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The neural tides of sleep and consciousness revealed by single-pulse electrical brain stimulation

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Abstract

Wakefulness and sleep arise from global changes in brain physiology that may also govern the flow of neural activity between cortical regions responsible for perceptual processing vs planning and action. To test whether and how the sleep/wake cycle affects the overall propagation of neural activity in large-scale brain networks, we applied single-pulse electrical stimulation (SPES) in patients implanted with intracranial EEG electrodes for epilepsy surgery. SPES elicited cortico-cortical spectral responses at high-gamma frequencies (CCSRHG, 80-150 Hz). which indexes changes in neuronal population firing rates. Using event-related causality analysis (ERC), we found that the overall patterns of neural propagation among sites with CCSRHG were different during wakefulness and different sleep stages. For example, stimulation of frontal lobe elicited greater propagation toward parietal lobe during slow wave sleep than during wakefulness. During REM sleep, we observed a decrease in propagation within frontal lobe, and an increase in propagation within parietal lobe, elicited by frontal and parietal stimulation, respectively. These biases in the directionality of large-scale cortical network dynamics during REM sleep could potentially account for some of the unique experiential aspects of this sleep stage. Together these findings suggest that the regulation of conscious awareness and sleep is associated with differences in the balance of neural propagation across large-scale frontalparietal networks.

Key words: brain waves, human electrocorticography, high-gamma activity, effective connectivity, causal interactions

Statement of Significance

While awake, different areas of the brain are involved in perception of the environment versus planning and organizing behavior, but they are in constant communication. While asleep, the brain processes information gathered during wakefulness, but how different brain areas interact during sleep, is poorly understood. To elucidate these interactions, we activated the brain with weak electrical pulses during wakefulness and sleep, and we analyzed how this activation spread through the brain. We observed greater propagation of activity from frontal to parietal lobe during slow wave sleep, and decreased propagation within frontal lobe, but increased propagation within parietal lobe, during REM sleep. These suggest that wakefulness and sleep are associated with different patterns of propagation of neural activity across brain networks.

Introduction

The profound neurophysiological changes that occur during the sleep-wake cycle may modify the overall direction by which neural activity is propagated among various areas. For example, previous studies have shown that low frequency oscillations characteristic of slow wave, or non-REM, sleep tend to travel in an anteroposterior direction. In contrast, this flow may reverse during wakefulness. Reminiscent of the tides of the ocean, circadian rhythms in the brain may be accompanied by cyclical variations in the kind and strength of neural interactions across large-scale brain networks. However, little is known whether these changes in neural interactions can only be observed at low frequencies, and how they are linked to brain function. Modulation of causal relationships between neural networks may be one of the mechanisms that the human brain employs, not only for efficient processing of information, but also for regulation of sleep and wakefulness. Furthermore, if significant differences exist in the predominant direction of flows in non-REM sleep, in which we are usually unconscious, versus wakefulness, these differences may reflect mechanisms regulating conscious awareness.

To address these questions, we retrospectively analyzed the data obtained in our previous study. We applied single-pulse electrical stimulation (SPES) through subdural electrodes that were implanted for presurgical evaluation in patients with intractable partial epilepsy, and we recorded electrophysiological responses to SPES in the rest of the implanted electrodes. SPES of the human cortex has been shown to evoke electrophysiological responses at distant sites. These responses may include cortico-cortical evoked potentials (CCEP), which consist of phase-locked responses revealed by averaging signals in the time domain, and cortico-cortical spectral responses (CCSRs), which are obtained by averaging in the frequency domain and are usually dominated, particularly at high frequencies, by non-phase-locked responses. Nowadays, many

researchers are using SPES to expand our understanding about human cortical networks and their input/output characteristics, often using CCEPs to estimate effective connectivity between stimulus sites and recording sites. ¹²⁻¹⁵ CCSRs, a term firstly introduced by Gkogkidis et al., ¹¹ occur at a variety of frequencies, including high-gamma (HG) bands. For example, Maliia et al. recently investigated high-gamma responses to SPES in human physiologic and pathological networks. ¹⁶ High-gamma activity is also widely used as an index of changes in neuronal population firing rates. ¹⁷⁻¹⁹ Thus, a CCSR^{HG} induced by SPES reflects changes in neuronal firing rates elicited by simulated external input, via direct or indirect anatomical connections with the stimulation site. ^{9,20-26} CCSR^{HG} generator mechanisms are still debated, ²⁷ but the changes in neural behavior reported in stimulation studies with single unit recordings or dye imaging are similar to those of CCSR^{HG}. ²⁸⁻³⁰ Because SPES is task-free and does not require patient cooperation, CCEP and CCSR recorded by SPES can be used to study how sleep alters the dynamic characteristics of human brain networks.

We used event-related causality (ERC) analysis of high-gamma activity induced by single-pulse electrical stimulation (SPES)³¹⁻³⁵ to estimate the magnitude and directionality of neural propagation within and between different brain lobes, and we compared the overall patterns of these induced neural propagations during wakefulness and different sleep stages. ERC analysis, a multichannel extension of the Granger causality concept,³⁶ has previously been used to study the dynamics of neural propagation during word production tasks,^{8,37-39} and to study both ictal and interictal activity in epileptogenic networks.⁴⁰ Here, we applied it to high-gamma activity induced by SPES. To the best of our knowledge, this is not only the first application of event-related causality analysis to stimulus-induced high-gamma activity, but as such, it is the

first study to measure the predominant directionality of internally generated neural propagation in human cortical networks during wakefulness and sleep.

Methods

This study is based largely on the dataset that were used in our previous studies. ^{9,41} The method of recording SPES data across wakefulness and sleep, sleep staging, and the anatomic localization of electrodes, are described in detail in our previous publications. ^{9,31,42} The outline of this study is depicted in **Figure 1**. We have determined sleep stages according to the standard Rechtchaffen & Kales (R&K) sleep staging criteria. ⁴³ Data analysis of high gamma activity and ERC analysis were performed using a custom data analysis pipeline (using Fortran77, C++, and R) developed at Johns Hopkins. All other analyses were performed using in-house scripts in Matlab software (version 2018a, MathWorks, Natick, MA).

Patients

Thirteen patients who underwent invasive presurgical evaluation for intractable partial epilepsy since February 2010 through October 2013 in Kyoto University Hospital provided written informed consent to the study. The protocol conformed to the Declaration of Helsinki and was approved by the Ethics Committee of the Institute (IRB#443). Among them, eight patients were eligible for this study (see **Table 1** for the patients' profiles) according to the criteria mentioned below in detail. The locations of the electrodes were confirmed by an MRI image after implantation, and they were linearly co-registered into a pre-implantation image. For the purpose of 3D display in figures in each patient, the gray matter segmentation was done for pre-

implantation image using Freesurfer software (http://surfer.nmr.mgh.harvard.edu/) and presented in FSL view (http://www.fmrib.ox.ac.uk.fsl/fslview).

Recording signals induced by SPES during sleep

During the daytime, direct electrical stimulation was applied in a bipolar manner to a pair of adjacently placed subdural platinum electrode (2.3 mm in diameter, 1 cm in interelectrode distance, AD-TECH, Wisconsin), using a constant-current stimulator (MS-120B/MEE-1232, Nihon Kohden, Tokyo, Japan). The single-pulse electrical stimuli (square wave pulse: 0.3-ms duration) were delivered in alternating polarity at a fixed frequency of 1 Hz. CCEPs obtained by stimulating the majority of the implanted electrodes during daytime were used for identifying the electrodes to be stimulated during sleep. One or two stimulation sites were selected for the nighttime CCEP recordings in each patient according to the following criteria: 1. sites away from the epileptic focus and 2. sites that produced discrete CCEP responses in the adjacent (local CCEP field) and/or distant (remote CCEP fields) regions. Thus, SPES was delivered to the cortical surface during wakefulness, light sleep, slow wave sleep, and REM sleep. In each recorded electrode of each stimulus sites, we calculated CCSRHGs. We used the highest intensity (maximum: 12 mA) at which 1. patients did not notice the stimulation and no symptoms were evoked, 2. the adjacent electrodes did not show excessive artifacts over a dynamic range that would prohibit recording, and 3. no afterdischarges were induced. Intensity was adjusted in increments of 1 or 2 mA. One stimulation set included 50-100 pulses. In general, 200-400 electrical pulses were delivered during each sleep stage that is judged on-line according to aforementioned sleep staging criteria. Electrocorticograms (ECoGs) were referenced to a scalp electrode placed on the skin over the mastoid process contralateral to the implantation site and

were sampled at 1000 Hz (in Subject 4 and 8) or 2000 Hz (in the other subjects) with a bandpass filter of 0.08–300 Hz or 0.08–600 Hz, respectively. ECoG signals were aligned to SPES (analysis window: –300 to +700 ms) offline. Examples of CCSR^{HG}s at two recording sites with corresponding CCEPs during different sleep stages are shown in **Supplementary Figure 1**.

Selection of the SPES-affected sites for ERC analysis

The following criteria were used for selecting eligible patients and sites (channels) to be analyzed with multivariate autoregressive modeling (MVAR). Finally, the patients in whom SPES was delivered to the cortical surface in frontal (7 sites in 6 patients in total) or parietal lobe (5 sites in 3 patients in total) across sleep stages were eligible for this study (**Table 1**). In one patient, SPES was delivered independently to frontal and parietal lobes.

First, for ERC analysis at least 50 trials were required, and the number of trials had to be the same across sleep stages in one patient. It reassured us that the analyses were not biased by different or insufficient numbers of trials. Trials were separated according to the stimulation polarity. Therefore, for each sleep stage at least 100 trials were used (50 of each polarity), and the first N trials (N = the least number of eligible trials across sleep stages) within each sleep stage were used for analysis, after removing trials with artifacts.

Second, we chose patients with midpoints of stimulation sites in frontal or parietal lobe to further investigate anterior-posterior interactions in sleeping brain. Patients in whom the pair of electrodes used for stimulation traversed central sulcus were excluded, because more than one lobe could be stimulated at the same time, which would lead to ambiguous results.

Third, to diminish the potential influence of pathological activities, we didn't include recordings from sites that were identified by certified clinicians as seizure onset zone, or containing frequent spikes (> 1/120 s), or that showed baseline shift, or artifacts.

Data sampled at 2000 Hz were down-sampled to 1000 Hz. Then, recorded signals were band-passed filtered at 70-170 Hz with a Butterworth filter of order 6. Next, the time-frequency energy distribution of SPES-induced activity, i.e., CCSR, at time intervals between +100 ms and +550 ms after stimulation were computed using matching pursuit (MP) algorithm. Haseline was set at -300 \sim -100 ms before stimulation. Data between -100 ms and +100 ms were excluded from analyses to avoid artifacts introduced by stimulation pulse. The sites that showed significant CCSR increase (between 80 and 150 Hz) (p < .05 by MP) in any sleep stage and in either stimulation polarity were included in ERC analyses.

We previously showed that SPES elicits CCSR^{HG} decrease right after stimulation at recording sites that had strong connectivity to the stimulation site (measured by CCEPs) and that these HG decreases are potentiated during sleep. ^{9,20} We excluded sites showing significantly decreased CCSR^{HG} immediately after stimulation. This reduced the risk of including sites with prolonged post-stimulus inhibition (high-gamma decrease), which would have affected the baseline.

ERC analysis

ERC method is a multichannel extension of Granger causality concept, which states that an observed time series x(t) causes another series y(t), if knowledge of x(t)'s past significantly improves prediction of y(t). ERC gives an estimate of changes in the intensity and direction of neural activity propagation between recording sites as a function of frequency, in comparison to

baseline activity. A greater ERC value $_{X \to Y}$ infers that high-gamma activity from site X causally influences site Y by Granger criteria. 8,37 To detect propagation between two sites, the ERC method analyzes the similarity of signals recorded at both sites and the time delays between them (for simplicity, one can think about delayed, or lagged, correlations). If high-gamma power is suppressed in one of the sites after SPES, it can reflect a decrease of neural input to the site, or inhibition of local activity caused by an increase of neural input to the site. As it operates on the activity of an entire neuronal population, the ERC method cannot distinguish between these two situations. We think that it is unlikely that SPES-induced high-gamma power suppression would be propagated to other sites, and that if this were to happen at all, it would occur early in our analysis window. For these reasons, we excluded sites in which SPES induced high-gamma power suppression.

For each dataset MVAR model order was determined using Akaike Information Criterion (AIC), and the coefficients were computed for 20 ms shifting window, overlapped by 5 ms over 650 ms trial (including baseline -300 to -100 ms and response period +100 to +550 ms). For each set of trials with the same stimulation polarity in each site, ERC values of post-stimulus period were compared to baseline epoch.⁸

Statistical group analysis

In each patient, all ERC values were normalized to the highest ERC value, resulting in the highest value of ERC being 1, which allowed averaging over patients. Normalized ERC values reaching significance (p < .05, Bonferroni corrected) for flows within and between three regions of interest (network nodes); frontal, parietal, and temporal-occipital, were pooled across patients, in frontal or parietal stimulation group separately, and used as an index of directed interaction

between the nodes (**Supplementary Figure 1** shows the examples of ERC values calculated between two electrodes). Our previous study did not show the difference of propensity of CCEP/CCSRs between different stimulation intensity. Therefore ERC results obtained for different stimulation intensities (4-12 mA) were pooled together in this study (see **Table 1**).

Our null hypothesis is that no difference in high-gamma propagation patterns is found between sleep stages. We used nonparametric permutation test for statistical analysis. First, we randomly shuffled ERC values between four sleep stages within each directed connection. Second, paired t-test between any two sleep stages was applied across time points (100-500 ms) and selected all the samples where the t-value satisfied p < .01 lasting for 20 ms or more. Third, we calculated the sum of the t-values in each cluster and made cluster-level statistics. Lastly, only the highest statistic from all the comparisons between any two sleep stages was taken. This procedure was repeated 10,000 times and the null distribution of the maximum cluster-level statistics was compared with the original cluster-level statistics in each propagation, respectively. A p-value threshold was defined < .05. See **Supplementary Figure 2** for an illustrative pipeline of the analysis.

Results

The ERC results of eight patients were combined after normalization of data (see **Table 1** for the patients' profile and **Figure 2** for electrodes that were included for analysis) and statistically analyzed. Significant SPES-induced changes in the propagation of high-gamma activity within and between regions of interest, were compared between sleep stages (**Figure 3** and **Figure 4**).

Changes in the pattern of induced neural propagation during slow wave sleep

Stimulation of frontal lobe during slow wave sleep, induced greater propagation from frontal to parietal lobe (F \rightarrow P, 245-280 ms, p < .05 by permutation test), as compared to the stimulation during wakefulness (**Figure 3**. Examples in which we didn't observe any significant ERC values between wakefulness and other sleep stages in F \rightarrow P are shown in **Supplementary Figure 3**.). No other significant differences in the patterns of neural propagation were found between slow wave sleep and wakefulness.

The results of stimulating parietal lobe during slow wave sleep were not significantly different from those during wakefulness.

Changes in the pattern of evoked neural interactions during REM sleep

Compared to wakefulness, frontal stimulation during REM sleep induced less high-gamma propagation within frontal lobe (at 110-140 ms, **Figure 4 top**). During wakefulness SPES induced immediate propagation within frontal network, while the frontal network did not exhibit a significant increase in propagation during REM. No other significant changes in neural propagation were found between REM and wakefulness.

Compared to light sleep, parietal stimulation during REM sleep resulted in greater propagation within parietal lobe (at 435-460 ms, **Figure 4 bottom**). Interestingly, no significant difference in propagation was found when compared to wakefulness. Overall, these findings suggest that the frontal network is "mute" during REM, while the parietal network is active.

Discussion

We observed changes in the overall pattern of neural propagation across large-scale human cortical networks when SPES was applied to frontal or parietal lobe during wakefulness and

sleep stages. We found that different sleep stages affected the overall pattern of neural propagation among brain network. For example, stimulation of frontal lobe elicited larger early propagation from frontal to parietal lobe $(F \rightarrow P)$ during slow wave sleep as compared to wakefulness (Figure 3). This result is in line with previous studies of passive scalp EEG showing propagation of low frequency activity (< 30 Hz) from anterior to posterior regions during slow wave sleep, 1-3 and extends those findings to high-gamma propagation. It is important to note that high-gamma activity has been shown to reflect the overall firing rates in neural populations. 19 Thus, our findings suggest a natural brain tide, reflecting a bias in propagation between frontal and parietal neuronal populations, changing according to sleep stage. During slow wave sleep, slow wave activities originate in the frontal lobe and tend to travel posteriorly¹-^{3,46} although they are partly produced and regulated locally.^{6,47-49} Source modeling of slow wave activity using a high-density EEG showed that this flow is mediated by a cingulate highway.³ It is difficult to directly relate our results to slow wave activity itself. However, our results support the notion that in slow wave sleep, the readout from processing in the frontal lobe is conveyed in a top-down direction to the parietal lobe. This may facilitate the modification of synaptic strengths to consolidate processed information.^{50,51} In frontal lobe epilepsy, seizures occur predominantly during slow wave sleep. 52,53 Propagation of neuronal activity from frontal lobe to parietal lobe during slow wave sleep, as observed here, may also facilitate seizure propagation in this sleep stage in frontal lobe epilepsy.

On the other hand, when we are awake, information from the periphery is largely processed in a bottom-up manner, and is encoded in widespread cortical regions.⁶ REM sleep is similar to wakefulness in terms of its cortico-cortical connectivity, and cortical excitability.^{9,54} Indeed, humans may be partially conscious during the dreams of REM sleep.⁵⁵ However, there

are clear experiential differences between wakefulness and REM sleep, and previous reports have attempted to account for these differences. For example, a positron emission tomography study showed decreased hypometabolism in frontal association areas including lateral orbital and dorsolateral prefrontal cortices during REM sleep. ⁵⁶ In the present study, we observed a reduction in early SPES-induced neural propagations within the frontal lobe during REM sleep (**Figure 4**). Consistent with the PET study, this suggests that few active processes are running in frontal cortex. Moreover, we observed enhanced network interactions within parietal lobe during REM sleep. Together, these findings may explain the complex and realistic egocentric experiences that characterize dreaming REM sleep, as well as some of their bizarre characteristics, perhaps owing to the lack of top down executive control on these experiences.

In previous studies, we and other researchers have observed inhibition of high-gamma activity immediately after stimulation, and decreased phase-locked activity between low-gamma and stimulation-induced slow waves during slow wave sleep, 9.23 suggesting that loss of information integration leads to unconsciousness during slow wave sleep, based on information integration theory. 9.23,58 However, our present results suggest that causal interactions could occur across "frontal-parietal" networks 9 during sleep. Siclari et al. recently found evidence for "hot zones" in the posterior human brain that when activated, elicit dream-like experiences, whether during non-REM or REM sleep. 55 They did not analyze how neuronal activity propagated from the hot zones, though. Humans are unconsciousness and/or amnestic during slow wave sleep. Based on our findings in the present study, we speculate that the propagation of neural processing from frontal to parietal lobes, observed to be greater during slow wave sleep than during wakefulness, reflects processes that are either incompatible with conscious awareness or

simply do not support it. However, the mechanisms responsible for this difference in the overall tides of neural propagation are unknown.

It would be noteworthy to compare the SPES-induced high-gamma propagation to our CCEP results. However, the two techniques are more complementary than comparable. The first sharp negative deflection of CCEP, called the N1, at latencies of 10-50 ms, is believed to reflect direct cortico-cortical connection. It was impossible to analyze SPES-induced high-gamma propagation earlier than 100 ms after stimulation in this study, because the MVAR model used by ERC method requires stationarity of the modeled process, while SPES introduces a strong perturbation. With this in mind, our analysis focused on connectivity among cortical networks involved in SPES-induced activities although we often observe CCSR^{HG} decrease early after stimulation. A more sophisticated method is required in the future to look at induced activities earlier after stimulation.

A few more concerns need to be addressed in this study. First, all our participants were patients with intractable partial epilepsy. Furthermore, they were all treated with different regimens of anti-seizure drugs, and it has recently been reported that some of these drugs can influence TMS-evoked EEG responses in healthy subjects. Both confounds could have influenced our results independently. Second, we did not define sleep stages throughout all sleep, but only during SPES sessions. Comparisons between ERC with and without SPES during particular sleep stages may be considered in future analyses but are beyond the scope of this report. Lastly, to reduce the risk of our baseline being affected by preceding SPESs at 1 second intervals, we excluded sites within the seizure onset zone, in which stronger and more prolonged inhibition of high-gamma can sometimes be elicited by SPES. We also excluded sites with frequent epileptiform discharges. Nevertheless, previous studies have failed to show significant

differences in CCEPs and CCSRs for SPES at 1 Hz vs. 0.5 Hz.^{12,14,23} The duration of interstimulus intervals for SPES is constrained by the amount of time available for testing during sleep. Establishing an efficient and patient-friendly SPES protocol during sleep is an important goal for future studies.

Herein we studied, for the first time, the strength and directionality of neural propagation, estimated with event-related causality analyses, elicited among different brain regions in response to external input delivered artificially during wakefulness and various sleep stages. We observed differences between interlobar neural propagations elicited during wakefulness and different sleep stages. Although slow wave sleep is believed to be accompanied by a breakdown of connectivity across cortex, we observed greater propagation from frontal lobe to parietal lobe during this sleep stage than during wakefulness, suggesting that this pattern of network interaction is either counterproductive to conscious awareness or reflects the absence of bottomup input to executive areas in frontal lobe from perceptual and integrative areas in parietal lobe. Moreover, during REM sleep, we observed a decrease in propagation within frontal lobe, and an increase in propagation within parietal lobe, relative to wakefulness and light sleep, respectively. These differences could potentially account for some of the unique experiential aspects of this sleep stage. Together our findings suggest that the regulation of conscious awareness and sleep is associated with differences in the balance of neural propagation across large-scale frontalparietal networks. Our results may help to understand the human physiology of sleep in terms of directed neural propagation among large scale cortical networks, and could help recast the pathophysiological mechanisms governing sleep-related behavioral disorders and the effects of sleep on epilepsy in these terms. Further investigations of causal interaction across large-scale

cortical networks during wakefulness and different sleep stages are needed, including analyses of the effects of lesions, neurobehavioral disorders, and sleep disorders on these interactions.

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References

- Kaminski M, Blinowska K, Szclenberger W. Topographic analysis of coherence and propagation of EEG activity during sleep and wakefulness. Electroencephalogr Clin Neurophysiol. 1997; 102 (3): 216-227.
- 2. Massimini M, Huber R, Ferrarelli F, Hill S, Tononi G. The sleep slow oscillation as a traveling wave. J Neurosci. 2004; 24 (31): 6862-6870.
- 3. Murphy M, Riedner BA, Huber R, Massimini M, Ferrarelli F, Tononi G. Source modeling sleep slow waves. Proc Natl Acad Sci U S A. 2009; 106 (5): 1608-1613.
- 4. Diekelmann S, Born J. The memory function of sleep. Nat Rev Neurosci. 2010; 11 (2): 114-126.
- 5. Maquet P. The role of sleep in learning and memory. Science. 2001; 294 (5544): 1048-1052.
- 6. Tononi G, Cirelli C. Sleep and the price of plasticity: from synaptic and cellular homeostasis to memory consolidation and integration. Neuron. 2014; 81 (1): 12-34.
- 7. Bassett DS, Sporns O. Network neuroscience. Nat Neurosci. 2017; 20 (3): 353-364.
- 8. Korzeniewska A, Crainiceanu CM, Kus R, Franaszczuk PJ, Crone NE. Dynamics of event-related causality in brain electrical activity. Hum Brain Mapp. 2008; 29 (10): 1170-1192.
- 9. Usami K, Matsumoto R, Kobayashi K, et al. Sleep modulates cortical connectivity and excitability in humans: Direct evidence from neural activity induced by single-pulse electrical stimulation. Hum Brain Mapp. 2015; 36 (11): 4714-4729.
- 10. Matsumoto R, Kunieda T, Nair D. Single pulse electrical stimulation to probe functional and pathological connectivity in epilepsy. Seizure. 2017; 44: 27-36.
- 11. Gkogkidis CA, Wang X, Schubert T, et al. Closed-loop interaction with the cerebral cortex using a novel micro-ECoG-based implant: the impact of beta vs. gamma stimulation

- frequencies on cortico-cortical spectral responses. Brain-Computer Interfaces. 2017; 4 (4): 214-224.
- 12. Entz L, Toth E, Keller CJ, et al. Evoked effective connectivity of the human neocortex. Hum Brain Mapp. 2014; 35 (12): 5736-5753.
- 13. Keller CJ, Bickel S, Entz L, et al. Intrinsic functional architecture predicts electrically evoked responses in the human brain. Proc Natl Acad Sci U S A. 2011; 108 (25): 10308-10313.
- 14. Keller CJ, Honey CJ, Entz L, et al. Corticocortical evoked potentials reveal projectors and integrators in human brain networks. J Neurosci. 2014; 34 (27): 9152-9163.
- 15. Keller CJ, Honey CJ, Megevand P, Entz L, Ulbert I, Mehta AD. Mapping human brain networks with cortico-cortical evoked potentials. Philos Trans R Soc Lond B Biol Sci. 2014; 369 (1653).
- 16. Maliia MD, Donos C, Barborica A, et al. High frequency spectral changes induced by single-pulse electric stimulation: Comparison between physiologic and pathologic networks. Clin Neurophysiol. 2017; 128 (6): 1053-1060.
- 17. Buzsaki G, Wang XJ. Mechanisms of gamma oscillations. Annu Rev Neurosci. 2012; 35: 203-225.
- 18. Crone NE, Korzeniewska A, Franaszczuk PJ. Cortical gamma responses: searching high and low. Int J Psychophysiol. 2011; 79 (1): 9-15.
- 19. Ray S, Crone NE, Niebur E, Franaszczuk PJ, Hsiao SS. Neural correlates of high-gamma oscillations (60-200 Hz) in macaque local field potentials and their potential implications in electrocorticography. J Neurosci. 2008; 28 (45): 11526-11536.
- 20. Kobayashi K, Matsumoto R, Matsuhashi M, et al. Different Mode of Afferents Determines the Frequency Range of High Frequency Activities in the Human Brain: Direct

- Electrocorticographic Comparison between Peripheral Nerve and Direct Cortical Stimulation. PLoS One. 2015; 10 (6): e0130461.
- 21. Kobayashi K, Matsumoto R, Matsuhashi M, et al. High frequency activity overriding cortico-cortical evoked potentials reflects altered excitability in the human epileptic focus. Clin Neurophysiol. 2017; 128 (9): 1673-1681.
- 22. Matsuzaki N, Juhasz C, Asano E. Cortico-cortical evoked potentials and stimulation-elicited gamma activity preferentially propagate from lower- to higher-order visual areas. Clin Neurophysiol. 2013; 124 (7): 1290-1296.
- 23. Pigorini A, Sarasso S, Proserpio P, et al. Bistability breaks-off deterministic responses to intracortical stimulation during non-REM sleep. Neuroimage. 2015; 112: 105-113.
- 24. Usami K, Matsumoto R, Kobayashi K, et al. Phasic REM Transiently Approaches
 Wakefulness in the Human Cortex-A Single-Pulse Electrical Stimulation Study. Sleep. 2017;
 40 (8). DOI:10.1093/sleep/zsx1077.
- 25. van 't Klooster MA, van Klink NE, van Blooijs D, et al. Evoked versus spontaneous high frequency oscillations in the chronic electrocorticogram in focal epilepsy. Clin Neurophysiol. 2017; 128 (5): 858-866.
- 26. Usami K, Milsap GW, Korzeniewska A, et al. Cortical Responses to Input From Distant Areas are Modulated by Local Spontaneous Alpha/Beta Oscillations. Cereb Cortex. 2019; 29 (2): 777-787.
- 27. Valentin A, Alarcon G. Single pulse electrical stimulation and high-frequency oscillations, a complicated marriage. Clin Neurophysiol. 2017; 128 (6): 1026-1027.

- 28. Alarcon G, Martinez J, Kerai SV, et al. In vivo neuronal firing patterns during human epileptiform discharges replicated by electrical stimulation. Clin Neurophysiol. 2012; 123 (9): 1736-1744.
- 29. Kozyrev V, Eysel UT, Jancke D. Voltage-sensitive dye imaging of transcranial magnetic stimulation-induced intracortical dynamics. Proc Natl Acad Sci U S A. 2014; 111 (37): 13553-13558.
- 30.Li CL, Chou SN. Cortical intracellular synaptic potentials and direct cortical stimulation. J Cell Comp Physiol. 1962; 60: 1-16.
- 31. Matsumoto R, Nair DR, LaPresto E, et al. Functional connectivity in the human language system: a cortico-cortical evoked potential study. Brain. 2004; 127 (Pt 10): 2316-2330.
- 32. Greenlee JD, Oya H, Kawasaki H, et al. A functional connection between inferior frontal gyrus and orofacial motor cortex in human. J Neurophysiol. 2004; 92 (2): 1153-1164.
- 33. Matsumoto R, Nair DR, LaPresto E, Bingaman W, Shibasaki H, Luders HO. Functional connectivity in human cortical motor system: a cortico-cortical evoked potential study. Brain. 2007; 130 (Pt 1): 181-197.
- 34. Lacruz ME, Garcia Seoane JJ, Valentin A, Selway R, Alarcon G. Frontal and temporal functional connections of the living human brain. Eur J Neurosci. 2007; 26 (5): 1357-1370.
- 35. Jimenez-Jimenez D, Abete-Rivas M, Martin-Lopez D, et al. Incidence of functional bitemporal connections in the human brain in vivo and their relevance to epilepsy surgery.

 Cortex. 2015; 65: 208-218.
- 36. Granger CWJ. Investigating Causal Relations by Econometric Models and Cross-Spectral Methods. Econometrica. 1969; 37 (3): 424-438.

- 37. Flinker A, Korzeniewska A, Shestyuk AY, et al. Redefining the role of Broca's area in speech.

 Proc Natl Acad Sci U S A. 2015; 112 (9): 2871-2875.
- 38. Korzeniewska A, Franaszczuk PJ, Crainiceanu CM, Kus R, Crone NE. Dynamics of large-scale cortical interactions at high gamma frequencies during word production: event related causality (ERC) analysis of human electrocorticography (ECoG). Neuroimage. 2011; 56 (4): 2218-2237.
- 39. Nishida M, Korzeniewska A, Crone NE, et al. Brain network dynamics in the human articulatory loop. Clin Neurophysiol. 2017; 128 (8): 1473-1487.
- 40. Korzeniewska A, Cervenka MC, Jouny CC, et al. Ictal propagation of high frequency activity is recapitulated in interictal recordings: effective connectivity of epileptogenic networks recorded with intracranial EEG. Neuroimage. 2014; 101: 96-113.
- 41. Usami K, Matsumoto R, Kobayashi K, et al. Phasic REM transiently approaches wakefulness in the human cortex a single-pulse electrical stimulation study. Sleep. 2017; 40 (8): DOI: 10.1093/sleep/zsx1077.
- 42. Matsumoto R, Imamura H, Inouchi M, et al. Left anterior temporal cortex actively engages in speech perception: A direct cortical stimulation study. Neuropsychologia. 2011; 49 (5): 1350-1354.
- 43. Rechtschaffen A, Kales A. A Manual of Standardized Terminology, Techniques and Scoring System for Sleep Stages of Human Subjects. Los Angeles: UCLA Brain Information Service/Brain Research Institute; 1968.
- 44. Mallat SG, Zhang ZF. Matching Pursuits with Time-Frequency Dictionaries. Ieee T Signal Proces. 1993; 41 (12): 3397-3415.

- 45. Maris E, Oostenveld R. Nonparametric statistical testing of EEG- and MEG-data. J Neurosci Methods. 2007; 164 (1): 177-190.
- 46. Steriade M, Nunez A, Amzica F. A novel slow (< 1 Hz) oscillation of neocortical neurons in vivo: depolarizing and hyperpolarizing components. J Neurosci. 1993; 13 (8): 3252-3265.
- 47. Huber R, Ghilardi MF, Massimini M, et al. Arm immobilization causes cortical plastic changes and locally decreases sleep slow wave activity. Nat Neurosci. 2006; 9 (9): 1169-1176.
- 48. Landsness EC, Crupi D, Hulse BK, et al. Sleep-dependent improvement in visuomotor learning: a causal role for slow waves. Sleep. 2009; 32 (10): 1273-1284.
- 49. Nir Y, Staba RJ, Andrillon T, et al. Regional slow waves and spindles in human sleep. Neuron. 2011; 70 (1): 153-169.
- 50. Steriade M, Timofeev I. Neuronal plasticity in thalamocortical networks during sleep and waking oscillations. Neuron. 2003; 37 (4): 563-576.
- 51. Esser SK, Hill SL, Tononi G. Sleep homeostasis and cortical synchronization: I. Modeling the effects of synaptic strength on sleep slow waves. Sleep. 2007; 30 (12): 1617-1630.
- 52. Herman ST, Walczak TS, Bazil CW. Distribution of partial seizures during the sleep--wake cycle: differences by seizure onset site. Neurology. 2001; 56 (11): 1453-1459.
- 53. Kaleyias J, Loddenkemper T, Vendrame M, et al. Sleep-wake patterns of seizures in children with lesional epilepsy. Pediatr Neurol. 2011; 45 (2): 109-113.
- 54. Massimini M, Ferrarelli F, Murphy M, et al. Cortical reactivity and effective connectivity during REM sleep in humans. Cogn Neurosci. 2010; 1 (3): 176-183.
- 55. Siclari F, Baird B, Perogamvros L, et al. The neural correlates of dreaming. Nat Neurosci. 2017; 20 (6): 872-878.

- 56. Braun AR, Balkin TJ, Wesensten NJ, et al. Dissociated pattern of activity in visual cortices and their projections during human rapid eye movement sleep. Science. 1998; 279 (5347): 91-95.
- 57. Hobson JA, Pace-Schott EF, Stickgold R. Dreaming and the brain: toward a cognitive neuroscience of conscious states. Behav Brain Sci. 2000; 23 (6): 793-842; discussion 904-1121.
- 58. Tononi G. Consciousness as integrated information: a provisional manifesto. Biol Bull. 2008; 215 (3): 216-242.
- 59. Bor D, Seth AK. Consciousness and the prefrontal parietal network: insights from attention, working memory, and chunking. Front Psychol. 2012; 3: 63.
- 60. Yamao Y, Matsumoto R, Kunieda T, et al. Intraoperative dorsal language network mapping by using single-pulse electrical stimulation. Hum Brain Mapp. 2014; 35 (9): 4345-4361.
- 61. Premoli I, Biondi A, Carlesso S, Rivolta D, Richardson MP. Lamotrigine and levetiracetam exert a similar modulation of TMS-evoked EEG potentials. Epilepsia. 2017; 58 (1): 42-50.

FIGURE LEGENDS

Figure 1-The schema of analysis. (1) Single-pulse electrical stimulation (SPES) at a fixed frequency of 1 Hz with alternating polarity was applied to frontal or parietal cortex (this representative figure shows one of the frontal stimulation group) via two neighboring subdural electrodes across sleep stages (wakefulness, light sleep, slow wave sleep, and REM sleep.). (2) High-gamma cortico-cortical spectral responses (CCSR^{HG}) were defined as statistically significant increases in high-gamma (80-150 Hz) power within 100-550 ms of stimulation, relative to baseline (-300 to -100 ms before stimulation). Any ECoG site with a CCSRHG during wakefulness or any sleep stage was considered to be part of network dynamics elicited by SPES. These ECoG sites were grouped by anatomy into frontal, parietal, and temporo-occipital lobe for further analysis. (3) Patterns of high-gamma propagation within the network were analyzed by the eventrelated causality (ERC) method in each sleep stage. ERC values were normalized for group analysis. (4) The data were pooled across subjects in each stimulation group (frontal or parietal) in each sleep stage. (5) We investigated whether ERC values between sleep stages change in each propagation pattern.

Figure 2-The electrodes in which CCSR^{HG} increase was found from +100 to +550 ms after stimulation, (A) in the frontal stimulation group, (B) in the parietal stimulation group. The subdural electrodes were all linearly co-registered onto MRI taken before implantation in each subject. Each angle of the brain view was arranged so that all analyzed electrodes could be seen. The Sylvian fissure is indicated by a red arrowhead in some subjects for better understanding of the orientation.

Figure 3-Schematic illustrates a category of neural propagation, induced by SPES (frontal or parietal stimulation), that was significantly different between sleep stages. Stimulation sites are shown by a red lightning bolt. Right graphs depict the time course in unit of seconds after stimulation (x-axis) of ERC values (y-axis) of each specific propagation within or between regions. Each waveform is shown by mean \pm 1.96*S.E.M. Black bar with an asterisk above the waveforms shows the period that showed significant difference by permutation test using cluster-level statistics. The average magnitude of significant differences (min = -0.246, max = 0.085), during the time interval marked with black line and asterisk, is shown by the color and thickness (same thickness for the positive and negative changes) of the arrow in the left figure. Note that herein we found that sleep stage changed ERC values in frontal-to-parietal propagation after frontal stimulation (N = 82) and that it significantly occurred between wakefulness and slow wave sleep.

Figure 4-Schematic illustrates categories of neural propagation, induced by SPES, that were significantly different between sleep stages (N = 256 for propagation within the frontal lobe after frontal stimulation; N = 118 for propagation within the parietal lobe after parietal stimulation). The other conventions are the same as in Figure 3.

Table 1. Patient profile

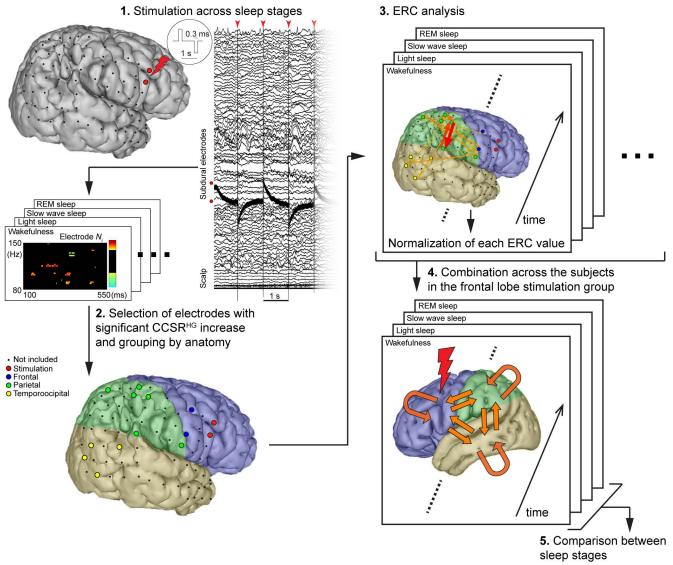
Patient No,	Epilepsy	Drug	Electrode-	Stimulus	Midpoint	No. of electrodes that showed		
age / gender,	classification		implanted side	intensity	of	significant CCSR ^{HG} increase and		rease and
handedness	/ etiology		/ No. of	(mA)	Stimulus	were used for ERC analysis		
			recording					
			electrodes					
						Frontal	Parietal	Temporo-
								Occipital
Frontal stimu	llation group							
1. 22F, R	R. FLE / FCD	CBZ, LTG	R, L/50	12	L MFG	5		5
2. 22M, R	L FLE, TLE /	CBZ, VPA	L / 60	10	L FP	3	1	4
	FCD							
3-1. 34M, R	R PLE, TLE /	CBZ, LEV,	R / 73	4	R preCG	2	4	6
	Traumatic	TPM, ZNS						
	injury, HS							
3-2.			R / 73	10	R PMv	2	6	4

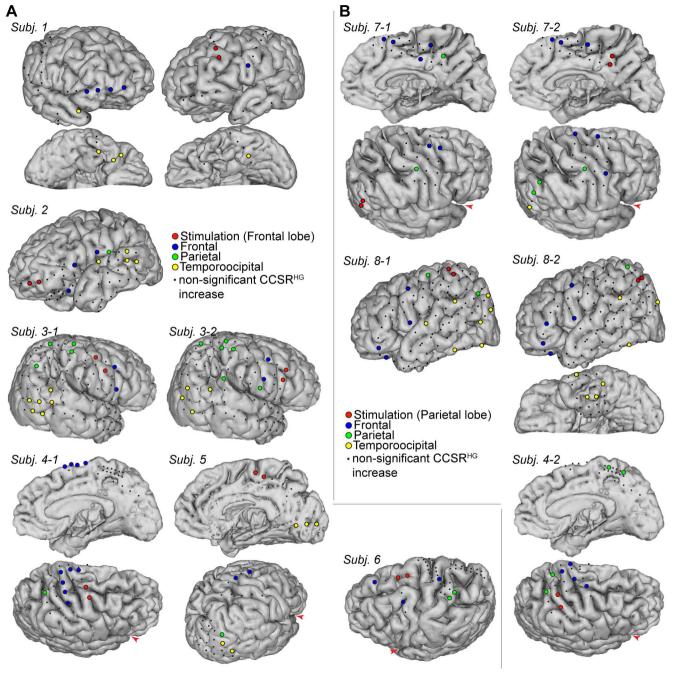
44 455 5	D D7 E /	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	D / = 1	4.0		10		
4-1. 27F, R	R PLE / tumor	CBZ, CZP,	R / 54	10	R MFG	10	1	
		LEV						
		LL V						
5. 27F, L	R PLE / FCD	CBZ, LEV,	R / 51	8	R SMA	2	1	5
		C DIJE						
		fosPHT,						
		PHT						
6.	L FLE / FCD	PHT, PB,	L/52	10	L PM	3	2	
		CLB, LEV						
		CLD, LEV						
Parietal stimulation group								
7-1. 44M, R	R FLE / tumor	CBZ, LEV,	R / 44	10	R AG	6	2	
		TDM VDA						
		TPM, VPA						
7-2.			R / 44	10	R precuneus	6	3	1
					_			
8-1. 29M, L	L mesial TLE,	CLB, PHT,	L/98	10	L SMG	4	2	7
	TLE / HS	ZNS						
	TLE / HS	ZIND						
8-2.			L/98	10	LAG	5	1	7
4-2.	See 4	4-1	R /54	10	R postCG	4	4	

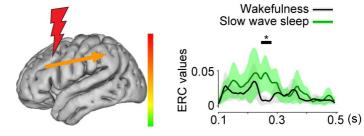
Abbreviation: **epilepsy classification** / **etiology** FLE//PLE/TLE: frontal/parietal temporal lobe epilepsy, FCD: focal cortical dysplasia, HS: hippocampal sclerosis.

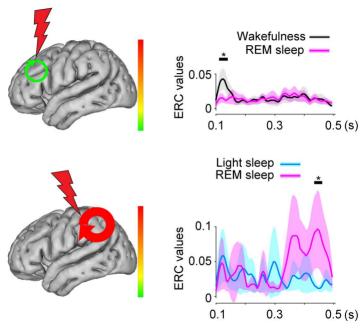
Drugs CBZ: carbamazepine, LTG: lamotrigine, VPA: valproic acid, LEV: levetiracetam, TPM: topiramate, ZNS: zonisamide, CZP: clonazepam, CLB: clobazam, PHT: phenytoin, PB: phenobarbital, PB: phenobarbital.

Stimulus site MFG: middle frontal gyrus, FP: frontal pole, preCG: precentral gyrus, PMv: ventral part of premotor area, SMA: supplementary motor area, AG: angular gyrus, LO: lateral part of occipital cortex, SMG: supramarginal gyrus, postCG: postcentral gyrus.









Supplementary Information

Feb. 1st, 2019

The neural tides of sleep and consciousness revealed by single-pulse electrical brain stimulation

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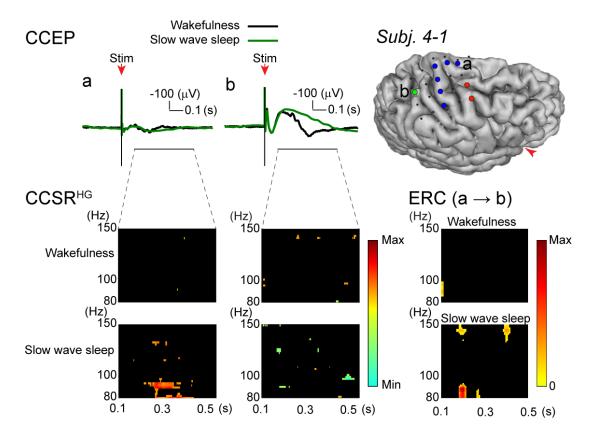
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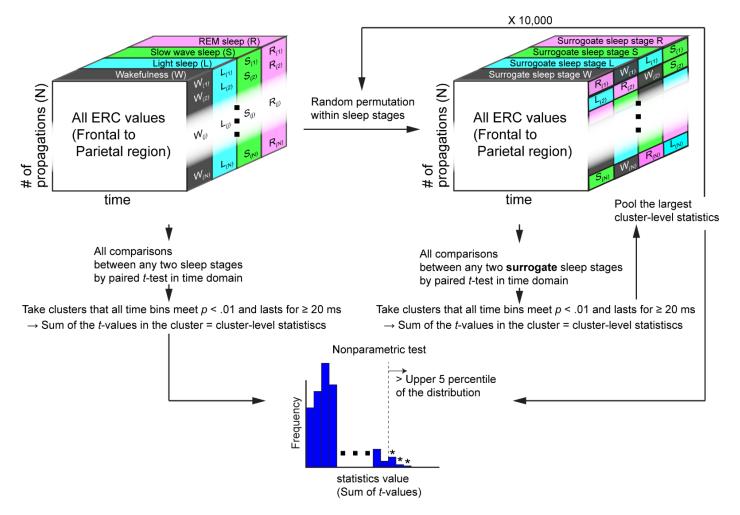
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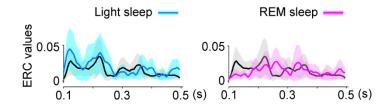
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Supplementary Figure 1- Examples of CCEP waveforms of two electrodes in one participant (a and b in Subj. 4-1) during wakefulness and slow wave sleep, corresponding CCSR^{HG} and ERC values. The underbar in CCEP shows the period where CCSR^{HG} below were calculated compared with baseline of -0.3~-0.1 s before stimulation. Note that sleep modulates CCEP waveforms as shown in our previous study (Usami K. et al., 2015, Hum Brain Mapp). Sleep also modulates CCSR^{HG} and ERC values in relation to a and b. The other conventions are the same as Fig. 2. Statistically significant values are colored and depicted in CCSR^{HG} and ERC on the black background. Note that the magnitudes of CCSR^{HG} does not directly correlate to the ERC values.



Supplementary Figure 2- An illustrative pipeline of the statistical analysis of this study. In this example, ERC values of propagation from frontal to parietal region are statistically analyzed.



Supplementary Figure 3- Comparison between wakefulness vs. light sleep or REM sleep in ERC values of propagation from frontal to parietal. The conventions are the same as Fig. 3. Note that no statistical difference was detected in these comparisons.