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# Wild Bees Preferentially Visit Rudbeckia Flower Heads with Exaggerated Ultraviolet Absorbing Floral Guides

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## RESEARCH ARTICLE

# Wild bees preferentially visit *Rudbeckia* flower heads with exaggerated ultraviolet absorbing floral guides

Lisa Horth\*, Laura Campbell and Rebecca Bray

## ABSTRACT

Here, we report on the results of an experimental study that assessed the visitation frequency of wild bees to conspecific flowers with different sized floral guides. UV absorbent floral guides are ubiquitous in Angiosperms, yet surprisingly little is known about conspecific variation in these guides and very few studies have evaluated pollinator response to UV guide manipulation. This is true despite our rich understanding about learning and color preferences in bees. Historical dogma indicates that flower color serves as an important long-range visual signal allowing pollinators to detect the flowers, while floral guides function as close-range signals that direct pollinators to a reward. We initiated the work presented here by first assessing the population level variation in UV absorbent floral guides for conspecific flowers. We assessed two species, *Rudbeckia hirta* and *R. fulgida*. We then used several petal cut-and-paste experiments to test whether UV floral guides can also function to attract visitors. We manipulated floral guide size and evaluated visitation frequency. In all experiments, pollinator visitation rates were clearly associated with floral guide size. Diminished floral guides recruited relatively few insect visitors. Exaggerated floral guides recruited more visitors than smaller or average sized guides. Thus, UV floral guides play an important role in pollinator recruitment and in determining the relative attractiveness of conspecific flower heads. Consideration of floral guides is therefore important when evaluating the overall conspicuousness of flower heads relative to background coloration. This work raises the issue of whether floral guides serve as honest indicators of reward, since guide size varies in nature for conspecific flowers at the same developmental stage and since preferences for larger guides were found. To our knowledge, these are the first cut-and-paste experiments conducted to examine whether UV absorbent floral guides affect visitation rates and pollinator preference.

**KEY WORDS:** Bee, *Rudbeckia*, Flower, Pollination, Opsin, Vision, Pollinator

## INTRODUCTION

A comprehensive, high quality literature exists regarding the sensory and cognitive abilities of bees. This research includes

extensive data on essential tasks associated with foraging, like visual choice, learning and memory (e.g. von Frisch, 1914; Daumer, 1956; Daumer, 1958; Backhaus, 1991; Giurfa et al., 1996; Dyer et al., 2008; Dyer et al., 2011; Spaethe et al., 2001). Bees and humans both have trichromatic vision, but bee vision is short wave shifted compared to humans. Retinal photoreceptors of bees are short (SWS), medium (MWS), and long wavelength sensitive (LWS) and are also classified as ultraviolet (UV), blue, and green sensitive. Data from 43 species of hymenoptera demonstrate maximal receptor sensitivity ( $\lambda$  max) at ~340 nm, 430 nm, and 535 nm, respectively (Peitsch et al., 1992; Menzel and Blakers, 1976; Menzel and Backhaus, 1991 and references therein).

The color space perceived by bees has been determined using mathematical modeling and multidimensional scaling analysis that employed empirical color-choice test results (Backhaus et al., 1987; Backhaus, 1991; Chittka et al., 1992). The color differences potentially detectable by bees were calculated for different chromatic dimensions (e.g. uv/blue-green and blue/uv-green axes) (Backhaus, 1991). Neurons with the same photoreceptor antagonism have been identified in the medulla and lobula of the bee brain (Kien and Menzel, 1977a; Kien and Menzel, 1977b). The spectral sensitivities predicted for hypothetical color opponent coding (COC) cells compared well to empirical measures for antagonistically coding neurons (Backhaus, 1991).

For bees to be able to detect flowers in the distance, contrast between floral spectral reflectance and background reflectance is necessary (e.g. Kevan, 1978; Chittka and Menzel, 1992). Green (LWS) receptor contrast is particularly important for distance-based detection, which is believed to occur prior to chromatic contrast (Giurfa et al., 1996). Green contrast is important for motion processing (Srinivasan and Lehrer, 1984) and detection of object edges (Lehrer et al., 1990). In honeybees, green contrast is detected by LWS receptors when the visual angle is small (5–15°), whereas chromatic traits are detected by SWS and MWS receptors at greater visual angles (>15°) (Giurfa et al., 1997; Giurfa and Vorobyev, 1998; Spaethe et al., 2001; Dyer et al., 2011).

From an evolutionary perspective, a trade off was found between foraging accuracy and decision time for bumble bees (*Bombus L. terrestris*). When foraging in a virtual flower meadow where ‘flowers’ had similar colors but different rewards, bees were more accurate when they took longer to make foraging choices (Chittka et al., 2003). Fast and slow bees did learn to slow down and improve accuracy with aversion stimuli (Chittka et al., 2003). However, fast bees had higher nectar collection rates (Burns, 2005), so costs are associated with slow, accurate decision-making.

In nature, competition for pollination is known to contribute to natural selection on floral traits (Caruso, 2000) and visual cues clearly affect conspecific floral attractiveness to pollinators. Field studies have shown that yellow flowered wild radishes (*Raphanus raphanistrum* L.) had higher pollination rates than white ones (Stanton et al., 1989) and deep blue flowered montane larkspur

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(*Delphinium nelsonii* Greene, *Delphinium nuttallianum* Pritz. ex Walp) had higher pollination rates than albinos (Waser and Price, 1981). Recent work with model flowers showed that bumble bees (*B. terrestris*) have innate preference for high spectral purity (Rohde et al., 2013). The value of colorful floral traits that we see is less contentious than the value of traits invisible to us but visible to bees.

The scientific community has ranged the gamut from intense interest in UV traits (Bennett and Cuthill, 1994), to discounting what was considered exaggerated attention and promoting the idea that UV is not a special channel for communication (e.g. Chittka et al., 1994). Middle ground was reached with demonstrations that floral UV reflection can be important, though sometimes no more important than blue, green or red reflection (Kevan et al., 2001). Despite the fact that UV reflective and absorptive floral patterns are widespread in nature and visible to pollinators (von Frisch, 1967; Daumer, 1958; Eisner et al., 1969; Silberglied, 1979; Höglund et al., 1973; Burkhardt, 1982; McCrea and Levy, 1983; Peitsch et al., 1989; Menzel and Backhaus, 1991; Chittka and Menzel, 1992; Peitsch et al., 1992; Bennett et al., 1996; Dyer, 1996; Briscoe et al., 2003; Winter et al., 2003; Skorupski and Chittka, 2010) few studies have empirically manipulated UV floral guides to assess bee visitation frequency or preference.

Early on Daumer elucidated the point that primary colors combine to create ‘novel’ colors visible to bees but not humans (Daumer, 1956; Daumer, 1958; see also Backhaus, 1991; Giurfa et al., 1995). Later, when Waser and Price were studying foraging economics, they painted albino montane larkspurs (Waser and Price, 1985). They showed that adding UV absorbent blue paint to albino sepals and guard petals that typically reflect UV enhanced visitation and decreased foraging time. So did performing this manipulation on albino nectariferous petals that typically absorb UV, making this a blue-only manipulation (Waser and Price, 1985).

Burr et al. later discussed the function of UV reflection as a long-range landing site cue (Burr et al., 1995). Then Eisner illustrated how ‘human-yellow’ on the distal half of *Rudbeckia hirta* petals was UV reflective (Eisner, 2002) and referred to this as ‘bee purple’ (originally described by Daumer, 1956; Daumer, 1958). Around this time Johnson and Anderson conducted field studies with the African potato (or star grass, *Hypoxis hemerocallidea* Fisch. & C. A. Mey) and showed that, when floral UV reflectance was obscured with sunscreen, fewer honey bees (*A. mellifera scutella*) approached and landed on flowers (Johnson and Andersson, 2002).

Burr et al. also discussed the use of very small UV absorptive areas on flowers as guides for orientation to reward (Burr et al., 1995). Such orientation cues have been called honey- or nectar-guides and will hereafter be referred to as floral guides. UV absorbance is quite common in these floral guides. In a tropical field study, UV absorbent banner petals were repositioned on legume (*Caesalpinia eriostachys* Benth. and *Parkinsonia aculeata* L.) flowers to successfully demonstrate their orientation function (Jones and Buchmann, 1974). Chemically, flavonol glucosides are primarily responsible for the large, UV absorbing floral guide in the floral heads of some asters, including *Rudbeckia hirta* L. (Thompson et al., 1972; Schlangen et al., 2009), one of the species used in the work we present here.

Our studies involved wild, cultivated and empirically manipulated asters (*R. hirta* and *R. fulgida*). First, we report novel findings regarding the distribution of floral guide sizes in natural populations and cultivars. We hypothesize that UV

absorbent floral guides are important for attracting pollinators, not just for orientation, and that larger guides will be more attractive than smaller ones. To test these ideas we compare pollinator visitation rates after enlarging and diminishing natural UV-absorbent orientation cues. We focus on Halictidae (sweat bees), the relatively poorly studied, yet numerically abundant, cosmopolitan generalist pollinators frequently found in urban ecosystems (Dikmen, 2007). This work was completed using a ‘cut-and-paste’ design akin to Andersson’s widowbird tail-length studies (Andersson, 1982). This is a timely demonstration that the size of UV absorbent floral guides clearly affects native bee visitation rates and that these floral guides may not be used solely for orientation toward a reward, as once believed, but also play a role in pollinator recruitment.

## RESULTS

### I. Floral specimens and relevant techniques for measuring UV floral guides

Color and ultraviolet absorbing *Rudbeckia hirta* photographs are presented in Fig. 1A,B, along with the spectrophotometric graph of floral reflectance (Fig. 1C) showing the different reflectance patterns in the distal and proximal portions of *R. hirta* ligules. Relevant measurements and the photographs of ‘cut-and-paste’ treatments for each experiment are presented in the associated Results sections below.

### II. Floral guide size-distributions for three data sets: greenhouse, urban, and wild *R. hirta*

#### Greenhouse flowers

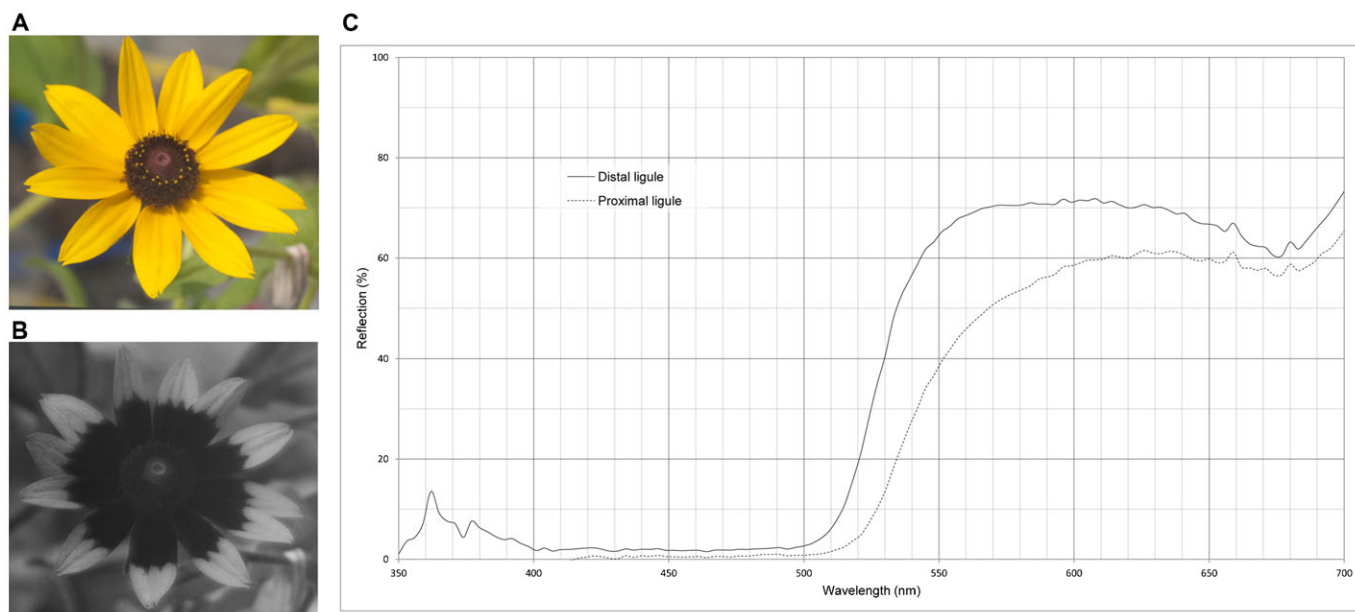
The average UV floral guides in greenhouse plants covered 62% (standard error 1.3%) of the petal length for flowers from the first plant and 57% (standard error 0.8%) of the petal length for flowers from the second plant. These averages differed ( $T=3.24$ ,  $d.f.=16$ , and  $P=0.005$ ) despite the plants’ shared environmental conditions. Floral guide size variation was greater between plants than for flowers on a single plant.

#### Naturalized urban flowers

The average UV floral guide in the naturalized plants covered 51% (standard error 1.23%) of the total petal surface area. However, there was a 27% range in this estimate, with a min–max of 39–66%. A strong correlation existed between floral head size and floral guide size ( $r=0.928$ ,  $n=25$ ,  $P<0.001$ ). This simply means that bigger flowers had more UV absorptive petal surface area than littler flowers, which says nothing about the relative size of floral guides. Perhaps more compelling is the result that arises when we address the relative amount of the petal surface that is comprised of floral guide. No correlation existed between the flower head size and the proportion of UV absorptive petal surface area ( $r=0.051$ ,  $n=25$ ,  $P=0.810$ ). In other words, larger flowers did not have relatively larger floral guides than smaller flowers.

#### Wild flowers

The average UV floral-guide in the wild plants covered 44% (standard error 1.69) of the total petal surface area. This was 7% smaller than the urban population average guide size. However, there was a 39% range in this estimate, with a min–max of 26–64%. This range was 11% more than the urban population range. Like the urban population, no correlation existed between the floral head size and the proportion of UV absorptive petal surface area ( $r=0.191$ ,  $n=21$ ,  $P=0.431$ ). Meaning once again, larger flowers did not have relatively larger floral guides than smaller flowers.

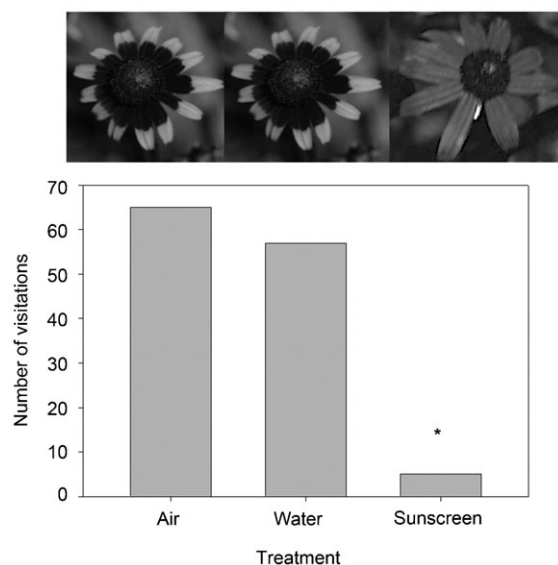


**Fig. 1. Spectrophotometric reflectance.** (A) Native *Rudbeckia hirta* as seen with color photography. (B) Native *R. hirta* as seen with ultraviolet (UV) photography. (C) Spectrophotometric reflectance plot of the proximal (...) and distal (—) portions of the *R. hirta* ligules, or petals. Relative reflection (%) is plotted as a function of wavelength (nm).

### III. Insect visitation response to floral guide manipulations

#### a. Sunscreen masked floral guides on *R. hirta*

Sunscreen obscured the floral guide typically found on *R. hirta* petals, and diminished UV reflection too (Fig. 2). The fine mist sunscreen dried quickly and left treated flowers dry. Fewer insects visited the sunscreen treatment than the other treatments. A total of 127 insects were observed visiting flowers in this experiment (five visited the ‘sunscreen’ treatment, 57 visited the



**Fig. 2. Results of a sunscreen experiment on the grounds of an urban campus.** The *R. hirta* floral head treatment UV photographs are displayed atop each data bar. Note the masking of the lack of bold UV absorbent floral guide in the sunscreen treatment. The asterisk represents a significant difference in the number of visitors to the ‘sunscreen’ treatment relative to the other treatments.

‘water misted’ treatment and 65 visited the ‘air sprayed’ treatment; Table 1). The Chi-square test was highly significant refuting the null hypothesis of an equal distribution of visitors among treatments ( $\chi^2_{(0.05,2)}=50.1$ ,  $P<0.001$ , two tail test). Post-hoc pairwise comparison tests indicated a difference between the number of visitors to the ‘sunscreen’ treatment when compared to the ‘water misted’ ( $\Delta\chi^2=27.85>3.8$ ,  $P<0.05$ ), and the ‘air sprayed’ treatment ( $\Delta\chi^2=20.80>3.8$ ,  $P<0.05$ ). The community of visitors was diverse and included the following 10 families: Scoliidae ( $n=45$ ), Apidae (28), Halictidae (18), Syrphidae (13), Megachilidae (13), Hesperidae (2), Sphecidae (3), Culicidae (1), Muscidae (2), Vespidae (1) and an unidentified fly (1). Since Scoliidae are generally considered minor pollinators, removing this family, as well as the single visitors that were not likely major pollinators (e.g. Culicidae), and repeating the test did not affect the outcome. Visitors to the ‘sunscreen’ treatment included four families: Syrphidae (1), Megachilidae (1), Muscidae (2) and Vespidae (1).

#### b. Augmented floral guides on *R. fulgida*

A total of 60 insects were observed visiting flowers in this experiment (23 visited the ‘enhanced’ (double ring) floral guide treatment, nine visited the ‘diminished’ (no guide) treatment, and 28 visited the ‘wild-type cut-and-paste control’; Table 2; Fig. 3). The Chi-square test was highly significant refuting the null hypothesis of an equal distribution of visitors among treatments ( $\chi^2_{(0.05,2)}=9.7$ ,  $P=0.008$ , two tail test). This result was not

**Table 1. Results from Experiment IIIa**

Treatment	Observed	Expected
Sunscreen	5	42.3
Water	57	42.3
Air	65	42.3

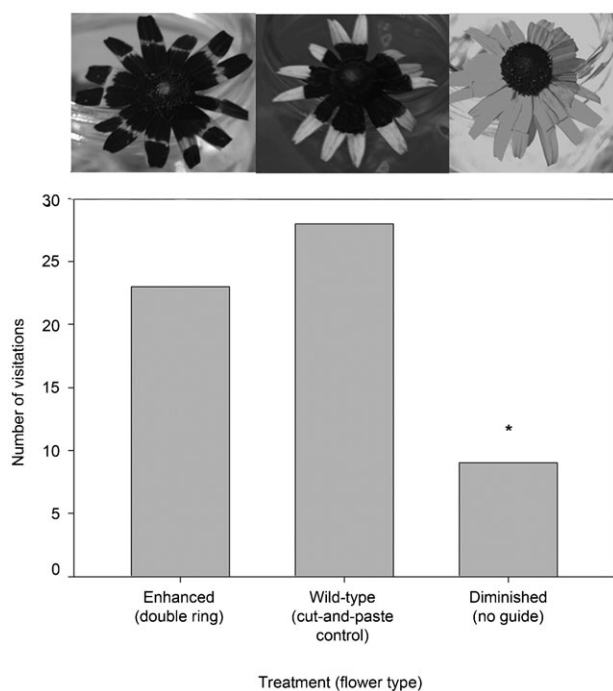
Sunscreen was used to eliminate the UV absorbent floral guide on *Rudbeckia hirta*. Misted water and sprayed air served as controls.

**Table 2. Results of Experiments IIIb,c**

Treatment	IIIb		IIIc	
	Observed	Expected	Observed	Expected
Enhanced guide	20	23	25	31
Diminished guide	20	9	25	13
Wild-type cut-and-paste control	20	28	25	26
Unmanipulated 'natural' flower	N/A	N/A	25	30

Petal 'cut-and-paste' was used to alter the UV absorbent floral guide size on *R. fulgida*.

attributable to a positional effect of the treatments (i.e. flower location in array,  $\chi^2_{(0.05,2)}=0.7$ ,  $P=0.705$ ). Post-hoc pairwise comparison tests indicated a difference between the number of visitors to the 'enhanced' treatment when compared to the 'diminished' treatment ( $\Delta\chi^2=5.6>3.8$ ,  $P<0.05$ ). Post-hoc results might appear counterintuitive, because the test is based on a squared deviation from the expectation. The squared deviation was smallest for the 'enhanced' treatment since it deviated least from the expected value, making the  $\Delta\chi^2$  largest when comparing this treatment to the 'diminished' treatment because it had the greatest deviation from the expected value. The frequency of visitors to the 'enhanced' treatment increased over time but decreased for the control (after Day 1). Frequency of visitors for Days 1 to 6 to the 'enhanced' treatment were: 0.250, 0.140, 0.160, 0.375, 0.500, 0.625, and to the 'control': 0.633, 0.714, 0.600, 0.375, 0.375, 0.250).



**Fig. 3. Results of an urban home garden experiment.** The *R. fulgida* floral head treatment UV photographs are displayed atop each data bar. Here, the 'enhanced' floral guide creates two concentric rings of UV absorbance, and the 'diminished (no guide)' treatment lacks the classic UV absorbing floral guide. The asterisks represent a significant difference in the number of visitors to the 'enhanced' treatment relative to the 'diminished' treatment.

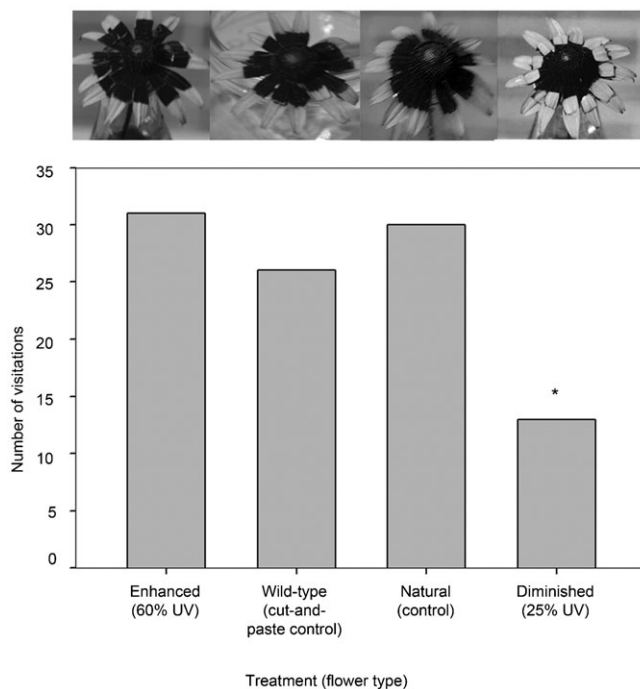
The community of visitors to this home garden experiment was less diverse than to the urban campus experiment. Here, Halictidae (sweat bees) comprised 75% (45/60) of the visitors and the number of their visits was similarly distributed across treatments each day. Substantially fewer individuals visited this garden from seven additional families: Papilionidae (3), Scoliididae (3), Apidae (2), Syrphidae (2), Megachilidae (2), Sphecidae (2), and Hesperidae (1).

### c. Slightly enlarged and diminished floral guides on *R. fulgida*

A total of 100 insects were observed visiting flowers in this experiment (31 visited the 'enhanced' (UV absorbent guide was ~60% of petal surface area) floral guide treatment, 13 visited the 'diminished' (~25%) floral guide, 26 visited the 'wild-type cut-and-paste control', and 30 visited the 'natural' flower; Table 2; Fig. 4). All insects were either major or minor pollinators and therefore included in the analysis. The Chi-square test was significant refuting the null hypothesis of an equal distribution of visitors among treatments ( $\chi^2_{(0.05,3)}=8.2$ , d.f.=3,  $P=0.041$ ). Post-hoc pairwise comparison tests indicated a difference in the number of visitors to the 'diminished' treatment and all other treatments ( $\Delta\chi^2=4.76>3.84$ ,  $P<0.05$ ;  $\Delta\chi^2=5.74>3.84$ ,  $P<0.05$ ;  $\Delta\chi^2=4.32>3.84$ ,  $P<0.05$ ). The community of visitors was diverse and included the following nine families: Apidae (33), Halictidae (33), Hesperidae (9), Lycaenidae (6), Nymphalidae (6), Syrphidae (6), Pieridae (2), Hesperidae (2), and Scoliididae (1).

### d. Oversized floral guides on *R. hirta*

In June, a total of 31 insects were observed visiting flowers in this experiment (20 visited the 'enhanced' (90%) floral guide, five



**Fig. 4. Results of an urban preserve experiment.** The *R. fulgida* floral head treatment UV photographs are displayed atop each data bar. Here the 'enhanced' floral guide treatment is slightly enlarged, with ~60% of the petal surface area UV absorbent and the 'diminished' floral guide treatment has ~25% of the petal surface area UV absorbent. The asterisk represents a significant difference in the number of visitors to the 'diminished (25% UV)' treatment relative to the other treatments.

**Table 3. Results of Experiment III**

Treatment	June 2011		Sept. 2011		Oct. 2011	
	Observed	Expected	Observed	Expected	Observed	Expected
Enhanced guide	20	10.3	18	10.6	13	7
Wild-type cut-and-paste control	5	10.3	9	10.6	3	7
Unmanipulated 'natural' flower	6	10.3	5	10.6	5	7

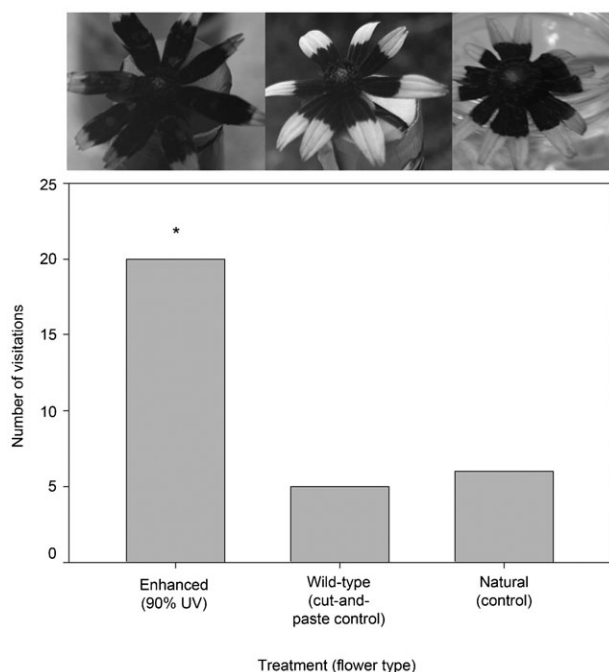
Petal 'cut-and-paste' was used to enlarge the UV absorbent floral guide size on *R. hirta*.

visited the 'wild-type cut-and-paste control', and six visited the 'natural' flower; Table 3; Fig. 5). The Chi-square test was highly significant refuting the null hypothesis of an equal distribution of visitors among treatments ( $\chi^2=13.6$ ,  $P=0.0011$ ). Post-hoc pairwise comparison tests indicated a difference in the number of visitors to the 'enhanced' treatment compared to the 'wild-type cut-and-paste control' and 'natural' flower ( $\Delta\chi^2=6.30>3.84$ ,  $P<0.05$  and  $7.24>3.84$ ,  $P<0.05$ ). The total visitation times were 472 seconds to the 'enhanced' treatment, 50 seconds to the 'wild-type cut-and-paste control' and 62 seconds to the 'natural' flower. Mean visitation times were 23.5 (s.d.=27.0), 10.0 (s.d.=9.59) and 10.3 (s.d.=9.11) seconds, respectively. ANOVA analysis indicated that mean visitation times were not significantly different (Sums of Squares (SS) between groups=8.8088, SS within groups=70.54, Total=78.6, d.f.=2, 27 and 29, respectively; Mean Square (MS) between groups=4.04, within group 2.61,  $F=1.548$  and  $P=0.231$ ). The community of visitors decreased in diversity and included two families: Halictidae (30), and Apidae (1).

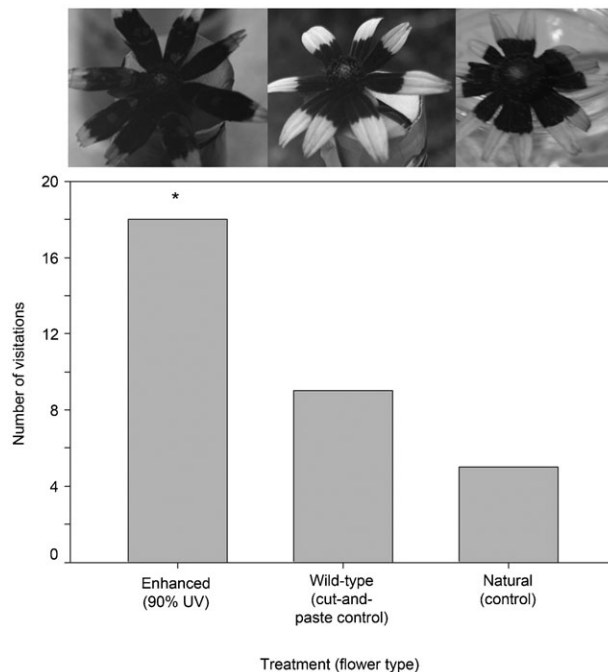
In September, a total of 32 insects were observed visiting flowers in this experiment (18 visited the 'enhanced' floral guide,

nine visited the 'wild-type cut-and-paste control' and five visited the 'natural' flower; Table 3; Fig. 6). The Chi-square test was again highly significant refuting the null hypothesis of an equal distribution of visitors among treatments ( $\chi^2=8.3$ ,  $P=0.016$ ). Post-hoc pairwise comparisons again indicated a difference in the number of visitors to the 'enhanced' treatment compared to the 'wild-type cut-and-paste control' ( $\Delta\chi^2=4.78>3.84$ ,  $P<0.05$ ), but not to the 'natural' flower ( $\Delta\chi^2=2.03<3.84$ ,  $P>0.05$ ). The total visitation times were 192 seconds to the 'enhanced' treatment, 75 seconds to the 'wild-type cut-and-paste control' and 71 seconds to the 'natural' flower. Mean visitation times were 10.6 (s.d.=13.6), 8.22 (s.d.=7.34), and 14.6 (s.d.=20.0) seconds, respectively. ANOVA analysis indicated that mean visitation times were not significantly different (SS between groups=0.081, SS within groups=4.93, Total=5.01, d.f.=2, 29 and 31, respectively. MS between groups=0.040, within group 0.170,  $F=0.237$  and  $P=0.791$ ). The community of visitors included three families: Halictidae (17), Apidae (12) and Syrphidae (3).

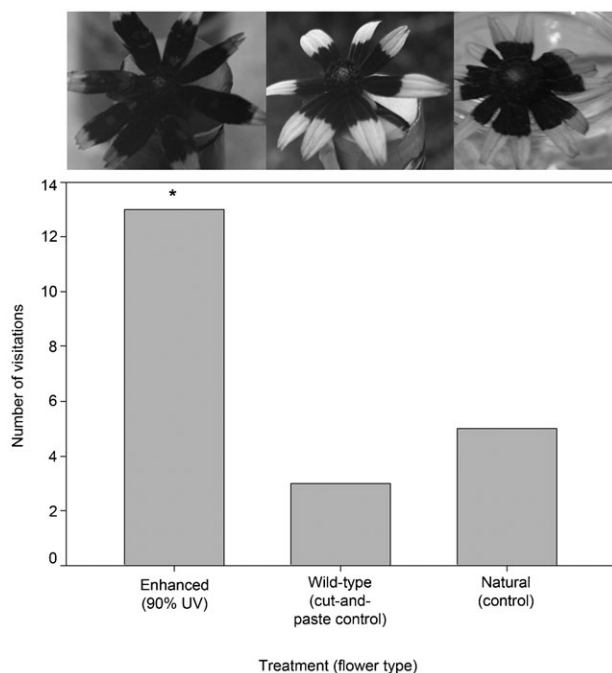
In October, a total of 21 visitors were observed visiting flowers in this experiment (13 visited the 'enhanced' floral guide, three



**Fig. 5. Results of an urban zoo grounds experiment.** The *R. hirta* floral head treatment UV photographs are displayed atop each data bar. Here the 'enhanced' floral guide treatment is very large, with ~90% of the petal surface area UV absorbent. The asterisk represents the significant difference in the number of visitors to the 'enhanced (90%)' treatment relative to the other treatments.



**Fig. 6. Results of an urban zoo grounds experiment.** The *R. hirta* floral head treatment UV photographs are displayed atop each data bar. Here the 'enhanced' floral guide treatment is very large, with ~90% of the petal surface area UV absorbent. The asterisk represents the significant difference in the number of visitors to the 'enhanced (90%)' treatment relative to the 'control' flower.



**Fig. 7. Results of an urban zoo grounds experiment.** The *R. hirta* floral head treatment UV photographs are displayed atop each data bar. Here the 'enhanced' floral guide treatment is very large, with ~90% of the petal surface area UV absorbent. The asterisk represents the significant difference in the number of visitors to the 'enhanced (90%)' treatment relative to the 'natural' flower.

visited the 'wild-type cut-and-paste control' and five visited the 'natural' flower; Table 3; Fig. 7). The Chi-square test was again highly significant refuting the null hypothesis of an equal distribution of visitors among treatments ( $\chi^2=8.0$ ,  $P=0.018$ ). Post-hoc pairwise comparison tests indicated a difference in the number of visitors to the 'enhanced' treatment compared to the 'natural' flower ( $\Delta\chi^2=4.57>3.84$ ,  $P<0.05$ ) but not to the 'wild-type cut-and-paste control' ( $\Delta\chi^2=2.86<3.84$ ,  $P<0.05$ ). The community of visitors increased slightly in diversity compared to the two prior experiments due to an influx of butterflies and included five families: Halictidae (15), Nymphalidae (2), Pieridae (2) Papilionidae (1), and Apidae (1). The one-day food color mark recapture study yielded no repeat visitors on Day 2 but captive insects retained their color.

## DISCUSSION

Despite the ubiquity of UV absorbent floral guides in Angiosperms, very little work has addressed the association between pollinator visitation frequency and UV floral guide traits, possibly because these traits were assumed to serve orientation purposes only. We show clearly that wild bees are more frequently attracted to *Rudbeckia* with oversized UV absorbing floral guides than to conspecific flowers with guide sizes closer to the average found on wild and cultivated flowers. Bees also demonstrate less recruitment to flowers with very small guides. We also found that sunscreen treated flowers with masked UV traits were avoided by pollinators, which is consistent with prior work of the same nature (Johnson and Andersson, 2002). However, we did not control for potential alternative chemosensory cues (e.g. sunscreen scent) so the sunscreen experiment must be interpreted with caution.

Our work shows that floral guides functioned in distance-based recruitment. Preference for relatively large floral guides held true across our experiments despite the fact that in nature, bigger flowers do not have relatively larger floral guides than smaller flowers. Daumer (Daumer, 1956; Daumer, 1958) and others (e.g. Burr et al., 1995) posited that UV reflectance attracts pollinators from a distance and that floral guides are used for close-range reward-orientation. While this is true, and could perhaps be particularly pertinent to floral species without radial symmetry, or with particular types of floral guides, we have also shown that in *Rudbeckia*, when we decrease the proportion of the floral head surface area that is UV reflective and increase the size of the UV absorbent guides, we see higher recruitment.

Chittka et al. have shown that pure UV reflecting flowers are very rare (Chittka et al., 1994). They comprised less than 4% of over 500 species surveyed. It is logical, based upon what we know about insect vision, that purely UV reflecting flowers would not be attractive to bees. Chittka et al. found that if flowers reflected UV they also tended to reflect red (Chittka et al., 1994), which may aid in recruitment of some insect pollinators. The reflectance curves for ~17% of their surveyed flowers, resemble our data for the proximal half of the *Rudbeckia* petal surface (e.g. human yellow and UV absorbent). Chittka et al. label this reflectance pattern 'green' for bees (Chittka et al., 1994). The reflectance curves for ~13% of their surveyed flowers resemble our data for the distal half of the *Rudbeckia* petal surface (e.g. human yellow and UV reflective). Chittka et al. label this pattern 'UV-green' for bees (Chittka et al., 1994). These reflectance spectra are predictably distinguishable from environmental background material including leaves, soil and rock, much of which may appear more gray to bees (Daumer, 1958; Kevan, 1978; Menzel and Shmida, 1993).

The visitation preferences that we identified are generalizable for at least three floral species (*R. hirta*, *R. fulgida*, and unpublished data for *Ranunculus bulbosus* L. Ranunculaceae). Bees are capable of learning fine color differences through trial and error (Reser et al., 2012); however, the minimal variation in 'human yellow' seen between the distal and proximal portions of the *Rudbeckia* petals are not predicted to have driven our results for untrained bees where we did not link this difference to reward or aversion resources. Bees generally take longer to discriminate between similar colors than dissimilar ones and the neural pathway invoked for learning fine differences demonstrates plasticity, unlike the pathway for coarse color discrimination, which functions more rapidly and is hard-wired (Dyer et al., 2011). The wild bees recruited in our work may have had prior experience in nature. So, we purposefully used multiple urban settings with unique surrounding vegetation, including different floral species at different experimental sites, so that prior learning would not bias our results. However, additional research, perhaps using 'cut-and-paste' style treatments with UV absorbent petals from another species, placed atop *Rudbeckia* flowers, might reinforce our results and eliminate the confounding issue associated with yellow coloration.

Since different types of illumination can affect bee vision (Arnold and Chittka, 2012) conducting experiments across a variety of lighting regimes may be important. However, brightness has generally been found to be an unimportant variable when chromatic differences provide for discrimination. Detecting brightness mandates training (Backhaus, 1991) and appears to matter most for small object identification (Spaethe et al., 2001).

From a behavioral perspective, flower color plays an important role in flower constancy. Flower constant pollinators visit only a subset of the species or morphs extant, despite the presence of alternative, untapped rewards that may be bypassed en route to the preferred resource (Waser, 1986). Flower color also plays an essential role when flower constant bees shift their preference to novel focal phenotypes (Chittka et al., 1997; Chittka et al., 1999). Whether UV floral guide size could contribute to flower constancy remains to be investigated.

In this work, we identified a normal distribution for floral guide size in wild and cultivated populations, though additional data would now be useful to determine whether cultivating plants disrupts the typical floral guide size found in wild plants. It is interesting to note that the mean floral guide size was smaller in the wild population than the cultivated one, and that this wild population had a larger variance in guide size. This comparison raises an important point: traits essential to pollinators can be altered when we cultivate plants, whether we can literally see these traits or not. To address whether, and how much, we alter UV floral guides when we cultivate flowers, more wild and cultivated populations should be surveyed for both UV guide size and for population-level variation in this trait.

Our work also prompts some fundamental questions like ‘Does co-evolution maintain UV vision in bees?’, ‘Does this result in natural selection for the maintenance of UV traits in flowers?’. Retention of UV opsins is ancient and widespread in insects (Briscoe and Chittka, 2001; Kevan et al., 2001), suggesting constraints on evolutionary changes in these genes. Since simple mutations in opsins can shift visual capability, the long-term persistence of UV vision suggests an adaptive value to this trait.

Global declines of commercial and wild bees raise serious pollination-service and economic concerns (Watanabe, 1994; Steffan-Dewenter et al., 2005; Biesmeijer et al., 2006; Brown and Paxton, 2009; Frankie et al., 2009; Gross, 2011; Bommarco et al., 2012; Cameron et al., 2011). This means that advancing our knowledge about the relative attractiveness of conspecific flowers to pollinators is relevant now more than ever. Urban landscapes and sprawl are increasing globally, which decreases the available natural habitat for native pollinations and often replaces it with small, landscaped parcels (Olujimi, 2009). Wild bees are rarely studied in this expanding modern habitat, making experimental work evaluating the pollination behavior of native bees in urban landscapes particularly timely. Recent work has shown that conscientious landscape management can increase bee diversity (Ahrné et al., 2001). Our work now shows that UV absorbent floral guides can play an important role in pollinator recruitment as well.

## MATERIALS AND METHODS

### I. Floral specimens and relevant techniques for measuring UV floral guides

#### Floral specimens

*R. hirta* and *R. fulgida* are Asters. They produce one, large floral head, that is comprised of many central black florets, commonly called true flowers. These tiny true flowers are surrounded by long ligulate florets that function like large petals. They are yellow to humans. Generally, the proximal half of these ‘petals’ is comprised of UV absorptive floral guide. In UV photography, this presents as a bold, dark ring around the center of the entire floral head, where the UV is absorbed. Early work refers to this UV pattern as a ‘nectar guide’ (Daumer, 1958; Eisner et al., 1969; Thompson et al., 1972). The distal portion of these petals is UV reflective and ‘human yellow’. The UV reflectance peaks at ~360 nm as can be seen on spectrophotometric reflectance graphs (Thompson et al., 1972; Schlangen et al., 2009).

### Photography and image analysis

UV photography was conducted using a Baader U-filter. This filter transmits UV wavelengths from 325 to 369 nm, with maximal penetration in the UV-A range at 360 nm (Savazzi, 2011). The filter was used with an AF Micro Nikkor 60 mm lens, several mounts, and a Nikon D70 DSLR digital camera. UV photographs were downloaded to Nikon Capture NX2 software and then transferred to Image J 1.46 for quantitative analyses. Image J is an open access processing and analysis program written in Java that can be downloaded from National Institutes of Health (<http://rsbweb.nih.gov/ij>). In Image J, a phenotypic trait, like petal surface area can be traced and digitized. The circumscribed area can be calculated and the resultant pixel measurement for the trait can be converted to a meaningful scale (e.g. mm<sup>2</sup>) based upon a reference scale used in the picture (here, a ruler). Multiple traits were measured including petal length, petal surface area, floral-head area, UV floral guide length, and proportion of petal surface area comprised of UV floral guide. All statistical tests were performed in SPSS.

### II. Floral-guide size-distributions for three data sets: greenhouse, urban and wild *R. hirta*

#### Greenhouse flowers

In November 2010, native *R. hirta* seeds were obtained from the Ornamental Plant Germplasm Center of The Ohio State University. In March 2011, 236 seeds were planted under full-spectrum 12L: 12D lighting in the Kaplan Orchid Conservatory of Old Dominion University (ODU, Norfolk, VA, USA, 36.885441N, -76.307466W). After seven weeks, plants were transferred the ODU greenhouse until flowering. In May 2011, flowers were used in a pilot study assessing UV-absorptive floral-guide length (mm) variation. Since plants were grown in the same conditions, variation may represent the genetic component of phenotypic diversity. Floral guides were measured for 18 flowers and two plants and these measurements were compared with an independent samples t-test. The t-test assumption regarding equal variance was met (Levene’s test  $F=0.00$ ,  $P=1.0$ ).

#### Naturalized urban flowers

In July 2010, we assessed the average sized UV absorptive floral guide by measuring the proportion of petal surface area that was comprised of floral guide (mm<sup>2</sup>) relative to the entire petal surface area, for each flower. We also evaluated the distribution of these guide sizes for this naturalized *R. hirta* population. Measurements were made for 30 flowers using Image J on photographs of all flowers. Flowers measured were located in a large, conspecific population on the ODU urban campus.

To evaluate the shape of the distribution for UV floral guide sizes, we used Shapiro-Wilks’ test. Consistent with many quantitative traits, the Shapiro-Wilks’ test indicated that UV guide sizes were normally distributed in these naturalized flowers ( $W=0.984$ , d.f.=27,  $P=0.938$ ).

Next, we used Pearson’s correlation coefficient to determine whether there was a relationship between the floral head width (in mm), and the total size of the floral guide (area in mm<sup>2</sup>), because this allows us to address questions like ‘Is UV guide size correlated with overall flower size?’. Since a significant result could merely represent a positive correlation between the total floral head size and amount of petal surface area that was floral guide, we next considered whether there was a correlation between the total floral head size and the proportion of the total petal surface area that was comprised of floral guide. This allows us to address whether larger floral heads have relatively larger guides (or greater percentage of petal surface area covered by UV guide) than smaller floral heads.

#### Wild flowers

In July 2011, we assessed the floral guide size distribution in the wild flower population for comparison to the naturalized flowers. Thirty flowers growing on a precipitous rocky outcrop in montane Colorado (39.44419N, 105.74084W) were evaluated in the same manner as the naturalized urban flowers. Based upon the Shapiro-Wilks’ test, the shape of the distribution of UV floral guide sizes for this population, like the urban one, was also normally distributed ( $W=0.958$ , d.f.=21,  $P=0.467$ ).



### III. Insect visitation response to floral guide manipulations

In the following series of experiments, floral guide size was manipulated in one of two species (*R. hirta* and *R. fulgida*) to determine whether insect visitation rates differed when floral guides were removed, enhanced and diminished. No flowers were reused in any trials.

#### a. Sunscreen masked floral guides on *R. hirta*

In July and August 2010, we conducted a sunscreen study with *R. hirta* from the urban population. Three treatments were used, two new flowers per treatment, for 15 trials. Thus, six floral heads located in the garden were haphazardly identified for use for each trial after visual matching for size and stage. Haphazard selection should randomize error variance. For treatment one, two floral heads were ‘sunscreen’ misted (Ocean Potion Suncare Instant Dry SPF 70 Mist) to mask the floral guide entirely. Treatment two was ‘water misted’ as a control for moisture (the true sunscreen control of sunscreen minus the active ingredient was unavailable). Treatment three was ‘air-sprayed’ as control for potential loss of pollen during water misting.

This study site was intentionally selected to assay the community of insects naturally attracted to cultivars in a large, landscaped urban garden. The garden was situated in front of a five-story building and comprised of a large tract of *R. hirta* (12 m×1.5 m), a smaller tract of *Echinacea purpurea* (5'×2') behind the *R. hirta*, and a few *Hosta* cultivars on the garden edges.

All trials were completed on hot summer full-sun days a few days apart from one another. At ~10:00 am, one hour trials were initiated and the total number of insect visitations was recorded. To be counted, the insect had to alight on a flower head then demonstrate some visual evidence of foraging or pollen collection. First landings were the sole landings counted for the few insects that were observed landing twice so as to avoid repeated measures. Insects were identified for this and subsequent experiments based upon our own expertise and guidebooks (e.g. Arnett and Jacques, 1981; Evans, 2007; Michener et al., 1994; Michener, 2007). Visitation data were analyzed with a Chi-square test and a post-hoc pairwise comparison test that assessed homogeneity of proportions (Cox and Key, 1993). The post-hoc test used the absolute value of the difference between two cell contributions in paired tests. The difference is distributed as Chi square with one degree of freedom so the  $\Delta$  Chi square between a pair of treatments was compared to 3.84 to assess statistical significance at the  $P=0.05$  level.

#### b. Augmented floral guides on *R. fulgida*

In August 2010, the first of a series of ‘cut-and-paste’ experiments was conducted in a diverse urban home garden (~6 m×6 m) in Norfolk, VA. This garden was more complex than the above one, comprised of scores of cultivated *R. fulgida* plants surrounded by a variety of additional flowering perennial species. Treatment *R. fulgida* (Goldsturm) cultivars from a chain store were used in these experiments to evaluate insect response to experimentally diminished and enhanced floral guides.

To create the ‘cut-and-paste’ treatments ~10 individual floral heads were collected for each trial. All petals were removed from heads, then cut sagittally at the point where the UV floral guide ended (about half way down the petal). UV photographs were taken to determine whether subsequent trimming was necessary to ensure that pieces were entirely floral guide or completely devoid of floral guide. The appropriate petal pieces were glued onto fresh floral heads contingent upon treatment.

Two ‘cut-and-paste’ treatments and one control were freshly constructed for each trial. Treatment one had a ‘diminished’, or nearly no, floral guide and was created by gluing the petal pieces devoid of floral guide atop the proximal portion of an unmanipulated flower head. Treatment two had an ‘enhanced’ or double floral guide, where an additional concentric ring of floral guide was added distally to the natural guide by pasting small floral guide petal pieces on another unmanipulated flower head. For the ‘wild-type cut-and-paste control’ flower, two sets of petal pieces, one with floral guide, and one devoid of it, were glued atop the petals of a final unmanipulated floral head, recreating the common wild-type pattern.

Each floral head (with stalk ~20 mm) was placed into a clear glass bottle on the ground ~3 meters away from the conspecific flowerbed.

Six trials were conducted from about noon until 2:00 pm for six days and each ended after 10 insect visitations. Treatment locations were rotated in the bottle line-up in subsequent trials (treatment on the left in trial 1 was positioned in the middle for trial 2, and on the right for trial 3, and so forth) such that each trial was comprised of a novel treatment series. Trials were conducted from August 21–27 in Norfolk, VA (36.886246N, –76.289351W). The time, treatment visited, weather, and visitor type were recorded. Visitation data were analyzed with a Chi-square test and a post-hoc pairwise comparison test, as above.

#### c. Slightly enlarged and diminished floral guides on *R. fulgida*

In a less cultivated, more natural setting at an urban sanctuary, a similar ‘cut-and-paste’ experiment was completed. *R. fulgida* were again altered to create two treatments and one ‘wild-type cut-and-paste’ control. Treatment one was ‘enhanced’, here to be slightly enlarged from the natural floral guide to ~60% of the petal surface area. Treatment two was ‘diminished’, here to be ~25% of the petal surface. Treatment three was the same ‘wild-type cut-and-paste control’ used previously. Additionally, a ‘natural’ unmanipulated flower head was added as a true control for our ‘cut-and-paste’ procedure. Treatment bottles were positioned near the *Rudbeckia* plot of the ‘meadow flower bed’ at Weyanoke Bird and Wildflower Sanctuary in Norfolk, VA (36.87373N, –76.307068W). This floral bed contained scores of *Rudbeckia*, along with a number of additional annual and perennial meadow flowers, some naturalized. The experiment was conducted from mid-September until mid-October in 2010 from ~noon until 2:00 pm. Data were recorded and statistics performed as above.

#### d. Oversized floral guides on *R. hirta*

At this point we knew small floral guides were least desirable and we wanted to focus on whether large, uninterrupted guides were preferred. As well, to determine if our results held for more than one species, the native *R. hirta* seed grown in the greenhouse were used for this study. This work was conducted on a grassy knoll, further away (~25 m) from a smaller, mixed species floral garden at an urban zoo. This site allowed us to determine whether there were differences in the families recruited to this more monoculture-like urban habitat. Treatment one was another enhanced floral guide, here quite oversized, and comprising ~90% of the petal surface. Treatment two was the ‘wild-type cut-and-paste control’ from above and treatment three was again a natural flower. Nine consecutive trials ran in full sun from ~10:00 am to 2:00 pm per day in June of 2011. In addition to the Chi-square test and post-hoc tests described above here the cumulative (or total) and mean visitation times were also recorded for all treatments. The variance in mean visitation times was large though homogenous (Levene’s statistic=0.056, d.f.<sub>1</sub>=2, d.f.<sub>2</sub>=27,  $P=0.946$ ) and these data were square root transformed prior to ANOVA.

This ‘oversized’ treatment experiment was repeated in September of 2011 for seven days. Here, insects were captured with mesh nets and removed after landing to ensure no repeat measures were included in the data set. The variance in mean visitation times was large though homogenous (Levene’s statistic=0.009, d.f.<sub>1</sub>=2, d.f.<sub>2</sub>=29,  $P=0.992$ ) and these data were  $\log_{10}$  transformed prior to ANOVA. Before we started this experiment, we conducted a food-color mark and recapture pilot study on Halictidae ( $n=20$ ) collected during a one-day trial. No marked visitors were identified the subsequent day but food-color marks remained on bees held for 24 hrs.

This ‘oversized’ treatment experiment was repeated again in October of 2011, but here the design was a linear array. Thirty *R. hirta* flower heads (three treatments ×10 flowers per treatment) were positioned ~60 mm apart from one another in one long array, where treatment order repeated sequentially, and visitation monitored for a longer, single day time period. All of these experiments were conducted at the Virginia Zoo (36.8794675N, –76.274154W) in Norfolk, VA.

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### Competing interests

The authors have no competing interests to declare.

### Author contributions

L.H. and R.B. conceived and designed the experiments, L.C. performed the experiments, L.H. analyzed the data and L.C., L.H., and R.B. wrote the manuscript.

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