| 1 | Supporting Information for |
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| 2 | Response of the Aerodyne Aerosol Mass Spectrometer to Inorganic Sulfates and |
| 3 | Organosulfur Compounds: Applications in Field and Laboratory Measurements |
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28 S1. Details of Chamber Experiment and Field Measurements

29 S1.1. Chamber Experiment

30 Isoprene photooxidation SOA under low-NO condition were generated in the Georgia Tech Environmental Chamber (GTEC) facility. Details of the facility have been described in Boyd et 31 al.¹ and details of the experiment have been described in Tuet et al..² Briefly, the experiment was 32 performed at 25 °C under dry (RH < 5 %) condition. Prior to the experiment, the chamber was 33 flushed with zero air for ~24 h. Seed aerosol was first introduced by atomizing 15 mM AS solution 34 and seed volume concentration was stabilized at ~ $25 \,\mu m^3/cm^3$. Aerosol volume concentrations 35 and distributions were measured using a scanning mobility particle sizer (SMPS; TSI) consisting 36 of a differential mobility analyzer (DMA; TSI 3040) and a condensation particle counter (CPC; 37 TSI 3775). Isoprene (99%, Sigma-Aldrich) was injected into a glass bulb and zero air was passed 38 over the solution until it evaporated. The initial concentration of isoprene was 97 ppb. H₂O₂ (50% 39 aqueous solution, Sigma-Aldrich) was then injected as an OH precursor. Once the concentrations 40 41 of all species stabilized, UV lights were turned on to initiate photooxidation. The photolysis of H_2O_2 yielded an OH concentration on the order of 10^6 molecules/cm³ under low-NO conditions. 42

43 S1.1. Field Measurements

The Centreville measurements were performed in Centreville, Alabama (USA), from 01 June to 15 July 2013 during the Southern Oxidant and Aerosol Study (SOAS) with the GT AMS.³, ⁴ The Centreville site is a rural site located in a forested area, with high biogenic emissions, especially isoprene and monoterpenes. Biogenic volatile organic compounds (BVOC)-derived organosulfur species have been identified as an abundant contributor to total OA in the southeastern US,^{5, 6} making this location ideal for organosulfur compound measurements. The Mace Head measurements were performed at the Mace Head Global Atmosphere Watch research station, Ireland, from 12 July 2010 to 9 September 2010 with the Galway AMS.⁷ This is a ground site located on the west coast of Ireland and facing westward to the northeast Atlantic, where clean marine air coming onshore can be perturbed by (mostly local) anthropogenic sources. In the summertime high oceanic biological activity results in abundant atmospheric MSA at this site.

The Polarstern measurements were performed on the German research vessel (RV) Polarstern during a cruise from Cape Town, Republic of South Africa to Bremerhaven, Germany, from 20 April to 20 May 2011 with the TROPOS AMS. ⁸ The cruise took place during the autumn in the Southern hemisphere, when both dimethyl sulphide and MSA concentrations are expected to be relatively low, and spring in the Northern hemisphere, where phytoplankton blooms were often encountered, resulting in high MSA concentrations. This spatial contrast was reflected in the MSA concentration time series.

The Wintertime Investigation of Transport, Emissions, and Reactivity (WINTER) aircraft campaign took place out of the NASA Langley Research Center (Hampton, VA) from February 1 to March 15, 2015 aboard the National Center for Atmospheric Research (NCAR) C-130 aircraft, with the Boulder AMS.⁹ One campaign objective was to look at the temporal evolution of power plant plumes under low temperature/stagnant conditions typical of the winter months in the northeastern US.

69 S2. Uncertainty Analysis for Sulfate Apportionment Method

Recall that ΣHSO_{AS}, ΣHSO_{OS/SS}, and ΣHSO_{MSA} are calculated by Equation (10) in the
main text. Including the error of each term we can get:

$$72 \begin{bmatrix} \Sigma HSO_{AS} \pm \delta \Sigma HSO_{AS} \\ \Sigma HSO_{OS/SS} \pm \delta \Sigma HSO_{OS/SS} \\ \Sigma HSO_{MSA} \pm \delta \Sigma HSO_{MSA} \end{bmatrix} = \begin{bmatrix} (f_{HSO_3} \pm \delta f_{HSO_3})_{AS} + (f_{HSO_3} \pm \delta f_{HSO_3})_{OS/SS} + (f_{HSO_3} \pm \delta f_{HSO_3})_{MSA} \end{bmatrix}^{-1} \begin{bmatrix} (HSO_3 \pm \delta HSO_3)_{meas} \\ (f_{H_2SO_4} \pm \delta f_{H_2SO_4})_{AS} + (f_{H_2SO_4} \pm \delta f_{H_2SO_4})_{OS/SS} + (f_{H_2SO_4} \pm \delta f_{H_2SO_4})_{MSA} \\ 1 & 1 \end{bmatrix}^{-1} \begin{bmatrix} (HSO_3 \pm \delta HSO_3)_{meas} \\ (H_2SO_4 \pm \delta H_2SO_4)_{meas} \\ (\Sigma HSO \pm \delta \Sigma HSO)_{meas} \end{bmatrix}$$

For laboratory calibration of standard compounds, δf_{HSO_3} and $\delta f_{H_2SO_4}$ of pure standard compounds (AS and MSA) were calculated as the standard deviation during calibration when signals were stable, while δf_{HSO_3} and $\delta f_{H_2SO_4}$ of OS/SS were calculated as the standard deviation of all OS and SS compounds calibrated in this study. For uncertainties in the measured mass concentratin of sulfate fragments, $\delta \Sigma$ HSO was calculated by error propagation:

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$$\delta \Sigma \text{HSO} = \sqrt{(\delta \text{SO}^+)^2 + (\delta \text{SO}_2^+)^2 + (\delta \text{SO}_3^+)^2 + (\delta \text{HSO}_3^+)^2 + (\delta \text{H}_2 \text{SO}_4^+)^2}$$
(S2)

80 The uncertainties of apportionment results (
$$\Sigma$$
HSO_{AS}, Σ HSO_{OS/SS}, and Σ HSO_{MSA}) can be
81 then assessed via Monte Carlo approach. For the scaling factor, $\left(\frac{\Sigma$ HSO}{total sulfate}\right)_{standard}, used to

convert ΣHSO signals from above calculations to total sulfate signals, the uncertainty was acquired
from the standard calibration. The uncertainties of IE determination (~ 10%), CE determination (~
30%), and RIE determination (~ 15%) were also encapsulated.¹⁰

Uncertainties for field measurement data were calculated and shown as error bars in Figure
S7. 1000 sets of parameters were randomly generated for every point and used as inputs in equation
(10). The campaign average "AS sulfate", "OS/SS Sulfate", and "MSA Sulfate" uncertainties are

26%, 45%, and 23% for Centreville measurements, 36%, 38%, and 90% for Mace Head
measurements, 31%, 86%, and 41% for Polarstern measurements.

90 S3. UMR Data Analysis

In addition to the HR sulfate mass spectra analysis, we also explored the plausibility of 91 deconvolving sulfate signals for UMR data using signals allocated to sulfate at m/z 48 (SO4 48, 92 counterpart of SO⁺), m/z 64 (SO4 64, counterpart of SO₂⁺), m/z 80 (SO4 80, counterpart of SO₃⁺), 93 m/z 81 (SO4_81, counterpart of HSO₃⁺), and m/z 98 (SO4_98, counterpart of H₂SO₄⁺). The results 94 after normalization are shown in Figure S9. For most standard compounds, the results resemble 95 those of HR analysis, and sulfate mass spectra of AS, OS/SS, MSA can still be distinguished by 96 the ion fractions of SO4_81 (f_{81} , counterpart of f_{HSO_3}) and SO4_98 (f_{98} , counterpart of $f_{H_2SO_4}$). 97 Sulfate apportionment was applied to UMR data of laboratory-generated binary mixtures, isoprene 98 99 photooxidation experiment, and one of the field measurements (Centreville SOAS data). For both binary mixtures (MSA+AS, OS+AS), "MSA sulfate" (or "OS Sulfate") to "AS sulfate" molar ratio 100 calculated by the apportionment method correlates well with MSA (or OS) to AS molar ratio in 101 the particles, with $R^2 = 0.998$ and $R^2 = 0.995$, respectively, while UMR analysis overestimates the 102 ratio by a factor of ~2 (Figure S10). For the isoprene photooxidation experiment, "OS sulfate" 103 and "AS sulfate" calculated from UMR data are very similar to HR calculations (Figure S11). For 104 Centreville measurements, 77% of the "OS sulfate" calculated from UMR data are negative values 105 106 (Figure S12), indicating the UMR-based sulfate apportionment may not perform as well as HR data in complex ambient conditions, but we can still conclude from the analysis of UMR data that 107 "AS sulfate" is the dominant source for sulfate signals in AMS at Centreville. 108





Figure S1 Fractions of organosulfur fragments produced by standard organosulfur compounds in
the AMS as a function of the molecular weight of carbon backbones. The data points are colored
by their carbon backbone structures and bonding types.



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Figure S2 Peak fit of main sulfate fragments for OS-10.





Figure S3 Peak fit of main sulfate fragments and CH₃SO₂⁺ for MSA.



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Figure S4 Peak fit of main sulfate fragments for Centreville measurements.



Figure S5 RIE_{SO4} calibration results for MSA and sodium ethyl sulfate (OS-2). The collection
efficiency (CE) of 1 was applied to AMS data considering that atomized organosulfur particles
were liquid droplets. The slope is acquired with intercept forced through zero.



Figure S6 (a) "MSA sulfate" to "AS sulfate" ratio calculated by sulfate apportionment method as
a function of MSA / AS molar ratio in particles. (b) "OS sulfate" to "AS sulfate" ratio calculated
by sulfate apportionment method as a function of OS / AS molar ratio in particles for OS-15 / AS
mixture. The slopes and intercepts are obtained by orthogonal distance regression (ODR). The
Pearson's R is obtained by linear least-squares fit.



135 Figure S7 $f_{H_2SO_4}$ vs. f_{HSO_3} for ambient measurements and time series of "AS sulfate", "OS/SS

sulfate" and "MSA sulfate" for (a) Centreville; (b) Mace Head; (c) Polarstern. OS and SS standard
calibrations are from GT AMS, while MSA and AS standard calibrations are from the AMS that
was used for the corresponding ambient measurements.



140 factor for Centreville measurements. The AMS factor time series are from Xu et al. The Pearson's

141 R is obtained by linear least-squares fit.





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Figure S10 HR and UMR comparisons of (a) "MSA sulfate" to "AS sulfate" ratio as a function of MSA / AS molar ratio in particles; (b) "OS sulfate" to "AS sulfate" ratio calculated by sulfate apportionment method as a function of OS / AS molar ratio in particles for OS-15 / AS mixture. The slopes and intercepts are obtained by orthogonal distance regression (ODR). The Pearson's R is obtained by linear least-squares fit.

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Figure S11 HR and UMR comparisons of (a) "OS sulfate" and (b) "AS sulfate" for the chamber 159 isoprene photooxidation experiment. The slopes and intercepts are obtained by orthogonal distance 160 regression (ODR). The Pearson's R is obtained by linear least-squares fit. 161



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Figure S12 HR and UMR comparisons of (a) "OS sulfate" and (b) "AS sulfate" for Centreville 164 measurements. The slopes and intercepts are obtained by orthogonal distance regression (ODR). 165 The Pearson's R is obtained by linear least-squares fit. 166

| Notation | Description |
|---------------------------------|--|
| AS | Ammonium sulfate |
| MSA | Methanesulfonic acid |
| OS | Organosulfur compounds except for MSA |
| SS | Sodium sulfate |
| Acidic AS | 1:1 ammonium sulfate / sulfuric acid |
| Sulfate fragments | H _x SO _y ⁺ ions produced in AMS of both inorganic and organic origins |
| Organosulfur fragments | $C_xH_yO_zS^+$ ions produced in AMS by organosulfur compounds |
| ΣΗSO | The sum of SO ⁺ , SO ₂ ⁺ , SO ₃ ⁺ , HSO ₃ ⁺ , and H ₂ SO ₄ ⁺ signals |
| fH ₂ so ₄ | The fraction of $H_2SO_4^+$ signal in ΣHSO |
| fhso3 | The fraction of HSO_3^+ signal in ΣHSO |
| "MSA Sulfate" | H _x SO _y ⁺ signals measured in AMS that attributed to MSA by sulfate apportionment method |
| "AS Sulfate" | H _x SO _y ⁺ signals measured in AMS that attributed to AS by sulfate apportionment method |
| "OS/SS Sulfate" | H _x SO _y ⁺ signals measured in AMS that attributed to OS/SS by sulfate apportionment method |
| | |

| Compound | Molecular Structure | Family | Source |
|--|--|---------------|--|
| Sodium methyl sulfate (OS-1) | O S O [−] Na ⁺ | Organosulfate | Commercial (99%) |
| Sodium ethyl sulfate (OS-2) | O_S O⁻Na ⁺ | Organosulfate | Commercial (96.31%) |
| Potassium glycolic acid sulfate (OS-3) | | Organosulfate | Lab Synthesized ¹¹ (> 98%) |
| Potassium hydroxyacetone sulfate (OS-4) | о о о о к+ | Organosulfate | Lab Synthesized ¹¹ (> 98%) |
| Sodium n-heptyl sulfate (OS-5) | O Na ⁺ | Organosulfate | Commercial (99%) |
| Sodium n-octyl sulfate (OS-6) | 0_0 0'Na ⁺ | Organosulfate | Commercial (> 95%) |
| Potassium o-cresol sulfate (OS-7) | | Organosulfate | Lab Synthesized ¹² (> 98%) |
| Potassium p-cresol sulfate (OS-8) | о о к+ 0 0 К+ | Organosulfate | Lab Synthesized ¹² (> 98%) |
| Potassium m-cresol sulfate (OS-9) | о о к+ | Organosulfate | Lab Synthesized ¹² (> 98%) |
| Sodium benzyl sulfate (OS-10) | O ⁻ Na ⁺ | Organosulfate | Lab Synthesized ¹² (> 98%) |

| Potassium 4- nitrophenyl sulfate (OS-11) | 0 ₂ N 0 0 ⁻ К ⁺ | Organosulfate | Commercial (> 98%) |
|---|---|----------------------|-----------------------|
| Potassium 4- hydroxy-3- methoxyphenylglycol sulfate (OS-12) | HO HO OCH ₃ O S O ⁻ K ⁺ | Organosulfate | Commercial (> 98%) |
| Sodium 1- butanesulfonate (OS-13) | | Sulfonate | Commercial (> 99%) |
| Sodium benzenesulfonate (OS-14) | O Na ⁺ | Sulfonate | Commercial (97%) |
| Methanesulfonic acid (MSA) | 0 о к+ | Sulfonic Acid | Commercial (> 99%) |
| Ethanesulfonic acid (OS-15) | O S O Na ⁺ | Sulfonic Acid | Commercial (95%) |
| Ammonium Sulfate (AS) | (NH ₄) ₂ SO ₄ | Inorganic Sulfate | Commercial (> 99%) |
| Sodium Sulfate (SS) | Na ₂ SO ₄ | Inorganic Sulfate | Commercial (> 99%) |

| Name | fso | fso ₂ | fso3 | $f_{\rm HSO_3}$ | $f_{\rm H_2SO_4}$ |
|--------------|--------|------------------|--------|-----------------|-------------------|
| OS-1 | 0.3760 | 0.6014 | 0.0215 | 0.0010 | 0.0000 |
| OS-2 | 0.3821 | 0.5620 | 0.0528 | 0.0028 | 0.0003 |
| OS-3 | 0.3686 | 0.6139 | 0.0174 | 0.0000 | 0.0002 |
| OS-4 | 0.3755 | 0.6060 | 0.0184 | 0.0001 | 0.0000 |
| OS-5 | 0.3655 | 0.5976 | 0.0364 | 0.0004 | 0.0000 |
| OS-6 | 0.3864 | 0.5453 | 0.0671 | 0.0009 | 0.0002 |
| OS-7 | 0.3772 | 0.5790 | 0.0433 | 0.0004 | 0.0001 |
| OS-8 | 0.3735 | 0.5922 | 0.0336 | 0.0005 | 0.0002 |
| OS-9 | 0.3754 | 0.5931 | 0.0310 | 0.0005 | 0.0000 |
| OS-10 | 0.3664 | 0.6037 | 0.0285 | 0.0012 | 0.0002 |
| OS-11 | 0.3684 | 0.5983 | 0.0333 | 0.0000 | 0.0000 |
| OS-12 | 0.3807 | 0.5705 | 0.0481 | 0.0006 | 0.0001 |
| OS-13 | 0.3754 | 0.6167 | 0.0061 | 0.0017 | 0.0001 |
| OS-14 | 0.3712 | 0.6171 | 0.0044 | 0.0072 | 0.0001 |
| OS-15 | 0.3796 | 0.6115 | 0.0021 | 0.0068 | 0.0000 |
| MSA | 0.4040 | 0.5358 | 0.0014 | 0.0587 | 0.0001 |
| AS | 0.3338 | 0.4223 | 0.1596 | 0.0546 | 0.0297 |
| Acidic AS | 0.3494 | 0.4667 | 0.0949 | 0.0567 | 0.0324 |
| SS | 0.3773 | 0.5914 | 0.0294 | 0.0013 | 0.0006 |

 Table S4 Sulfate Fragmentation Table

| Ion | HR_frag_sulfate (AS) | HR_frag_sulfate (MSA, OS, SS) |
|--|--|--|
| O 0.04*HR_frag_sulfate[{H2O}] | | - |
| HO 0.25*HR_frag_sulfate[{H2O}] | | - |
| j18O 0.00205499*HR_frag_sulfate[{O}] | | - |
| H2O 0.67*HR_frag_sulfate[{SO2}], 0.67*HR_frag_sulfate[{SO}] | | - |
| Hj18O 0.00205499*HR_frag_sulfate[{HO}] | | - |
| H2j18O 0.00205499*HR_frag_sulfate[{H2O}] | | - |
| S | 0.21*HR_frag_sulphate[{SO2}], 0.21*HR_frag_sulphate[{SO}], 0.068*HR_frag_sulphate[{HSO3}], 0.068*HR_frag_sulphate[{HSO3}] | 0.21*HR_frag_sulphate[{SO2}], 0.21*HR_frag_sulphate[{SO}], 0.068*HR_frag_sulphate[{HSO3}], 0.068*HR_frag_sulphate[{HSO3}] |
| j33S | 0.00789557*HR_frag_sulphate[{S}] | 0.00789557*HR_frag_sulphate[{S}] |
| j34S | 0.0447416*HR_frag_sulphate[{S}] | 0.0447416*HR_frag_sulphate[{S}] |

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