

96 Eyes: Parallel Fourier Ptychographic Microscopy for high-throughput screening

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ABSTRACT

This document provides supplementary information to the “96 Eyes: Parallel Fourier Ptychographic Microscopy for high-throughput screening,” [year], pp. [page].

Supplementary Information

1 Parallel FPM acquisition and reconstruction

2 A parallel image acquisition technique is proposed here. Four
3 (4) frame grabbers are simultaneously controlled by individual
4 processes in the workstation, each is run in individual
5 central processor core. One of the process supervises the
6 illumination system to implement step, and then sends out
7 trigger signal to all other processes to perform image acquisition
8 and storage. As shown in Figs. S1(b-c), the ratio of the
9 number of image sensors to the number of running processes
10 is equal to 24, that corresponds to four set of 24-to-1
11 multiplexers for a total of the 96 image sensors. With respect
12 to target applications, such ratio can be varied to optimize
13 the overall data throughput within the allowable bandwidth
14 of the interface.

15 Another data throughput challenge preciously not addressed
16 in previous studies (e.g. EmSight¹) is the requirement
17 of segmenting the image data into tiles on the fly. If the
18 images are first saved and segmented later, both the imaging
19 system and the graphical processor(s) will be idling, thus it
20 limits the overall image restoration throughput. Our study
21 shows that our system can finish writing the raw image data
22 within 2 minutes, yet it takes around 20 minutes to reorganize
23 (i.e. read, segment, and write) the raw data from/to the hard
24 drive. This challenge can be addressed with a in-memory parallel
25 data storage strategy accessible by all running processes,
26 which houses a four-dimensional image data “hypercube”
27 with a dimensions of “number of illumination angles” times
28 “number of image sensors” times “image height” times “image
29 width”. The hypercube is pre-segmented into chunks
30 of dimensions (in our case, it is $1 \times 96 \times 256 \times 256$). For
31 each unique illumination pattern, the incoming image data
32 of all image sensors are simultaneously sorted, indexed, and

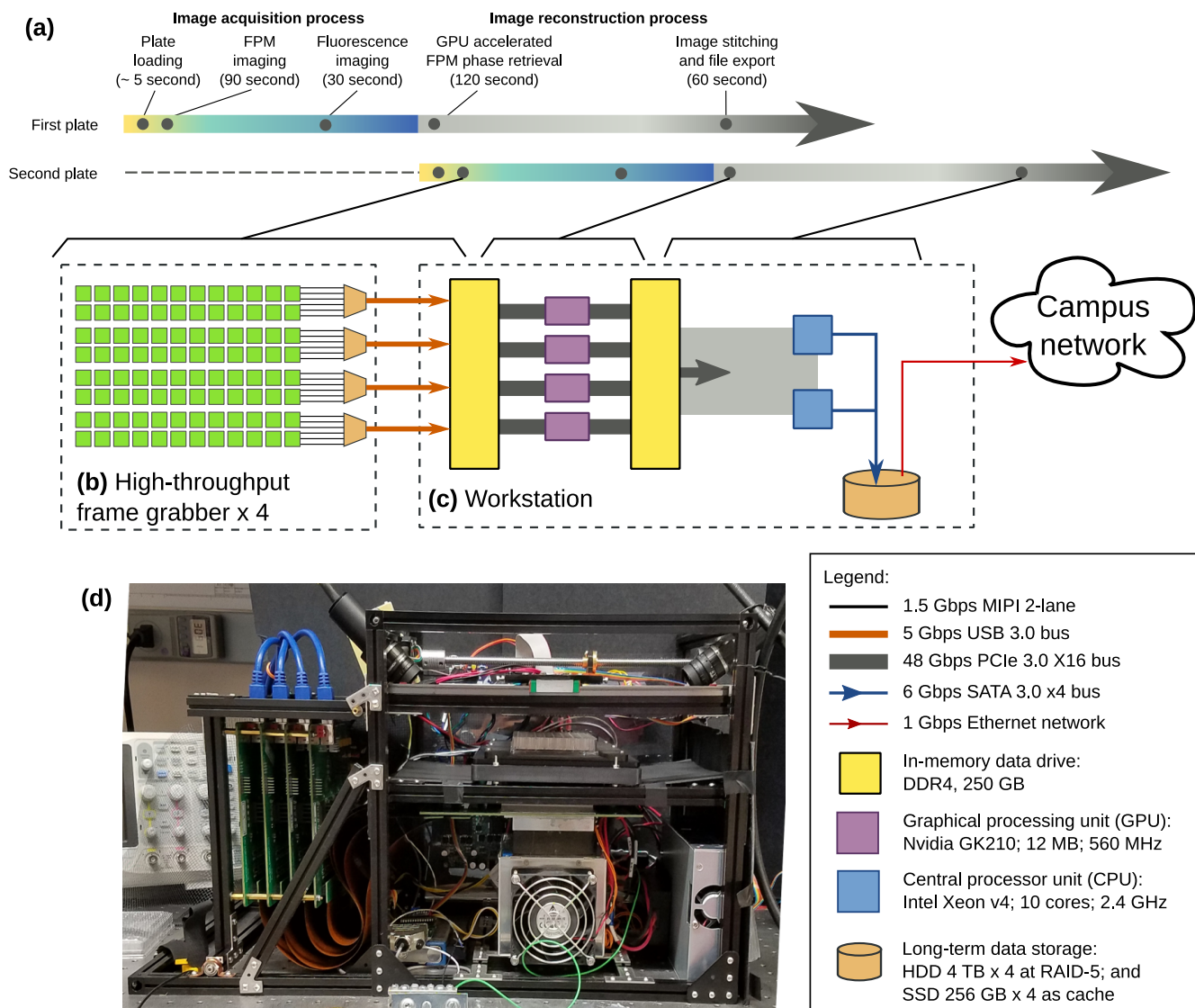
segmented online by the file system. The individual chunks
of the hypercube are then written to the hard drive in a linear
layout, which facilitates the image segment loading and
restoration method in the next step. In short, by sorting
and segmenting the incoming image data on the fly, it helps
saving the precious data bandwidth.

Enabled by the data alignment of the image chunks and
the identical illumination pattern across all image sensors,
multiple image segments can be restored by the graphical
processor in a massively parallel manner. The corresponding
image segments for all image sensors (i.e. at identical locations
in the image FOV) can be processed simultaneously as they
possess an identical set of illumination conditions. This
substantially reduces the GPU idling time because a chunk
in the data “hypercube” only requires one set of function
calls and data read/write instead of 96 (for 96 image sensors),
reducing the processing overhead.

If only a single 96-well plate is imaged and analyzed, the
back-to-back data acquisition (one layer of phase image plus
10 z-layers of fluorescence image) and processing pipeline
requires $(90 + 30 + 120) = 240 \text{ s} \approx 4 \text{ min}$ to complete. However,
if multiple plates are involved in one batch of study, the
acquisition stages and the reconstruction stages can be performed
simultaneously (Fig. S1), reducing the overall imaging
time to around 120 second per plate.

LED position calibration

Fourier ptychographic algorithm requires accurate illumination
angles from different LEDs in order to register the raw
images in the Fourier domain. Because of the presence of
liquid meniscus in the 96-well culture plate, the LEDs appears
to be much closer to the object than they are physically
located, altering the incident angles of the light rays on the
object. Here, we present the ray tracing method to estimate



Supplementary Figure S1. Parallel FPM acquisition and reconstruction process.(a) Timeline of plate image acquisition and reconstruction processes two consecutive plates. Since the reconstruction process can be done offline, the second plate can be loaded and imaged while the workstation is reconstructing the images of the first plate. (b) and (c) Four (4) high-throughput frame grabbers streams raw images to the internal memory buffers of the workstation through the high speed links. (d) Front view of the 96 Eyes hardware.

66 the incident angles due to refraction.

67 First, we consider the case when the liquid interface is
 68 devoid of meniscus. Let us denote the vertical distance be-
 69 tween the object and the light source by h_a , and the liquid
 70 medium (refractive index = n) height above the object by h_b .
 71 For a light ray from a single LED passing through the flat
 72 air-to-liquid interface [inset of Supp. Fig. S2], the angle of
 73 illumination on the sample θ is governed by

$$n \sin \theta = \frac{x_a - x_b - \delta}{\sqrt{(h_a - h_b)^2 + (x_a - x_b - \delta)^2}} \quad \text{Snell's law} \quad (\text{S1})$$

$$\sin \theta = \frac{\delta}{\sqrt{h_b^2 + \delta^2}} \quad \text{Geometry} \quad (\text{S2})$$

74 The close form solution of $\sin \theta$ exists, but it involves find-
 75 ing the root of a fourth order polynomial derived from the
 76 above equations. Instead, we numerically solve for δ with
 77 the following root-finding algorithm

$$\delta^0 = \frac{(x_a - x_b)h_b}{h_a} \quad (\text{S3})$$

$$\delta^{k+1} = g(\delta^k)$$

$$\text{subject to } g(\delta) = \frac{1}{n} \left[\frac{(x_a - x_b - \delta) \sqrt{h_b^2 + \delta^2}}{\sqrt{(h_a - h_b)^2 + (x_a - x_b - \delta)^2}} \right], \quad (\text{S4})$$

78 which guarantees to converge for $|x_a - x_b| < h_a - h_b$. The
 79 illumination angle can be now be evaluated by substituting
 80 δ^K into Eq. S2 for a large number K .

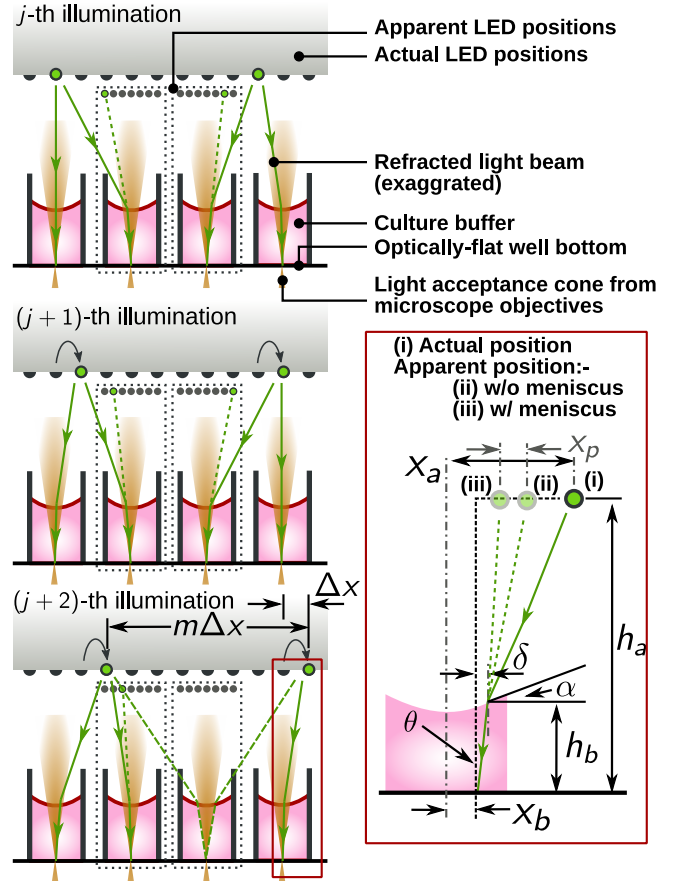
81 Next, we analyze the changes to the optical light path in
 82 the presence of meniscus. The meniscus introduces a tilted
 83 air-to-liquid interface at an angle $\alpha(x_b)$, which is a function
 84 of the lateral position x_b from the center of the well on the
 85 culture plate. Trying to incorporate this variable to the ray-
 86 tracing model will add unnecessary complexity to Eq. S1.
 87 Therefore, we linearize the meniscus effect by introducing a
 88 parallax shift x_p , with

$$x_p(x_b) = (h_a - h_b)[\tan(\alpha(x_b) + \theta) - \tan \theta] \approx cx_b, \quad (\text{S5})$$

89 for some constant $c > 0$. The meniscus-compensated illu-
 90 mination angle θ is now approximated by modifying Eq. S4
 91 with $x_a \mapsto x_a - x_p(x_b) \approx x_a - cx_b$.

92 Speed improvement factor and the design criteria 93 of the parallel illumination scheme

94 Without parallel illumination, only a single camera is ac-
 95 tive at any instance of image acquisition. Let f be the
 96 effective frame rate of a single camera. For the 96-well
 97 plate, the total acquisition time required is equal to $f^{-1} \times$
 98 number of wells \times number of illumination = $4704f^{-1}$. Par-
 99 allel illumination scheme instead utilizes a 2D lattice illu-
 100 mination pattern with a source-to-source separation of m LEDs



Supplementary Figure S2. Detailed illustration of the parallel illumination scheme of 96 Eyes. The source-to-source separation is chosen to maximize the effective acquisition rate, as well as avoiding interference. This is made possible by making sure that only one single LED is responsible for brightfield illumination for any camera and for any time instance of ptychographic image acquisition. Inset: definition of symbols for LED position calibration.

with a LED-to-LED separation of Δx . The total number of illumination is now reduced to m^2 . Hence, the effective acquisition time is equal to $f^{-1} \times m^2 \times$ number of cameras \times (number of frame grabber cards) $^{-1} = 24m^2f^{-1}$, for four frame grabber cards. The speed improvement is given as

$$\frac{4704f^{-1}}{24m^2f^{-1}} = 196m^{-2}. \quad (\text{S6})$$

In the following paragraph, we will compute the lower limit of value m . For all time instances, we only allow one LED to fulfill the brightfield illumination condition with respect to the object (Fig. S2). Let us denote the vertical distance between the object and the light source by h_a , and the liquid medium (refractive index = n) height of h_b . The following conditions have to be fulfilled in addition to Eqs. (S1) and

113 (S2):

$$n \sin \theta \geq \text{NA} \quad (\text{S7})$$

$$x_b = 0 \quad (\text{S8})$$

$$x_a = m \Delta x / 2 \quad \text{subject to } m = 2^M, \quad (\text{S9})$$

114 for a given numerical aperture (NA) of the microscope ob-
 115 jective, and some integer M . Here a power of two is pre-
 116 ferred because it simplifies the electronic design of the LED
 117 matrix. For our system with $h_a = 33 \text{ mm}$, $h_b = 3 \text{ mm}$ and
 118 $\Delta x = 3 \text{ mm}$, we picked $m = 8$. This implies a conservative
 119 speed improvement of at least 3 times. Compared to mechan-
 120 ical scanning system which has a much lower effective frame
 121 rate f , the speed improvement can be up to 8 times compared
 122 to commercially available instruments.

123 Modification to the Fourier ptychography phase re- 124 trieval algorithm

125 **Forward model for our imaging system** Let us denote
 126 a segment of the object to be reconstructed by $u \in \mathbb{C}^n$, a
 127 two-dimensional image with $n^{1/2} \times n^{1/2}$ pixels. We also
 128 denote the j -th illuminated low-resolution intensity image
 129 of the object by $I_j \in \mathbb{R}_+^m$, with $m^{1/2} \times m^{1/2}$ pixels (u and
 130 I_j are both written as a vector by a lexicographical order).
 131 It can be shown that $I_j = |\mathcal{F}^H \text{diag}(p) \mathbf{Q}_j \mathcal{F} u|^2$. The pupil
 132 function $p \in \mathbb{C}^m$ can be considered as the circular aperture
 133 at the back aperture plane of the imaging system. Binary
 134 matrix $\mathbf{Q}_j \in \mathbb{R}^{m \times n}$ depicts the downsampling of the object
 135 u by cropping a region of m pixels in Fourier space cor-
 136 responding to the j -th position of the light source. What
 137 we measure is a stack of low-resolution intensity images
 138 $I_j = |\mathcal{F}^H \psi_j|^2 = |\mathcal{F}^H \text{diag}(p) \mathbf{Q}_j \mathcal{F} u|^2 \in \mathbb{R}_+^m$, $j = 1, 2, \dots, k$,
 139 where the hyperscript H denotes a Hermitian conjugate. The
 140 operation $\text{diag}(a)b$ represents the element-by-element mul-
 141 tiplication² between two vectors a, b .

142 In reality, the measured sequence of low-resolution images
 143 are corrupted by (i) the ambient light level $I_b > 0$, (ii) angular
 144 dependency of LED intensity $w_j > 0$, (iii) background inter-
 145 ference of suspended particulates in the liquid $\phi_{\text{dust}} \in \mathbb{R}^n$;
 146 and (iv) dark current and readout noise of the sensor $n_j \in \mathbb{R}^m$.
 147 Therefore, we modified the forward model to

$$I_j = w_j |\mathcal{F}^H \text{diag}(p) \mathbf{Q}_j \mathcal{F} \text{diag}(e^{i\phi_{\text{dust}}}) u|^2 + I_b + n_j, \quad (\text{S10})$$

148 Minimizer with partial spatial coherence constraint

149 Since the lens aberration is almost completely unknown, one
 150 has to solve a blind ptychographic phase retrieval problem
 151 with an amplitude constraint²

$$\begin{aligned} & \min_{\{w_j\}, p, u} \sum_{j=1}^N f_j(w_j, p, u) \\ \Leftrightarrow & \min_{\{w_j\}, p, u} \sum_{j=1}^N \left\| |\mathcal{F}^H \text{diag}(p \sqrt{w_j}) \mathbf{Q}_j \mathcal{F} u| - \sqrt{I_j - I_b} \right\|_2^2, \end{aligned} \quad (\text{S11})$$

Algorithm 1 Pseudo-code of the phase retrieval algorithm
 for 96 Eyes system

1. **Inputs:** segments of low resolution images I_j and am-
 bient light level I_b of the corresponding camera.
 2. Initialize local pupil functions p_ℓ for all L segments of
 the object.
 3. Estimate the global pupil function $\sum p/L :=$
 $(1/L) \sum_{\ell=1}^L p_\ell$.
 4. **FPM-EPRY algorithm:** Run the phase retrieval algo-
 rithm for the ℓ -th segment with pupil function recovery.
 - (a) **Initialize** $u^0 := \sqrt{I_0 - I_b}$,
 $p^0 := \sum p/L$ and $w_j^0 := 1$ for $j \in [1, N]$.
 - (b) For the k -th iteration,
 - i. Evaluate $j = \text{mod}(k, N) + 1$.
 - ii. **Object update:** solve $u^{k+1} =$
 $\arg \min_u f_j(w_j^k, p^k, u)$.
 - iii. **Weighting update:** when $k \leq 3N$, solve
 $w_j^{k+1} = \arg \min_w f_j(w, p^k, u^{k+1})$. Otherwise,
 $w_j^{k+1} = w_j^k$.
 - iv. **Pupil update:** when $k > 3N$, solve
 $p^{k+1} = \arg \min_p f_j(w_j^{k+1}, p, u^{k+1})$. Other-
 wise, $p^{k+1} = p^k$.
 - (c) Repeat step (4b) until $k = K$.
 - (d) Update the object estimate $u_\ell := u^K$ and local
 pupil function estimate $p_\ell := p^K$.
 5. Repeat steps (3)–(4) one more time.
 6. **Separate u from $e^{i\phi_{\text{dust}}}$** by digitally high-passing the
 phase component of u_ℓ with an inverted Gaussian
 blur kernel. The amplitude component is preserved as
 $|u_\ell^{\text{cell}}| := |u_\ell|$.
 7. **Stitch** the recovered image segments u_ℓ for all $\ell \leq L$.
 8. **Outputs:** amplitude and phase component of the
 stitched image u , the global pupil function \bar{p} and the
 local aberrations $\{p_\ell\}$.
-

152 Because of the limited number of low-resolution images (=21)
 153 in the measurement, the estimated pupil function p^{est} cannot
 154 be efficiently separated from the estimated object $\mathcal{F}u^{\text{est}}$ in
 155 the Fourier domain. This shortcoming is compounded by
 156 the fact that the target biological specimen is a weak phase
 157 object, where most of the information in the Fourier domain
 158 is concentrated in that of the un-scattered transmitted light.
 159 To suppress the crosstalk between the two, we utilize a finite
 160 number ($L > 0$) of overlapping segments of the object u_ℓ and
 161 the corresponding local pupil p_ℓ to enforce the partial spatial
 162 coherence constraint. That is, the above minimizer is further
 163 subject to

$$\sum_{\ell=1}^L \|p_\ell - \sum p/L\|_2^2 \leq \epsilon^{\text{tol}}, \quad (\text{S12})$$

164 for a “global” average pupil function $\sum p/L = (1/L)\sum_{\ell=1}^L p_\ell$
 165 and tolerance value $\epsilon^{\text{tol}} > 0$.

166 **Background estimation** To recover the average level of
 167 the ambient light level I_b , we capture the images when all
 168 light sources are switched off. The value of I_b for a partic-
 169 ular CMOS sensor is then set to be the pixel average of the
 170 captured dark image.

171 **Separation of the non-uniform illumination profile of
 LEDs and the pupil function** From Eq. (S11), it is known
 172 that the pupil function p cannot be efficiently separated from
 173 the factor w_j . Therefore, the factor w_j is optimized only for
 174 the first three iterations³, while the recovery of p is post-
 175 poned until the fourth iteration.

176 **Separation of cells and background interference** The
 177 out-of-focus suspended particulates show up as blurred shad-
 178 ows in the sequence of low-resolution images [Supp. Fig. S3].
 179 We utilize this property to estimate ϕ_{dust} by applying a Gaus-
 180 sian blur of the recovered object phase. The morphological
 181 information of the cells can be extracted from the phase
 182 difference between the recovered field u^{est} and $e^{i\phi_{\text{dust}}}$.

183 It is noted that there are existing algorithms that specializes
 184 in separation of the object from out-of-focus noise⁴.

185 **Choice of adaptive step size for pupil recovery** While
 186 the object update in Step 4(b)ii of Algorithm 1 is solved by
 187 the time-honored Gaussian-Newton algorithm⁵, the pupil
 188 update in Step 4(b)iv of Algorithm 1 is instead solved by the
 189 gradient descent method⁶, with

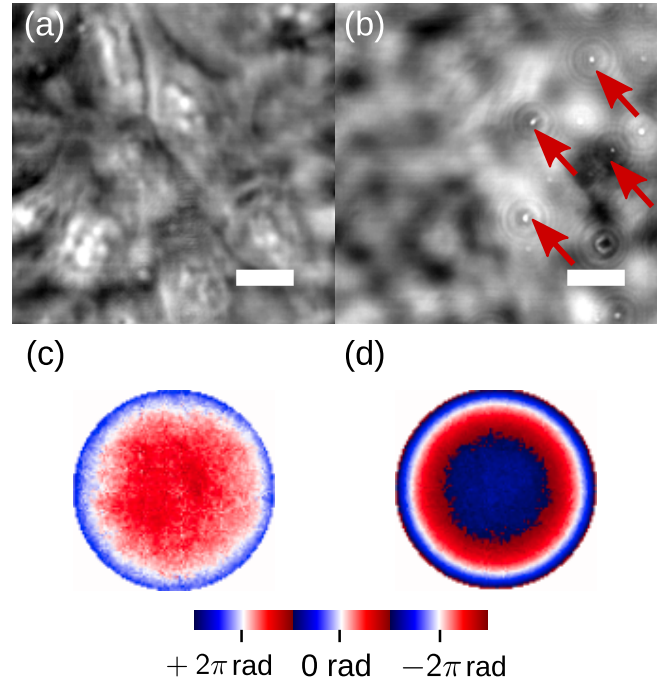
$$\begin{aligned} p^{k+1} &= \arg \min_p f_j(w_j^k, p, u^k) \\ &= p^k + \gamma \text{diag}(\bar{s}^k) \times \\ &\quad \left[\mathcal{F} \text{diag} \left(\frac{\sqrt{I_j}}{|\mathcal{F}^H g_j(w^k, p^k, s^k)|} \right) \mathcal{F}^H g_j(w^k, p^k, s^k) - \right. \\ &\quad \left. g_j(w^k, p^k, s^k) \right], \end{aligned} \quad (\text{S13})$$

191 where $s^k = \mathcal{F}u^k$ and $g_j(w^k, p^k, s^k) = \text{diag}(p^k \sqrt{w_j^k}) \mathbf{Q}_j s^k$ for
 192 a step size of $\gamma \in \mathbb{R}_+^m$. Because of the choice of parallel il-

lumination in our 96 Eyes system, all of the captured data
 are brightfield images. For a weak phase object, most of the
 incoming light rays remains un-scattered, that result in a
 strong peak in the Fourier domain. If the step size γ is a
 constant, the recovered pupil will be corrupted with a con-
 stellations like artifact [Fig. S4(a)]. Therefore, we heuristically
 adjust the step size with

$$\gamma = \left[\text{diag} \left((1 - \beta) |s^k| + \beta \|s^k\|_\infty \right) \right]^{-1}, \quad (\text{S14})$$

where $\|s\|_\infty$ denotes the maximum amplitude of the complex-
 valued signal s . Effectively, the step size γ normalizes the
 value of $\text{diag}(\bar{s}^k)$. The non-dimensional number $\beta \in [0, 1]$
 adjusts the relative strength of normalization of signal \bar{s}^k .
 When $\beta = 0$, the Fourier domain of the object $s^k = \mathcal{F}u^k$ will
 be completely normalized. In the main text, the value is set
 to be $\beta = 10^{-6}$, around twice the order-of-magnitude of an 8-
 bit image. This helps smooth the pupil function [Fig. S4(b)] as
 well as reduce the reconstruction residual [Fig. S4(c)], defined



Supplementary Figure S3. Particulates outside of the focal plane introduce background interference. (a) Phase component of the recovered complex wavefront, showing the the U2OS cell line almost buried in the phase fluctuation; (b) phase image of dust particles on the underside of the well plate, reconstructed by digital refocusing of the recovered complex wavefront. (c) Recovered phase component of system aberration; and (d) pupil function used for digital refocusing.

$$\epsilon^k = \frac{\sum_{j=1}^N \left\| |\mathcal{F}^H g_j(w^k, p^k, s^k)| - \sqrt{I_j - I_b} \right\|_2^2}{\sum_{j=1}^N \left\| \sqrt{I_j - I_b} \right\|_2^2}. \quad (\text{S15})$$

Improving the dynamic range of fluorescence images with two-stage digital averaging

Because of the limited photo-sensitivity and bit depth of our choice of consumer-grade CMOS sensor, we adopted the digital averaging approach to enhance the signal-to-noise ratio of the sensor. The digital averaging technique is also known as *dithering* in audio digitization community⁷⁻⁹, where the band-limited signal of interest is mixed with an artificial out-of-band noise on the input side of the analog-to-digital conversion circuit to reduce the quantization error. Our technique is also very similar to *halftoning* of digital images¹⁰, where an artificial pepper noise is added to simulate grayscale images out of a black-and-white display device. In contrast,

the noise source for our CMOS sensors in the 96 Eyes system cannot be precisely controlled. Notably, similar digital averaging approaches has been proposed before for radiometry studies¹¹. However, the underlying principle is poorly understood. Here, we provide a theoretical framework to offer to explain the dynamic range improvement of our fluorescence images with digital averaging.

Forward model For a fluorophore concentration $c(x, y)$ illuminated by an uniform intensity I_0 , the imaging system in the fluorescence channel is empirically modeled as

$$\begin{aligned} I(x, y, t) &= \lfloor g_{\text{amp}} \eta c(x, y) I_0 + g_{\text{amp}} n_{\text{dark}}(x, y, t) + n_{\text{amp}}(t) \rfloor \\ &= g_{\text{amp}} \eta c(x, y) I_0 + n_{\text{amp}}(t) + \\ &\quad g_{\text{amp}} n_{\text{dark}}(x, y, t) + \epsilon(x, y, t), \end{aligned} \quad (\text{S16})$$

where the non-dimensional factor η is a product of (i) quantum efficiency of the fluorophore, (ii) photon collection efficiency of the microscope objectives, and (iii) quantum efficiency of the photosensing circuit in the CMOS sensor. The amplifier with gain $g_{\text{amp}} > 0$ naturally comes with an additive power-line noise $n_{\text{amp}}(t)$. The round-off operator $\lfloor \cdot \rfloor$ denotes the quantization process, which in turn can be modeled as an additive quantization error $\epsilon(x, y, t) \in [-0.5, +0.5)$. Here, the photon noise is assumed to be negligible compared to dark current noise.

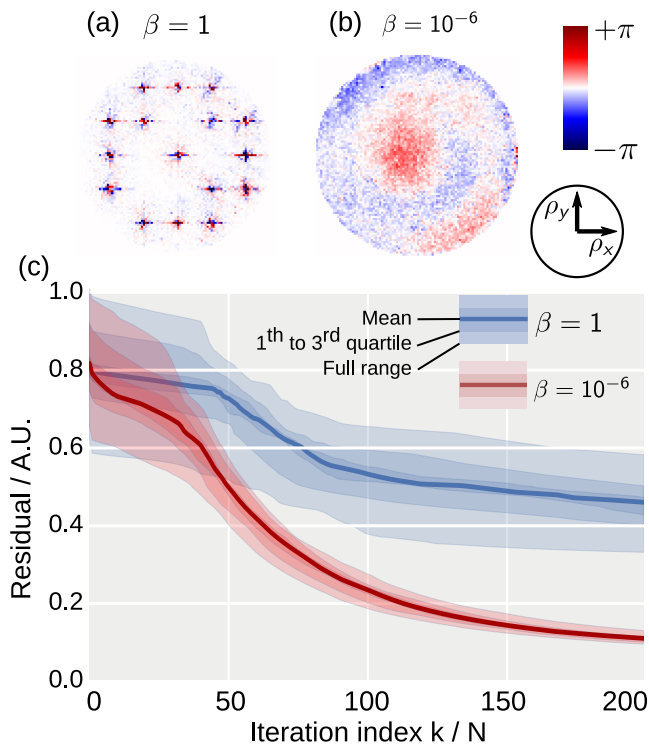
In rolling shutter mode, rows of pixels are read out at a traversal rate of v , so the amplifier noise is mapped to the vertical axis of the j -th image $I_j(x, y)$.

$$\begin{aligned} I_j(x, y) &= g_{\text{amp}} \eta c(x, y) I_0 + n_{\text{amp}}(y, j) + \\ &\quad g_{\text{amp}} n_{\text{dark}}(x, y, j) + \epsilon_j(x, y), \end{aligned} \quad (\text{S17})$$

where $n_{\text{dark}}(x, y, j) = n_{\text{dark}}(x, y, t)|_{t=t_j+y/v}$ and $n_{\text{amp}}(y, j) = n_{\text{amp}}(t = y/v + jH/v)$ for H rows of pixels in the CMOS sensor.

Suppressing both dark current noise and quantization error with digital averaging Consumer-grade CMOS sensors, designed for daylight applications, have a much higher quantization error than the dark current noise. For a amplifier gain value g_{amp} at unity, the dark current noise component is typically always rounded-off to zero. In other words, direct digital averaging of multiple frames I_j at unity gain usually do not result in reduction of quantization error. However, typical biological specimen is known to have a low fluorophore concentration. Concerns about photobleaching also limit the illumination intensity I_0 . Therefore, the amplifier gain g_{amp} must be boosted sufficiently to utilize the full quantization range of the CMOS sensor.

The dark current noise is known to possess a Gaussian distribution¹², i.e. $n_{\text{dark}}(t) \sim \mathcal{N}(0, \sigma_{\text{dark}}^2)$ (symbols x, y are omitted for clarity). This also applies to the power-line noise, where $n_{\text{amp}}(t) \sim \mathcal{N}(0, \sigma_{\text{amp}}^2)$. For a sufficient gain with $g\sigma_{\text{dark}} \gg 0.5$, the probability density function of I_j is given



Supplementary Figure S4. Adaptive step size

improves pupil function recovery. (a) Recovered phase component of pupil function with a constant step size, i.e. at $\beta = 1$, compared to (b) at $\beta = 10^{-6}$. Symbols (ρ_x, ρ_y) are the local coordinates of the pupil function. (c) Comparison of reconstruction residuals by applying phase retrieval to all segments ($L = 80$) of the cell sample captured from one camera. With our method, the residual reduces by around one-third (after $k/N = 200$ iterations) with a much smaller spread, demonstrating a more robust object and pupil co-recovery.

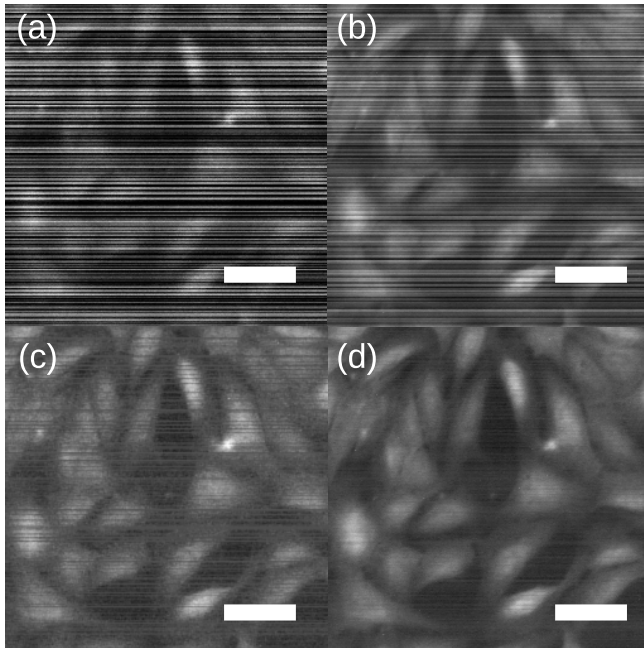
$$P(I_j = a) = \begin{cases} \frac{1}{A} \int_{a-0.5}^{a+0.5} \exp\left(\frac{-(I - g_{\text{amp}}\eta c I_0)^2}{(\sigma_{\text{amp}}^2 + g_{\text{amp}}^2 \sigma_{\text{dark}}^2) \sqrt{2}}\right) dI & \text{if } a \text{ is an integer,} \\ 0 & \text{otherwise.} \end{cases} \quad (\text{S18})$$

268 The scaling factor A is defined such that $\int_{-\infty}^{\infty} P(I_j = a) da =$
269 1. By averaging a sufficient number of frames, i.e.

$$I_1(x, y) := \frac{1}{N} \sum_{j=1}^N I(x, y, t_j), \quad (\text{S19})$$

270 both the dark noise and the quantization error can be re-
271 duced. For instance, it can be shown that $\lim_{N \rightarrow \infty} I_1(x, y) =$
272 $\int_{-\infty}^{+\infty} a P(I_j = a) da = g_{\text{amp}}\eta c(x, y) I_0$, which is independent
273 of both noise terms.

274 **Suppressing the power-line noise** For our digital-
275 averaged fluorescence signal captured by the 96 Eyes system,
276 we can still observe the presence of row-wise intensity fluctu-
277 ation [Supp. Fig. S5(b)] originated from the power-line
278 noise above the quantization level, modeled as $I_1(x, y) \approx$
279 $g_{\text{amp}}\eta c(x, y) I_0 + n_{\text{amp}}(y)$. This is caused by the power-line
280 noise in the 96-in-1 camera board.



Supplementary Figure S5. Improving the dynamic range of fluorescence images with digital averaging.

(b) Single frame at gain $g_{\text{amp}} = 8$; (c) averaging 10 frames at gain $g_{\text{amp}} = 8$; Suppressing the band-like pattern noise for (c) a single frame, and (d) the digital average of 10 frames. All images are contrast-stretched to highlight the background noise and artifacts. Scale bar: $20 \mu\text{m}$.

281 The size constraint of the 96-well culture plate limits the
282 available real estate on the printed-circuit board for electronic
283 filters, especially the decoupling capacitors. To further sup-
284 press such row-wise fluctuations, we apply the same digital
285 averaging technique to isolate it from the fluorescence signal
286 $c(x, y)$. Here, we assume that the fluorophore concentration
287 possesses a Gaussian distribution $c(x, y) \sim \mathcal{N}(\mu_c, \sigma_c^2)$ for
288 $\mu_c > 0$, $\sigma_c \ll \mu_c$. By taking a row-wise average of pixels of
289 $I_1(x, y)$, we have

$$I_2(y) := \frac{1}{W} \sum_{i=1}^W I_1(x_i, y) \approx g_{\text{amp}}\eta \mu_c I_0 + n_{\text{amp}}(y), \quad (\text{S20})$$

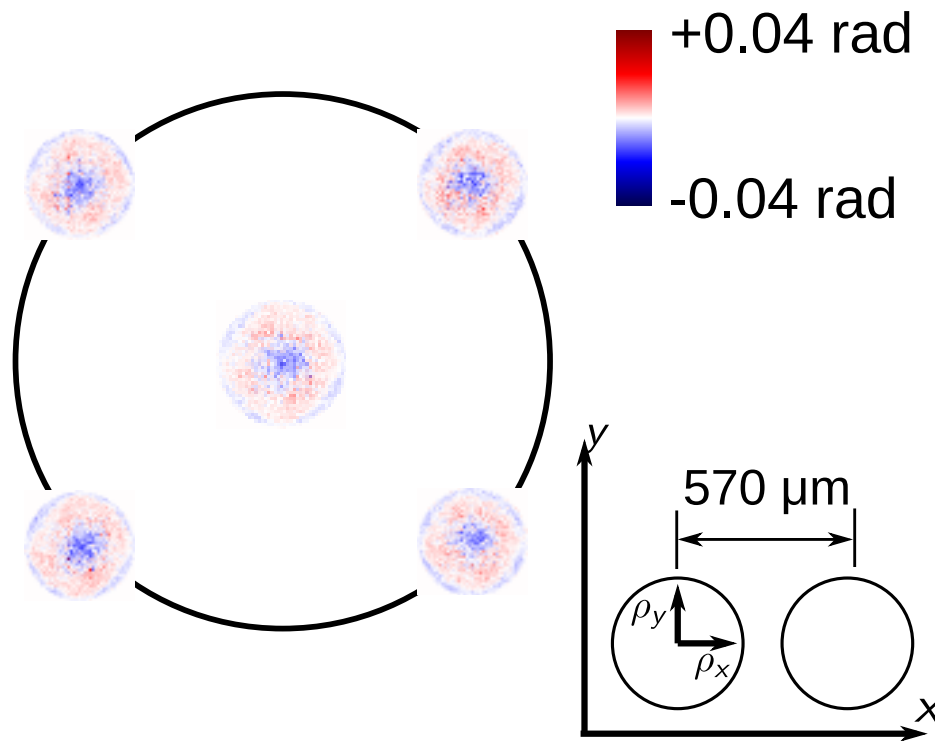
290 for W pixels along individual rows of the image. Since we
291 only care about the morphology of the biological cells stained
292 with the fluorophore, the average fluorophore concentration
293 μ_c can be eliminated as well. Hence, the recovered fluores-
294 cence image is given as

$$c^{\text{est}}(x, y) := c(x, y) - \mu_c \approx \frac{I_1(x, y) - I_2(y)}{g_{\text{amp}}\eta I_0}. \quad (\text{S21})$$

295 In practice, the signal $c(x, y)$ does not fit well with the Gaus-
296 sian process assumption. The row-wise averaging operation
297 Eq. S20 is replaced with row-wise median operation to reduce
298 sensitivity to extreme values in $c(x, y)$.

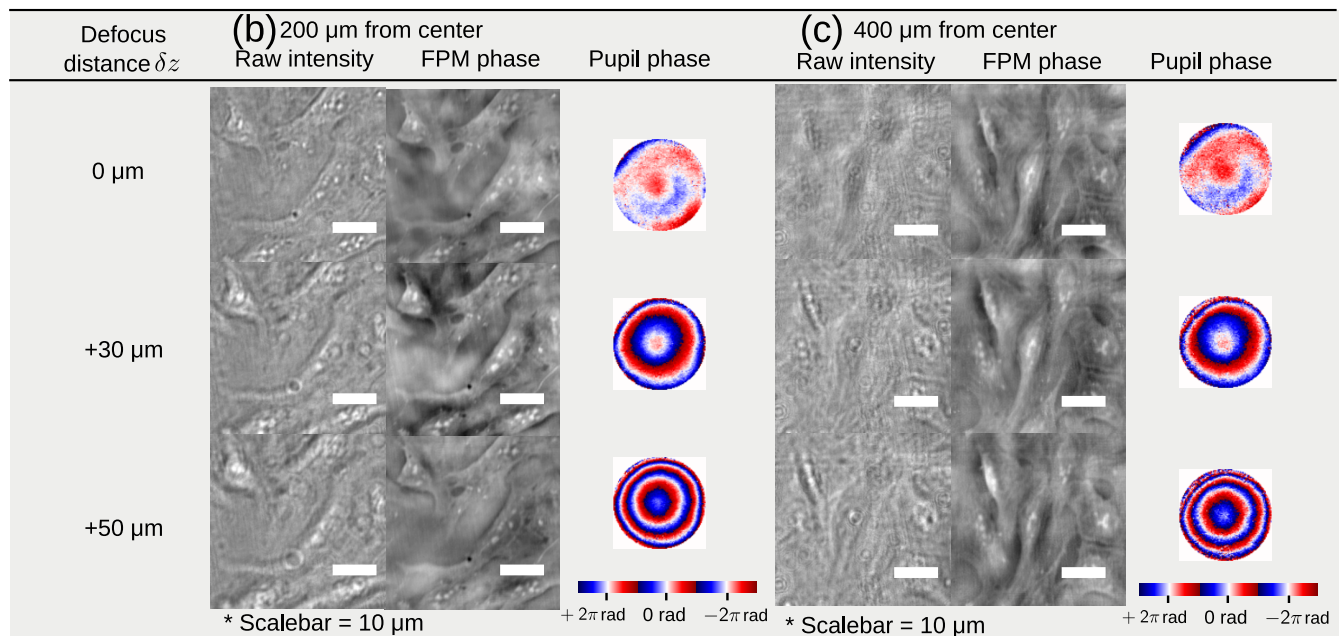
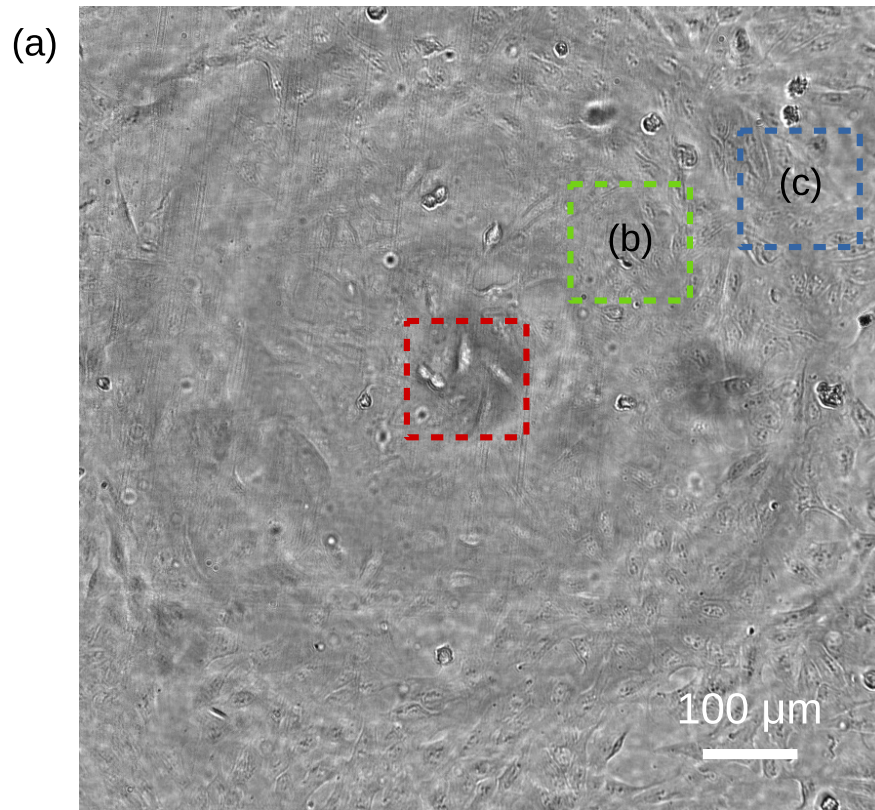
References

1. Kim, J., Henley, B. M., Kim, C. H., Lester, H. A. & Yang, C. Incubator embedded cell culture imaging system (emsight) based on fourier ptychographic microscopy. *Biomed. Opt. Express* 7, 3097–3110, DOI:10.1364/BOE.7.003097 (2016).
2. Yeh, L.-H., Dong, J., Zhong, J., Tian, L., Chen, M., Tang, G., Soltanolkotabi, M. & Waller, L. Experimental robustness of Fourier ptychography phase retrieval algorithms. *Optics Express* 23, 33214, DOI:10.1364/OE.23.033214 (2015).
3. Pan, A., Zhang, Y., Zhao, T., Wang, Z., Dan, D., Lei, M. & Yao, B. System calibration method for fourier ptychographic microscopy. *Journal of Biomedical Optics* 22, DOI:10.1117/1.JBO.22.9.096005 (2017).
4. Hou, L., Wang, H., Sticker, M., Stoppe, L., Wang, J. & Xu, M. Adaptive background interference removal for fourier ptychographic microscopy. *Applied Optics* 57, 1575, DOI:10.1364/AO.57.001575 (2018).
5. Tian, L., Li, X., Ramchandran, K. & Waller, L. Multiplexed coded illumination for fourier ptychography with an led array microscope. *Biomedical Optics Express* 5, 2376, DOI:10.1364/BOE.5.002376 (2014).
6. Ou, X., Zheng, G. & Yang, C. Embedded pupil function recovery for fourier ptychographic microscopy. *Optics Express* 22, 4960, DOI:10.1364/OE.22.004960 (2014).

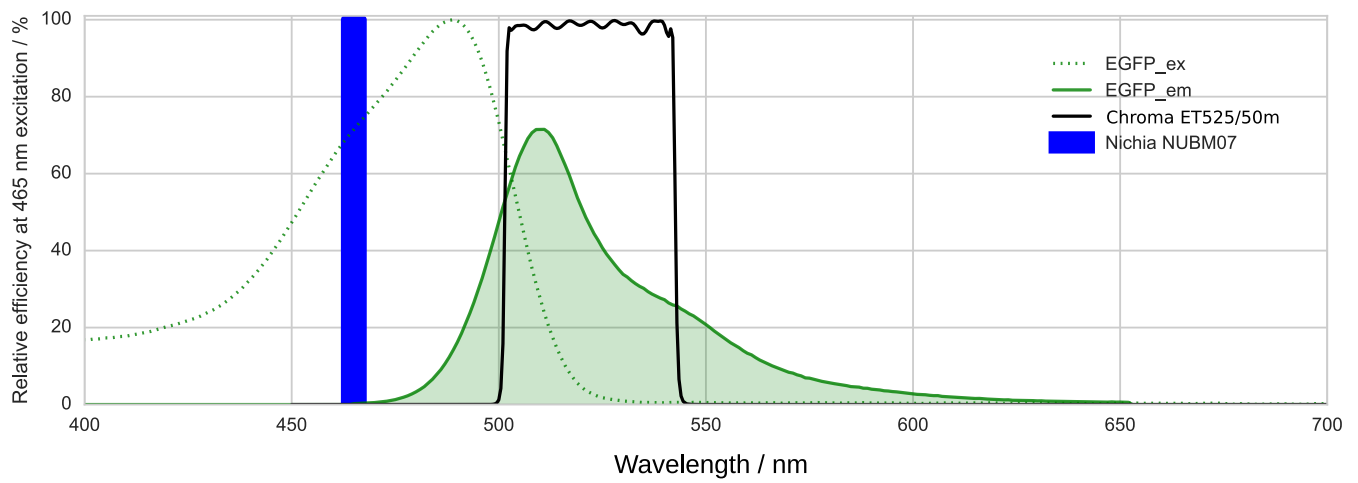


Supplementary Figure S6. Surface flatness of the polystyrene samples, analyzed by the scientific-grade FPM.

7. Widrow, B. Statistical analysis of amplitude-quantized sampled-data systems. *Part II: Applications and Industry Transactions of the American Institute of Electrical Engineers* **79**, 555–568, DOI:[10.1109/TAI.1961.6371702](https://doi.org/10.1109/TAI.1961.6371702) (1961).
8. Schuchman, L. Dither signals and their effect on quantization noise. *IEEE Transactions on Communication Technology* **12**, 162–165, DOI:[10.1109/TCOM.1964.1088973](https://doi.org/10.1109/TCOM.1964.1088973) (1964).
9. Vanderkooy, J. & Lipshitz, S. P. Resolution below the least significant bit in digital systems with dither. *Journal of the Audio Engineering Society* **32**, 106–113 (1984).
10. Roberts, L. Picture coding using pseudo-random noise. *IRE Transactions on Information Theory* **8**, 145–154, DOI:[10.1109/TIT.1962.1057702](https://doi.org/10.1109/TIT.1962.1057702) (1962).
11. Balsam, J., Bruck, H. A., Kostov, Y. & Rasooly, A. Image stacking approach to increase sensitivity of fluorescence detection using a low cost complementary metal-oxide-semiconductor (CMOS) webcam. *Sensors and Actuators, B: Chemical* **171-172**, 141–147, DOI:[10.1016/j.snb.2012.02.003](https://doi.org/10.1016/j.snb.2012.02.003) (2012).
12. Nakamura, J. *Image Sensors and Signal Processing for Digital Still Cameras* (CRC Press, 2005).



Supplementary Figure S7. Computationally refocused phase images at off-axis locations. (a) Raw intensity image of the entire field-of-view of the U2OS cell line. Also shown are the FPM Phase reconstruction (b) halfway from the edge of the field-of-view; and (c) close to the edge of the field-of-view.



Supplementary Figure S8. Spectra of laser, fluorophore (eGFP) and filter set for fluorescence microscopy. The multimode diode laser (Nichia NUBM07) is filtered with a laser clean up filter of a 5 nm bandwidth (Semrock FF01-465/5).