Journal of Mind and Medical Sciences

Volume 5 | Issue 1

Article 15

2018

Melatonin attenuates oxidative stress and modulates inflammatory response after experimental burn trauma

Minka Hristova Medical University of Varna, Department of Physiology and Pathophysiology, hristova_minka@abv.bg

Ganka Bekyarova Medical University of Varna, Department of Physiology and Pathophysiology, ganka.bekyarova@gmail.com

Milena Atanasova Medical University of Pleven, Department of Biology, m.atanasova@mu-pleven.bg

Maria Tzaneva Medical University of Varna, Department of General and Clinical Pathology, maria.tzaneva@mu-varna.bg

Follow this and additional works at: https://scholar.valpo.edu/jmms Part of the <u>Disease Modeling Commons</u>, and the <u>Medical Physiology Commons</u>

Recommended Citation

Hristova, Minka; Bekyarova, Ganka; Atanasova, Milena; and Tzaneva, Maria (2018) "Melatonin attenuates oxidative stress and modulates inflammatory response after experimental burn trauma," *Journal of Mind and Medical Sciences*: Vol. 5 : Iss. 1, Article 15. DOI: 10.22543/7674.51.P93100

Available at: https://scholar.valpo.edu/jmms/vol5/iss1/15

This Research Article is brought to you for free and open access by ValpoScholar. It has been accepted for inclusion in Journal of Mind and Medical Sciences by an authorized administrator of ValpoScholar. For more information, please contact a ValpoScholar staff member at scholar@valpo.edu.

Research Article



Melatonin attenuates oxidative stress and modulates inflammatory response after experimental burn trauma

Minka Hristova^{1*}, Ganka Bekyarova¹, Milena Atanasova², Maria Tzaneva³

¹Medical University of Varna, Department of Physiology and Pathophysiology ²Medical University of Pleven, Department of Biology ³Medical University of Varna, Department of General and Clinical Pathology

Abstract

Introduction. Thermal injury activates an inflammatory response. Melatonin possesses antioxidant and anti-inflammatory properties. The objective of the present work was to study melatonin effects on the inflammatory response under conditions of oxidative stress during the early stage of thermal injury.

Materials and methods. We used 24 white male rats of Wistar breed, randomly divided into three experimental groups. Group one was the control, group two was inflicted with burn trauma, and group three was inflicted with burn trauma, with melatonin application following the thermal injury. Melatonin was applied twice in doses of 10 g/kg b.m. immediately after the burn trauma and again at 12 hours. Plasma levels of tumor necrosis-factor- α (TNF- α), a pro-inflammatory mediator, and of interleukin-10 (II-10), an anti-inflammatory mediator, were examined and their ratio was calculated. The levels of malondialdehyde (MDA), an oxidative stress marker, were also estimated.

Results. Thermal trauma significantly increased plasma TNF- α levels (δ <0.01) and TNF- α /IL-10 ratio but did not change IL-10 ones. Plasma MDA concentrations were significantly elevated as well (δ <0.0001). Melatonin application significantly reduced TNF- α (δ <0.05), increased IL-10 (δ <0.05), down-regulated TNF- α /IL-10 ratio and changed MDA concentrations (δ <0.01).

In conclusion, our results show that local alteration induces oxidative stress and inflammatory response with TNF- α /IL-10 disbalance. Melatonin modulates this response and attenuates oxidative stress in experimental burn injury.

Keywords : melatonin, inflammatory response, TNF-α, IL-10, burn
Highlights · Thermal trauma significantly increased plasma TNF-α levels and TNF-α /IL-10 ratio but did not change IL-10.
· Melatonin attenuates oxidative stress and changes the disbalance between the pro- and anti-inflammatory mediators in favor of the anti-inflammatory ones.
To cite this article: Hristova M, Bekyarova G, Atanasova M, Tzaneva M. Melatonin attenuates oxidative stress and modulates inflammatory response after experimental burn trauma. J Mind Med Sci. 2018; 5(1): 93-100. DOI: 10.22543/7674.51.P93100

*Corresponding author: Minka Hristova Aleksandrova, Medical University of Varna, Department of Physiology and Pathophysiology, 55 Marin Drinov St., 9002 Varna, Bulgaria e-mail: <u>hristova minka@abv.bg</u>

Introduction

Severe thermal trauma can lead to the development of systemic inflammatory response syndrome (SIRS) and sepsis. Generalization of the pathophysiological manifestations causes increased morbidity, poly-organ insufficiency, and mortality among burned patients (1). Locally, in a burn-induced wound, numerous cells such as neutrophils and macrophages are activated (2, 3). Their count in systemic circulation also increases. (4). These are the source of cytokines with pro-inflammatory action, activate the inflammatory cascade and, as along with them, synthesize and liberate cytokines with thev antiinflammatory action. The cellular response also underlies the generation of free radicals (5) which induce lipid peroxidation, cell membrane damage, and apoptosis (6, 7). The induced postburn oxidative and nitrosative remote organ damage disturbs immune system balance (8), contributes to immunosuppression development, and enhances the risk for the development of systemic inflammatory response syndrome (SIRS) and sepsis (9, 10).

Tumor necrosis factor- α (TNF- α) is a cytokine presenting with a variety of biological effects (11) and acting as a central inflammation mediator in sepsis, trauma, and burn (12-14). TNF- α induces gene expression of a series of pro-inflammatory cytokines and is capable of selfinduction (11, 15).

Interleukin-10 (IL-10) has initially been described as an inhibitory factor for the synthesis of cytokines (TNF- α , IL-1, IL-6), chemokine, and adhesion molecules in the monocytes/macrophages and neutrophils (16-18). TNF- α reduction is considered the most important suppressive role of IL-10 (19). Data regarding how the elevated concentration of the pro-inflammatory mediators of thermal trauma prevail are presented in the recent literature, but there are relatively few investigations of the anti-inflammatory mediators.

Melatonin (N-acetyl-5-methoxytryptamine) is mainly a secretory product of the pineal gland. Its functions in the organism relate to numerous physiological and pathological processes. Melatonin exerts direct antioxidant effects via free radical scavenging and indirectly stimulates the activities of antioxidant enzymes, superoxide dismutase (SOD), glutathione peroxidase (GPx), glutathione reductase (GRd), and catalase (20). Melatonin may exert an antiinflammatory effect as well, by restricting the action of the free radicals and inhibiting the nuclear factor kappa-lightchain-enhancer of activated B cells (NF-kB) and related target genes, which participate in immunity and inflammation (21). Melatonin application has been found to reduce the manifestation of the systemic inflammatory response in experimental and clinical investigations (22-24). However, there are few studies regarding the melatonin effect on IL-10 levels under the conditions of experimental thermal trauma (25, 26).

Herewith we hypothesize that melatonin modifies cytokine secretion and modulates the systemic inflammatory response after burn trauma. To test this hypothesis, we examined melatonin effects on plasma levels of TNF- α , a pro-inflammatory mediator, and of Il-10, an anti-inflammatory mediator, during the early stage of thermal trauma.

Materials and Methods

Experimental design

The experimental procedure was approved by the Home Office for Care and Use of Laboratory Animals, and experiments were performed in accordance with the European Communities Council Directives 86/609/EEC. Age-matched male rats weighing between 220 and 250g fasting for 12 h were allowed free access to water before injury.

Animals were housed in individual wire-bottomed cages at 20° C and offered rat chow and water ad libitum. They were kept in dark/light cycles (12:12 h; lights on at 8:00 am) to ensure a satisfactory photoperiod. After light ether inhalation, general anesthesia was intraperitoneally performed using thiopental (30 mg/kg). In order to accomplish 30% of a third-degree burn, boiling water (98° C) was applied on the back of the animals for 10 sec. For those rats subjected to burn injury, 4 mL of physiological saline was intraperitoneally applied for immediate resuscitation after the trauma. No animals died within the first 24 h of the post-burn period. Twenty-four male rats were randomly assigned to three groups of 8 animals each: control, non-burned and non-treated (C), vehicle-treated burned group (B), and melatonin treated burned group (B+M).

Melatonin treatment

Melatonin (N-acetyl-5-methoxytryptamine, Merck, Germany) in a dose of 10 mg/kg body weight (b.w.) dissolved in vehicle, or vehicle alone (2% ethyl alcohol diluted in physiological saline in a dose 5 ml/kg) was administered to the appropriate group. Melatonin and vehicle were applied i.p. twice, immediately after burns in the morning between 8:00 a.m. and 9:00 a.m. and 12 hours after burn injury. All animals were given buprenorphine (0.3 mg /kg i.p. b.w.) twice daily for pain control post burn.

Animals from the all groups were anesthetized with thiopental and euthanized 24 h after burns.

Biochemical analysis

Blood was taken from the jugular vein and heparinized. Plasma was separated by centrifugation at 800 x g rpm for 10 min and aliquots were stored at -80oC until analysis. Plasma lipid peroxidation was assayed by MDA levels detected by thiobarbituric acid (TBA) reactivity as described by Porter et al. (27). Results were expressed as nmol MDA/mL plasma, using the extinction coefficient of MDA-TBA complex at 532 nm = $1.56 \times 10-5 \text{ cm}-1 \text{ M}-1$ solution.

Determination of plasma cytokine levels

Plasma levels of TNF α and IL-10 were determined by enzyme-linked immunosorbent assay (ELISA) using Gen-Probe Diaclone SAS kits (Besancon Cedex, France). Results were reported as pg/mL.

Statistical analysis

Statistical analyses utilized Graphpad Prism version 6.0. The results are shown as mean \pm SEM and box plots. Significance was determined by unpaired Student's t test or the nonparametric Mann-Whitney-U-test. A P-value less than 0.05 two-tailed was considered significant.

Results

Examination of MDA in thermal trauma and melatonin effect

Plasma MDA levels were significantly increased by 39% (p<0.0001) in the burned rats compared to the control group (Fig. 1). Melatonin treatment significantly inhibited the elevation in plasma MDA level (p<0.01) and restored control values.

Examination of TNF-\alpha in thermal trauma and melatonin effect

TNF- α levels increased significantly by 115% (p<0.01) in plasma of burned rats compared with controls (Fig. 2). Plasma TNF- α concentration decreased following melatonin treatment by 41% (p<0.05) in the burned rats but was 74% higher (p<0.01) relative to control rats.

Examination of IL-10 in thermal trauma and melatonin effect

Plasma IL-10 level did not change significantly in burned rats when compared to controls (Figure 3). Melatonin significantly elevated this level by 50% (p<0.05) in burned rats and was higher than that of the control rats.

Examination of TNF- α /IL-10 ratio in thermal trauma and melatonin effect

This ratio was higher by 114% (p<0.05) in the experimental group than the control group (Fig. 4). Melatonin treatment reduced this ratio by 37% (p<0.05), tending to restore values comparable to those of the control group.

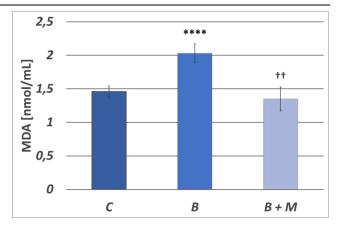


Figure 1. Effect of melatonin on MDA levels in plasma after burns. (C) control group; (B) burned, non - treated group; (B+M) burned, treated with melatonin group. Results are given as the mean \pm SEM. *****p<0.0001 vs. control group; ^{††}p<0.01 vs. burned, non - treated group.

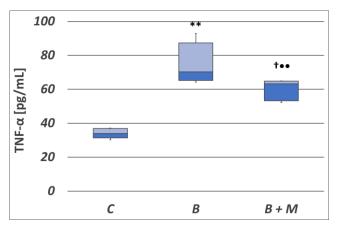


Figure 2. Effect of melatonin on TNF- α levels in plasma after burns. (C) control group; (B) burned, non - treated group; (B+M) burned, treated with melatonin group. Results are given as box plot, with median, 25th-and 75th-percentile values, min and max values. **p<0.01 vs. control group; †p<0.05 vs. burned, non - treated group; ••p <0.01 vs. control group.

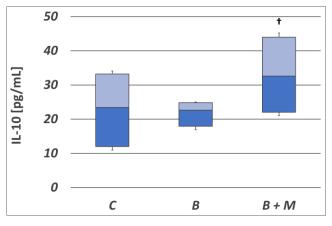


Figure 3. Effect of melatonin on IL-10 levels in plasma after burns. (C) control group; (B) burned, non - treated group; (B+M) burned, treated with melatonin group. Results are given as box plot, with median, 25th- and 75th-percentile values, min and max values. $\dagger p < 0.05$ vs. burned, non - treated group.

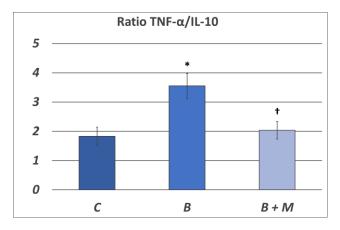


Figure 4. Effect of melatonin on TNF- α /IL-10 ratio in plasma after burns. (C) control group; (B) burned, non - treated group; (B+M) burned, treated with melatonin group. Results are given as the mean \pm SEM. *p<0.05 vs. control group; †p<0.05 vs. burned, non - treated group.

Discussions

Thermal injury induces local tissue damage. The activated pro-inflammatory cells such as neutrophils and macrophages synthesize and liberate large amounts of chemokines, cytokines, adhesion molecules, and alarmines which when entering the circulation can cause a systemic inflammatory response (28). The enhanced production of pro-inflammatory cytokines, lipid peroxides, and acutephase proteins, along with activation of the polymorphonuclear cells plays an important role in the systemic inflammatory response induced by thermal trauma (29, 30). Results from the present study demonstrate elevated plasma TNF-a concentrations after burn injury, providing evidence for the role of this mediator in the early local and systemic inflammatory response.

TNF-α represents not only an inflammation-inducing mediator but also an important factor in the course of the inflammation (31). It induces the expression of other inflammatory mediators such as IL-1a, IL-1β, IL-6 and IL-8, which form a cytokine network, adhesion molecules, acute phase proteins (32) and II-10 as well (33). TNF- α augments ROS production by the pro-inflammatory cells. ROS (reactive oxygen species) activate lipid peroxidation and cause cell membrane damage in thermal trauma (6, 34). Both TNF- α and ROS enhance the expression of NF-kB, a transcription factor responsible for the production of other pro-inflammatory mediators with cytotoxic action. A correlation exists between MDA and TNF-a in liver during burns (35) as well as between plasma TNF- α concentration and the degree of thermal damage (36). Similar data have been obtained in clinical and experimental investigations of thermal injury (37, 38). But there are contradictory data about the changes of tissue and plasma TNF- α levels. Some investigators have failed to establish alterations in burned animals compared with not-burned ones, most probably due to the experimental design and the duration of the examination period (39).

Our results demonstrate that the level of the proinflammatory mediator significantly increases while that of the anti-inflammatory mediator shows no change. There are contradictory data about IL-10 concentrations in burn injury. While some authors report results similar to ours (40), others report elevated concentrations of two antiinflammatory cytokines, IL-2 and IL-10, reaching their peak values during the initial hours after thermal trauma (41). The primary role of IL-10 is the suppression of the production of the pro-inflammatory mediators and the regulation of the inflammatory response (42). It seems possible that IL-10 accomplished its effects though the inhibition of the nuclear factor NF-kB (43) and through pathways that do not depend on NF-kB, such as activation of nuclear factor (erythroidderived 2)-like 2 (Nrf2) and heme oxygenase (HO-1) (44).

The examination of the balance between pro- and antiinflammatory cytokines serves as a predictive marker in clinical practice (45). The mortality of the patients with sepsis is related to elevated TNF- α and IL-10 concentrations (46, 47). The IL-10: TNF- α ratio is low in surviving patients (47), so it has been assumed that TNF- α /IL-10 disbalance represents one of the triggering factors for the development of SIRS and polyorgan failure in the initial stage after burn injury (48). Further research is needed to clarify these alterations during the various stages of the severe thermal trauma. It is known that antioxidant application inhibits the systemic inflammatory changes in thermal injury (49) and other diseases in which pathogenesis of systemic inflammation is involved (50).

Our results demonstrate a disbalance between the proand anti-inflammatory cytokines under the conditions of oxidative stress induced by burn trauma. Melatonin application results in the establishment of a tendency towards restoration of balance more similar to baseline levels in control rats. The data also indicate that melatonin reduces plasma TNF- α level and enhances plasma IL-10, a finding consistent with other authors who have also established that anti-inflammatory melatonin action is related to reduced plasma TNF- α concentration and increased plasma IL-10 (51, 52). This effect is most probably due to melatonin's inhibitory effect on NF-kB (53), also evident from thermal trauma (26). As such, melatonin improves the pro-/anti-inflammatory balance and restricts the manifestations of the systemic inflammation.

In the present study, plasma MDA levels were significantly increased, thus demonstrating severe lipid peroxidation following considerable burn injury, a finding consistent with other studies (54, 55). The pathophysiological alterations are most likely a consequence of ischemia/reperfusion injury and polymorphonuclear cell activation both locally and in the systemic circulation 3. resulting in free-radical overgeneration (56, 4). On the other hand, the depletion of plasma glutathione and antioxidant enzymes has been shown as a cause for the manifestations of the systemic oxidative response and the aggravation of the pathological processes in thermal trauma (57). Melatonin administration significantly decreases plasma 4. MDA levels, a consequence of the antioxidant and freeradical-scavenging capacities of melatonin and its metabolites (20).

Conclusions

Melatonin attenuates oxidative stress and changes the disbalance between the pro- and anti-inflammatory mediators in favor of the anti-inflammatory ones. Therefore, melatonin, by restricting the lipid peroxidation and by modulating the inflammatory response, can counteract the systemic inflammation and the subsequent development of sepsis and polyorganic insufficiency. These results confirm the broad therapeutic potential of melatonin and substantiate its possible application for the treatment of critical pathological conditions of the organism.

Acronyms and abbreviations

TNF-α: Tumor necrosis-factor-α II-10: Interleukin-10 MDA: Malondialdehyde SIRS: Systemic inflammatory response syndrome SOD: Superoxide dismutase GPx: Glutathione peroxidase GRd: Glutathione reductase NF-kB: Nuclear factor kappa-light-chain-enhancer of activated B cells Nrf2: Nuclear factor (erythroid-derived 2)-like 2 HO-1: Heme oxygenase

References

- Wolf SE, Rose JK, Desai MH, Mileski JP, Barrow RE, Herndon DN. Mortality determinants in massive pediatric burns. An analysis of 103 children with > or = 80% TBSA burns (> or = 70% full-thickness). *Ann Surg.* 1997; 225(5): 554–69. PMID: 9193183
- Foubert P, Gonzalez AD, Teodosescu S, Berard F, Doyle-Eisele M, Yekkala K, Tenenhaus M, Fraser JK. Adipose-Derived Regenerative Cell Therapy for Burn Wound Healing: A Comparison of Two Delivery Methods. Adv Wound Care (New Rochelle). 2016; 5(7): 288-98. PMID: 27366590, DOI: 10.1089/wound.2015.0672

- Sun LT, Friedrich E, Heuslein JL, Pferdehirt RE, Dangelo NM, Natesan S, Christy RJ, Washburn NR. Reduction of burn progression with topical delivery of (antitumor necrosis factor-α)-hyaluronic acid conjugates. *Wound Repair Regen*. 2012; 20(4): 563-72. PMID: 22712482, DOI: 10.1111/j.1524-475X.2012.00813.x
- 4. Dinsdale RJ, Devi A, Hampson P, Wearn CM, Bamford AL, Hazeldine J, Bishop J, Ahmed S, Watson C, Lord JM Moiemen N, Harrison P. Changes in novel haematological parameters following thermal injury: A prospective observational cohort study. *Sci Rep.* 2017; 7(1): 3211. PMID: 28607467, DOI: 10.1038/s41598-017-03222-w
- Wiggins-Dohlvik K, Han MS, Stagg HW, Alluri H, Shaji CA, Oakley RP, Davis ML, Tharakan B. Melatonin inhibits thermal injury-induced hyperpermeability in microvascular endothelial cells. J Trauma Acute Care Surg. 2014; 77(6): 899-905. PMID: 25051382,

DOI: 10.1097/TA.00000000000346

- Al-Roujayee AS. Naringenin improves the healing process of thermally-induced skin damage in rats. *J Int Med Res.* 2017; 45(2): 570-82. PMID: 28415935, DOI: 10.1177/0300060517692483
- Sehirli O, Sener E, Sener G, Cetinel S, Erzik C, Yeğen BC. Ghrelin improves burn-induced multiple organ injury by depressing neutrophil infiltration and the release of pro-inflammatory cytokines. *Peptides*. 2008; 29(7): 1231-40. PMID: 18395937, DOI: 10.1016/j.peptides.2008.02.012
- Avlan D, Taşkinlar H, Tamer L, Camdeviren H, Ozturhan H, Oztürk C, Aksöyek S. Protective effect of trapidil against oxidative organ damage in burn injury. *Burns.* 2005; 31(7): 859-65. PMID: 15963644, DOI: 10.1016/j.burns.2005.04.013
- Schwacha MG, Chaudry IH. The cellular basis of postburn immunosuppression: macrophages and mediators. *Int J Mol Med.* 2002; 10(3): 239–43. PMID: 12165794
- O'Dea KP, Porter JR, Tirlapur N, Katbeh U, Singh S, Handy JM, Takata M. Circulating Microvesicles Are Elevated Acutely following Major Burns Injury and Associated with Clinical Severity. *PLoS One.* 2016; 11(12): e0167801. PMID: 27936199, DOI: 10.1371/journal.pone.0167801
- 11. Ellerin T, Rubin RH, Weinblatt ME. Infections and anti-tumor necrosis factor alpha therapy. *Arthritis Rheum.* 2003; 48(11): 3013-22. PMID: 14613261, DOI: 10.1002/art.11301
- Klein D, Einspanier R, Bolder U, Jeschke MG. Differences in the hepatic signal transcription pathway and cytokine expression between thermal injury and sepsis. *Shock.* 2003; 20(6): 536-43. PMID: 14625478, DOI: 10.1097/01.shk.0000093345.68755.98

- 13. Kothari N, Bogra J, Abbas H, Kohli M, Malik A, 23. Lowes DA, Webster NR, Murphy MP, Galley HF. Kothari D, Srivastava S, Singh PK. Tumor Necrosis Factor gene polymorphism results in high TNF level in sepsis and septic shock. Cytokine. 2013; 61(2): 676-81. PMID: 23317877, DOI: 10.1016/j.cyto.2012.11.016
- 14. Roumen RM, Hendriks T, van der Ven-Jongekrijg J, Nieuwenhuijzen GA, Sauerwein RW, van der Meer JW, Goris RJ. Cytokine patterns in patients after major vascular surgery, hemorrhagic shock, and severe blunt trauma. Relation with subsequent adult respiratory distress syndrome and multiple organ failure. Ann Surg. 1993; 218(6): 769-76. PMID: 8257227
- 15. Turner NA, Mughal RS, Warburton P, O'Regan DJ, Ball SG, Porter KE. Mechanism of TNFα-induced IL- 1α , IL-1 β and IL-6 expression in human cardiac fibroblasts: Effects of statins and thiazolidinediones. Cardiovasc Res. 2007; 76(1): 81-90. PMID: 17612514, DOI: 10.1016/j.cardiores.2007.06.003
- 16. Howard M, Muchamuel T, Andrade S, Menon S. Interleukin 10 protects mice from lethal endotoxemia. J Exp Med. 1993; 177(4): 1205-8. PMID: 8459215
- 17. Asadullah K, Sterry W, Volk HD. Interleukin-10 Therapy—Review of a New Approach. Pharmacol Rev. 2003; 55(2): 241-69. PMID: 12773629, DOI: 10.1124/pr.55.2.4
- 18. de Waal Malefyt R, Abrams J, Bennett B, Figdor CG, de Vries JE. Interleukin 10(IL-10) inhibits cytokine synthesis by human monocytes: an autoregulatory role of IL-10 produced by monocytes. J Exp Med. 1991; 174(5): 1209-20. PMID: 1940799
- 19. Ward NS, Casserly B, Ayala A. The compensatory anti-inflammatory response syndrome (CARS) in critically ill patients. Clin Chest Med. 2008; 29(4): 617-25. PMID: 18954697, DOI: 10.1016/j.ccm.2008.06.010
- 20. Reiter RJ, Tan DX, Terron, MP, Flores LJ, Czarnocki Z. Melatonin and its metabolites: new findings regarding their production and their radical scavenging actions. Acta Biochim Pol. 2007; 54(1): 1-9. PMID: 17351668
- 21. Laothong U, Pinlaor P, Hiraku Y, Boonsiri P, Prakobwong S, Khoontawad J, Pinlaor S. Protective effect of melatonin against Opisthorchis viverriniinduced oxidative and nitrosative DNA damage and liver injury in hamsters. J Pineal Res. 2010; 49(3): 271-82. PMID: 20626588, DOI: 10.1111/j.1600-079X.2010.00792.x
- 22. Alamili M, Bendtzen K, Lykkesfeldt J, Rosenberg J, Gögenur I. Melatonin suppresses markers of inflammation and oxidative damage in a human daytime endotoxemia model. J Crit Care. 2014; 29(1): 184.e9-184.e13. PMID: 24140166, DOI: 10.1016/j.jcrc.2013.09.006

- Antioxidants that protect mitochondria reduce interleukin-6 oxidative and stress, improve mitochondrial function, and reduce biochemical markers of organ dysfunction in a rat model of acute sepsis. Br J Anaesth. 2013; 110(3): 472-80. PMID: 23381720, DOI: 10.1093/bja/aes577
- Srinivasan V, Mohamed M, Kato H. Melatonin in 24. bacterial and viral infections with focus on sepsis: a review. Recent Pat Endocr Metab Immune Drug Discov. 2012; 6(1): 30-9. PMID: 22264213
- 25. Bekyarova G, Tancheva S, Hristova M. Protective effect of melatonin against oxidative hepatic injury after experimental thermal trauma. Methods Find Exp Clin Pharmacol. 2009; 31(1): 11-4. PMID: 19357793, DOI: 10.1358/mf.2009.31.1.1338411
- 26. Bekyarova G, Apostolova M, Kotzev I. Melatonin protection against burn-induced hepatic injury by down-regulation of nuclear factor kappa B activation. Int J Immunopathol Pharmacol. 2012; 25(3): 591-6. PMID: 23058009.

DOI: 10.1177/039463201202500305

- Porter NA, Nixon JR. Isaac R. Cyclic peroxidase and 27. thiobarbituric assay. Biochim Biophys Acta. 1976; 441(3): 596-9. PMID: 3040114
- Bianchi ME. DAMPs, PAMPs and alarmins: all we 28. need to know about danger. J Leukoc Biol. 2007; 81(1): 1-5. PMID: 17032697, DOI: 10.1189/jlb.0306164
- 29. Friedl HP, Till GO, Trentz O, Ward PA. Roles of histamine, complement and xanthine oxidase in thermal injury of skin. Am J Pathol. 1989; 135(1): 203-17. PMID: 2570531
- 30. Rawlingson A, Shendi K, Greenacre SA, England TG, Jenner AM, Poston RN, Halliwell B, Brain SD. Functional Significance of Inducible Nitric Oxide Synthase Induction and Protein Nitration in the Thermally Injured Cutaneous Microvasculature. Am J Pathol. 2003; 162(4): 1373-80. PMID: 12651629, DOI: 10.1016/S0002-9440(10)63933-8
- 31. Ravat F, Payre J, Peslages P, Fontaine M, Sens N. Burn: An inflammatory process. Pathol Biol (Paris). 2011; 59(3): e63-72. PMID: 20116940, DOI: 10.1016/j.patbio.2009.12.001
- 32. Turner NA, Mughal RS, Warburton P, O'Regan DJ, Ball SG, Porter KE. Mechanism of TNFa-induced IL- 1α , IL-1 β and IL-6 expression in human cardiac fibroblasts: Effects of statins and thiazolidinediones. Cardiovasc Res. 2007; 76(1): 81-90. PMID: 17612514, DOI: 10.1016/j.cardiores.2007.06.003
- 33. Standiford TJ, Strieter RM, Lukacs NW, Kunkel SL. Neutralization of IL-10 increases lethality in

endotoxemia. Cooperative effects of macrophage inflammatory protein-2 and tumor necrosis factor. *J Immunol.* 1995; 155(4): 2222-9. PMID: 7636269

- Sehirli O, Sener E, Sener G, Cetinel S, Erzik C, Yeğen BC. Ghrelin improves burn-induced multiple organ injury by depressing neutrophil infiltration and the release of pro-inflammatory cytokines. *Peptides*. 2008; 29(7): 1231-40. PMID: 18395937, DOI: 45. 10.1016/j.peptides.2008.02.012
- 35. Bekyarova G, Atanasova M, Tzaneva M, Dimitrova A. Melatonin modulates inflammatory response and suppresses burn-induced apoptotic injury. *J Mind Med Sci.* 2017; 4(1): 59-66. DOI: 10.22543/7674.41.P5966
- Agay D, Andriollo-Sanchez M, Claeyssen R, Touvard L, Denis J, Roussel AM, Chancerelle Y. Interleukin-6, TNF-alpha and interleukin-1 beta levels in blood and tissue in severely burned rats. *Eur Cytokine Netw.* 2008; 19(1): 1-7. PMID: 18299267, DOI: 10.1684/ecn.2008.0113
- Kim HS, Kim JH, Yim H, Kim D. Changes in the Levels of Interleukins 6, 8, and 10, Tumor Necrosis Factor Alpha, and Granulocyte-colony Stimulating Factor in Korean Burn Patients: Relation to Burn Size and Postburn Time. *Ann Lab Med.* 2012; 32(5): 339-44. PMID: 22950069,

DOI: 10.3343/alm.2012.32.5.339

- Finnerty CC, Przkora R, Herndon DN, Jeschke MG. Cytokine expression profile over time in burned mice. *Cytokine*. 2009; 45(1): 20-5. PMID: 19019696, DOI: 49. 10.1016/j.cyto.2008.10.005
- 39. Gauglitz GG, Finnerty CC, Herndon DN, Mlcak RP, Jeschke MG. Are serum cytokines early predictors for the outcome of burn patients with inhalation injuries who do not survive? *Crit Care*. 2008; 12(3): R81. PMID: 18564432, DOI: 10.1186/cc6932
- Kawakami M, Kaneko N, Anada H, Terai C, Okada Y. Measurement of interleukin-6, interleukin-10, and tumor necrosis factor-alpha levels in tissues and plasma after thermal injury in mice. *Surgery*. 1997; 121(4): 440-8. PMID: 9122875
- Jeschke MG, Einspanier R, Klein D, Jauch KW. Insulin Attenuates the Systemic Inflammatory Response to Thermal Trauma. *Mol Med.* 2002; 8(8): 443-50. PMID: 12435855
- 42. Asadullah K, Sterry W, Volk HD. Interleukin-10 therapy-review of a new approach. *Pharmacol Rev.* 2003; 55(2): 241-69. PMID: 12773629, DOI: 10.1124/pr.55.2.4
- 43. Schottelius AJ, Mayo MW, Sartor RB, Baldwin AS Jr. Interleukin-10 signaling blocks inhibitor of kappaB kinase activity and nuclear factor kappaB DNA binding. *J Biol Chem.* 1999; 274(45): 31868-74. PMID: 10542212

 Lee IT, Luo SF, Lee CW, Wang SW, Lin CC, Chang CC, Chen YL, Chau LY, Yang CM. Overexpression of HO-1 protects against TNF-alpha-mediated airway inflammation by down-regulation of TNFR1dependent oxidative stress. *Am J Pathol.* 2009; 175(2): 519-32. PMID: 19608869,

DOI: 10.2353/ajpath.2009.090016

- Taniguchi T, Koido Y, Aiboshi J, Yamashita T, Suzaki S, Kurokawa A. The ratio of interleukin-6 to interleukin-10 correlates with severity in patients with chest and abdominal trauma. *Am J Emerg Med.* 1999; 17(6): 548-51. PMID: 10530532
- 46. Cesur S, Şengül A, Kurtoğlu Y, Kalpakçı Y, Özel SA, Bilgetürk A. Erdem H, Aslan T, Kınıklı S, Eyigün CP, Bıyıklı Z. Prognostic value of cytokines (TNF-α, IL-10, Leptin) and C-reactive protein serum levels in adult patients with nosocomial sepsis. *J Microbiol Infect Dis.* 2011; 1(3): 101-9.

DOI: 10.5799/ahinjs.02.2011.03.0024

- 47. Gogos CA, Drosou E, Bassaris HP, Skoutelis A. Proversus anti-inflammatory cytokine profile in patients with severe sepsis: a marker for prognosis and future therapeutic options. *J Infect Dis.* 2000; 181(1): 176-80. PMID: 10608764, DOI: 10.1086/315214
- Jaffer U, Wade RG, Gourlay T. Cytokines in the systemic inflammatory response syndrome: a review. *HSR Proc Intensive Care Cardiovasc Anesth.* 2010; 2(3): 161-75. PMID: 23441054
- Toklu HZ, Tunali-Akbay T, Erkanli G, Yüksel M, Ercan F, Sener G. Silymarin, the antioxidant component of Silybum marianum, protects against burn-induced oxidative skin injury. Burns. 2007; 33(7): 908-16. PMID: 17521818,

DOI: 10.1016/j.burns.2006.10.407

50. Liao YR, Lin JY. Quercetin intraperitoneal administration ameliorates lipopolysaccharide-induced systemic inflammation in mice. *Life Sci.* 2015; 137: 89-97. PMID: 26209141,

DOI: 10.1016/j.lfs.2015.07.015

- 51. Carrillo-Vico A, Lardone PJ, Naji L, Fernández-Santos JM, Martín-Lacave I, Guerrero JM, Calvo JR. Beneficial pleiotropic actions of melatonin in an experimental model of septic shock in mice: regulation of pro-/anti-inflammatory cytokine network, protection against oxidative damage and anti-apoptotic effects. *J Pineal Res.* 2005; 39(4): 400-8. PMID: 16207296, DOI: 10.1111/j.1600-079X.2005.00265.x
- 52. Ochoa JJ, Diaz-Castro J, Kajarabille N et al. Melatonin supplementation ameliorates oxidative stress and inflammatory signaling induced by strenuous exercise in adult human males. *J Pineal Res.* 2011; 51(4): 373–80. PMID: 21615492, DOI: 10.1111/j.1600-079X.2011.00899.x

- 53. Sasaki M, Jordan P, Joh T, Itoh M, Jenkins M, Pavlick K, Minagar A, Alexander SJ. Melatonin reduces TNFa induced expression of MAdCAM-1 via inhibition of NF-kappaB. BMC Gastroenterol. 2002; 2: 9. PMID: 56. 12003644
- 54. Cherng JY, Liu CC, Shen CR, Lin HH, Shih MF. Beneficial effects of Chlorella-11 peptide on blocking LPS-induced macrophage activation and alleviating Immunopathol Pharmacol. 2010; 23(3): 811-20. PMID: 20943052,

DOI: 10.1177/039463201002300316

55. Hoşnuter M, Gürel A, Babucçu O, Armutcu F, Kargi E, Işikdemir A. The effect of CAPE on lipid peroxidation and nitric oxide levels in the plasma of rats following thermal injury. Burns. 2004; 30(2): 121-5. PMID: 15019118,

DOI: 10.1016/j.burns.2003.09.022

Hernekamp JF, Hu S, Schmidt K, Walther A, Lehnhardt M, Kremer T. Methysergide attenuates systemic burn edema in rats. Microvasc Res. 2013; 89: 115-21. PMID: 23669653,

DOI: 10.1016/j.mvr.2013.03.002

thermal injury-induced inflammation in rats. Int J 57. Al-Jawad FH, Sahib AS, Al-Kaisy AA. Role of Antioxidants in the Treatment of Burn Lesions. Ann Burns Fire Disasters. 2008; 21(4): 186-91. PMID: 21991135