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Vol. 11(5), pp. 371-378, 4 February, 2016 DOI: 10.5897/AJAR2015.10333 Article Number: C9D1DAA57053 ISSN 1991-637X Copyright ©2016 Author(s) retain the copyright of this article http://www.academicjournals.org/AJAR

African Journal of Agricultural Research

Full Length Research Paper

Morphological characterization of pearl millet hybrids [*Pennisetum glaucum* (L.) R. Br.] and their parents

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Received 22 August, 2015; Accepted 20 November, 2015

The present study was undertaken to characterize pearl millet [*Pennisetum glaucum* (L.) R. Br.] hybrids and their parents on the basis of morphological descriptors with the objective to identify key diagnostic characters of the genotypes. A set of 24 pearl millet genotypes (7 hybrids and their 17 parental lines) was evaluated in randomized block design (RBD) with three replications at two locations, Chaudhary Charan Singh Haryana Agricultural University (CCS HAU) Hisar and Regional Research Station (RRS) Bawal (Rewari) under irrigated and rainfed conditions, respectively, during kharif, 2011. Observations were recorded for 28 morphological and yield characters. All genotypes were classified into different groups based on each character. Nodal pubescence, nodal pigmentation, spike shape, spike density, spike tip sterility, sheath pubescence and spikelet glum colour distinguished all the 24 genotypes by assigning them key diagnostic features that would certainly help the plant breeders, seed growers and seed certification agencies to use these diagnostic characters. Hybrids HHB 216, HHB 226, HHB 117 could be differentiated by bristle length, spikelet glume colour and spike tip sterility. Majority of the morphological characters found to be dominant in the hybrids were contributed by the male parent.

Key words: Pearl millet, morphological descriptor, characterization, key diagnostic character.

INTRODUCTION

Pearl millet [*Pennisetum glaucum* (L.) R. Br.] is extensively cultivated for grain as well as fodder in the dry areas of north western and southern India. It is the fourth most important staple food crop after rice, wheat and sorghum primarily grown for grain and fodder production (Yadav et al., 2007). Pearl millet has an ability to grow in environments of low and erratic rainfall, high temperature and low soil fertility. Therefore, pearl millet is the main source of food and fodder for the poor farming communities which are habitant to these environments. With its ability to adopt to diverse agro ecological conditions, it occupies a unique position in the world agriculture.

Pearl millet has wide genetic diversity which is of little value unless it is characterized, evaluated and documented properly to enhance its utilization in crop improvement. In order to utilize effectively the available genetic diversity, the material must be properly

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S/N	Hybrids	Year of release	Female parent	Male parent	Maintainer
1	HHB 94	2000	ICMA 89111	G 73-107	ICMB 89111
2	HHB 117	2004	HMS 7A	H 77/ 29-2	HMS 7B
3	HHB 67 Imp	2005	ICMA 843-22	H 77/833-2-202	ICMB 843-22
4	HHB 197	2007	ICMA 97111	HBL 11	ICMB 97111
5	HHB 216	2010	HMS 37A	HTP 3/13	HMS 37B
6	HHB 223	2010	ICMA 94555	HBL 11	ICMB 94555
7	HHB 226	2010	ICMA 843-22	HBL 11	ICMB 843-22

Table 1. List of pearl millet hybrids and their parents studied.

characterized and catalogued. With the proliferation of newly developed varieties in important cultivated crops, the task of establishing the identity of these varieties and maintaining their seed lots has become a major concern. Characterization of varieties is thus of significance for the purpose of establishment and verification of identity and assessment of varietal purity for seed production and certification. Apart from this, characterization of pearl millet genotypes is also required for their protection under Plant Varieties Protection (PVP) legislation, because varietal testing for distinctness, uniformity and stability (DUS) is the basis for grant of protection of new plant varieties under the Protection of Plant Varieties and Farmers' Rights Act, 2001 (PPV and FR Act, 2001). Thus, the objective of present study was to characterize pearl millet hybrids and their parents using morphological descriptors developed as DUS descriptors (All India Coordinated Pearl Millet Improvement Project (AICPMIP), 2006).

MATERIALS AND METHODS

The present investigation was carried out at the research farm of The Department of Genetics and Plant Breeding, Chaudhary Charan Singh Haryana Agricultural University, Hisar under irrigated condition and Regional Research Station, CCS Haryana Agricultural University, Bawal (Rewari) under rainfed conditions. The experiment was conducted during the kharif season, 2011. The seed material for the present study consisted of 24 genotypes (7 hybrids, 6 CMS lines, 6 B lines, 5 restorers) of pearl millet as given in Table 1. A set of 24 genotypes was evaluated in randomized block design (RBD) with three replications under irrigated conditions. The plot size was one row of 4 m length with row to row distance of 50 cm and plant to plant 15 cm. Recommended agronomic practices were followed to raise the crop. The observations were recorded on 5 plants in each replication at specified stages of crop growth period when the characters under study had full expression. Pooled data from both locations were used to classify the genotypes. Flow charts were also prepared depicting key diagnostic characters related to genotypes.

RESULTS AND DISCUSSION

The identification of a pearl millet genotype is established by using a set of morphological characteristics. These

characteristics are also useful to establish distinctness, uniformity and stability of the new variety, based on which the variety is given protection. In the present study, 28 morphological characteristics listed in the DUS test guidelines and morphological descriptors for pearl millet were explored for varietal description of pearl millet. Among the 24 pearl millet genotypes, considerable variation was observed between the genotypes for all the important attributes under study at both locations except anthocynin coloration of first leaf sheath, number of nodal tillers, spike girth, stem thickness, leaf mid rib colour and ligule hair. All the qualitative traits, as expected, were found similar over the locations (Kumar et al., 1993). The characterization of pearl millet hybrids and their parents under study is presented in Table 2. Key diagnostic characters of each genotype are assigned through flow chart in Figure 1.

In the present study, sheath pubescence was absent in all the genotype except HMS 7A and HMS 7B. On the basis of number of productive tillers, genotypes were classified in three categories: low, medium and high, having four, sixteen and four genotypes, respectively. On the basis of anther colour, genotypes were classified in two groups: yellow and brown, all the genotypes had yellow anther colour except HHB 94, ICMB 97111 and G 73-107 which possessed brown anther colour. One hybrid (HHB 67 Improved) and eight parental lines showed absence of nodal pubescence whereas all other fifteen genotypes showed presence of nodal pubescence. Among 24 genotypes of pearl millet under study, eleven expressed green nodal pigmentation, four expressed brown nodal pigmentation, six had red nodal pigmentation, two had purple nodal pigmentation and G 73-107 exhibited whitish nodal pigmentation. All the genotypes showed green internode pigmentation except ICMA 97111 and ICMB 97111 which exhibited purple internode pigmentation. All the genotypes showed complete ear head exertion type except HMS 37A and HMS 37B (partial ear head exertion). Only HHB 117 had brown spikelet glum colour whereas all the other genotypes of pearl millet under study were having green spikelet glum colour. HHB 67 improved and its male parent H77/833-2-202 showed purple bristle colour whereas HHB 197, HHB 223, HHB 226, HHB 216 and

Plant descriptors	Expressions	No. of genotypes	Genotypes Hybrids	Parental lines
Anthocyanin coloration of first leaf sheath	Absent	24	HHB 67 Improved, HHB 197, HHB 223, HHB 226, HHB 94, HHB 117, HHB 216	ICMA 843-22, ICMB 843-22, ICMA 97111, ICMB 97111, ICMA 89111, ICMB 89111, ICMA 94555, ICMB 94555, HMS 37A, HMS 37B, HMS 7A, HMS 7B, HBL 11, H 77/833-2-202, H 77/29-2, G 73-107, HTP 3/13
	Low (2-3)	4	-	ICMA 94555, ICMB 94555, HMS 37A, HMS 37B
Productive tillers	Medium (3-6)	16	HHB 67 Improved, HHB 197, HHB 223	ICMA 843-22, ICMB 843-22, ICMA 97111, ICMB 97111, ICMA 89111, ICMB 89111, ICMB 89111, HMS 7A, HMS 7B, HBL 11, H 77/833-2-202, H 77/29-2, G 73-107, HTP 3/13
	High (>6)	4	HHB 226, HHB 94, HHB 117, HHB 216	
Nodal tillers	No tillers	24	HHB 67 Improved, HHB 197, HHB 223, HHB 226, HHB 94, HHB 117, HHB 216	ICMA 843-22, ICMB 843-22, ICMA 97111, ICMB 97111, ICMA 89111, ICMB 89111, ICMB 89111, ICMB 94555, ICMB 94555, HMS 37A, HMS 37B, HMS 7A, HMS 7B, HBL 11, H 77/833-2-202, H 77/29-2, G 73-107, HTP 3/13
	Present	2	-	HMS 7A HMS 7B
Sheath pubescence	1 TOOM	-		
	Absent	22	HHB 67 Improved, HHB 197, HHB 223, HHB 226, HHB 94, HHB 117, HHB 216	ICMA 843-22, ICMB 843-22, ICMA 97111, ICMB 97111, ICMA 89111, ICMB 89111, ICMA 94555, ICMB 94555, HMS 37A, HMS 37B, HBL 11, H 77/833-2- 202, H 77/29-2, G 73-107, HTP 3/13
	Medium(51-60)	3		H 77/833.2.202 H 77/20-2 C 73.107
Leaf length (cm)	Long (61-70)	13	ННВ 226	ICMA 843-22, ICMB 843-22, ICMA 97111, ICMB 97111, ICMA 89111, ICMB 89111, ICMB 89111, ICMB 89111, IMS 37A, IMS 37B, IMS 7A, IMS 7B, IBL 11, ITP 3/13
	Very long (>70)	8	HHB 67 Improved, HHB 197, HHB 223, HHB 94, HHB 117, HHB 216	ICMA 94555, ICMB 94555
	No. (12)	2		
	Narrow (<3)	3	-	H 77/833-2-202, G 73-107, HBL 11
Leaf width (cm)	Medium (3-4)	15	HHB 67 Improved, HHB 197, HHB 223, HHB 94, HHB 226	ICMA 843-22, ICMB 843-22, ICMA 97111, ICMB 97111, ICMA 89111, ICMB 89111, ICMB 89111, IMS 37A, IMS 37B, IMS 7A, IMS 7B
	Broad (>4)	6	HHB 117, HHB 216	ICMA 94555, ICMB 94555,HTP 3/13, H 77/29-2
				ICMA 942 22 ICMD 942 22 ICMA 07111 ICMA 90111 ICMD 90111 ICMA
Anther colour	Yellow	21	HHB 67 Improved, HHB 197, HHB 223, HHB 226, HHB 117, HHB 216	94555, ICMB 94555, HMS 37A, HMS 37B, HMS 7A, HMS 7B, HBL 11, H 77/833-2-202, H 77/29-2, HTP 3/13
	Brown	3	ННВ 94	ICMB 97111, G 73-107
Number of nodes	Low (< 11)	22	HHB 67 Improved, HHB 197, HHB 223, HHB 226, HHB 94	ICMA 843-22, ICMB 843-22, ICMA 97111, ICMB 97111, ICMA 89111, ICMB 89111, ICMA 94555, ICMB 94555, HMS 37A, HMS 37B, HMS 7A, HMS 7B, HBL 11, H 77/833-2-202, H 77/29-2, G 73-107, HTP 3/13
	Medium (11-15)	2	HHB 117, HHB 216	

Table 2. Characterization of pearl millet hybrids and their parents on the basis of morphological descriptors.

Table 2. Contd.

	Present	15	HHB 197, HHB 223, HHB 226, HHB 94, HHB 117, HHB 216	ICMA 843-22, ICMB 843-22, HMS 37A, HMS 37B, HBL 11, H 77/833-2-202, H
Nodal pubescence	Absort	٥		ICMA 97111, ICMB 97111, ICMA 89111, ICMB 89111, ICMA 94555, ICMB
	Absent	9		94555, HMS 7A, HMS 7B
Internode pigmentation	Green	22	HHB 67 Improved, HHB 197, HHB 223, HHB 226, HHB 94, HHB 117, HHB 216	ICMA 843-22, ICMB 843-22, ICMA 89111, ICMB 89111, ICMA 94555, ICMB 94555, HMS 37A, HMS 37B, HMS 7A, HMS 7B, HBL 11, H 77/833-2-202, H 77/29-2, G 73-107, HTP 3/13
	Purple	2		ICMA 97111, ICMB 97111
	Whitish	1		G 73-107
Nodal pigmentation	Green	11	HHB 67 Improved, HHB 197, HHB 223, HHB 226, HHB 117, HHB 216	ICMA 94555, ICMB 94555, HBL 11, H 77/29-2, HTP 3/13
	Brown Red Pumle	4 6 2	HHB 94 -	ICMA 843-22, ICMB 843-22, H 77/833-2-202 HMS 37A, HMS 37B, HMS 7A, HMS 7B, ICMA 89111, ICMB 89111 ICMA 97111, ICMB 97111
	Fulple	2		
Ear head exertion	Complete	22	HHB 67 Improved, HHB 197, HHB 223, HHB 226, HHB 94, HHB 117, HHB 216	ICMA 843-22, ICMB 843-22, ICMA 89111, ICMB 89111, ICMA 94555, ICMB 94555, HMS 7A, HMS 7B, HBL 11, H 77/833-2-202, H 77/29-2, G 73-107, HTP 3/13, ICMA 97111, ICMB 97111
	Partial	2	-	HMS 37A, HMS 37B
Spike length (cm)	Small (11-20)	15		ICMA 843-22, ICMB 843-22, ICMA 89111, ICMB 89111, ICMA 94555, ICMB 94555, HMS 7A, HMS 7B, HBL 11, H 77/833-2-202, H 77/29-2, G 73-107, HTP 3/13, HMS 37A, HMS 37B
	Intermediate (21-30)	9	HHB 67 Improved, HHB 197, HHB 223, HHB 226, HHB 94, HHB 117, HHB 216	ICMA 97111, ICMB 97111,
Spikelet glume colour	Green	23	HHB 67 Improved, HHB 197, HHB 223, HHB 226, HHB 94, HHB 216	ICMA 843-22, ICMB 843-22, ICMA 97111, ICMB 97111, ICMA 89111, ICMB 89111, ICMA 94555, ICMB 94555, HMS 37A, HMS 37B, HMS 7A, HMS 7B, HBL 11, H 77/833-2-202, H 77/29-2, G 73-107, HTP 3/13
	Brown	1	HHB 117	-
	Chart (< 0 E)	0	LIUD 67 Improved	11 77/022 2 202
Bristle length (cm)	Medium (0.5-2.0)	2 6	HHB 197, HHB 223, HHB 226, HHB 216	HBL 11, HTP 3/13
Spike girth (mm)	Medium (16-30)	24	HHB 67 Improved, HHB 197, HHB 223, HHB 226, HHB 94, HHB 117, HHB 216	ICMA 843-22, ICMB 843-22, ICMA 97111, ICMB 97111, ICMA 89111, ICMB 89111, ICMA 94555, ICMB 94555, HMS 37A, HMS 37B, HMS 7A, HMS 7B, HBL 11, H 77/833-2-202, H 77/29-2, G 73-107, HTP 3/13
Bristle colour	Purple Brown	2 6	HHB 67 Improved HHB 197, HHB 223, HHB 226, HHB 216	H 77/833-2-202 HBL 11, HTP 3/13

	Conical	13	HHB 223, HHB 94	ICMA 843-22, ICMB 843-22, ICMA 89111, ICMB 89111, ICMA 94555, ICMB 94555, HMS 37A, HMS 37B, HMS 7A, HMS 7B, G 73-107
Spike shape	Cylindrical	5	HHB 67 Improved HHB 197	ICMA 97111 ICMB 97111 HBI 11
	Candle	5	HHB 226 HHB 117 HHB 216	H 77/29-2 HTP 3/13
		1		H 77/833_2_2002
	Lanceolate	I	-	11777033-2-202
Spike tip sterility	Present	16	HHB 197, HHB 223, HHB 94, HHB 216	ICMA 843-22, ICMB 843-22, ICMA 89111, ICMB 89111, ICMA 94555, ICMB 94555, HMS 37A, HMS 37B, HMS 7A, HMS 7B, HBL 11, HTP 3/13
	Absent	8	HHB 67 Improved, HHB 226, HHB 117	ICMA 97111, ICMB 97111, H 77/833-2-202, H 77/29-2, G 73-107,
	Compact	9	HHB 67 Improved, HHB 197, HHB 226, HHB 117, HHB 216	ICMA 843-22, ICMB 843-22, HMS 7A, HMS 7B,
Spike density	Semi-compact	4	HHB 223, HHB 94	H 77/29-2, HBL 11
	Loose	11	-	ICMA 97111, ICMB 97111, ICMA 89111, ICMB 89111, ICMA 94555, ICMB 94555, HMS 37A, HMS 37B, H 77/833-2-202, G 73-107, HTP 3/13
	Short (101-150)	6	•	ICMA 94555, ICMB 94555, HMS 7A, HMS 7B, ICMA 843-22, ICMB 843-22,
Plant height (cm)	Medium (151-200)	10	HHB 67 Improved,	ICMA 89111, ICMB 89111, HMS 37A, HMS 37B, HBL 11, H 77/833-2-202, H 77/29-2, G 73-107, HTP 3/13
	Tall (201-250)	5	HHB 197, HHB 223, HHB 226,	ICMA 97111. ICMB 97111
	Verv tall (>250)	3	HHB 94 HHB 117 HHB 216	-
	Vory tan (* 200)	U		
Stem thickness (cm)	Medium (0.6-1.0)	24	HHB 67 Improved, HHB 197, HHB 223, HHB 226, HHB 94, HHB 117, HHB 216	ICMA 843-22, ICMB 843-22, ICMA 97111, ICMB 97111, ICMA 89111, ICMB 89111, ICMA 94555, ICMB 94555, HMS 37A, HMS 37B, HMS 7A, HMS 7B, HBL 11, H 77/833-2-202, H 77/29-2, G 73-107, HTP 3/13
Leaf mid rib colour	White	24	HHB 67 Improved, HHB 197, HHB 223, HHB 226, HHB 94, HHB 117, HHB 216	ICMA 843-22, ICMB 843-22, ICMA 97111, ICMB 97111, ICMA 89111, ICMB 89111, ICMA 94555, ICMB 94555, HMS 37A, HMS 37B, HMS 7A, HMS 7B, HBL 11, H 77/833-2-202, H 77/29-2, G 73-107, HTP 3/13
Ligule hair	Present	24	HHB 67 Improved, HHB 197, HHB 223, HHB 226, HHB 94, HHB 117, HHB 216	ICMA 843-22, ICMB 843-22, ICMA 97111, ICMB 97111, ICMA 89111, ICMB 89111, ICMA 94555, ICMB 94555, HMS 37A, HMS 37B, HMS 7A, HMS 7B, HBL 11, H 77/833-2-202, H 77/29-2, G 73-107, HTP 3/13
Soud covoring	Exposed	13	HHB 67 Improved, HHB 223, HHB 226, HHB 94, HHB 117, HHB 216	ICMA 97111, ICMB 97111, HBL 11, H 77/833-2-202, H 77/29-2, G 73-107, HTP 3/13
	Intermediate	11	HHB 197	ICMA 843-22, ICMB 843-22, ICMA 89111, ICMB 89111, ICMA 94555, ICMB 94555, HMS 37A, HMS 37B, HMS 7A, HMS 7B

Table 2. Contd.

	Low (<15)	2	-	HMS 7A, HMS 7B
Grain yield (g/plant)	Medium (16-30)	12	-	ICMA 843-22, ICMB 843-22, ICMA 89111, ICMB 89111, ICMA 94555, ICMB 94555, HMS 37A, HMS 37B, HMS 7A, HMS 7B, HBL 11, H 77/833-2-202, G 73- 107, HTP 3/13
	High (31-50)	4	HHB 67 Improved	ICMA 97111, ICMB 97111, H 77/29-2
	Very high (>50)	6	HHB 223, HHB 226, HHB 94, HHB 117, HHB 216 HHB 197	-
	Low (<200)	1		
	LOW (~200)	4	-	
	Medium (201-300)	7		ICMA 843-22, ICMB 843-22, ICMA 89111, ICMB 89111, ICMA 94555, ICMB 94555, H 77/833-2-202,
Fresh fodder weight (g/plant)	High (301-500)	11	HHB 67 Improved HHB 223, HHB 226, HHB 117, HHB 197	ICMA 97111, ICMB 97111, HBL 11, G 73-107, HTP 3/13, H 77/29-2
	Very high (>500)	2	HHB 94, HHB 216	-
	Low (<100)	2		HMS 7A, HMS 7B
	Medium (101-200)	12	-	ICMA 843-22, ICMB 843-22, ICMA 89111, ICMB 89111, ICMA 94555, ICMB 94555, HMS 37A, HMS 37B, H 77/833-2-202, H 77/29-2, G 73-107, HBL 11
Dry fodder weight (g/plant)	High (201-300)	7	HHB 67 Improved HHB 223, HHB 226, HHB 197	ICMA 97111, ICMB 97111, HTP 3/13
	Very high (>300)	2	HHB 94, HHB 216, HHB 117	-

their male parent HBL 11 and HTP 3/13 had brown bristles. Spike shape classified all the genotypes into four categories: conical, cylindrical, candle and lanceolate. H 77/833-2-202 had lanceolate spike shape, five genotype showed candle spike shape, another five showed cylindrical spike shape and the remaining thirteen were conical in spike shape. Spike tip sterility was present in sixteen genotypes and absent in the remaining eight genotypes. Nine genotypes showed compact spike density, four showed semi compact and the remaining eleven genotypes had loose spike.

Information outlined in Table 2 can effectively be used to find out distinct features of pearl millet genotypes. For example, HHB 117 was the only genotype having brown spikelet glum colour. The HMS 7A and HMS 7B were the only genotype having sheath pubescence. The HHB 94, ICMB 97111 and G 73-107 had brown anther colour. Whitish nodal pigmentation is the unique feature of G 73-107 and purple nodal pigmentation is the unique feature of ICMA 97111 and ICMB 97111. It is also found that partial ear head exertion type is



Figure 1. Flow chart diagram based on qualitative characters to identify the key diagnostic characters of pearl millet genotypes.

only found in HMS 37A and HMS 37B. H 77/833-2-202 was the only genotype having lanceolate spike shape. These are the qualitative characters and they can help in clear cut identification of particular genotype (Gupta et al., 2011).

Observations on plant height (short, medium, tall, very tall), leaf length (medium, long, very long), leaf width (narrow, medium, broad), number of nodes (low, medium), spike length (small, intermediate), bristle

length (short, medium), grain yield/plant (low, medium, high, very high), fresh fodder weight/plant and dry fodder weight/plant (low, medium, high, very high) were recorded and the data was pooled over the locations. These could be grouped into distinct classes and could be useful for varietal identification and genetic purity testing.

In the Figure 1, flow chart was prepared using qualitative characters to find out distinctive features of all

the genotypes (Rana, 2004). Nodal pigmentation was considered as primary character keeping in view the early identification of genotype during plant growth. Nodal pigmentation classified all the genotypes into five groups green, whitish, red, purple and brown. The restorer line G 73-107 could easily be distinguished at early stage of crop growth by having whitish nodal pigmentation. The ICMA 97111 and ICMB 97111 could easily be distinguished by purple nodal pigmentation. Other three nodal pigmentation classes were further separated using nodal pubescence and spike shape. At this level H easily be 77/833-2-202 could distinguished by combination of two characters, that is, nodal pigmentation (brown) and spike shape (lanceolate). In the same way, distinguishing features of all the 24 genotypes were identified through this flow chart based on seven qualitative characters (nodal pigmentation, nodal pubescence, spike shape, spike density, spikelet glume colour, spike tip sterility and sheath pubescence). For example, HHB 94 having a path of three characters spike shape (conical), spike density (semi compact) and nodal pigmentation (brown) is unique for this genotype. It means no other genotype can follow or repeat this path, so this unique path revealed the key diagnostic features of HHB 94. In this flow chart, the CMS line and maintainer line have similar pathway because these two lines are isogenic lines. For example, HMS 7A and HMS 7B have the same key diagnostic features and common path in flow charts.

It may also be concluded that majority of the qualitative/morphological characters found to be dominant in the hybrids were contributed by the male parent. Bristles (present/absent), nodal pubescence, seed covering were found as dominant characters. The study revealed that the morphological characters studied (Table 2), can be used to characterize and catalogued pearl millet genotypes. The key diagnostic characters assigned to the genotypes may also be used for maintaining their seed lots and verification of identity and assessment of varietal purity for seed production, certification and quality control.

Conflict of Interests

The authors have not declared any conflict of interests.

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