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Cytokines as a stressor: implications for depressive illness

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Abstract

Stressful events have been implicated in the provocation of depressive illness. Inasmuch as immunological challenge, and particularly cytokine administration, engender neuroendocrine and central neurochemical changes reminiscent of those provoked by psychogenic stressors, it was suggested that immune activation may also contribute to affective illness. The present report provides a brief overview of the neurochemical sequelae of acute and repeated interleukin-1 β (IL-1 β), tumour necrosis factor- α (TNF- α) and IL-2 treatment, describes some of the synergisms associated with these treatments, as well as their potential interactions with psychogenic stressors. In addition, a discussion is provided concerning the fact that cytokines, like stressors, may have time-dependent proactive effects, so that re-exposure to the treatments provoke greatly augmented neurochemical changes (sensitization). Given that the effects of cytokines are evident within hypothalamic, as well as extrahypothalamic sites, including various limbic regions, it is suggested that cytokines may impact on emotional changes, including depression.

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Introduction

Stressful events, coupled with the inability to cope adequately with such insults, may be fundamental in the provocation of affective disorders (Griffiths et al., 2000), and may exacerbate or promote physical pathologies, including those related to cardiovascular illness and immune dysfunction (Herbert and Cohen, 1993). The findings that communication may occur between the immune and central nervous systems (Blalock, 1984) prompted the proposition that depression may be influenced by immunological processes, just as psychological stressors have such an effect. While several potential routes of communication exist between the immune and central nervous systems, it has been suggested that cytokines, signalling molecules of the immune system, may act as immunotransmitters (Dunn, 2001). In this respect, cytokines engender central neurochemical changes, much like those elicited by psychological stressors (Anisman and Merali 1999; Dunn, 1993; 2001), thereby promoting affective disorders (Leonard, 2001; Maes, 1999). In addition to

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their indirect effects, central expression of several cytokines may be elicited by stressors, various insults related to brain injury, and by endotoxin challenge, and may thus impact on mood states. The present review will argue that:

- (1) Immune challenge (and particularly cytokine release), like stressful events, provokes central neurochemical changes that favour the development of affective disturbances.
- (2) Affective disturbances are more likely to emerge if an immune insult is superimposed on a background of stressful events (synergistic effects).
- (3) Stressors and cytokines may promote the sensitization of central neurochemical processes, and may thus proactively influence the response to subsequently encountered insults.

Relation between stressors and affective state

Depressive illness has been associated with antecedent stressful life events (Abramson et al., 1978; Brown and Harris, 1978; Brown et al., 1987; Monroe and Depue, 1991; Paykel, 2001), including not only major traumas, but also the accumulation of slight stressors (Kanner et al., 1981). While the development of depression may stem from the cognitive disturbances





associated with stressful events or failure experiences (Abramson et al., 1978), it is equally possible that the affective illness follows from the stressor-elicited neurochemical alterations (Anisman et al., 1991; Weiss and Simson, 1989).

The impact of stressor experiences appears to be dependent upon experiential factors, the individual's ability to cope with the stressor through behavioural means, and by the characteristics of the stressor itself (e.g. severity, chronicity, predictability). Among other things, these factors may influence the appraisal of the stressor and the behavioural response to it (Lazarus, 1993), as well as neurochemical functioning (Anisman et al., 1991). Additionally, the individual's stressor history (including early life trauma) may affect the subsequent response to a stressor, thereby influencing vulnerability to illness (Brown and Harris, 1989; Roy, 1985). In this respect, it was suggested (Post, 1992) that the neurochemical substrates of the illness may evolve over time following stressor exposure and over repeated illness episodes (sensitization effects). Thus, while early depressive episodes may be associated with stressful events, once the neurochemical systems are sensitized, even inocuous events may elicit adverse outcomes (Kendler et al., 2000; Lewinsohn et al., 1999; Solomon et al., 2000). As will be described shortly, stressors and cytokine treatments share several common effects, including the sensitization of neurochemical systems. In light of the parallel effects of these treatments, the possibility ought to be considered that activation of the inflammatory response system might also influence affective consequences of later stressor encounters.

Stress, depression and neurochemical status

Depression has been attributed to a variety of neurochemical disturbances, including alterations of norepinephrine (NE), dopamine (DA) and serotonin (5-HT), or their receptors (Maes and Meltzer, 1995; Schatzberg and Schildkraut, 1995) as well as various hormonal alterations (Plotsky et al., 1995). Coupled with the disparate symptoms of the illness and the variability in response to treatments, it is likely that depression is a biochemically heterogeneous disorder, wherein the substrates for the illness varies across individuals.

It appears that aversive events induce several of the central neurotransmitter alterations (e.g. variations of NE, DA and 5-HT turnover and levels, as well as receptor regulation) thought to subserve the depressive syndrome. Moreover, many of the variables considered important in promoting human depression

(e.g. stressor controllability, predictability, chronicity), influence stressor-provoked amine alterations in animals (Anisman et al., 1991; Weiss and Simson, 1989). These neurochemical changes may be of adaptive significance, in that they may blunt the psychological or physical impact of stressors, facilitate responses to deal with the challenge (Anisman et al., 1991), and stimulate processes which prevent excessive physiological activation (Munck et al., 1984). With continued exposure to a stressor, compensatory increases of amine synthesis may ensue, and concentrations of the amine may be increased (Anisman et al., 1991; Deutch and Roth, 1990; Herman and Cullinan, 1997; Irwin et al., 1986; Puglisi-Allegra et al., 1991). Despite the presumed adaptive consequences of the enhanced neuronal functioning, it is thought that under such conditions the wear and tear on the system may become excessive (e.g. allostatic load), increasing vulnerability to pathology (McEwen, 1998). In this respect, it may be particularly appropriate to evaluate the impact of chronic, unpredictable stressors (including psychogenic, neurogenic and systemic insults) on depressive states.

Cytokines as neuromodulators

Although cytokines are relatively large molecules, they may gain access to the brain and have direct actions on CNS processes. In particular, interleukin-1 β (IL-1 β) (Banks et al., 1989) and tumour necrosis factor- α (TNF- α) (Gutierrez et al., 1993) have saturable carriermediated transport mechanisms, which pump these cytokines into the brain. Further, entry into the brain may occur at circumventricular areas, which lack an efficient blood-brain barrier (BBB) (Banks et al., 1989; Quagliarello et al., 1991), ultimately reaching various brain nuclei through a process of volume diffusion (Konsman et al., 2000; Laflamme and Rivest, 1999; Lee et al., 1998). Interestingly, IL-1 β and TNF- α themselves may disrupt the BBB (Quagliarello et al., 1991), thereby increasing accessibility of the CNS. Parenthetically, the view was also expressed that cytokines may affect the BBB by the induction of adhesion molecules, such as ICAM-1 and VCAM-1 in the brain endothelium and astrocytes, which may guide inflammatory leucocytes into the brain parenchyma (Merrill and Benveniste, 1996). Moreover, cytokines and stressors may increase BBB permability by increasing the expression of vasoactive and inflammatory factors, such as cyclooxygenase-2 (COX-2; rate-limiting enzyme for the synthesis of the pyrogenic, prostaglandins) and histamine at cerbrovascular sites (Esposito et al., 2001; Mark et al., 2001). Additionally, stressors may influence the trafficking of cytokines into the brain parenchyma;

as such challenges may also alter BBB permeability (Esposito et al., 2001). Once present within the brain, cytokines can provoke a functional response by binding to specific receptors at hypothalamic and extra-hypothalamic brain regions (Cunningham and De Souza, 1993; Kinouchi et al., 1991; Laflamme and Rivest, 1999; Schobitz et al., 1994) or by other processes that stimulate neurotransmitter functioning. Indeed, both IL-1 and TNF- α may have functions similar to classical neurotransmitters through their modulation of neuronal Ca²⁺ channels and activation of intracellular second-messenger systems (Tancredi et al., 1992).

In response to systemic insults, cytokines and other immune factors (such as endotoxin) can bind to the endothelium of the brain microvasculature, which will in turn produce signalling mediators, such as histamine, nitric oxide (NO), nuclear factor kappa B (NFκB; a signal transduction protein common to many cytokines, including IL-1 β and TNF- α) and COX-2 (Chao et al., 1995; O'Connor and Coogan, 1999; Rivest et al., 2000). Indeed, TNF- α was reported to increase histamine, NFkB and COX-2 expression as well as prostaglandin release at cerebral endothelial capillaries (Blais and Rivest, 2001; Igaz et al., 2001; Mark et al., 2001). These mediators may be responsible for the altered synthesis/activity of other cytokines, neurotransmitters and hormones as well as central metabolic processes. For instance, elevated central prostaglandins may stimulate hypothalamic-pituitary-adrenal (HPA) activity and provoke febrile responses (Parsadaniantz et al., 2000; Roth et al., 2002). Additionally, through induction of these factors, and perpetuation of the inflammatory response, pro-inflammatory cytokines may influence cellular plasticity and neurodegeneration. In fact, chronic stressors have recently been suggested to promote neurodegeneration through activation of a TNF- α cascade involving increased NF κ B expression and downstream activation of NO (Madrigal et al., 2002).

In addition to direct actions, cytokines may influence CNS processes indirectly through stimulation of afferent fibres of the vagus nerve (Dantzer et al., 1996; Maier and Watkins, 1998; Watkins et al., 1995). Receptors for IL-1 are present on the nodose ganglion which sends afferent projections to the brainstem nucleus tractus solitarius (NTS), and following cytokine or endotoxin challenge, *c-fos* expression is elevated in these regions (Ek et al., 1998; Gaykema et al., 1998). It appears that activation of vagal branches may stimulate the de-novo synthesis of cytokines within brainstem and hypothalamic nuclei (Dantzer et al., 2001; Gaballec et al., 1995; Hopkins and Rothwell, 1995).

Central cytokine distribution and the impact of varied challenges

Although their levels in brain are admittedly low under basal conditions, cytokines and their receptors are endogenous to the brain, having been identified within neuronal cell bodies, microglia and astrocytes (Cunningham and De Souza, 1993; Kinouchi et al., 1991; Laflamme and Rivest, 1999; Nistico and De Sarro, 1991). Levels of cytokines and their receptors are influenced by various stressors as well as by immunological and neurological insults. For instance, IL-1 β protein levels were increased in several brain regions in response to stressors (Nguyen et al., 1998, 2000). Moreover, elevated IL-1 β , IL-6 and/or TNF- α mRNA was observed in the CNS following immobilization stress (Minami et al., 1991; Yabuuchi et al., 1996), systemic lipopolysaccharide (LPS) administration (Ban et al., 1992; Breder et al., 1994; Buttini and Boddeke, 1995; Gaballec et al., 1995; Gatti and Bartfai, 1993; Laye et al., 1994; Liu et al., 1996), central LPS injection (De Simoni et al., 1995; Quan et al., 1994; Rajora et al., 1997), viral infection (Sato et al., 1997), brain injury, tumours, cerebral ischaemia and seizure (Buttini et al., 1994; Giulian and Robertson, 1990; Hopkins and Rothwell, 1995; Ilyin et al., 1999; Minami et al., 1990; Rothwell and Hopkins, 1995; Taupin et al., 1993; Yabuuchi et al., 1993). Further, peripherally administered cytokines may even stimulate their own expression within the brain, thus potentially placing these molecules in the vicinity of their receptors (Butinni and Boddeke, 1995; Cunningham and De Souza, 1993). Parenthetically, it will be recognized that some of the aforementioned insults were essentially of an acute nature, while others involved chronic repercussions. At present, insufficient information exists concerning the relative effects of acute and chronic stressors on central cytokine activity.

Elevated central cytokine levels in response to insults have been associated with neurological and behavioural changes, including somnolence (Krueger et al., 1998), cognitive disturbances (Fiore et al., 1996), and disruptions of eating as well as cachexia (Plata-Salamán, 1998). Similarly, these signs are evident in response to exogenous central cytokine application. The severity of these signs depends on the nature, chronicity and magnitude of the challenge, all of which will influence central levels of cytokines.

In considering the central actions of the pro-inflammatory or stimulatory cytokines, it should be considered that anti-inflammatory or inhibitory signals are also present, including IL-4, IL-10 (Szczepanik et al., 2001), IL-1 receptor antagonist (IL-1Ra) (Licinio and Wong, 1997) as well as soluble receptors for IL-1 β (Colotta et al., 1994) and TNF- α (Shohami et al., 1999). It is not the level of pro-inflammatory cytokine, per se, that determines the impact of cytokine activation in the brain, but the balance between the pro- and antiinflammatory signals (Licinio and Wong, 1997; Plata-Salaman et al., 1998). Additionally, since most stimulatory cytokine receptor-bearing cells need only a few ligands to become biologically active, a substantial accumulation of their inhibitory counterparts will be needed to suppress the activating signal. In effect, when evaluating immune insults, it ought to be considered that the magnitude and chronicity of the challenge will impact on the neurological manifestations associated with cytokine activation. Highly toxic challenges (e.g. LPS) and chronic ailments (e.g. tumours) will shift the balance toward the pro-inflammatory cytokines and promote more detrimental consequences (Plata-Salaman et al., 1998), while milder and acute challenges (e.g. single, low doses of a cytokine) will shift the balance toward the anti-inflammatory cytokines and have more subdued repercussions.

Cytokine-induced activation of the HPA axis

Like stressors, various immunogenic and cytokine challenges (e.g. LPS and IL-1 β) stimulate HPA activity (Brebner et al., 2000; Del Rey and Besedovsky, 1992; Dunn, 1992; Turnbull and Rivier, 1999). Systemic and intracerebroventricular (i.c.v.) IL-1 β administration increased c-fos expression within the paraventricular nucleus (PVN) of the hypothalamus corticotropinreleasing hormone (CRH) neurons (Ericsson et al., 1994; Vellucci et al., 1995) and increased the expression of CRH and arginine vasopressin (AVP) secretion in PVN neurons of the hypothalamus (Lee and Rivier, 1994; Mandrup-Poulsen et al., 1995; Tilders et al., 1993). Moreover, IL-1 β infusion into the median eminence (site of CRH terminals from neurons originating within the PVN) increased AVP and CRH secretion and provoked elevated plasma adrenocorticotropic hormone (ACTH) and corticosterone levels (McCoy et al., 1994; Watanobe and Takebe, 1993). Predictably, CRH antagonists attenuated the IL-1 β -induced ACTH changes (Saperstein et al., 1992). While, the effects of IL-1 β on HPA activity have primarily been assessed in acute preparations, continuous systemic infusion (via osmotic minipumps) of this cytokine over 1 wk provoked a persistent increase of plasma ACTH and corticosterone levels (Sweep et al., 1992). Moreover, as will be discussed later, acute and chronic systemic IL-1 β administration may result in a long-lasting increase of CRH and AVP co-expression within the external zone of the median eminence, and thus may have protracted

repercussions with respect to the impact of later challenges (Tilders and Schmidt, 1998). Finally, in addition to IL-1 β , HPA activity was increased by other proinflammatory cytokines, including IL-6 and TNF- α (Bernardini et al., 1990; Brebner et al., 2000; Dunn, 2001; Hayley et al., 1999; Rothwell and Hopkins, 1995; Zhou et al., 1996), and these effects were blocked by CRH antiserum (Bernardini et al., 1990; Turnbull et al., 1997).

Central neurochemical effects elicited by activation of the inflammatory response system or cytokine challenge

Although the central neurotransmitter effects of various cytokines have been assessed, greatest attention has been devoted to the impact of IL-1 β . Following its systemic administration, IL-1 β stimulated c-fos expression in several stressor-sensitive brain regions, including the PVN, bed nucleus of the stria terminalis, and central nucleus of the amygdala (Day et al., 1999; Ericsson et al., 1997; Xu et al., 1999). Moreover, this treatment increased NE activity within the PVN, medial basal and lateral hypothalamic nuclei (Dunn, 2001; Kaur et al., 1998; Lacosta et al., 1998a,b), and increased DA utilization within the hypothalamus and prefrontal cortex (Kabiersch et al., 1988; Masana et al., 1990) and 5-HT activity within the hypothalamus, prefrontal cortex and hippocampus (Brebner et al., 2000; Carmelia et al., 1991; Dunn, 2001; Zalcman et al., 1994). In vivo, systemic IL-1 β administration increased hypothalamic NE and 5-HT release from the nucleus accumbens and the hippocampus, respectively (Merali et al., 1997; Smagin et al., 1996; Song et al., 1999).

Paralleling the actions of systemic treatment, i.c.v. IL-1 β increased hippocampal 5-HT release (Linthorst et al., 1995), while direct application of IL-1 β into the rat anterior hypothalamus increased the release of NE, 5-HT and DA (Shintani et al., 1993). Similarly, when directly injected into the medial basal hypothalamus, IL-1 β augmented 5-HT and DA release (Mohankumar and Quadri, 1993; Mohankumar et al., 1993), and local injection of IL-1 β increased NE release within the medial prefrontal cortex (Kamikawa et al., 1998).

Like IL-1 β , systemic TNF- α administration also increased central monoamine activity, including NE and 5-HT activity within the PVN, central amygdala, locus coeruleus and prefrontal cortex (Hayley et al., 1999), and altered tryptophan levels within the hippocampus and hypothalamus (Dunn, 2001; Leonard, 2001). Similarly, i.c.v. TNF- α stimulated amine turnover, particularly within hypothalamic nuclei (PVN and

median eminence/arcuate nucleus) (Hayley et al., In Press).

Most of the studies described thus far have pointed to cytokine treatment provoking increase monoamine release. Yet, it will be recognized that depressive illness has typically been considered to reflect a downregulation of monoamine functioning. However, two issues need to be considered in relating the effects of cytokine treatments to depression. First, in addition to affecting monoamine release, it was reported that, in vitro, IL-1 provokes activation of the 5-HT transporter, thus enhancing reuptake of 5-HT from the synaptic cleft (Ramamoorthy et al., 1995). Thus, the availability of 5-HT may be diminished, depending on the cytokine dosage.

Secondly, the aforementioned studies assessed the consequences of acute cytokine administration. There is presently insufficient information available concerning the impact of chronic cytokine treatment on neuroendocrine and central neurotransmitter functioning, and limited data are available concerning the proactive effects of cytokines on neurochemical activity. This is particularly critical, as immune activation stemming from bacterial or viral insults, as well as cytokine immunotherapy, involve sustained and persistent alterations of cytokine activity. Thus, to obtain a more realistic index of the effects relevant to depression, it may be more productive to assess the impact of chronic activation of the inflammatory response system or repeated administration of cytokines. It has been shown that sustained systemic or i.c.v. IL-1 β administration provokes marked and persistent HPA activation (Sweep et al., 1992; Van der Meer et al., 1996) while chronic central IL-1 β administration promotes activation of hypothalamic-pituitary-gonadal axis (Rivest et al., 1993). Indeed, chronic IL-1 β administration promoted sustained variations of CRH, CRH receptors and pro-opiomelanocortin gene expression coupled with elevated secretion of ACTH, β -endorphin and corticosterone (Parsadaniantz et al., 1997). Importantly, it was also shown that continuous intravenous infusion of IL-1 β not only augmented c-fos expression within specific hypothalamic nuclei (PVN and supraoptic nucelus), but also provoked such effects within the central amygdala. While the hypothalamic effects were attenuated by pretreatment with a cyclo-oxygenase inhibitor, this was not the case within the central amygdala (Niimi et al., 1996). Thus, hypothalamic and amygdala alterations likely involve different mechanisms. At any rate, the finding that chronic cytokine treatments may have protracted repercussions is consistent with the elevated HPA activity evident in some forms of depression. However, it is still not certain

what immediate or protracted effects are induced by chronic cytokine treatments on the activity of monoamines that are believed to contribute to affective illness.

Owing to the redundancies and pleiotropic nature of cytokine networks, it is important not only to consider their individual effects, but also to determine the interactive effects of cytokines on central processes. In particular, cytokines may act in a synergistic fashion, such that their co-administration results in effects greater than the sum of their individual effects. In fact, co-administration of either TNF- α and IL-1 β or IL-6 and IL-1 β provoked a synergistic increase of HPA, but not central monoamine, activity (Brebner et al., 2000; Perlstein et al., 1993; Zhou et al., 1996). Interestingly, cytokines and stressors may also have synergistic effects, as observed with respect to the in-vivo release of 5-HT from mesolimbic brain sites (Merali et al., 1997). In particular, it has been shown that the increased invivo NE and 5-HT release within the hippocampus and nucleus accumbens elicited by IL-1 β was greatly augmented following the application of a mild stressor (air puff) (Merali et al., 1997; Song et al., 1999). Moreover, in chronically stressed animals the administration of LPS promoted a greater IL-1 response, although this effect was not necessarily accompanied by elevated ACTH or corticosterone secretion (Mekaouche et al., 1994). Nevertheless, it seems reasonable to suppose that synergisms may occur between stressor and cytokine treatments, as they do between different cytokines, so that the response of endogenous neurochemical systems will be exaggerated. Incidentally, it might be noted at this juncture that the behavioural effects of cytokine treatments may be altered by environmental stimuli, particularly those contextual cues that involve a stressful component. For instance, reactivity in response to the T-cell superantigen, staphylococcal enterotoxin B, was enhanced by exposure to novel stimuli (Kawashima and Kusnecov, 2002), and IL-2 similarly provoked behavioural activation in response to a novel stimulus (Zalcman et al., 1998, Zalcman, 2001), and increased locomotor activity under anxietyprovoking conditions (on a plus maze) (Petitto et al., 1997). Similarly, the sickness-inducing effects elicited by cytokines, such as IL-1, may not be evident in novel environments, particularly when this environment was stressful (Lacosta et al., 1999). Thus, we argued that while cytokines could produce marked illness, in a novel environment that may signal danger, animals do not have the luxury of expressing sickness behaviours. In a like fashion, one can imagine that the anhedonic or depressogenic action of cytokines may be contextdependent.

As indicated earlier, it has been proposed that depressive mood results when monoamine utilization exceeds synthesis, resulting in insufficient amine concentrations to meet demands exerted by new or ongoing stressors. In a similar fashion, it ought to be considered that following chronic activation of the inflammatory response system the effects of stressors may be augmented, and moreover in chronically stressed animals the central effects of cytokine or immune challenges may be appreciably enhanced, thereby favouring the development of mood disturbances.

The role of cytokines in the stress response: immune activation as a stressor

As already indicated, 'processive' stressors (i.e. psychogenic or neurogenic stimuli or events that involve appraisal processes or higher order sensory cortical processing) share several effects with those elicited by systemic (metabolic) insults, such as bacterial or viral infection (Herman and Cullinan, 1997). While, different neural circuits are activated in response to processive and systemic stressors, these insults trigger some common end points, such as HPA activation. It was suggested that while processive stressors engender such outcomes via activation of limbic mechanisms, systemic stressors may have more direct effects through actions at the hypothalamus (Herman and Cullinan, 1997). Indeed, exposure to a novel environment (processive stressor) increased c-fos mRNA within limbic regions (e.g. lateral septum and medial amygdaloid nucleus) to a much greater extent than did inhalation of ether vapours (systemic stressor), although both stressors had comparable effects within the hypothalamus (Emmert and Herman, 1999). Further, while footshock and cytokine challenge were both shown to increase c-fos within the PVN, catecholamine denervation was only effective in attenuating the effects of the cytokine treatment (Li et al., 1996). Similarly, the HPA-stimulating effects of IL-1 β were attenuated by ablation of aminergic fibres originating from the medulla or removal of the area postrema (a brainstem, circumventricular organ) (Ericsson et al., 1994; Lee et al., 1998). Interestingly, Shintani et al. (1995) demonstrated that pretreatment with IL-1Ra, blocked the hypothalamic NE alterations and the plasma ACTH increases induced by immobilization, suggesting that cytokines may play a role in the regulation of the stress response elicited by processive types of stressors.

In relating the effects of stressors and cytokines to emotional states, it is important to consider the effects on limbic regions that have been implicated in anxiety and depression (e.g. the amygdala and prefrontal cortex). It appears likely that different nuclei of the amygdala, and particularly CRH neuronal activity within these sites, contributes to the maintenance of anxiety (LeDoux, 2000). In this respect, the basolateral nucleus of the amygdala may play a prominent role in the initial processing of fearful stimuli, while the central nucleus may be more important for the generation of behavioural outputs to contend with the challenge (Davis, 1992). Consistent with a role of IL-1 β in promoting anxiety-related responses (Anisman and Merali, 1999), it was reported that as in the case of psychogenic stressors, IL-1 β increased CRH mRNA expression at the amygdala and bed nucleus of the stria terminalis (Day et al., 1999; Lee and Rivier, 1998; Makino et al., 1999; Sawchenko et al., 1996). Moreover, lesions of the central amygdala attenuated the plasma corticosterone and ACTH responses elicited by IL-1 β , supporting the possibility that amygdaloid-PVN communication may be important in regulating cytokineelicited HPA responses (Xu et al., 1999).

While these data point to the similarity between the effects of stressors and immunogenic stimuli, it seems that stressors and cytokines may activate different regions of the amygdala. Ericsson et al. (1994) reported profound c-fos activation of the central amygdala in response to footshock, while systemic IL-1 β provoked the strongest effect within the medial nucleus. Little information exists concerning the role of the medial amygdala in the response to cytokines, relative to those elicited by stressors; however, it was suggested that the medial nucleus may play a more important role than the central nucleus in the neuroendocrine response to restraint stress (Davas et al., 1999). Given that cytokines and stressors may contribute differentially to amygdaloid stimulation, these challenges may affect different phases of anxiety/fear responses.

Although cytokines influence amygdala neuronal activity, there is limited information concerning the anxiogenic effects of cytokine treatments. The i.c.v. administration of IL-1 β or TNF- α elicited anxiogenic-like effects, but this has been evaluated in only a limited number of situations (Connor et al., 1998). Moreover, it is not clear that the anxiety-like effects associated with IL-1 β treatment stemmed from the central actions elicited by the cytokine, and instead may have been secondary to the malaise engendered by the treatment.

Cytokine and stressor sensitization effects

In addition to their immediate behavioural and neurochemical effects, stressful events may influence the organism's responses to later challenges. In fact, a variety of stressors, including immunological stimuli, prime biological systems so that an augmented response is elicited by later exposure to the same or somewhat different challenge (sensitization) (Anisman et al., 2001; Tilders and Schmidt, 1998). As indicated earlier, the development of a stressor-elicited sensitization effect is believed to have important repercussions for behavioural pathology, particularly with respect to the recurrence of depression (approx. 30–50% of patients suffer recurrence within 1 yr). Given that cytokines may induce neurochemical effects similar to those provoked by processive stressors, it ought to be considered that cytokine alterations (and sensitization to the effects of cytokines) might similarly contribute to depressive episodes.

Stressor effects

Stressor-provoked sensitization effects have been demonstrated with respect to NE activity within the hypothalamus, hippocampus and amygdala, and DA utilization within the prefrontal cortex (Anisman et al., 1991; Finlay et al., 1997; Jordan et al., 1994). Also, cross-sensitization has been demonstrated wherein exposure to a particular stimulus enhances the response to a subsequently applied stimulus of a different form, including pharmacological challenges (Anisman et al., 1993; Finlay et al., 1997; Flores et al., 2000; Gresch et al., 1994; Hayley et al., 1999; Tilders and Schmidt, 1998). Like acute challenges, chronic stressors influence responses to subsequent stressor encounters. In animals exposed to a chronic cold stressor regimen, later application of tail shock augmented NE release within the hippocampus and prefrontal cortex and DA release from cortically projecting neurons (Gresch et al., 1994; Jedema et al., 1999; Nisenbaum and Abercrombie, 1992).

As in the case of amine variations, stressors may have protracted effects on HPA functioning. It has been suggested that such changes stem from phenotypic alterations of CRH terminals within the median eminence, wherein co-localization of AVP and CRH occurs (Bartanusz et al., 1993). Specifically, a chronic stressor regimen has been shown to provoke a progressive increase of AVP stores in these CRH terminals (Bartanusz et al., 1993; de Goeij et al., 1992a,b; Schmidt et al., 1995). As CRH and AVP synergistically stimulate pituitary ACTH release, the neuroendocrine response to subsequent challenges ought to be increased.

Cytokine sensitization: HPA effects

Cytokines, such as IL-1 β and TNF- α , provoke behavioural and neurochemical sensitization effects, just as traditional stressors do. These cytokines provoke the sensitization of HPA activity such that later re-exposure to the cytokine resulted in augmented neuropeptide and hormonal activity. Interestingly, the emergence the sensitization was dependent on the passage of time following the initial challenge (Hayley et al., 1999, 2001a; Schmidt et al., 1995). In particular, while the cytokines increased the co-localization of CRH and AVP within the median eminence, this outcome only became apparent 4 d after IL-1 β administration and peaked 1–2 wk following the treatment (Schmidt et al., 1995). A second administration of IL-1 β (11 d after the initial challenge) elevated plasma ACTH and corticosterone levels (Schmidt et al., 1995), suggesting that the cytokine provoked a functionally hyper-responsive HPA axis.

Systemic administration of TNF- α also induced a time-dependent sensitization of HPA activity. However, the temporal profile of induction of the hypothalamic peptide immunoreactivity did not parallel the expression of the corticosterone sensitization. Specifically, while median eminence CRH and AVP immunoreactivities were maximally elevated 7-14 d following the initial TNF- α injection, the neuropeptide expression was comparable to baseline by 28 d (Hayley et al., 2001a). However, re-exposure to TNF- α 28 d following initial administration of the cytokine provoked a pronounced sensitization of corticosterone release, an effect that was absent at earlier re-exposure intervals (1, 7 and 14 d) (Hayley et al., 1999). Thus, factors in addition to hypothalamic CRH and AVP may influence the TNF- α -induced hormonal sensitization. For instance, the cytokine may have had direct actions upon the adrenal or pituitary gland, both of which contain high TNF- α receptor densities (Kobayashi et al., 1997). Similarly, other peripherally acting inflammatory factors (e.g. histamine) may mediate some of the sensitizing effects of TNF- α . Indeed, we recently observed that systemic antihistamine treatment (H1 and H2 antagonists) ameliorated both the corticosterone and sickness sensitization effects of TNF- α (Kelly et al., 2001). As well, in animals treated with TNF- α , later i.c.v. re-exposure to the cytokine did not sensitize HPA activity, irrespective of whether the initial injection was centrally or peripherally administered, suggesting that peripheral factors and/or targets are involved (Hayley et al., 2002).

It is important to note that mice displaying the TNF- α -induced HPA sensitization also showed signs of marked illness (e.g. ptosis, piloerection, lethargy, cyanosis of the extremities) (Hayley et al., 2001b). Although it is tempting to speculate that stress or other factors associated with the illness may be related to the provocation of the corticosterone sensitization, we

observed that that the two processes were independent of one another (Hayley et al., 2001b).

Cytokine sensitization: central monoamine alterations

In addition to the immediate neurotransmitter effects, re-exposure to TNF- α augmented monoamine activity in a region-specific and time-dependent fashion (Hayley et al., 1999, 2002). Within the PVN, TNF- α induced a sensitization of NE activity, which followed a time-course similar to that observed with respect to the corticosterone and sickness sensitization effects, becoming progressively more pronounced at longer intervals following initial cytokine treatment (Hayley et al., 1999). In contrast, the sensitization of NE activity within the prefrontal cortex and central amygdala was evident at 1 d following the initial treatment with the cytokine, but not at longer intervals. However, a TNF- α provoked sensitization of 5-HT activity was apparent within the central amygdala and prefrontal cortex upon re-exposure to the cytokine after an intermediate interval (7-14 d) following pretreatment (Hayley et al., 1999), paralleling the increased CRH-AVP co-localization within the median eminence (Hayley et al., 2001a).

Although i.c.v. TNF- α did not sensitize corticosterone activity, this treatment did elicit a sensitization of NE and DA utilization within the PVN and median eminence/arcuate nucleus complex (ME/ARC). Moreover, mice that initially received intraperitoneal (i.p.) mTNF- α and later challenged via i.c.v. administration with this cytokine displayed greatly increased 5-HT and DA activity within the ME/ARC, as well as augmented NE activity within the locus coeruleus. Interestingly, unlike the effects of peripheral cytokine administration, when administered i.c.v., the sensitization occurred largely within hypothalamic nuclei (Hayley et al., 2002).

Cytokines and depression

Major depression in humans, particularly in patients presenting with severe melancholic illness, has been associated with variations of immune functioning, including a reduction of mitogen-stimulated lymphocyte proliferation and reduced natural-killer (NK) cell activity (Herbert and Cohen, 1993; Irwin, 1999; Maes, 1995, 1999). Contrary to the position that depression was associated with the suppression of non-specific immunity, the view was advanced that affective disturbances may be secondary to activation of the inflammatory immune response (Licinio and Wong, 1997; Maes, 1995, 1999). In this respect, depressed patients were found to present with signs of immune activation reminiscent of an acute phase response, including increased plasma concentrations of complement proteins, C3 and C4, IgM, and positive acute phase proteins, haptoglobin, α_1 -antitrypsin, α_1 and α_2 macroglobulin, whereas negative acute phase proteins were reduced (Maes, 1999; Nieto et al., 2000; Rothermundt et al., 2001; Sluzewska, 1999). Further, major depressive illness was accompanied by elevated activated T cells $(CD_{25} + and HLA-DR+)$, secretion of neopterin, prostaglandin E2 and thromboxane (Maes, 1995, 1999). Parenthetically, although cortisol has often been shown to be immunosuppressive, the changes in immune cell populations are correlated with elevated urinary cortisol levels (Maes et al., 1994). Interestingly, in a dexamethasone suppression test, non-supressors were also resistant to the immune effects of dexamethasone (Maes et al., 1994), suggesting that the absence of negative feedback by cortisol on the HPA axis in depression could extend to the dysregulation in the levels of immune markers observed in the condition.

Commensurate with the view that cytokines are fundamental in depression, it was reported that severe affective illness was accompanied by elevated circulating cytokines or their soluble receptors, including IL-2, soluble IL-2 receptors (sIL-2R), IL-1 β , IL-1Ra, IL-6, soluble IL-6 receptors (sIL-6R), and γ -interferon $(\gamma$ -IFN) (Berk et al., 1997; Frommberger et al., 1997; Maes, 1995, 1999; Mullar and Ackenheil, 1998; Nassberger and Traskman-Bendz, 1993; Sluzewska et al., 1995; Smith, 1991; Song et al., 1994) as well as increased production of IL-1 β , IL-6 and TNF- α in response to mitogen challenge (Anisman et al., 1999a, b; Maes, 1995, 1999). Interestingly, with alleviation of depression in response to antidepressant treatment, normalization was evident with respect to levels of IL-1 β , IL-6 and α_1 -acid glycoprotein (Frommberger et al., 1997; Sluzewska et al., 1995), whereas no such changes were apparent concerning the up-regulated production of sIL-2R, IL-6 and sIL-6R in major depression (Maes, 1999). Similarly, antidepressant treatment did not diminish the elevated serum levels of the IL-6, or that of anti-inflammatory cytokines, IL-10 and IL-1Ra (Kubera et al., 2000). In effect, these cytokines may act simply as trait markers of the illness (Anisman et al., 1999b; Maes, 1999), although it ought to be considered that sustained treatment may be necessary to achieve normalization of cytokine functioning.

The aforementioned studies, in the main, are correlational, and thus it cannot be determined whether the cytokine elevations are secondary to the illness (i.e. being directly or indirectly brought on by the depression), or contribute to the provocation of the disorder. Yet, it has been reported that in humans undergoing immunotherapy high doses of IL-2 and IFN- α induce neuropsychiatric symptoms, including depression. As such, these data imply that the affective changes were related to the cytokine treatment (Capuron et al., 1998; Caraceni et al., 1992; Denicoff et al., 1987; Maes et al., 2001; Meyers and Valentine, 1995). Importantly, it was reported (Musselman et al., 2001) that depressive symptoms provoked by IFN- α were attenuated by treatment with the selective serotonin reuptake inhibitor, paroxetine. Together, these findings provide good reason to suspect an aetiological role for cytokines in depressive illness. Yet, it is important to consider that the populations being assessed in the latter studies were undergoing considerable strain (e.g. the distress regarding their illness), and hence depression may reflect the interactive effects of the distress and the cytokine treatments.

TNF- α signalling and depression

Recent clinical studies have suggested that elevated circulating TNF-a or its p55 receptor may be associated with psychiatric illness (Maes, 1999) and such an effect could come about by virtue of the cytokine's effects on central monoamine turnover (Ignatowski et al., 1997). Indeed, antidepressant medication has been reported to alter levels of TNF- α in brain regions such as the hippocampus and locus coeruleus (Ignatowski et al., 1997). Commensurate with the proposition that TNF- α may be involved in stressor- or depressive-like states, we found that TNF- α sensitized expression of the immediate early gene, c-fos, in several stressor-sensitive brain regions. Specifically, re-exposure to the cytokine 7 or 14 d following its initial i.p. pretreatment resulted in a pronounced increase of Fos immunoreactive protein within the PVN, supraoptic nucleus and central amygdala.

The fact that TNF- α sensitized 5-HT activity within stressor-sensitive brain regions (Hayley et al., 1999, 2002) may reflect cytokine-provoked neuroplastic changes, which may be reminiscent of those thought to characterize clinical depression (Altar, 1999; Manji et al., 2001). Further to this point, recent studies suggest that major affective disorders probably involve alterations in pathways typically activated by cytokines (Chen et al., 2001). In particular, the mitogen-activated protein (MAP) kinase pathway, which is involved in cytokine signalling [e.g. IL-6, brain-derived neurotrophic factor (BDNF)], as well as pathways involving the cAMP response-binding element (CREB) factor, which has important neuroplastic effects, are both proposed to play a role in the long-term neurochemical changes evident in depression (Guillin et al., 2001). Indeed, while stressors (which may be associated with the provocation of depression) reduce the expression of neurotrophic products of these pathways, antidepressants stimulate these trophic factors. For instance, chronic antidepressant treatments increased the central expression of cAMP and CREB and some of their target genes, including the neurotrophic cytokine, BDNF (Vaidya and Duman, 2001). Moreover, stressorinduced reductions of central BDNF levels were ameliorated by antidepressant pretreatment (Vaidya and Duman, 2001). Supporting a direct beneficial role for neurotrophic cytokines in depression, infusion of either BDNF or the closely related neurotrophin-3 (NT-3) into the dentate gyrus produced an antidepressant effect, as assessed in animal models of depression (forced swim and learned helplessness), that was comparable to that provoked by traditional chemical antidepressants (Shirayama et al., 2002). In human post-mortem studies, increased hippocampal BDNF expression was associated with chronic antidepressant regimens (Chen et al., 2001). Thus, it may be useful to consider alternate antidepressant treatments that affect BDNF or other trophic factors that may engender a certain degree of protection or resilency of monoaminergic cells in the face of everyday stressors. In particular, it is significant that the recent report by Shirayama et al. (2002) indicated that a single bilateral infusion of BDNF had antidepressant effects similar to that provoked by a chronic antidepressant regimen. The possibility exists that traditional antidepressants come to exert their beneficial effects only after neutrophic factors and related messenger pathways have been sufficiently 'primed'. If this is the case, then alternate neurotrophic antidepressants may circumvent the problem of long-term daily administration required for traditional antidepressant efficacy.

Stimulation of these cAMP and MAPkinase-dependent pathways can also induce the anti-apoptotic factor, bcl-2, which has been suggested to have beneficial actions in depressive disorders (Chen et al., 2000; Manji et al., 2001), just as it has a neuroprotective role in neurodegenerative states (Guillin et al., 2001). With respect to cytokines and depression, it appears that TNF- α provokes bcl-2 expression, as well as proapoptotic factors, suggesting that a delicate balance between the expression of death regulatory signals may ultimately determine the consequences of the cytokine, with respect to cellular survival and resiliency. It may be the case that TNF- α priming induces changes in these intracellular pathways (e.g. phosphorylation states) that 'set the stage' for the aberrant neurochemical responses to subsequent challenges that impinge upon this sensitized system.

Conclusions

It seems likely that behavioural and neurochemical plasticity is essential for an animal to cope with environmental demands. Various challenges, including stressful and immunological stimuli, may provoke central neurochemical alterations that may be of adaptive significance. Yet, some of these effects may also increase vulnerability to behavioural disturbances, particularly mood disorders. Further, it appears that both stressors and activation of the inflammatory response system may prime biological systems so that augmented responses are elicited upon later challenges with either the same or somewhat different stimuli (Tilders and Schmidt, 1998). As a result, these treatments do not only have immediate repercussions, but may exert long-lasting neurochemical consequences, which may impact on behavioural processes.

Several sources of evidence are, in fact, consistent with the supposition that activation of the inflammatory response system contributes to depression. For instance, depressive illness has been associated with increased levels of several cytokines and their soluble receptors (Maes, 1999), endotoxin challenge elicits signs of depression in both animals and human studies (Reichenberg et al., 2001; Yirmiya et al., 1999), and such effects in animals are attenuated by tricyclic antidepressant treatments (Shen et al., 1999; Yirmiya et al., 1999). Similarly, immunotherapy (IL-2 and IFN- α) has been shown to elicit depressive symptoms (Capuron et al., 2001) that are attenuated by antidepressant medication (Musselman et al., 2001). Clearly the behavioural tests are still somewhat limited, but coupled with the neurochemical findings, a prima-facie case exists supporting a cytokine link in the aetiology of some instances of depression. Of course, it ought to be underscored that while activation of the inflammatory response system may favour the development of depressive symptoms, this does not necessarily imply that instances of depression necessarily involve immune activation.

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References

Abramson LY, Seligman MEP, Teasdale JD (1978). Learned helplessness in humans: critique and reformulation. *Journal* of Abnormal Psychology 87, 49–74.

- Altar CA (1999). Neurotrophins and depression. *Trends Pharmacological Science* 20, 59–61.
- Anisman H, Merali Z (1999). Anhedonic and anxiogenic effects of cytokine exposure. *Advances in Experimental and Medical Biology* 461, 199–233.
- Anisman H, Ravindran AV, Griffiths J, Merali Z (1999a). Endocrine and cytokine correlates of major depression and dysthymia with typical or atypical features. *Molecular Psychiatry* 4, 182–188.
- Anisman H, Ravindran AV, Griffiths J, Merali Z (1999b). Interleukin-1 beta production in dysthymia before and after pharmacotherapy. *Biological Psychiatry* 46, 1649–1655.
- Anisman H, Zalcman S, Shanks N, Zacharko RM (1991).
 Multisystem regulation of performance deficits induced by stressors: an animal model of depression. In: Boulton A, Baker G, Martin-Iverson M. (Eds.), *Neuromethods: Animal Models of Psychiatry, II* (pp. 1–59). New Jersey: Humana Press.
- Anisman H, Hayley S, Merali Z (2001). Behavioral and central neurochemical consequences of cytokine challenge. In:
 Bienenstock J, Gorzynski R, Berczi I (Eds.), *Neuroimmune Biology: New Foundation of Biology* (pp. 141–162). New York: Elsevier Science.
- Anisman H, Zalcman S, Zacharko RM (1993). The impact of stressors on immune and central neurotransmitter activity: bidirectional communication. *Reviews in the Neurosciences* 4, 147–180.
- Ban EM, Haour F, Lenstra R (1992). Brain interleukin-1 gene expression induced by peripheral lipopolysaccharide administration. *Cytokine* 4, 48–54.
- Banks WA, Kastin AJ, Durham DA (1989). Bidirectional transport of interleukin-1 alpha across the blood brain barrier. *Brain Research Bulletin* 23, 437–443.
- Bartanusz V, Jezova D, Bertini LT, Tilders FJ, Aubry JM, Kiss JZ (1993). Stress-induced increase in vasopressin and corticotropin-releasing factor expression in hypophysiotrophic paraventricular neurons. *Endocrinology* 132, 895–902.
- Berk M, Wadee AA, Kuschke RH, O' Neill-Kerr A (1997). Acute phase proteins in major depression. *Journal of Psychosomatic Research* 43, 529–34.
- Bernardini R, Kamilaris TC, Calogero AE, Johnson EO, Gomez MT, Gold PW, Chrousos GP (1990). Interactions between tumor necrosis factor-*α*, hypothalamic corticotropinreleasing hormone, and adrenocorticotropin secretion in the rat. *Endocrinology* 126, 2876–2881.
- Blais V, Rivest S (2001). Inhibitory action of nitric oxide on circulating tumor necrosis factor-induced NF-kappaB activity and COX-2 transcription in the endothelium of the brain capillaries. *Journal of Neuropathological Experimental Neurology* 60, 893–905.
- Blalock JE (1984). The immune system as a sensory organ. Journal of Immunology 132, 1067–1070.
- Brebner K, Hayley S, Merali Z, Anisman H (2000). Synergistic effects of interleukin-1b, interleukin-6 and tumor necrosis factor-a: central monoamine, corticosterone and behavioral variations. *Neuropsychopharmacology* 22, 566–580.

Breder CD, Hazuka C, Ghayur T, Klug C, Huginin M, Yasuda K, Teng M, Saper, CD (1994). Regional induction of tumor necrosis factor a expression in the mouse brain. *Proceedings of the National Acedemy of Sciences USA 91*, 11393–11397.

Brown GW, Bifulco A, Harris TO (1987). Life events, vulnerability and onset of depression: some refinements. *British Journal of Psychiatry* 150, 30–42.

Brown GW, Harris TO (1978). Social Origins of Depression: A Study of Psychiatric Disorder in Women. New York: Free Press.

Brown GW, Harris TO (1989). *Life Events and Illness*. New York: Guilford Press.

Buttini M, Boddeke H (1995). Peripheral lipopolysaccharide stimulation induces interleukin-1β messenger RNA in rat brain microglial cells. *Neuroscience* 65, 523–530.

Buttini M, Sauter A, Boddeke HW (1994). Induction of interleukin-1 beta mRNA after focal cerebral ischaemia in the rat. *Molecular Brain Research* 23, 126–134.

Capuron L, Ravaud A, Gualde N, Bosmans E, Dantzer R, Maes M, Neveu PV (2001). Association between immune activation and early depressive symptoms in cancer patients treated with interleukin-2-based therapy. *Psychoneuroendocrinology* 26, 797–808.

Capuron L, Ravaud A, Radat F, Dantzer R, Goodall G (1998). Affects of interleukin-2 and alpha-interferon cytokine immunotherapy on the mood and cognitive performance of cancer patients. *Neuroimmunomodulation* 5, 9.

Caraceni A, Martini C, Belli F, Mascheroni L, Rivoltini L, Arienti F, Cascinelli N (1992). Neuropsychological and neurophysiological assessment of the central effects of interleukin-2 administration. *European Journal of Cancer 29A*, 1266–1269.

Carmelia G, Pietro G, De Simoni MG (1991). Activation of the hypothalamic serotonergic system by central interleukin-1. *European Journal of Pharmacology* 209, 139–140.

Chao CC, Hu S, Ehrlich L, Peterson PK (1995). Interleukin-1 and tumor necrosis factor-alpha synergistically mediate neurotoxicity: involvement of nitric oxide and of N-methyl-D-aspartate receptors. *Brain, Behavior and Immunity 9*, 355–365.

Chen B, Dowlatshahi D, MacQueen GM, Wang JF, Young LT (2001). Increased hippocampal BDNF immunoreactivity in subjects treated with antidepressant medication. *Biological Psychiatry 50*, 260–265.

Chen G, McCuskey RS, Reichlin S (2000). Blood interleukin-6 and tumor necrosis factor-alpha elevation after intracerebroventricular injection of *Escherichia coli* endotoxin in the rat is determined by two opposing factors: peripheral induction by LPS transferred from brain to blood and inhibition of peripheral response by a brainmediated mechanism. *Neuroimmunomodulation 8*, 59–69.

Colotta F, Dower SK, Sims JE, Mantovani A (1994). The type II 'decoy' receptor: a novel regulatory pathway for interleukin-1. *Immunology Today 15*, 562–566.

Connor TJ, Song C, Leonard BE, Merali Z, Anisman H (1998). An assessment of the effects of central interleukin-1, -2, -6, and tumor necrosis factor-*α* administration on some behavioral, neurochemical, endocrine and immune parameters in the rat. *Neuroscience* 84, 923–933.

Cunningham Jr. ET, DeSouza EB (1993). Interleukin-1 receptors in the brain and endocrine tissue. *Immunology Today* 14, 171–176.

Dantzer R, Bluthe RM, Aubert A, Goodall G, Bret-Dibat J-L, Kent S, Goujon E, Laye S, Parnet P, Kelley KW (1996). Cytokine actions on behavior. In: Rothwell NJ (Ed.), Cytokines and the Nervous System (pp. 117–144). London: Landes.

Dantzer R, Bluthe RM, Castanon N, Chauvet N, Capuron L, Goodall G, Kelley KW, Konsman J-P, Laye S, Parnet P, Pousset F (2001). Cytokine effects on behavior. In: Ader R, Felten DL, Cohen N (Eds.), *Psychoneuroimmunology*, vol. 2 (pp. 703–727). New York: Academic Press.

Davis M (1992). The role of the amygdala in fear and anxiety. Annual Review of Neuroscience 15, 353–375.

Day HE, Curran EJ, Watson Jr. SJ, Akil H (1999). Distinct neurochemical populations in the rat central nucleus of the amygdala and bed nucleus of the stria terminalis: evidence for their selective activation by interleukin-1beta. *Journal of Comparative Neurology* 413, 113–128.

Dayas CV, Buller KM, Day TA (1999). Neuroendocrine responses to an emotional stressor: evidence for involvement of the medial but not the central amygdala. *European Journal of Neuroscience 11*, 2312–2322.

de Goeij DC, Dijkstra H, Tilders FJH (1992a). Chronic psychosocial stress enhances vasopressin, but not corticotropin-releasing factor, in the external zone of the median eminence of male rats: relationship to subordinate status. *Endocrinology* 131, 847–853.

de Goeij DC, Jezova D, Tilders FJH (1992b). Repeated stress enhances vasopressin synthesis in corticotropin releasing factor neurons in the paraventricular nucleus. *Brain Research* 577, 165–168.

Del Rey A, Besedovsky HO (1992). Metabolic and neuroendocrine effects of pro-inflammatory cytokines. *European Journal of Clinical Investigation 1* (Suppl.), 10–15.

Denicoff KD, Rubinow DR, Papa MZ, Simpson L, Seipp LA, Lotze MT, Chang AE, Rosenstein D, Rosenberg SA (1987). The neuropsychiatric effects of treatment with interleukin-2 and lymphokine-activated killer cells. *Annals of Internal Medicine* 107, 293–300.

De Simoni MG, Del Bo R, De Luigi A, Simard S, Forloni G (1995). Central endotoxin induces different patterns of interleukin (IL)-1 beta and IL-6 messenger ribonucleic acid expression and IL-6 secretion in the brain and periphery. *Endocrinology* 136, 897–902.

Deutch AY, Roth RH (1990). The determinants of stressinduced activation of the prefrontal cortical dopamine system. In: Uylings HBM, Van Eden CG, De Bruin JPC, Corner MA, Feenstra MGP (Eds.), *Progress in Brain Research*, vol. 85 (pp. 367–403). New York: Elsevier.

Dunn AJ (1992). The role of interleukin-1 and tumor necrosis factor alpha in the neurochemical and neuroendocrine responses to endotoxin. *Brain Research Bulletin 6*, 807–812.

- Dunn AJ (1993). Infection as a stressor: a cytokine-mediated activation of the hypothalamo–pituitary–adrenal axis? *Ciba Foundation Symposium 172*, 226–239.
- Dunn AJ (2001). Effects of cytokines and infections on brain neurochemistry. In: Ader R, Felten DL, Cohen N (Eds.), *Psychoneuroimmunology*, vol. 2 (pp. 649–666). New York: Academic Press.
- Ek M, Kurosawa M, Lundeberg T, Ericsson A (1998). Activation of vagal afferents after intravenous injection of interleukin-1beta: role of endogenous prostaglandins. *Journal of Neuroscience* 18, 9471–9479.
- Emmert MH, Herman JP (1999). Differential forebrain c-fos mRNA induction by ether inhalation and novelty: evidence for distinctive stress pathways. *Brain Research* 845, 60–67.
- Ericsson A, Arias C, Sawchenko PE (1997). Evidence for an intramedullary prostaglandin-dependent mechanism in the activation of stress-related neuroendocrine circuitry by intravenous interleukin-1. *Journal of Neuroscience* 17, 7166–7179.
- Ericsson A, Kovacs KJ, Sawchenko PE (1994). A functional anatomical analysis of central pathways subserving the effects of interleukin-1 on stress-related neuroendocrine neurons. *Journal of Neuroscience* 14, 899–913.
- Esposito P, Gheorghe D, Kandere K, Pang X, Connolly R, Jacobson S (2001). Acute stress increases permeability of the blood–brain barrier through activation of mast cells. *Brain Research 888*, 117–127.
- Finlay JM, Jedema HP, Rabinovic AD, Mana MJ, Zigmond MJ, Sved AF (1997). Impact of corticotropin-releasing hormone on extracellular norepinephrine in prefrontal cortex after chronic cold stress. *Journal of Neurochemistry* 69, 144–150.
- Fiore M, Probert L, Kollias G, Akassoglou K, Alleva E, Aloe L (1996). Neurobehavioral alterations in developing transgenic mice expressing TNF-alpha in the brain. *Brain, Behavior and Immunity* 10, 126–138.
- Flores C, Samaha AN, Stewart J (2000). Requirement of endogenous basic fibroblast growth factor for sensitization to amphetamine. *Journal of Neuroscience 20*, RC55.
- Frommberger UH, Bauer J, Haselbauer P, Fraulin A, Riemann D, Berger M (1997). Interleukin-6-(IL-6) plasma levels in depression and schizophrenia: comparison between the acute state and after remission. *European Archives of Psychiatry and Clinical Neurosciences* 247, 228–233.
- Gaballec M-M, Griffais R, Fillion G, Haour F (1995). Expression of interleukin-1 α , interleukin-1 β and interleukin 1 receptor antagonist mRNA in mouse brain: regulation by bacterial lipopolysaccharide (LPS) treatment. *Molecular Brain Research* 31, 122–130.
- Gatti S, Bartfai T (1993). Induction of tumor necrosis factoralpha mRNA in the brain after peripheral endotoxin treatment: comparison with interleukin-1 family and interleukin-6. *Brain Research* 624, 291–294.
- Gaykema RP, Goehler LE, Tilders FJ, Bol JG, McGorry M, Fleshner M, Maier SF, Watkins LR (1998). Bacterial endotoxin induces fos immunoreactivity in primary

afferent neurons of the vagus nerve. *Neuroimmuno-modulation 5,* 234–240.

- Giulian D, Robertson C (1990). Inhibition of mononuclear phagocytes reduces ischemic injury in the spinal cord. *Annals of Neurology* 1, 33–42.
- Gresch PJ, Sved AF, Zigmond MJ, Finlay JM (1994). Stressinduced sensitization of dopamine and norepinephrine efflux in medial prefrontal cortex of the rat. *Journal of Neurochemistry* 63, 575–583.
- Griffiths J, Ravindran AV, Merali Z, Anisman H (2000). Dysthymia: neurochemical and behavioral perspectives. *Molecular Psychiatry* 5, 242–261.
- Guillin O, Diaz J, Carroll P, Griffon N, Schwartz JC, Sokoloff P (2001). BDNF controls dopamine D3 receptor expression and triggers behavioural sensitization. *Nature* 411, 86–89.
- Gutierrez EG, Banks WA, Kastin AJ (1993). Murine tumor necrosis factor alpha is transported from blood to brain in the mouse. *Journal of Neuroimmunology* 47, 169–176.
- Hayley S, Brebner K, Lacosta S, Merali Z, Anisman H (1999). Sensitization to the effects of tumor necrosis factor-*α*: neuroendocrine, central monoamine and behavioral variations. *Journal of Neuroscience* 19, 5654–5665.
- Hayley S, Lacosta S, Merali Z, van Rooijen N, Anisman H (2001b). Central monoamine and plasma corticosterone changes induced by a bacterial endotoxin: sensitization and cross-sensitization effects. *European Journal of Neuroscience* 13, 1155–1165.
- Hayley S, Staines W, Merali Z, Anisman H (2001a). Timedependent sensitization of corticotropin-releasing hormone, arginine vasopressin and c-fos immunoreactivity within the mouse brain in response to tumor necrosis factor-alpha. *Neuroscience* 106, 137–148.
- Hayley S, Wall P, Anisman H (2002). Sensitization to the neuroendocrine, central monoamine and behavioural effects of murine tumor necrosis factor-alpha: peripheral and central mechanisms. *European Journal of Neuroscience* 15, 1061–1076.
- Herbert TB, Cohen S (1993). Stress and immunity in humans: a meta-analytic review. *Psychosomatic Medicine* 55, 364–379.
- Herman JP, Cullinan WE (1997). Neurocircuitry of stress: central control of hypothalamo-pituitary-adrenocortical axis. *Trends in Neuroscience* 20, 78–84.
- Hopkins SJ, Rothwell NJ (1995). Cytokines and the nervous system. *Trends in Neuroscience* 18, 83–88.
- Igaz P, Novak I, Lazaar E, Horvath B, Heninger E, Falus A (2001). Bidirectional communication between histamine and cytokines. *Inflammatory Research* 50, 123–128.
- Ignatowski TA, Noble BK, Wright JR, Gorfien JL, Heffner RR, Spengler RN (1997). Neuronal-associated tumor necrosis factor (TNFa): its role in noradrenergic functioning and modification of its expression following antidepressant drug administration. *Journal of Neuroimmunology* 79, 84–90.
- Ilyin SE, Gayle D, Gonzalez-Gomez I, Miele ME, Plata-Salaman CR (1999). Brain tumor development in rats is associated with changes in central nervous system cytokine and neuropeptide systems. *Brain Research Bulletin* 48, 363–373.

Irwin J, Ahluwalia P, Anisman H (1986). Sensitization of norepinephrine activity following acute and chronic footshock. *Brain Research* 379, 98–103.

Irwin M (1999). Immune correlates of depression. *Advances in Experimental Medicine and Biology* 461, 1–24.

Jedema HP, Sved AF, Zigmond MJ, Finlay JM (1999). Sensitization of norepinephrine release in medial prefrontal cortex: effect of different chronic stress protocols. *Brain Research* 830, 211–217.

Jordan S, Kramer GL, Zukas PK, Petty F (1994). Previous stress increases in vivo biogenic amine response to swim stress. *Neurochemistry Research* 19, 1521–1525.

Kabiersch A, Del Rey A, Honegger CG, Besedovsky HO (1988). Interleukin-1 induces changes in norepinephrine metabolism in the rat brain. *Brain, Behavior and Immunity 2*, 267–274.

Kamikawa H, Hori T, Nakane H, Aou S, Tashiro N (1998). IL-1beta increases norepinephrine level in rat frontal cortex: involvement of prostanoids, NO, and glutamate. *American Journal of Physiology* 275, R803–810.

Kaur D, Cruess DF, Potter WZ (1998). Effect of IL-1alpha on the release of norepinephrine in rat hypothalamus. *Journal of Neuroimmunology* 90, 122–127.

Kawashima N, Kusnecov AW (2002). Effects of staphylococcal enterotoxin A on pituitary-adrenal activation and neophobic behavior in the C57BL/6 mouse. *Journal of Neuroimmunology* 123, 41–49.

Kelly O, Hayley S, Kokkinidis L, Anisman H (2001).
Histaminergic modulation of the central sensitizing effects of tumor necrosis factor-*α*. *Society for Neurosciences* 27.
Abst. no. 634.12.

Kendler KS, Thornton LM, Gardner CO (2000). Stressful life events and previous episodes in the etiology of major depression in women: an evaluation of the 'kindling' hypothesis. *American Journal of Psychiatry* 157, 1243–1251.

Kinouchi K, Brown G, Pasternak G, Donner DB (1991). Identification and characterization of receptors for tumor necrosis factor-α in the brain. *Biochemical and Biophysical Research Communications* 181, 1532–1538.

Kobayashi H, Fukata J, Murakami N, Usui T, Ebisui O, Muro S, Hanaoka I, Inoue K, Imura H, Nakao K (1997). Tumor necrosis factor receptors in the pituitary cells. *Brain Research* 758, 45–50.

Konsman JP, Tridon V, Dantzer R (2000). Diffusion and action of intracerebroventricularly injected interleukin-1. *Neuroscience* 101, 957–967.

Krueger JM, Fang J, Taishi P, Chen Z, Kusikata T, Gandi J (1998). Sleep: a physiologic role for IL-1 beta and TNF-alpha. *Annals of the New York Academy of Sciences 856*, 148–159.

Kubera M, Kenis G, Bosmans E, Zieba A, Dudek D, Nowak G, Maes M (2000). Plasma levels of interleukin-6, interleukin-10, and interleukin-1 receptor antagonist in depression: comparison between the acute state and after remission. *Polish Journal of Pharmacology* 52, 237–241.

Lacosta S, Merali Z, Anisman H (1998a). Effects of interleukin- 1β and mild stress on alterations of norepinephrine,

dopamine and serotonin neurotransmission: a regional microdialysis study. *Brain Research 761*, 225–235.

Lacosta S, Merali Z, Anisman H (1998b). Influence of interleukin-1 on exploratory behaviors, plasma ACTH and cortisol, and central biogenic amines in mice. *Psychopharmacology* 137, 351–361.

Lacosta S, Merali Z, Anisman H (1999). Influence of acute and repeated interleukin-2 administration on spatial learning, locomotor activity, exploratory behaviors and anxiety. *Behavioral Neuroscience* 113, 1030–1041.

Laflamme N, Rivest S (1999). Effects of systemic immunogenic insults and circulating proinflammatory cytokines on the transcription of the inhibitory factor kappaB alpha within specific cellular populations of the rat brain. *Journal of Neurochemistry* 73, 309–321.

Laye S, Parnet P, Goujon E, Dantzer R (1994). Peripheral administration of lipopolysaccharide induces the expression of cytokine transcripts in the brain and pituitary of mice. *Molecular Brain Research* 27, 157–162.

- Lazarus RS (1993). Coping theory and research: past, present and future. *Psychosomatic Medicine* 55, 234–247.
- Lewinsohn PM, Allen NB, Seeley JR, Gotlib IH (1999). First onset versus recurrence of depression: differential processes of psychosocial risk. *Journal of Abnormal Psychology* 108, 483–489.
- LeDoux JE (2000). Emotion circuits in the brain. Annual Review of Neuroscience 23, 155–184.

Lee HY, Whiteside MB, Herkenham M (1998). Area postrema removal abolishes stimulatory effects of intravenous interleukin-1beta on hypothalamic–pituitary–adrenal axis activity and c-fos mRNA in the hypothalamic

paraventricular nucleus. *Brain Research Bulletin 46*, 495–503. Lee S, Rivier C (1994). Hypophysiotropic role and

hypothalamic gene expression of corticotropin-releasing factor and vasopressin in rats injected with interleukin-1 beta systemically or into the brain ventricles. *Journal of Neuroendocrinology 6*, 217–224.

Leonard BE (2001). The immune system, depression and the action of antidepressants. *Progress in Neuropsychopharmacology and Biological Psychiatry* 25, 767–780.

Li HY, Ericsson A, Sawchenko PE (1996). Distinct mechanisms underlie activation of hypothalamic neurosecretory neurons and their medullary catecholaminergic afferents in categorically different stress paradigms. *Proceedings of the National Academy of Sciences USA 93*, 2359–2364.

Licinio J, Wong ML (1997). Interleukin 1 receptor antagonist gene expression in rat pituitary in the systemic inflammatory response syndrome: pathophysiological implications. *Molecular Psychiatry* 2, 99–103.

Linthorst ACE, Flachskamm C, Muller-Preuss P, Holsboer F, Reul JMHM (1995). Effect of bacterial endotoxin and interleukin-1 β on hippocampal serotonergic neurotransmission, behavioral activity, and free corticosterone levels: an in vivo microdialysis study. *Journal* of Neuroscience 15, 2920–2934.

Liu L, Kita T, Tanaka N, Kinoshita Y (1996). The expression of tumor necrosis factor in the hypothalamus after treatment with lipopolysaccharide. *International Journal of Experimental Pathology* 77, 37–44.

- Madrigal JL, Hurtado O, Moro MA, Lizasoain I, Lorenzo P, Castrillo A, Bosca L, Leza JC (2002). The increase in TNFalpha levels is implicated in NF-kappaB activation and inducible nitric oxide synthase expression in brain cortex after immobilization stress. *Neuropsychopharmacology 26*, 155–163.
- Maes M (1995). Evidence for an immune response in major depression: a review and hypothesis. *Progress in Neuro-Psychopharmacology and Biological Psychiatry* 19, 11–38.
- Maes M (1999). Major depression and activation of the inflammatory response system. *Advances in Experimental and Medical Biology* 461, 25–46.
- Maes M, Capuron L, Ravaud A, Gualde N, Bosmans E, Egyed B, Dantzer R, Neveu, PJ (2001). Lowered serum dipeptidyl peptidase IV activity is associated with depressive symptoms and cytokine production in cancer patients receiving interleukin–2–based immunotherapy. *Neuropsychopharmacology* 24, 130–140.
- Maes M, Meltzer HY (1995). The serotonin hypothesis of major depression. In: Bloom FE, Kupfer DJ (Eds.), *Psychopharmacology: The Fourth Generation of Progress* (pp. 933–944.) New York: Raven Press.
- Maes M, Meltzer HY, Stevens W, Cosyns P, Blockx P (1994). Multiple reciprocal relationships between *in vivo* cellular immunity and hypothalamic–pituitary–adrenal axis in depression. *Psychological Medicine* 24, 167–177.
- Maier SF, Watkins LR (1998). Cytokines for psychologists: implications of bidirectional immune-to-brain communication for understanding behavior, mood, and cognition. *Psychological Review* 105, 83–107.
- Makino S, Asaba K, Nishiyama M, Hashimoto K (1999). Decreased type 2 corticotropin-releasing hormone receptor mRNA expression in the ventromedial hypothalamus during repeated immobilization stress. *Neuroendocrinology* 70, 160–167.
- Mandrup-Poulsen T, Nerup J, Reimers JI, Pociot F, Andersen HU, Karlsen A, Bjerre U, Bergholdt R (1995). Cytokines and the endocrine system. I. The immunoendocrine network. *European Journal of Endocrinology* 133, 660–671.
- Manji HK, Drevets WC, Charney DS (2001). The cellular neurobiology of depression. *Nature Medicine* 7, 541–547.
- Mark KS, Trickler WJ, Miller DW (2001). Tumor necrosis factor-alpha induces cyclooxygenase-2 expression and prostaglandin release in brain microvessel endothelial cells. Journal of Pharmacological and Experimental Therapeutics 297, 1051–1058.
- Masana MI, Heyes MP, Mefford IN (1990). Indomethacin prevents increased catecholamine turnover in rat brain following systemic endotoxin challenge. *Progress in Neuro-Psychopharmacology and Biological Psychiatry* 14, 609–621.
- McCoy JG, Matta SG, Sharp BM (1994). Prostaglandins mediate the ACTH response to interleukin-1-beta instilled into the hypothalamic median eminence. *Neuroendocrinology* 60, 426–435.

- McEwen BS (1998). Stress, adaptation, and disease. Allostasis and allostatic load. *Annals of the New York Academy Sciences* 840, 33–44.
- McEwen B, Brinton R, Chao H, Coirini H, Gannon M, Gould E (1990). The hippocampus: a site for modulatory interaction between steroid hormones, neurotransmitters and neuropeptides. In: Muller E, Macloed R (Eds.), *Neuroendocrine Perspectives*, vol. 8 (pp. 93–131). New York: Springer.
- McEwen BS, Seeman T (1999). Protective and damaging effects of mediators of stress. Elaborating and testing the concepts of allostasis and allostatic load. *Annals of the New York Academy Sciences 896*, 30–47.
- Mekaouche M, Givalois L, Barbanel G, Siaud P, Maurel D, Malaval F, Bristow AF, Boissin J, Assenmacher I, Ixart G (1994). Chronic restraint enhances interleukin-1-beta release in the basal state and after an endotoxin challenge, independently of adrenocorticotropin and corticosterone release. *Neuroimmunomodulation* 1, 292–299.
- Merali Z, Lacosta S, Anisman H (1997). Effects of interleukin-1 β and mild stress on alterations of norepinephrine, dopamine and serotonin neurotransmission: a regional microdialysis study. *Brain Research* 761, 225–235.
- Merrill JE, Benveniste EN (1996). Cytokines in inflammatory brain lesions: helpful and harmful. *Trends in Neuroscience 8*, 331–338.
- Meyers CA, Valentine AD (1995). Neurological and psychiatric adverse effects of immunological therapy, *CNS Drugs* 3, 56–68.
- Minami M, Kuraishi Y, Yamaguchi T, Nakai S, Hirai Y, Satoh M (1990). Convulsants induce interleukin-1β messenger RNA in rat brain. *Biochemical and Biophysical Research Communications* 171, 832–837.
- Minami M, Kuraishi Y, Yamaguchi T, Nakai S, Hirai Y, Satoh M (1991). Immobilization stress induces interleukin- 1β mRNA in rat hypothalamus. *Neuroscience Letters* 123, 254–256.
- Mohankumar PS, Quadri SK (1993). Systemic administration of interleukin-1 stimulates norepinephrine release in the paraventricular nucleus. *Life Science* 52, 1961–1967.
- Mohankumar PS, Thyagarajan S, Quadri SK (1993). Interleukin-1 β increases 5-hydroxyindoleacetic acid release in the hypothalamus in vivo. *Brain Research Bulletin* 31, 745–748.
- Monroe SM, Depue RA (1991). Life stress and depression. In: Becker J, Kleinman A (Eds.), *Psychosocial Aspects of Depression* (pp. 101–130). Hillsdale, NJ: Erlbaum.
- Mullar N, Ackenheil M (1998). Psychoneuroimmunology and the cytokine action in the CNS: implications for psychiatric disorders. *Progress in Neuro-Psychopharmacoloy and Biological Psychiatry* 22, 1–33.
- Munck A, Guyre PM, Holbrook NJ (1984). Physiological functions of glucocorticoids in stress and their relation to pharmacological actions. *Endocrine Review* 5, 25–44.
- Musselman DL, Lawson DH, Gumnick JF, Manatunga A, Penna S, Goodkin R, Greiner K, Nemeroff CB, Miller AH (2001). Paroxetine for the prevention of the depression

and neurotoxicity induced by high dose interferon alpha. *New England Journal of Medicine 344, 961–966.*

Nassberger L, Traskman-Bendz L (1993). Increased soluble interleukin-2 receptor concentrations in suicide attempters. *Acta Psychiatrica Scandinavica 88*, 48–52.

Nguyen KT, Deak T, Owens SM, Kohno T, Fleshner M, Watkins LR, Maier SF (1998). Exposure to acute stress induces brain interleukin-1 protein in the rat. *Journal of Neuroscience 18*, 2239–2246.

Nguyen KT, Deak T, Will MJ, Hansen MK, Hunsaker BN, Fleshner M, Watkins LR, Maier SF (2000). Time course and corticosterone sensitivity of the brain, pituitary, and serum interleukin-1beta protein response to acute stress. *Brain Research* 859, 193–201.

Nieto E, Vieta E, Alvarez L, Torra M, Colom F, Gastó C (2000). Alpha-1-acid glycoprotein in major depressive disorder. Relationships to severity, response to treatment and imipramine plasma levels. *Journal of Affective Disorders* 59, 159–164.

Niimi M, Sato M, Wada Y, Takahara J, Kawanishi K (1996). Effect of central and continuous intravenous injection of interleukin-1 beta on brain c-fos expression in the rat: involvement of prostaglandins. *Neuroimmunomodulation 3*, 87–92.

Nisenbaum LK, Abercrombie ED (1992). Enhanced tyrosine hydroxylation in hippocampus of chronically stressed rats upon exposure to a novel stressor. *Journal of Neurochemistry* 58, 276–281.

Nistico G, De Sarro G (1991). Is interleukin 2 a neuromodulator in the brain? *Trends in Neuroscience* 14, 146–150.

O'Connor JJ, Coogan AN (1999). Actions of the proinflammatory cytokine IL-1β on central synaptic transmission. *Experimental Physiology* 84, 601–614.

Parsadaniantz SM, Batsche E, Gegout-Pottie P, Terlain B, Gillet P, Netter P, Kerdelhue B (1997). Effects of continuous infusion of interleukin 1 beta on corticotropin-releasing hormone (CRH), CRH receptors, proopiomelanocortin gene expression and secretion of corticotropin, betaendorphin and corticosterone. *Neuroendocrinology 65*, 53–63.

Parsadaniantz SM, Lebeau A, Duval P, Grimaldi B, Terlain B, Kerdelhue B (2000). Effects of the inhibition of cyclooxygenase 1 or 2 or 5-lipoxygenase on the activation of the hypothalamic–pituitary–adrenal axis induced by interleukin-1beta in the male rat. *Journal of Neuroendocrinology* 12, 766–773.

Paykel ES (2001). Stress and affective disorders in humans. Seminars in Clinical Neuropsychiatry 6, 4–11.

Perlstein RS, Whitnall MH, Abrams JS, Moughey EH, Neta R (1993). Synergistic roles of interleukin-6, interleukin-1, and tumor necrosis factor in the adrenocorticotropin response to bacterial lipopolysaccharide in vivo. *Endocrinology* 132, 946–952.

Petitto JM, McCarthy DB. Rinker CM, Huang Z, Getty T (1997). Modulation of behavioral and neurochemical measures of forebrain dopamine function in mice by species-specific interleukin-2. *Journal of Neuroimmunology* 73, 183–190. Plata-Salaman CR (1998). Cytokine-induced anorexia: Behavioral, cellular, and molecular mechanisms. Annals of the New York Academy Sciences 856, 160–170.

Plata-Salaman CR, Ilyin SE, Gayle D, Flynn MC (1998). Gram-negative and Gram-positive bacterial products induce differential cytokine profiles in the brain: analysis using an integrative molecular-behavioral in vivo model. *International Journal of Molecular Medicine* 1, 387–397.

Plotsky PM, Owens MJ, Nemeroff CB (1995). Neuropeptide alterations in mood disorders. In: Bloom FE, Kupfer DJ (Eds.), *Psychopharmacology: The Fourth Generation of Progress* (pp. 971–981). New York: Raven Press.

Post RM (1992). Transduction of psychosocial stress into the neurobiology of recurrent affective disorder. *American Journal of Psychiatry* 149, 999–1010.

Puglisi-Allegra S, Imperato A, Angelucci L, Cabib S (1991). Acute stress induces time-dependent responses in dopamine mesolimbic system. *Brain Research* 554, 217–222.

Quagliarello VJ, Wisplwey B, Long Jr. WJ, Sheld WM (1991). Recombinant interleukin-1 induces meningitis and bloodbrain barrier injury in the rat. *Journal of Clinical Investigations* 87, 1360–1366.

Quan N, Sundar SK, Weiss JM (1994). Induction of interleukin-1 in various brain regions after peripheral and central injections of lipopolysaccharide. *Journal of Neuroimmunology* 49, 125–134.

Rajora N, Boccoli G, Burns D, Sharma S, Catania A, Lipton JM (1997). a-MSH modulates local and circulating tumor necrosis factor-*a* in experimental brain inflammation. *Journal of Neuroscience* 15, 2181–2186.

Ramamoorthy S, Ramamoorthy JD, Prasad PD, Bhat GK, Mahesh VB, Leibach FH, Ganapathy V (1995). Regulation of the human serotonin transporter by interleukin-1 beta. *Biochemical Biophysical Research Communications* 216, 560–567.

Reichenberg A, Yirmiya R, Schuld A, Kraus T, Haack M, Morag A, Pollmacher T (2001). Cytokine-associated emotional and cognitive disturbances in humans. *Archives* of *General Psychiatry* 58, 445–452.

Rivest S, Lacroix S, Vallières L, Nadeau S, Zhang J, Laflamme N (2000). How the blood talks to the brain parenchyma and the paraventricular nucleus of the hypothalamus during systemic inflammatory and infectious stimuli. *Proceedings of the Society of Experimental* and Biological Medicine 223, 22–38.

Rivest S, Lee S, Attardi B, Rivier C (1993). The chronic intracerebroventricular infusion of interleukin-1 beta alters the activity of the hypothalamic–pituitary– gonadal axis of cycling rats. I. Effect on LHRH and gonadotropin biosynthesis and secretion. *Endocrinology* 133, 2424–2430.

Roth J, Hubschle T, Pehl U, Ross G, Gerstberger R (2002). Influence of systemic treatment with cyclooxygenase inhibitors on lipopolysaccharide-induced fever and circulating levels of cytokines and cortisol in guinea-pigs. *Pflügers Archiv* 443, 411–417.

Rothermundt M, Arolt V, Peters M, Gutbrodt H, Fenker J, Kersting A, Kirchner H (2001). Inflammatory markers in major depression and melancholia. *Journal of Affective Disorders* 63, 93–102.

Rothwell NJ, Hopkins SJ (1995). Cytokines and the nervous system II: actions and mechanisms of action. *Trends in Pharamcological Science 18*, 130–136.

Roy A (1985). Early parental separation and adult depression. *Archives of General Psychiatry* 42, 987–991.

Saperstein A, Brand H, Audhya T, Nabriski D, Hutchinson B, Rosenweig S, Hollander CS (1992). Interleukin-1β mediates stress-induced immunosuppression via corticotropinreleasing factor. *Endocrinology* 130, 152–159.

Sato S, Reiner SL, Jensen MA, Roos RP (1997). Central nervous system mRNA expression following Theilers murine encephalomyelitis virus infection. *Journal of Neuroimmunology* 76, 213–223.

Sawchenko PE, Brown ER, Chan RK, Ericsson A, Li HY, Roland BL, Kovcs KJ (1996). The paraventricular nucleus of the hypothalamus and the functional neuroanatomy of visceromotor responses to stress. *Progress in Brain Research* 107, 201–222.

Schatzberg AF, Schildkraut JJ (1995) Recent studies on norepinephrine systems on mood disorders. In: Bloom FE, Kupfer DJ (Eds.), *Psychopharmacology: The Fourth Generation* of Progress (pp. 911–920). New York: Raven.

Schmidt ED, Janszen AWJW, Wouterlood FG, Tilders FJH (1995). Interleukin-1 induced long-lasting changes in hypothalamic corticotropin-releasing hormone (CRH) neurons and hyperresponsiveness of the hypothalamic– pituitary–adrenal axis. *Journal of Neuroscience* 15, 7417–7426.

Schobitz B, De Kloet ER, Holsboer F (1994). Gene expression and function of interleukin 1, interleukin 6, and tumor necrosis factor in the brain. *Progress in Neurobiology* 44, 397–432.

Shen Y, Connor TJ, Nolan Y, Kelly JP, Leonard BE (1999). Differential effect of chronic antidepressant treatments on lipopolysaccharide-induced depressive-like behavioural symptoms in the rat. *Life Science* 65, 1773–1786.

Shintani F, Kanba S, Nakaki T, Nibuya M, Kinoshita N, Suzuki E, Yagi G, Kato R, Asai M (1993). Interleukin-1 β augments release of norepinephrine, dopamine and serotonin in the rat anterior hypothalamus. *Journal of Neuroscience* 13, 3574–3581.

Shintani F, Nakaki T, Kanba S, Sato K, Yagi G, Shiozawa M, Aiso S, Kato R, Asai M (1995). Involvement of interleukin-1 in immibilization stress-induced increase in plasma adrenocorticotropic hormone and in release of hypothalamic monoamines in the rat. *Journal of Neuroscience* 15, 1961–1970.

Shirayama Y, Chen AC, Nakagawa S, Russell DS, Duman RS (2002). Brain-derived neurotrophic factor produces antidepressant effects in behavioral models of depression. *Journal of Neuroscience* 22, 3251–3261.

Shohami E, Ginis I, Hallanbeck JM (1999). Dual role of tumor necrosis factor alpha in brain injury. *Cytokine Growth Factor Review 10*, 119–130.

Sluzewska A (1999). Indicators of immune activation in depressed patients. *Advances in Experimental Medicine and Biology* 461, 59–74. Sluzewska A, Rybakowski JK, Laciak M, Mackiewicz A, Sobieska M, Wiktorowicz K (1995). Interleukin-6 serum levels in depressed patients before and after treatment with fluoxetine. *Annals of the New York Academy of Sciences 762*, 474–476.

Smagin GN, Swiergiel AH, Dunn AJ (1996). Peripheral administration of interleukin-1 increases extracellular concentrations of norepinephrine in rat hypothalamus: comparison with plasma corticosterone. *Psychoneuroendocrinology* 21, 83–93.

Smith RS (1991). The macrophage theory of depression. *Medical Hypotheses 35, 298–306.*

Solomon DA, Keller MB, Leon AC, Mueller TI, Lavori PW, Shea MT, Coryell W, Warshaw M, Turvey C, Maser JD, Endicott J (2000). Multiple recurrences of major depressive disorder. *American Journal of Psychiatry* 157, 229–233.

Song C, Dinan T, Leonard BE (1994). Changes in immunoglobulin, complement and acute phase protein levels in depressed patients and normal controls. *Journal* of Affective Disorders 30, 283–288.

Song C, Merali Z, Anisman H (1999). Variations of nucleus accumbens dopamine and serotonin following systemic interleukin-1, interleukin-2 or interleukin-6 treatment. *Neuroscience 88*, 823–836.

Sweep CG, Van der Meer MJM, Hermus AR, Smals AG, Van der Meer JWM, Pesman GJ, Willemsen SJ, Benraad TJ, Kloppenborg PW (1992). Chronic stimulation of the pituitary–adrenal axis in rats by interleukin-1 β infusion: in vivo and in vitro studies. *Endocrinology* 130, 1153–1164.

Szczepanik AM, Funes S, Petko W, Ringheim GE (2001). IL-4, IL-10 and IL-13 modulate A-beta(1-42)-induced cytokine and chemokine production in primary murine microglia and a human monocyte cell line. *Journal of Neuroimmunology* 113, 49–62.

Tancredi V, DArcangelo G, Grassi F, Tarroni P, Palmier G, Santoni A, Eusebi F (1992). Tumor necrosis factor alters synaptic transmission in rat hippocampal slices. *Neuroscience Letters* 146, 176–178.

Taupin V, Toulmond S, Serrano A, Benavides J, Zavala F (1993). Increase in IL-6, IL-1 β and TNF α levels in rat brain following traumatic lesion. *Journal of Neuroimmunology* 42, 177–186.

Tilders FJH, Schmidt ED (1998). Interleukin-1-induced plasticity of hypothalamic CRH neurons and long-term stress hyperresponsiveness. *Annals of the New York Academy Sciences 840*, 65–73.

Tilders FJH, Schmidt ED, De Goeij DCE (1993). Phenotypic plasticity of CRF neurons during stress. *Annals of the New York Academy Sciences* 697, 39–52.

Turnbull AV, Pitossi FJ, Lebrun J-J, Lee S, Meltzer JC, Nance DM, del Rey A. Besedovsky H, Rivier C (1997). Inhibition of tumor necrosis factor-*α* within the CNS markedly reduces the plasma adrenocorticotropin response to peripheral local inflammation in rats. *Journal of Neuroscience* 17, 3262–3273.

Turnbull AV, Rivier CL (1999). Regulation of the hypothalamic–pituitary–adrenal axis by cytokines: Actions and mechanisms of action. *Physiology Reviews* 79, 1–71. Vaidya VA, Duman RS (2001). Depression – emerging insights from neurobiology. Brain Medical Bulletin 57, 61–79.

- van der Meer MJ, Sweep CG, Pesman GJ, Tilders FJ, Hermus AR (1996). Chronic stimulation of the hypothalamus–pituitary–adrenal axis in rats by interleukin 1beta: central and peripheral mechanisms. *Cytokine 8*, 910–919.
- Vellucci SV, Parrott RF, da Costa AC, Ohkura S, Kendrick KM (1995). Increased body temperature, cortisol secretion, and hypothalamic expression of c-fos, corticotrophin releasing hormone and interleukin-1 beta mRNAs, following central administration of interleukin-1 beta in the sheep. *Brain Research and Molecular Brain Research* 29, 64–70.
- Watanobe H, Takebe K (1993). Intrahypothalamic infusion with interleukin-1 beta stimulates the release of corticotropin-releasing hormone and arginine vasopressin and the plasma adrenocorticotropin in freely moving rats: a comparative perfusion of the paraventricular nucleus and the median eminence. *Neuroendocrinology* 57, 593–599.
- Watkins LR, Maier SF, Goehler LE (1995). Cytokine-to-brain communication: a review, analysis of alternative mechanisms. *Life Science* 11, 1011–1026.
- Weiss JM, Simson PE (1989). Electrophysiology of the locus coeruleus: implications for stress-induced depression. In: Koob GF, Ehlers CL, Kupfer DJ (Eds.), Animal Models of Depression. Boston: Birkhauser.
- Xu Y, Day TA, Buller KM (1999). The central amygdala modulates hypothalamic–pituitary–adrenal axis responses

to systemic interleukin-1beta administration. *Neuroscience* 94, 175–183.

- Yabuuchi K, Maruta E, Minami M, Satoh M (1996). Induction of interleukin-1 β mRNA in the hypothalamus following subcutaneous injections of formalin into the rat hind paws. *Neuroscience Letters* 207, 109–112.
- Yabuuchi K, Minami M, Katsumata S, Satoh M (1993). In situ hybridization study of interleukin-1β mRNA induced by kainic acid in the rat brain. *Molecular Brain Research 20*, 153–161.
- Yirmiya R, Weidenfeld J, Pollak Y, Morag M, Morag A, Avitsur R, Barak O, Reichenberg A, Cohen E, Shavit Y, Ovadia H (1999). Cytokines, 'depression due to a general medical condition,' and antidepressant drugs. *Advances in Experimental and Medical Biology* 461, 283–316.
- Zalcman S, Murray L, Dyck DG, Greenberg AH, Nance DM (1998). Interleukin-2 and –6 induce behavioral-activating effects in mice. *Brain Research 811*, 111–121.
- Zalcman S, Green-Johnson JM, Murray L, Nance DM, Dyck D, Anisman H, Greenberg A (1994). Cytokine-specific central monoamine alterations induced by interleukin (IL)-1, IL-2 and IL-6. *Brain Research* 643, 40–49.
- Zalcman SS (2001). Interleukin-2 potentiates novelty- and GBR 12909-induced exploratory activity. *Brain Research 899*, 1–9.
- Zhou DH, Shanks N, Riechman SE, Liang RM, Kusnecov AW, Rabin BS (1996). Interleukin-6 modulates interleukin-1 and stress-induced activation of the hypothalamic– pituitary–adrenal axis in male rats. *Neuroendocrinology* 63, 227–236.