019. Climate change and emerging infectious diseases: Evolutionary complexity in action. Current Opinions in Systems Biology 13:75-81.
- BROOKS, D. R., E. P. HOBERG, W. A. BOEGER, S. L. GARDNER, K. E. GALBREATH,
 D. HERCZEG, H. H. MEJÍA-MADRID, S. E. RÁCZ, AND A. T. DURSAHINHAN.
 2014. Finding them before they find us: Informatics, parasites, and environments in accelerating climate change.
 Comparative Parasitology 81:155–164.
- BROOKS, D. R., AND E. P. HOBERG. 2007. How will global climate change affect parasite host assemblages? Trends in Parasitology 23:27–30.
- BROWNSTEIN, J. S., T. R. HOLFORD, AND D. FISH. 2005. Effect of Climate Change on Lyme Disease Risk in North America. EcoHealth 2:38–46.
- BUDKE, C. M., A. CASULLI, P. KERN, AND D. A. VUITTON. 2017. Cystic and alveolar echinococcosis: Successes and continuing challenges. PLOS Neglected Tropical Diseases 11:e0005477.
- CAMPBELL, L. P., C. LUTHER, D. MOO-LLANES, J. M. RAMSEY, R. DANIS-LOZANO, AND A. T. PETERSON. 2015. Climate change influences on global distributions of dengue and chikungunya virus vectors. Philosophical Transactions of the Royal Society: Biological Sciences 370:20140135–20140135.
- CHAPMAN, A. D., AND J. WIECZOREK (EDS.). 2006. Guide to Best Practices for Georeferencing. Copenhagen: Global Biodiversity Information Facility.
- CHETAN, N., K. K. PRAVEEN, AND G. K. VASUDEVA. 2014. Delineating ecological boundaries of hanuman langur species complex in peninsular India using MaxEnt modeling approach. PLoS ONE 9:1–11.
- CIVITELLO, D. J., J. COHEN, H. FATIMA, N. T. HALSTEAD, J. LIRIANO, T. A. MCMAHON, C. N. ORTEGA, E. L. SAUER, T. SEHGAL, S. YOUNG, AND J. R. ROHR. 2015. Biodiversity inhibits parasites: Broad evidence

for the dilution effect. Proceedings of the National Academy of Sciences 112:8667–8671.

- DA SILVA, A. M. 2010. Human echinococcosis: A neglected disease. Gastroenterology Research and Practice 2010:583297.
- Davidson, R. K., A. Lavikainen, S. Konyaev, J. Schurer, A. L. Miller, A. Oksanen, K. Skirnisson, and E. Jenkins. 2016. *Echinococcus* across the north: Current knowledge, future challenges. Food and Waterborne Parasitology 4:39–53.
- ELITH, J., C. H. GRAHAM, R. P. ANDERSON, M. DUDÍK, S. FERRIER, A. GUISAN, R. J. HIJMANS, F. HUETTMANN, J. R. LEATHWICK, A. LEHMANN, J. LI, L. G. LOHMANN, B. A. LOISELLE, G. MANION, C. MORITZ, M. NAKAMURA, Y. NAKAZAWA, J. M. M. OVERTON, A. T. PETERSON, S. J. PHILLIPS, K. RICHARDSON, R. SCACHETTI-PEREIRA, R. E. SCHAPIRE, J. SOBERÓN, S. WILLIAMS, M. S. WISZ, AND N. E. ZIMMERMANN. 2006. Novel methods improve prediction of species' distributions from occurrence data. Ecography 29:129–151.
- ELITH, J., S. J. PHILLIPS, T. HASTIE, M. DUDÍK, Y. E. CHEE, AND C. J. YATES. 2011. A statistical explanation of MaxEnt for ecologists. Diversity and Distributions 17:43–57.
- ESCOBAR, L. E., AND M. E. CRAFT. 2016. Advances and limitations of disease biogeography using ecological niche modeling. Frontiers in Microbiology 7:1–21.
- ESTRADA-PEÑA, A., R. S. OSTFELD, A. T. PETERSON, R. POULIN, AND J. DE LA FUENTE. 2014. Effects of environmental change on zoonotic disease risk: an ecological primer. Trends in Parasitology 30:205–14.
- FICK, S. E., AND HIJMANS, R. J. 2017. WorldClim 2: new 1-km spatial resolution climate surfaces for global land areas. International Journal of Climatology 37:4302–4315.
- FILZ, K. J., AND T. SCHMITT. 2015. Niche overlap and host specificity in parasitic *Maculinea* butterflies (Lepidoptera: Lycaenidae) as a measure for potential extinction risks under climate change. Organisms Diversity and Evolution 15:555–565.
- FOURCADE, Y., J. O. ENGLER, D. RÖDDER, AND J. SECONDI. 2014. Mapping species distributions with MAXENT using a geographically biased sample of presence data: A performance assessment of methods for correcting sampling bias. PLoS ONE 9:1–13.
- GARDNER, S. L., A. T. DURSAHINHAN, G. R. RÁCZ, N. BATSAIKHAN, AND S. GANZORIG. 2013. Sylvatic species of *Echinococcus* from rodent intermediate hosts in Asia and South America. Occasional Papers, Museum of Texas Tech University 318:1–13.
- GARDNER, S. L., AND G. D. SCHMIDT. 1988. Cestodes of the genus *Hymenolepis* Weinland, 1858 *sensu stricto* from pocket gophers *Geomys* and *Thomomys* spp. [Rodentia: Geomyidae] in Colorado and Oregon, with a discriminant analysis of four species of *Hymenolepis*. Canadian Journal of Zoology 66: 896-903.
- GESY, K. M., J. M. SCHURER, A. MASSOLO, S. LICCIOLI, B. T. ELKIN, R. ALISAUSKAS, AND E. J. JENKINS. 2014. Unexpected diversity of the cestode *Echinococcus multilocularis* in wildlife in Canada. International Journal for Parasitology: Parasites and Wildlife 3:81–87.
- GIANNINI, T. C., D. S. CHAPMAN, A. M. SARAIVA, I. ALVES-DOS-SANTOS, AND J. C. BIESMEIJER. 2013. Improving species distribution models using biotic interactions: a case study of parasites, pollinators and plants. Ecography 36:649–656.
- HARRISON, D. J. 1992. Dispersal Characteristics of Juvenile Coyotes in Maine. The Journal of Wildlife Management 56:128–138.
- HAVERKOST, T. R., S. L. GARDNER, AND A. T. PETERSON. 2010. Predicting the distribution of a parasite using the ecological niche

model, GARP Predicción de la distribución de un parásito usando el modelo de nicho ecológico, GARP. Revista Mexicana de Biodiversidad 81:895–902.

- HILL, N. J., A. J. TOBIN, A. E. RESIDE, J. G. PEPPERELL, AND T. C. L. BRIDGE. 2016. Dynamic habitat suitability modelling reveals rapid poleward distribution shift in a mobile apex predator. Global Change Biology 22:1086–1096.
- IUCN. 2016. The IUCN Red List of Threatened Species. Version 2016-1.
- KERN, P., A. M. DA SILVA, O. AKHAN, B. MÜLLHAUPT, K. A. VIZCAYCHIPI, C. BUDKE, AND D. A. VUITTON. 2017. The echinococcoses: diagnosis, clinical management and burden of disease. Advances in parasitology 96:259-369.
- KRAMER-SCHADT, S., J. NIEDBALLA, J. D. PILGRIM, B. SCHRÖDER, J. LINDENBORN, V. REINFELDER, M. STILLFRIED, I. HECKMANN, A. K. SCHARF, D. M. AUGERI, S. M. CHEYNE, A. J. HEARN, J. ROSS, D. W. MACDONALD, J. MATHAI, J. EATON, A. J. MARSHALL, G. SEMIADI, R. RUSTAM, H. BERNARD, R. ALFRED, H. SAMEJIMA, J. W. DUCKWORTH, C. BREITENMOSER-WUERSTEN, J. L. BELANT, H. HOFER, AND A. WILTING. 2013. The importance of correcting for sampling bias in MaxEnt species distribution models. Diversity and Distributions 19:1366–1379.
- KRITICOS, D. J., V. JAROŠIK, AND N. OTA. 2014. Extending the suite of bioclim variables: a proposed registry system and case study using principal components analysis. Methods in Ecology and Evolution 5:956–960.
- KUHN, T., S. CUNZE, J. KOCHMANN, AND S. KLIMPEL. 2016. Environmental variables and definitive host distribution: a habitat suitability modelling for endohelminth parasites in the marine realm. Scientific Reports 6:30246.
- Lass, A., B. Szostakowska, P. MYJAK, AND K. KORZENIEWSKI. 2015. The first detection of *Echinococcus multilocularis* DNA in environmental fruit, vegetable, and mushroom samples using nested PCR. Parasitology Research 114:4023–4029.
- LEIBY, P. D., AND D. C. KRITSKY. 1972. *Echinococcus multilocularis*: a possible domestic life cycle in central North America and its public health implications. Journal of Parasitology 58:1213–1215.
- LEIBY, P. D., AND O. W. OLSEN. 1964. The cestode *Echinococcus multilocularis* in foxes in North Dakota. Science 145:1066.
- MANTER, H. W. 1966. Parasites of fishes as biological indicators of recent and ancient conditions. Pp. 59-71 *in* Host-Parasite Relationships. (McCauley, J. E. Ed.). Oregon State University Press, Corvallis, Oregon, USA.
- MASSOLO, A., S. LICCIOLI, C. BUDKE, AND C. KLEIN. 2014. *Echinococcus multilocularis* in North America: the great unknown. Parasite 21:73.
- MUSCARELLA, R., P. J. GALANTE, M. SOLEY-GUARDIA, R. A. BORIA, J. M. KASS, M. URIARTE, AND R. P. ANDERSON. 2014. ENMeval: An R package for conducting spatially independent evaluations and estimating optimal model complexity for Maxent ecological niche models. Methods in Ecology and Evolution 5:1198–1205.
- Nakao, M., N. Xiao, M. Okamoto, T. Yanagida, Y. Sako, and A. Ito. 2009. Geographic pattern of genetic variation in the fox tapeworm *Echinococcus multilocularis*. Parasitology International 58:384–389.
- NORRIS, D. 2014. Model thresholds are more important than presence location type: Understanding the distribution of lowland tapir (*Tapirus terrestris*) in a continuous Atlantic forest of southeast Brazil. Tropical Conservation Science 7:529–547.

- NYLIN, S., S. Agosta, S. Bensch, W. A. Boeger, M. P. Braga, D. R. Brooks, M. L. Forister, P. A. Hambäck, E. P. Hoberg, T. Nyman, A. Schäpers, A. L. Stigall, C. W. Wheat, M. Österling, and N. Janz. 2018. Embracing Colonizations: A New Paradigm for Species Association Dynamics. Trends in Ecology and Evolution 33:4–14.
- OSTFELD, R. S., AND J. L. BRUNNER. 2015. Climate change and *lxodes* tick-borne diseases of humans. Philosophical Transactions of the Royal Society of London. Series B, Biological sciences 370:20140051.
- PANDEY, R., AND M. PAPES. 2018. Changes in future potential distributions of apex predator and mesopredator mammals in North America. Regional Environmental Change 18:1223–1233.
- PETERSON, A. T., M. PAPES, AND J. SOBERÓN. 2008. Rethinking receiver operating characteristic analysis applications in ecological niche modeling. Ecological Modelling 213:63–72.
- PETERSON, A. T., AND J. SOBERÓN. 2012. Species distribution modeling and ecological niche modeling: getting the concepts right. Natureza and Conservação, 10:102–107.
- PHILLIPS, S. J., R. P. ANDERSON, AND R. E. SCHAPIRE. 2006. Maximum entropy modeling of species geographic distributions. Ecological Modelling 190:231–259.
- PHILLIPS, S. J., AND M. DUDIK. 2008. Modeling of species distributions with Maxent: new extensions and a comprehensive evaluation. Ecography 31:161–175.
- PHILLIPS, S. J., M. DUDIK, AND R. E. SCHAPIRE. 2017. [Internet] Maxent software for modeling species niches and distributions (Version 3.4.1). <I: http://biodiversityinformatics.amnh.org/open_source/maxent/> (June 11, 2018).
- PICKLES, R. S. A., D. THORNTON, R. FELDMAN, A. MARQUES, AND D. L. MURRAY. 2013. Predicting shifts in parasite distribution with climate change: A multitrophic level approach. Global Change Biology 19:2645–2654.
- RADOSAVLJEVIC, A., AND R. P. ANDERSON. 2014. Making better Maxent models of species distributions: Complexity, overfitting and evaluation. Journal of Biogeography 41:629–643.
- RAES, N., M. C. ROOS, J. W. F. SLIK, E. E. VAN LOON, AND H. TER STEEGE. 2009. Botanical richness and endemicity patterns of Borneo derived from species distribution models. Ecography 32:180–192.
- RAGHAVAN, R. K., D. G. GOODIN, G. A. HANZLICEK, G. ZOLNERWICH, M. W. DRYDEN, G. A. ANDERSON, AND R. R. GANTA. 2016. Maximum Entropy-Based Ecological Niche Model and Bio-Climatic Determinants of Lone Star Tick (*Amblyomma americanum*) Niche. Vector-Borne and Zoonotic Diseases X:vbz.2015.1837.
- RAUSCH, R. L., AND S. H. RICHARDS. 1971. Observations on parasitehost relationships of *Echinococcus multilocularis* Leuckart, 1863, in North Dakota. Canadian Journal of Zoology 49:1317–1330.
- RAUSCH, R. L., AND E. L. SCHILLER. 1954. Studies on the helminth fauna of Alaska. XXIV. *Echinococcus sibiricensis* n. sp., from St. Lawrence Island. Journal of Parasitol 40:659–662.
- Romig, T., D. Thoma, AND A. K. Weible. 2006. *Echinococcus multilocularis--*а zoonosis of anthropogenic environments? Journal of Helminthology 80:207–212.
- SAGE, K. M., T. L. JOHNSON, M. B. TEGLAS, N. C. NIETO, AND T. G. SCHWAN. 2017. Ecological niche modeling and distribution of *Ornithodoros hermsi* associated with tick-borne relapsing fever in western North America. PLoS Neglected Tropical Diseases 11.

- SAMY, A. M., L. P. CAMPBELL, AND A. TOWNSEND-PETERSON. 2014. Leishmaniasis transmission: Distribution and coarse-resolution ecology of two vectors and two parasites in Egypt. Revista da Sociedade Brasileira de Medicina Tropical 47:57–62.
- Schurer, J. M., K. M. Gesy, B. T. Elkin, and E. J. Jenkins. 2014. *Echinococcus multilocularis* and *Echinococcus canadensis* in wolves from western Canada. Parasitology 141:159–163.
- Schweiger, A., R. W. Ammann, D. Candinas, P. Clavien, J. Eckert, B. Gottstein, N. Halkic, B. Muellhaupt, B. M. Prinz, J. Reichen, P. E. Tarr, P. E. Torgerson, and P. Deplazes. 2007. Human alveolar echinococcosis after fox population increase, Switzerland. Emerging Infectious Diseases 13:878–882.
- SEARCY, C. A., AND H. B. SHAFFER. 2016. Do ecological niche models accurately identify climatic determinants of species ranges? American Naturalist 187:423–435.
- SEHGAL, R. N. M., W. BUERMANN, R. J. HARRIGAN, C. BONNEAUD, C. LOISEAU, A. CHSER, I. SEPIL, G. VALKIŪNAS, T. IEZHOVA, S. SAATCHI, AND T. B. SMITH. 2011. Spatially explicit predictions of blood parasites in a widely distributed African rainforest bird. Proceedings of the Royal Society B: Biological Sciences 278:1025–1033.
- SOBERON, J. 2007. Grinnellian and Eltonian niches and grographic distribution of species. Ecology Letters 10:1115–1123.
- STAUBACH, C., L. HOFFMANN, V. J. SCHMID, M. ZILLER, K. TACKMANN, AND F. J. CONRATHS. 2011. Bayesian space-time analysis of *Echinococcus multilocularis*-infections in foxes. Veterinary Parasitology 179:77–83.
- STENSGAARD, A. S., J. UTZINGER, P. VOUNATSOU, E. HÜRLIMANN, N. SCHUR, C. F. SAARNAK, C. SIMOONGA, P. MUBITA, N. B. KABATEREINE, L. A. TCHUEM TCHUENTÉ, C. RAHBEK, AND T. K. KRISTENSEN. 2013. Large-scale determinants of intestinal schistosomiasis and intermediate host snail distribution across Africa: Does climate matter? Acta Tropica 128:378–390.
- TAKUMI, K., A. DE VRIES, M. L. CHU, J. MULDER, P. TEUNIS, AND J. VAN DER GIESSEN. 2008. Evidence for an increasing presence of *Echinococcus multilocularis* in foxes in the Netherlands. International Journal for Parasitology 38:571–578.
- TORGERSON, P. R. 2013. The emergence of echinococcosis in central Asia. Parasitology 140:1667–1673.
- Torgerson, P. R., K. Keller, M. Magnotta, and N. Ragland. 2010. The global burden of alveolar echinococcosis. PLoS Neglected Tropical Diseases 4.
- TREWHELLA, W. J., S. HARRIS, F. E. AND MCALLISTER. 1988. Dispersal distance, home-range size and population density in the Red Fox (*Vulpes vulpes*): A Quantitative Analysis. The Journal of Applied Ecology 25:423–434.
- TUFTS, D. M., N. BATSAIKHAN, A. T. DURSAHINHAN, AND S. L. GARDNER. 2016. Identification of *Taenia* metacestodes from Mongolian mammals using multivariate morphometrics of the rostellar hooks. Erforschung Biologischer Ressourcen Der Mongolei (Halle/Saale) 13:361-375.
- VEIT, P., B. BILGER, V. SCHAD, J. SCHÄFER, W. FRANK, AND R. LUCIUS. 1995. Influence of environmental factors on the infectivity of *Echinococcus multilocularis* eggs. Parasitology 110:79–86.
- WARREN, D. L., WRIGHT, A. N., SEIFERT, S. N., AND H. B SHAFFER. 2014. Incorporating model complexity and spatial sampling bias into ecological niche models of climate change risks faced by 90 California vertebrate species of concern. Diversity and distributions 20:334–343.

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