

Supplemental Information S4

Species-level predation network uncovers high prey specificity in a Neotropical army ant community

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Figure S1. Sampling effect in army ant-prey network. Influence of rarefaction of sampling effort on calculations of (A) network-level specialization H_2' and (B) modularity metric Q . The number of interactions has been reduced by randomly deleting one interaction per army ant species per deletion step. Black dot shows H_2' and Q of the original army ant-prey matrix, red dots the effect of down-sampling (50 randomizations per step).

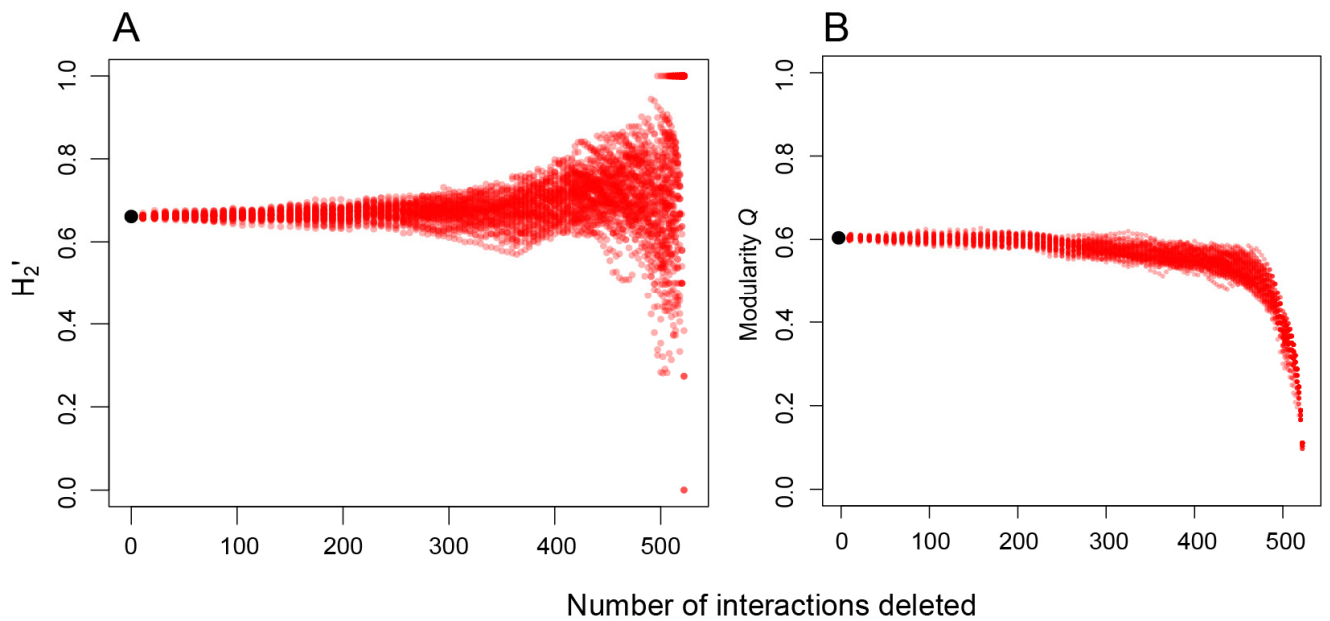


Figure S2. Collection events per time of the day. Histogram showing the number of collection events per hour of the day. Details about the time of the day for each collection event and species is given in Supporting information S2.

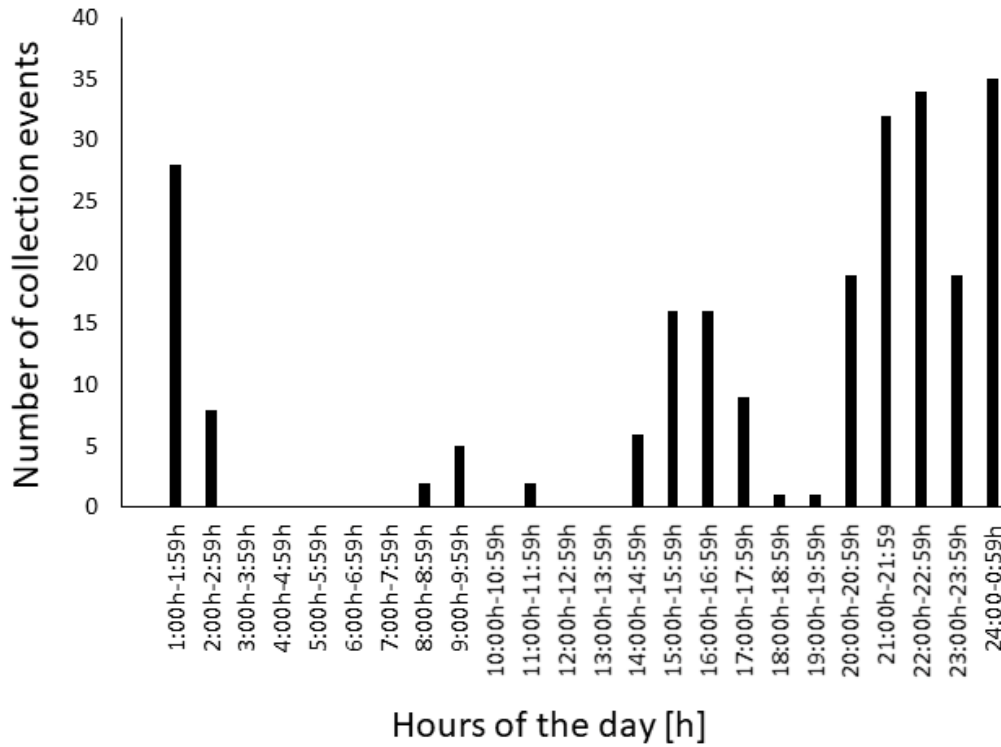


Figure S3. Prey composition. Composition of developmental stages of ant prey items. Different shades of gray depict adult, egg, larva and pupal stages. Total number of ant prey items per species is shown in parentheses.

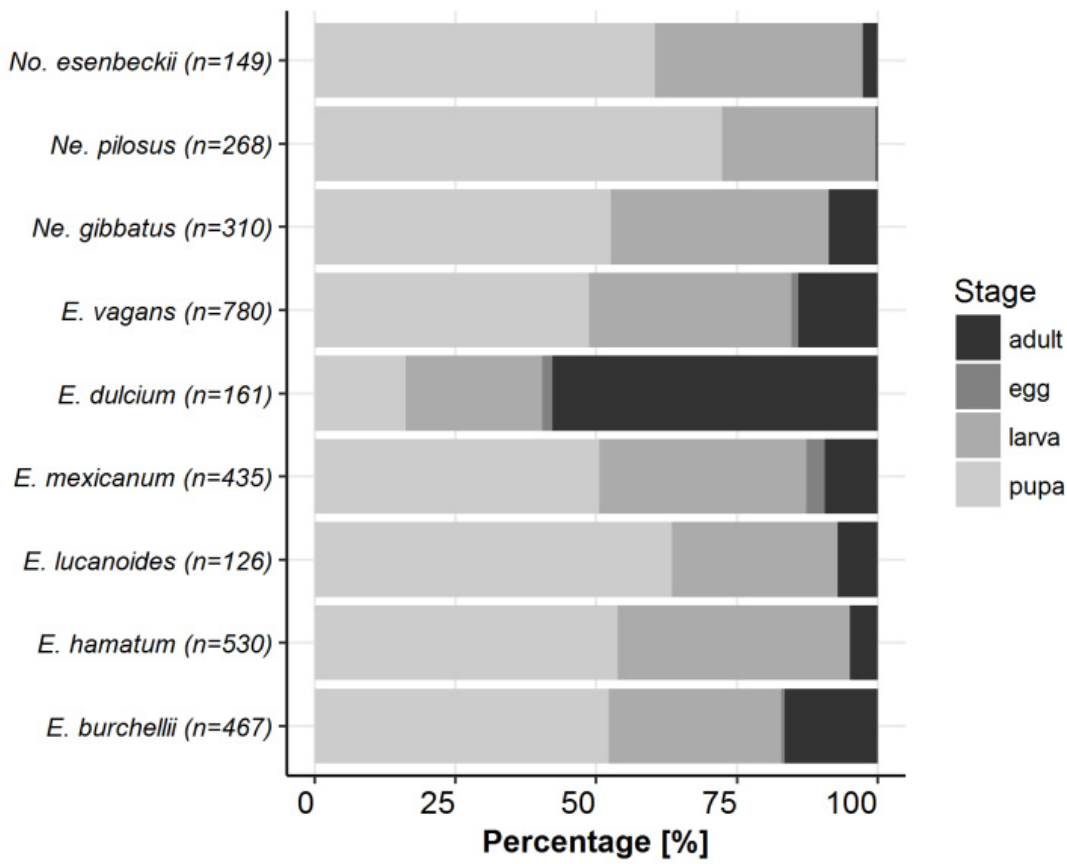


Figure S4. Distribution of *COI* barcode length fragments in base pairs.

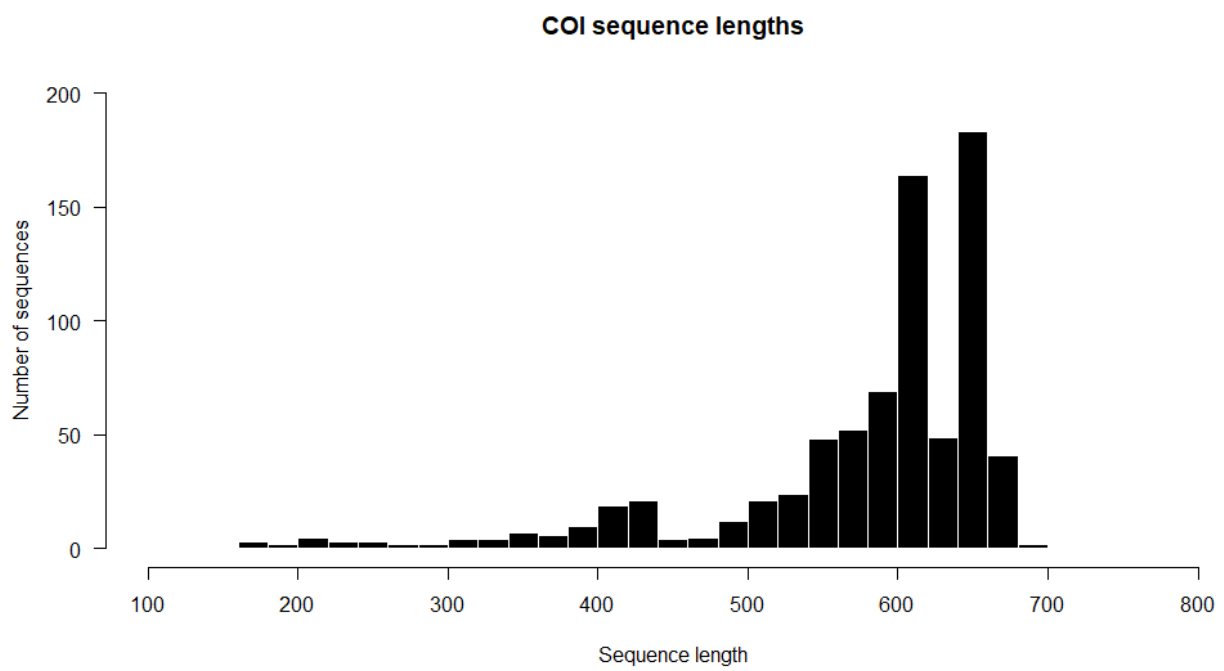


Figure S3. Possible intraguild predation in *Neivamyrmex*. We observed a case of possible intraguild predation at La Selva during the night of the 6th of April 2017 at 01:00h. *Neivamyrmex* cf. *iridescens* was swarming, seemingly uncoordinated, over a larger area. Workers carried their brood around and placed many of them in open brood caches. It first looked like an emigration to us, but less coordinated. Then we noted a second army ant species at the scene, *Neivamyrmex* cf. *swainsonii*. Smaller fights between both species were observed, and several *Ne. cf. swainsonii* workers carried away brood of *Ne. cf. iridescens*. Because we only observed this once, we cannot conclude with certainty that intraguild predation is a common behavior in this species. More detailed observations of the food habits in both species, *Neivamyrmex* cf. *swainsonii* and other *Neivamyrmex* species are urgently needed. Left: *Neivamyrmex* cf. *iridescens* with brood pile. Right: Fight of *Ne. cf. swainsonii* worker with *Ne. cf. iridescens* worker.



Table S1. *COI* forward and reverse primer combinations used in this study. Primer combinations with successful *COI* amplification are given for each ant genus. Primer combinations with most reliable *COI* amplification are marked with an asterisk and highlighted in bold. Annealing temperatures varied between 45°C and 55°C.

| Ant genus | Primer combinations (Forward primer /reverse primer) |
|-----------------------|---|
| <i>Acanthognathus</i> | LCO1490 / HCO2198 |
| <i>Acromyrmex</i> | LCO1532 / HCO2198* , LCO1532 / HCO1938, LCO1532 / AcromyR1 |
| <i>Anochetus</i> | dgLCO1490 / CrematoR1, LCO1490 / HCO2198* , LCO1490 / dgHCO2198, LepF1 / LepR1, MLepF1 / LepR1 |
| <i>Aphaenogaster</i> | LCO1490 / HCO2198 |
| <i>Apterostigma</i> | dgLCO1490 / CrematoR1, dgLCO1490 / dgHCO2198, LCO1490 / HCO2198* , LepF1 / LepR1 |
| <i>Azteca</i> | dgLCO1490 / AcromyR1, dgLCO1490 / CrematoR1, LCO1490 / HCO2198* |
| <i>Basiceros</i> | LepF1 / LepR1 |
| <i>Camponotus</i> | CampoF2 / CampoR2, CampoF2 / dgHCO2198* , CampoF2 / HCO2198, dgLCO1490 / AcromyR1, dgLCO1490 / CrematoR1, dgLCO1490 / dgHCO2198, LCO1490 / HCO2198* , LCO1490 / dgHCO2198, MLepF1 / LepR1 |
| <i>Crematogaster</i> | dgLCO1490 / CrematoR1* , dgLCO1490 / LepR1, LCO1490 / CrematoR1, LepF1 / CrematoR1, LCO1490 / HCO2198, LepF1 / LepR1 |
| <i>Cyphomyrmex</i> | dgLCO1490 / CrematoR1* , LCO1490 / HCO2198* , LepF1 / LepR1 |
| <i>Dolichoderus</i> | LCO1490 / HCO2198 |
| <i>Ectatomma</i> | LCO1490 / HCO2198* , LepF1 / LepR1, MLepF1 / LepR1 |
| <i>Gnamptogenys</i> | dgLCO1490 / CrematoR1, LCO1490 / dgHCO2198, LCO1490 / HCO2198* , LepF1 / LepR1, |
| <i>Hypoponera</i> | LCO1490 / HCO2198 |
| <i>Mayaponera</i> | dgLCO1490 / CrematoR1, LCO1490 / HCO2198* , LepF1 / LepR1 |
| <i>Megalomyrmex</i> | LCO1490 / HCO2198 |
| <i>Neoponera</i> | CampoF2 / dgHCO2198, dgLCO1490 / CrematoR1, LCO1490 / dgHCO2198, LCO1490 / HCO2198* , LepF1 / LepR1, MLepF1 / LepR1 |
| <i>Nylanderia</i> | CampoF2 / dgHCO2198, dgLCO1490 / CrematoR1* , LCO1490 / HCO2198* , LepF1 / LepR1 |
| <i>Odontomachus</i> | CampoF2 / dgHCO2198, dgLCO1490 / CrematoR1, LCO1490 / dgHCO2198, LCO1490 / HCO2198* , LepF1 / LepR1 |
| <i>Pachycondyla</i> | dgLCO1490 / CrematoR1, LCO1490 / dgHCO2198* , LCO1490 / HCO2198* , LepF1 / LepR1, MLepF1 / LepR1, |
| <i>Pheidole</i> | dgLCO1490 / AcromyR1, dgLCO1490 / CrematoR1, LCO1490 / HCO2198* , LCO1490 / dgHCO2198, LepF1 / LepR1, MLepF1 / LepR1, MLepF1 / dgHCO2198 |
| <i>Platythyrea</i> | LCO1490 / HCO2198* , LepF1 / LepR1, MLepF1 / LepR1 |
| <i>Sericomyrmex</i> | LepF1 / LepR1, MLepF1 / LepR1* |
| <i>Solenopsis</i> | dgLCO1490 / CrematoR1, LCO1490 / HCO2198 |
| <i>Strumigenys</i> | dgLCO1490 / CrematoR1* , MLepF1 / LepR1, LCO1490 / dgHCO2198, LCO1490 / HCO2198* |
| <i>Tapinoma</i> | LCO1490 / HCO2198 |
| <i>Trachymyrmex</i> | dgLCO1490 / CrematoR1, LCO1490 / dgHCO2198, LCO1490 / HCO2198* , MLepF1 / LepR1 |
| <i>Wasmannia</i> | dgLCO1490 / CrematoR1, LCO1490 / HCO2198* |

Table S2. COI primers used in this study.

| Primer name | Forward/ reverse primer | Sequence (5' –3') | Source |
|-------------|----------------------------|-------------------------------|-------------|
| AcromyR1 | Reverse | CTCCGGCAAGAACGRGRAGGGAAAGRA | This study |
| CampoF2 | Forward | TTTGCAATYTGATCWGGWATAATTGGATC | This study |
| CampoR2 | Reverse | CCYCCYCCBGAWGGRTCRAARAA | This study |
| CrematoR1 | Reverse | GGRTCTCCYCTCCDGMGGRTC | This study |
| dgLCO1490 | Forward | GGTCAACAAATCATAAAGAYATYGG | [1] |
| dgHCO2198 | Reverse | TAAACTTCAGGGTGACCAAARAAAYCA | [1] |
| LCO1490 | Forward | GGTCAACAAATCATAAAGATATTGG | [2] |
| LCO1532 | Forward | TCAGGTATAGTAGGATCTGCCA | C. Rabeling |
| HCO1938 | Reverse | GCACCAAGAATGGATGATATACCTGC | C. Rabeling |
| HCO2198 | Reverse | TAAACTTCAGGGTGACCAAAAAATCA | [2] |
| LepF1 | Forward | ATTCAACCAATCATAAAGATATTGG | [3, 4] |
| LepR1 | Reverse | TAAACTTCTGGATGTCCAAAAAATCA | [3, 4] |
| MLepF1 | Forward | GCTTCCCACGAATAAATAATA | [4] |

References

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4. Hajibabaei M, Janzen DH, Burns JM, Hallwachs W, Hebert PDN. DNA barcodes distinguish species of tropical Lepidoptera. *Proceedings of the National Academy of Sciences of the United States of America*. 2006;103:968–971.

Table S3. DNA barcode identification of non-ant prey. Non-ant barcodes were compared to the reference library of the 'Barcode of Life Data' System (BOLD) in September 2018. GenBank accession numbers of non-ant barcodes are given in Supporting information S2. We added several non-ant prey items that we obtained from a preliminary study about army ant diets at the same collection site (all sample IDs starting with the code 'cvb'; for details see Supporting information S2).

| Sample ID | Best hit in BOLD | Order | Family | Similarity to reference [%] | GenBank/BO LD number | Collected from... |
|---------------------|----------------------------------|-------------------|--------------------|-----------------------------|----------------------|--------------------------|
| PH_EB58_ca_1_02 | <i>Metapolybia cingulata</i> | Hymenoptera | Vespidae | 92.59 | GU596904 | <i>Eciton burchellii</i> |
| PH_EB58_undet_np_03 | <i>Synoeca septentrionalis</i> | Hymenoptera | Vespidae | 92.76 | Early release | <i>Eciton burchellii</i> |
| PH_EB64_nonant_1_05 | <i>Scolopocryptops mexicanus</i> | Scolopendromorpha | Scolopocryptopidae | 84.56 | JX422679 | <i>Eciton burchellii</i> |
| cvb777prey022 | <i>Digitops indicus</i> | Scolopendromorpha | Scolopendridae | 84.26 | JX531857 | <i>Eciton burchellii</i> |
| cvb777prey006 | <i>Rhiginia cinctiventris</i> | Hemiptera | Reduviidae | 99.37 | CNCHB1508 | <i>Eciton burchellii</i> |
| cvb777prey015 | <i>Labiduridae</i> | Dermaptera | Labiduridae | 87.93 | Private | <i>Eciton burchellii</i> |
| cvb709prey003 | Blattodea | Blattodea | n.a. | 83.75 | Private | <i>Eciton burchellii</i> |
| cvb709prey001 | <i>Eublaberus posticus</i> | Blattodea | Blaberidae | 91.39 | MF136388 | <i>Eciton burchellii</i> |
| cvb708prey002 | Ectobiidae | Blattodea | Ectobiidae | 99.66 | Private | <i>Eciton burchellii</i> |
| cvb777prey013 | Scarabaeidae | Coleoptera | Scarabaeidae | 90.10 | Private | <i>Eciton burchellii</i> |
| cvb777prey014 | Carabidae | Coleoptera | Carabidae | 100.00 | Private | <i>Eciton burchellii</i> |
| cvb777prey004 | Fulgoridae | Hemiptera | Fulgoridae | 100.00 | Private | <i>Eciton burchellii</i> |
| cvb595prey002 | <i>Panchlora</i> sp. CC-2017 | Blattodea | Blaberidae | 98.31 | KY741983 | <i>Eciton dulcium</i> |
| PH_EH60_undet_1_02 | Phoridae | Diptera | Phoridae | 98.63 | Early release | <i>Eciton hamatum</i> |
| cvb011prey004 | <i>Ugyops</i> | Hemiptera | Delphacidae | 79.47 | Early release | <i>Eciton hamatum</i> |
| cvb400prey001 | <i>Protopolybia</i> | Hymenoptera | Vespidae | 89.09 | AY918914 | <i>Eciton hamatum</i> |
| PH_EL34m_ca_1_01 | <i>Synoeca septentrionalis</i> | Hymenoptera | Vespidae | 93.37 | Early release | <i>Eciton lucanoides</i> |
| PH_EM49_ec_1_03 | <i>Metapolybia cingulata</i> | Hymenoptera | Vespidae | 92.79 | GU596904 | <i>Eciton mexicanum</i> |