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Soil Sampling and Analysis Plan for the 3718-F Alkali Metal Treatment and Storage Facility Closure Activities

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U.S. Department of Energy Contract DE-AC06-96RL13200

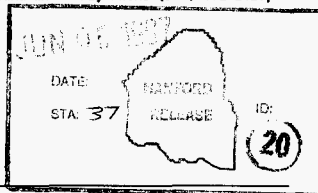
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Key Words: Dangerous Waste, Sampling, Analysis, Soils, Cleanup Levels,
Field Work

Abstract: The plan defines the soil sampling and analysis to be
performed at the 3718-F TSD to satisfy the Hanford Facility RCRA Permit
Conditions.

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HNF-SD-ENV-AP-004, Rev. 0

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SOIL SAMPLING AND ANALYSIS PLAN FOR THE 3718-F ALKALI METAL
TREATMENT AND STORAGE FACILITY CLOSURE ACTIVITIES

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44 A DATA VALIDATION: INORGANIC DATA VALIDATION REQUIREMENTS A-I

LIST OF TERMS

1		
2		
3		
4	CERCLA	<i>Comprehensive Environmental Response, Compensation and Liability Act of</i>
5		<i>1980</i>
6	CLARC	Cleanup Levels and Risk Calculation
7	CRDL	Contracted Required Detection Limit
8	Ecology	Washington State Department of Ecology
9	EII	Environmental Investigation Instructions
10	ICP	Inductively Coupled Plasma
11	IRIS	Integrated Risk Information System
12	MDL	Method Detection Limit
13	MTCA	Model Toxics Control Act
14	NIOSH	National Institute for Occupational Safety and Health
15	RCRA	<i>Resource Conservation and Recovery Act of 1976</i>
16	RTECS	Registry of Toxic Effects of Chemical Substances
17	TSD	treatment, storage, and disposal
18	WAC	Washington Administrative Code

1 SOIL SAMPLING AND ANALYSIS PLAN FOR THE 3718-F ALKALI METAL
2 TREATMENT AND STORAGE FACILITY CLOSURE ACTIVITIES
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6 1.0 PURPOSE
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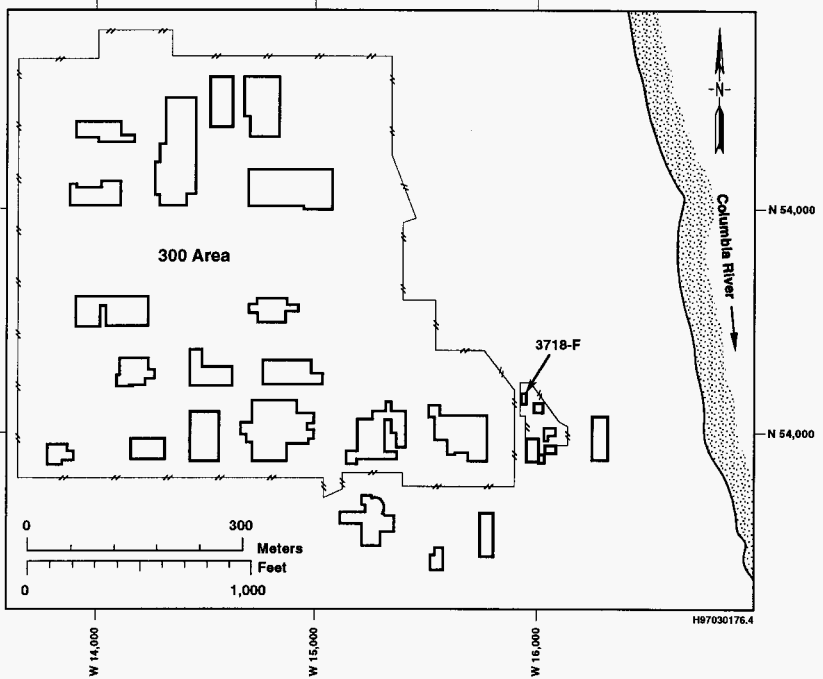
9 Amendment V.13.B.b to the approved closure plan (DOE-RL 1995a) requires that a
10 soil sampling and analysis plan be prepared and submitted to the Washington State
11 Department of Ecology (Ecology) for review and approval. Amendment V.13.B.c
12 requires that a diagram of the 3718-F Alkali Metal Treatment and Storage Facility
13 unit (the treatment, storage, and disposal [TSD] unit) boundary that is to be
14 closed, including the maximum extent of operation, be prepared and submitted as part
15 of the soil sampling and analysis plan. This document describes the sampling and
16 analysis that is to be performed in response to these requirements and amends the
17 closure plan. Specifically, this document supersedes Section 6.2, lines 43-46, and
18 Section 7.3.6 of the closure plan.
19

20 Results from the analysis will be compared to cleanup levels identified in the
21 closure plan. These cleanup levels will be established using residential exposure
22 assumptions in accordance with the Model Toxics Control Act (MTCA) Cleanup
23 Regulation (Washington Administrative Code [WAC] 173-340) as required in Amendment
24 V.13.B.1. Results of all sampling, including the raw analytical data, a summary of
25 analytical results, a data validation package, and a narrative summary with
26 conclusions will be provided to Ecology as specified in Amendment V.13.B.e. The
27 results and process used to collect and analyze the soil samples will be certified
28 by a licensed professional engineer. These results and a certificate of closure for
29 the balance of the TSD unit, as outlined in Chapter 7.0 of the approved closure plan
30 (storage shed, concrete pad, burn building, scrubber, and reaction tanks), will
31 provide the basis for a closure determination.
32
33
34

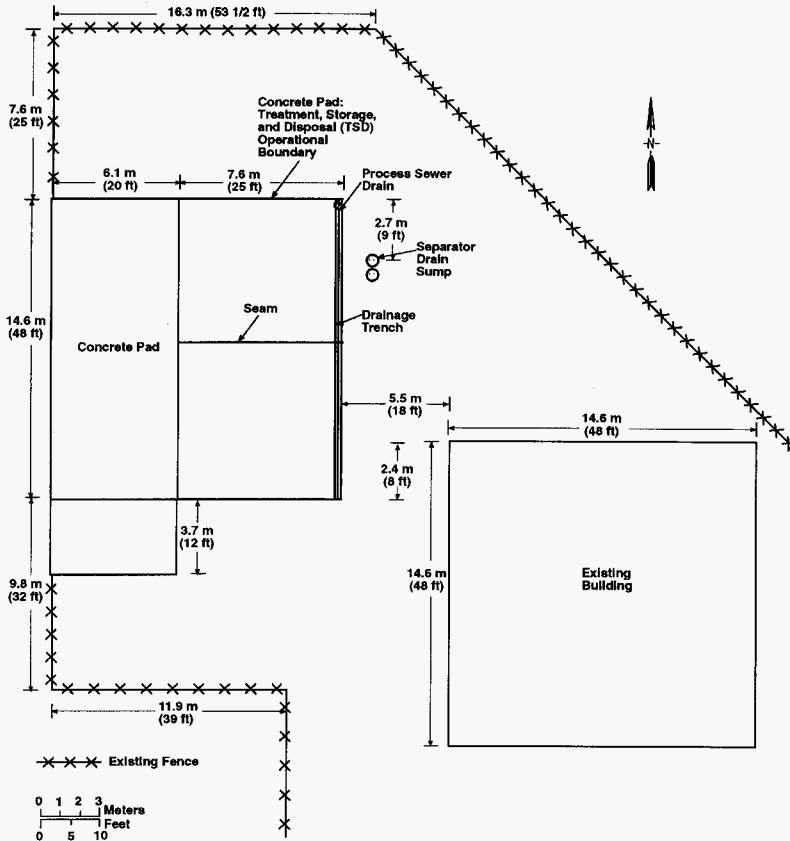
35 2.0 BACKGROUND: SITE DESCRIPTION AND STATUS
36
37

38 The 3718-F Facility is located in the southeast quadrant of the 300 Area.
39 The location of the TSD unit is shown in Figure 1. The TSD unit began treatment of
40 alkali metal waste in 1968 and continued through June 1987. Storage activities
41 began in 1968 and continued through May 1989. Equipment contaminated with sodium
42 was cleaned using baths of either water, methanol, isopropanol, or 2-butoxy ethanol.
43

44 In accordance with Amendment V.13.B.c, Figure 2 provides a diagram of the
45 TSD unit boundary to be closed. The area to be closed includes the buildings and
46 treatment tanks that are located on the concrete pad, the concrete pad, and the
47 surrounding soil within 3 meters of the concrete pad (see Figure 2). The maximum
48 extent of normal operation extended to the edge of the concrete pad, as described in



1 Figure 1. Map of 300 Area Showing Location of 3718-F Facility.



H97090178.2

1 Figure 2. Treatment, Storage, and Disposal Boundary of the 3718-F Facility as
 2 Defined in the Part A, Form 3 Permit.

1 the Part A, Form 3, "3718-F Alkali Metal Treatment and Storage Area," Rev. 4
2 (DOE-RL 1988). It has been assumed that some spillage or unintentional release of
3 product or waste (used cleaning reagent) may have flowed to the soil along the east
4 side of the concrete pad.
5

6 Three stainless steel tanks were used to treat laboratory-scale components.
7 All components treated at the TSD unit had been placed in a flowing liquid alkali
8 metal (sodium, lithium, potassium) environment before treatment. In most cases,
9 this was a liquid sodium environment. When the sodium was drained or allowed to
10 cool, some of the alkali metal adhered to the surface of the stainless steel. The
11 alkali metal, most often sodium, was removed from these components using an alcohol
12 bath in the stainless steel tanks or reaction vessels at the TSD unit. The
13 components were cleaned by immersion in one of several alcohols, such as methanol,
14 isopropanol, or 2-butoxy ethanol (Dowanol). Dowanol was most commonly used because
15 the reaction rate with sodium is slower than the reaction rate with lighter alcohols
16 and water. The majority of components that were treated in the steel tanks were
17 fabricated from stainless steel. Some of the components may have been fabricated
18 from carbon steel. The components consisted of piping, valves, sealed instruments,
19 and sealed electromagnetic pumps. The reaction process (treatment or cleaning)
20 typically would take several days depending on the amount of sodium coated on the
21 component, the presence of tight crevices, and the strength of the cleaning agent
22 (Dowanol). The final step of the cleaning process consisted of a water bath in a
23 separate tank.
24

25 From a chemistry perspective, alcohol and water are used commonly by industry
26 to remove sodium from stainless steel. The reaction process with alcohol produces
27 alkoxides and hydrogen. Although these alkoxides are strong bases, they are not
28 concentrated sufficiently to cause any damage to the steel during the reaction
29 process. The reaction of water with sodium produces sodium hydroxide and hydrogen.
30 The sodium hydroxide produced in the reaction process is not sufficiently caustic to
31 cause measurable corrosion of the stainless steel. Therefore, the reaction in the
32 treatment tanks most likely did not result of the release of nickel or chromium.
33 Visual inspection of the remaining stainless steel tank at the TSD unit showed no
34 evidence of pitting, corrosion, or oxidation. All alkali metal byproducts
35 eventually will form a carbonate when exposed to environmental media (soil, air, and
36 water).
37

38 Periodically, the used cleaning reagents (water and alcohols) were replaced or
39 replenished. Before 1985, these solutions were drained onto the concrete pad
40 through a valve in the bottom of the tanks. The concrete pad sloped toward a trough
41 that was located along the east side of the pad. The spent reagent would flow
42 across the pad to the trough and from the trough to the 300 Area Process Sewer.
43 Beginning in 1985, all spent alcohol solutions were packaged in approved containers
44 and handled as dangerous waste. The used water continued to be drained to the
45 process sewer. The TSD unit ceased treatment in 1987.
46

1 Clean closure of the buildings and treatment tanks has been completed. Closure
2 of these components followed the process outlined in Chapter 7.0 of the approved
3 closure plan. A certificate of closure for these components has been received from
4 a licensed professional engineer and placed in the Administrative Record. The
5 characterization of the soil remains and is the subject of this sampling and
6 analysis plan. As stated previously, the results and process that will be used to
7 plan, collect, and analyze soil samples will be certified by an independent licensed
8 professional engineer. These two certifications along with the results from the
9 soil sampling and analysis will provide the basis for a closure determination.

10 11 12 13 3.0 OBJECTIVE 14 15

16 The objective of the sampling and analysis is to characterize the surface and
17 shallow subsurface soils at the 3718-F Facility. The characterization activities
18 were designed (1) to determine if the operation of the TSD unit released any
19 dangerous waste to the soil that surrounds the TSD unit in sufficient quantities
20 that would pose a threat to human health and the environment; and (2) if so, to
21 define the nature and extent of contamination. The results of the characterization
22 will be compared to cleanup levels that will be quantified using the MTCA
23 methodology. Based on this analysis, a recommendation on the method of closure will
24 be submitted to Ecology.

25 26 27 28 4.0 CLEANUP LEVELS 29 30

31 In this section, soil cleanup levels are quantified for the three dangerous
32 waste constituents identified in the approved closure plan (DOE-RL 1995a). The
33 dangerous waste constituents include: sodium carbonate, lithium carbonate, and
34 potassium carbonate. The cleanup levels were established in accordance with the
35 residential exposure requirements provided in MTCA (WAC 173-340) Methods A or B
36 (see Amendment V.13.B.1) or the use of a natural background concentration as
37 specified in WAC 173-340-700(4)(d). Because soil cleanup levels have not been
38 tabulated for all of the dangerous waste constituents (Method A), cleanup levels
39 were assigned using either the natural background concentration, where appropriate,
40 or the Method B guidelines.

41
42 The continual wetting and drying of sediments under the natural environmental
43 conditions of the Hanford Site has resulted in the gradual increase of carbonates in
44 the first few meters of the vadose zone. As a result, sediments across the Hanford
45 Site contain significant amounts of carbonates.
46
47

4.1 SODIUM CARBONATE

In support of environmental restoration activities, a study was performed to establish the naturally occurring background concentrations for most nonradioactive analytes that reside in the Hanford Soils (DOE-RL 1995b). The chemical composition of over 170 soil samples from 22 locations on the Hanford Site and 3 locations adjoining the Hanford Site were used as a basis to establish the natural background concentrations. For sodium, the concentration ranged from 98.7 milligrams of metal per kilogram of soil (mg/kg) to 6,060 mg/kg. The mean concentration for the sodium analyte was calculated to be 480 mg/kg.

Using the statistical guidance provided in WAC 173-340-708(11)(d) of MTCA, a representative sitewide background concentration of 1,390 mg/kg for sodium was calculated for the 95 percent confidence interval for the 95th percentile (i.e., coverage of 95 percent and a tolerance coefficient of 95 percent). (See Chapter 6.0, Section 6.3.2.3 [DOE-RL 1995b]).

A soil cleanup level for sodium has not been tabulated in the "Model Toxics Control Act Cleanup Levels and Risk Calculation (CLARC)" document (Ecology 1996). A reference dose for sodium carbonate was not found in the Integrated Risk Information System (IRIS). Furthermore, little information on risk to human health from exposure to this constituent was found in the literature. As a point of reference, the LD₅₀ for rat (oral) was found in the National Institute for Occupational Safety and Health (NIOSH) *Registry of Toxic Effects of Chemical Substances* (RTECS) database to be 4,090 mg/kg or toxic category D.

Following the MTCA guidance (selection of the most stringent condition), a soil cleanup level of 1390 mg/kg is assigned to sodium.

4.2 LITHIUM CARBONATE

As discussed in the two previous sections, carbonates are abundant in the soil at the Hanford Site. A soil cleanup level for lithium was established using the same process outlined for sodium. Using the sitewide background concentration data (DOE-RL 1995b), the concentration for lithium was measured from 68 samples, which established a range from 34 mg/kg to 38 mg/kg. The mean was calculated to be 34.1 mg/kg. Using the tolerance interval approach, a soil cleanup level of 37 mg/kg was assigned to lithium.

Lithium carbonate is used for the treatment of some psychological disorders. The NIOSH RTECS database states that daily dosages of 1,200 mg to 1,800 mg have been used without serious side effects. The LD₅₀ for rat (oral) was found in the NIOSH RTECS database to be 525 mg/kg or toxic category D.

Following the MTCA guidance, a soil cleanup level of 37 mg/kg is assigned to lithium. This cleanup level is well below the health hazard as determined by the NIOSH RTECS database.

4.3 POTASSIUM CARBONATE

A soil cleanup level for potassium was established using the same process outlined for both lithium and sodium. The background concentration for potassium was measured from 117 samples, which established a range from 851 mg/kg to 3,780 mg/kg. The mean was calculated to be 1,414 mg/kg. Using the tolerance interval approach, a soil cleanup level of 3,090 mg/kg was assigned to potassium.

A reference dose for humans was not found in the IRIS database. The LD₅₀ for rat (oral) was found to be 1,870 mg/kg or toxic category D (NIOSH RTECS).

Following the MTCA guideline, a soil cleanup level of 3,090 mg/kg is assigned to potassium.

5.0 FIELD WORK

A phased approach will be used to characterize the soil. Phase I will consist of collecting a limited number of soil samples to determine whether dangerous waste that had resulted from the operations of the TSD unit was released to the soil column. To complete this phase, soil samples will be collected from three locations representative of worst-case contamination pathways. If these samples indicate that contaminants of concern are present above cleanup levels, the sampling and analysis plan will be revised to include a second phase of sampling, which would address the full nature and extent of soil contamination.

As currently planned, the field work will be performed by RUST Federal Services Northwest (RFS), Sampling and Mobile Laboratory (SML) personnel. The field work will be performed under the direction of RUST Federal Services of Hanford, Inc. (RFSH), Hanford Analytical Services Program, Sample Management Group (SMG). The SMG has the responsibility to coordinate all work performed in both the laboratory and the field. The coordination and control of these activities is administered through a Sample Authorization Form. The SMG will prepare and issue the Sample Authorization Form. This Sample and Analysis Plan will be used by the SMG to prepare the Sample Authorization Form.

5.1 SAMPLING PROCEDURES

The activities associated with implementing this sampling and analysis plan will be conducted in accordance with existing Environmental Investigation Instructions (EII) (WHC 1988) procedures as follows:

- EII 5.2, Soil and Sediment Sampling

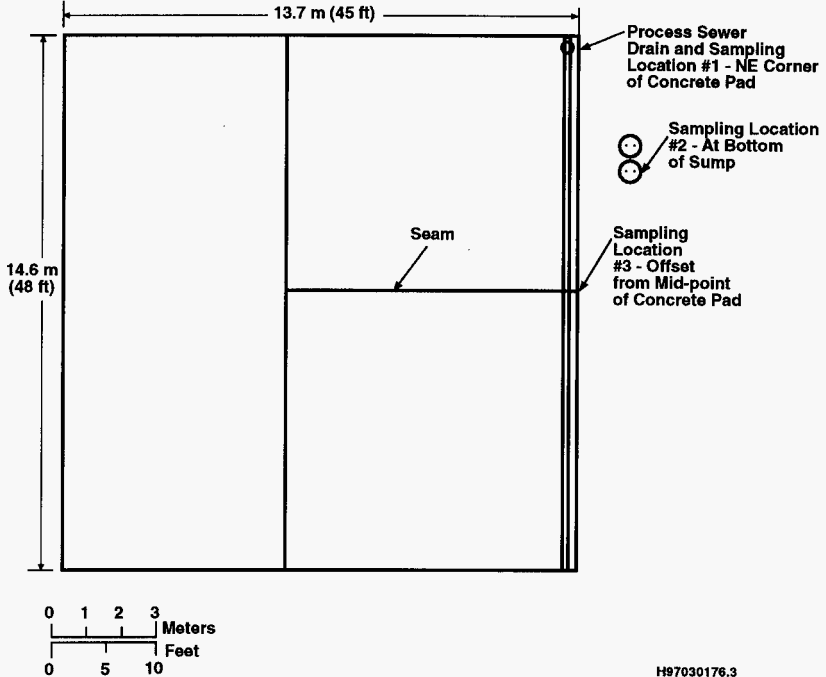
- 1 • EII 1.1, Hazardous Worker Site Entry Requirements (graded approach and as
2 required through EII 5.2)
- 3
- 4 • EII 1.13, Environmental Readiness Review (graded approach and as required
5 through EII 5.2)
- 6
- 7 • EII 1.5, Field Logbooks (as required through EII 5.2)
- 8
- 9 • EII 5.1, Chain of Custody (as required through EII 5.2)
- 10
- 11 • EII 5.4, Field Cleaning and/or Decontamination (as required through EII 5.2)
- 12
- 13 • EII 5.5, Laboratory Cleaning of RCRA/CERCLA Sampling Equipment (as required
14 through EII 5.2)
- 15
- 16 • EII 5.11, Sample Packaging and Shipping (as required through EII 5.2).
- 17
- 18

19 5.2 PHASE I ANALYTES OF CONCERN

20
21 As defined in the closure plan, the analytes of concern consist of the alkali
22 metal carbonates (sodium carbonate, potassium carbonate, and lithium carbonate) and
23 the alcohols used for cleaning (methanol, isopropanol, 2-butoxy ethanol). Sampling
24 for the three alcohols is unnecessary because these chemicals breakdown extremely
25 fast in the natural environment when exposed to sunlight and biological activities.
26 The biological half-life for methanol is estimated to be 17.8 days. The biological
27 half-life for both isopropanol and 2-butoxy ethanol is less than 24 hours.
28 Therefore, for Phase I it was decided to focus all sampling and analysis activities
29 towards the quantification of total sodium, total lithium, and total potassium. The
30 holding time for inorganic chemicals will be limited to 180 days.
31

32 5.3 PHASE I SAMPLING LOCATIONS

33
34
35 The sampling locations of the TSD Unit are shown in Figure 3. For
36 Phase I, three sampling locations along the east side of the concrete pad were
37 selected. One of these locations is at the northeast corner of the concrete slab
38 (location 1). A second location is in the Separator Drain Sump (location 2). A
39 third location is directly opposite the location of the southern treatment tank
40 (location 3). Sampling location 3 is located slightly south of the concrete
41 expansion joint that runs east to west near the middle of the pad.



DESCRIPTION OF SAMPLING LOCATIONS

1
 2 Two sample locations are at the northeast corner of the concrete pad. One of these
 3 locations (location 1) is in the soil next to the process sewer drain and the second
 4 (location 2) is at the bottom of the separator drain sump, which is located
 5 approximately 3 meters (9 feet) south of the northeast corner. A third sample
 6 location (location 3) is near the mid-point of the concrete pad, which is directly
 7 south of the expansion joint that runs east/west. All samples will be collected at
 8 a sample interval from 15 centimeters (6 inches) to 46 centimeters (18 inches).
 9 Two duplicate soil samples will be collected; one at the northeast corner and the
 10 other at sample location 3. All material placed in the sample container will be
 11 less than 12 millimeters (0.5 inches) in diameter.

12
 13 Figure 3. Soil Sampling Locations.

5.4 SOIL SAMPLING

Soil samples will be collected at 3 locations. The sump drain sample (location 2) will be collected at the bottom of the sump from 15 to 46 centimeters (6 to 18 inches) in depth. The samples collected adjacent to the concrete pad (locations 1 and 3) will be 15 to 46 centimeters (6 to 18 inches) in depth from the soil surface. Field duplicate samples will be collected at sampling locations 1 and 3 (a total of 2) and will be co-located with samples 1 and 3. Stainless steel hand tools, preferably an auger if soil and other conditions permit its use, will be used to collect the soil samples. If an auger is used, the sample material taken from the barrel of the auger will be placed in a stainless steel bowl and a representative sample will be taken from this material and placed in the sample container. Care will be taken to collect the representative sample over the full depth of the sample interval (approximately 30 centimeters). If an auger is used, the diameter of the soil sample (plug) will be from 75 to 125 millimeters depending on the nominal size of the auger. If hand tools are used (e.g., trowel or shovel), the material will be placed in a stainless steel bowl, and a representative soil sample will be collected from this material and placed in the sample container.

Clean equipment will be used for collecting each sample. If sampling equipment need decontamination, the equipment will be decontaminated with an appropriate detergent and triple rinsed. All rinsate will be collected and properly disposed. All samples will be placed in new, EPA-approved, pre-cleaned polyethylene or plastic sample containers. The sample size will be specified on the Sample Authorization Form based on contractual arrangements with the analytical laboratory that will be used. It is assumed that a minimum sample size of 500 milligrams will be required. A minimum sample size of 200 milligrams will be required for analytical purposes or as specified on the Sample Authorization Form. If possible, each sample container will be filled unless otherwise specified on the Sample Authorization Form. Care will be taken not to place any large rocks in the sample container. No material with a diameter larger than 12 millimeters (approximately 0.5 inches) should be placed in the sample containers. If the soil sample contains numerous rocks that are greater than 12 millimeters, the soil sample shall be placed in a stainless steel bowl and the larger rocks removed before placing the sample in the sample container. If additional material is needed to fill the sample container, the material will be collected from the sides of the sample excavation.

All samples will be labeled and sealed. Each sample will be assigned an unique sample number by the Sample Management Group. The sample number will be entered into the Hanford Environmental Information System (HEIS). Additional information to be placed on the label will include but not be limited to the following: project name or identifier, name and signature of samplers, and the date and time that the sample was collected. A chain-of-custody form will be established to trace the sample from the time of collection through shipping and analysis. Information to be retained on the chain-of-custody form will include but not be limited to the following: project name, sample number, and the date, time, names, and signatures of all individuals involved in sample transfers.

1 The sample labeling information and chain-of-custody record will be provided as
 2 part of the validation package. A record of all field activities will be maintained
 3 and documented in a field logbook in accordance with EII 1.5 requirements.
 4

5 All samples will be sized in accordance with the analytical needs specified in
 6 Chapter 6.0. If requested, a split sample will be provided to Ecology for their
 7 analysis. All samples, including split samples and duplicate samples, are to be
 8 collected in accordance with EII 5.2, "Soil and Sediment Sampling" (see Chapter 5.0,
 9 Section 5.1). Co-located duplicate samples will be collected at soil sampling
 10 locations 1 and 3. A summary of the soil samples to be collected is provided in
 11 Table 1.
 12
 13

14 Table 1. A Summary of Soil Sampling for Inorganics.

Location	Sample Type	No. Samples
1	Soil sample collected at northeast corner of pad, interval 6 to 18 inches	1
2	Soil sample collected at bottom of sump drain (see Figure 3), interval 6 to 18 inches	1
3	Soil sample collected near mid-point of concrete pad, interval 6 to 18 inches	1
1	Duplicate sample at location 1 (co-located) (QA/QC)	1
3	Duplicate sample at location 3 (co-located) (QA/QC)	1

17
 18
 19
 20
 21 QA/QC = quality assurance/quality control
 22
 23
 24

25 **6.0 LABORATORY ANALYSIS**
 26
 27

28 The soil samples will be analyzed for total lithium, potassium, and sodium.
 29 The constituents of concern will be analyzed in accordance with SW-846, Method 6010,
 30 Inductively Coupled Plasma - Atomic Emission Spectroscopy (ICP) Metals (EPA 1986).
 31 Table 2 provides a summary of Contract Required Detection Limit (CRDL) and
 32 Laboratory Method Detection Limit (MDL) for soil using Method 6010. These values
 33 are compared to the range in natural concentrations found across the Hanford Site.
 34 Method 6010 is appropriate for this range of data. All samples will be prepared
 35 using Method 3050 (Acid Digestion). All analysis will be performed by an Ecology-
 36 accredited laboratory under contract with the SMG. Care will be taken to limit the
 37 analysis to fine grain material (less than 2 millimeters).

Table 2. Detection Limits Using Method 6010.

Analyte	Detection Limits (mg/kg)		Background Concentration, 95% UCL (mg/kg)
	CRDL	MDL	
Sodium	1.4	2.2	1,390
Potassium	TBD	158	3,090
Lithium	0.2	0.98	37

CRDL Contract Required Detection Level: "The minimum level of detection acceptable under the contractual statement of work" (WHC 1993)

MDL Method Detection Level: "The minimum concentration of a compound that can be measured and reported with 99% confidence that the value is above zero" (WHC 1993)

7.0 QUALITY CONTROL

All procurement and coordination of field sampling activities and laboratory services will be the responsibility of the SMG. The SMG will assign sample numbers, provide sampling specifications to both the field and analytical laboratory, maintain the responsibility for shipping and receiving samples, and provide a validated data package in accordance with the following guidelines.

7.1 QUALITY CONTROL IN THE FIELD

Field quality control includes the following:

- Co-located duplicate soil samples will be collected at sample locations 1 and 3. These soil samples will be collected at a soil depth of 15 to 46 centimeters (6 to 18 inches).

Based on sample size, method of collection, media sampled, and constituents analyzed, the following applies:

- No trip blanks will be required because there will be no analysis for volatile organics
- One equipment blank will be required. This blank will be used to detect any contamination associated with the sampling equipment.

The equipment blank will be prepared using deionized water. Deionized water will be poured over or through the sampling device in the field and collected in a sample container. The collected sample will be returned to the SMG as a sample.

1 All field samples will be collected in accordance with EII 5.2, "Soil and
 2 Sediment Sampling." All sample packing and shipping will be performed in accordance
 3 with EII 5.11, "Sample Packing and Shipping." All field cleaning and
 4 decontamination of soil sampling equipment will be performed in accordance with EII
 5 5.4, "Field Cleaning and/or Decontamination." All field personnel will be trained
 6 in accordance with EII 1.7, "Qualifications and Training." No material larger than
 7 12 millimeters will be collected in the field and placed in the sample containers.
 8
 9

10 7.2 QUALITY CONTROL IN THE LABORATORY

11
 12 All soil samples will be analyzed for total sodium, lithium, and potassium.
 13 The constituents will be analyzed in accordance with SW-846, Method 6010, for ICP
 14 metals. All soil samples and associated quality control samples will be analyzed in
 15 the same batch. Additionally, one laboratory control sample, one laboratory
 16 duplicate, one method blank, and one matrix spike will be prepared and processed as
 17 part of the sample batch. All laboratory quality control samples will be prepared
 18 using Method 3050 (Acid Digestion). All laboratory work will be performed by an
 19 Ecology-approved accredited laboratory and will be managed by SMG. All quality
 20 control criteria will be in accordance with Chapter 8.0, "Inorganic Data Validation
 21 Requirements", *Data Validation Procedures for Chemical Analysis* (WHC 1993). A copy
 22 of Chapter 8.0 has been provided as Appendix A for purposes of information.
 23
 24
 25
 26
 27
 28

29 8.0 DATA VALIDATION

30 Data validation will be performed to Level D standards for inorganic chemicals,
 31 as defined in Chapter 8.0, "Inorganic Data Validation Requirements" (WHC 1993).
 32 A description of the criteria that will be used for validation is provided as
 33 Appendix A. Level D validation for ICP metals consists of the following:
 34

- 35 • Verification of required deliverables
- 36 • Verification of requested versus reported analyses
- 37 • Verification of lack of transcription errors
- 38 • Evaluation and quantification of results based on:
 - 39 - analytical holding times
 - 40 - matrix spikes
 - 41 - laboratory control samples
 - 42 - laboratory duplicates
 - 43 - analytical method blanks
 - 44 - matrix spike recoveries
 - 45 - initial and continuing instrument calibrations
 - 46 - calculational checks.

1 The specific criteria for validation of inorganic metals analyses for both 6000 and
2 7000 Series Method is provided in Appendix A. As stated in condition V.13.B.e, "The
3 results of all sampling shall be submitted to Ecology. These submittals shall
4 include the raw analytical data, a summary of analytical results, a data validation
5 package, and a narrative summary with conclusions."
6
7
8

9.0 PATH FORWARD

9
10
11 A discussion of the path forward that includes a description of tasks and
12 planned deliverables is provided in this section.
13
14
15

9.1 SCHEDULE

16
17 Task 1: Revise and submit Sampling and Analysis Plan to Ecology for approval

18
19 Task 2: Within 30 days of Ecology's approval (See Task 1), collect field
20 samples
21
22

23 Task 3: Within 45 days of completing Task 2, receive laboratory results
24

25 Task 4: Within 28 days of completing Task 3, complete validation package
26

27 Task 5: Within 14 days of completing Task 4, complete draft Data Evaluation
28 Report (see Deliverables) and submit to Ecology for review and comment
29

30 Task 6: Prepare closure package for certification or prepare Phase II
31 sampling and analysis plan.
32
33

9.2 DELIVERABLES

34
35 Deliverable 1: The sampling and analysis plan will be submitted to Ecology for
36 approval
37
38

39 Deliverable 2: Provide Ecology with a copy of the validation data package
40 including all supporting and raw data
41

42 Deliverable 3: A Data Evaluation Report will be prepared and submitted to
43 Ecology. This report will include a summary of the raw data and analytical
44 results, and a recommendation on future action
45

46 Deliverable 4: Prepare closure package for certification or a Phase II
47 sampling and analysis plan.
48

10.0 REFERENCES

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APPENDIX A
DATA VALIDATION REQUIREMENTS FOR INORGANIC SAMPLES

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APPENDIX A

DATA VALIDATION REQUIREMENTS FOR INORGANIC SAMPLES

1

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INORGANIC ANALYSIS DATA VALIDATION CHECKLIST

VALIDATION LEVEL:	A	B	C	D	E
PROJECT:			DATA PACKAGE:		
VALIDATION:		LAB:		DATE:	
CASE:			SDG:		
ANALYSES PERFORMED					
<input type="checkbox"/> CLP/CP	<input type="checkbox"/> CLP/GFAA	<input type="checkbox"/> CLP/Hg	<input type="checkbox"/> CLP/Cyanide	<input type="checkbox"/>	<input type="checkbox"/>
<input type="checkbox"/> SW-846/CP	<input type="checkbox"/> SW-846/GFAA	<input type="checkbox"/> SW-846/Hg	<input type="checkbox"/> SW-846 Cyanide	<input type="checkbox"/>	<input type="checkbox"/>
SAMPLES/MATRIX					

1. DATA PACKAGE COMPLETENESS AND CASE NARRATIVE

Is technical verification documentation present? Yes No N/A

Is a case narrative present? Yes No N/A

Comments: _____

2. HOLDING TIMES

Are sample holding times acceptable? Yes No N/A

Comments: _____

INORGANIC ANALYSIS DATA VALIDATION CHECKLIST

3. INSTRUMENT PERFORMANCE AND CALIBRATIONS

Were initial calibrations performed on all instruments? Yes No N/A
Are initial calibrations acceptable? Yes No N/A
Are ICP interference checks acceptable? Yes No N/A
Were ICV and CCV checks performed on all instruments? Yes No N/A
Are ICV and CCV checks acceptable? Yes No N/A

Comments: _____

4. BLANKS

Were ICB and CCB checks performed for all applicable analyses? Yes No N/A
Are ICB and CCB results acceptable? Yes No N/A
Were preparation blanks analyzed? Yes No N/A
Are preparation blank results acceptable? Yes No N/A
Were field/trip blanks analyzed? Yes No N/A
Are field/trip blank results acceptable? Yes No N/A

Comments: _____

5. ACCURACY

Were spike samples analyzed? Yes No N/A
Are spike sample recoveries acceptable? Yes No N/A
Were laboratory control samples (LCS) analyzed? Yes No N/A
Are LCS recoveries acceptable? Yes No N/A

Comments: _____

INORGANIC ANALYSIS DATA VALIDATION CHECKLIST

6. PRECISION

Were laboratory duplicates analyzed?	Yes	No	N/A
Are laboratory duplicate samples RPD values acceptable?	Yes	No	N/A
Were ICP serial dilution samples analyzed?	Yes	No	N/A
Are ICP serial dilution %D values acceptable?	Yes	No	N/A
Are field duplicate RPD values acceptable?	Yes	No	N/A
Are field split RPD values acceptable?	Yes	No	N/A

Comments: _____

7. FURNACE AA QUALITY CONTROL

Were duplicate injections performed as required?	Yes	No	N/A
Are duplicate injection %RSD values acceptable?	Yes	No	N/A
Were analytical spikes performed as required?	Yes	No	N/A
Are analytical spike recoveries acceptable?	Yes	No	N/A
Was MSA performed as required?	Yes	No	N/A
Are MSA results acceptable?	Yes	No	N/A

Comments: _____

8. REPORTED RESULTS AND DETECTION LIMITS

Are results reported for all requested analyses?	Yes	No	N/A
Are all results supported in the raw data?	Yes	No	N/A
Are results calculated properly?	Yes	No	N/A
Do results meet the CRDLs?	Yes	No	N/A

Comments: _____

Matrix Spike Recovery (% R)

$$\frac{SSR-SR}{SA} \times 100 \text{ where,} \quad \text{D-1}$$

SSR = spike sample result
 SR = sample result
 SA = spike added

Relative Percent Difference (RPD)

MS/MSD analysis RPD:

$$RPD = \frac{|MSR - MSDR|}{(MSR + MSDR) / 2} \times 100 \text{ where,} \quad \text{D-2}$$

MSR = matrix spike recovery
 MSDR = matrix spike duplicate recovery

Duplicate sample analysis RPD:

$$RPD = \frac{|S-D|}{(S+D)/2} \times 100 \text{ where,} \quad \text{D-3}$$

S = sample concentration
 D = duplicate sample concentration

Relative Standard Deviation (%RSD)

$$\% RSD = \frac{SD}{\bar{x}} \times 100 \text{ where,} \quad \text{D-4}$$

SD = Standard deviation of the initial five response factors per compound, calculated as

$$\sqrt{\frac{\sum_{i=1}^n (x_i - \bar{x})^2}{n-1}} \text{ and,} \quad \text{D-5}$$

\bar{x} = mean of initial five response factors

Relative Response Factor (RRF) (volatiles and semivolatiles)

$$RRF = \frac{A_x}{A_{is}} \times \frac{C_{is}}{C_x} \text{ where,} \quad D-6$$

- A_x = area of the characteristic ion measured
 A_{is} = area of the characteristic ion measured for the associated internal standard
 C_x = concentration (in ng for volatiles and ng/ μ l for semivolatiles) of the characteristic compound of interest
 C_{is} = concentration (in ng for volatiles and ng/ μ l for semivolatiles) of the characteristic associated interval standard

Percent Difference (% D, volatile and semivolatile RRF values)

$$\%D = [(RRF_i - RRF_o)/RRF_i] \times 100 \text{ where,} \quad D-7$$

RRF_i = the average RRF from the initial calibration

RRF_o = the RRF value from the continuing calibration

Surrogate Recoveries (%R)

$$\% R = (Q_d/Q_a) \times 100 \text{ where,} \quad D-8$$

Q_d = quantity of surrogate determined from the analysis

Q_a = quantity of surrogate added to the sample

Results Calculations (volatile analysis)

- Water Samples, μ g/L

$$\frac{(A_x)(I_s)(D_i)}{(A_{is})(RRF)(V_o)} \text{ where,} \quad D-9$$

A_x = area of the quantitation ion (EICP) for the compound measured

A_{is} = area of the quantitation ion (EICP) for the specified internal standard

I_s = amount of internal standard added in nanograms (ng)

- RRF = relative response factor from the ambient temperature purge of the calibration standard
- V_o = volume of water purged in milliliters (mL)
- D_f = dilution factor. The dilution factor for the analysis of water samples for volatiles is defined as the ratio of the number of mL of water purged (V_o) to the number of mL of the original sample used for purging. For example, if 2.5 mL of sample is diluted to 5.0 mL with reagent water and purged, $D_f = 5.0 \text{ mL}/2.5 \text{ mL} = 2.0$. If no dilution is performed, $D_f = 1.0$.

- Soil/sediment samples, low level, $\mu\text{g}/\text{Kg}$

$$\frac{(A_x)(I_s)}{(A_s)(RRF)(W_s)(D)} \text{ where,} \quad \text{D-10}$$

- A_x, I_s, A_s, RRF are as given for water samples above.
- $D = (100 - \% \text{moisture})/100$
- $W_s = \text{weight of sample added to the purge tube in grams (g)}$.

- Soil/sediment samples, medium level, $\mu\text{g}/\text{Kg}$

$$\frac{(A_x)(I_s)(1000)(D_f)}{(A_s)(RRF)(V_s)(W_s)(D)} \text{ where} \quad \text{D-11}$$

- A_x, I_s, A_s, RRF are as given for water samples above
- $V_t = \text{total volume of methanol extract in mL. Note: this volume is typically 10.0 mL, even though only 1.0 mL is transferred to a vial for storage as specified in the SOW}$
- $V_s = \text{volume of the aliquot of methanol extract in } \mu\text{L added to reagent water for purging}$
- $W_s = \text{weight of soil extracted, in grams}$
- $D = \text{same as given for low level soils above}$
- $D_f = \text{Dilution factor. The dilution factor for the analysis of soil/sediment samples for volatiles by the } \underline{\text{medium level}} \text{ method is defined as the ratio of the number of nanogram of methanol added to the reagent water for purging, (i.e. } V_s, \text{ above), to the number of nanogram of methanol } \underline{\text{extract}} \text{ of the sample contained in volume } V_s. \text{ The dilution factor is equal to 1.0 in all cases other than those requiring dilution of the methanol extract. The factor of 1,000 in the numerator converts the value of } V_t \text{ from mL to } \mu\text{L.}$

Result Calculations (semivolatiles analysis)

- Water samples, $\mu\text{g/L}$

$$\frac{(A_x)(I_s)(V_i)(D_i)}{(A_s)(RRF)(V_o)(V_i)} \text{ where,} \quad \text{D-12}$$

- A_x = area of the quantitation ion (EICP) for the compound measured
 A_s = area of the quantitation ion (EICP) for the specified internal standard
 I_s = amount of internal standard added in nanograms (ng)
 V_o = volume of water extracted in milliliters (mL)
 V_i = volume of extract injected in microliters (μL)
 V_c = volume of concentrated extract in μL
 RRF = relative response factor from the daily calibration standard
 D_i = dilution factor. The dilution factor for analysis of water samples for semivolatiles is defined as follows:

$$\frac{\mu\text{L of most conc. extracted used for dilution} + \mu\text{L clean solvent}}{\mu\text{L most conc. extract used to make dilution}}$$

If no dilution is performed, $D_i = 1.0$

- Soil samples, $\mu\text{g/Kg}$

$$\frac{(A_x)(I_s)(V_i)(D_i)}{(A_s)(RRF)(V_i)(W_s)(D)} \text{ where,} \quad \text{D-13}$$

A_x , I_s , A_s , RRF , V_i , V_i , D_i are as given for water samples above

- D = $(100 - \% \text{moisture})/100$
 W_s = weight of sample extracted in grams (g).

Resolution (pesticide analysis)

$$\text{Resolution} = (P_v/P_h) \times 100 \text{ where,} \quad \text{D-14}$$

- P_v = the peak height of the valley of the larger peak
 P_h = the peak height of the smaller peak being resolved.

Percent Difference (%D, pesticides/PCB analysis)

$$\%D = (RT_i - RT_s) / RT_i \times 100 \text{ where,} \quad \text{D-15}$$

RT_i = the absolute retention time of DBC in the initial standard (EVAL A) mixture

RT_s = the absolute retention time of DBC in the sample and subsequent standards

Percent Breakdown (pesticides/PCB analysis)

$$\%Breakdown = \frac{\text{Total Degradation Peak Area}}{\text{Total Peak Area}} \times 100 \text{ where,} \quad \text{D-16}$$

total degradation peak area = the sum of the peak area for DDE and DDD or endrin aldehyde and endrin ketone

total peak area = sum of all associated peak areas for DDE, DDD and DDT or endrin, endrin aldehyde and endrin ketone.

DBC Retention Time Percent Difference (pesticide/PCB analysis)

$$\%D = \frac{RT_i - RT_s}{RT_i} \times 100 \text{ where,} \quad \text{D-17}$$

RT_i = Absolute retention time of dibutylchloroendate in the initial standard (Evaluation Standard Mix A)

RT_s = Absolute retention time of dibutylchloroendate in the subsequent standard and sample analyses

Relative Response Factors (RRF_n and RRF_m, dioxin analysis)

$$RRF_n \text{ (unlabeled)} = \frac{A_x \times Q_{is}}{Q_x \times A_{is}} \text{ and } RRF_m \text{ (labeled)} = \frac{A_{is} \times Q_{rs}}{Q_{is} \times A_{rs}} \quad \text{D-18}$$

$$RSD = \frac{RRF_m}{RRF_s} \text{ where,} \quad \text{D-19}$$

where,

- A_x = sum of the integrated ion abundances of the quantitation ions for the unlabeled PCDDs/PCDFs
- A_{is} = sums of the integrated ion abundances of the quantitation ions for the labeled internal standards
- A_{rs} = sum of the integrated ion abundances of the quantitation ions for the labeled recovery standards
- Q_{is} = quantity of the internal standard injected (picograms pg)
- Q_{rs} = quantity of the recovery standard injected (pg)
- Q_x = quantity of the unlabeled PCDD/PCDF analyte injected (pg)
- RRF_m = mean of the five response factors for the particular PCDD/PCDF analyte
- RRF_s = standard deviation of the five response factors for the particular PCDD/PCDF analyte

Results Calculations (inorganics analysis)

- Water, µg/L = C x D where, D-20
 - C = concentration, µg/L from calibration curve
 - D = dilution factor (if any)
- Soil, mg/Kg = (C x D) / (W x S) where, D-21
 - C = concentration, mg/L from calibration curve
 - D = volume in mL of final extract volume
 - W = weight of sample in grams
 - S = (100 - %moisture)/100

Serial Dilution (%D, inorganics analysis)

$$\%D = [I - S / I] \times 100 \text{ where,} \quad \text{D-22}$$

- I = analyte concentration before dilution
- S = serial dilution analyte concentrations

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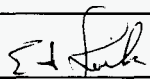
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From	E. F. Loika, Director, FFTF Project 	N2-51	File No. Or Ref.: BWHC-9754727
Subj	ISSUANCE OF SOIL SAMPLING AND ANALYSIS PLAN FOR THE 3718-F ALKALI METAL TREATMENT AND STORAGE FACILITY CLOSURE ACTIVITIES, HFN-SD-ENV-AP-004, REV.0, MAY 1997		Date:

The 3718-F Alkali Metal Treatment and Storage Facility Closure Plan was included in Revision 3 of the Hanford Facility Resource Conservation and Recovery Act of 1976 Permit in November 1996. Included at that time was a unit-specific condition that required the collection and analysis of soil samples. This condition (Permit Condition V.13.B.b) states, "the Department shall be provided, for review and approval, a soil sampling and analysis plan at least 30 days prior to initiating actual sampling. Such a plan shall include a schedule for conducting sampling events. The analytical results of the sampling event will be used to determine if corrective action will be required to close the 3718-F Alkali Metal Treatment and Storage Facility."

In response to this requirement, the subject soil sampling and analysis plan was submitted to the Washington State Department of Ecology for review and comment at the Project Managers' Meeting held March 13, 1997. Approval of the soil sampling and analysis plan was received on April 24, 1997.

The subject plan was prepared by Rust Federal Services of Hanford, Inc. (RFSH), Environmental Services Department for B&W Hanford Company (BWHC).

Should you have any questions or comments, please contact Mr. T. A. Dillhoff (BWHC) of my staff at (509) 376-0441 or Mr. J. C. Sonnichsen (RFSH) at (509) 376-9956.

sms

Attachment

BWHC-9754727

ATTACHMENT

HFN-SD-ENV-AP-004
"Soil Sampling and Analysis Plan for the
3718-F Alkali Metal Treatment and Storage Facility
Closure Activities"

Consisting of 47 pages,
including cover page

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