Modulatory Effects of Pregnancy on Inflammatory Bowel Disease

Janine van der Giessen, MsC, MD¹, Vivian W. Huang, MsC, MD, PhD², C. Janneke van der Woude, MD, PhD¹ and Gwenny M. Fuhler, MsC, PhD¹

The disease course of autoimmune diseases such as rheumatoid arthritis is altered during pregnancy, and a similar modulatory role of pregnancy on inflammatory bowel disease (IBD) has been proposed. Hormonal, immunological, and microbial changes occurring during normal pregnancy may interact with the pathophysiology of IBD. IBD consists of Crohn's disease and ulcerative colitis, and because of genetic, immunological, and microbial differences between these disease entities, they may react differently during pregnancy and should be described separately. This review will address the pregnancy-induced physiological changes and their potential effect on the disease course of ulcerative colitis and Crohn's disease, with emphasis on the modulation of epithelial barrier function and immune profiles by pregnancy hormones, microbial changes, and microchimerism.

Clinical and Translational Gastroenterology 2019;10:e-00009. https://doi.org/10.14309/ctg.000000000000000

INTRODUCTION

Inflammatory bowel disease (IBD), comprising Crohn's disease (CD) and ulcerative colitis (UC), is a group of chronic diseases of the gastrointestinal tract that affects men and women in their reproductive years of life. IBD and IBD therapies can have an impact on fertility, pregnancy outcomes, and fetal/neonatal health. *Vice versa*, the changes in hormones and in the immune system that occur during pregnancy may also influence IBD activity.

There is a clear link between the female reproductive cycle and the gastrointestinal tract, as demonstrated by several studies reporting an increase in gastrointestinal symptoms among women with IBD and irritable bowel syndrome before and during the menstrual period (1,2) and in changes to the menstrual function among women with IBD (3). Physiological changes that occur during the menstrual period include changes in hormones, cytokines, and immune profiles, which may affect gastrointestinal motility, inflammation, and sensitivity (4). Similar changes also occur during pregnancy, and a modulatory role of pregnancy on inflammatory disease behavior has therefore been the topic of research for many years. The most convincing amelioration of (auto)inflammatory disease during pregnancy is observed in rheumatoid arthritis (RA), where symptoms abate during pregnancy, and flares are commonly observed postpartum (5,6). With many of the underlying pathogenic mechanisms (genetics, intestinal microbiome alterations, and immune shifts) overlapping with IBD, resulting in several shared treatment options (7-9), it is not surprising that a disease modulatory role for pregnancy in IBD has also been speculated upon. Nevertheless, conflicting results of the effect of pregnancy in IBD have been observed. One study showed that patients with both CD and UC experienced fewer flares in the 3 years postpartum as compared to their flare

rate before pregnancy (10). A 10-year follow-up study confirmed that relapse rates decreased in UC (from 0.34 to 0.18 flares per year) and CD (from 0.76 to 0.12 flares per year) after pregnancy (11). In addition, it appears safe to stop anti-tumor necrosis factor alpha (TNF- α) treatment in pregnant patients with IBD, without increasing the risk of flares (12,13). However, these data are disputed by a study of Pedersen et al. (14), who showed that pregnant women with CD have a similar disease course during and after pregnancy as compared to nonpregnant women with CD. In contrast, women with UC have a higher risk of relapse during pregnancy (relative risk (RR) 2.19) and postpartum (RR 6.22), compared to nonpregnant women with UC. The course of IBD activity during pregnancy is closely related to disease activity preconception (15), with women who conceive during a time of active disease having twice the risk (RR 2.0) of disease flare during pregnancy compared to those who conceive during a time of remission. Although the often-reported medication nonadherence during pregnancy may be a confounding factor (16), disease course during pregnancy appears to be related to the type of IBD, suggesting a true relationship between pregnancy and disease activity.

In this review, we summarize the current knowledge regarding the interaction between reproductive physiology and IBD pathophysiology, and propose explanations for the clinical observations of IBD behavior during the reproductive period. We describe the pathological alterations in barrier function, immunology, and microbiome in IBD and discuss how these factors are modulated during pregnancy. A better understanding of these complex interactions and clinical observations will aid clinicians and researchers in improving the management of IBD during pregnancy, and optimize maternal and neonatal outcomes.

© 2019 The Author(s). Published by Wolters Kluwer Health, Inc. on behalf of The American College of Gastroenterology

¹Department of Gastroenterology and Hepatology, Erasmus Medical Center, Rotterdam, the Netherlands; ²Department of Gastroenterology and Hepatology, Mount Sinai Hospital, Toronto, Canada. **Correspondence:** Janine van der Giessen. E-mail: j.vandergiessen@erasmusmc.nl. **Received August 16, 2018; accepted January 7, 2019; published online March 19, 2019**

van der Giessen et al.

PATHOGENESIS OF IBD

IBD is a multifactorial disease, in which an altered immune response toward the intestinal microflora results in chronic inflammation of the intestinal tract. In addition to environmental factors (hygiene, smoking, diet, etc.), genetic susceptibility plays an important role in IBD, and large genome-wide association studies have identified more than 200 genetic loci associated with an increased risk of developing IBD (17,18). Interestingly, attempts at identifying common underlying mechanisms based on these loci have uncovered an important role for (innate) immunity and bacterial handling in IBD susceptibility: Many of the identified risk genes can be classified in pathways affecting epithelial barrier function, innate immune cell function, or adaptive immunity. All of these processes are critical at the contact interface between host and bacteria, underscoring the importance of these interactions in IBD development (18).

Epithelial barrier function in IBD

The first obstacle for bacterial invasion is represented by the intestinal epithelial barrier, which, although not traditionally regarded as part of the immune system, is now gaining recognition as part of the first-line innate immune defense. Bacteria are physically separated from the actual barrier cells through the production of a mucous layer and the release of antimicrobial peptides, therein, by goblet cells and Paneth cells, respectively. Disease predisposing genetic variants in mucin genes may contribute to alterations in the mucus layer in patients with IBD (19). With the mucosal layer breached, bacterial components have an increased chance to reach the epithelial cell layer. In response, different immune cells at the mucosa/luminal interface produce inflammatory cytokines, such as interferon gamma (IFN- γ) and TNF- α , which can inhibit antiapoptotic proteins and promote apoptotic processes, resulting in a weakening of the epithelial lining and an increased translocation of pathogens (20). This process, referred to as "leaky gut," is already seen in healthy firstdegree relatives of patients and therefore appears to be one of the disease initiating events in patients with IBD. Barrier dysfunction is worsened during active disease, when there is an additional reduction in tight junctions, which regulate the epithelial permeability (21). Thus, the overall weakening of the barrier function in IBD results in an enhanced exposure of the mucosa to bacterial components, which stimulates the attraction of immune cells and perpetuates inflammation.

Epithelial barrier function during pregnancy

Female reproductive hormones fluctuate during the normal menstrual cycle, with estrogen reaching peak levels before ovulation and progesterone reaching peak levels during the luteal phase of the cycle. Fluctuations of these hormones even on estrus scale already appear to affect bowel health. The gut epithelium expresses receptors for both estrogen (estrogen receptor α and β) and progesterone (22), and data in animal models show that paracellular permeability is decreased during the estrogen dominant phase of the cycle as compared to the progesterone dominant phase, consistent with an improved barrier integrity in response to estrogen (23). Gut epithelial cells in female rats are also more resistant to injury and inflammation than in male rats, and application of estrogen to male gut cells abrogates the enhanced inflammatory susceptibility in these male cells (24). Furthermore, progesterone receptor expression is increased in

constipated persons, suggesting that even though progesterone does not have a direct effect on barrier integrity (23), it may affect ion and water transport in the gut (25).

With these relatively small systemic fluctuations in hormone levels already impacting epithelial barrier function in an antiinflammatory and diarrhea-reducing manner, it is tempting to speculate that similar actions on a larger scale take place during pregnancy. Estrogen and progesterone levels increase rapidly during the first trimester, causing some of the nausea women experience. Estrogen peak levels are reached during the third trimester, accounting for the vascularization of the placenta and uterus, supporting the development of the fetus and the development of the milk duct. Interestingly, although low levels of 17 beta-estradiol (17β-EE) decreased paracellular permeability of vascular endothelial cells, high levels of EE increased the permeability, as a result of biphasic modulation of the tight junction molecule occludin (26). Thus, although epithelial barrier function has not been investigated during pregnancy, it is possible that studies performed during estrus are not reflective of a protective role of estrogen during pregnancy. Nevertheless, in irritable bowel syndrome, a link between increased gastrointestinal symptoms at the lowest estrogen levels of the menstrual cycle and reduced complaints during pregnancy is suggestive of a positive effect of pregnancy hormones on intestinal health (27).

IMMUNITY IN IBD

Innate immunity

The immune system represents a complex interplay of different cell types aimed at defending the human body from pathogenic microorganisms. Innate immunity is the first-line defense of the body against infections and includes monocyte/macrophages, granulocytes, and dendritic cells (DCs) (Table 1). These cells, constitutively present in body tissues, act as sentinels of the body by indiscriminate uptake (phagocytosis) and digestion of pathogens. Increased numbers of granulocytes, macrophages, and DCs have been observed in intestinal lesions in IBD. These cells may contribute to exacerbation of disease by releasing damaging reactive oxygen species and increasing local proinflammatory cytokine levels. An inherent alteration in bacterial responses of these cells appears to be present in IBD, which may contribute to the pathogenesis (28-30). For instance, macrophages from patients with IBD show increased proinflammatory and decreased anti-inflammatory cytokines when stimulated with bacteria (31). Epithelial wound healing in colitis models requires the presence of specialized M2 macrophages (32,33), which, in contrast to proinflammatory M1 macrophages, release anti-inflammatory interleukin 10 (IL-10) and contribute to tissue remodeling. In mucosa from patients with IBD, a shift toward M1 macrophages at the expense of M2 macrophages is observed (34), which may contribute to an impaired mucosal healing and prolonged inflammation.

Adaptive immunity

Presentation of pathogenic antigens on the cell surface of DCs and macrophages cells in the context of major histocompatibility complex (MHC) II molecules can subsequently activate cells of the adaptive immune system, in particular CD4+ T-helper cells (Th cells). On antigen stimulation, T cells differentiate into different subsets, depending on the local cytokine milieu. These include Th1, Th2, Th17, and regulatory T cells (Tregs), which each fulfill different functions and produce different cytokines.

2

Table 1. Changes of the immune system during pregnancy and IBD

	Cell type	Subtype	Function	Changes in IBD	Changes during pregnancy	Conceivable effect of changes during pregnancy on IBD
Epithelial barrier	Goblet cells Paneth cells		Production of a mucous layer Release of antimicrobial peptides	Decrease of mucus layer in IBD Decrease of antimicrobial peptides in IBD	Improved barrier integrity in response to estrogen	Positive effect, mainly due to increase estrogen
Innate immune system; adaptive immune system	Monocytes	Macrophages and dendritic cells (DCs)	Phagocytosis and digestion of pathogens Antigen presentation and activation of the adaptive immune system DCs express IDO1, which induces apoptosis of CD8+ T cells and promotes differentiation of CD4+ T cells to Tregs	Increased in IBD Skewing of macrophages from M2 (important for wound healing) to M1 (inflammatory) phenotype	Decrease from early pregnancy to mid-gestation Skewing toward M2 wound healing macrophages at placental interface Fetal tolerance via Tregs	Positive effects through circulating M2 macrophages and Tregs and their cytokines
	Granulocytes		Phagocytosis and digestion of pathogens Release a number of different effector molecules at site of infection	Increased in IBD	Decrease from early pregnancy to mid-gestation	Decrease during pregnancy may have a positive effect on IBD course
	Natural killer (NK) cells		Secrete cytokines such as IFN-γ and TNF-α, which act on macrophages and DCs Ability to kill tumor cells without any priming or prior activation	Increased in CD	Increase of placental and decidual NK cells	Local effect during pregnancy, so probably no influence on the course of IBD
	Innate lymphoid cells (ILCs)	ILC1 (IFN-γ, TNF-α) ILC2 (IL-4, IL-5, IL-9, IL-13) ILC3 (IL-17, IL-22, TNF-α)	Secrete immunoregulatory cytokines	ILC1 increased in CD ILC2 increased in IBD ILC3 increased in IBD	First trimester: Increase of ILC1 and ILC3	Negative effect on CD, potentially beneficial effect on UC

Modulatory Effects of Pregnancy on IBD

REVIEW ARTICLE

Table 1. (continued)						
	Cell type	Subtype	Function	Changes in IBD	Changes during pregnancy	Conceivable effect of changes during pregnancy on IBD
	T cells	T-helper 1 (Th1) (<i>IL-2</i> , <i>IL-12</i> , IFN-γ, <i>TNF-α</i>) T-helper 2 (Th2) (<i>IL-4</i> , <i>IL-5</i> , <i>IL-6</i> , <i>IL-9</i> , <i>IL-10</i> , <i>IL-13</i>) T-helper 17 (Th17) (<i>IL-17</i> , <i>IL-21</i> , <i>IL-22</i>) Regulatory T cells (Tregs) (<i>TGF-β</i> , <i>IL-35</i> , <i>IL-10</i>)	Cytotoxicity, antitumor and antiviral responses Antibody mediated immunity by stimulating B cells Protect cell surfaces by removing extracellular bacteria Regulate the function of other T-cell subsets and thereby repress inflammatory processes	Increased in CD Increased in UC Increased in UC Increased in IBD	First trimester: Increased in local tissue Second/third trimester: Increase in local tissue Second trimester: Increased	UC more likely to flare during pregnancy CD, not UC, may benefit from the shift to Th2 phenotype Unclear
Hormones	HCG Progesterone		Mediates early expansion of Tregs Modulates DC responses by inducing IDO1 expression Reduces inflammatory cytokines such as IL-17 while increasing IL-10 levels <i>In vitro</i> , hCG is able to stimulate peripheral blood DC subsets to maintain a tolerant phenotype Decrease of proinflammatory mediators (i.e., TNF-α, IL-6, IL-		Increase in first trimester Increase during pregnancy, with a decrease before labor	HCG or its peptides may contribute to the amelioration of inflammatory processes
	Estrogen		1β , NO) Increases IL-10 production by macrophages and monocytes Decreases inflammatory cytokine production (i.e., TNF-α and IFN-γ) Inhibits NO synthase activity Decreases the recruitment of inflammatory cells		Increase during pregnancy	Conflicting data in animal studies and human studies regarding contraceptive use

van der Giessen et al.

lable 1. (continued)						
	Cell type	Subtype	Function	Changes in IBD	Changes during pregnancy	Conceivable effect of changes during pregnancy on IBD
Microbiome	Bacteria and their metabolites			Reduced diversity of the bacteria present Decrease of anti-inflammatory <i>Firmicutes</i> (i.e., <i>Faecalibacterium prausnitzii</i>) Increase of proteobacteria and Bacteroidetes phylum members CD show a less stable microbiome and reduced diversity compared to patients with UC	Third trimester: Dysbiosis, resembling a state of low-grade inflammation of the gastrointestinal tract	Worsening of IBD because of increase of dysbiotic changes seen in IBD during pregnancy
Microchimerism	Immune cells, stem cells	O O	existence of two genetically ferent populations of cells in e individual	Maternal microchimerism is not increased in patients with IBD Fetal microhimerism in IBD pregnancy remains to be investigated	Whole cells cross the placenta from mother to child and vice versa	Fetal microchimerism in IBD pregnancy remains to be investigated, but adverse effects speculated in (auto)immune diseases
CD, Crohn's disease; HCG, hume	an chorionic gonadotropin; IBD,	, inflammatory bowel c	disease; INF, interferon gamma;	TNF- α , tumor necrosis factor alpha; l	UC, ulcerative colitis.	

Modulatory Effects of Pregnancy on IBD 5

Distinct cytokine expression differences and T-cell subset activities have been observed between patients with CD and UC (35), and an overactivation of the adaptive immune response with mucosal infiltrating T cells is evident, with the effectivity of targeted therapies against T cells underscoring the importance of this cell compartment in disease activity. Although IFN- γ and IL-17A cytokine expression, representative of Th1 and Th17 cells, respectively, are increased the lamina propria in CD, the Th2 cytokines IL-4, IL-5, and IL-13 are increased in UC (36). Under normal circumstances, Th1 and Th2 cells are in a dynamic equilibrium, with an imbalance resulting in either Th1 or Th2 dominant diseases. Although a gross simplification, CD is now generally regarded as a Th1/Th17 disease, whereas UC is considered as a Th2/Th17 disease.

Tolerance

Of course, with the number of bacteria being equal to the number of human cells in the body (37), it is imperative that the immune system does not respond to all bacteria present. Immune tolerance development is therefore key to a successful symbiosis with our commensal microflora and is largely mediated by Tregs, which suppress T-cell activation through production of cytokines such as IL-10 and transforming growth factor β (38). In addition, on IFN- γ stimulation, DCs express the enzyme indoleamine 2,3,-dioxygenase (IDO1), which converts the essential amino acid tryptophan into kynurenine. This has the dual effect of inducing apoptosis of CD8+ T cells by tryptophan depletion, and skewing CD4+ T cells to Treg differentiation (39,40). Although theoretically it might be expected that regulatory T-cell functions would be decreased in intestinal inflammation, the reverse has been observed: In IBD, both the number of Tregs and their differentiation-inducing agent IDO1 are increased in mucosal IBD biopsies compared to patients with non-IBD (41,42). In part, this seems a (failed) compensatory mechanism, with Tregs from the peripheral blood being recruited to inflamed mucosal area (42). Nevertheless, many experimental models show the benefits of redirecting the Th/Treg balance and suggest that Tregs may be a suitable target for treatment (43). Phenotypic alterations associated with reduced tolerance induction have also been observed for DCs and macrophages in IBD (44,45).

Innate lymphoid cells

In addition to innate myeloid cells, a family of lymphoid-derived cells with innate properties exists, which includes natural killer (NK) cells and a relatively recently identified subset of cells called innate lymphoid cells (ILCs) (46) (Table 1). These cells are enriched at the intestinal mucosa, but unlike "real" lymphocytes, they do not require antigen recognition in MHC II context, but rather rely on myeloid-derived cytokines and natural cytotoxicity receptors for their activation (47). Increased numbers of IL-17 and IFN- γ producing NK cells and ILCs have been observed in mucosal biopsies from patients with CD (48,49), but not from patients with UC (50). Although several experimental models have now highlighted the importance of ILCs for IBD pathology (reviewed in (51)), and NK cell populations are a target for treatments such as 6-mercaptopurine and azathioprine (48,52), cytokine disturbances in IBD have traditionally been linked to a skewing in adaptive immune responses, in particular those represented by Th-cell subsets.

Immunity during pregnancy

Many excellent reviews have already been written on the immunological changes taking place during pregnancy (53–56), the main findings of which are summarized here.

During pregnancy, an MHC mismatched fetus is present in the mother, which, despite the presence of a placental barrier, still affects the maternal immune system. Thus, induction of tolerance against paternal antigens appears to lay at the heart of immunological changes in successful pregnancies. Immune cells infiltrate the placenta during pregnancy, around 70% of which consists of NK cells. Unlike peripheral NK cells, placental NK cells are not cytotoxic, but help decidualization, angiogenesis, immune tolerance, and fetal development by producing growth factors (57,58). Decidual NK cells may possess both immuneactivating and regulatory properties (59), and although their presence is beneficial during early pregnancy, their persistence or failure to switch to a different phenotype in later pregnancy is associated with adverse pregnancy outcomes (60,61).

The remainder of placental immune cells consists mostly of macrophages and T cells, including Tregs. Macrophages in the decidua show a distinct M2 phenotype and are a major source of placental anti-inflammatory IL-10 and show reduced T cellactivating properties compared to their peripheral blood counterparts (62). They (as well as DCs and trophoblasts) are an important source of the soluble IDO1 enzyme, which contributes to the generation of Tregs and establishment of fetal tolerance (63). It has been postulated that a shift from inflammatory Th1 to more permissive Th2 cytokine profiles is required for a successful pregnancy (64,65). IL-25, an IL-17 family member expressed by decidual T-cells, NK cells, Tregs, and macrophages, stimulates the production of IL-4 and IL-10 in decidual T cells, thereby contributing to a Th2 environment in first-term placentas (66). Furthermore, human term placentas show increased levels of Th2 cytokines compared to preterm placentas (67,68). However, it is increasingly accepted that a healthy pregnancy depends on the maternal immune system to adapt to the different stages of pregnancy, and that proinflammatory processes are also required for the tissue remodeling, which is essential for decidua formation and labor induction (53). For instance, despite the presence of IL-10, the first trimester of pregnancy is also characterized by the presence of a proinflammatory Th1 immune profile for the successful implantation of the blastocyst, and IL-6, IL-8, and TNF- α are present at the implantation site (55). The source of these cytokines may be Th1 cells (69), although ILC1 and specialized ILC3 cells have also been observed in first-term placentas (70). Cell subsets shift during pregnancy, with the presence of macrophages declining from early to mid gestation, whereas Tcell frequencies increase during this time interval (71). Term labor and delivery appears to require low-level, well-controlled inflammatory processes (72). Correspondingly, placental IL-10 levels decrease toward labor (73), and rat models indicate an increase IL-6, TNF- α , and IL-1 β in term placentas (74).

In toto, the current general consensus suggests that implantation requires a Th1 response, followed by a shift toward a Th2 phenotype for the main duration of pregnancy and again a Th1 milieu toward partition (75). It should be noted, however, that much of the data come from animal studies, which may not necessarily reflect the human situation as in contrast to human placentas, T cells represent a rare population in mouse placentas (76).

Systemic immunological effects of pregnancy on IBD?

Differences in disease behavior between CD and UC during pregnancy and peripartum may potentially be explained by intrinsic differences in the immune pathways that lead to each disease. As seen above, pregnancy is associated with immunological changes at the fetal/maternal interface, with a predominantly Th2/tolerogenic phenotype. Thus, it is tempting to speculate that a Th2 shift during pregnancy ameliorates disease in those patients in whom Th1 responses dominate (such as CD), while aggravating disease in Th2 dominant patients (mainly UC). Nevertheless, the maternal peripheral immune system is still capable of mounting a robust immune response to pathogenic antigens (77), and the question therefore remains to what extent placental immunological changes can affect immunological processes at distant body sites.

Levels of Th1 and Th2 patterns in utero generally appear to be mirrored by ratios in peripheral blood (53), although most data are derived from studies comparing pregnancy outcomes, and hence blood is usually obtained at only one timepoint, often postpartum. There are conflicting data on modulation of serum cytokine levels in healthy pregnant women, with some studies reporting a significant decrease of proinflammatory Th1 cytokines (e.g., IL-8, IL-12, IFN- γ , and TNF- α) from first to third trimester in healthy pregnant women (78), and others showing no difference or even an increase (79,80). For Th2 cytokines, even less is known, with one study reporting a stable level of IL-4 and IL-5 during pregnancy (81). Thus, it is unclear to what extent pregnancy induces peripheral cytokine changes, which may influence inflammatory diseases. However, ample evidence suggests that peripheral blood cell subsets at least are altered in normal pregnancy. For instance, stimulated peripheral blood mononuclear cells from pregnant women produce less Th1 and Th2 cytokines compared to healthy controls, in particular during second trimester, whereas levels increased postpartum, suggesting that systemic alterations in cell sensitivity exist during pregnancy, which may contribute to decreased (auto)immunity during pregnancy and increased flaring thereof, afterward (82). The peripheral blood percentage of Tregs also peaks during the second trimester of pregnancy, and in vitro, these Tregs are capable of reducing T-cell activation in response to DCs (83). Because development of Tregs during pregnancy appears to be related to the presence of fetal alloantigens rather than pregnancy hormones (84) and the Treg recognition receptor repertoire differs per organ (85), it is uncertain to what extent pregnancyinduced circulating Tregs would be useful at the inflamed mucosa. Nevertheless, much is unclear regarding mucosal Treg antigen recognition (86), and the fact that peripheral blood Treg levels drop during inflammation suggest that general recruitment of Tregs to inflammatory sites occurs (42). Their presence there may potentially contribute to modulation of inflammatory processes through production of inhibitory cytokines or suppression of DC maturation.

Similar to Tregs, NK cells present in preimplantation endometrium show a different receptor repertoire compared to peripheral blood NK cells in the same women (87). However, it has also been reported that in the first trimester of pregnancy, progesterone-dependent expression of the receptor T cell immunoglobulin and mucin-domain–containing-3 on peripheral blood NK cell confers immunosuppressive properties (88), and it is conceivable that these cells also reach the intestinal mucosa where they may modulate disease activity. The most important early immune modulator in pregnancy is now acknowledged to be human chorionic gonadotropin (hCG), which mediates early expansion of regulatory T cells (Tregs), modulates DC responses by inducing IDO1 expression, and reduces inflammatory cytokines such as IL-17 while increasing IL-10 levels (89). Importantly, many of these effects occur in peripheral blood from nonpregnant patients receiving hCG for their in vitro fertilization treatment. In vitro, hCG is able to stimulate peripheral blood DC subsets to maintain a tolerant phenotype (90). Several cleaved or "nicked" forms of hCG exist in vivo, and studies have shown that such hCG peptides show antiinflammatory properties in a host of mouse models, including lipopolysaccharides-induced septic shock, polymicrobial sepsis, hemorrhagic shock, and diabetes (91-96). Administration of hCG peptides also inhibited neutrophil recruitment and inflammatory markers such as IL-6 and TNF- α (89,96,97). The same authors also showed that human graft vs host disease at the skin was successfully treated with hCG, which corresponded with increased IDO1 expression in peripheral mononuclear cells, and also IL-10 serum levels and Treg upregulation (98). With hCG administration being able to prevent autoimmune diabetes in mice by downregulating Th1 responses (99), the use of hCG to control the autoimmune processes in RA and Sjogren's disease has been suggested (100), and it is tempting to speculate that hCG may also positively affect IBD.

The high amount of progesterone throughout pregnancy not only results in the laxity of the ligaments and joints, but is also thought to suppress the maternal immunologic response to fetal antigens and allows implantation in the endometrium. Progesterone reduces proinflammatory mediators (i.e., TNF- α , IL-6, IL-1 β , and nitric oxide (NO)) and increases IL-10 production by macrophages and monocytes (101). Application of progesterone in a temporomandibular joint inflammation model of ovariectomized rats reduced synovial inflammation and levels of TNF- α and IL-1 β (102). In a rat colitis model, progesterone ameliorated disease activity through reduction of TNF- α levels in colon and blood (103). Nevertheless, conflicting data have also been reported. In a chemically induced model of colitis 2,4,6-trinitrobenzene sulfonic acid, the progesterone dominant luteal phase of the menstrual cycle was associated with increased severity of colitis and treatment of ovariectomized animals with progesterone similarly increased disease severity, whereas estrogen decreased colitis (104). Indeed, anti-inflammatory properties have often been ascribed to estrogen, because it decreases inflammatory cytokine production (i.e., TNF- α and IFN- γ) inhibits NO synthase activity and decreases the recruitment of inflammatory cells (105,106). However, animal studies of IBD and the effect of estrogen also show inconsistent findings. Improvement of stool scores in human leukocyte antigen-B27 transgenic rats with chronic diarrhea was noted after treatment with 17α -ethynyl- 17β -EE (107). Similarly, estrogen reduced TNF- α , IL-1 β , and IL-6 levels, as well as inflammation in diverse rat models of colitis (108). Verdu et al. (109) found that a supraphysiological dose of 17β -EE has an anti-inflammatory effect in a dextran sodium sulfate murine model for colitis but a proinflammatory effect in the dinitrobenzene sulfonic acid colitis model. Clinical human studies of IBD and sex hormones focus mainly on postmenopausal women and/or oral contraception use. Kane et al. (110) described a protective effect of estrogen on the bowel in women with IBD, whereas Khalili et al.

REVIEW ARTICLE

(111) showed that postmenopausal women who use oral contraceptives had a higher risk of developing UC, but not CD. A meta-analysis of Cornish et al. (112) demonstrated that with time of exposure to oral contraceptives, the risk of developing CD was increased, and when contraceptives were stopped, the risk decreased again to that of the normal population.

It is clear that there is conflicting data on IBD and levels of sex hormones and that it is difficult to translate these clinical studies to the situation in a pregnant patient with IBD. Different immune cells may react in an opposite manner to different concentrations of estrogen and progesterone, and expression patterns of receptors of these hormones may vary under inflammatory conditions, precluding robust predictions on the overall effect of progesterone and estrogen on autoimmune disease.

MICROBIOME IN IBD AND PREGNANCY

As mentioned before, IBD is thought to arise in consequence of an altered immune response toward intestinal bacteria. We now know that the microbiome of patients with IBD is substantially altered as compared to healthy controls, and that inflamed regions show further microbial deregulation as compared to noninflamed regions (113-115). This so-called dysbiosis includes a reduced diversity of the bacteria present, in particular in patients with CD, with a noted decrease of anti-inflammatory Firmicutes (i.e., Faecalibacterium prausnitzii) and an increase of proteobacteria and Bacteroidetes phylum members (i.e., Bacteroides fragilis) (116-119). The host-microbiome interaction is reciprocal, and it is as yet unclear whether dysbiosis in IBD presents the chicken or the egg in the etiology of disease. Nevertheless, the general consensus now favors a causative role for the microbiome in disease initiation, because animal models indicate that bacterial presence is required for colitis development and that colitis may be conferred by transplantation of inflammation-associated feces (120).

Pregnancy is also accompanied by intestinal microbial changes. These changes induce a metabolic state that may be beneficial during pregnancy, as concluded by Koren et al. (121). They described that, in the first trimester of pregnancy, the gut microbiota is similar in many aspects to that of healthy nonpregnant controls. However, in the third trimester, a dysbiosis was observed, resembling a state of low-grade inflammation of the gastrointestinal tract. This dysbiosis was accounted for by the presence of Proteobacteria and Actinobacteria and was not related to body mass index (before pregnancy), antibiotic use, diet, or the presence of gestational diabetes. The overall diversity of the bacteria was also reduced at T3. These data would suggest that microbial changes that occur during normal pregnancy fortify dysbiotic changes seen in IBD, and would aggravate disease activity. Interestingly, patients with CD show a less stable microbiome and reduced diversity compared to patients with UC (122), and it is conceivable that further alterations during pregnancy therefore have less of an effect on CD disease activity as compared to UC. Of note, there are several studies which show that diet shapes the microbiome, and in particular, western diets are associated with IBD (123,124). Because it is commonly appreciated that women change their diet during pregnancy, it is important to take this into account in future studies.

It is clear that while microbial changes during pregnancy and the host defense mechanisms are both changed during pregnancy and IBD, the interactions are complex, reciprocal, and double edged, hampering interpretations of the observed changes.

MICROCHIMERISM AND α -FETOPROTEIN

The placental exchange of maternal and fetal gases, nutrients, metabolic waste products, and antibodies is well described. However, in addition to these small molecules, it is also possible for whole cells to cross the placenta from mother to child and vice versa. Such coexistence of 2 genetically different populations of cells in one individual is termed (micro)chimerism. Cellular transport is bidirectional (125,126), with maternal cells detected in 24%-42% of fetal-derived samples, and fetal cells were detected in 26%-51% of mothers (127,128). Fetal cells, which can be detected as early as 7 weeks gestation, are known to persist for some time after delivery (129). In fact, microchimerism has been observed in mothers up to 38 years after pregnancy, and in offspring well into adult life (130,131). In addition to fetal mature T-cells, CD34+ progenitor cells enter the maternal bloodstream during pregnancy, which retain their multilineage potential and can become adult hematopoietic cells of all linages and epithelial cells (131,132). With the potential for these cells to assert effector functions and affect the maternal immune system, the functional immunological consequences of these microchimers in health and disease are gaining interest (133,134). Male fetal cell-derived T-cell clones isolated from parous women show proliferation and IL-4 production in response to ex vivo stimulation with maternal T cells and MHC antigens, and this effect was higher in patients with systemic sclerosis, suggesting that these offspring T cells show a Th2 profile and could play a pathogenic role in immune disease (135). Interestingly, increased microchimerism was observed in peripheral blood mononuclear cells from patients with the autoimmune disease scleroderma, which has a peak incidence in women after childbearing years, again suggesting that such microchimerism may contribute in autoimmune disease (131). However, patients with either Grave's disease or Hashimoto's thyroiditis, two other autoimmune diseases associated with pregnancy, have reduced microchimerism as compared to healthy controls (136). Furthermore, microchimerism has been investigated in systemic lupus erythematosus (SLE), Sjogren's syndrome, multiple sclerosis, and RA, and may be either protective or harmful (133). In RA, where there is a clear beneficial effect of pregnancy on disease course, it had been suggested that accumulation of fetal T cells, which appear around gestational week 13, may dampen the maternal immune response, and that this effect weakens over time, because of senescence of these cells (137). The presence of fetal cells in maternal tissues correlates to the presence of maternal Tregs, which may account for some of the dampening of inflammatory disease during pregnancy (134). Although the exact effect of microchimerism on (auto)immune disease is as yet unclear, it is tempting to speculate that it may also play a role in IBD and affect disease course during pregnancy. Thus far, only maternal microchimerism has been studied in IBD, which did not appear to be increased in patients with IBD (138,139). Fetal microchimerism in IBD pregnancy remains to be investigated.

Another potential fetal source contributing to maternal (auto) immune response is α -fetoprotein (AFP), a protein produced by the yolk sac and fetal liver, which can be detected in the maternal serum from week 14 of pregnancy onward. AFP was shown to bind to autoantibodies produced in patients with the autoimmune disorder myasthenia gravis (MG), and it was thus speculated that circulating levels of AFP in the second and third trimester of pregnancy could induce clinical remission in patients with myasthenia gravis during these times (140). The immunomodulatory

effects of AFP extend beyond antibody binding (141). Studies indicate that AFP ameliorates a mouse model of multiple sclerosis (MS), through, among others, inhibition of Th1 cytokine production (142). It has been speculated that AFP may be used for the treatment of myasthenia gravis, MS, autoimmune uveitis, and psoriasis (143). Thus far, however, the potential role of AFP in IBD remains unexplored.

MODULATION OF IBD RESPONSE TO MEDICATION THROUGH PREGNANCY

Disease activity in women with IBD may also be modulated by pharmacokinetic changes induced by pregnancy and through interaction of IBD medication with the placenta, which may modulate the clinical effectivity of these drugs. However, although drugs such as the thiopurine 6-thioguanine nucleotide and 5-aminosalicylic acid are known to cross the placenta, this does not seem to influence therapeutic levels in the mothers (144). Less is known about the effects on patient and child outcomes of biologicals, the most recent IBD medications. The earliest of these are the anti-TNF- α treatments (infliximab, adalimumab, golimumab, and certolizumab pegol), with vedolizumab (\alpha 4\beta 7 integrin blocker) and ustekinumab (IL-12/ IL-23 blocker) following suit. From week 20 onward, maternal immunoglobulins (Igs) are transported across the placental barrier, to provide immunoprotection to the fetus (145). Transport of Igs is mediated by the neonatal fragment crystallizable (Fc) receptor (FcRn), which binds to the Fc region present in all antibodies, including therapeutic monoclonal antibodies. Certolizumab pegol does not undergo this FcRnmediated transfer across the placenta, because it lacks an IgG Fc region and therefore does not bind FcRn. Owing to the passive diffusion across the placenta, the levels of certolizumab pegol reaching the fetus are probably much lower when compared to infliximab and adalimumab (146). We and others have previously shown that infliximab and adalimumab levels in cord blood exceed levels present in serum from mothers treated with these medications (147,148), suggesting that active transport of these antibodies over the placental barrier may decrease bioavailability of the antibodies in the mother. As serum drug levels of these therapeutic antibodies correspond to clinical outcomes for patients with IBD, modulation of these levels through placental transport could potentially result in disease relapse (149,150). Thus far, however, maternal infliximab levels during pregnancy were shown to be increased, whereas adalimumab levels remained stable. Nevertheless, pharmacokinetic changes of these therapeutic antibodies on pregnancy have only been studied in a small cohort of patients, and larger studies are needed (151).

SUMMARY

The observation that the disease course of several (auto)immune diseases are altered during pregnancy suggests that there is an interaction between physiological changes taking place during pregnancy and pathophysiology of these diseases (Figure 1). For IBD, this relationship appears more apparent for UC than CD, which may be due to the fact that CD and UC have differential underlying genetic susceptibilities, immune profiles, and microbial changes. Genetic variants affecting cellular innate immunity are associated more with CD, whereas UC-specific single nucleotide polymorphisms affect epithelial barrier function genes (152). Furthermore, patients with CD show a more Th1 dominant



Figure 1. Interaction between physiological changes during pregnancy and the pathophysiology of inflammatory bowel disease.

cytokine profile and less stable microbiome compared to patients with UC, where Th2 responses appear more prevalent. Pregnancy modulates these disease-underlying mechanisms to a different extent at different timepoints during gestation, which may further explain why disease modification is not always apparent. Nevertheless, several conclusions may be inferred from our current understanding of pregnancy-induced physiological changes. Overall, a beneficial effect of pregnancy on epithelial barrier function seems apparent, with relatively small fluctuations of pregnancy hormones already affecting the gut barrier. Furthermore, an overall image of induction of tolerance and suppression of immune responses during gestation is arising. With a predominant shift toward a Th2 phenotype, many reviews have speculated that, in particular, Th1-mediated diseases such as RA and CD may benefit from these pregnancy-induced changes, whereas Th2-mediated diseases (such as SLE and UC) might be negatively affected (6,153,154). HCG, estrogen, and progesterone rise rapidly during pregnancy and have shown several antiinflammatory actions in animal models. Although most of these changes would be compatible with improvement of IBD activity, it has also been demonstrated that pregnant patients with SLE have lower levels of estrogen and progesterone in the third trimester of pregnancy compared to healthy controls, suggesting that some patient groups may benefit less from rises in pregnancy hormones (155). This also might be the case in UC, but studies to support this hypothesis are lacking. Finally, changes in the microbiome occurring during normal pregnancy do not appear to be beneficial to patients with IBD, but again, it is unclear to what extent these changes are modulated by pregnancy hormones and to what extent microbial alterations are present in pregnant patients with IBD. Thus, immune regulation in both pregnancy and IBD are complicated and not static. Whether or not IBD course is affected by pregnancy may depend on individual patient's characteristics, including ongoing disease activity before conception, their microbiome and hormone/diet-induced changes therein, and genetic underlying risk factors. Predicting which patients may experience reduced disease burden or increased disease activity during pregnancy and postpartum requires a better insight into the physiology and pathology of pregnancy and IBD.

CONFLICTS OF INTEREST

Guarantor of the article: J. van der Giessen, MsC, MD. **Specific author contributions:** J.v.d.G. wrote manuscript; V.W.H. wrote manuscript; C.J.v.d.W. devised study and corrected manuscript; G.M.F. devised study and wrote manuscript. All authors read and approved the final manuscript.

Financial support: None of the authors have anything to disclose. **Potential competing interests:** None.

REFERENCES

- Bernstein MT, Graff LA, Avery L, et al. Gastrointestinal symptoms before and during menses in healthy women. BMC Womens Health 2014;14:14.
- Lim SM, Nam CM, Kim YN, et al. The effect of the menstrual cycle on inflammatory bowel disease: A prospective study. Gut Liver 2013;7(1): 51–7.
- 3. Saha S, Zhao YQ, Shah SA, et al. Menstrual cycle changes in women with inflammatory bowel disease. Inflamm Bowel Dis 2014;20(3):534–40.
- Bharadwaj S, Kulkarni G, Shen B. Menstrual cycle, sex hormones in female inflammatory bowel disease patients with and without surgery. J Dig Dis 2015;16(5):245–55.
- Barrett JH, Brennan P, Fiddler M, et al. Does rheumatoid arthritis remit during pregnancy and relapse postpartum? Results from a nationwide study in the United Kingdom performed prospectively from late pregnancy. Arthritis Rheum 1999;42(6):1219–27.
- de Man YA, Dolhain RJEM, van de Geijn FE, et al. Disease activity of rheumatoid arthritis during pregnancy: Results from a nationwide prospective study. Arthritis Rheum 2008;59(9):1241–8.
- Lees CW, Barrett JC, Parkes M, et al. New IBD genetics: Common pathways with other diseases. Gut 2011;60(12):1739–53.
- 8. Rhodes J, Collins P. Lessons for inflammatory bowel disease from rheumatology. Dig Liver Dis 2006;38(3):157–62.
- Diamanti AP, Manuela Rosado M, Laganà B, et al. Microbiota and chronic inflammatory arthritis: An interwoven link. J Transl Med 2016; 14(1):233.
- Castiglione F, Pignata S, Morace F. Effect of pregnancy on the clinical course of a cohort of women with IBD. Ital J Gastroenterol 1996;28(4): 199–204.
- Riis L, Vind I, Politi P, et al. Does pregnancy change the disease course? A study in a European cohort of patients with inflammatory bowel disease. Am J Gastroenterol 2006;101:1539–45.
- de Lima A, Zelinkova Z, van der Ent C, et al. Tailored anti-TNF therapy during pregnancy in patients with IBD: Maternal and fetal safety. Gut 2016;65(8):1261–8.
- 13. Zelinkova Z, van der Ent C, Bruin KF, et al. Effects of discontinuing antitumor necrosis factor therapy during pregnancy on the course of inflammatory bowel disease and neonatal exposure. Clin Gastroenterol Hepatol 2013;11(3):318–21.
- Pedersen N, Bortoli A, Duricova D, et al. The course of inflammatory bowel disease during pregnancy and postpartum: A prospective European ECCO-EpiCom study of 209 pregnant women. Aliment Pharmacol Ther 2013;38(5):501–12.
- Abhyankar A, Ham M, Moss AC. Meta-analysis: The impact of disease activity at conception on disease activity during pregnancy in patients with inflammatory bowel disease. Aliment Pharmacol Ther 2013;38(5): 460–6.
- 16. Julsgaard M, Nørgaard M, Lodberg Hvas C, et al. Scandinavian Journal of Gastroenterology influence of medical treatment, smoking and disease activity on pregnancy outcomes in Crohn's disease influence of medical treatment, smoking and disease activity on pregnancy outcomes in Crohn's disease. Scand J Gastroenterol 2014;49:302–8.
- Cleynen I, Boucher G, Jostins L, et al. Inherited determinants of Crohn's disease and ulcerative colitis phenotypes: A genetic association study. Lancet 2016;387(10014):156–67.
- 18. Jostins L, Ripke S, Weersma RK, et al. Host-microbe interactions have shaped the genetic architecture of inflammatory bowel disease. Nature 2012;491(7422):119–24.
- Cornick S, Tawiah A, Chadee K. Roles and regulation of the mucus barrier in the gut. Tissue barriers 2015;3(1–2):e982426.
- Shen L, Su L, Turner JR. Mechanisms and functional implications of intestinal barrier defects. Dig Dis 2009;27(4):443–9.

- Gassler N, Rohr C, Schneider A, et al. Inflammatory bowel disease is associated with changes of enterocytic junctions. Am J Physiol Gastrointest Liver Physiol 2001;281(1):G216–28.
- Konstantinopoulos PA, Kominea A, Vandoros G, et al. Oestrogen receptor beta (ERbeta) is abundantly expressed in normal colonic mucosa, but declines in colon adenocarcinoma paralleling the tumour's dedifferentiation. Eur J Cancer 2003;39(9):1251–8.
- Braniste V, Leveque M, Buisson-Brenac C, et al. Oestradiol decreases colonic permeability through oestrogen receptor beta-mediated upregulation of occludin and junctional adhesion molecule-A in epithelial cells. J Physiol 2009;587(Pt 13):3317–28.
- 24. Homma H, Hoy E, Xu DZ, et al. The female intestine is more resistant than the male intestine to gut injury and inflammation when subjected to conditions associated with shock states. Am J Physiol Liver Physiol. 2005;288(3):G466–72.
- Xiao ZL, Pricolo V, Biancani P, et al. Role of progesterone signaling in the regulation of G-protein levels in female chronic constipation. Gastroenterology. 2005;128(3):667–75.
- Ye L, Martin TÁ, Parr C, et al. Biphasic effects of 17-beta-estradiol on expression of occludin and transendothelial resistance and paracellular permeability in human vascular endothelial cells. J Cel Physiol. 2003; 196(2):362–9.
- 27. Mulak A, Taché Y, Larauche M. Sex hormones in the modulation of irritable bowel syndrome. World J Gastroenterol. 2014;20(10):2433–48.
- Steinbach EC, Plevy SE. The role of macrophages and dendritic cells in the initiation of inflammation in IBD. Inflamm Bowel Dis. 2014;20(1): 166–75.
- 29. Somasundaram R, Deuring JJ, Van Der Woude CJ, et al. Linking risk conferring mutations in NCF4 to functional consequences in Crohn's disease. Gut. 2012;61(7):1097.
- Somasundaram R, Nuij VJAA, Van Der Woude CJ, et al. Peripheral neutrophil functions and cell signalling in Crohn's disease. PLoS One. 2013;8(12):e84521.
- Campos N, Magro F, Castro AR, et al. Macrophages from IBD patients exhibit defective tumour necrosis factor-α secretion but otherwise normal or augmented pro-inflammatory responses to infection. Immunobiology. 2011;216(8):961–70.
- Cosín-Roger J, Ortiz-Masiá D, Calatayud S, et al. The activation of Wnt signaling by a STAT6-dependent macrophage phenotype promotes mucosal repair in murine IBD. Mucosal Immunol. 2016;9(4):986–98.
- Van Welden S, De Vos M, Wielockx B, et al. Haematopoietic prolyl hydroxylase-1 deficiency promotes M2 macrophage polarization and is both necessary and sufficient to protect against experimental colitis. J Pathol. 2017;241(4):547–58.
- Lissner D, Schumann M, Batra A, et al. Monocyte and M1 macrophageinduced barrier defect contributes to chronic intestinal inflammation in IBD. Inflamm Bowel Dis. 2015;21(6):1297–305.
- Chen ML, Sundrud MS. Cytokine networks and T-cell subsets in inflammatory bowel diseases. Inflamm Bowel Dis. 2016;22(5):1157–67.
- 36. Fuss IJ, Neurath M, Boirivant M, et al. Disparate CD4+ lamina propria (LP) lymphokine secretion profiles in inflammatory bowel disease. Crohn's disease LP cells manifest increased secretion of IFN-gamma, whereas ulcerative colitis LP cells manifest increased secretion of IL-5. J Immunol. 1996;157(3):1261–70.
- 37. Sender R, Fuchs S, Milo R. Revised estimates for the number of human and bacteria cells in the body. PLoS Biol. 2016;14(8):e1002533.
- Wirnsberger G, Hinterberger M, Klein L. Regulatory T-cell differentiation versus clonal deletion of autoreactive thymocytes. Immunol Cel Biol. 2011;89(1):45–53.
- Munn DH, Mellor AL. Indoleamine 2,3 dioxygenase and metabolic control of immune responses. Trends Immunol. 2013;34(3):137–43.
- 40. Mondanelli G, Bianchi R, Pallotta MT, et al. A relay pathway between arginine and tryptophan metabolism confers immunosuppressive properties on dendritic cells. Immunity. 2017;46(2):233–44.
- Sznurkowska K, Żawrocki A, Sznurkowski J, et al. Indoleamine 2,3dioxygenase and regulatory t cells in intestinal mucosa in children with inflammatory bowel disease. J Biol Regul Homeost Agents. 2017;31(1): 125–31.
- 42. Maul J, Loddenkemper C, Mundt P, et al. Peripheral and intestinal regulatory CD4+CD25 high T cells in inflammatory bowel disease. Gastroenterol. 2005;128(7):1868–78.
- 43. Yamada A, Arakaki R, Saito M, et al. Role of regulatory T cell in the pathogenesis of inflammatory bowel disease. World J Gastroenterol. 2016;22(7):2195–205.

- 44. Elshal M, Aldahlawi A, Saadah O, et al. Reduced dendritic cells expressing CD200R1 in children with inflammatory bowel disease: Correlation with Th17 and regulatory T cells. Int J Mol Sci. 2015; 16(12):28998–9010.
- Kamada N, Hisamatsu T, Okamoto S, et al. Unique CD14+ intestinal macrophages contribute to the pathogenesis of Crohn's disease via IL-23/IFN-γ axis. J Clin Invest. 2008;118(6):2269–80.
- Hazenberg MD, Spits H. Human innate lymphoid cells. Blood. 2014; 124(5):700–9.
- 47. Kruse PH, Matta J, Ugolini S, et al. Natural cytotoxicity receptors and their ligands. Immunol Cel Biol. 2014;92(3):221–9.
- Steel AW, Mela CM, Lindsay JO, et al. Increased proportion of CD16+ NK cells in the colonic lamina propria of inflammatory bowel disease patients, but not after azathioprine treatment. Aliment Pharmacol Ther. 2011;33(1):115–26.
- Bernink JH, Peters CP, Munneke M, et al. Human type 1 innate lymphoid cells accumulate in inflamed mucosal tissues. Nat Immunol. 2013;14(3):221–9.
- Geremia A, Arancibia-Cárcamo CV, Fleming MPP, et al. IL-23–responsive innate lymphoid cells are increased in inflammatory bowel disease. J Exp Med. 2011;208(6):1127–33.
- 51. Geremia A, Arancibia-Cárcamo CV. Innate lymphoid cells in intestinal inflammation. Front Immunol. 2017;8:1296.
- Yusung S, McGovern D, Lin L, et al. NK cells are biologic and biochemical targets of 6-mercaptopurine in Crohn's disease patients. Clin Immunol. 2017;175:82–90.
- 53. Saito S, Nakashima A, Shima T, et al. Th1/Th2/Th17 and regulatory T-cell paradigm in pregnancy. Am J Reprod Immunol. 2010;63(6):601–10.
- 54. Mor G, Cardenas I. The immune system in pregnancy: A unique complexity. Am J Reprod Immunol. 2010;63(6):425–33.
- Mor G, Cardenas I, Abrahams V, et al. Inflammation and pregnancy: The role of the immune system at the implantation site. Ann NY Acad Sci. 2011;1221:80–7.
- Veenstra van Nieuwenhoven AL, Heineman MJ, Faas MM. The immunology of successful pregnancy. Hum Reprod Update. 2003;9(4): 347–57.
- 57. Faas MM, de Vos P. Uterine NK cells and macrophages in pregnancy. Placenta. 2017;56:44–52.
- Fu B, Zhou Y, Ni X, et al. Natural killer cells promote fetal development through the secretion of growth-promoting factors. Immunity. 2017; 47(6):1100–13.e6.
- Costa ML, Robinette ML, Bugatti M, et al. Two distinct myeloid subsets at the term human fetal–maternal interface. Front Immunol. 2017;8: 1357.
- Vivier E, Tomasello E, Baratin M, et al. Functions of natural killer cells. Nat Immunol. 2008;9(5):503–10.
- Yougbaré I, Tai WS, Zdravic D, et al. Activated NK cells cause placental dysfunction and miscarriages in fetal alloimmune thrombocytopenia. Nat Commun. 2017;8(1):224.
- Svensson-Arvelund J, Ernerudh J. The role of macrophages in promoting and maintaining homeostasis at the fetal-maternal interface. Am J Reprod Immunol. 2015;74(2):100–9.
- 63. La Rocca C, Carbone F, Longobardi S, et al. The immunology of pregnancy: Regulatory T cells control maternal immune tolerance toward the fetus. Immunol Lett. 2014;162(1):41–8.
- 64. Wegmann TG, Lin H, Guilbert L, et al. Bidirectional cytokine interactions in the maternal-fetal relationship: Is successful pregnancy a TH2 phenomenon? Immunol Today. 1993;14(7):353–6.
- Sykes L, MacIntyre DA, Yap XJ, et al. The Th1:th2 dichotomy of pregnancy and preterm labour. Mediators Inflamm. 2012;2012:967629.
- 66. Zhang Y, Wang Y, Li MQ, et al. IL-25 promotes Th2 bias by upregulating IL-4 and IL-10 expression of decidual γδT cells in early pregnancy. Exp Ther Med. 2018;15(2):1855–62.
- El-Shazly S, Makhseed M, Azizieh F, et al. Increased expression of proinflammatory cytokines in placentas of women undergoing spontaneous preterm delivery or premature rupture of membranes. Am J Reprod Immunol. 2004;52(1):45–52.
- Makhseed M, Raghupathy R, El-Shazly S, et al. Pro-inflammatory maternal cytokine profile in preterm delivery. Am J Reprod Immunol. 2003;49(5):308–18.
- Nancy P, Erlebacher A. T cell behavior at the maternal-fetal interface. Int J Dev Biol. 2014;58(2–4):189–98.
- Vacca P, Montaldo E, Croxatto D, et al. Identification of diverse innate lymphoid cells in human decidua. Mucosal Immunol. 2015;8(2):254–64.

- Kwan M, Hazan A, Zhang J, et al. Dynamic changes in maternal decidual leukocyte populations from first to second trimester gestation. Placenta. 2014;35(12):1027–34.
- Romero R, Espinoza J, Gonçalves LF, et al. Inflammation in preterm and term labour and delivery. Semin Fetal Neonatal Med. 2006;11(5): 317–26.
- Hanna N, Hanna I, Hleb M, et al. Gestational age-dependent expression of IL-10 and its receptor in human placental tissues and isolated cytotrophoblasts. J Immunol. 2000;164(11):5721–8.
- 74. Mark PJ, Lewis JL, Jones ML, et al. The inflammatory state of the rat placenta increases in late gestation and is further enhanced by glucocorticoids in the labyrinth zone. Placenta 2013;34(7):559-66
- glucocorticoids in the labyrinth zone. Placenta. 2013;34(7):559–66. 75. Mor G, Aldo P, Alvero AB. The unique immunological and microbial aspects of pregnancy. Nat Rev Immunol. 2017;17(8):469–82.
- Erlebacher A. Immunology of the maternal-fetal interface. Annu Rev Immunol. 2013;31(1):387–411.
- Racicot K, Kwon JY, Aldo P, et al. Understanding the complexity of the immune system during pregnancy. Am J Reprod Immunol. 2014;72(2): 107–16.
- 78. Doria A, Cutolo M, Ghirardello A, et al. Effect of pregnancy on serum cytokines in SLE patients. Arthritis Res Ther. 2012;14(2):R66.
- 79. Neuteboom RF, Verbraak E, Sa Voerman J, et al. First trimester interleukin 8 levels are associated with postpartum relapse in multiple sclerosis. 2018;15(11):1356–58.
- Christian LM, Porter K. Longitudinal changes in serum proinflammatory markers across pregnancy and postpartum: Effects of maternal body mass index. Cytokine. 2014;70(2):134–40.
- Vassiliadis S, Ranella A, Papadimitriou L, et al. Serum levels of pro- and anti-inflammatory cytokines in non-pregnant women, during pregnancy, labour and abortion. Mediat Inflamm. 1998;7(2):69–72.
- Shimaoka Y, Hidaka Y, Tada H, et al. Changes in cytokine production during and after normal pregnancy. Am J Reprod Immunol. 2000;44(3): 143–7.
- 83. Somerset DA, Zheng Y, Kilby MD, et al. Normal human pregnancy is associated with an elevation in the immune suppressive CD25+ CD4+ regulatory T-cell subset. Immunology. 2004;112(1):38–43.
- Zhao J, Zeng Y, Liu Y. Fetal alloantigen is responsible for the expansion of the CD4+CD25+ regulatory T cell pool during pregnancy. J Reprod Immunol. 2007;75(2):71–81.
- Lathrop SK, Bloom SM, Rao SM, et al. Peripheral education of the immune system by colonic commensal microbiota. Nature. 2011; 478(7368):250–4.
- Gratz IK, Campbell DJ. Organ-specific and memory treg cells: Specificity, development, function, and maintenance. Front Immunol. 2014;5:333.
- Feyaerts D, Kuret T, van Cranenbroek B, et al. Endometrial natural killer (NK) cells reveal a tissue-specific receptor repertoire. Hum Reprod. 2018;33:441–51.
- Li Y, Zhang J, Zhang D, et al. Tim-3 signaling in peripheral NK cells promotes maternal-fetal immune tolerance and alleviates pregnancy loss. Sci Signal. 2017;10(498):eaah4323.
- Koldehoff M, Katzorke T, Wisbrun NC, et al. Modulating impact of human chorionic gonadotropin hormone on the maturation and function of hematopoietic cells. J Leukoc Biol. 2011;90(5):1017–26.
- Sauss K, Ehrentraut S, Zenclussen AC, et al. The pregnancy hormone human chorionic gonadotropin differentially regulates plasmacytoid and myeloid blood dendritic cell subsets. Am J Reprod Immunol. 2018; 79(4):e12837.
- Khan NA, Khan A, Savelkoul HF, et al. Inhibition of diabetes in NOD mice by human pregnancy factor. Hum Immunol. 2001;62(12):1315–23.
- Khan NA, Khan A, Savelkoul HFJ, et al. Inhibition of septic shock in mice by an oligopeptide from the beta-chain of human chorionic gonadotrophin hormone. Hum Immunol. 2002;63(1):1–7.
- van der Zee M, van den Berg JW, van Holten-Neelen C, et al. The betahuman chorionic gonadotropin-related peptide LQGV exerts antiinflammatory effects through activation of the adrenal gland and glucocorticoid receptor in C57BL/6 mice. J Immunol. 2010;185(9): 5066–73.
- van den Berg JW, Dik WA, van der Zee M, et al. The β-human chorionic gonadotropin-related peptide LQGV reduces mortality and inflammation in a murine polymicrobial sepsis model. Crit Care Med. 2011;39(1):126–34.
- 95. van der Zee M, Dik WA, Kap YS, et al. Synthetic human chorionic gonadotropin-related oligopeptides impair early innate immune

responses to Listeria monocytogenes in Mice. J Infect Dis. 2010;201(7): 1072-80.

- 96. van den Berg HR, Khan NA, van der Zee M, et al. Synthetic oligopeptides related to the β -subunit of human chorionic gonadotropin attenuate inflammation and liver damage after (trauma) hemorrhagic shock and resuscitation. Shock. 2009;31(3):285–91.
- 97. Khan NA, Susa D, van den Berg JW, et al. Amelioration of renal ischaemia-reperfusion injury by synthetic oligopeptides related to human chorionic gonadotropin. Nephrol Dial Transpl. 2009;24(9): 2701–8.
- Elmaagacli AH, Ditschkowski M, Steckel NK, et al. Human chorionic gonadotropin and indolamine 2,3-dioxygenase in patients with GVHD. Bone Marrow Transpl. 2014;49(6):800–5.
- Khil LY, Jun HS, Kwon H, et al. Human chorionic gonadotropin is an immune modulator and can prevent autoimmune diabetes in NOD mice. Diabetologia. 2007;50(10):2147–55.
- Rao CV. Potential therapy for rheumatoid arthritis and sjögren syndrome with human chorionic gonadotropin. Reprod Sci. 2016;23(5): 566–71.
- 101. Preciado-Martínez E, García-Ruíz G, Flores-Espinosa P, et al. Progesterone suppresses the lipopolysaccharide-induced proinflammatory response in primary mononuclear cells isolated from human placental blood. Immunol Invest. 2018;47(2):181–95.
- 102. Xue XT, Kou XX, Li CS, et al. Progesterone attenuates temporomandibular joint inflammation through inhibition of NF-κB pathway in ovariectomized rats. Sci Rep. 2017;7(1):15334.
- 103. Karatepe O, Altiok M, Battal M, et al. The effect of progesterone in the prevention of the chemically induced experimental colitis in rats. Acta Cir Bras. 2012;27(1):23–9.
- 104. Houdeau E, Moriez R, Leveque M, et al. Sex steroid regulation of macrophage migration inhibitory factor in normal and inflamed colon in the female rat. Gastroenterology. 2007;132(3):982–93.
- 105. Lei B, Mace B, Dawson HN, et al. Anti-inflammatory effects of progesterone in lipopolysaccharide-stimulated BV-2 microglia. PLoS One. 2014;9(7):e103969.
- 106. Straub RH. The complex role of estrogens in inflammation. Endocr Rev. 2007;28(5):521–74.
- 107. Harnish DC, Albert LM, Leathurby Y, et al. Beneficial effects of estrogen treatment in the HLA-B27 transgenic rat model of inflammatory bowel disease. Am J Physiol Liver Physiol. 2004;286(1):G118–25.
- Hajj Hussein I, Eid A, Maksoud R, et al. Estrogens control inflammation in experimental colitis. J Biol Regul Homeost Agents. 2014;28(2):213–24.
- Verdú EF, Deng Y, Bercik P, et al. Modulatory effects of estrogen in two murine models of experimental colitis. Am J Physiol Liver Physiol. 2002; 283(1):G27–36.
- 110. Kane S. Activity of IBD during pregnancy. Nat Rev Gastroenterol Hepatol. 2013;10(10):571–2.
- Khalili H, Higuchi LM, Ananthakrishnan AN, et al. Hormone therapy increases risk of ulcerative colitis but not Crohn's disease. Gastroenterol. 2012;143(5):1199–206.
- 112. Cornish JA, Tan E, Simillis C, et al. The risk of oral contraceptives in the etiology of inflammatory bowel disease: A meta-analysis. Am J Gastroenterol. 2008;103(9):2394–400.
- 113. Walker AW, Sanderson JD, Churcher C, et al. High-throughput clone library analysis of the mucosa-associated microbiota reveals dysbiosis and differences between inflamed and non-inflamed regions of the intestine in inflammatory bowel disease. BMC Microbiol. 2011;11(1):7.
- 114. Lupp C, Robertson ML, Wickham ME, et al. Host-mediated inflammation disrupts the intestinal microbiota and promotes the overgrowth of Enterobacteriaceae. Cell Host Microbe. 2007;2(2): 119–29.
- 115. Darfeuille-Michaud A, Boudeau J, Bulois P, et al. High prevalence of adherent-invasive Escherichia coli associated with ileal mucosa in Crohn's disease. Gastroenterology. 2004;127(2):412–21.
- Peterson DA, Frank DN, Pace NR, et al. Metagenomic approaches for defining the pathogenesis of inflammatory bowel diseases. Cell Host Microbe. 2008;3(6):417–27.
- 117. Sokol H, Pigneur B, Watterlot L, et al. Faecalibacterium prausnitzii is an anti-inflammatory commensal bacterium identified by gut microbiota analysis of Crohn's disease patients. Proc Natl Acad Sci. 2008;105(43): 16731–6.

- Sokol H, Seksik P, Furet JP, et al. Low counts of Faecalibacterium prausnitzii in colitis microbiota. Inflamm Bowel Dis. 2009;15(8): 1183–9.
- Manichanh C, Rigottier-Gois L, Bonnaud E, et al. Reduced diversity of faecal microbiota in Crohn's disease revealed by a metagenomic approach. Gut. 2006;55(2):205–11.
- Taurog JD, Richardson JA, Croft JT, et al. The germfree state prevents development of gut and joint inflammatory disease in HLA-B27 transgenic rats. J Exp Med. 1994;180(6):2359–64.
- Koren O, Goodrich JK, Cullender TC, et al. Host remodeling of the gut microbiome and metabolic changes during pregnancy. Cell. 2012; 150(3):470–80.
- 122. Pascal V, Pozuelo M, Borruel N, et al. A microbial signature for Crohn's disease. Gut. 2017;66(5):813–22.
- 123. Cheng L, Jin H, Qiang Y, et al. High fat diet exacerbates dextran sulfate sodium induced colitis through disturbing mucosal dendritic cell homeostasis. Int Immunopharmacol. 2016;40:1–10.
- 124. Statovci D, Aguilera M, MacSharry J, et al. The impact of western diet and nutrients on the microbiota and immune response at mucosal interfaces. Front Immunol. 2017;8:838.
- Iverson GM, Bianchi DW, Cann HM, et al. Detection and isolation of fetal cells from maternal blood using the flourescence-activated cell sorter (FACS). Prenat Diagn. 1981;1(1):61–73.
- Herzenberg LA, Bianchi DW, Schröder J, et al. Fetal cells in the blood of pregnant women: Detection and enrichment by fluorescence-activated cell sorting. Proc Natl Acad Sci USA. 1979;76(3):1453–5.
- 127. Lo YMD, Lau TK, Chan LYS, et al. Quantitative analysis of the bidirectional fetomaternal transfer of nucleated cells and plasma DNA. Clin Chem. 2000;46(9):1301–9.
- 128. Lo YM, Lo ES, Watson N, et al. Two-way cell traffic between mother and fetus: Biologic and clinical implications. Blood. 1996;88:4390–5.
- 129. Hyodo M, Samura O, Fujito N, et al. No correlation between the number of fetal nucleated cells and the amount of cell-free fetal DNA in maternal circulation either before or after delivery. Prenat Diagn. 2007;27(8): 717–21.
- Maloney S, Smith A, Furst DE, et al. Microchimerism of maternal origin persists into adult life. J Clin Invest. 1999;104(1):41–7.
- 131. Evans PC, Lambert N, Maloney S, et al. Long-term fetal microchimerism in peripheral blood mononuclear cell subsets in healthy women and women with scleroderma. Blood. 1999;93(6):2033–7.
- 132. Khosrotehrani K, Johnson KL, Cha DH, et al. Transfer of fetal cells with multilineage potential to maternal tissue. JAMA. 2004;292(1):75.
- Nelson JL. The otherness of self: Microchimerism in health and disease. Trends Immunol. 2012;33(8):421–7.
- Kinder JM, Stelzer IA, Arck PC, et al. Immunological implications of pregnancy-induced microchimerism. Nat Rev Immunol. 2017;17(8): 483–94.
- 135. Scaletti C, Vultaggio A, Bonifacio S, et al. Th2-oriented profile of male offspring T cells present in women with systemic sclerosis and reactive with maternal major histocompatibility complex antigens. Arthritis Rheum. 2002;46(2):445–50.
- Cirello V, Rizzo R, Crippa M, et al. Fetal cell microchimerism: A protective role in autoimmune thyroid diseases. Eur J Endocrinol. 2015; 173(1):111–8.
- 137. Guthrie KA, Dugowson CE, Voigt LF, et al. Does pregnancy provide vaccine-like protection against rheumatoid arthritis? Arthritis Rheum. 2010;62(7):1842–8.
- 138. Suskind DL, Kong D, Stevens AM, et al. Maternal microchimerism in pediatric inflammatory bowel disease. Chimerism. 2011;2(2):50–4.

- Boniotto M, Berti I, Santon D, et al. Absence of maternal microchimerism in very early onset inflammatory bowel disease R1. J Gastroenterol Hepatol. 2006;21(6):1082–4.
- Brenner T, Beyth Y, Abramsky O. Inhibitory effect of alpha-fetoprotein on the binding of myasthenia gravis antibody to acetylcholine receptor. Proc Natl Acad Sci USA. 1980;77(6):3635–9.
- Chakraborty M, Mandal C. Immuno-suppressive effect of human alphafetoprotein: A cross species study. Immunol Invest. 1993;22(5): 329–39.
- 142. Irony-Tur-Sinai M, Grigoriadis N, Lourbopoulos A, et al. Amelioration of autoimmune neuroinflammation by recombinant human alpha-fetoprotein. Exp Neurol. 2006;198(1):136–44.
- Dudich E. MM-093, a recombinant human alpha-fetoprotein for the potential treatment of rheumatoid arthritis and other autoimmune diseases. Curr Opin Mol Ther. 2007;9(6):603–10.
- 144. de Boer NKH, Jarbandhan SVA, de Graaf P, et al. Azathioprine use during pregnancy: Unexpected intrauterine exposure to metabolites. Am J Gastroenterol. 2006;101(6):1390–2.
- 145. Simister NE. Placental transport of immunoglobulin G. Vaccine. 2003; 21(24):3365–9.
- Clowse MEB, Wolf DC, Förger F, et al. Pregnancy outcomes in subjects exposed to certolizumab pegol. J Rheumatol. 2015;42(12):2270–8.
- 147. Zelinkova Z, De Haar C, De Ridder L, et al. High intra-uterine exposure to infliximab following maternal anti-TNF treatment during pregnancy. Aliment Pharmacol Ther. 2011;33(9):1053–8.
- 148. Mahadevan U, Wolf DC, Dubinsky M, et al. Placental transfer of anti-tumor necrosis factor agents in pregnant patients with inflammatory bowel disease. Clin Gastroenterol Hepatol. 2013;11(3): 286–92.
- Davidov Y, Ungar B, Bar-Yoseph H, et al. Association of induction infliximab levels with clinical response in perianal Crohn's disease. J Crohn's Colitis. 2016;11(5):jjw182.
- Nakase H. Editorial: Therapeutic drug monitoring for anti-TNF agentshas it all been said? Author's reply. Aliment Pharmacol Ther. 2017; 46(11–12):1114–5.
- 151. Seow CH, Leung Y, Vande Casteele N, et al. The effects of pregnancy on the pharmacokinetics of infliximab and adalimumab in inflammatory bowel disease. Aliment Pharmacol Ther. 2018;45(10):1329–38.
- Ek WE, D'Amato M, Halfvarson J. The history of genetics in inflammatory bowel disease. Ann Gastroenterol. 2014;27(4):294–303.
- 153. Hazes JMW, Coulie PG, Geenen V, et al. Rheumatoid arthritis and pregnancy: Evolution of disease activity and pathophysiological considerations for drug use. Rheumatol. 2011;50(11):1955–68.
- Petri M, Howard D, Repke J. Frequency of lupus flare in pregnancy. The Hopkins lupus pregnancy center experience. Arthritis Rheum. 1991; 34(12):1538–45.
- 155. Doria A, Cutolo M, Ghirardello A, et al. Steroid hormones and disease activity during pregnancy in systemic lupus erythematosus. Arthritis Rheum. 2002;47(2):202–9.

Open Access This is an open-access article distributed under the terms of the Creative Commons Attribution-Non Commercial-No Derivatives License 4.0 (CCBY-NC-ND), where it is permissible to download and share the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal.