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Developmental parameters of a southern mountain pine beetle population reveal potential source of latitudinal differences in generation time

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Title: Developmental parameters of a southern mountain pine beetle population reveal potential source of latitudinal differences in generation time

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1 Abstract

2 Mountain pine beetle (*Dendroctonus ponderosae*, Hopkins) is a major disturbance agent
3 in pine ecosystems of western North America. Adaptation to local climates has resulted in
4 primarily univoltine generation time across a thermally diverse latitudinal gradient. We
5 hypothesized that voltinism patterns have been shaped by selection for slower developmental
6 rates in southern populations inhabiting warmer climates. To investigate traits responsible for
7 latitudinal differences we measured lifestage-specific development of southern mountain pine
8 beetle eggs, larvae and pupae across a range of temperatures. Developmental rate curves were fit
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11 optimal development of southern individuals occurred at higher temperatures, with higher
12 development thresholds, as compared with northern individuals. Observed developmental rates
13 of the southern and northern populations were similar across studied lifestages at 20 °C, and
14 southern lifestages were generally faster at temperature extremes (10, 27 °C). At 25 °C southern
15 fourth instars were significantly slower than northern fourth instars. Our results suggest that
16 evolved traits in the fourth instar and remaining unstudied lifestage, teneral (i.e., pre-emergent)
17 adult, likely influence latitudinal differences in mountain pine beetle generation time.

18

19 **Keywords:** *Dendroctonus ponderosae*, development rate, mountain pine beetle, phenology,
20 latitudinal gradient

21

22 **1. Introduction**

23 As poikilotherms, insect development rates and thresholds are temperature-dependent
24 (Taylor 1981). In seasonal environments evolved adaptations in these and related physiological
25 traits, including diapause and quiescence, serve to synchronize developmental timing with local
26 climates (Tauber and Tauber 1976; Danks 1987), and to enhance mate-finding and host plant
27 feeding (Forrest and James 2011; Li *et al.* 2011). For species that inhabit highly seasonal and
28 cold environments, these strategies can also reduce the probability that lifestages vulnerable to
29 cold-induced mortality are present during winter. Due to their significant influence on population
30 success, thermally-dependent fitness traits commonly vary with environmental conditions along
31 latitudinal gradients, particularly in species with extensive distributions (Deutsch *et al.* 2008).
32 Understanding intraspecific trait variability is key to predicting population responses in a
33 changing climate.

34 Mountain pine beetle (*Dendroctonus ponderosae*, Hopkins, Coleoptera: Curculionidae,
35 Scolytinae) is a bark beetle native to mountainous areas of western North America with an
36 expansive distribution that ranges from Baja Norte, Mexico to northern British Columbia and
37 Alberta (Dowle *et al.* 2017; Cooke and Carroll, 2017). Mountain pine beetle feeds on, reproduces
38 in, and, when populations are at outbreak levels, kills pine (*Pinus*) trees. Mountain pine beetle
39 was responsible for 5.2 Mha of pine mortality in the western United States (US) between 1997
40 and 2012 (Hicke *et al.* 2016). In addition to the availability of suitable host trees, weather that
41 supports appropriate seasonal timing and synchronous adult emergence are essential for
42 population outbreaks (Logan and Bentz 1999; Safranyik and Carroll 2006). Pines have evolved
43 resins and other defensive compounds to resist attack (Franceschi *et al.* 2005; Boone *et al.* 2011),
44 and synchronous adult emergence facilitates a mass attack on individual trees that can occur

45 more quickly than a tree can mobilize its defenses (Berryman *et al.* 1985). Successful attacks on
46 the largest and often better defended trees with the thickest phloem (i.e., food for developing
47 larvae) can lead to increased offspring and ultimately a population outbreak (Raffa *et al.* 2008).
48 In northern US mountain pine beetle, synchronous adult emergence is achieved by temperature-
49 dependent physiological strategies including a facultative prepupal diapause (Bentz and Hansen
50 2017) and lifestage-specific developmental rates and thresholds (Bentz *et al.* 1991; Powell and
51 Logan 2005; Régnière *et al.* 2012).

52 In addition to facilitating synchronicity in lifestage timing, evolved temperature-
53 dependent physiological strategies control the time required to complete a generation and
54 subsequently the number of generations that can be completed annually (i.e., voltinism). A
55 lifecycle that is appropriately timed and results in one generation per year is considered
56 univoltine, two generations in a single year is considered bivoltine, and semivoltine generations
57 occur when two years are required for a single generation. Mountain pine beetle adult emergence
58 typically occurs in mid to late summer across its range and univoltinism is considered the most
59 optimal strategy (Logan and Bentz 1999; Safranyik and Carroll 2006). Bivoltinism has not been
60 observed at the warmest or most southern extent of the current mountain pine beetle range in the
61 US (Hopkins 1909; Bentz *et al.* 2014; Bentz and Powell 2014; BJB unpublished). A combination
62 of univoltine and semivoltine strategies, however, can be found in outbreak-level populations at
63 the highest elevations (Bentz *et al.* 2014), demonstrating that semivoltinism is a viable strategy
64 in the coldest areas (Weed *et al.* 2015).

65 The pervasiveness of univoltinism found across the range of mountain pine beetle in the
66 western US masks thermally-dependent strategies that evolved as a result of climatic differences
67 across latitudes (Bentz *et al.* 2014), as revealed in common garden studies. Median generation

68 time of southern US populations was significantly longer than mountain pine beetle from
69 northern US populations when reared at the same constant temperature (Bentz *et al.* 2001; Bentz
70 *et al.* 2011; Bracewell *et al.* 2013). Countergradient variation, a type of phenotypic plasticity
71 wherein the evolutionary response to a gradient is opposite of the ecological response, is not
72 uncommon in species with large geographic ranges (Conover and Schultz 1995). The observed
73 longer generation time in southern mountain pine beetle is likely a result of selection pressure to
74 maintain univoltinism despite a warmer climate. To predict the impact of continued climate
75 warming on population success, an understanding of the strategies and lifestages responsible for
76 developmental differences between southern and northern mountain pine beetle populations is
77 critical.

78 Our goal was to describe temperature-dependent lifestage-specific developmental times
79 and thresholds for a southern US mountain pine beetle population. We then compare our results
80 with previously described developmental data for a northern US population (Régnière *et al.*
81 2012). Using maximum posterior likelihood estimation, we fit observed data on time to complete
82 each lifestage across a range of temperatures to the same seven parameter rate function used by
83 Régnière *et al.* (2012), adding a Bayesian prior to the procedure to increase stability in the model
84 fits. We also used transfer treatments to facilitate timely data collection and increase survival at
85 extreme temperatures, and developed a method of assessing the reliability and effectiveness of
86 those treatments. Our comparison of lifestage-specific thermal responses for northern and
87 southern mountain pine beetle populations provides a platform for increased understanding of
88 evolved developmental differences across latitudes, in addition to the development of a
89 phenology model for southern populations that can be used in predicting range-wide population
90 success in a changing climate.

91

92 **2. Methods**93 *2.1 Experimental Materials and Design*

94 To obtain fresh phloem material to infest with mountain pine beetle parents from a
95 southern population, one live, un-infested southwestern white pine (*Pinus strobiformis* Engelm.)
96 was harvested on 3 May, 2016 near Flagstaff, Arizona (AZ) (35.36272, -111.7439) and cut into
97 50-55 cm bolts. Bolts were transported to the US Forest Service Rocky Mountain Research
98 Station (RMRS) laboratory in Logan, Utah (UT) and bolt ends were waxed (Gulf Wax, Roswell,
99 GA) to retain moisture and then stored at 0 °C for up to 2 months. Unmated adults for infesting
100 the bolts of host tree material were acquired by harvesting a mountain pine beetle-infested
101 southwestern white pine on 4 May, 2016 near Flagstaff, AZ (35.35506, -111.6132). The infested
102 tree was cut into bolts 45-50 cm long and transported to the RMRS laboratory in Logan, UT.
103 Bolts ends were waxed to retain moisture and stored at 0 °C when not used to produce brood.
104 Eight bolts from the infested tree were immediately placed in incubators (Percival Scientific,
105 Perry, IA) (4 bolts per incubator) at ~20 °C to facilitate brood development and adult emergence.
106 Adults were collected daily and kept at 4 °C in Petri dishes for 1-7 days before use. Moistened
107 filter paper was placed in each dish to reduce desiccation. Beetles were sexed using secondary
108 sex characteristics on the seventh tergite (Lyon 1958).

109 Phloem sandwiches were used to monitor lifestage-specific development following
110 methods found in Bentz *et al.* (1991) and Hansen *et al.* (2001). Sandwiches were initiated with
111 eggs, and development of each individual was monitored on a daily basis until the adult stage
112 was reached or mortality occurred. To obtain eggs, several un-infested bolts were manually
113 infested with male/female pairs by inserting first a female then a male into holes drilled

114 vertically into the phloem. Wire mesh screen was placed over each hole to prevent parent beetle
115 escape. The bolts were inverted and incubated at room temperature for 10-12 days before peeling
116 the bark to expose egg galleries. Eggs were collected from three 1.5 cm gallery sections, starting
117 with the most recently completed gallery, and eggs were considered to be 1, 2, or 3 days old
118 respectively based on preliminary data on rate of gallery construction and oviposition (McManis
119 2018).

120 To obtain phloem for sandwiches, the outer bark was stripped from several un-infested
121 bolts with a sterilized draw knife. Phloem pieces were cut into six-inch squares using a sterilized
122 knife and carefully peeled from the bolt. Peeled phloem squares were vacuum packed
123 (FoodSaver, Sunbeam Products, Boca Raton, FL) and refrigerated for 1-2 days before use.
124 Phloem sandwiches were assembled using tools sterilized in 95% ethanol to reduce
125 contamination. On the cambial surface of the phloem, seven evenly spaced niches for eggs were
126 cut along the centerline of the phloem parallel to the grain with a sterilized dissecting probe. For
127 each phloem sandwich eggs of similar age were used (i.e., 1, 2 or 3 days old). Phloem containing
128 eggs was placed between sterilized glass and sterilized plexiglass plates, with plexiglass against
129 the bark side and glass against the cambial side with the eggs. These `sandwiches' were clamped
130 on each edge and the edges secured with tape (Nexcare 3M, St. Paul, MN) and parafilm (Bemis,
131 Neenah, WI).

132 Completed phloem sandwiches (hereafter, "plates") were numbered and placed upright in
133 racks in 26 cm diameter plastic desiccators (Bel-Art™ SP Scienceware™, Fisher Scientific,
134 Pittsburg, PA), with a 5% sodium chloride solution in the bottom to maintain constant humidity
135 (~ 93%), and the desiccators were placed in incubators. Individual eggs were numbered from 1-7
136 for each plate by writing on the glass next to the current location of the individual. There were

137 seven plates per desiccator, and two desiccators per temperature, for a total of 98 individuals at
138 each temperature, 10, 15, 20, 25, 27, 28, 29 and 30 °C. Experimental temperatures were spread
139 asymmetrically across the previously developed rate curve for a northern population (Régnière *et*
140 *al.* 2012) to ensure sufficient data to resolve the upper and lower developmental thresholds.
141 Because the slope of the development rate curve at temperatures lower than the expected optimal
142 (~25 °C) is shallow, we included temperatures every 5 °C between 10 and 25 °C. Expecting a
143 sudden sharp drop in development rate above the optimal, we included temperatures every 1 °C
144 between 27 and 30 °C.

145 Plates were inspected under a dissecting microscope every 24 hours and larval head
146 capsule width recorded. When present, discarded head capsule exuviae, indicating a recent molt
147 to a new instar, was also recorded for each individual. In the absence of head capsule exuviae,
148 increases in head capsule width of at least 0.5 mm between individual observations were
149 recorded as advancement to the next instar. An individual was considered a pupa when a loss of
150 larval body morphology and the presence of proto-wing structures were observed. An individual
151 was considered an adult when adult structures were present (e.g. legs and elytra) and
152 sclerotization began (i.e., the individual turned from a creamy white to light brown). From these
153 data, the number of days to complete each instar/lifestage at a particular temperature was
154 calculated for each individual. Individuals that failed to complete a lifestage (i.e., died or were
155 still alive at the end of the experiment) or transitioned between lifestages while hidden beneath
156 the surface of the phloem (so that the exact date of transition was unknown) were included as
157 censored data *sensu* Régnière *et al.* (2012).

158

159 *2.2 Transfer Treatment Implementation*

160 Based on estimates of development time from preliminary data, in addition to previously
161 published data on a northern population (Régnière *et al.* 2012), total development time for
162 individuals at or below 15 °C and above 27 °C would require an extended amount of time, in
163 addition to the likelihood of reduced survival. To reduce these effects, we used transfer
164 treatments. Transfer treatments assume that thermal history does not influence development time,
165 and they increase the probability of observing lower and upper thermal thresholds (Régnière *et al.*
166 2012). In transfer treatments, plates spent part of the time at the treatment temperature and part
167 of the time at 25 °C. Transfer treatments were used for 10, 15, 29 and 30 °C and included 98
168 individuals (14 plates) per temperature.

169 For each lifestage, individuals were kept at the treatment temperature for approximately
170 seven days before transfer to 25 °C. Seven days was chosen as a compromise between
171 accelerating data collection and ensuring that, even where rates were lowest, a non-trivial
172 amount of an individual's development (i.e. 10-15%) would be completed at the treatment
173 temperature. If an individual in the plate had already advanced to the next lifestage, the plate was
174 left at the treatment temperature for another seven days. Plates were transferred from 25 °C back
175 to the treatment temperature one day after the most advanced individual completed the current
176 lifestage. There were seven individuals per plate, and the individual in the most advanced
177 lifestage was used to determine if and when a plate should be transferred. This insured that all
178 individuals spent at least seven days per lifestage at the treatment temperature, although some
179 individuals spent more time.

180 For comparison, a constant temperature control at 10 °C with 49 individuals that were not
181 transferred was established. After 382 days, the majority of these individuals had not completed

182 development to the pupal stage. Data on eggs, and first, second and third instars were used to
 183 compare development time of transferred and non-transferred individuals at 10 °C.

184

185 2.3 Model Development

186 Although Régnière *et al.* (2012) used a lognormal error distribution for northern
 187 population data, a normal distribution was a better description of variability among southern
 188 individual development times/rates at a given temperature. Lognormal error is multiplicative and
 189 lower rates correspond to lower variance regardless of temperature (Régnière *et al.* 2012).
 190 Conversely, normal error is additive and does not scale with development times/rates and was a
 191 better fit to the variability among southern individuals. Using normally-distributed error also
 192 allowed for the possibility that an individual's upper and lower threshold may vary relative to the
 193 median threshold in the population, whereas lognormal error assumes fixed upper and lower
 194 developmental thresholds for all individuals.

195 For the n^{th} individual, there is a mismatch (ε_n) between the observed development rate
 196 and the modeled mean rate due to individual variation in rate, which we assumed to be normally
 197 distributed with variance σ^2 . Therefore an individual's rate of development, $r_n(T, \boldsymbol{\theta})$, relates to
 198 the mean rate, $r_o(T, \boldsymbol{\theta})$, as

$$199 \quad r_n(T, \boldsymbol{\theta}) = r_o(T, \boldsymbol{\theta}) + \varepsilon_n, \quad \varepsilon_n \sim N(0, \sigma^2), \quad (1)$$

200 where T is temperature, $\boldsymbol{\theta}$ is a vector of parameters, and $r_o(T, \boldsymbol{\theta})$ is the rate function

$$201 \quad r(T, \boldsymbol{\theta}) = \psi \left[e^{\omega(T-T_b)} - \left(\frac{T_m-T}{T_m-T_b} \right) e^{-\frac{\omega(T-T_b)}{\Delta_b}} - \left(\frac{T-T_b}{T_m-T_b} \right) e^{\omega(T_m-T_b) - \frac{T_m-T}{\Delta_m}} \right], \quad (2)$$

202 previously used by Régnière *et al.* (2012). In this rate function, T_m and T_b correspond to the
 203 upper and lower temperature thresholds, respectively, and the remaining parameters are shape

204 parameters. The observed development time, t_n , of individual n at constant temperature (T)
 205 gives an observed rate, $r_n(T, \theta) = \frac{1}{t_n}$, and therefore the likelihood of observing t_n is

$$206 \quad L_n = \frac{e^{-\frac{(r_o(T, \theta) - \frac{1}{t_n})^2}{2\sigma^2}}}{\sqrt{2\pi\sigma^2}}. \quad (3)$$

207 The negative log likelihood for the single observation, t_n , (after multiplying the numerator and
 208 denominator by t_n^2) becomes

$$209 \quad NLL_n = \frac{(r_o(T, \theta)t_n - 1)^2}{2\sigma^2 t_n^2} + \frac{1}{2} \ln(2\pi\sigma^2). \quad (4)$$

210

211 2.4 Transfer Treatment Data

212 For individuals that were transferred between temperatures to accelerate development and
 213 increase survival, fitting their data to the rate curve was more complicated. Integrating equation
 214 (1) gives

$$215 \quad \int_0^t r_n(T, \theta) dt = \int_0^t r_o(T, \theta) dt + \int_0^t \varepsilon_n dt. \quad (5)$$

216 For an individual that spent some time (t_{n1}) at a treatment temperature (T_1) and some time (t_{n2})
 217 at T_2 (25 °C) this results in

$$218 \quad \int_0^{(t_{n1}+t_{n2})} r_o(T, \theta) dt = r_o(T_1, \theta)t_{n1} + r_o(T_2, \theta)t_{n2}. \quad (6)$$

219 The integral from 0 to time of completion is always one,

$$220 \quad \int_0^{(t_{n1}+t_{n2})} r_n(T, \theta) dt = 1, \quad (7)$$

221 and therefore

$$222 \quad 1 = r_o(T_1, \theta)t_{n1} + r_o(T_2, \theta)t_{n2} + \varepsilon_n(t_{n1} + t_{n2}). \quad (8)$$

223 Solving for ε_n and using $\varepsilon_n \sim N(0, \sigma^2)$, the negative log likelihood for observing ($t_{n1} + t_{n2}$)
 224 becomes

$$225 \quad NLL_n = \frac{(r_o(T_1, \theta)t_{n1} + r_o(T_2, \theta)t_{n2} - 1)^2}{2\sigma^2(t_{n1} + t_{n2})^2} + \frac{1}{2} \ln(2\pi\sigma^2). \quad (9)$$

226

227 *2.5 Testing Consistency of Transfer Treatment Data*

228 Developmental rates for transferred and control individuals at 10 °C were compared
 229 based on developmental deviance. Developmental deviance (Δ_n) is a measure of how observed
 230 developmental time for an individual differs from the median time in a particular lifestage. For
 231 individuals in a transfer treatment being moved between 10 and 25 °C, we first calculated the
 232 median rate of development at 10 °C, R_{10} , using observed development rates for individuals at a
 233 constant 10 °C, and the median rate of development at 25 °C, R_{25} , using observed development
 234 rates for individuals at a constant 25 °C. The Δ_n for an individual is calculated as

$$235 \quad \Delta_n = t_{10}R_{10} + t_{25}R_{25}, \quad (10)$$

236 where t_{10} and t_{25} are time spent at 10 °C and 25 °C, respectively. The Δ_n for individuals at a
 237 constant 10 °C who were not transferred is calculated the same way, except $t_{25} = 0$, so

$$238 \quad \Delta_n = t_{10}R_{10}. \quad (11)$$

239 If development rate is not affected by thermal history of an individual, then the distributions of
 240 Δ_n values for individuals at a constant 10 °C and transferred individuals will not be significantly
 241 different. If development rate is affected by thermal history, then there will be a significant
 242 difference in the Δ_n values between treatment groups. Because sample sizes were relatively
 243 small and somewhat skewed, we used a non-parametric Wilcoxon Rank-Sum test (W) to compare
 244 groups (R Core Team 2015).

245

246 *2.6 Censored Data*

247 Censored data represents individuals who failed to complete their current lifestage while
 248 data collection was ongoing or transitioned between lifestages while unobservable beneath the
 249 phloem such that the exact duration of the lifestage was unknown. Therefore, their total
 250 development time is at least as long as the observation time, but could have been longer. The
 251 probability (P) the observed time for a censored data point is less than the mean development
 252 time for that temperature is

$$253 \quad P\left(t_n < \frac{1}{r_o(T, \theta)}\right) = P(r_o(T, \theta)t_n < 1) = P(r_o(T, \theta)t_n - 1 < 0). \quad (12)$$

254 Since $1 - r_o(T, \theta)t_n = \varepsilon_n t_n$ and $\varepsilon_n t_n \sim N(0, \sigma^2 t_n^2)$,

$$255 \quad P(r_o(T, \theta)t_n - 1 < 0) = F(r_o(T, \theta)t_n; \sigma^2 t_n^2), \quad (13)$$

256 where F is the normal CDF with variance $\sigma^2 t_n^2$. The negative log likelihood for a censored
 257 observation, t_n , is then

$$258 \quad NLL_n = -\ln[F(r_o(T, \theta)t_n; \sigma^2 t_n^2)]. \quad (14)$$

259

260 *2.7 Adding a Bayesian Prior to the Negative Log Likelihood*

261 A Bayesian prior was used to improve the stability of the fit for upper and lower
 262 threshold parameters. This weights the maximum likelihood fit using prior information and
 263 confidence in the prior. According to Bayes' Theorem, the posterior distribution of the vector of
 264 parameters, θ , satisfies

$$265 \quad P(\theta|t_n) = \frac{1}{P(t_n)} P(t_n|\theta)P(\theta) \quad (15)$$

266 In this expression $P(t_n|\theta)$ is the likelihood, $P(\theta)$ is a prior distribution of the parameters and
 267 $P(t_n)$ is an unknown constant that can be ignored. For example, assuming the prior distribution of
 268 the upper threshold (T_u) is normal we can write the posterior likelihood as

269
$$P(T_u) = e^{-NLL(\theta|t_n)} \left(\frac{e^{-\frac{(T_u-U)^2}{2\sigma_u^2}}}{\sqrt{2\pi\sigma_u^2}} \right), \quad (16)$$

270 where U and σ_u^2 are the mean and variance of the prior distribution. The posterior probability of
 271 observing t_n is therefore

272
$$P(t_n) \propto e^{-NLL(\theta|t_n)} P(T_u) P(T_l), \quad (17)$$

273 where $P(T_u)$ and $P(T_l)$ are, respectively, the prior probability of particular upper and lower
 274 thresholds. Assuming $T_u \sim \text{Normal}(U, \sigma_u^2)$ and $T_l \sim \text{Normal}(L, \sigma_l^2)$, the negative log posterior
 275 (NLP) is

276
$$NLP(\theta|t_n) = NLL(\theta|t_n) + \frac{(T_u-U)^2}{2\sigma_u^2} + \frac{(T_l-L)^2}{2\sigma_l^2} + K. \quad (18)$$

277 Here K is a constant which is independent of parameter values. Prior means for the upper
 278 thresholds were chosen based on high observed mortality in the data at 30°C, and prior variances
 279 were set at $\sigma_u^2 = 0.125$ and $\sigma_l^2 = 0.50$, reflecting the differing steepness of the rate curves
 280 approaching upper and lower thresholds. Where consistent with our data, the lower threshold
 281 means for the northern population (Régnière *et al.* 2012) were used as priors for the southern
 282 population. If a fit generated by minimizing NLP resulted in a steep drop off in the curve
 283 unsupported by our data, the prior mean was stepped down by 0.5 °C up to six times (at most a
 284 3 °C decrease), the fit was rerun, and the parameters associated with the lowest NLP was kept.

285

286 2.8 Descriptive Statistics and Pairwise Comparison of Observed Developmental Times

287 Using our data for the southern population and data for the northern population published
 288 in Régnière *et al.* (2012), we compared observed developmental time in each population for egg,
 289 larva, and pupa at temperatures used in both studies (i.e., 10, 20, 25, 27 °C). Data were analyzed
 290 using a generalized linear model with a Poisson distribution (SAS Institute Inc., v9.4). Post-hoc

291 pairwise comparisons were tested with a Tukey-Kramer adjustment for multiple comparisons.
292 We tested for size differences between the fourth instar with and without a fifth instar using a
293 similar analyses based on a normal distribution.

294

295 3. Results

296 When individuals across all treatments were considered, mortality was greatest in the
297 fourth instar (35%) and egg (29%). Mortality for all lifestages was lowest at 20 °C, and no
298 individuals survived the pupal stage at 29 or 30 °C (Fig 1). Although four instars have
299 historically been described for mountain pine beetle (Amman and Cole 1983; Rosenberger *et al.*
300 2018), Myrholm and Langor (2016) recently observed individuals with up to seven instars. We
301 observed a fifth instar in 57 individuals (~14% of fourth instar individuals) at temperatures \geq
302 15 °C. Headcapsule size of fifth instar (mean = 1.39 ± 0.09 , N = 51) was larger than the size of
303 fourth instar (mean = 1.26 ± 0.08 , N = 348) ($\chi^2 = 101.23$, $P < 0.0001$). A headcapsule size was
304 not measured for those individuals inappropriately oriented within phloem sandwiches. Of those
305 individuals with a fifth instar, size of the fourth instar (mean = 1.16 ± 0.08 , N = 48) was smaller
306 than fourth instars that did not molt to a fifth instar (mean = 1.27 ± 0.07 , N = 300) ($\chi^2 = 32.50$, P
307 < 0.0001). Due to limited data, model parameters for a fifth instar could not be estimated.

308 Developmental deviance, and therefore developmental rates, did not differ significantly
309 for second instars that were either transferred from 10 to 25 °C or kept at a constant 10 °C ($P =$
310 0.427). However, developmental deviance of eggs, first, and third instar larvae were significantly
311 different between transferred individuals and those kept at constant 10 °C (Fig. 2). We also
312 observed reduced variability among transferred eggs, relative to eggs kept at a constant
313 temperature (Fig. 2). There was insufficient constant temperature data to test for developmental

314 differences between transferred and non-transferred individuals in the fourth instar, and
315 subsequently pupa, because the majority of non-transferred fourth instars held at a constant
316 10 °C did not pupate. In contrast, a majority of the individuals transferred between 10 and 25 °C
317 did pupate. Differences in pupation rates between transferred and non-transferred individuals are
318 most likely a result of prepupal facultative diapause development that is sped up with warm
319 temperatures (Bentz and Hansen 2017). Due to observed differences, model fits were performed
320 separately for temperature data with and without transfers for all stages except second instar.

321 Using constant temperature and censored data (i.e., no transfer data), estimated
322 parameters from the fit to equation (2) differed among lifestages (Table 1). In particular, the
323 lower developmental threshold (T_b) for the fourth instar was estimated to be substantially higher
324 (~15 °C) than all other life stages which ranged from ~4.6 to 6.3 °C (Table 1). The estimated
325 upper developmental thresholds (T_m) were similar among lifestages and ranged from 30.8 to
326 31.9 °C. Optimal developmental rate was estimated to be between 24.8 and 26.5 °C for all
327 lifestages (Fig. 3). At the optimal rate, first instars developed approximately twice as fast as eggs,
328 third instars and pupae, and almost four times as fast as fourth instars. When data on individuals
329 that were transferred among temperatures were included in parameter estimation, predicted
330 development rates differed slightly from fits using constant temperature data, particularly in the
331 fourth instar (Table 2; Fig. 4). Fourth instars exposed to 25 °C during part of their development
332 (i.e., a transfer treatment) had positive development at temperatures ≤ 15 °C (Fig. 4) in contrast
333 to fourth instars that were kept at a constant 10 or 15 °C, where no development was observed
334 (Fig. 3).

335 We were interested in comparing lifestage-specific observed developmental times and
336 fitted rate curves of the southern population with those previously described for a northern

337 population (Bentz *et al.* 1991, Régnière *et al.* 2012, Bentz and Powell 2014). The same phloem
338 sandwich methodology was used to collect data for both populations. At a constant 10 °C,
339 observed development time of southern population eggs and second instars was faster than
340 northern individuals in the same lifestages, and southern third instars developed slower than
341 northern third instars (Table 3). No individuals in either population completed fourth instar
342 development (i.e., pupated) at 10 °C without some period of development at a warmer
343 temperature. There were no significant differences in observed development time between the
344 populations in any lifestage at 20 °C (Table 3). At 25 °C southern fourth instars developed
345 significantly slower than northern fourth instars, and at 27 °C southern second and third instars
346 developed faster than northern individuals (Table 3). When fitted development rate curves for
347 each population were compared, using only not-transferred data, estimated upper thresholds were
348 higher for southern compared to northern individuals across all lifestages (Fig. 5). Lower
349 thresholds were similar between the populations in all lifestages except the fourth instar, where
350 southern individuals developed at a lower temperature (Fig. 5). Estimated development rates of
351 southern second and third instars were higher than northern second and third instars at all
352 temperatures.

353

354 **4. Discussion**

355 Our goal was to describe temperature-dependent lifestage-specific developmental times
356 and thresholds for a southern mountain pine beetle population. We then compared these
357 developmental data from a southern population with previously described developmental data for
358 a northern population that was collected using the same phloem sandwich method (Régnière *et al.*
359 2012). Previous research showed that in common garden experiments, median generation time of

360 a southern population was significantly longer (~73 days) than that of a northern population at
361 22 °C (Bracewell *et al.* 2013), and we were interested in identifying the lifestage(s) and evolved
362 traits that may differ between the populations. Estimated upper developmental thresholds and
363 optimal development rates were at slightly higher temperatures for southern compared to
364 northern individuals across all lifestages. Although northern data was not collected at
365 temperatures ≥ 28 °C, the slowing trend in northern individuals at 27 °C suggests optimal
366 development occurs at slightly higher temperatures in southern individuals. Southern fourth
367 instars that were not transferred to 25 °C during development pupated (i.e., completed the fourth
368 instar) at a lower temperature than did northern fourth instars given the same treatment, a result
369 also found by Bentz and Hansen (2017). Both populations have a facultative prepupal (i.e., last
370 stage of the fourth instar) diapause, and induction can occur at higher temperatures in northern
371 (15 to 17 °C) compared with southern populations (< 15 °C) (Bentz and Hansen 2017). Southern
372 fourth instars developed slower than northern fourth instars, but only significantly so at 25 °C. At
373 all other temperatures and lifestages, southern individuals generally developed faster than
374 northern individuals.

375 Given the longer total development time from egg through adult emergence of southern
376 compared to northern individuals at a constant 22.5 °C (Bracewell *et al.* 2013), we hypothesized
377 that selection would act on one or more lifestages to slow development rates and thereby
378 maintain univoltinism despite warmer habitat temperatures. Although development was
379 generally faster in southern individuals, our result that southern fourth instars developed slower
380 (~ 5 days) than northern fourth instars at 25 °C highlights that physiological aspects of the fourth
381 instar may partially explain generation time differences between the populations. In addition to
382 the lifestages monitored in our study, total development time as reported by Bracewell *et al.*

383 (2013) included oviposition and teneral (i.e., pre-emergent) adult development through
384 emergence. McManis (2018) showed that oviposition is slightly slower in southern compared to
385 northern mountain pine beetle (18.5 vs 12 days at 21 °C). This time difference, combined with
386 the few days difference in fourth instar, contribute only marginally to the observed median
387 difference of 73 days in total generation time. A potential additional explanation for differences
388 in generation time observed by Bracewell *et al.* (2013) is trait differences between the
389 populations in the unstudied teneral adult stage. Differences in evolved traits such as adult
390 development and maturation rates, and temperature thresholds for emergence from beneath the
391 bark could result in differences in adult emergence timing and generation time. Because teneral
392 adults feed on spores of fungal associates to obtain vital nutrients prior to emergence (Six and
393 Paine 1998), differences in fungal acquisition or species composition could also play a role in
394 developmental differences between the populations (Addison *et al.* 2014).

395 A proportion of individuals (~14%) molted to a fifth instar prior to pupation. Plasticity in
396 the number of instars an insect may go through prior to pupation can be influenced by multiple
397 environmental factors including temperature and food quality and quantity (Esperk *et al.* 2007).
398 Myrholm and Langor (2016) recently observed mountain pine beetles with up to seven instars,
399 and the headcapsule sizes of the additional instars were between that of instar 3 and instar 4.
400 They suggested that inadequate nutrition results in extra instars and that larvae must attain a
401 threshold size to initiate metamorphosis (Nijhout 1994). In contrast to Myrholm and Langor
402 (2016), the size of the additional instar we observed was larger than the size of fourth instars, and
403 therefore was the last instar prior to pupation. Moreover, individuals that molted to a fifth instar
404 were smaller as a fourth instar than were individuals that pupated following the fourth instar. Our
405 results concur with Myrholm and Langor (2016) that a size threshold for pupation likely exists in

406 mountain pine beetle, and that additional instars may serve as a compensatory mechanism in
407 adverse conditions (Esperk *et al.* 2007). Collectively, however, results suggest that the timing of
408 supernumerary instars is not fixed.

409 Measuring insect development near thresholds can be difficult. We used temperature
410 transfer treatments to increase survival at extreme temperatures and reduce the time required for
411 measurements when development is slowed (Régnière *et al.* 2012). An assumption of transfer
412 treatments is that past thermal history does not affect future developmental rate. We found that
413 only second instars transferred between 10° and 25 °C met this assumption of no difference, with
414 first and third instars developing faster than those kept at a constant 10 °C and transferred eggs
415 developing more slowly. The apparent slowing of development in transferred eggs is likely due
416 to the difficulty of accurately aging eggs, rather than the transfer treatment alone. For first and
417 third instars, however, pulses of warm temperature (25 °C) during development at 10 °C resulted
418 in slightly faster development. Future study is needed to determine if this effect is due to the
419 relatively brief exposure of transferred individuals to their treatment temperature, or if it reflects
420 more complex physiological processes.

421 A physiological-based description of an organism's thermal response can provide a
422 robust framework for making predictions of potential range shifts due to climatic changes. A
423 major benefit of mechanistic, relative to statistical, models is that inherent biological behaviors
424 can emerge. Our description of developmental responses of a southern US mountain pine beetle
425 population provide a foundation for incorporating evolved geographic variation in mountain pine
426 beetle lifecycle timing into predictive models. Our findings suggest that future research should
427 focus on physiological traits in the fourth instar and teneral adult lifestages to further our

428 understanding of intraspecific fitness trait differences that drive population success across the
429 expansive mountain pine beetle range.

430

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439

440 **References**

- 441 Addison, A., Powell, J., Bentz, B., and Six, D. 2014. Integrating models to investigate critical
442 phenological overlaps in complex ecological interactions: The mountain pine beetle-fungus
443 symbiosis. *Journal of Theoretical Biology*, **368**: 55-66.
- 444 Bentz, B., and Hansen, E. M. 2017. Evidence for a prepupal diapause in the mountain pine beetle
445 (*Dendroctonus ponderosae*). *Environmental Entomology*, **47**:175-183, doi:
446 10.1093/ee/nvx192.
- 447 Bentz, B., and Powell, J. A. 2014. Mountain pine beetle seasonal timing and constraints to
448 bivoltinism. *The American Naturalist*, **184**:787-796.
- 449 Bentz, B., Vandygriff, J., Jensen, C., Coleman, T., Maloney, P., Smith, S., Grady, A., and Schen-
450 langenheim, G. 2014. Mountain pine beetle voltinism and life history characteristics across

- 451 latitudinal and elevational gradients in the western United States. *Forest Science*, **60**:434–
452 449.
- 453 Bentz, B.J., Bracewell, R.R., Mock, K.E., and Pfrender, M.E. 2011. Genetic architecture and
454 phenotypic plasticity of thermally-regulated traits in an eruptive species, *Dendroctonus*
455 *ponderosae*. *Evolutionary Ecology*, **25**: 1269–1288. doi:10.1007/s10682-011-9474-x.
- 456 Bentz, B.J., Logan, J.A., and Amman, G.D. 1991. Temperature-dependent development of the
457 mountain pine beetle (Coleoptera: Scolytidae) and simulation of its phenology. *Canadian*
458 *Entomology*, **123**:1083–1094. doi:10.4039/Ent1231083-5.
- 459 Bentz, B.J., Logan, J.A., and Vandygriff, J.C. 2001. Latitudinal variation in *Dendroctonus*
460 *ponderosae* (Coleoptera: Scolytidae) development time and adult size. *The Canadian*
461 *Entomologist*, **133**:375–387.
- 462 Berryman, A.A., Dennis, B., Raffa, K.F., and Stenseth, N.C. 1985. Evolution of optimal group
463 attack, with particular reference to bark beetles (Coleoptera: Scolytidae). *Ecology*, **66**:898–
464 903. doi:10.2307/1940552.
- 465 Boone, C.K., Aukema, B.H., Bohlmann, J., Carroll, A.L., and Raffa, K.F. 2011. Efficacy of tree
466 defense physiology varies with bark beetle population density: a basis for positive feedback
467 in eruptive species. *Canadian Journal of Forest Research*, **41**:1174–1188. doi:10.1139/x11-
468 041.
- 469 Bracewell, R.R., Pfrender, M.E., Mock, K.E., and Bentz, B.J. 2013. Contrasting geographic
470 patterns of genetic differentiation in body size and development time with reproductive
471 isolation in *Dendroctonus ponderosae* (Coleoptera: Curculionidae, Scolytinae). *Annals of*
472 *the Entomological Society of America*, **106**:385–391. doi:10.1603/AN12133.
- 473 Conover, D.O., and Schultz, E.T. 1995. Phenotypic similarity and the evolutionary significance

- 474 of countergradient variation. *Trends in Ecology & Evolution*, **10**:248–252.
475 doi:10.1016/S0169-5347(00)89081-3.
- 476 Cooke, B.J., and Carroll, A.L. Predicting the risk of mountain pine beetle spread to eastern pine
477 forests: Considering uncertainty in uncertain times. *Forest Ecology and Management*, **396**:
478 11-25.
- 479 Danks, H. V. 1987. Insect dormancy: an ecological perspective. *In* Biological. Biological Survey
480 of Canada Monograph Series, No.1). Entomological Society Canada 439,
481 <http://www.biology.ualberta.ca/bsc/pdf/contents.pdf>.
- 482 Deutsch, C.A., Tewksbury, J.J., Huey, R.B., Sheldon, K.S., Ghalambor, C.K., Haak, D.C., and
483 Martin, P.R. 2008. Impacts of climate warming on terrestrial ectotherms across latitude.
484 *Proceedings of the National Academy of Sciences of the United States of America*,
485 **105**:6668–6672. doi:10.1073/pnas.0709472105.
- 486 Dowle, E.J., Bracewell, R.R., Pfrender, M.E., Mock, K.E., Bentz, B.J., and Ragland, G.J. 2017.
487 Reproductive isolation and environmental adaptation shape the phylogeography of
488 mountain pine beetle (*Dendroctonus ponderosae*). *Molecular Ecology*, **26**:6071-6084.
- 489 Esperk, T., Tammaru, T., and Nylin, S. 2007. Intraspecific variability in number of larval instars
490 in insects. *Journal of Economic Entomology*, **100**:627-645.
- 491 Forrest, J.R.K., and James, D.T. 2011. An examination of synchrony between insect emergence
492 and flowering in Rocky Mountain meadows. *Ecological Monographs*, **81**:469–491.
493 doi:10.1890/10-1885.1.
- 494 Franceschi, V.R., Franceschi, V.R., Krokene, P., Krokene, P., Christiansen, E., Christiansen, E.,
495 Krekling, T., and Krekling, T. 2005. Anatomical and chemical defenses of conifer bark
496 against bark beetles and other pests. *New Phytologist*, **167**:353–375. doi:10.1111/j.1469-

- 497 8137.2005.01436.x.
- 498 Hansen, E.M., Bentz, B.J., and Turner, D.L. 2001. Physiological basis for flexible voltinism in
499 the spruce beetle (Coleoptera: Scolytidae). *The Canadian Entomologist*, **133**:805–817.
500 doi:10.4039/Ent133805-6.
- 501 Hicke, J.A., Meddens, A.J.H., and Kolden, C.A. 2016. Recent tree mortality in the western
502 United States from bark beetles and forest fires. *Forest Science*, **62**:141–153.
503 doi:10.5849/forsci.15-086.
- 504 Hopkins, A.D. 1909. Contributions Toward a Monograph of the Scolytid Beetles. In USDA
505 Bureau of Entomology. Washington D.C.
- 506 Li, J.L., Johnson, S.L., and Banks Sobota, J. 2011. Three responses to small changes in stream
507 temperature by autumn-emerging aquatic insects. *Journal of the North American*
508 *Benthological Society*, **30**:474–484. doi:10.1899/10-024.1.
- 509 Logan, J.A., and Bentz, B.J. 1999. Model analysis of mountain pine beetle (Coleoptera:
510 Scolytidae) seasonality. *Environmental Entomology*, **28**:924–934. doi:10.1093/ee/28.6.924.
- 511 Lyon, R.L. 1958. A useful secondary sex character in *Dendroctonus* bark beetles. *The Canadian*
512 *Entomologist*, **90**:582–584. doi:10.4039/Ent90582-10.
- 513 McManis, A.E. 2018. Phenology of a southern population of mountain pine beetle
514 (*Dendroctonus ponderosae*). Masters Thesis. Utah State University, Logan, UT.
- 515 Myrholm, C.L., and Langor, D.W. 2016. Assessment of the impact of symbiont Ophiostomatales
516 (Fungi) on mountain pine beetle (Coleoptera: Curculionidae) performance on a jack pine
517 (Pinaceae) diet using a novel *in vitro* rearing method. *The Canadian Entomologist*, **148**:68-
518 82.
- 519 Nijhout, H.F. 1994. *Insect hormones*. Princeton University Press, Princeton, NJ.

- 520 Powell, J.A., and Logan, J.A. 2005. Insect seasonality: Circle map analysis of temperature-driven
521 life cycles. *Theoretical Population Biology*, **67**:161–179. doi:10.1016/j.tpb.2004.10.001.
- 522 R Core Team. 2015. R: A Language and Environment for Statistical Computing. doi:ISBN 3-
523 900051-07-0.
- 524 Raffa, K.F., Aukema, B.H., Bentz, B.J., Carroll, A.L., Hicke, J. a., Turner, M.G., and Romme,
525 W.H. 2008. Cross-scale drivers of natural disturbances prone to anthropogenic
526 amplification: The dynamics of bark beetle eruptions. *BioScience*, **58**:501-517.
527 doi:10.1641/B580607.
- 528 Régnière, J., Powell, J., Bentz, B., and Nealis, V. 2012. Effects of temperature on development,
529 survival and reproduction of insects: Experimental design, data analysis and modeling.
530 *Journal of Insect Physiology*, **58**:634–647. doi:10.1016/j.jinsphys.2012.01.010.
- 531 Safranyik, L., and Carroll, A. 2006. The biology and epidemiology of the mountain pine beetle
532 in lodgepole pine forests.. *The Mountain Pine Beetle: A Synthesis of Its Biology,*
533 *Management and Impacts on Lodgepole Pine*, Pp. 3–66. doi:10.1016/j.giec.2010.09.011.
- 534 Six, D., and Paine, T. 1998. Effects of mycangial fungi and host tree species on progeny survival
535 and emergence of *Dendroctonus ponderosae*. *Environmental Entomology*, **27**:1392–1401.
- 536 Tauber, M.J., and Tauber, C.A. 1976. Insect seasonality: Diapause maintenance, termination, and
537 postdiapause development. *Annual Review of Entomology*, **21**:81-107.
538 doi:10.1146/annurev.en.21.010176.000501.
- 539 Taylor, F. 1981. Ecology and evolution of physiological time in insects. *The American Naturalist*,
540 **117**:1–23. doi:10.1086/283683.
- 541 Weed, A.S., Bentz, B.J., Ayers, M.P., and Holmes, T.P. 2015. Geographically variable response
542 of *Dendroctonus ponderosae* to winter warming in the western United States. *Landscape*

543 Ecology, **30**:1075–1093. doi:10.1007/s10980-015-0170-z.

544

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545 **Figure Captions:**

546 Figure 1: Proportion mortality of southern population mountain pine beetle lifestages in phloem
547 sandwiches at a range of constant temperatures (°C) throughout the experimental period.

548 Figure 2: Comparison of developmental deviance for individual eggs and first, second and third
549 instars in transfer treatments (i.e., transferred between 10 and 25 °C) versus a constant
550 temperature control at 10 °C. Based on a Wilcoxon Rank-Sum test, transferred individuals
551 developed significantly differently than individuals at a constant 10 °C in eggs ($W = 2100$, P
552 <0.005), and first ($W = 684.5$, $P = 0.005$) and third ($W = 353$, $P = 0.005$) instars. Boxes
553 represent the third and first quartile (25th and 75th percentiles), whiskers extend up to 1.5 times
554 the interquartile range from the top (bottom) of the box to the furthest data point, and the midline
555 is the median.

556 Figure 3. Model-predicted and observed lifestage-specific developmental rates for a southern
557 mountain pine beetle population based on constant temperature and censored data. ‘Censored’
558 data represent individuals that did not complete the lifestage at a given temperature. Data for the
559 fourth instar includes prepupal rates. All point sizes are on a log scale, with larger points
560 corresponding to more highly repeated observations. Dashed lines are ± 1 sigma, the variance
561 parameter associated with model fit. Note differences among plots in y-axis scale.

562 Figure 4. Model-predicted and observed lifestage-specific developmental rates for a southern
563 mountain pine beetle population based on constant temperature and transfer data. ‘Transfer’ data
564 represent individuals that were transferred between the treatment temperature and a constant
565 25 °C. Data for the fourth instar includes prepupal rates. All point sizes are on a log scale, with

566 larger points corresponding to more highly repeated observations. Dashed lines are ± 1 sigma, the
567 variance parameter associated with model fit. Note differences among plots in y-axis scale.

568 Figure 5. Comparison of predicted development rates of northern and southern mountain pine
569 beetle populations. Southern population predictions were based on estimated parameters in the
570 current study using non-transferred constant temperature and censored data (see Figure 3).
571 Northern population predictions were published in Régnière et al. (2012; Figure 4), also using
572 non-transferred constant temperature and censored data. Estimated upper (T_m) and lower (T_b)
573 development thresholds for each population and lifestage are in Table 1. Also shown are the
574 observed median development rates (\pm SD) of each population at the common experimental
575 temperatures (see Table 3 for observed development times).

576 Table 1: Lifestage-specific parameters for the rate curve (equation 2) for a southern mountain
 577 pine beetle population using constant temperature data and censored data (see Figure 3). Data
 578 from transfer treatments was excluded from the model fits for eggs, first instar, third instar, and
 579 fourth instar. T_b and T_m are lower and upper thresholds for development, Δ_b and Δ_m are the
 580 width of thermal transitions from normal to negligible development at the lower and upper
 581 thresholds, ω describes the expected exponential acceleration of rate with temperature, ψ is
 582 proportional to the maximum development rate, and σ is the variance.

Parameter	Eggs	First Instar	Second Instar	Third Instar	Fourth Instar	Pupae
ψ	0.0326	0.0521	0.0431	0.017	0.0545	0.0166
ω	0.2045	0.1517	0.1374	0.1856	0.1694	0.1658
T_b	6.0251	4.6029	5.9791	6.0115	14.9999	6.3504
T_m	31.9309	31.7661	31.8337	31.2656	31.4364	30.8041
Δ_b	0.5410	0.0117	0.0413	0.0	0.0	0.0
Δ_m	5.5031	5.4256	4.4534	4.3079	5.2947	3.5426
σ	0.038	-0.2182	0.1524	0.1650	0.1354	0.0673

583

584

585 Table 2: Lifestage-specific parameters for the rate curve (equation 2) for a southern mountain
 586 pine beetle population including constant temperature data and data from transfer treatments (see
 587 Figure 4). Second instar parameters are equivalent to those in Table 1 because there were no
 588 differences between transfer and constant temperature treatments. T_b and T_m are lower and upper
 589 thresholds for development, Δ_b and Δ_m are the width of thermal transitions from normal to
 590 negligible development at the lower and upper thresholds, ω describes the expected exponential
 591 acceleration of rate with temperature, ψ is proportional to the maximum development rate, and σ
 592 is the variance.

Parameter	Eggs	First Instar	Second Instar	Third Instar	Fourth Instar	Pupae
Ψ	0.0306	0.0433	0.0431	0.0417	0.0044	0.0179
Ω	0.1914	0.1965	0.1374	0.1406	0.2791	0.1494
T_b	6.5976	4.6428	5.9791	4.3248	10.0	5.6187
T_m	31.8135	31.7810	31.8337	31.6171	31.4294	30.7638
Δ_b	0.9239	1.0369	0.0413	1.3224	0.1478	0.2520
Δ_m	5.5441	5.3945	4.4534	6.2150	3.2194	3.1587
σ	0.0349	0.1938	0.1524	0.1411	0.1139	0.0726

593

594 Table 3: Comparison of median development time (days) for southern (this study) and northern
 595 (Régnière *et al.* 2012) mountain pine beetle populations at a constant 10, 20, 25 and 27 °C.
 596 Shown are the median days, standard deviation (SD), sample size (N) and test results (z) using a
 597 generalized linear model with a Tukey's post-hoc multiple comparison (Adjusted *P*) testing for
 598 differences between populations. No individuals in either population completed instar 4 or pupal
 599 development at a constant 10 °C.

Lifestage	Northern Days (SD)	Southern Days (SD)	z	Adj <i>P</i> > z
10°C				
Egg	29 (1.6) N = 20	17 (5.2) N = 32	-7.80	< 0.0001
First Instar	15 (2.9) N = 18	14 (4.1) N = 36	-1.55	1.0
Second Instar	33 (49.3) N = 13	18 (29.6) N = 27	-16.13	< 0.0001
Third Instar	63 (37.6) N = 9	166 (64.9) N = 7	17.77	< 0.0001
Fourth Instar	NA	NA	NA	NA
Pupa	NA	NA	NA	NA
20°C				
Egg	7 (0.8) N = 32	6 (0.9) N = 81	-2.54	0.9355
First Instar	3 (0.7) N = 19	4 (1.6) N = 69	1.87	0.9998
Second Instar	4 (2.0) N = 36	4 (1.2) N = 61	-0.09	1.0
Third Instar	6 (2.3) N = 46	6 (1.4) N = 60	-1.14	1.0
Fourth Instar	11 (3.5) N = 61	14 (2.5) N = 68	3.48	0.2506
Pupa	6 (1.1) N = 91	7 (0.5) N = 67	2.75	0.8326
25°C				
Egg	6 (0.6) N = 28	5 (0.7) N = 141	-1.56	1.0
First Instar	3 (0.8) N = 17	3 (2.7) N = 119	1.21	1.0
Second Instar	4 (1.4) N = 11	3 (3.1) N = 107	-1.91	0.9997
Third Instar	7 (3.3) N = 9	5 (1.9) N = 110	-1.48	0.9999
Fourth Instar	7 (4.7) N = 19	12 (6.3) N = 112	5.48	< 0.0001

Pupa	5 (1.4) N = 9	5 (0.5) N = 99	0.28	1.0
<i>27°C</i>				
Egg	6 (0.5) N = 22	4 (0.9) N = 73	-2.80	0.8007
First Instar	4 (1.1) N = 21	2 (2.1) N = 63	-2.39	0.9741
Second Instar	7 (3.7) N = 16	3 (3.7) N = 61	-5.42	< 0.0001
Third Instar	17 (9.3) N = 6	5 (2.4) N = 59	-11.50	< 0.0001
Fourth Instar	NA	14 (3.6) N = 54	NA	NA
Pupa	NA	5 (0.5) N = 42	NA	NA

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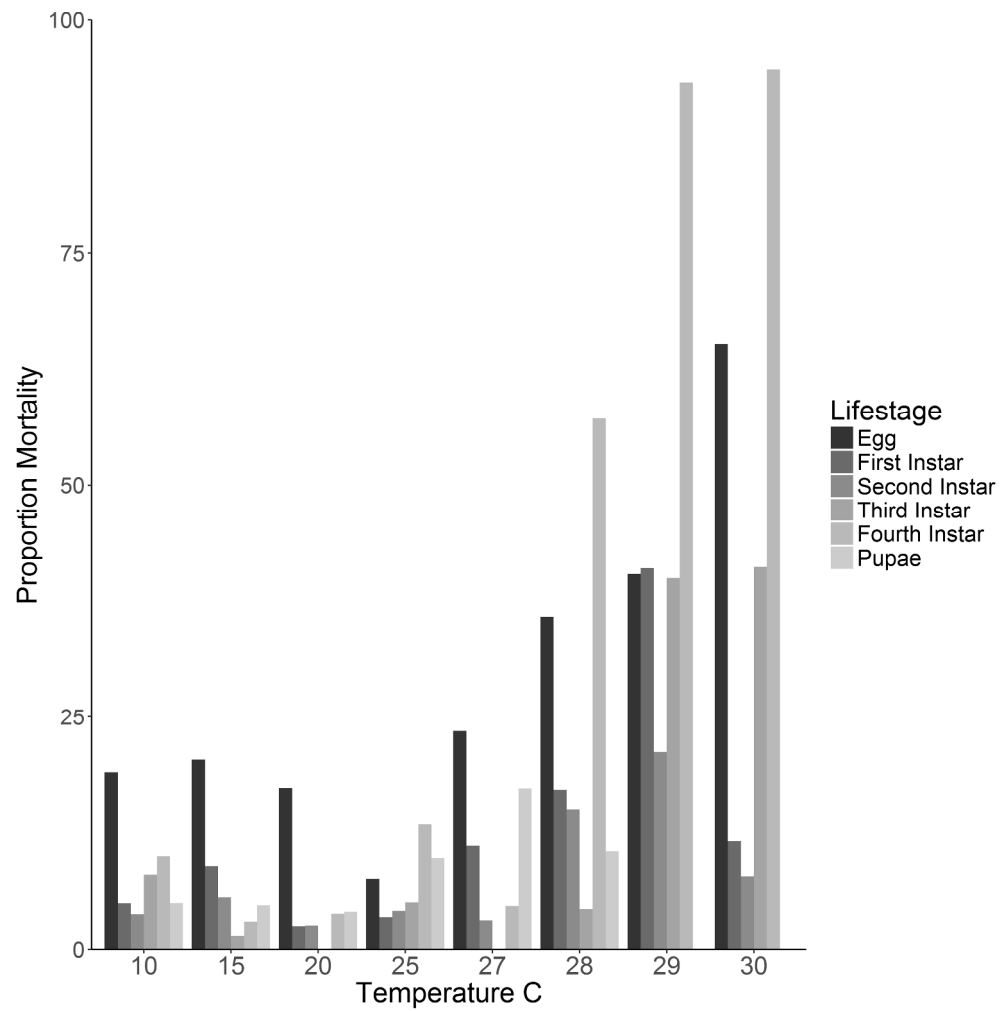


Figure 1: Proportion mortality of southern population mountain pine beetle lifestages in phloem sandwiches at a range of constant temperatures (°C) throughout the experimental period.

254x254mm (300 x 300 DPI)

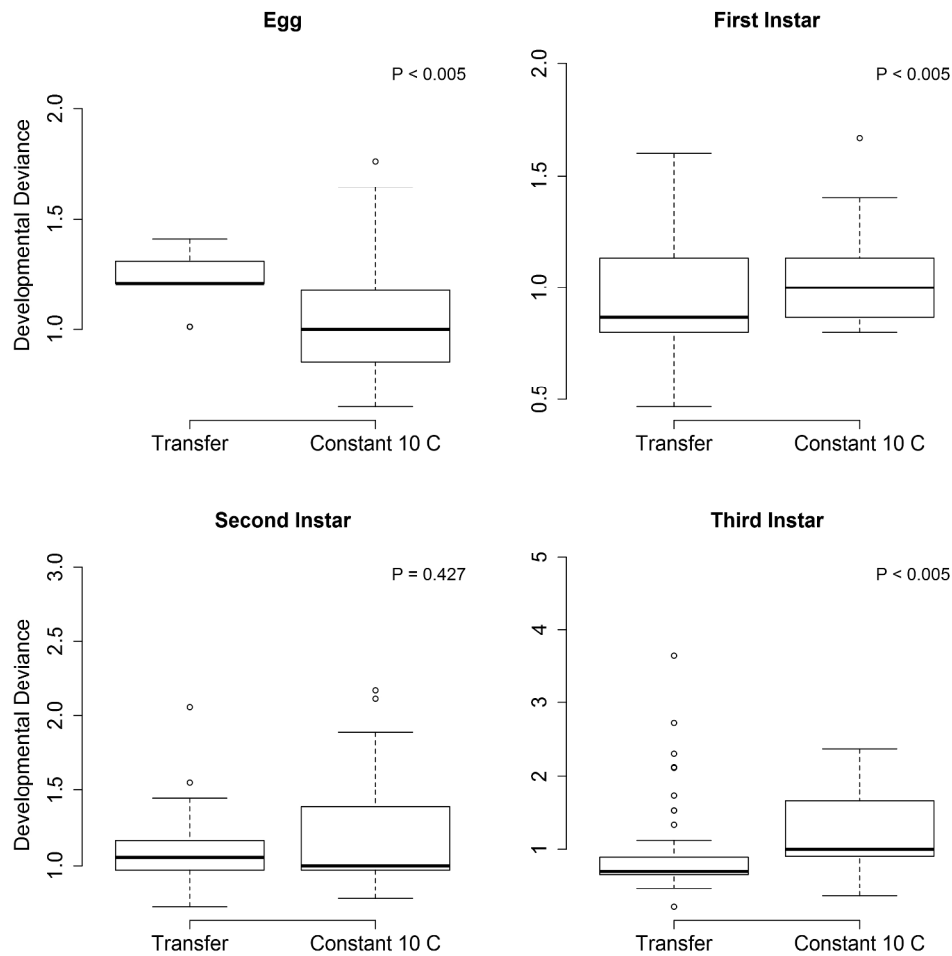


Figure 2: Comparison of developmental deviance for individual eggs and first, second and third instars in transfer treatments (i.e., transferred between 10 and 25 °C) versus a constant temperature control at 10 °C. Based on a Wilcoxon Rank-Sum test, transferred individuals developed significantly differently than individuals at a constant 10 °C in eggs ($W = 2100$, $P < 0.005$), and first ($W = 684.5$, $P = 0.005$) and third ($W = 353$, $P = 0.005$) instars. Boxes represent the third and first quartile (25th and 75th percentiles), whiskers extend up to 1.5 times the interquartile range from the top (bottom) of the box to the furthest data point, and the midline is the median.

254x254mm (300 x 300 DPI)

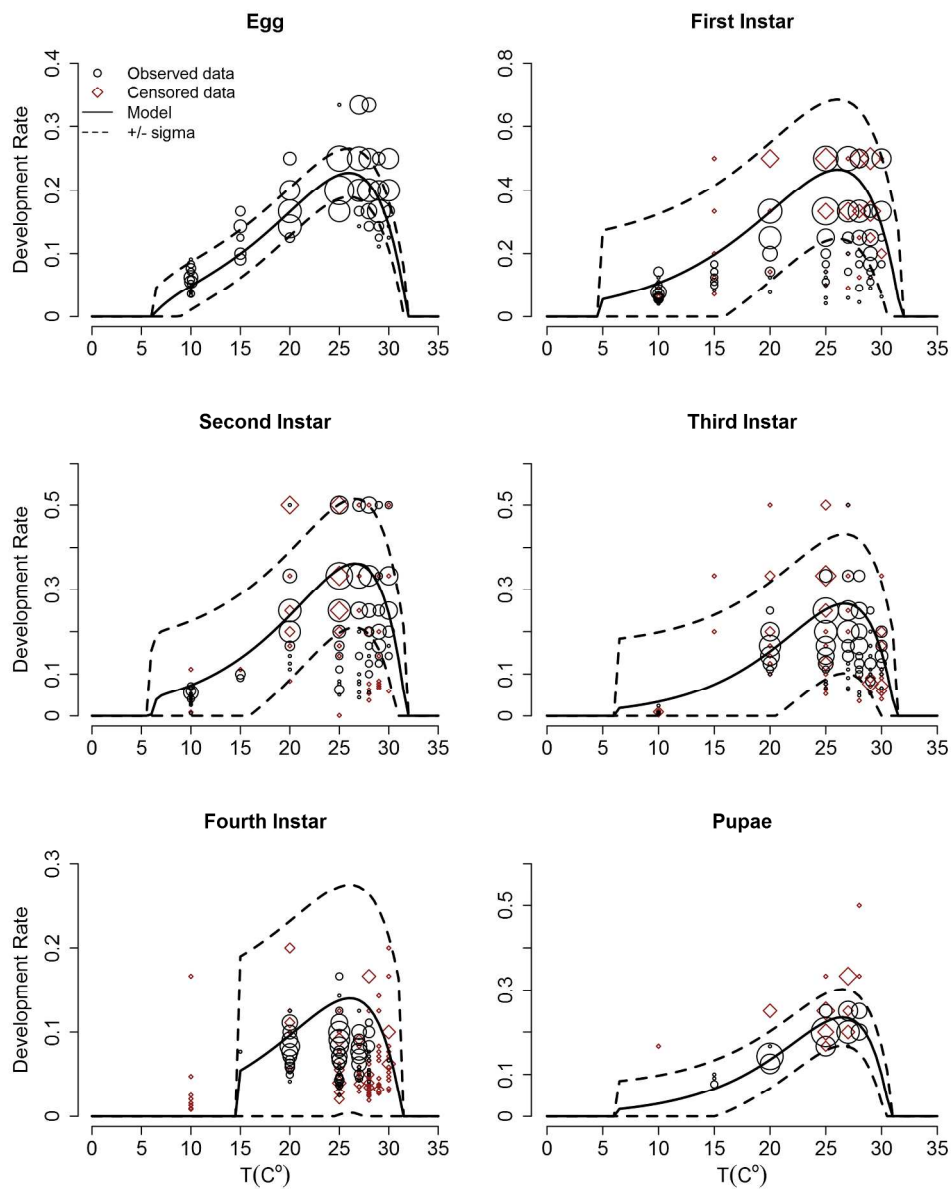


Figure 3. Model-predicted and observed lifestage-specific developmental rates for a southern mountain pine beetle population based on constant temperature and censored data. 'Censored' data represent individuals that did not complete the lifestage at a given temperature. Data for the fourth instar includes prepupal rates. All point sizes are on a log scale, with larger points corresponding to more highly repeated observations. Dashed lines are ± 1 sigma, the variance parameter associated with model fit. Note differences among plots in y-axis scale.

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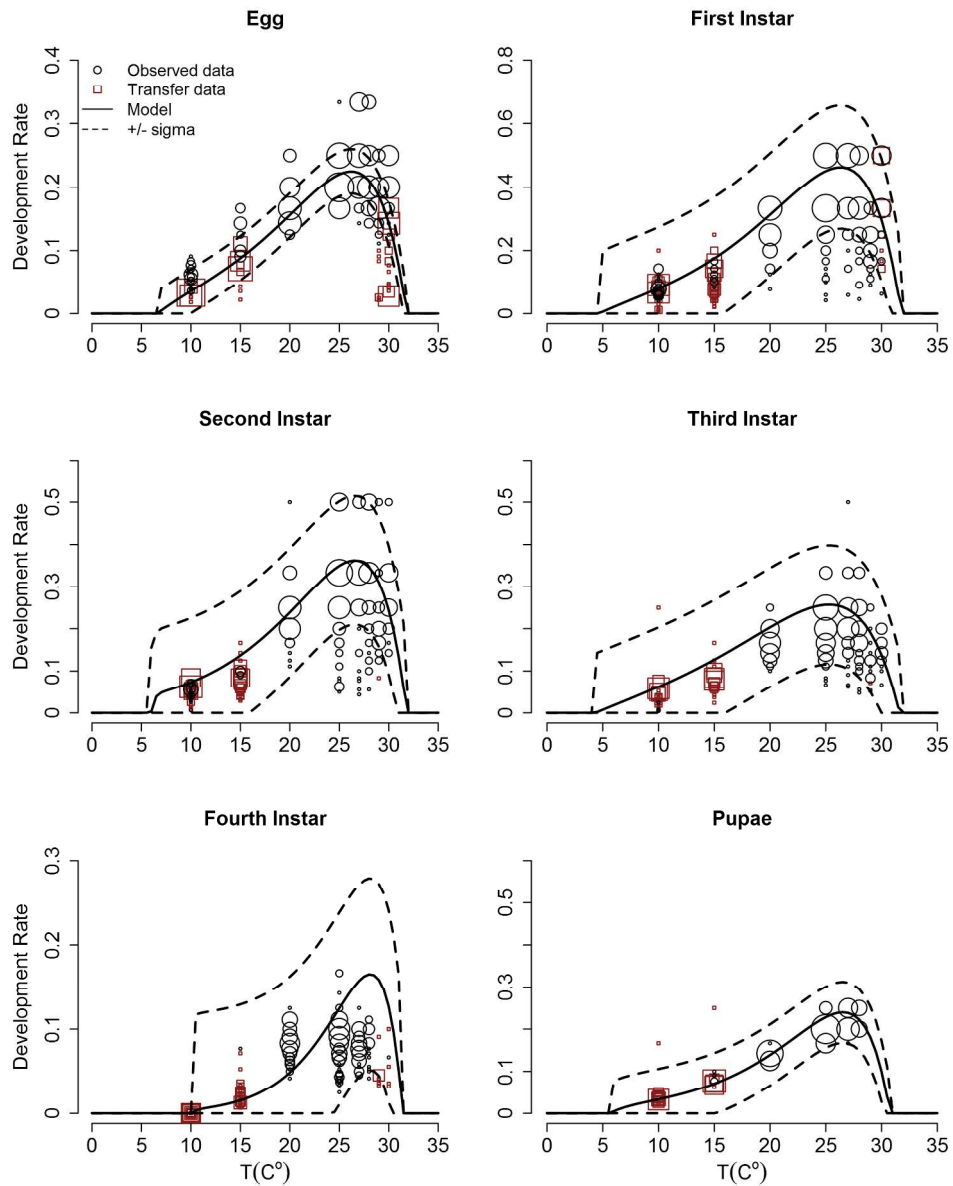


Figure 4. Model-predicted and observed lifestage-specific developmental rates for a southern mountain pine beetle population based on constant temperature and transfer data. 'Transfer' data represent individuals that were transferred between the treatment temperature and a constant 25 °C. Data for the fourth instar includes prepupal rates. All point sizes are on a log scale, with larger points corresponding to more highly repeated observations. Dashed lines are ± 1 sigma, the variance parameter associated with model fit. Note differences among plots in y-axis scale.

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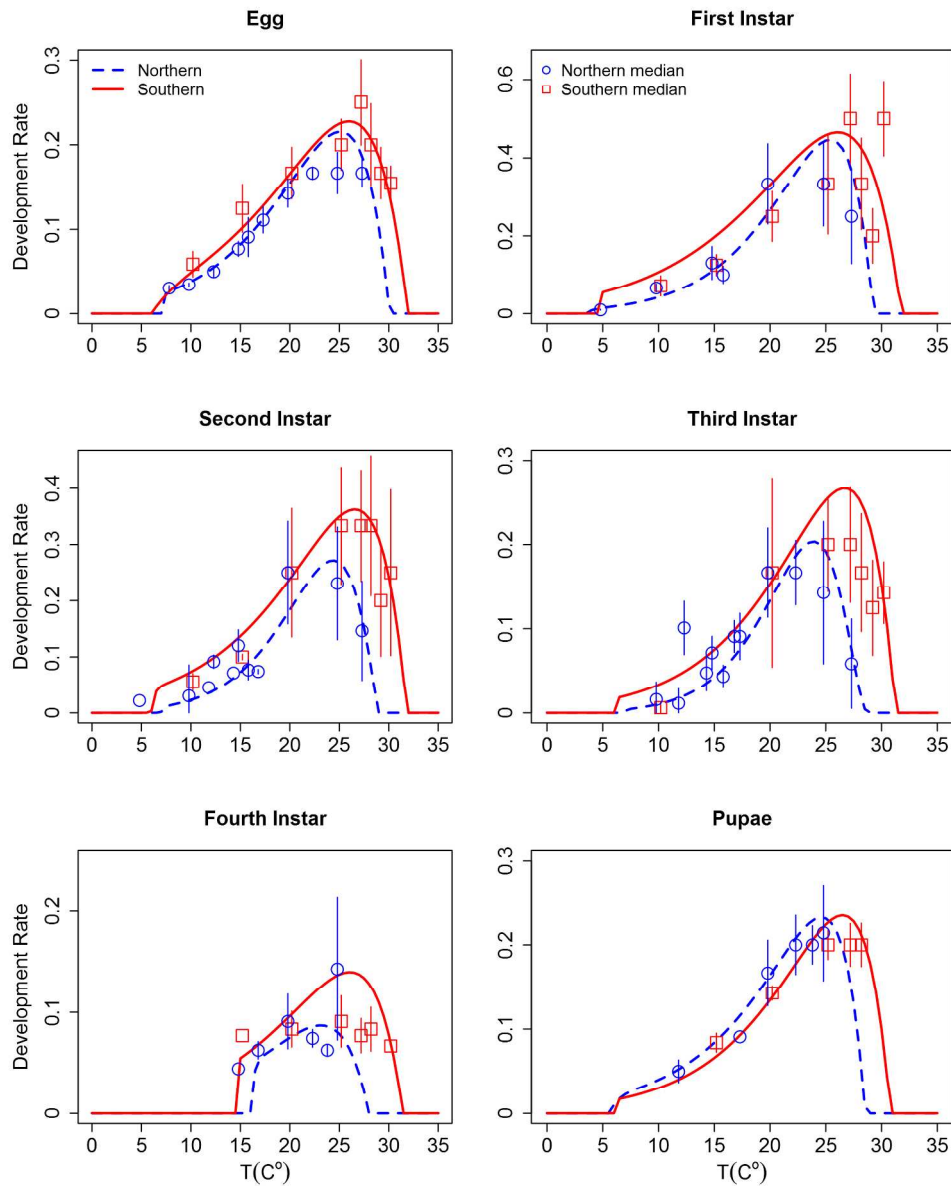


Figure 5. Comparison of predicted development rates of northern and southern mountain pine beetle populations. Southern population predictions were based on estimated parameters in the current study using non-transferred constant temperature and censored data (see Figure 3). Northern population predictions were published in Régnière et al. (2012; Figure 4), also using non-transferred constant temperature and censored data. Estimated upper (T_m) and lower (T_b) development thresholds for each population and lifestage are in Table 1. Also shown are the observed median development rates (\pm SD) of each population at the common experimental temperatures (see Table 3 for observed development times).

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