Thrombosis and Sickle Cell Disease

Lucia De Franceschi, M.D.,¹ Maria Domenica Cappellini, M.D.,² and Oliviero Olivieri, M.D.¹

ABSTRACT

Sickle cell disease (SCD) is characterized by the presence of sickle hemoglobin, which has the unique property of polymerizing when deoxygenated. The pathophysiology of acute and chronic clinical manifestations of SCD have shown the central role of dense, dehydrated red cells in acute and chronic clinical manifestations of this pathology. Recent studies have indicated that SCD is characterized by a hypercoagulable state that contributes to the vaso-occlusive events in microcirculation, leading to acute and chronic sickle cell-related organ damage. This review discusses, in the context of SCD, (1) abnormalities in the coagulation system, (2) perturbation of platelet activation and aggregation, (3) vascular endothelial dysfunction, (4) the contribution of cell inflammatory responses, and (5) the connection with nitric oxide metabolism. We also review the available studies on the therapeutic approaches in clinical management of hypercoagulability in SCD.

KEYWORDS: Hypercoagulability, vascular endothelial dysfunction, inflammation, neutrophils, dense red cells

Sickle cell disease (SCD; Online Mendelian Inheritance in Man [OMIM] No. 603903) is an autosomal recessive genetic red cell disorder with a worldwide distribution that results from a point mutation $(\beta^{S}, 6V)$ in codon 6, with the insertion of value in place of glutamic acid, leading to the production of a defective form of hemoglobin (hemoglobin S [HbS]). In the United States ~75,000 people have SCD. In Europe, immigration from developing countries has increased the prevalence of SCD through the second half of the 20th century, and now almost 20,000 to 25,000 SCD patients have been registered.¹⁻⁴ Sickle hemoglobin (HbS) shows peculiar biochemical properties, polymerizing when deoxygenated. Studies of the kinetics of HbS polymerization following deoxygenation have shown that the kinetics of polymer formation is a high-order exponential function of hemoglobin concentration, thus demonstrating the crucial role of cellular HbS concentration in the phenomenon of sickling. 5,6

HbS polymerization is associated with a reduction in cell ion and water content (cell dehydration), increased red cell density, and further acceleration of HbS polymerization.⁵⁻⁷ Pathophysiological studies have shown that the dense, dehydrated red cells play a central role in acute and chronic clinical manifestations of SCD, in which intravascular sickling in capillaries and small vessels leads to vaso-occlusive and impaired blood flow.^{6,8} The persistent membrane damage associated with HbS polymerization also favors the generation of distorted rigid cells and further contributes to vaso-occlusive events and cell destruction in the peripheral circulation. These damaged dense sickle red cells also show a loss of phospholipid asymmetry with externalization of phosphatidylserine (PS; Fig. 1), which is believed to play a significant role in promoting macrophage recognition with removal of

¹Department of Medicine, University of Verona, Verona, Italy; ²Policlinico Foundation, Istituto di Ricovero e Cura a Carattere Scientifico (IRCCS), University of Milan, Milan, Italy.

Address for correspondence and reprint requests: Lucia De Franceschi, M.D., Department of Medicine, University of Verona, Policlinico GB Res8i, P.le L Scuro, 10, 37134 Verona, Italy (e-mail: lucia.defranceschi@ univr.it).

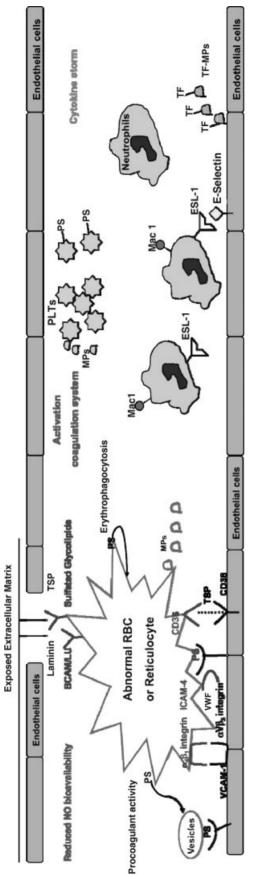
Coagulopathies and Thrombosis: Usual and Unusual Causes and

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erythrocytes (erythrophagocytosis) and activation of coagulation. Setty et al have recently shown that increased PS-exposing red cells are associated with thrombin generation as well as increased tissue factor (TF) expression, more likely due to the increased circulating hemoglobin than as a direct connection between PS-exposing red cells and vascular endothelium.^{9–11}

A complex perturbation of hemostasis has been reported in SCD both under steady state and during acute events. The vaso-occlusive events in the microcirculation result from a complex and still partially known scenario involving the interactions among different cell types, including dense, dehydrated sickle cells, reticulocytes, abnormally activated endothelial cells, leukocytes, platelets, and plasma factors such as coagulation system, cytokines and oxidized proinflammatory lipids.^{12–20} Clinical manifestations of the prothrombotic state of sickle cell patients include venous thromboembolism, in situ thrombosis, and stroke, associated with an higher risk of thrombotic complications in patients splenectomized or with functional hyposplenism.^{6,21–27}

COAGULATION SYSTEM AND SICKLE CELL DISEASE

Studies in SCD have shown increased prothrombin fragment 1.2 (F1.2), thrombin-antithrombin complexes,

plasma fibrinogen products, D-dimer, and decreased factor V, suggesting an enhanced thrombin generation and supporting a chronic thrombophilic state in SCD patients that is further amplified during acute events (Table 1).^{13,17,28–38} Sickle cell patients also show abnormal (decreased) levels of factor (F) VII and activated FVII compared with normal subjects, most likely due to increased TF activity that promotes accelerated FVII turnover.^{37,39} Moreover, in sickle cell patients under steady state conditions, decreased FXII and FIX have been observed, possibly related to activation of the intrinsic coagulation pathway⁴⁰ (Table 1).

Ataga et al recently reported high levels of thrombin-antithrombin complex, prothrombin fragment F1 + 2, and D-dimer, associated with an activation profile of vascular endothelium (i.e., soluble vascular endothelial cell adhesion molecule) in sickle cell subjects with pulmonary hypertension compared with normal controls.⁴¹ It is interesting to note that these authors also observed a correlation between the rate of hemolysis and the hypercoagulability state in SCD patients with pulmonary hypertension.⁴¹ However, in another cohort of SCD patients with mild pulmonary hypertension, van Beers et al reported no association of the hypercoagulative state of SCD with the early phase of pulmonary hypertension,⁴² suggesting that more complex events are involved in the pathogenesis of pulmonary hypertension in SCD.

 Table 1
 Platelets and Coagulation System in Sickle Cell Disease

Platelet Parameters and	Alterations	References	
Coagulation System			
Platelet activation (CD62, CD63, GPIIb/IIIa)	Increased	Tomer et al, ^{29,36} Foulon et al, ⁶⁴ Browne et al, ⁶⁵	
		Wun et al, ⁶⁹ Famodu and Oduwa, ⁷⁰ Lee et al ⁷¹	
Platelet aggregation	Increased	Kenny et al, ⁶⁶ Westwick et al, ⁶⁷ Winichagoon et al ⁶⁸	
Phosphatidyl serine-rich platelets	Increased	d Tomer et al ^{29,36}	
Thrombin-antithrombin complex	Increased	Peters et al, ²⁸ Rickles and O'Leary, ³¹ Stuart and Setty, ³² Green and Scott, ³³ Richardson et al, ³⁵ Tomer et al, ³⁶ Kurantsin-Mills et al, ³⁷ Ataga et al, ⁴¹ van Beers et al ⁴²	
Prothrombin fragment $F1 + 2$	Increased	Peters et al, ²⁸ Tomer et al, ³⁶ Ataga et al, ⁴¹ van Beers et al ⁴²	
Plasmin-antiplasmin complex	Increased	Tomer et al ^{29,36}	
FV	Decreased	Leslie et al ³⁰	
FVII and FVIIa	Accelerated turnover	Kurantsin-Mills et al, ³⁷ Hagger et al ³⁹	
FXII and FIX	Decreased	Branch and Rodgers ⁴⁰	
Fibrinogen and fibrin-fibrinogen complex	Increased	Adam et al, ¹⁷ Leslie et al ³⁰	
Fibrinopeptide A	Increased	Green and Scott, ³³ Kurantsin-Mills et al, ³⁷ Westerman et al ³⁸	
D-dimer	Increased	Adam et al, ¹⁷ Ataga et al, ⁴¹ van Beers et al, ⁴² Francis and Haywood ¹²⁰	
Protein C and S	Decreased	Green and Scott, ³³ Westerman et al, ³⁸ Tam, ⁴⁵ el-Hazmii et al ⁴⁶ Karayalcin and Lanzkowsky, ⁴⁶ Kuypers et al, ⁴⁸ Lane et al, ⁴⁹ Francis and Haywood ¹²⁰	
Plasminogen activator inhibitor	Increased	Tomer et al, ³⁶ Westerman et al, ³⁸ Nsiri et al ^{50,51}	

GP, glycoprotein.

A possible role in activating the clotting system with thrombin generation in sickle cell patients is also played by the circulating TF-positive microparticles (MP) derived from red cells, platelets, endothelial cells, and monocytes (Fig. 1). Shet et al reported increased TF-positive MPs in sickle cell patients in both steady state and during acute events compared with normal controls, thereby suggesting a possible contribution of TF-positive MPs to the sickle cell prothrombotic state.⁴³ In a 2009 study, van Beers et al observed higher levels of circulating MPs from erythrocytes and platelets in SCD patients under steady state compared with normal controls, further increasing during acute vaso-occlusive events.⁴⁴ These authors did not find TF-MPs as previously reported by Shet et al,43 but they observed that the levels of circulating MPs strongly correlate with hemolysis, von Willebrand factor (VWF), D-dimer, and F1+2 levels, supporting a role of MPs in the prothrombotic state of sickle cell patients.⁴⁴

Studies on natural anticoagulant in patients with SCD showed low levels of protein C and S, suggesting a possible perturbation in either their synthesis related to liver disease or a consumption by increased TF and thrombin production. The relative deficiency of protein C and S was reported to have a clinical impact on the risk of developing stroke in children with SCD.^{33,34,38,45–47} However, variable levels of protein S and C were observed in sickle cell patients during acute events compared with steady state, suggesting a more complex biological scenario.^{33,34,38,45–47} In addition, the increased percentage of circulating sickle red cells exposing PS might bind protein S, most likely contributing to protein S reduction.^{48,49}

The prothrombotic state of SCD is also associated with abnormalities in the fibrinolytic system, mainly characterized by increased plasma levels of plasminogen activator inhibitor (PAI)-1 in both steady state and during sickle acute events compared with the normal population.^{36,38,50,51} Because the synthesis of PAI-1 is increased in activated or damaged endothelial cells and also secreted by activated platelets,⁵² the increased PAI-1 levels in sickle cell patients suppressing the normal fibrinolytic system might participate in the pathogenesis of vaso-occlusive events in SCD. Although studies have been performed on coagulation system activation during acute events in SCD, inconclusive data have been reported thus far.^{34,36,38,53,54}

Studies on thrombophilic deoxyribonucleic acid mutations have been performed in patients with SCD to assess their possible impact on sickle cell thrombotic events. Factor V Leiden and the prothrombin variant (FII G20210A) have been evaluated in sickle cell patients of African descent. Because the frequency of these two alleles is low in the African descendent population, the contribution of these two thrombophilic mutations on thrombotic clinical manifestation of SCD seems to be limited.^{55–58} However, in sickle cell patients from eastern Saudi Arabia and Lebanon, a nonsignificantly higher frequency of FII G20210A was observed compared with normal controls.⁵⁹ Moreover, an association between factor V Leiden and venous thrombotic events was shown in Iranian patients with SCD, supporting a different impact of this thrombophilic mutation within sickle cell patients from different ethnic groups.^{60,61}

Studies on methylene tetrahydrofolate reductase (MTHFR) polymorphisms were performed in patients with SCD, but no association between any MTHFR mutation and thrombotic events was high-lighted.^{55,57,62,63}

PLATELETS AND SICKLE CELL DISEASE

Studies on the role of platelets in clinical manifestations of SCD on both steady state and acute events have been partially characterized, and much still remains to be investigated (Table 1). Increased production of thromboxane-A2 and prostaglandin metabolites associated with decreased platelet trombospondin-1 levels, suggesting a chronic activation of platelets, was shown in urine from SCD patients.^{64,65} Other studies also showed increased platelet aggregation in SCD.66-68 Increased platelet activation markers such as P-selectin (CD62), CD63. and activated glycoprotein (GP)IIb/IIIa as well as increased plasma soluble factors as platelet factor (PF)-3, -4 and β-thromboglobulin and platelet-derived soluble CD40 ligand (sCD40L) were reported in SCD patients using a cytofluorimetric approach.36,69-71 Villagra et al suggested that the relative reduction in nitric oxide (NO) bioavailability might participate in triggering platelet activation as well as increasing platelet adhesive properties in patients with SCD.⁷² PS-rich platelets have also been described in SCD patients; these show an increased binding to annexin V that might participate in activation of the coagulation system.³⁶ Recently, Proença-Ferreira et al reported an increased ability of platelets from subjects with SCD to adhere to fibrinogen through modulation of platelet's cyclic adenosine monophosphate content via phosphodiesterase-3A activity associated with increased $\alpha_{IIb}\beta_{3-}$ integrin activation.⁷³ It is interesting to note that in clinical management of SCD patients during acute pain events, the platelet count first decreases, followed by a paradoxical increase associated with higher plasma levels of the platelet products PF4 and β-thromboglobulin (βTG) . This suggests a further amplification of platelet activation during acute events.^{33,74–76} The reduction in platelet content and then the rebound during resolution of acute events have been related to functional asplenia of patients with SCD.^{13,15,65}

Studies on polymorphisms of human platelet alloantigen (HPA), a complex of platelet glycoproteins with other cell-bound factors, showed a possible prothrombotic role in different thrombotic disorders and in sickle cell patients with cerebrovascular events.^{77–80} HPA polymorphisms result in platelet structural changes and/or levels of adhesion proteins. In a case-control study, Al-Subaie et al reported that the HPA-3 variant, which has an isoleucine-to-serine substitution close to the C-terminus of the GPIIb heavy chain, seems to be an independent risk factor for acute vaso-occlusive events in SCD (Table 2).⁸¹

However, no conclusive evidence is actually available on the real impact of thrombotic mutations in SCD mainly due to the limited number of studies and the differences in the genetic background of the sickle cell population studied.

VASCULAR ENDOTHELIUM DYSFUNCTION IN SICKLE CELL DISEASE

SCD patients have shown abnormally activated circulating endothelial cells that increase during acute vasoocclusive crisis, which is compatible with the presence of chronic vascular endothelial damage further worsening during acute events.^{8,54,82–85} Recent studies on the sickle cell-endothelium adhesive mechanism identified different interactions that may have particular relevance in the generation of acute vaso-occlusive events: (1) the integrin $\alpha \alpha_{4b}\beta_1$ receptor of fibronectin and VCAM-1, Eselectin, and P-selectin; (2) the thrombospondin (TSP)

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and/or collagen and receptor CD36, present on the surface of endothelial cells, platelets, and reticulocyterich subpopulations of normal and sickle erythrocytes; (3) the sulfate glycolipids, which bind TSP, VWF multimer, and laminin;^{6,86,87} (4) the Lutheran blood group proteins (BCAM/LU), whose expression is increased in red cells from SCD patients as is their binding to the α 5 subunit of laminin, a component of extracellular subendothelial matrix;^{88,89} (5) the ICAM-4 (Landsteiner-Weiner blood group glycoprotein), which binds $\alpha V\beta$ 3 integrin receptors on endothelial cells;^{90–93} and (6) the exposure of PS, detectable in a subpopulation of sickle red cells, which participates in sickle cell adhesion to activated endothelium^{9,94–97} (Fig. 1).

In SCD patients, increased levels of VWF and, in particular, large VWF multimers were observed and associated with acute vaso-occlusive events.^{98–101} The increased levels of circulating VWF multimers are related to the activity of the metalloprotease ADAMTS 13 (a disintegrin and metalloproteinase with thrombospondin domain 13) that cleaves the hyperadhesive ultra-large VWF under conditions of high fluid shear stress,^{102–104} playing an important role in maintaining the endothelial cell surface free from hyperadhesive ultra-large VWF.¹⁰⁵ Studt et al showed that free hemoglobin can inhibit ADAMTS 13 activity, affecting the VWF cleavage in patients with thrombocytopenic purpura.¹⁰⁶ Schnog et al⁹⁸ reported a decreased ADAMTS 13 activity in

Defenses

 Table 2 Genetic Modifiers of Platelet Activation and Endothelial Function with Effects in Sickle Cell Disease Acute

 Vaso-Occlusive Events

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Genetic Modifier	Target	Effects	References
Human platelets alloantigen (HPA) polymorphism: HPA-3 variant	C-terminus of GPIIb heavy chain on platelets	Independent risk factor for acute vaso-occlusive events	Al-Subaie et al ⁸¹
G1238C VCAM-1 single nucleotide polymorphism	Modulation of vascular endothelial adhesion molecule	Protective effects from stroke in SCD children	Taylor et al ¹⁰⁸
$TNF\text{-}\alpha$ and IL-4 polymorphism	Proinflammatory cytokines	Linked to risk of stroke in SCD children	Hoppe et al ¹⁰⁹
Endothelin-1 (ET-1) polymorphism: ET-1 T8002C	Vascular active molecule	Increased susceptibility to acute chest syndrome	Chaar et al ¹¹⁰
Endothelium nitric oxide synthase (eNOS) polymorphism: T-786C variant	Endothelial NO metabolism	Reduced susceptibility to develop acute chest syndrome in SCD children Increased susceptibility to develop acute chest syndrome in female patients with SCD	Chaar et al ¹¹⁰ Sharan et al ¹¹¹
Klotho (KL) gene polymorphism	β-Galactosidase like proteins involved in endothelial NO homeostasis	Linked to stroke, osteonecrosis, leg ulcers, and priapism	Baldwin et al, ¹¹² Sebastiani et al, ¹¹³ Nolan et al ¹¹⁴
Receptor tyrosine kinase Tie polymorphism	Kinase involved in signaling pathways for chemotaxis and NO metabolism	Linked to leg ulcers	Nolan et al ¹¹⁵
Polymorphism of proteins from bone morphogenic protein pathway	Proteins involved in signaling pathway to protect vascular endothelial surface	Linked to risk factor for stroke, priapism, leg ulcers	Baldwin et al, ¹¹² Sebastiani et al, ¹¹³ Elliott, ¹¹⁶

SCD, sickle cell disease; GP, glycoprotein; VCAM, vascular cell adhesion molecule; TNF, tumor necrosis factor; IL, interleukin; NO, nitric oxide.

sickle cell patients compared with normal controls as well as an associated reduced ADAMTS 13-to-VWF antigen ratio.^{99,107} The same authors further demonstrated an inverse relationship between ADAMTS 13 activity and extracellular hemoglobin levels, suggesting that hemoglobin might compete with VWF for ADAMTS 13. This would cause a relative deficiency of ADAMTS 13, which in turn would block the proteolysis of VWF and lead to the accumulation of hyperadhesive ultra-large VWF on the vascular endothelium surface.^{98,99,107} This mechanism might represent a new element contributing to the complex scenario of microvascular thrombosis in SCD.

Genetic modifiers with functional effects on vascular endothelium homeostasis were evaluated in few studies in patients with SCD (Table 2). Taylor et al showed a protective effect from stroke of G1238C VCAM1 single nucleotide polymorphism in sickle cell patients.¹⁰⁸ Polymorphisms of cytokines involved in proinflammatory responses, such as tumor necrosis factor $(TNF)\alpha$ and interleukin (IL)4, were correlated with the risk of stroke in children with SCD.¹⁰⁹ Polymorphisms of either endothelin-1 (ET-1) and endothelial-NO synthase (eNOS) were linked to the susceptibility of sickle cell patients to acute vaso-occlusive events.^{110,111} Gene polymorphisms in Klotho (KL), encoding for a β glucosidase-like protein involved in endothelial NO homeostasis, were linked to stroke and osteonecrosis in SCD as well as to leg ulcers.^{112–115} A polymorphism of the receptor tyrosine kinase Tie, a factor involved in NO metabolism and leukocyte chemotaxis, was identified as a possible additional genetic modifier associated with leg ulcers in SCD patients.^{112–115} Finally, polymorphisms of genes from bone morphogenic protein signaling pathway involved in protection of the vascular endothelial surface were associated with the risk for different acute vasoocclusive events including stroke, priapism, or leg ulcers.^{112,113,116} Although these studies show the possible role of genetic modifiers of endothelial homeostasis affecting acute vaso-occlusive events in SCD, further investigations should be performed to verify the real impact of these gene polymorphisms on larger SCD patient populations.

INFLAMMATION AND LEUKOCYTES PARTICIPATE IN SICKLE CELL VASO-OCCLUSIVE EVENTS

A chronic inflammatory state has been described in SCD patients characterized by increased plasma levels of acute phase proteins and soluble cytokines such as IL1 β , IL6, TNF- α , and endothelin-1 (ET-1) that are further elevated during acute vaso-occlusive events. These factors participate in leukocyte chemotaxis, modulate vascular tone, and contribute to sickle cell–related tissue damage.^{18,20,85,117–122} Recently, Enenstein et al reported an altered ratio of proinflammatory factor RelA to anti-

inflammatory factor KLF2 in sickle cell children with high risk of stroke, thus suggesting a complex prothrombotic network involving abnormal activated vascular endothelium and inflammation.¹²³

Inflammatory cells such as neutrophils were shown to participate in sickle cell-related vaso-occlusive events; these cells were able more efficiently to adhere to fibronectin and vascular endothelial cells than neutrophils from normal subjects. This phenomenon seems to be related to the higher surface expression of adhesion molecules used for transendothelial migration.^{124,125} In fact, $\beta 2$ integrin Mac 1 ($\alpha_M \beta_2$ or CD11b/CD18) was reported to be increased in neutrophils from SCD patients, suggesting a higher ability of SCD neutrophils to firmly adhere to vascular endothelium surface than normal controls with local reduction of blood flow, which is crucial in the development of acute vasoocclusive events.^{126,127} Hidalgo et al reported an interesting connection between neutrophils, red blood cells, and activated endothelial surface in developing vascular occlusion and hypoxic cell damage in microcirculation of lung from a mouse model of SCD.¹²⁸ These authors hypothesized that the interaction between these blood cell types and the activated endothelium might be coordinated by the interactions of endothelial E-selectin, neutrophil E-selectin ligand (ESL)-1, and the leukocyte integrin Mac1 ($\alpha_M\beta_2$ CD11b/CD18). The heterotypic aggregates are generated by the binding of ESL-1 on neutrophils to E-selecting on the vascular endothelium, which signals the activation of Mac 1 on neutrophils and in turn mediates the heterotypic association of neutrophils with sickle red blood cells.¹²⁸ This interesting pathway, E-selectin-ESL-Mac1, should be further studied in human subjects with SCD and might be explored as a possible target to design new pharmacological strategies in treatment sickle cell-related vasoocclusive events.

Increased iNKT cells, nonphagocytic inflammatory cells producing proinflammatory cytokines were observed in SCD patients, and it was proposed that iNKT cells might contribute to SCD vasculopathy, representing a possible additional risk factor for stroke in sickle cell patients.^{23,129}

THE NITRIC OXIDE CONNECTION

SCD is characterized by a relative reduction in NO bioavailability that contributes to abnormal endothelial activation and SCD organ damage.^{14,16,85,122,130–132} NO is a potent vasodilator and inhibitor of vascular remodeling, and it also affects the multistep cascade of events involved in leukocyte, platelet, and endothelial activation. NO is generated from L-Arginine by endothelial cells via constitutive and inflammatory inducible nitric oxide synthases. Moreover, chronic hemolysis leading to increase the plasma levels of hemoglobin (i.e., an efficient

NO buffer) contributes to reduce the levels of NO in SCD. Thus the NO-reduced bioavailability might contribute to the hypercoagulable state observed in patients with SCD.

ANTIPLATELET AND ANTICOAGULANT AGENTS IN SICKLE CELL DISEASE

Information regarding the use of antiplatelet agents or anticoagulants in the treatment of either acute or chronic clinical manifestation of SCD is still limited due to the small and relative low-quality design of the studies.^{17,133-138} The impact of antiplatelet agents such as aspirin or ticlopidine on either the frequency and/or duration of acute vaso-occlusive crisis were evaluated in a few studies.^{17,133-138} The results were not conclusive because either no differences or limited ameliorations in the frequency and duration of acute vaso-occlusive events in SCD patients were observed.^{17,133–138} Tomer et al recently reported a reduction of pain episodes and in plasma levels of F1 + 2, D-dimer, and the plasminantiplasmin complex in a small cohort of sickle cell patients supplemented with n-3 fatty acid, suggesting a possible correlation between a reduced prothrombotic state and the rate of acute sickle cell-related events.²⁹

Anticoagulant treatment with heparin has been considered as an additional therapeutic approach to block sickle cell adhesion to endothelial cells through the P-selectin pathway^{132,139–141} or binding to TSP that can mediate the interactions between sickle erythrocytes and the vascular endothelial surface. A double-blind randomized trial with tinzaparin in SCD patients during acute vaso-occlusive events documented a reduction of their severity and duration.^{9,142} Studies with a low dose of warfarin or acenocoumarol were reported to slightly reduce the frequency of acute pain events with decreased thrombin generation and fibrinolysis, however without reaching a significant clinical amelioration. This therapy was also associated with frequent bleeding complications.^{98,133,143,144}

Thrombolytic agents in the treatment of stroke in SCD are generally precluded due to the high risk of possible hemorrhage as a complication of thrombolytic therapy in patients presenting with moya-moya disease or pseudoxanthoma elasticum tissue abnormalities.^{23,145,146}

CONCLUSION

The prothrombotic state in SCD contributes to acute and chronic clinical manifestations. Studies have shown abnormalities in the coagulation system, perturbation of platelet activation and aggregation, increased adherence of neutrophils, increased nonphagocytic iNKT cells, and abnormal red cells that more easily adhere on an abnormally activated vascular endothelial surface in SCD. Although progress has been made in characterizing the hypercoagulable state in this challenging disorder, more remains to be investigated, both related to the pathogenesis of vaso-occlusive events and the use of antiplatelet and anticoagulant treatments for a more effective clinical management of SCD patients.

REFERENCES

- Modell B, Darlison M. Global epidemiology of haemoglobin disorders and derived service indicators. Bull World Health Organ 2008;86(6):480–487
- Modell B, Darlison M, Birgens H, et al. Epidemiology of haemoglobin disorders in Europe: an overview. Scand J Clin Lab Invest 2007;67(1):39–69
- Weatherall DJ. The global problem of genetic disease. Ann Hum Biol 2005;32(2):117–122
- Weatherall DJ, Clegg JB. Inherited haemoglobin disorders: an increasing global health problem. Bull World Health Organ 2001;79(8):704–712
- Eaton WA, Hofrichter J. Sickle cell hemoglobin polymerization. Adv Protein Chem 1990;40:63–279
- Steinberg MH. Management of sickle cell disease. N Engl J Med 1999;340(13):1021–1030
- Ballas SK, Smith ED. Red blood cell changes during the evolution of the sickle cell painful crisis. Blood 1992;79(8): 2154–2163
- Solovey AA, Solovey AN, Harkness J, Hebbel RP. Modulation of endothelial cell activation in sickle cell disease: a pilot study. Blood 2001;97(7):1937–1941
- Gayen Betal S, Setty BN. Phosphatidylserine-positive erythrocytes bind to immobilized and soluble thrombospondin-1 via its heparin-binding domain. Transl Res 2008; 152(4): 165–177
- Setty BN, Betal SG, Zhang J, Stuart MJ. Heme induces endothelial tissue factor expression: potential role in hemostatic activation in patients with hemolytic anemia. J Thromb Haemost 2008;6(12):2202–2209
- Setty BN, Rao AK, Stuart MJ. Thrombophilia in sickle cell disease: the red cell connection. Blood 2001;98(12):3228– 3233
- Rosenbaum C, Peace D, Rich E, Van Besien K. Granulocyte colony-stimulating factor-based stem cell mobilization in patients with sickle cell disease. Biol Blood Marrow Transplant 2008;14(6):719–723
- Ataga KI, Cappellini MD, Rachmilewitz EA. Betathalassaemia and sickle cell anaemia as paradigms of hypercoagulability. Br J Haematol 2007;139(1):3–13
- Ataga KI, Orringer EP. Hypercoagulability in sickle cell disease: a curious paradox. Am J Med 2003;115(9): 721–728
- Cappellini MD. Coagulation in the pathophysiology of hemolytic anemias [review]. Hematology Am Soc Hematol Educ Program 2007:74–78
- Morris CR. Mechanisms of vasculopathy in sickle cell disease and thalassemia. Hematology Am Soc Hematol Educ Program 2008:177–185
- Adam SS, Key NS, Greenberg CS. D-dimer antigen: current concepts and future prospects. Blood 2009;113(13):2878– 2887
- Archer DR, Stiles JK, Newman GW, et al. C-reactive protein and interleukin-6 are decreased in transgenic sickle cell mice fed a high protein diet. J Nutr 2008;138(6):1148–1152

- Ou J, Ou Z, Jones DW, et al. L-4F, an apolipoprotein A-1 mimetic, dramatically improves vasodilation in hypercholesterolemia and sickle cell disease. Circulation 2003;107(18): 2337–2341
- Lanaro C, Franco-Penteado CF, Albuqueque DM, Saad ST, Conran N, Costa FF. Altered levels of cytokines and inflammatory mediators in plasma and leukocytes of sickle cell anemia patients and effects of hydroxyurea therapy. J Leukoc Biol 2009;85(2):235–242
- Austin H, Key NS, Benson JM, et al. Sickle cell trait and the risk of venous thromboembolism among blacks. Blood 2007;110(3):908–912
- Stein PD, Beemath A, Meyers FA, Skaf E, Olson RE. Deep venous thrombosis and pulmonary embolism in hospitalized patients with sickle cell disease. Am J Med 2006;119(10):897; e7–e11
- Verduzco LA, Nathan DG. Sickle cell disease and stroke. Blood 2009;114(25):5117–5125
- 24. James AH, Jamison MG, Brancazio LR, Myers ER. Venous thromboembolism during pregnancy and the postpartum period: incidence, risk factors, and mortality. Am J Obstet Gynecol 2006;194(5):1311–1315
- Prengler M, Pavlakis SG, Prohovnik I, Adams RJ. Sickle cell disease: the neurological complications. Ann Neurol 2002;51(5):543–552
- Adedeji MO, Cespedes J, Allen K, Subramony C, Hughson MD. Pulmonary thrombotic arteriopathy in patients with sickle cell disease. Arch Pathol Lab Med 2001;125(11):1436– 1441
- Haque AK, Gokhale S, Rampy BA, Adegboyega P, Duarte A, Saldana MJ. Pulmonary hypertension in sickle cell hemoglobinopathy: a clinicopathologic study of 20 cases. Hum Pathol 2002;33(10):1037–1043
- Peters M, Plaat BE, ten Cate H, Wolters HJ, Weening RS, Brandjes DP. Enhanced thrombin generation in children with sickle cell disease. Thromb Haemost 1994;71(2):169–172
- Tomer A, Kasey S, Connor WE, Clark S, Harker LA, Eckman JR. Reduction of pain episodes and prothrombotic activity in sickle cell disease by dietary n-3 fatty acids. Thromb Haemost 2001;85(6):966–974
- Leslie J, Langler D, Serjeant GR, Serjeant BE, Desai P, Gordon YB. Coagulation changes during the steady state in homozygous sickle-cell disease in Jamaica. Br J Haematol 1975;30(2):159–166
- Rickles FR, O'Leary DS. Role of coagulation system in pathophysiology of sickle cell disease. Arch Intern Med 1974;133(4):635-641
- 32. Stuart MJ, Setty BN. Hemostatic alterations in sickle cell disease: relationships to disease pathophysiology. Pediatr Pathol Mol Med 2001;20(1):27–46
- Green D, Scott JP. Is sickle cell crisis a thrombotic event? Am J Hematol 1986;23(4):317–321
- Francis RB Jr. Elevated fibrin D-dimer fragment in sickle cell anemia: evidence for activation of coagulation during the steady state as well as in painful crisis. Haemostasis 1989; 19(2):105–111
- Richardson SG, Matthews KB, Stuart J, Geddes AM, Wilcox RM. Serial changes in coagulation and viscosity during sickle-cell crisis. Br J Haematol 1979;41(1):95–103
- Tomer A, Harker LA, Kasey S, Eckman JR. Thrombogenesis in sickle cell disease. J Lab Clin Med 2001;137(6):398–407
- Kurantsin-Mills J, Ofosu FA, Safa TK, Siegel RS, Lessin LS. Plasma factor VII and thrombin-antithrombin III levels

indicate increased tissue factor activity in sickle cell patients. Br J Haematol 1992;81(4):539–544

- Westerman MP, Green D, Gilman-Sachs A, et al. Antiphospholipid antibodies, proteins C and S, and coagulation changes in sickle cell disease. J Lab Clin Med 1999;134(4): 352–362
- Hagger D, Wolff S, Owen J, Samson D. Changes in coagulation and fibrinolysis in patients with sickle cell disease compared with healthy black controls. Blood Coagul Fibrinolysis 1995;6(2):93–99
- 40. Branch DW, Rodgers GM. Induction of endothelial cell tissue factor activity by sera from patients with antiphospholipid syndrome: a possible mechanism of thrombosis. Am J Obstet Gynecol 1993;168(1 Pt 1):206–210
- Ataga KI, Moore CG, Hillery CA, et al. Coagulation activation and inflammation in sickle cell disease-associated pulmonary hypertension. Haematologica 2008;93(1): 20–26
- 42. van Beers EJ, Spronk HM, Ten Cate H, et al; CURAMA study Group. No association of the hypercoagulable state with sickle cell disease related pulmonary hypertension. Haematologica 2008;93(5):e42–e44
- 43. Shet AS, Aras O, Gupta K, et al. Sickle blood contains tissue factor-positive microparticles derived from endothelial cells and monocytes. Blood 2003;102(7):2678–2683
- 44. van Beers EJ, Schaap MC, Berckmans RJ, et al; CURAMA study group. Circulating erythrocyte-derived microparticles are associated with coagulation activation in sickle cell disease. Haematologica 2009;94(11):1513–1519
- 45. Tam DA. Protein C and protein S activity in sickle cell disease and stroke. J Child Neurol 1997;12(1):19–21
- el-Hazmi MA, Warsy AS, Bahakim H. Blood proteins C and S in sickle cell disease. Acta Haematol 1993;90(3):114–119
- Karayalcin G, Lanzkowsky P. Plasma protein C levels in children with sickle cell disease. Am J Pediatr Hematol Oncol 1989;11(3):320–323
- Kuypers FA, Larkin SK, Emeis JJ, Allison AC. Interaction of an annexin V homodimer (Diannexin) with phosphatidylserine on cell surfaces and consequent antithrombotic activity. Thromb Haemost 2007;97(3):478–486
- Lane PA, O'Connell JL, Marlar RA. Erythrocyte membrane vesicles and irreversibly sickled cells bind protein S. Am J Hematol 1994;47(4):295–300
- Nsiri B, Gritli N, Bayoudh F, Messaoud T, Fattoum S, Machghoul S. Abnormalities of coagulation and fibrinolysis in homozygous sickle cell disease. Hematol Cell Ther 1996; 38(3):279–284
- Nsiri B, Gritli N, Mazigh C, Ghazouani E, Fattoum S, Machghoul S. Fibrinolytic response to venous occlusion in patients with homozygous sickle cell disease. Hematol Cell Ther 1997;39(5):229–232
- Yamamoto K, Saito H. A pathological role of increased expression of plasminogen activator inhibitor-1 in human or animal disorders. Int J Hematol 1998;68(4):371–385
- Key NS, Slungaard A, Dandelet L, et al. Whole blood tissue factor procoagulant activity is elevated in patients with sickle cell disease. Blood 1998;91(11):4216–4223
- Solovey A, Gui L, Key NS, Hebbel RP. Tissue factor expression by endothelial cells in sickle cell anemia. J Clin Invest 1998;101(9):1899–1904
- 55. Andrade FL, Annichino-Bizzacchi JM, Saad ST, Costa FF, Arruda VR. Prothrombin mutant, factor V Leiden, and thermolabile variant of methylenetetrahydrofolate reductase

among patients with sickle cell disease in Brazil. Am J Hematol 1998;59(1):46–50

- 56. Kahn MJ, Scher C, Rozans M, Michaels RK, Leissinger C, Krause J. Factor V Leiden is not responsible for stroke in patients with sickling disorders and is uncommon in African Americans with sickle cell disease. Am J Hematol 1997;54(1): 12–15
- Cumming AM, Olujohungbe A, Keeney S, Singh H, Hay CR, Serjeant GR. The methylenetetrahydrofolate reductase gene C677T polymorphism in patients with homozygous sickle cell disease and stroke. Br J Haematol 1999;107(3):569– 571
- Rees DC, Chapman NH, Webster MT, Guerreiro JF, Rochette J, Clegg JB. Born to clot: the European burden. Br J Haematol 1999;105(2):564–566
- Rahimi Z, Vaisi-Raygani A, Nagel RL, Muniz A. Thrombophilic mutations among Southern Iranian patients with sickle cell disease: high prevalence of factor V Leiden. J Thromb Thrombolysis 2008;25(3):288–292
- 60. Isma'eel H, Arnaout MS, Shamseddeen W, et al. Screening for inherited thrombophilia might be warranted among Eastern Mediterranean sickle-beta-0 thalassemia patients. J Thromb Thrombolysis 2006;22(2):121–123
- 61. Fawaz NA, Bashawery L, Al-Sheikh I, Qatari A, Al-Othman SS, Almawi WY. Factor V-Leiden, prothrombin G20210A, and MTHFR C677T mutations among patients with sickle cell disease in Eastern Saudi Arabia. Am J Hematol 2004;76(3):307–309
- 62. Zimmerman SA, Ware RE. Inherited DNA mutations contributing to thrombotic complications in patients with sickle cell disease. Am J Hematol 1998;59(4):267–272
- Adekile AD, Kutlar F, Haider MZ, Kutlar A. Frequency of the 677 C—> T mutation of the methylenetetrahydrofolate reductase gene among Kuwaiti sickle cell disease patients. Am J Hematol 2001;66(4):263–266
- 64. Foulon I, Bachir D, Galacteros F, Maclouf J. Increased in vivo production of thromboxane in patients with sickle cell disease is accompanied by an impairment of platelet functions to the thromboxane A2 agonist U46619. Arterioscler Thromb 1993;13(3):421–426
- 65. Browne PV, Mosher DF, Steinberg MH, Hebbel RP. Disturbance of plasma and platelet thrombospondin levels in sickle cell disease. Am J Hematol 1996;51(4):296–301
- Kenny MW, George AJ, Stuart J. Platelet hyperactivity in sickle-cell disease: a consequence of hyposplenism. J Clin Pathol 1980;33(7):622–625
- Westwick J, Watson-Williams EJ, Krishnamurthi S, et al. Platelet activation during steady state sickle cell disease. J Med 1983;14(1):17–36
- Winichagoon P, Fucharoen S, Wasi P. Increased circulating platelet aggregates in thalassaemia. Southeast Asian J Trop Med Public Health 1981;12(4):556–560
- Wun T, Paglieroni T, Rangaswami A, et al. Platelet activation in patients with sickle cell disease. Br J Haematol 1998;100(4):741–749
- Famodu AA, Oduwa D. Platelet count and platelet factor 3 (PF-3) availability in sickle cell disease. Br J Biomed Sci 1995;52(4):323–324
- Lee SP, Ataga KI, Orringer EP, Phillips DR, Parise LV. Biologically active CD40 ligand is elevated in sickle cell anemia: potential role for platelet-mediated inflammation. Arterioscler Thromb Vasc Biol 2006;26(7):1626– 1631

- 72. Villagra J, Shiva S, Hunter LA, Machado RF, Gladwin MT, Kato GJ. Platelet activation in patients with sickle disease, hemolysis-associated pulmonary hypertension, and nitric oxide scavenging by cell-free hemoglobin. Blood 2007; 110(6):2166–2172
- Proença-Ferreira R, Franco-Penteado CF, Traina F, Saad ST, Costa FF, Conran N. Increased adhesive properties of platelets in sickle cell disease: roles for alphaIIb beta3mediated ligand binding, diminished cAMP signalling and increased phosphodiesterase 3A activity. Br J Haematol 2010;149(2):280–288
- Mehta P, Mehta J. Abnormalities of platelet aggregation in sickle cell disease. J Pediatr 1980;96(2):209–213
- Haut MJ, Cowan DH, Harris JW. Platelet function and survival in sickle cell disease. J Lab Clin Med 1973;82(1): 44–53
- Buchanan GR, Holtkamp CA. Evidence against enhanced platelet activity in sickle cell anaemia. Br J Haematol 1983; 54(4):595–603
- 77. Kunicki TJ. The influence of platelet collagen receptor polymorphisms in hemostasis and thrombotic disease. Arterioscler Thromb Vasc Biol 2002;22(1):14–20
- Deckmyn H, Ulrichts H, Van De Walle G, Vanhoorelbeke K. Platelet antigens and their function. Vox Sang 2004; 87(Suppl 2):105–111
- Bray PF. Platelet glycoprotein polymorphisms as risk factors for thrombosis. Curr Opin Hematol 2000;7(5):284–289
- Castro V, Alberto FL, Costa RN, et al. Polymorphism of the human platelet antigen-5 system is a risk factor for occlusive vascular complications in patients with sickle cell anemia. Vox Sang 2004;87(2):118–123
- Al-Subaie AM, Fawaz NA, Mahdi N, et al. Human platelet alloantigens (HPA) 1, HPA2, HPA3, HPA4, and HPA5 polymorphisms in sickle cell anemia patients with vasoocclusive crisis. Eur J Haematol 2009;83(6):579–585
- Solovey A, Gui L, Ramakrishnan S, Steinberg MH, Hebbel RP. Sickle cell anemia as a possible state of enhanced antiapoptotic tone: survival effect of vascular endothelial growth factor on circulating and unanchored endothelial cells. Blood 1999;93(11):3824–3830
- Ortiz A. Circulating endothelial cells in sickle cell anemia. N Engl J Med 1998;338(16):1162; author reply 1162–1163
- Parise LV, Telen MJ. Erythrocyte adhesion in sickle cell disease. Curr Hematol Rep 2003;2(2):102–108
- De Franceschi L, Corrocher R. Established and experimental treatments for sickle cell disease. Haematologica 2004;89(3): 348–356
- Kaul DK, Tsai HM, Liu XD, Nakada MT, Nagel RL, Coller BS. Monoclonal antibodies to alphaVbeta3 (7E3 and LM609) inhibit sickle red blood cell-endothelium interactions induced by platelet-activating factor. Blood 2000;95(2): 368–374
- Barabino GA, Liu XD, Ewenstein BM, Kaul DK. Anionic polysaccharides inhibit adhesion of sickle erythrocytes to the vascular endothelium and result in improved hemodynamic behavior. Blood 1999;93(4):1422–1429
- Hines PC, Zen Q, Burney SN, et al. Novel epinephrine and cyclic AMP-mediated activation of BCAM/Lu-dependent sickle (SS) RBC adhesion. Blood 2003; 101(8):3281– 3287
- Murphy MM, Zayed MA, Evans A, et al. Role of Rap1 in promoting sickle red blood cell adhesion to laminin via BCAM/LU. Blood 2005;105(8):3322–3329

- Zennadi R, Hines PC, De Castro LM, Cartron JP, Parise LV, Telen MJ. Epinephrine acts through erythroid signaling pathways to activate sickle cell adhesion to endothelium via LW-alphavbeta3 interactions. Blood 2004;104(12):3774– 3781
- Zennadi R, De Castro L, Eyler C, Xu K, Ko M, Telen MJ. Role and regulation of sickle red cell interactions with other cells: ICAM-4 and other adhesion receptors. Transfus Clin Biol 2008;15(1-2):23–28
- Kaul DK, Liu XD, Zhang X, et al. Peptides based on alphaVbinding domains of erythrocyte ICAM-4 inhibit sickle red cell-endothelial interactions and vaso-occlusion in the microcirculation. Am J Physiol Cell Physiol 2006;291(5):C922– C930
- Mankelow TJ, Spring FA, Parsons SF, et al. Identification of critical amino-acid residues on the erythroid intercellular adhesion molecule-4 (ICAM-4) mediating adhesion to alpha V integrins. Blood 2004;103(4):1503–1508
- 94. Sabina RL, Wandersee NJ, Hillery CA. Ca2+-CaM activation of AMP deaminase contributes to adenine nucleotide dysregulation and phosphatidylserine externalization in human sickle erythrocytes. Br J Haematol 2009;144(3):434– 445
- Hebbel RP. Adhesion of sickle red cells to endothelium: myths and future directions. Transfus Clin Biol 2008;15(1-2): 14–18
- Kuypers FA, Styles LA. The role of secretory phospholipase A2 in acute chest syndrome. Cell Mol Biol (Noisy-le-grand) 2004;50(1):87–94
- Kuypers FA, de Jong K. The role of phosphatidylserine in recognition and removal of erythrocytes. Cell Mol Biol (Noisy-le-grand) 2004;50(2):147–158
- Schnog JJ, Hovinga JA, Krieg S, et al; CURAMA Study Group. ADAMTS13 activity in sickle cell disease. Am J Hematol 2006;81(7):492–498
- Krishnan S, Siegel J, Pullen GJr, Hevelow M, Dampier C, Stuart M. Increased von Willebrand factor antigen and high molecular weight multimers in sickle cell disease associated with nocturnal hypoxemia. Thromb Res 2008;122(4):455– 458
- Roberts DD, Williams SB, Gralnick HR, Ginsburg V. von Willebrand factor binds specifically to sulfated glycolipids. J Biol Chem 1986;261(7):3306–3309
- Kaul DK, Nagel RL, Chen D, Tsai HM. Sickle erythrocyteendothelial interactions in microcirculation: the role of von Willebrand factor and implications for vasoocclusion. Blood 1993;81(9):2429–2438
- Tsai HM. Current concepts in thrombotic thrombocytopenic purpura. Annu Rev Med 2006;57:419–436
- 103. Sadler JE. A new name in thrombosis, ADAMTS13. Proc Natl Acad Sci U S A 2002;99(18):11552–11554
- 104. Feys HB, Anderson PJ, Vanhoorelbeke K, Majerus EM, Sadler JE. Multi-step binding of ADAMTS-13 to von Willebrand factor. J Thromb Haemost 2009;7(12):2088– 2095
- 105. Turner NA, Nolasco L, Ruggeri ZM, Moake JL. Endothelial cell ADAMTS-13 and VWF: production, release, and VWF string cleavage. Blood 2009;114(24): 5102–5111
- 106. Studt JD, Hovinga JA, Antoine G, et al. Fatal congenital thrombotic thrombocytopenic purpura with apparent ADAMTS13 inhibitor: in vitro inhibition of ADAMTS13 activity by hemoglobin. Blood 2005;105(2):542–544

- 107. Zhou Z, Han H, Cruz MA, López JA, Dong JF, Guchhait P. Haemoglobin blocks von Willebrand factor proteolysis by ADAMTS-13: a mechanism associated with sickle cell disease. Thromb Haemost 2009;101(6):1070–1077
- Taylor JG VI, Tang DC, Savage SA, et al. Variants in the VCAM1 gene and risk for symptomatic stroke in sickle cell disease. Blood 2002;100(13):4303–4309
- 109. Hoppe C, Klitz W, D'Harlingue K, et al; Stroke Prevention Trial in Sickle Cell Anemia (STOP) Investigators. Confirmation of an association between the TNF(-308) promoter polymorphism and stroke risk in children with sickle cell anemia. Stroke 2007;38(8):2241–2246
- 110. Chaar V, Tarer V, Etienne-Julan M, Diara JP, Elion J, Romana M. ET-1 and ecNOS gene polymorphisms and susceptibility to acute chest syndrome and painful vasoocclusive crises in children with sickle cell anemia. Haematologica 2006;91(9):1277–1278
- 111. Sharan K, Surrey S, Ballas S, et al. Association of T-786C eNOS gene polymorphism with increased susceptibility to acute chest syndrome in females with sickle cell disease. Br J Haematol 2004;124(2):240–243
- 112. Baldwin C, Nolan VG, Wyszynski DF, et al. Association of klotho, bone morphogenic protein 6, and annexin A2 polymorphisms with sickle cell osteonecrosis. Blood 2005; 106(1):372–375
- Sebastiani P, Ramoni MF, Nolan V, Baldwin CT, Steinberg MH. Genetic dissection and prognostic modeling of overt stroke in sickle cell anemia. Nat Genet 2005; 37(4):435–440
- Nolan VG, Baldwin C, Ma Q, et al. Association of single nucleotide polymorphisms in klotho with priapism in sickle cell anaemia. Br J Haematol 2005;128(2):266–272
- 115. Nolan VG, Adewoye A, Baldwin C, et al. Sickle cell leg ulcers: associations with haemolysis and SNPs in Klotho, TEK and genes of the TGF-beta/BMP pathway. Br J Haematol 2006;133(5):570–578
- 116. Elliott L, Ashley-Koch AE, De Castro L, et al. Genetic polymorphisms associated with priapism in sickle cell disease. Br J Haematol 2007;137(3):262–267
- 117. Graido-Gonzalez E, Doherty JC, Bergreen EW, Organ G, Telfer M, McMillen MA. Plasma endothelin-1, cytokine, and prostaglandin E2 levels in sickle cell disease and acute vaso-occlusive sickle crisis. Blood 1998;92(7): 2551–2555
- Natarajan M, Udden MM, McIntire LV. Adhesion of sickle red blood cells and damage to interleukin-1 beta stimulated endothelial cells under flow in vitro. Blood 1996;87(11): 4845–4852
- Croizat H. Circulating cytokines in sickle cell patients during steady state. Br J Haematol 1994;87(3):592–597
- Francis RB Jr, Haywood LJ. Elevated immunoreactive tumor necrosis factor and interleukin-1 in sickle cell disease. J Natl Med Assoc 1992;84(7):611–615
- 121. Hibbert JM, Hsu LL, Bhathena SJ, et al. Proinflammatory cytokines and the hypermetabolism of children with sickle cell disease. Exp Biol Med (Maywood) 2005;230(1): 68–74
- Conran N, Costa FF. Hemoglobin disorders and endothelial cell interactions. Clin Biochem 2009;42(18):1824–1838
- 123. Enenstein J, Milbauer L, Domingo E, et al. Proinflammatory phenotype with imbalance of KLF2 and RelA: risk of childhood stroke with sickle cell anemia. Am J Hematol 2010;85(1):18–23

- 124. Fadlon E, Vordermeier S, Pearson TC, et al. Blood polymorphonuclear leukocytes from the majority of sickle cell patients in the crisis phase of the disease show enhanced adhesion to vascular endothelium and increased expression of CD64. Blood 1998;91(1):266–274
- 125. Canalli AA, Franco-Penteado CF, Saad ST, Conran N, Costa FF. Increased adhesive properties of neutrophils in sickle cell disease may be reversed by pharmacological nitric oxide donation. Haematologica 2008;93(4):605–609
- Lum AF, Wun T, Staunton D, Simon SI. Inflammatory potential of neutrophils detected in sickle cell disease. Am J Hematol 2004;76(2):126–133
- 127. Assis A, Conran N, Canalli AA, Lorand-Metze I, Saad ST, Costa FF. Effect of cytokines and chemokines on sickle neutrophil adhesion to fibronectin. Acta Haematol 2005; 113(2):130–136
- 128. Hidalgo A, Chang J, Jang JE, Peired AJ, Chiang EY, Frenette PS. Heterotypic interactions enabled by polarized neutrophil microdomains mediate thromboinflammatory injury. Nat Med 2009;15(4):384–391
- 129. Wallace KL, Marshall MA, Ramos SI, et al. NKT cells mediate pulmonary inflammation and dysfunction in murine sickle cell disease through production of IFN-gamma and CXCR3 chemokines. Blood 2009;114(3):667–676
- de Franceschi L, Baron A, Scarpa A, et al. Inhaled nitric oxide protects transgenic SAD mice from sickle cell diseasespecific lung injury induced by hypoxia/reoxygenation. Blood 2003;102(3):1087–1096
- Yang Y, Loscalzo J. Regulation of tissue factor expression in human microvascular endothelial cells by nitric oxide. Circulation 2000;101(18):2144–2148
- 132. Kato GJ, Martyr S, Blackwelder WC, et al. Levels of soluble endothelium-derived adhesion molecules in patients with sickle cell disease are associated with pulmonary hypertension, organ dysfunction, and mortality. Br J Haematol 2005;130(6):943–953
- Chaplin HJr, Alkjaersig N, Fletcher AP, Michael JM, Joist JH. Aspirin-dipyridamole prophylaxis of sickle cell disease pain crises. Thromb Haemost 1980;43(3):218–221
- Osamo NO, Photiades DP, Famodu AA. Therapeutic effect of aspirin in sickle cell anaemia. Acta Haematol 1981;66(2): 102–107

- Greenberg J, Ohene-Frempong K, Halus J, Way C, Schwartz E. Trial of low doses of aspirin as prophylaxis in sickle cell disease. J Pediatr 1983;102(5):781–784
- Semple MJ, Al-Hasani SF, Kioy P, Savidge GF. A doubleblind trial of ticlopidine in sickle cell disease. Thromb Haemost 1984;51(3):303–306
- 137. Cabannes R, Lonsdorfer J, Castaigne JP, Ondo A, Plassard A, Zohoun I. Clinical and biological double-blind-study of ticlopidine in preventive treatment of sickle-cell disease crises. Agents Actions Suppl 1984;15:199–212
- Zago MA, Costa FF, Ismael SJ, Tone LG, Bottura C. Treatment of sickle cell diseases with aspirin. Acta Haematol 1984;72(1):61–64
- 139. Mohan JS, Lip GY, Wright J, Bareford D, Blann AD. Plasma levels of tissue factor and soluble E-selectin in sickle cell disease: relationship to genotype and to inflammation. Blood Coagul Fibrinolysis 2005;16(3):209–214
- Wood K, Russell J, Hebbel RP, Granger DN. Differential expression of E- and P-selectin in the microvasculature of sickle cell transgenic mice. Microcirculation 2004;11(4): 377–385
- 141. Blum A, Yeganeh S, Peleg A, et al. Endothelial function in patients with sickle cell anemia during and after sickle cell crises. J Thromb Thrombolysis 2005;19(2):83–86
- 142. Qari MH, Aljaouni SK, Alardawi MS, et al. Reduction of painful vaso-occlusive crisis of sickle cell anaemia by tinzaparin in a double-blind randomized trial. Thromb Haemost 2007;98(2):392–396
- 143. Ahmed S, Siddiqui AK, Iqbal U, et al. Effect of low-dose warfarin on D-dimer levels during sickle cell vaso-occlusive crisis: a brief report. Eur J Haematol 2004;72(3):213–216
- 144. Salvaggio JE, Arnold CA, Banov CH. Long-term anticoagulation in sickle-cell disease. A clinical study. N Engl J Med 1963;269:182–186
- Dobson SR, Holden KR, Nietert PJ, et al. Moyamoya syndrome in childhood sickle cell disease: a predictive factor for recurrent cerebrovascular events. Blood 2002;99(9):3144– 3150
- 146. Aessopos A, Farmakis D, Loukopoulos D. Elastic tissue abnormalities resembling pseudoxanthoma elasticum in beta thalassemia and the sickling syndromes. Blood 2002;99(1): 30–35