



Supplement of

Low-volatility compounds contribute significantly to isoprene secondary organic aerosol (SOA) under high-NO $_x$ conditions

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S1 Comparisons between Kinetic Model and Experimental Results

Reaction	Rate (cm ³ molecule ^{-1} s ^{-1})	Source
$CH_3ONO + h\nu \rightarrow HCHO + HO_2 + NO$	$(1.4-2.3) \ge 10^{-4} \text{ s}^{-1}$	NA ^a
$CH_3ONO + OH \rightarrow H_2O + HCHO + NO$	$3 \ge 10^{-13} \ge 0.5$	See note ^b
$CH_3ONO + OH \rightarrow HCHO + HO_2 + HONO$	$3 \ge 10^{-13} \ge 0.5$	See note ^b
HO_2 + $\mathrm{NO}_2 \rightarrow \mathrm{HONO}$	$5 \ge 10^{-16}$	JPL
HO_2 + HCHO \rightarrow HOCH ₂ OO	9.7 x $10^{-15} \exp(625/T)$	IUPAC
$\mathrm{HOCH}_2\mathrm{OO} ightarrow \mathrm{HO}_2$ + HCHO	2.4 x $10^{12} \exp(-7000/T) \text{ s}^{-1}$	IUPAC
$\mathrm{HOCH}_2\mathrm{OO} + \mathrm{HO}_2 \rightarrow \mathrm{HMHP}$	$5.6 \ge 10^{-15} \exp(2300/T) \ge 0.5$	IUPAC
$\mathrm{HOCH}_2\mathrm{OO} + \mathrm{HO}_2 \rightarrow \mathrm{HCOOH}$	$5.6 \ge 10^{-15} \exp(2300/\text{T}) \ge 0.3$	IUPAC
$HOCH_2OO + HO_2 \rightarrow HCOOH + HO_2 + OH$	$5.60 \ge 10^{-15} \exp(2300/T) \ge 0.2$	IUPAC
$HOCH_2OO \rightarrow HCOOH$	$7 \ge 10^{-13} \ge RO_2$	IUPAC
$\mathrm{HOCH}_2\mathrm{OO} ightarrow \mathrm{CH}_2(\mathrm{OH})_2$	$7 \text{ x } 10^{-13} \text{ * RO}_2$	IUPAC
$\mathrm{HOCH}_2\mathrm{OO} ightarrow \mathrm{HCOOH} + \mathrm{HO}_2$	$5.50 \ge 10^{-12} \ge 2 \ge RO_2$	IUPAC
$HOCH_2OO + NO \rightarrow HCOOH + HO_2 + NO_2$	$5.60 \ge 10^{-12}$	IUPAC
HMHP + OH \rightarrow HOCH ₂ OO	$3.1 \ge 10^{-11} \ge 0.12$	Jenkin (2007)
$\rm HMHP + OH \rightarrow \rm HCOOH + OH$	$3.1 \ge 10^{-11} \ge 0.88$	Jenkin (2007)
HMHP + $h\nu \rightarrow$ HCOOH + HO ₂ + OH	$2.0 \text{ x } 10^{-7} \text{ s}^{-1}$	JPL
$OH + OH \rightarrow O$	$6.2 \ge 10^{-14} (T/298)^{2.6} \exp(945/T)$	IUPAC
$OH + NO_2 + M \rightarrow HOONO + M$	Termolecular	IUPAC
$\rm HOONO + M \rightarrow OH + NO_2 + M$	Termolecular	IUPAC
$OH + OH + M \rightarrow H_2O_2 + M$	Termolecular	IUPAC
$\mathbf{OH} + \mathbf{NO}_2 + \mathbf{M} ightarrow \mathbf{HNO}_3 + \mathbf{M}$	Termolecular	IUPAC
\mathbf{NO}_2 + $\mathbf{O}_3 ightarrow \mathbf{NO}_3$	1.4 x 10 ⁻¹³ exp(-2470/T)*0.97	Cantrell (1985)
\mathbf{NO}_2 + $\mathbf{O}_3 ightarrow \mathbf{NO}$	1.4 x 10 ⁻¹³ exp(-2470/T)*0.03	Cantrell (1985)
$NO_2 + NO_2 + M \rightarrow N_2O_4 + M$	Termolecular	IUPAC
$N_2O_4 + M \rightarrow NO_2 + NO_2 + M$	Termolecular	IUPAC
$\textbf{CISOPAO2 + NO} \rightarrow \textbf{CISOPAO + NO2}$	KRO2NO*0.88	Wennberg (2018)
$\textbf{CISOPAO2 + NO} \rightarrow \textbf{ISOPANO3}$	KRO2NO*0.12	Wennberg (2018)
$\mathbf{ISOPBO2} + \mathbf{NO} \rightarrow \mathbf{ISOPBNO3}$	KRO2NO*0.14	Wennberg (2018)
$\mathbf{ISOPBO2} + \mathbf{NO} \rightarrow \mathbf{ISOPBO} + \mathbf{NO2}$	KRO2NO*0.86	Wennberg (2018)

Table S1: Additional reactions and reaction rates included in the kinetic model, but not in MCM v3.3.1

Reaction	Rate (cm ³ molecule ^{-1} s ^{-1})	Source
$\mathbf{CISOPCO2} + \mathbf{NO} \rightarrow \mathbf{ISOPCNO3}$	KRO2NO*0.12	Wennberg (2018)
$\mathbf{ISOPDO2} + \mathbf{NO} \rightarrow \mathbf{ISOPDNO3}$	KRO2NO*0.13	Wennberg (2018)
$\mathbf{ISOPDO2} + \mathbf{NO} \rightarrow \mathbf{ISOPDO} + \mathbf{NO2}$	KRO2NO*0.87	Wennberg (2018)
$\mathbf{ISOPAO2} + \mathbf{NO} \rightarrow \mathbf{ISOPANO3}$	KRO2NO*0.12	Wennberg (2018)
$\mathbf{ISOPAO2} + \mathbf{NO} \rightarrow \mathbf{ISOPAO} + \mathbf{NO2}$	KRO2NO*0.88	Wennberg (2018)
$\mathbf{ISOPCO2} + \mathbf{NO} \rightarrow \mathbf{CISOPCO} + \mathbf{NO2}$	KRO2NO*0.88	Wennberg (2018)
$\mathbf{ISOPCO2} + \mathbf{NO} \rightarrow \mathbf{ISOPCNO3}$	KRO2NO*0.12	Wennberg (2018)
$\mathbf{OH} \textbf{+} \mathbf{ISOPBNO3} \rightarrow \mathbf{INB1O2}$	3.0 x 10 ⁻¹¹ * 0.75	Wennberg (2018)
$OH + ISOPBNO3 \rightarrow INB2O2$	3.0 x 10 ⁻¹¹ * 0.25	Wennberg (2018)
$OH + ISOPCNO3 \rightarrow INCO2$	8.0 x 10 ⁻¹¹	Wennberg (2018)
$OH + ISOPDNO3 \rightarrow INDO2$	4.15 x 10 ⁻¹¹ *0.9	Wennberg (2018)
$OH + ISOPDNO3 \rightarrow IND1O2$	4.15 x 10 ⁻¹¹ *0.1	Wennberg (2018)
IND1O2 + HO2 \rightarrow INDOOH	KRO2HO2*0.706	Wennberg (2018)
IND1O2 + NO \rightarrow INB1NO3	KRO2NO*0.104	Wennberg (2018)
IND1O2 + NO \rightarrow IND1O + NO2	KRO2NO*0.896	Wennberg (2018)
IND1O2 + NO3 \rightarrow IND1O + NO2	KRO2NO3	Wennberg (2018)
$IND1O2 \rightarrow IND1O$	8.00 x 10 ⁻¹³ *0.8*RO ₂	Wennberg (2018)
$IND1O2 \rightarrow INDOH$	8.00 x 10 ⁻¹³ *0.2*RO ₂	Wennberg (2018)
$\mathbf{IND1O} \rightarrow \mathbf{C58ANO3} + \mathbf{HO2}$	KDEC	Wennberg (2018)
$\textbf{CISOPAO} \rightarrow \textbf{HC4CCHO} + \textbf{HO2}$	KDEC	Wennberg (2018)
$\textbf{CISOPCO} \rightarrow \textbf{HC4ACHO} + \textbf{HO2}$	KDEC	Wennberg (2018)
ISOPAO \rightarrow HC4CCHO	KDEC	Wennberg (2018)
$OH + MPAN \rightarrow ACETOL + CO + NO3$	2.9 x 10 ⁻¹¹ * 0.25	Wennberg (2018)
$OH + MPAN \rightarrow HMML + NO3$	2.9 x 10 ⁻¹¹ * 0.75	Wennberg (2018)

Notes: HMHP = $CH_2(OH)(OOH)$ and all other names are identical to those used in MCMv3.3.1. All reactions in bold were already included in MCM v3.3.1, but have been revised for this work based on the source listed and as described in the text. ^{*a*} CH₃ONO photolysis was calculated from the GC-FID measurements. ^{*b*} Nielsen (1991), Cox (1980), and Jenkin (1988).

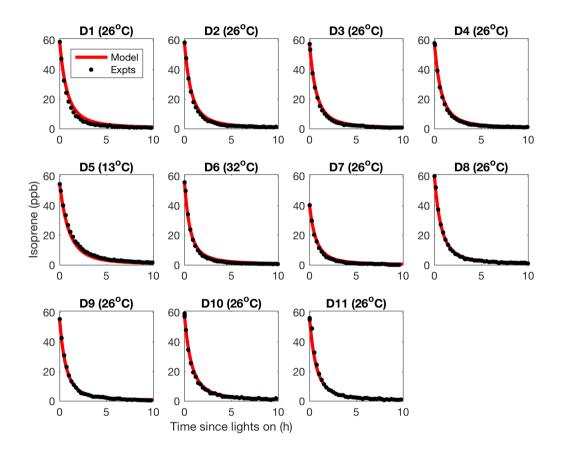


Figure S1. Isoprene observed (black) compared to simulated (red) for all LV (low volatility) pathway experiments.

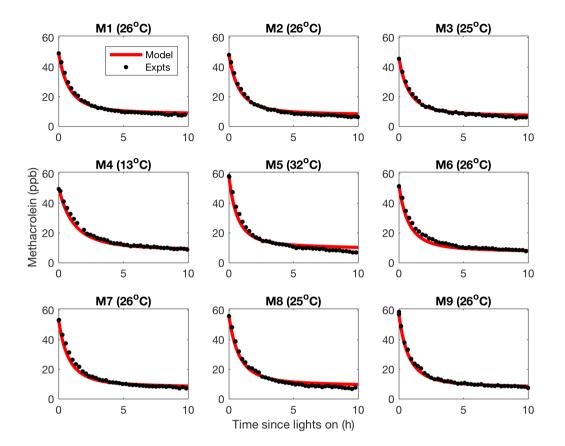


Figure S2. Methacrolein observed (black) compared to simulated (red) for all 2MGA (2-methyl glyceric acid and oligomers) pathway experiments.

As shown in Figure S3, NO₂ and NO compare reasonably well with the model in both the LV and 2MGA experiments. NO is measured using a Teledyne NO_x analyzer (T200) and NO₂ is measured using a luminol NO₂/acyl peroxynitrate analyzer developed by Fitz Aerometric Technologies. There is a large model bias in the NO₂/NO ratio (Figure S3), but this bias is largely caused by differences of only several ppb in the NO level between the model and observations. NO/NO_x and NO₂/NO_x are much more similar between the model and observations and are a more relevant metric for determining MPAN formation at high NO₂/NO ratios. Remaining biases are likely caused by unknown measurement interferences or unaccounted for wall deposition of NO_x reservoir species (e.g., N₂O₅, HNO₃, HO₂NO₂, etc.) in the kinetic model.

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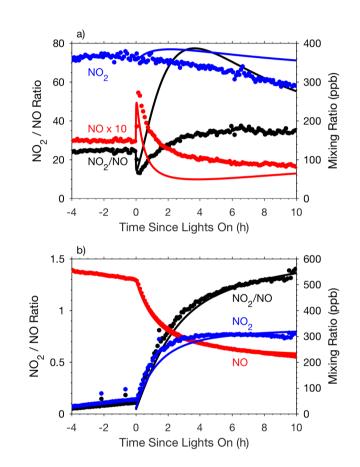


Figure S3. NO_2 (blue, right axis), NO (red, right axis), and NO_2 / NO ratio (black, left axis) for an example 2MGA experiment (M2, panel a) and an example LV experiment (D8, panel b) comparing experimental data (markers) and kinetic model results (lines). Note: in panel a, the NO mixing ratio is multiplied by 10 for ease of viewing.

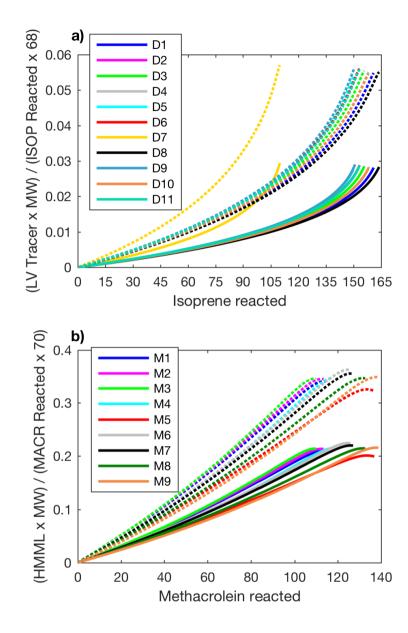


Figure S4. Simulated known gas-phase SOA precursors divided by VOC reacted as a proxy of SOA mass yield for all LV pathway experiments (panel a) and all 2MGA pathway experiments (panel b). The kinetic model confirms experimental conditions were similar enough to produce relatively consistent yields of known gas-phase SOA precursors for both systems. In panel a, all LV pathway gas-phase tracers listed in Table S2 with an estimated fraction in the particle-phase > 65% at 299 K are included as solid lines and > 5% as dashed lines. In panel b, HMML gas-phase SOA precursor is converted only based on the mass of HMML itself - 102 g/mol (solid lines) and 2MGA-nitrate mass - 165 g/mol (dashed lines). In both panels for all cases, the F_P values calculated in Table S2 are not used, so in this proxy for SOA yield all simulated gas-phase SOA precursors are assumed to exist 100% in the particle-phase.

Table S2: Estimated saturation mass concentration (C^*) and fraction in particle phase (F_P) for organic nitrates and dinitrates in MCM v3.3.1.

MCM Name	Structure	C*(299K)	C*(286K)	C*(305K)	F _P (299K)	F _P (286K)	F _P (305K)
IVOC at 299 K							
ISOP34NO3	O ₂ NO	6.59E+05	1.99E+05	1.09E+06	0	0	0
ISOPDNO3		4.47E+05	1.32E+05	7.47E+05	0	0	0
ISOPBNO3		4.47E+05	1.32E+05	7.47E+05	0	0	0
ISOPCNO3	HO ONO ₂	2.28E+05	6.40E+04	3.89E+05	0	0	0
ISOPANO3	HO ONO ₂	2.28E+05	6.40E+04	3.89E+05	0	0	0
MACRNB	ONO2	1.90E+05	5.15E+04	3.29E+05	0	0	0
HMVKANO3	O ₂ NO OH	9.35E+04	2.46E+04	1.64E+05	0	0.001	0
MACRNO3	OTONO2 OH	4.62E+04	1.13E+04	8.36E+04	0.001	0.002	0
MVKNO3		3.72E+04	9.17E+03	6.70E+04	0.001	0.002	0
C530NO3	O ONO2 OH	2.64E+04	6.16E+03	4.87E+04	0.001	0.004	0
C51NO3	OH ONO2	2.14E+04	5.03E+03	3.94E+04	0.001	0.004	0.001
C47CHO		3.06E+03	5.82E+02	6.14E+03	0.009	0.038	0.004

Table S2: Estimated saturation mass concentration (C^*) and fraction in particle phase (F_P) for organic nitrates and dinitrates in MCM v3.3.1.

MCM Name	Structure	C*(299K)	C*(286K)	C*(305K)	F _P (299K)	F _P (286K)	F _P (305K)
C4M2ALOHN	03 _{os}	1.85E+03	3.39E+02	3.79E+03	0.014	0.063	0.006
	O₂NO OH						
C58ANO3	O ₂ NO OH	4.36E+02	7.01E+01	9.42E+02	0.059	0.245	0.025
INDHCHO	O ₂ NO OH O OH	4.36E+02	7.01E+01	9.42E+02	0.059	0.245	0.025
INCNCHO		3.05E+02	4.68E+01	6.70E+02	0.082	0.327	0.034
INCCO		3.04E+02	4.82E+01	6.59E+02	0.082	0.32	0.035
SVOC at 299 I	K						
C58NO3		2.56E+02	3.93E+01	5.63E+02	0.096	0.366	0.041
C57NO3		2.56E+02	3.93E+01	5.63E+02	0.096	0.366	0.041
INANCHO		1.86E+02	2.74E+01	4.16E+02	0.128	0.453	0.054
INB1NACHO		1.31E+02	1.89E+01	2.95E+02	0.172	0.546	0.075
INB1NBCHO		1.31E+02	1.89E+01	2.95E+02	0.172	0.546	0.075
INANCO		1.05E+02	1.51E+01	2.37E+02	0.206	0.601	0.092

Table S2: Estimated saturation mass concentration (C^*) and fraction in particle phase (F_P) for organic nitrates and dinitrates in MCM v3.3.1.

MCM Name	Structure	C*(299K)	C*(286K)	C*(305K)	F _P (299K)	F _P (286K)	$F_P(305K)$
HMVKNO3	ОН	4.61E+01	6.33E+00	1.06E+02	0.371	0.782	0.183
INCNO3	02NO	1.32E+01	1.53E+00	3.27E+01	0.673	0.937	0.422
	O ₂ NO OH OH						
INANO3	O ₂ NO	7.67E+00	8.50E-01	1.93E+01	0.78	0.964	0.552
	HO ONO2 OH						
INB1NO3	HO HO	5.23E+00	5.66E-01	1.33E+01	0.838	0.976	0.642

Notes: C* is the saturation mass concentration in μ g cm⁻³ and F_P is the fraction of a compound estimated to be present in the particle phase.

In Table S2, vapor pressure is estimated using the vapor pressure and boiling point estimations from Nannoolal et al. (2004, 2008) using the online calculator located at: http://www.aim.env.uea.ac.uk/aim/ddbst/pcalc_main.php. The saturation mass concentration (C*) is calculated using the equation: $C^* = P^0 \gamma MW / (RT)$ where P^0 is the vapor pressure, γ is the activity coefficient, MW is the molecular weight, R is the gas constant, and T is the temperature (Seinfeld and Pandis, 2016). Here

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the activity coefficient (γ) is unknown and so assumed to be 1. The amount of each compound in the particle phase (F_P) is estimated using the equation: $F_P = (1 + C*/C_{OA})^{-1}$ where C_{OA} is the concentration of the organic aerosol (Seinfeld and Pandis, 2016). The reported F_P in Table S2 uses the measured 1 hr average C_{OA} after 10 hr of photooxidation - 27.16, 22.71, 23.87 µg cm⁻³ for experiments D3 (299 K), D5 (286 K), and D6 (305 K), respectively.

S2 Corrections for Particle Coagulation and Particle Wall Deposition

For each experiment, after all gases and particles were injected into the chamber, purified air was added to facilitate mixing. Photooxidation was delayed by 4 h, during which particle wall deposition was measured. The first 0.5 h of this 4 h period was not used in the particle wall deposition to ensure that air currents and particles/gases in the chamber had stabilized.

- 5 A numerical model, similar to that reported by Nah et al. (2017), Sunol et al. (2018), and Charan et al. (2018) was used to simulate Brownian diffusion, particle settling, and electrostatic effects. The numerical model based on the aerosol dynamic equation (e.g., Sunol et al. (2018)) assumes $\beta(D_p,t)$ follows the Crump and Seinfeld (1981) equation for a spherical chamber. Prior to injection, all ammonium sulfate particles were passed through a soft x-ray source (TSI Model 3088) in order to impart a consistent initial charge distribution with a net charge of zero. For the particle coagulation/wall deposition correction, the
- 10 initial charge distribution was assumed to be that computed by Leppa et al. (2017), which is an update to Lopez-Yglesias and Flagan (2013) and Wiedensohler (1988) and consistent with the charge distribution assumed in the DMA inversion (Section 2.2 of the main text). Only charges from -8 to 8 are considered, which is sufficient for the particles used in this study, which have a diameter range of 30 800 nm. All particles measured by the DMA were grouped into 15 size bins. The DMA collects data across 390 size bins, but reducing the size bin number decreased the analysis uncertainty by increasing the number of particles
- 15 per size bin. The only unknown parameters in the numerical model then become the mean electrostatic field experienced within the chamber (\bar{E}) and the chamber eddy diffusion coefficient (k_e). The numerical model determines k_e and \bar{E} by comparing the observed particle dynamics to that simulated and minimizing the optimization function J.

$$J = \int_0^{t_{final}} \sum_{D_p} \left(\left(\sum_{charges} N(D_p, t; ke, \bar{E})_{simulated} - N(Dp, t)_{observed} \right)^2 \frac{N(D_p, t)_{observed}}{\sum_{D_p} N(D_p, t)_{observed}} \right) dt \tag{1}$$

The particle wall deposition coefficients ($\beta(D_p,t)$) can then be extrapolated from k_e and \bar{E} . Tabulated k_e and \bar{E} values for all experiments are provided in Table S3. Only experiments with an inferred chamber electric field within the range verified by the control experiments (i.e., \bar{E} value < 15.7 V cm⁻¹) are reported. This consequently also removes experiments with abnormally high k_e values.

Experiment #	$k_e (\mathrm{s}^{-1})$	$\bar{E} (\mathrm{V}\mathrm{cm}^{-1})$				
Control Dry Experiments						
C1	0.05	5.1				
C2	0.10	8.9				
C3	0.06	7.0				
C4	0.21	15.7				
Control Humid	Experiment	S				
C5	0.07	7.9				
C6	0.02	5.8				
C7	0.11	11.8				
C8	0.15	15.3				
C9	0.31	20.9				
Experiments optimized for LV pathway						
D1	NA	NA				
D2	0.03	4.3				
D3	0.03	5.6				
D4	0.03	6.5				
D5	0.27	13.3				
D6	0.30	14.2				
D7	0.02	4.5				
D8	1.48	17.7				
D9	1.09	27.6				
D10	0.01	2.1				
D11	0	0				
Experiments of	otimized for	2MGA pathway				
M1	NA	NA				
M2	0.29	11.6				
M3	0.16	10.9				
M4	0.32	10.6				
M5	0.30	12.4				
M6	1.02	18.7				
M7	0.45	20.5				
M8	0.33	19.6				
M9	0	0				

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T_{1} L_{1} C_{2} C_{2} C_{1} C_{2} C_{1} C_{2} C_{2		1 1 (E) 1 1 1 1 1 CC C	$CC \cdot (I) C = II \cdot (I)$
Lable S3. Upfimized mean electro	Static field experienced within the d	r_{1} champer (F_{i}) and the eddy diffusion	(κ_{-}) for all experiments
indie bei optimized mean electro	studie neia experiencea wrann die s		coefficient (k_e) for all experiments.

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For experimental conditions, see Table 1 in main text.

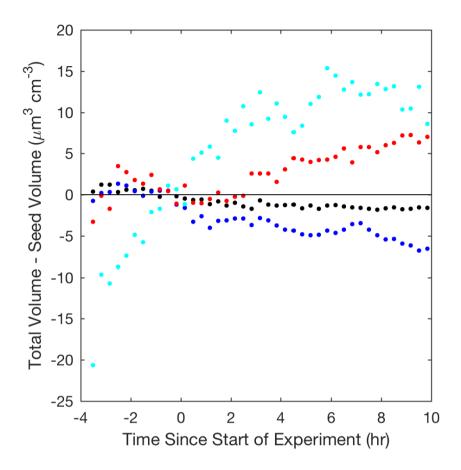


Figure S5. Total volume - seed volume (20 min averages) for the following particle wall deposition control experiments: C1 (V = 37 μ m³cm⁻³, •), C2 (109 μ m³cm⁻³, •), C3 (183 μ m³cm⁻³, •), and C4 (375 μ m³cm⁻³, •), respectively where V = initial corrected particle volume.

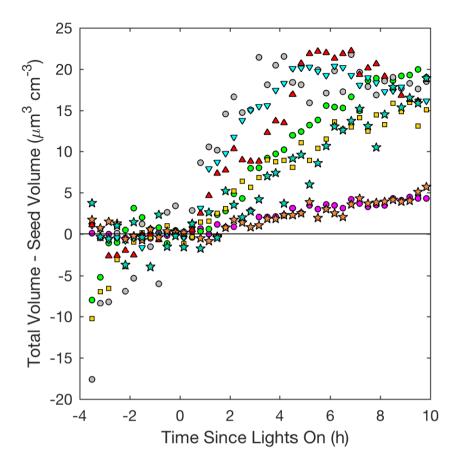


Figure S6. Total volume minus seed volume (20 min averages) as measured by DMA for all isoprene experiments with seed aerosol: seed surface area - D2 (SA = 1170 μ m² cm⁻³, •), D3 (SA = 3420 μ m² cm⁻³, •), & D4 (SA = 5770 μ m² cm⁻³, •), temperature - D5 (13 °C, \checkmark) and D6 (32 °C, \blacktriangle), isoprene loading - D7 (initial isoprene 110 μ g m⁻³, •), and new chamber with less wall charging - D10 (SA = 1580 μ m² cm⁻³, \bigstar) and D11 (SA = 4770 μ m² cm⁻³, \bigstar)

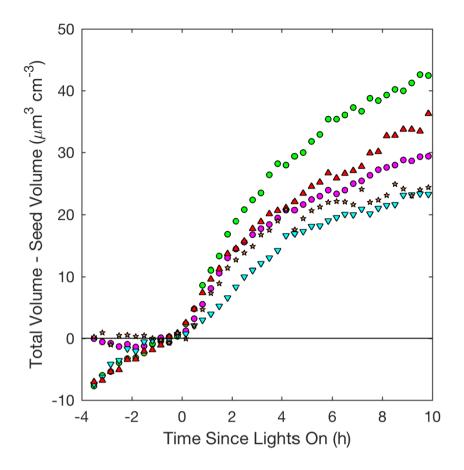


Figure S7. Total volume - seed volume (20 min averages) for all methacrolein experiments with seed aerosol: seed surface area - M2 (SA = 1640 μ m²cm⁻³, •), & M3 (SA = 2260 μ m²cm⁻³, •), temperature - M5 (13 °C, **v**) and M6 (32 °C, **A**), and new chamber with less wall charging - M9 (SA = 1910 μ m²cm⁻³, **★**).

S3 Additional Aerosol Composition Analysis from Aerosol Mass Spectrometer

The AMS results confirm that organic nitrates are present in the particle phase under dry conditions, but this technique is ill-suited for identifying specific organic nitrates present because $C_w H_x N_y O_z^+$ ions are produced in small yields (~5% of the nitrogen signal and <1% of the organic signal) and often occluded by more abundant peaks (Farmer et al., 2010). Organic

5 nitrates fragment with a distinctly higher NO⁺ / NO⁺₂ ratio than inorganic ammonium nitrate (Farmer et al., 2010). Considering ammonium nitrate fragments to an NO⁺ / NO⁺₂ ratio of 2.4 for the Caltech AMS, the NO⁺ / NO⁺₂ ratio of 4-8 (Figure S8) implies organic nitrates are present during all the LV and 2MGA pathway experiments performed under dry conditions. Under humid conditions, nitric acid partitions to the particle phase (Figure S8) and the NO⁺ / NO⁺₂ (~2) shifts closer to that of ammonium nitrate.

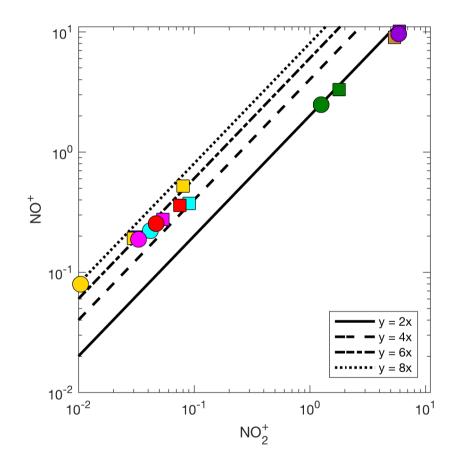


Figure S8. Average NO⁺ vs NO₂⁺ ratio from AMS during 10 h of photooxidation for LV pathway experiments (\odot) and 2MGA experiments (\Box) with colors indicating no initial seed aerosol (**blue**), seed aerosol (**magenta**), 13°C (**cyan**), 32°C (**red**), RH ~50% (**green**), RH ~70% (**brown**), RH ~80% (**purple**), new chamber with negligible wall charging (**gold**)

As explained in Section 4.1 of the main text, SOA mass yields measured by the DMA under humid conditions were not reported due to the need for more characterization of particle coagulation and wall loss under humid, high-NO, and high-nitric acid conditions. The AMS results can provide a qualitative understanding of the SOA mass formed under various conditions. Due to uncertainties in the collection efficiency (CE), the AMS results should not be used for quantification of SOA mass in

- 5 chamber experiments. Because humidity will enhance the CE (e.g., Docherty et al., 2013), for all humid experiments a Nafion dryer was used to dry the particles prior to AMS measurement. Changes in the CE due to differences in the organic composition between the experiments are possible. The AMS results are not corrected for particle wall loss. Additionally, as explained in Section 5.2 of the main text, an interference due to ammonium sulfate (Pieber et al., 2016) was subtracted from the organic signal. In general, this interference was higher and more variable in the humid experiments than the dry experiments.
- For the LV pathway experiments, increases in humidity increase the aerosol mass measured by the AMS (Figure S9a). Given the low collection efficiency for the LV pathway compared to that from the 2MGA pathway (Section 5.3 of the main text), this increase is likely explained by slight enhancements in the particle phase of compounds, to which the AMS is more sensitive such as glyoxal and methylglyoxal. Zhang et al. (2011) determined the isoprene high-NO SOA mass yield under dry conditions was ~ 2 times larger than that under humid conditions. Dommen et al. (2006) determined the isoprene SOA yield was not
- 15 dependent on RH from 2-85%; however, Zhang et al. (2011) reevaluated data from Dommen et al. (2006) and concluded that when comparing only experiments performed under similar conditions, the SOA mass yield under dry conditions is \sim 2 times greater than that formed under humid conditions. Here when comparing the AMS results for experiments M6 (RH = 47%) and M2 (RH = 8.9%), which have similar initial seed surface areas, SOA formation under dry conditions is \sim 1-2 times higher than under humid conditions depending on the time of oxidation (Figure S9b).
- In Figures S11-S14 and 9-10 in the main text, the average AMS mass spectra over the entire experiment (10h of photooxidation) is shown. The contribution of the highlighted fragments to the total tend to be fairly consistent over the entire 10h photooxidation period in both the LV and 2MGA pathway experiments.

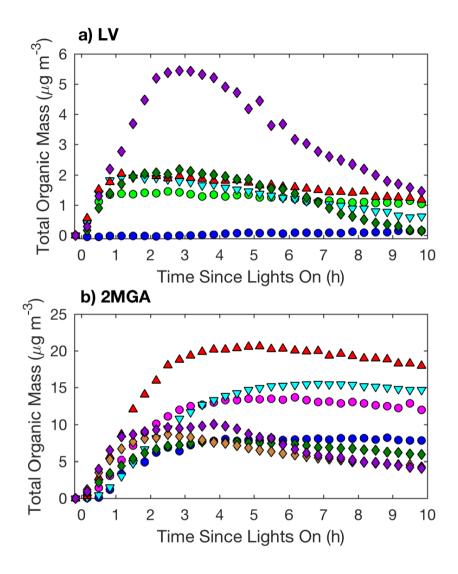


Figure S9. Total organic mass (20 minute averages) as measured by the AMS for LV pathway experiments (panel a): seed surface area - D1 (SA = 0 μ m² cm⁻³, •) & D3 (SA = 3420 μ m² cm⁻³, •), temperature - D5 (13 °C, \checkmark) & D6 (32 °C, \blacktriangle), and humidity - D8 (RH = 45%, \blacklozenge) & D9 (RH = 78%, \blacklozenge) and 2MGA pathway experiments (panel b): seed surface area - M1 (SA = 0 μ m² cm⁻³, •) & M2 (SA = 1640 μ m² cm⁻³, •), temperature - M4 (13 °C, \checkmark) & M5 (32 °C, \blacktriangle), and humidity - M6 (RH = 47%, \diamondsuit), M7(RH = 67%, \diamondsuit), & M8 (RH = 81%, \diamondsuit).

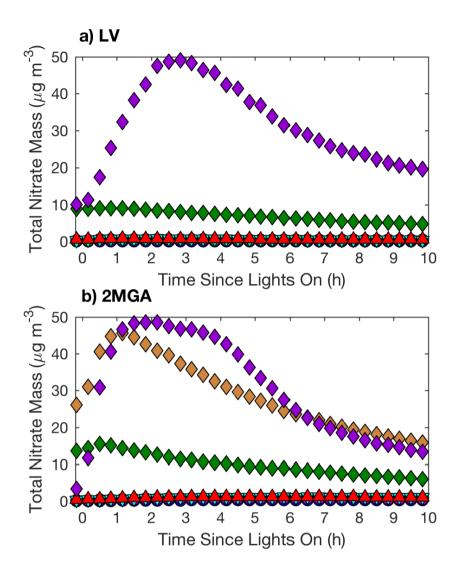


Figure S10. Total nitrate mass (20 minute averages) as measured by the AMS for LV pathway experiments (a): seed surface area - D1 (SA = 0 μ m² cm⁻³, •) & D3 (SA = 3420 μ m² cm⁻³, •), temperature - D5 (13 °C, •) & D6 (32 °C, •), and humidity - D8 (RH = 45%, •) & D9 (RH = 78%, •) and 2MGA pathway experiments (b): seed surface area - M1 (SA = 0 μ m² cm⁻³, • & M2 (SA = 1640 μ m² cm⁻³, •, temperature - M4 (13 °C, •) & M5 (32 °C, •), and humidity - M6 (RH = 47%, •), M7(RH = 67%, •), & M8 (RH = 81%, •).

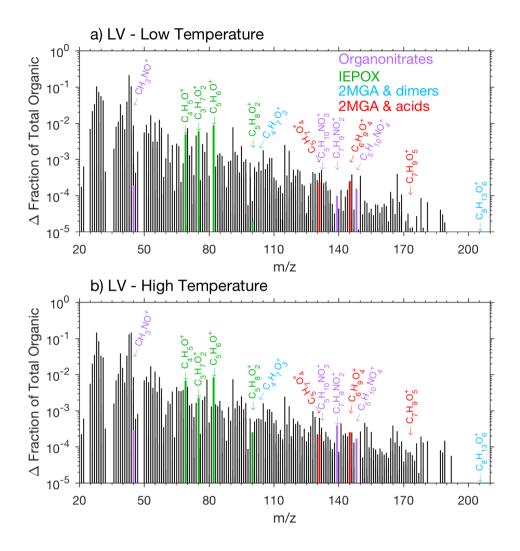


Figure S11. High resolution AMS mass spectra (averaged over 10h of photooxidation - the sulfate background) for experiment D5 (13 °C, panel a) and D6 (32 °C, panel b). Fragments are labeled as 2-MGA monomer/dimer (cyan), esterification of 2-MGA with acids (red), isoprene epoxydiol (IEPOX) tracers (dark green), and examples of organonitrate fragments - $C_x H_u NO_z$ (purple).

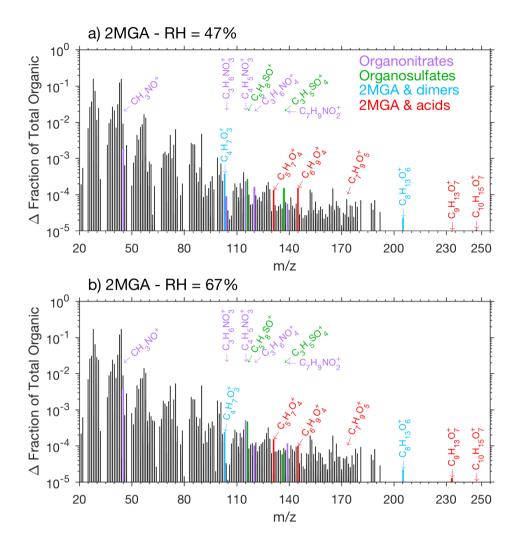


Figure S12. High resolution AMS mass spectra (averaged over 10h of photooxidation - the sulfate background) for experiment M6 (RH = 47%, panel a) and M7 (RH = 67%, panel b). Fragments are labeled as 2-MGA monomer/dimer (cyan), esterification of 2-MGA with acids (red), examples of organosulfate fragments (dark green), and examples of organonitrate fragments - $C_x H_y NO_z$ (purple).

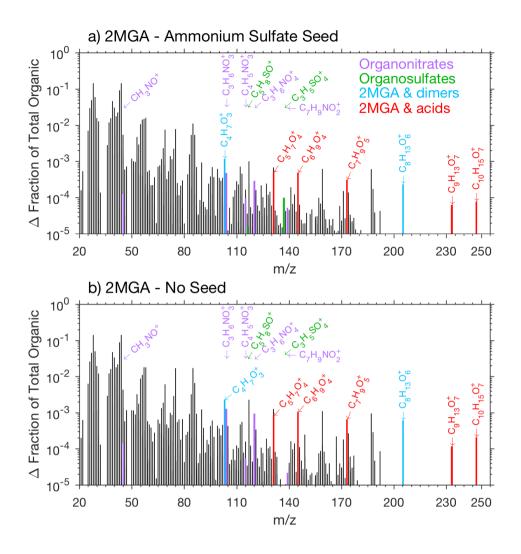


Figure S13. High resolution AMS mass spectra (averaged over 10h of photooxidation - the sulfate background) for experiment M2 (1640 μ m² cm⁻³, panel a) and AMS mass spectra (averaged over 10h of photooxidation) for experiment M1 (0 μ m² cm⁻³, panel b). Fragments are labeled as 2-MGA monomer/dimer (cyan), esterification of 2-MGA with acids (red), examples of organosulfate fragments (dark green), and examples of organonitrate fragments - C_xH_uNO_z (purple).

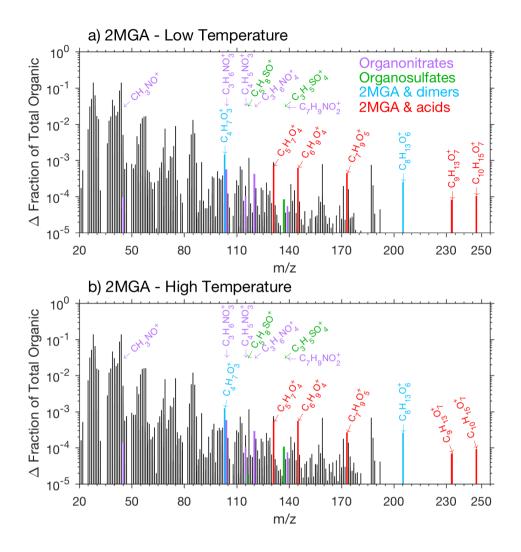


Figure S14. High resolution AMS mass spectra (averaged over 10h of photooxidation - the sulfate background) for experiment M4 (13 $^{\circ}$ C, panel a) and M5 (32 $^{\circ}$ C, panel b). Fragments are labeled as 2-MGA monomer/dimer (cyan), esterification of 2-MGA with acids (red), examples of organosulfate fragments (dark green), and examples of organonitrate fragments - C_xH_yNO_z (purple).

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