

In the format provided by the authors and unedited.

Sequential formation and resolution of multiple rosettes drive embryo remodelling after implantation

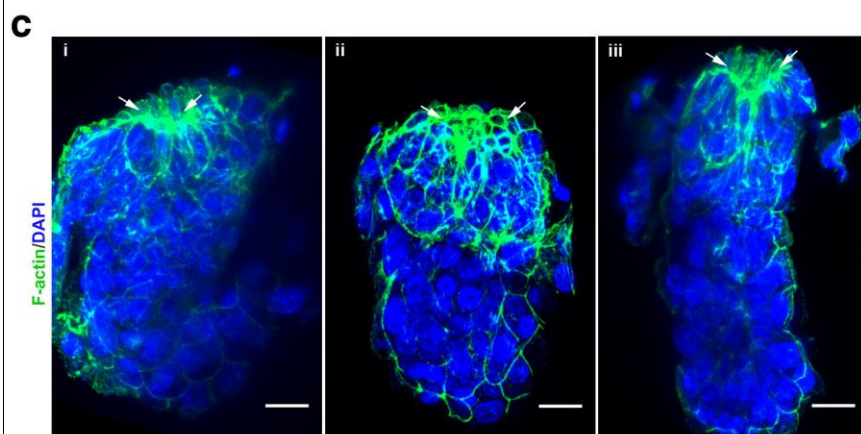
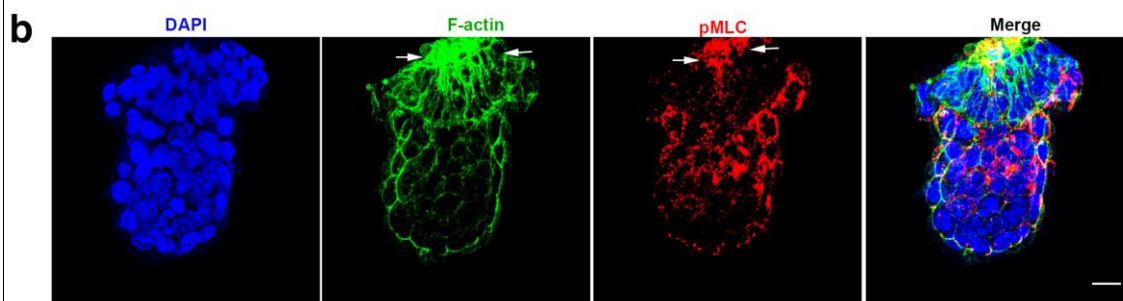
Neophytos Christodoulou^{1,4}, Christos Kyprianou^{1,4}, Antonia Weberling¹, Ran Wang², Guizhong Cui²,
Guangdun Peng², Naihe Jing^{2,3} and Magdalena Zernicka-Goetz^{1*}

¹Mammalian Embryo and Stem Cell Group, Department of Physiology, Development and Neuroscience, University of Cambridge, Cambridge, UK.

²State Key Laboratory of Cell Biology, CAS Center for Excellence in Molecular Cell Science, Shanghai Institute of Biochemistry and Cell Biology, Chinese Academy of Sciences, University of Chinese Academy of Sciences, Shanghai, China. ³School of Life Science and Technology, ShanghaiTech University,

Shanghai, China. ⁴These authors contributed equally: Neophytos Christodoulou, Christos Kyprianou. *e-mail: mz205@cam.ac.uk

a E5.0 decidua recovery → 30 min 4% PFA → embryo recovery

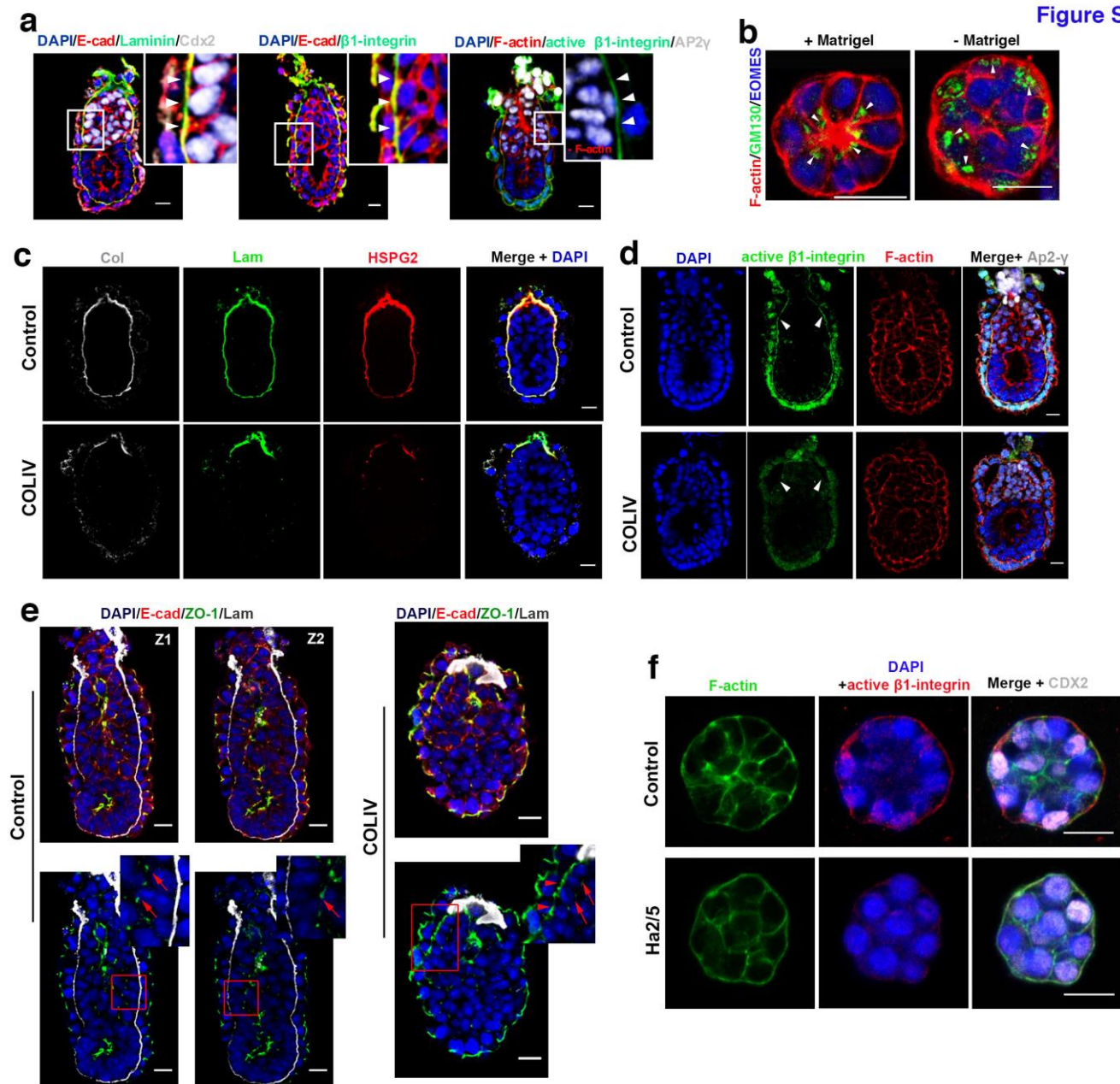


Supplementary Figure 1

Analysis of extra-embryonic ectoderm tissue folding in embryos derived after decidua fixation

(a) Experimental strategy for post-fixation embryo recovery. (b) Representative E5.0 recovered from a fixed decidua. Accumulation of actin and phosphorylated myosin (arrows) is evident at the proximal tip of extra-embryonic ectoderm cells in recovered embryos post-fixation. This indicates that apical actin accumulation and apical constriction tissue folding (see Fig. 1) is an active morphogenetic process and not a wound healing response stemming from the recovery of embryos from deciduae. (c) 3 representative examples of E5.0 embryos recovered from fixed deciduae. Arrows indicate actin apical accumulation at extra-embryonic ectoderm most proximal cells. Image represents 3 biological replicates. Scale bars= 20um.

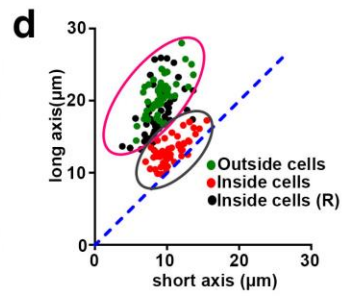
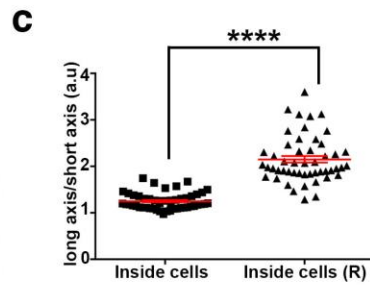
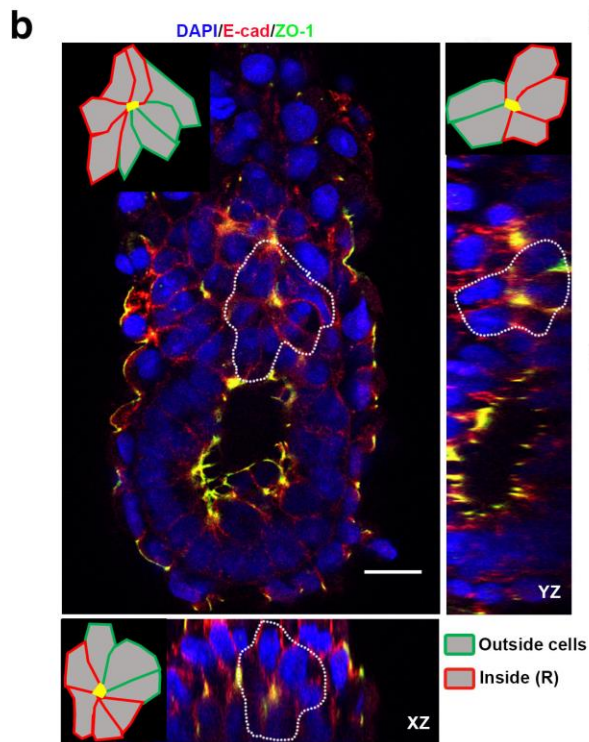
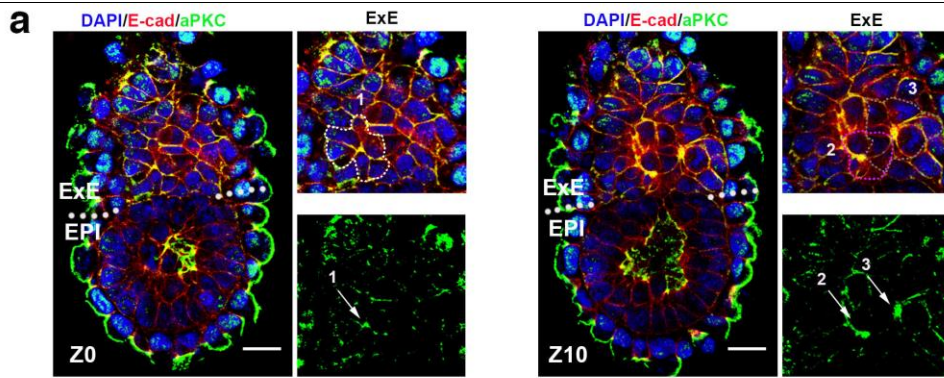
Figure S2



Supplementary Figure 2

ECM/ β 1-integrin during extra-embryonic ectoderm and TSCs polarization

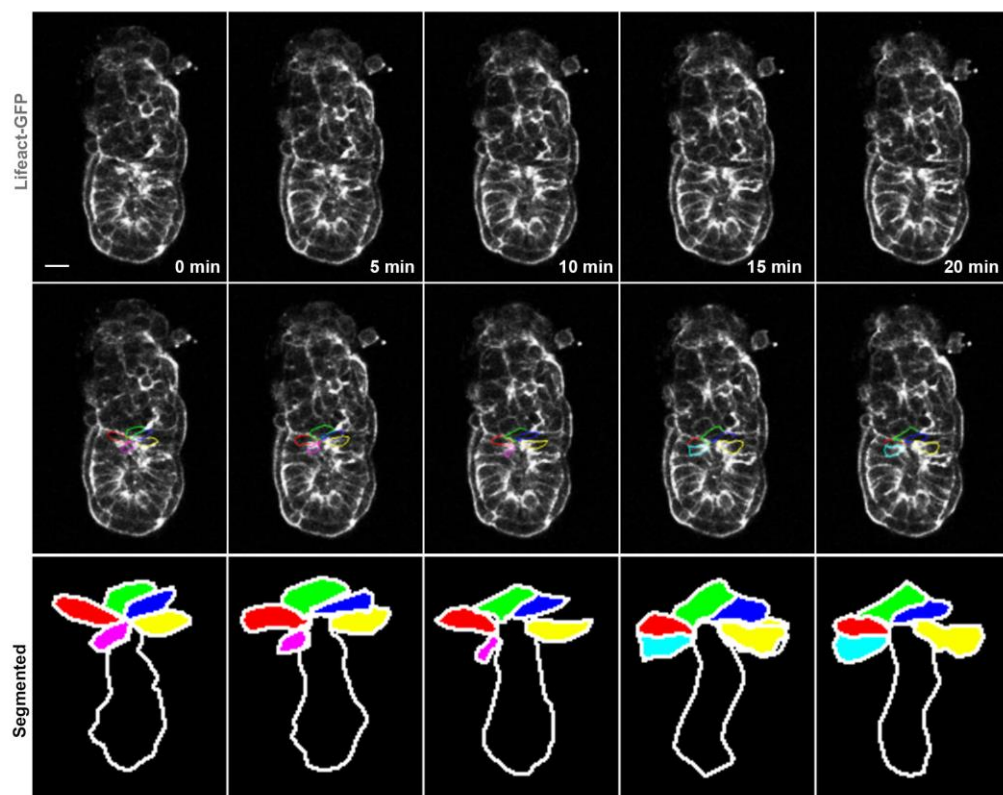
(a) ECM/ β 1-integrin localization in E5.5 embryo revealed by antibodies for laminin, β 1-integrin and active β 1-integrin (9EG7). Image representative of 10 embryos (b) Representative examples of TSC clumps culture in suspension in the presence or absence of Matrigel. In the absence of Matrigel TSCs fail to polarize. Specifically the cells fail to acquire columnar morphology (indicated by F-actin staining) and they don't display apical Golgi polarization (GM130 staining; arrowheads) compared with TSCs cultured in the presence of Matrigel. Image representative of 3 biological replicates (c) The effect of collagenase IV (COLIV) treatment on the surrounding basement membrane (assessed by laminin, collagen and perlecan staining) of E5.5 embryos cultured for 5h in the presence or absence of COLIV. Image representative of 5 biological replicates (d) β 1-integrin activation status (indicated by 9EG7 antibody staining) in control and COLIV treated embryos. White arrowheads indicate basal site of ExE outside cells. Image representative of 2 biological replicates (e) The effect of COLIV treatment on the localization of the tight junction's localisation assessed by ZO-1 staining in E5.5 embryos (two different z slices in control embryo; Z1 and Z2). Arrows point to apical site while arrowheads point towards the basal site. Image representative of 10 embryos (f) β 1-integrin activation status (indicated by 9EG7 antibody staining) in control and β 1-integrin blocking antibody (Ha2/5) treated TSCs in 3D culture; Image representative of 2 biological replicates. Scale bars=20um



Supplementary Figure 3

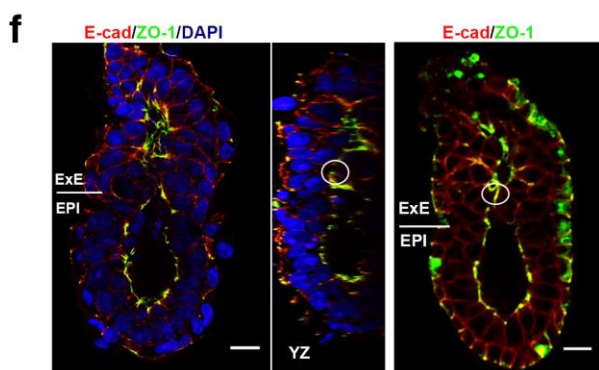
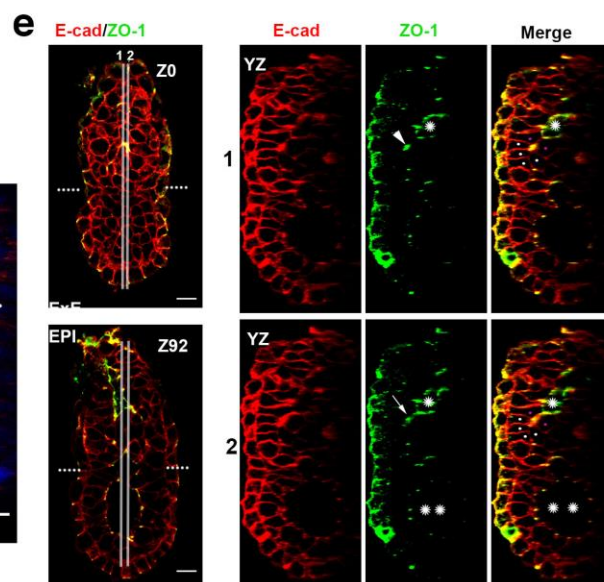
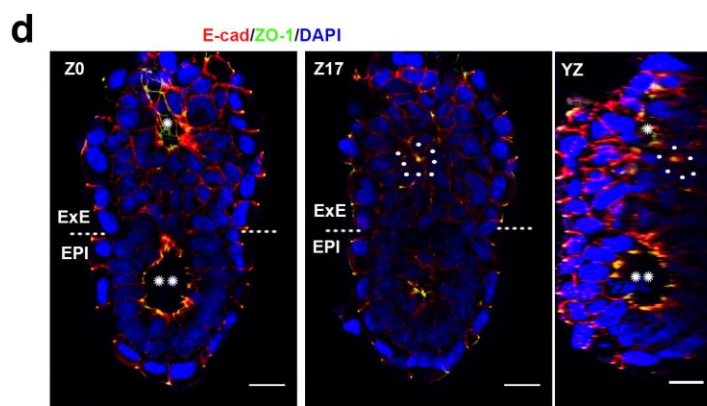
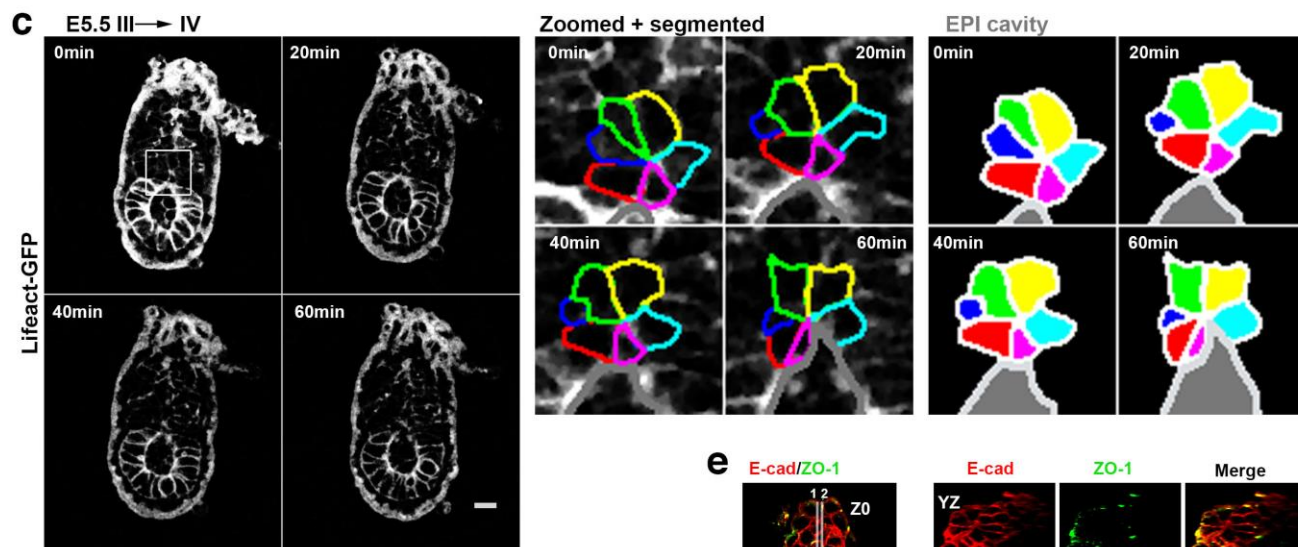
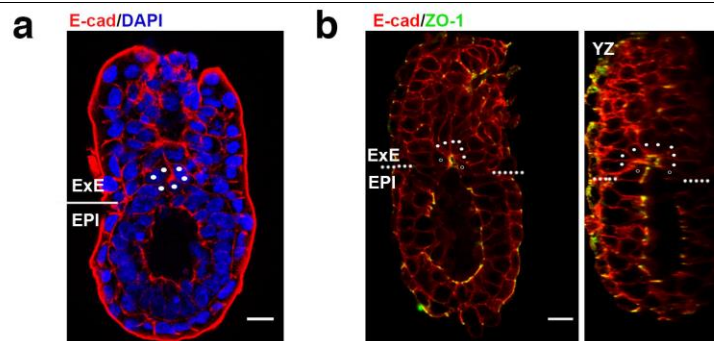
Characterization of extra-embryonic ectoderm cells

(a) Images of different Z slices (0.8 μm) from a representative E5.5 (Stage III) embryo. Extra-embryonic ectoderm (ExE) rosettes are outlined with a dashed line. Arrows denote rosettes' centre. Image representative of 20 embryos (b) Representative example of an ExE rosette (dashed outline) found in E5.5 embryo as revealed by cell morphology (E-cadherin) and polarization (ZO-1). Orthogonal views (XZ, YZ) of the embryo are displayed on the right and the bottom of the image with the segmented rosette as viewed in the respective orthoslice. Image representative of 20 embryos. (c) Cell aspect ratio comparison of outside, inside vs inside cells that contribute to rosettes (Inside cells, R). Two sided unpaired student t-test; **** $p < 0.0001$; mean \pm SEM; $n = 50$ inside cells and $n = 47$ inside rosette cells. (d) Plot of long and short axis. Two separate clusters are indicated (magenta and grey outlines). Cluster shown with magenta consists of outside cells and inside cells contributing to rosettes (inside cells R). Cluster shown with grey consists of inside cells that do not contribute to rosettes. Blue dashed line: $y = x$. $n = 50$ outside and inside cells and $n = 47$ inside rosette cells. Scale bars = 20 μm



Supplementary Figure 4**Epiblast remodelling upon hybrid rosette resolution**

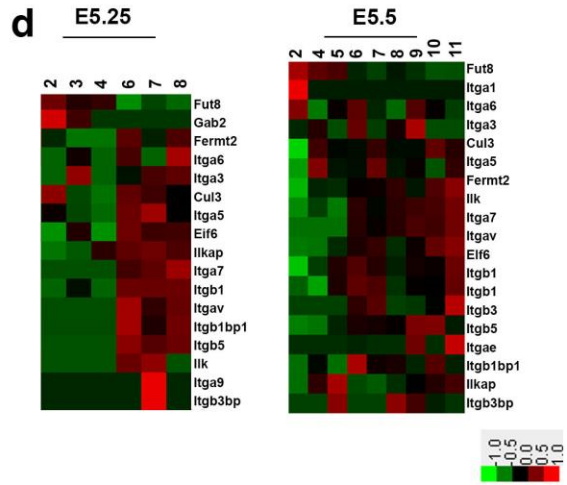
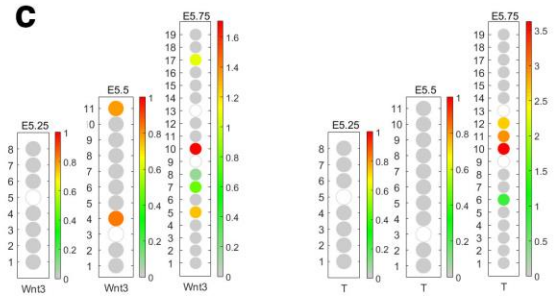
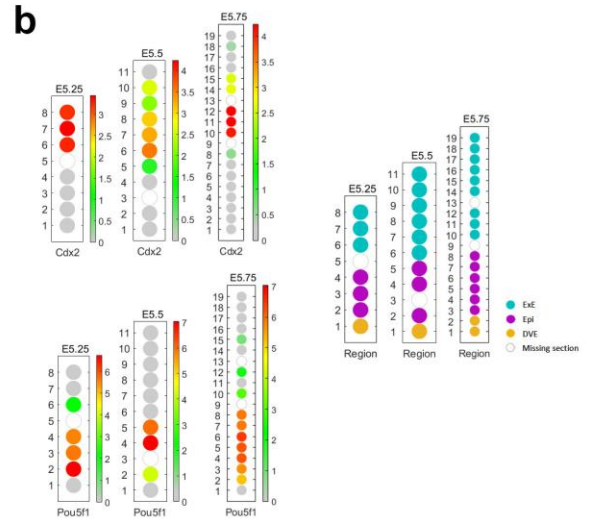
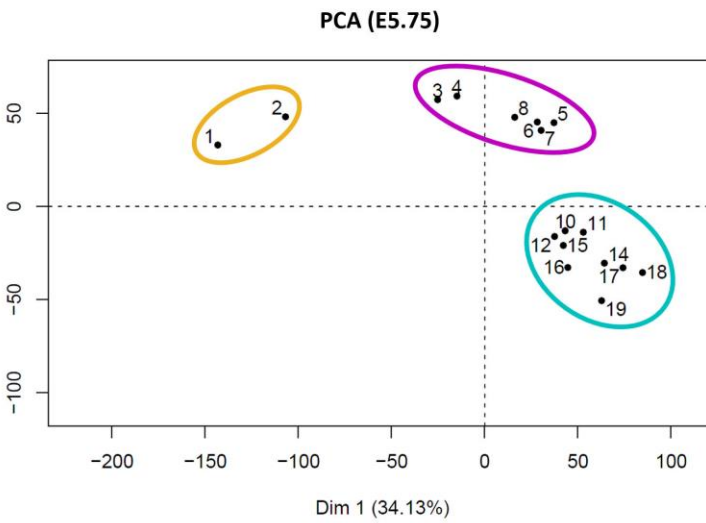
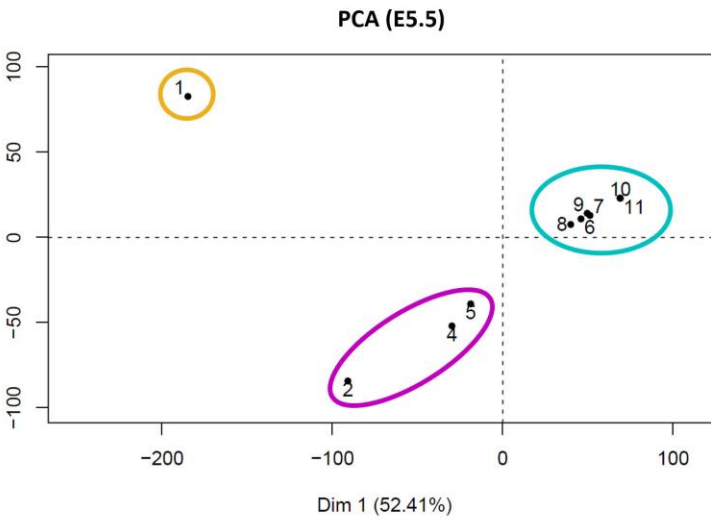
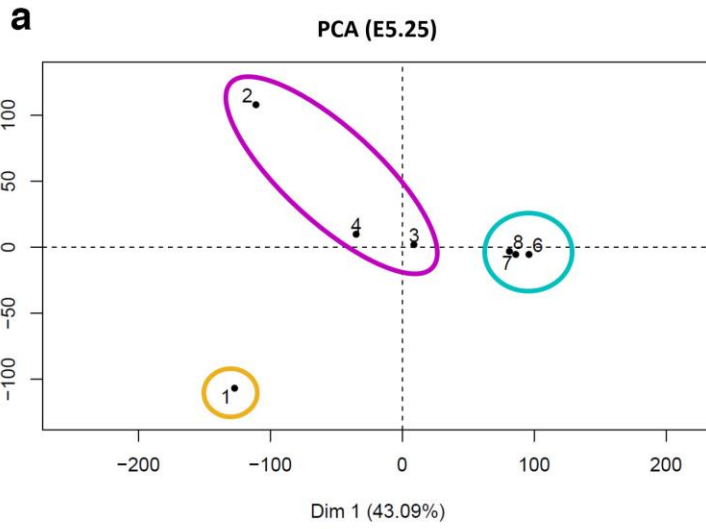
Stills from a time lapse movie of a LifeAct-GFP embryo showing epiblast EPI remodelling after hybrid rosette resolution. Cells are arbitrarily colour-coded for tracking. EPI: purple, yellow and cyan cells; ExE: blue, red and green cells. Image representative of 3 embryos. Scale bar=20um



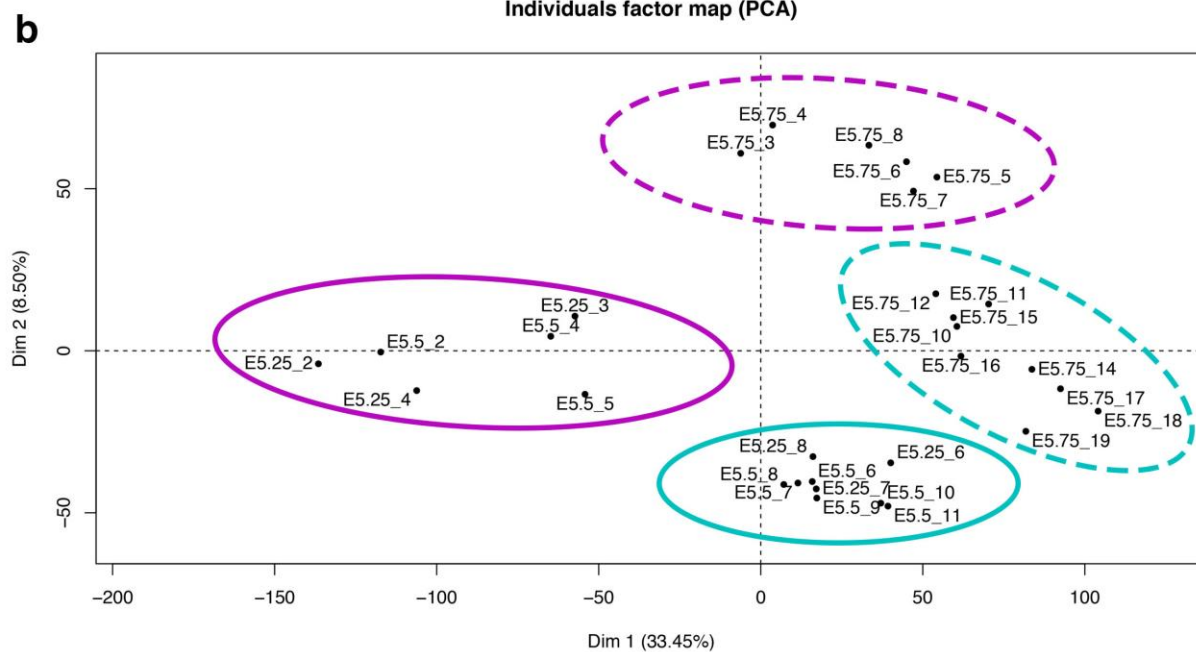
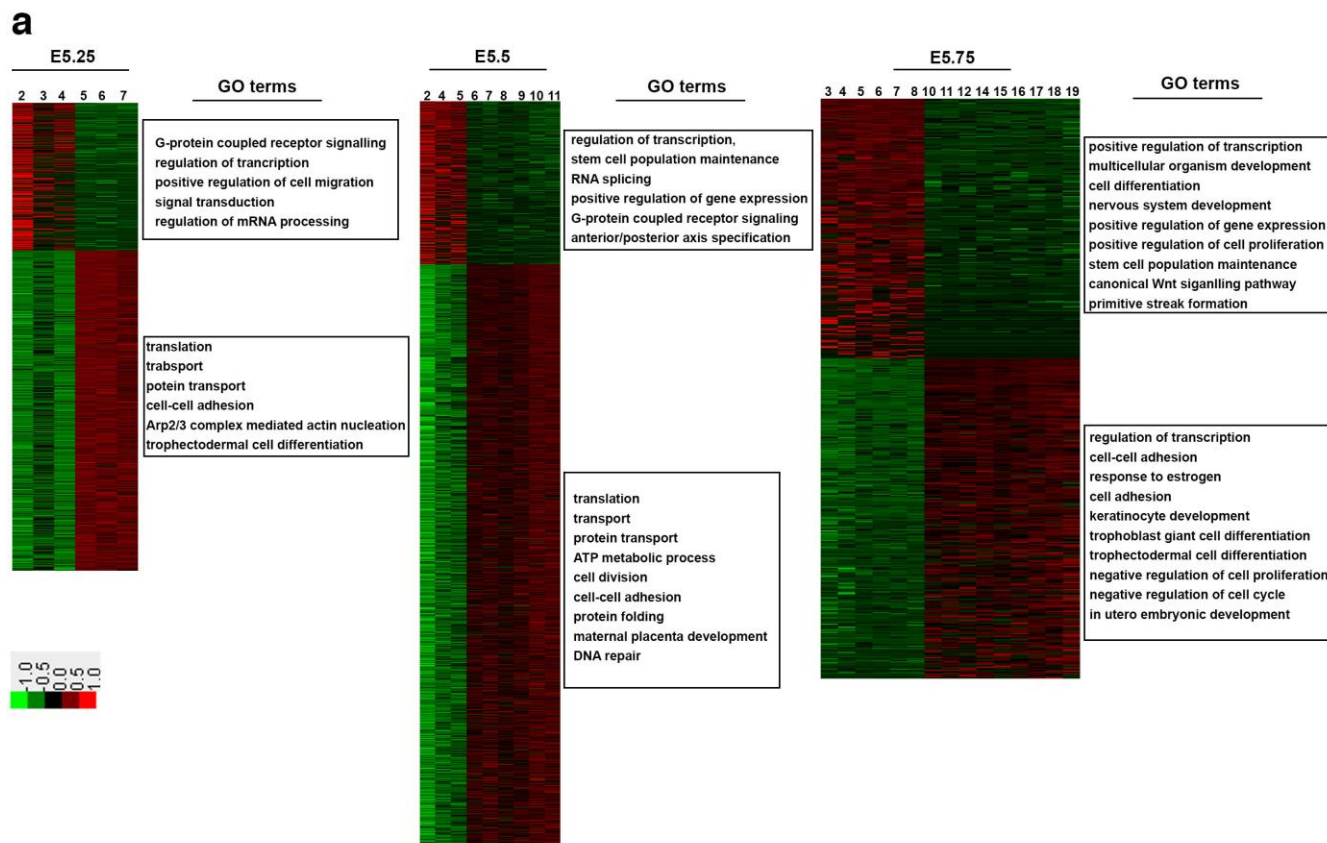
Supplementary Figure 5

Extra-embryonic ectoderm rosettes facilitate cavities extension and fusion

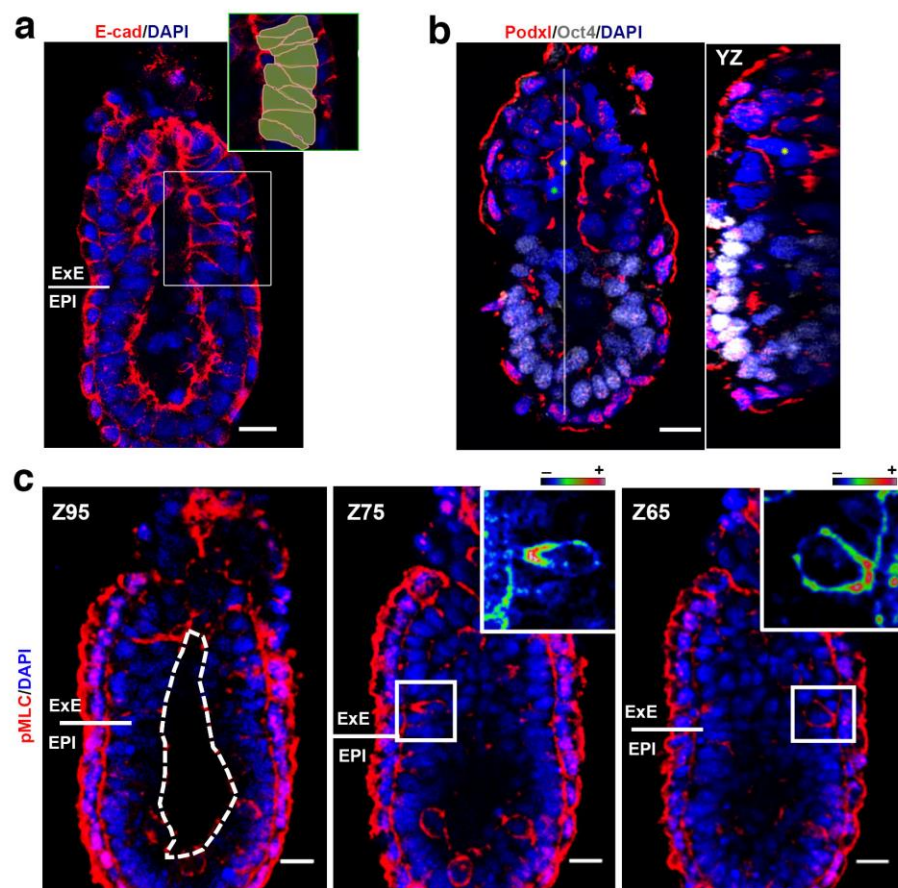
(a) Representative example of E5.5 (stage III) embryo stained for E-Cadherin. White dots label the cells contributing to the epiblast (EPI) cavity's proximal rosette (adjacent to EPI-ExE boundary). Image representative of 15 embryos. (b) Example of a resolving EPI cavity's proximal rosette (dots show cells contributing to the rosette) through loss of cell-cell contacts (cells marked with hollow dots) of ExE cells at the boundary. YZ projection is displayed on the right of the image. Image representative of 10 embryos (c) Stills from a time lapse movie of an E5.5 LifeAct-GFP embryo showing EPI cavity expansion after EPI cavity's proximal rosette resolution. Cells are arbitrarily color-coded for tracking. Image representative of 3 embryos (d) E5.5 (stage III) embryo stained for E-Cadherin and tight junction protein ZO-1. Two different z slices are presented (Z0 and Z17, Z step=0.8 μ m) one showing the ExE cavity and the other the overlaying rosette (white dots). YZ projection of rosette (white dots showing cells contributing to rosette) and ExE cavities shown on the far right of the panel. *=ExE cavity, **=EPI cavity. Image representative of 10 embryos. (e) Representative example of ExE cavity (asterisk in YZ projections, right panel) connected with the centre of an ExE cavity proximal rosette (arrowhead points to the centre of this rosette; magenta dots indicate cells contributing to both rosette and ExE cavity where white dots indicate the rest of rosette's cells) through a tract of ZO-1 (arrow). White dotted line = EPI-ExE boundary. *=ExE cavity, **=EPI cavity. Image representative of 10 embryos. (f) Two representative examples of an invading EPI cavity and an extended ExE cavity connected with a polarized tract of ZO-1 (white circle). Image representative of 6 embryos. Scale bars = 20 μ m



Supplementary Figure 6
Spatial transcriptome analysis of E5.25-E5.75 embryos
(a) Principal component analysis (PCA) of sections along the proximo-distal embryo axis from at E5.25, E5.5 and E5.75. Yellow outline: VE, Purple outline: epiblast (EPI), Cyan outline: extra-embryonic ectoderm (ExE). (b) Gene expression pattern for EPI (Pou5f1) and ExE (CDX2) markers from E5.25-E5.75. (c) Gene expression pattern for Wnt3 and T from E5.25-E5.75. (d) Differential gene expression analysis heat maps for GO term: integrin mediated signalling. E5.25: sections 2-4 EPI, sections 6-8 ExE ; E5.5: sections 2,4,5 EPI, sections 6-11 ExE.



Supplementary Figure 7
Differential gene expression analysis of embryonic and extra-embryonic compartments
(a)Differential gene expression analysis heat maps between epiblast (EPI) and extra-embryonic ectoderm (ExE) in E5.25, E5.5 and E5.75 embryos. Representative gene ontology (GO) terms enriched in each tissue are shown on the right of each heat map. E5.25: sections 2-4 EPI, sections 6-8 ExE; E5.5: sections 2,4,5 EPI, sections 6-11 ExE; E5.75 sections 3-8 EPI, sections 9-19 ExE. n=1 E5.25, n=1 E5.5 and n=1 E5.75 embryos (b) Combined principal component analysis (PCA) for embryonic and extra-embryonic sections from E5.25-E5.75. Purple outline: E5.25-E5.5 EPI; Cyan outline: E5.25-E5.5 ExE; Dashed purple outline: E5.75 EPI; Dashed cyan outline: E5.75 ExE. n=1 E5.25, n=1 E5.5 and n=1 E5.75 embryos



Supplementary Figure 8**Cell intercalation after initial cavities fusion results in the formation of a pseudostratified epithelium**

(a)ExE acquires a pseudostratified epithelium morphology shortly after unification of the cavities. Image representative of 10 embryos. (b)After cavities fusion ExE cells show intercalative behaviour (green and yellow asterisks). Image representative of 5 embryos. (c) 3 different optical Z slices (0.6um) of a representative example of an E5.75(stage IV) embryo. Dashed outline in left panel shows pro-amniotic cavity spanning through EPI and ExE. In different Z slices (middle and right panel) ExE cells intercalating towards the basement membrane can be identified. Polarized pMLC localization displays the polarized nature of this event. Insets in middle and right panel show magnified fluorescent intensity color coded images of ExE intercalating cells. Image representative of 5 embryos. Scale bars=20um

Supplementary Table 1: Processed RNA-seq data.**Supplementary Table 2: Statistics source data****Supplementary Movie 1. Tissue folding mediated extra-embryonic cavity formation**

Time lapse movie (2D view) of a representative E5.0 Lifeact-GFP transgenic embryo showing tissue bending mediated extra-embryonic cavity formation. Representative of 5 separate time-lapse movies Time interval=5min.

Supplementary Movie 2. Tissue folding mediated extra-embryonic cavity formation in 3D

Top 3D view of Movie S1 showing apical constriction mediated cell ingression during extra-embryonic ectoderm cavity formation. Time interval=5min Representative of 5 separate time-lapse movies.

Supplementary Movie 3. Apical constriction during extra-embryonic ectoderm tissue folding

Time lapse movie showing two different single cells from movie S2. Note the increase of medioapical actin before apical cell constriction. Time interval=5 min.

Supplementary Movie 4. 3D segmentation of representative extra-embryonic ectoderm rosettes

3D segmentation of a representative extra-embryonic ectoderm rosette. Each rosette cell is represented with different colour. White circle highlights the rosette center.

Supplementary Movie 5. 3D segmentation of neighboring extra-embryonic ectoderm rosettes

3D segmentation of a representative extra-embryonic ectoderm rosettes sharing cells. Each cell is represented with different colour.

Supplementary Movie 6. Hybrid rosette resolution precedes epiblast remodelling(1).

Time lapse movie of a representative E5.5 Lifeact-GFP transgenic embryo showing epiblast remodelling after hybrid rosette resolution. Time interval=20 min. Representative of 3 separate time-lapse movies

Supplementary Movie 7. Hybrid rosette resolution precedes epiblast remodelling(2)

Time lapse movie of a representative E5.5 Lifeact-GFP transgenic embryo showing epiblast remodelling after hybrid rosette resolution. Time interval=5 min. Representative of 3 separate time-lapse movies

Supplementary Movie 8. Laser ablation mediated hybrid rosette resolution.

Time lapse movie of a representative E5.5 Lifeact-GFP (stage II) transgenic embryo. The cell-cell interface connecting the hybrid rosette centre with the epiblast cavity was ablated via laser ablation (red bar). After ablation, the epiblast cavity has extended towards the centre of hybrid rosette, Time interval=1 min. Representative of 2 biological replicates.

Supplementary Movie 9. Polarized tract connects the centre of a rosette with the embryonic cavity.

Representative images through Z ($Z=0.9\mu\text{m}$) of the boundary between epiblast and extra-embryonic ectoderm of a representative E5.5 embryo. Polarized track extending from the EPI cavity towards the centre of an ExE rosette is evident. blue: DAPI, red:E-cad, green: ZO-1. White dots indicate ExE rosette cells. n=10 embryos

Supplementary Movie 10. ExE rosettes facilitate epiblast cavity extension(1).

Time lapse movie of a representative E5.5 mTmG transgenic embryo showing epiblast cavity expansion towards the ExE after ExE rosette resolution. Time interval=20 min. Representative of 3 separate time-lapse movies.

Supplementary Movie 11. ExE rosettes facilitate epiblast cavity extension(2).

Time lapse movie of a representative E5.5 LifeactGFP transgenic embryo showing epiblast cavity expansion towards the ExE after ExE rosette resolution. Time interval=20 min. Representative of 3 separate time-lapse movies

Supplementary Movie 12. ExE rosettes facilitate extra-embryonic cavity extension.

Time lapse movie of a representative E5.5 LifeactGFP transgenic embryo showing extra-embryonic ectoderm cavity extension towards the EPI after ExE rosette resolution. Time interval=5 min. Representative of 3 separate time-lapse movies.

Supplementary Movie 13. Rosette mediated embryonic and extra-embryonic cavities extension.

Time lapse movie of a representative E5.5 LifeactGFP transgenic embryo showing embryonic and extra-embryonic cavities extension after rosette resolution. Green outlines indicate embryonic (bottom) and extra-embryonic cavities (top). Yellow dots: cells contributing to resolving rosette during extra-embryonic cavity extension; Green dots: cells contributing to resolving rosette during embryonic cavity extension; Time interval=5 min. Representative of 2 separate time-lapse movies.

Supplementary Movie 14. Extra-embryonic ectoderm cell intercalation upon initial cavities fusion.

Time lapse movie of a representative E5.75 mTmG (stage IV → V) transgenic embryo showing ExE cell intercalation after embryonic (bottom) and extra-embryonic(top) cavities fusion. magenta: cavities, green: Intercalating ExE cell. Time interval=20 min. Representative of 2 separate time-lapse movies.