

1 **New Phytologist Supporting Information**
2 **Methods S1, Notes S1, Figs. S1 – S11, and Tables S1 – S13**

3
4 **Article title: Resolving the backbone of the Brassicaceae phylogeny for investigating**
5 **trait diversity**

6
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13 **Methods S1 – Supplementary Materials and Methods**

14 **Plant Material.** A total of 63 species with 65 accessions used in this study are listed in
15 Table S1. Fresh flash-frozen, silica-gel dried, or herbarium material were used for DNA
16 extraction using PowerPlant Pro DNA Isolation Kit (QIAGEN Inc., Hilden, Germany).
17 Three DNA samples were obtained from the Kew DNA collection
18 (<http://apps.kew.org/dnabank/introduction.html>). DNA samples with visible
19 impurities (green or brown tint, and/or flocculate) were additionally purified using
20 QIAquick MinElute columns (QIAGEN Inc., Hilden, Germany). For *Turritis glabra* L.,
21 total RNA was extracted from gibberellic acid (GA₃)-treated seedlings grown under
22 short day conditions using RNeasy Mini Kit (Qiagen, Hilden, Germany). For
23 comparative transcriptomics, eight species with contrasting leaf morphologies (simple
24 leaves – *Aethionema arabicum*, *Arabis alpina*, *Arabidopsis thaliana*, *Boechera stricta*, and
25 complex leaves – *Sisymbrium irio*, *Arabidopsis lyrata*, *Cardamine hirsuta*, *Capsella rubella*)
26 were grown under short day conditions (8 h light (20C)/16 h dark (16C) cycle) in a
27 growth chamber with humidity control. A triplicate of ~30 developing leaves (nodes 5
28 and 6) in an organogenetic stage (100 µm – 500 µm, depending on the species; one leaf
29 per plant) were manually dissected between 10 am and 12 pm, and total RNA was
30 extracted using RNeasy Plus Micro Kit (Qiagen, Hilden, Germany).

31 **Library Preparation and Sequencing.** Each DNA sample was sheared and selected to
32 modal lengths of ~275 bp using a dual-step SPRI bead cleanup. Illumina sequencing
33 libraries were prepared with the NEBNext Fast DNA Library Prep Kit (New England
34 Biolabs, Ipswich, Massachusetts, USA), and amplified using KAPA HiFi HotStart
35 ReadyMix (Kapa Biosystems, Wilmington, Massachusetts, USA) for 6 cycles using the

1 manufacturer's recommended thermal profile and dual P7 and P5 indexing primers (see
2 Kircher *et al.*, 2012 for primer configurations). After SPRI bead purification, 500 ng of
3 each library were enriched using the MYbaits-2 Target Enrichment system (MYcroarray,
4 Ann Arbor, Michigan, USA) following manual version 3.0. Bead-bound libraries were
5 additionally amplified for 12 cycles using universal P5/P7 primers, purified, and
6 combined equimolarly to 1 ng/ μ L for sequencing. 2 x 75 paired end reads were
7 generated on GAIIx (Illumina, San Diego, California, USA) in two separate runs at the
8 Cologne Center for Genomics at the University of Cologne, and the raw reads have
9 been deposited at the National Center for Biotechnology Information (NCBI) Sequence
10 Read Archive (Bioproject PRJNA518905).

11 Transcriptome libraries were prepared after mRNA enrichment with NEBNext Poly(A)
12 mRNA Magnetic Isolation Module (New England Biolabs, Ipswich, Massachusetts,
13 USA) with NEBNext Ultra™ Directional RNA Library Prep Kit (New England Biolabs,
14 Ipswich, Massachusetts, USA) for Illumina. Libraries were sequenced on HiSeq3000
15 (Illumina, San Diego, California, USA) as 2 x 150 paired end runs at the Genome Center
16 of the Max Planck Institute for Plant Breeding Research, Cologne, Germany, and raw
17 reads have been deposited at the NCBI Sequence Read Archive (Bioproject
18 PRJNA518905).

19 ***Obtaining corresponding exons from sequenced genomes.*** To complement our *de novo*
20 derived sequences, we included publicly available genome data from 16 Brassicaceae
21 species and two outgroups (Table S2). Their genomes were queried with the exon sets
22 for each of the three reference species using translated BLAT alignment (highest scoring
23 hit was retained if the number of called positions divided by the size of the reference
24 exons was greater than 40%). The target exons derived from each of the three references
25 were oriented and consensus was called after alignment with MAFFT 6.851b (Kato *et*
26 *al.*, 2002); non-overlapping fragments recovered from the different references were
27 discarded. This resulted in 1748 shared exons (out of 1827 target Brassicaceae exons)
28 with the immediate outgroup *Tarrenaya hassleriana* and 1101 shared exons with the more
29 distantly related outgroup *Carica papaya*. Plastid-coding sequences were obtained from
30 sequenced plastomes (Table S3) following the same protocol.

31

1 **Notes S1**

2 **Tree description**– Consistent with previous analyses (Beilstein *et al.*, 2006; Huang *et al.*,
 3 2015), *Aethionema* (Aethionemeae) is sister to the rest of Brassicaceae. Following
 4 *Aethionema*, the next branching event in the phylogeny separates representatives of the
 5 traditionally recognized Lineage III (clade E, Huang *et al.*, 2015) from the rest, with
 6 placement and composition universally supported across all nuclear datasets and
 7 analyses. Within Lineage III, the basal split included the sister taxa *Dontostemon*
 8 *integrifolius* (Dontostemoneae) and *Chorispora tenella* (Chorisporae) and their sister
 9 clade was composed of a grade of *Hesperis matronalis* (Hesperideae), *Neotorularia torulosa*
 10 (Euclidieae), *Bunias orientalis* (Buniadeae), and *Matthiola incana* (Anchonieae) + the
 11 unassigned to a tribe *Veselskya griffithiana*. The placements of *Hesperis matronalis* and
 12 *Neotorularia torulosa* alternated in the plastid phylogeny (Fig. S9), as reported in
 13 Mandáková *et al.*, 2017.

14 The next Brassicaceae clade, which we call Lineage IV (clade D, Huang *et al.*, 2015), is
 15 universally supported in all nuclear datasets and analyses, and consists of *Arabis alpina*
 16 (Arabideae) + *Pseudoturritis turrita* (Stevenieae), which are sisters to *Aurinia saxatilis*
 17 (Alysseae) + the unassigned to a tribe *Asperuginoides axillaris*. This clade includes the
 18 first (Arabideae) and the third (Alysseae) largest tribes in the family, harbors one-fifth
 19 of the family's species diversity, and has a center of diversity in the Mediterranean and
 20 the Irano-Turanian regions (Karl & Koch, 2013; Rešetnik *et al.*, 2013).

21 The next clade consists of representatives of the traditionally recognized Lineage I
 22 (clade A excluding *Macropodium* in Huang *et al.*, 2015). Biscutelleae (Özüdođru *et al.*,
 23 2017), which included *Biscutella laevigata* sister to *Heldreichia bupleurifolia* and *Lunaria*
 24 *rediviva* sister to *Ricotia cretica*, were strongly supported and tentatively placed as sister
 25 to Lineage I in the maximum likelihood analyses after removing genes that contribute
 26 disproportionately to the phylogenetic signal. The earliest split in Lineage I includes (1)
 27 Cardamineae (*Cardamine hirsuta*, *Leavenworthia alabamica*, and the unassigned to a tribe
 28 *Andrzeiowskia cardamine*), followed by (2) Lepidieae (*Lepidium hirtum* and *Lepidium*
 29 *meyenii*), (3) a clade consisting of *Descurainia artemisioides* (Descurainieae) sister to
 30 *Yinshania acutangula* (Yinshanieae) + *Smelowskia calycina* (Smelowskieae), (4) a clade of
 31 *Erysimum cheiri* (Erysimeae) + *Malcolmia maritima* (Malcolmieae), (5) the *Arabidopsis*
 32 segregate of Camelieae, (6) *Physaria purpurea* (Physarieae), (7) a clade of *Murbeckiella*

1 *pinnatifida* (Oreophytoneae) sister to *Turritis glabra* (Turritideae) + *Alyssopsis mollis*
 2 (Alyssopsidae), followed by (8) *Capsella rubella* and *Camelina sativa* (both Camelinae),
 3 (9) *Geococcus pusillus* (Microlepidieae) + the unassigned to a tribe *Hemilophia rockii* and
 4 *Dipoma iberideum*, and (10) *Boechera stricta* (Boechereae), *Halimolobos pubens*
 5 (Halimolobeae), and *Crucihimalaya himalaica* (Crucihimalayae). The support for this
 6 branching order was robust among analyses, with the notable conflict for the
 7 relationship among the last three clades between coalescent and concatenation analyses.
 8 We confirm the polyphyly of Camelinae based on nuclear markers (Bailey *et al.*, 2006;
 9 Beilstein *et al.*, 2006), as well as the close relationship between *Malcolmia maritima* and
 10 *Erysimum cheiri* (Moazzeni *et al.*, 2014). In contrast to prior conflicting positions for
 11 Lepidieae (Kagale *et al.*, 2014; Huang *et al.*, 2015), our placement immediately after the
 12 divergence of Cardamineae is robust and universally supported in all nuclear analyses.
 13 *Lepidium* position only became unstable after removing data from our most exclusive
 14 dataset (not shown).

15 Another major clade of Brassicaceae includes relatives of the traditionally recognized
 16 Lineage II and some members of the 'expanded' Lineage II, which is sister to a clade
 17 consisting of *Megacarpaea* spp. (Megacarpaeae) and *Lobularia maritima* (Anastaticae) +
 18 *Cochlearia officinalis* (Cochlearieae) + *Iberis linifolia* (Iberideae). This resolution is
 19 supported in the maximum likelihood analyses after excluding genes that contribute
 20 disproportionately to the phylogenetic signal but unstable in the taxon-pruning
 21 experiments. The position of *Cochlearia officinalis* differs among the maximum likelihood
 22 analyses; however, it is resolved as a successive sister to *Kernera saxatilis* + *Petrocallis*
 23 *pyrenaica* preceding the *Conringia* clade in all ASTRAL-II analyses. The branching order
 24 of Lineage II includes a grade of (1) *Aphragmus bouffordii* (Aphragmeae), (2) *Kernera*
 25 *saxatilis* (Kernereae) + the unassigned to a tribe *Petrocallis pyrenaica*, (3) the *Conringia*
 26 clade, consisting of *Conringia orientalis* (Conringieae) + *Noccaea vesicaria*
 27 (Coluteocarpeae) + the unassigned to a tribe *Arabis ottonis-schulzii*, (4) *Calepina irregularis*
 28 (Calepineae), (5) a clade including *Eutrema salsugineum* (Eutremeae) sister to *Peltaria*
 29 *alliacea* + *Thlaspi arvense* (both Thlaspideae) and *Camelinopsis campylopoda*, (6) the
 30 unassigned to a tribe *Schrenkiella parvula*, (7) a clade consisting of *Bivonaea lutea*
 31 (Bivonaeae) sister to the unassigned to a tribe taxa *Arabis josiae* + *Fourrea alpina*, (8)
 32 *Brassica* spp. (Brassicaceae), (9) *Isatis tinctoria* (Isatideae) + the unassigned to a tribe

1 *Conringia planisiliqua*, (10) *Stanleya elata* (Thelypodieae) and the unassigned to a tribe
2 *Raphanorhyncha crassa*, and (11) *Sisymbrium irio* (Sisymbrieae) sister to *Pseudofortuynia*
3 *esfandiarii* and the unassigned to a tribe *Ochthodium aegyptiacum*. Many of these
4 relationships are recovered in the plastid phylogeny, which also removes
5 *Orychophragmus* spp., a taxon not sampled for nuclear markers, from other Brassiceae.
6 Applying our phylogenomic approach at the infratribal level will help identify such
7 divergent lineages in other tribes and is an obvious next step towards describing the
8 global diversity of Brassicaceae.

9 Our results strongly support a novel clade, Lineage V, which includes 179 species in 21
10 genera primarily distributed in the Southern Hemisphere. The four exceptions are
11 *Idahoia scapigera* (North America), *Asta shaffneri* and *Scoliaxon mexicanus* (Mexico), and
12 *Subularia aquatica* (northern North American and northern Eurasia). This lineage
13 includes two monophyletic sister groups, (1) the New Zealand *Notothlaspi australe*
14 (Notothlaspidiae) sister to the South African *Heliophila coronopifolia* (Heliophileae) + the
15 unassigned to a tribe *Chamira circaeoides* and (2) the Mexican clade *Asta shaffneri* (Asteae)
16 + *Scoliaxon mexicanus* (Scoliaxoneae) and sister to the South American CES clade
17 (Salariato *et al.*, 2016) *Bryopsis diapensioides* (Eudemeae) + (*Cremolobus peruvianus*
18 (Cremolobeae) + *Schizopetalon walkeri* (Schizopetaleae)). The second lineage also
19 includes the unassigned to a tribe *Subularia aquatica* and *Idahoia scapigera*, which are
20 sometimes placed together as sisters to the rest in the maximum likelihood trees.
21 Although the circumboreal distribution of *S. aquatica* appears exceptional in this clade,
22 its sister species, *Subularia monticola*, is distributed in the highest elevation habitats of
23 Africa, the Ethiopian highlands and Mount Kilimanjaro in Tanzania, and may represent
24 the ancestral range of the genus. All species in Lineage V have exclusively simple
25 trichomes or are glabrous (except Schizopetaleae and *Scoliaxon*, which feature dendritic
26 trichomes). All have dehiscent fruits except four species of *Heliophila*, within which
27 indehiscent fruits evolved three times. None has auriculate leaves except for two species
28 of *Heliophila*, within which this feature evolved once. None of the species has leaves
29 dissected into leaflets, though entire, dentate, and deeply pinnately (but not palmately)
30 divided leaves are present. *Asta* and *Scoliaxon* in the Mexican clade share a unique
31 feature not found in the other ca. 4000 species of Brassicaceae, the presence of large,
32 ovate, white gland pairs at leaf and sometimes pedicel base. Because both genera form a

1 strongly supported clade in all analyses, the tribe Scoliaxoneae should be united with
2 the earlier-published Asteae.

3 **Placing taxa unassigned to a tribe**– Our sampling of Brassicaceae lineages allows
4 confirming previously suggested relationships and associating unassigned to a tribe
5 orphan taxa with related clades or tribes. The divergent positions of *Schrenkiella parvula*,
6 *Subularia aquatica* (and likely *S. monticola*), and *Idahoa scapigera* justify the erection of new
7 tribes, and the name Subularieae is available; new tribal names should be proposed for
8 *Idahoa* and *Schrenkiella* (Table S9). New tribes can be proposed for *Dipoma iberideum* +
9 *Hemilophia rockii* and *Arabis josiae* + *Fourrea alpina*, which are consistently recovered as
10 sister taxa. *Petrocallis pyrenaica* can be included in Kernereae. In other cases, additional
11 sampling of the closest relatives of unassigned to a tribe taxa may provide resolution.
12 More exhaustively sampled *Sisymbrium* is needed to establish its boundaries with
13 respect to *Pseudofortuynia esfandiarii* and *Ochthodium aegyptiacum* (Khosravi *et al.*, 2009).
14 Recent taxonomic work in the group proposed an expanded concept for *Sisymbrium* and
15 synonymization of *Pseudofortuynia esfandiarii* with *Sisymbrium leucocladum* (German &
16 Al-Shehbaz, 2018), raising interesting questions about the fruit evolution in this group.
17 Broader sampling in Coluteocarpeae is similarly needed to confirm the sister position of
18 *Arabis ottonis-schultzii*. Although *Chamira circaeoides* appears sister to *Heliophila*
19 *cornopifolia* in our analyses and has been previously chosen as an outgroup in infratribal
20 phylogenies of Heliophileae (Mandáková *et al.*, 2012), broader sampling within
21 *Heliophila*, an exceptionally diverse genus, is necessary to test whether *Chamira* could be
22 nested within *Heliophila* or placed as a distinct genus in the same tribe.

23 To test whether some of the newly placed taxa fell outside of the variation of their sister
24 tribes, we reconstructed the phylogenies of the respective tribes using broader taxon
25 sampling (Fig. S11, Table S8). *Asperuginioides axillaris* falls outside of the molecular
26 variation of the tribe Alysseae (Rešetnik *et al.*, 2013) based on the plastid marker *ndhF*
27 and additional taxon sampling in this part of the tree is necessary to determine its
28 placement. *Vesselskya griffithiana* is placed as sister to Anchoneae with *rbcl*, although
29 critical taxa (e.g., *Iskandera* spp.; Warwick *et al.*, 2007) were not sampled. *Andrzejowskia*
30 *cardamine* belongs to Cardamineae and is sister to *Barbarea verna* (in the plastid
31 phylogeny) or *Leavenworthia alabamica* (in the nuclear dataset). *Raphanorhyncha crassa*,
32 which is only known from its type specimen (Al-Shehbaz, 2012), is placed within

1 Thelypodieae but its placement is not firmly resolved with the molecular marker *ndhF*.
 2 Based on the high degree of sequence polymorphism we observed, it is likely of hybrid
 3 origin. *Conringia planisiliqua* does not belong to Conringieae but is instead a member of
 4 Isatideae in the clade of *Isatis tinctoria*, a finding supported on morphological grounds
 5 (except fruit type). *Camelinopsis campylopoda* (now *Pseudocamelina campylopoda*) is firmly
 6 placed within Thlaspidaeae and most closely related to *Thalspi arvensis* based on *ndhF*, a
 7 finding recently reported by Esmailbegi *et al.*, 2017; Esmailbegi *et al.*, 2018.

8

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 12 nuclear ribosomal ITS DNA sequences. *Ann Missouri Bot Gard* **94(1)**: 56–78.
 13

14 **Supplementary Figure Legends**

15
 16 **Figure S1. Phylogenetic reconstruction workflow.**

17
 18 **Figure S2. Plastid coding sequences and sequenced plastid genomes obtained from**
 19 **skimming. a,** Coverage among species. Complete plastid genomes were assembled for
 20 several species; maps for three of these (asterisks) are shown below. Note that *rps16*
 21 (yellow bar) has been lost in *Subularia aquatica*. **b,** Annotated plastid genome maps for
 22 *Arabis ottonis-schulzii*, *Pseudofortuynia esfandiarii* , and *Subularia aquatica* .
 23

24 **Figure S3. Density plots of metrics used to partition the exon set into 27 datasets after**
 25 **excluding the extreme outliers for each metric** (grey shading in **a, c,** and **d**). **a,** Long-
 26 branch (*LB*) attraction score, *LB* upper quartile. Exons with *LB* score >60 were excluded,
 27 the rest were partitioned into three increasingly exclusive sets, L1, L2, and L3, with
 28 cutoffs 60, 45, and 37. **b,** Heat map of *LB* attraction score of exons across taxa. Species
 29 and exons with a higher score (shades of green) are prone to long-branch attraction. **c,**
 30 Exon-specific saturation score (*S*), saturation R^2 . Exons with *S* score <0.6 were excluded,
 31 the rest were partitioned into three increasingly exclusive sets, S1, S2, and S3, with
 32 cutoffs 0.6, 0.7, and 0.85. **d,** Tree distances measured as ‘matching splits’ (*MS*). Exons
 33 with *MS* score >250 were excluded, the rest were partitioned into three increasingly
 34 exclusive sets, M1, M2, and M3, with cutoffs 250, 190, and 120. The inset depicts the tree
 35 space used to derive the tree distances.
 36

37 **Figure S4. Summary of the concatenation results represented by the tree space of the**
 38 **eight unique topologies (circles) from the maximum likelihood analysis, and the**

1 **topological differences among them.** The summary tree on top is derived from the
 2 most inclusive dataset, *S1L1M1*, and color-coding marks the branches on the tree that
 3 are alternatively resolved with the other datasets. The tree space is divided into two
 4 areas (blue dotted line) based on the position of *Idahoa* and *Subularia* with respect to the
 5 Mexican (*Asta* + *Scoliaxon*) + CES (*Cremolobus* + *Schizopetalon* + *Brayopsis*) clade. It is
 6 further divided into two areas (orange dotted line) based on the position of the
 7 Biscutelleae + (*Lobularia* + *Iberis*) clade as sister to either *Megacarpaea* or Lineage I.

8
 9 **Figure S5. Phylogenetic informativeness of the loci from the most inclusive datasets**
 10 **that produced unique topologies (*S1L1M1*, *S1L1M3*, *S1L2M2*, *S1L3M1*, *S1L3M3*,**
 11 ***S2L2M1*, *S3L1M1*, *S3L3M1*).** Exons with phylogenetic informativeness >10 deemed
 12 strong contributors were excluded from the subsequent concatenation analyses.

13
 14 **Figure S6. Summary of the concatenation results after excluding exons with**
 15 **phylogenetic informativeness score >10. a,** The summary tree in the upper left corner
 16 is derived from the most inclusive dataset, *S1L1M1R*, and color-coding marks the
 17 branches on the tree that are alternatively resolved with the other datasets. **b,** Tree
 18 distances derived from the maximum likelihood analyses before (blue) and after
 19 (yellow) exclusion of the strong contributors from the phylogenetic informativeness test
 20 mapped onto two parallel planes.

21
 22 **Figure S7. Pruning of taxa with unstable placement in different datasets and**
 23 **analyses.** Relationships among *Aethionema*, Lineage III and Lineage IV did not vary
 24 among datasets and analyses and these are not represented. **a,** Excluding *Idahoa*
 25 *scapigera*. **b,** Excluding *Subularia aquatica*. **c,** Excluding *Cochlearia officinalis*. **d,** Excluding
 26 *Megacarpaea* spp. **e,** Excluding *Lobularia maritima*. **f,** Excluding *Iberis linifolia*.

27
 28 **Figure S8. Summary of the ASTRAL-II results represented by the tree space of the**
 29 **seven unique topologies (circles), and the differences among them.** The summary tree
 30 on top is derived from the most inclusive dataset, *S1L1M1*, and color-coding marks the
 31 branches on the tree that are alternatively resolved with the other datasets.

32

1 **Figure S9. Placing unassigned to a tribe species with expanded taxon sampling of**
2 **associated sister tribes. a, *Asperuginoides axillaris*. b, *Raphanorhyncha crassa*. c, *Veselskya***
3 ***griffithiana*. d, *Camelinopsis campylopoda*. e, *Conringia planisiliqua*.**

4

5 **Figure S10. Plastid coding gene phylogeny (same tree depicted without and with**
6 **branch transformation). Bootstrap support is 100 unless indicated otherwise.**

7

8 **Figure S11. Comparison between tree topologies derived from nuclear and plastid**
9 **loci. Unmatched taxa between the two datasets are excluded, and multispecies genera**
10 **(except *Arabis* spp. and *Conringia* spp.) are trimmed to a single species.**

11

12

13 **Supplementary Tables**

14 **Table S1.** Taxon and voucher information for species with newly generated sequence
15 data.

16

17 **Table S2.** Accession information for species with sequenced nuclear genomes included
18 in this study.

19

20 **Table S3.** Accession information for species with sequenced plastid genomes.

21

22 **Table S4.** Sequencing reads and contig statistics for nuclear loci.

23

24 **Table S5.** Skimmed reads and whole plastome statistics.

25

26 **Table S6.** Concatenated dataset statistics.

27

28 **Table S7.** AU tests to evaluate the support of a given matrix for a given topology.

29 Values of support are color coded, with highest support in blue and lowest support in
30 red. Note that many of the unique topologies are indistinguishable based on the data.

31

- 1 **Table S8.** Accession information of marker genes used for the infratribal placement of
2 unassigned to a tribe taxa.
3
- 4 **Table S9.** Summary of the phylogenetic placement of unassigned to a tribe taxa and
5 taxonomic notes.
6
- 7 **Table S10.** Transcriptome assembly statistics.
8
- 9 **Table S11.** Genes that deviate from the background optimal expression value in the
10 family in the core Brassicaceae.
11
- 12 **Table S12.** Genes that deviate from the background optimal expression value in the
13 family along the branches leading to species with complex leaves.
14
- 15 **Table S13.** Genes and protein residues under positive selection in species with complex
16 leaves.

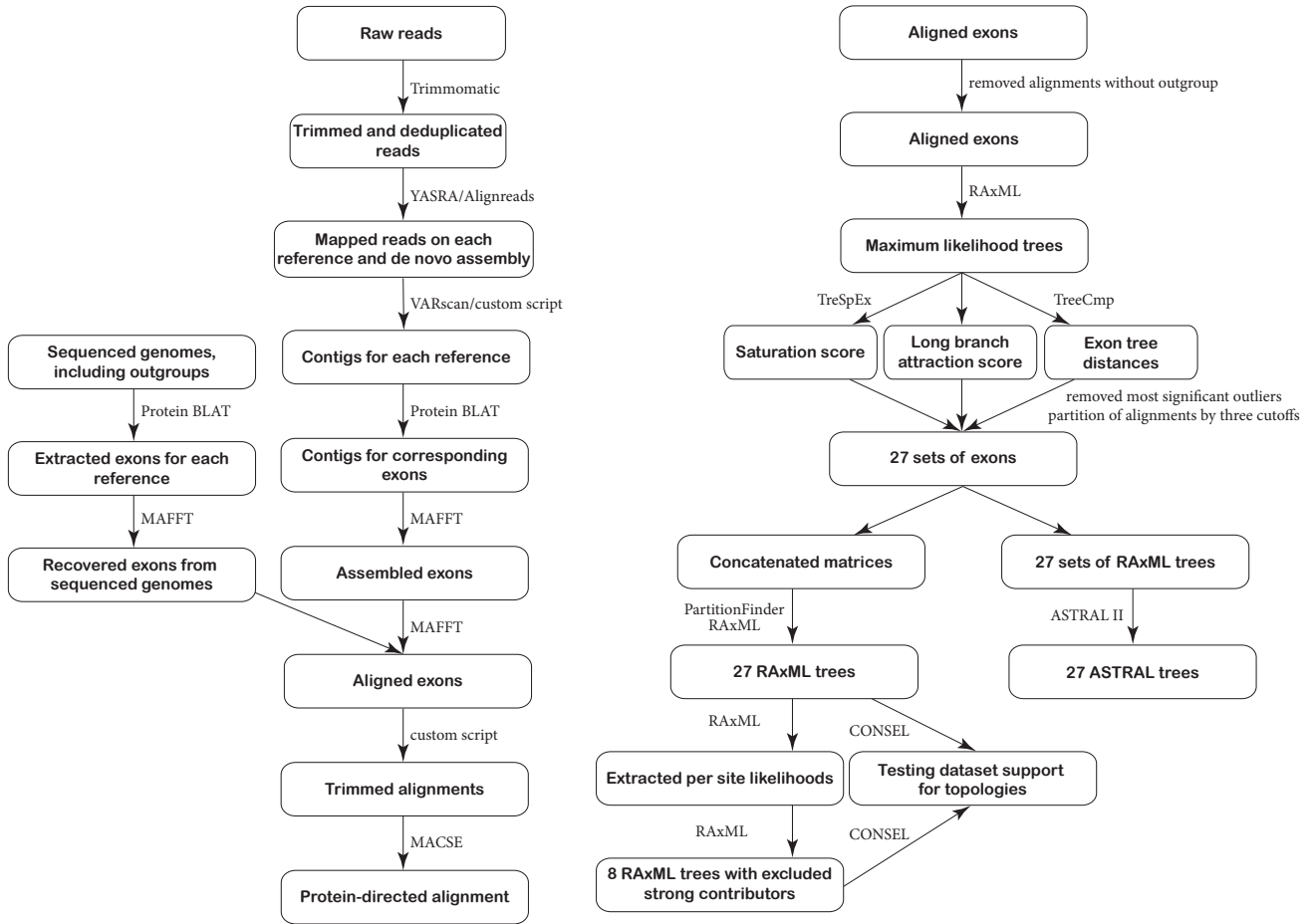


Figure S1. Phylogenetic reconstruction workflow.

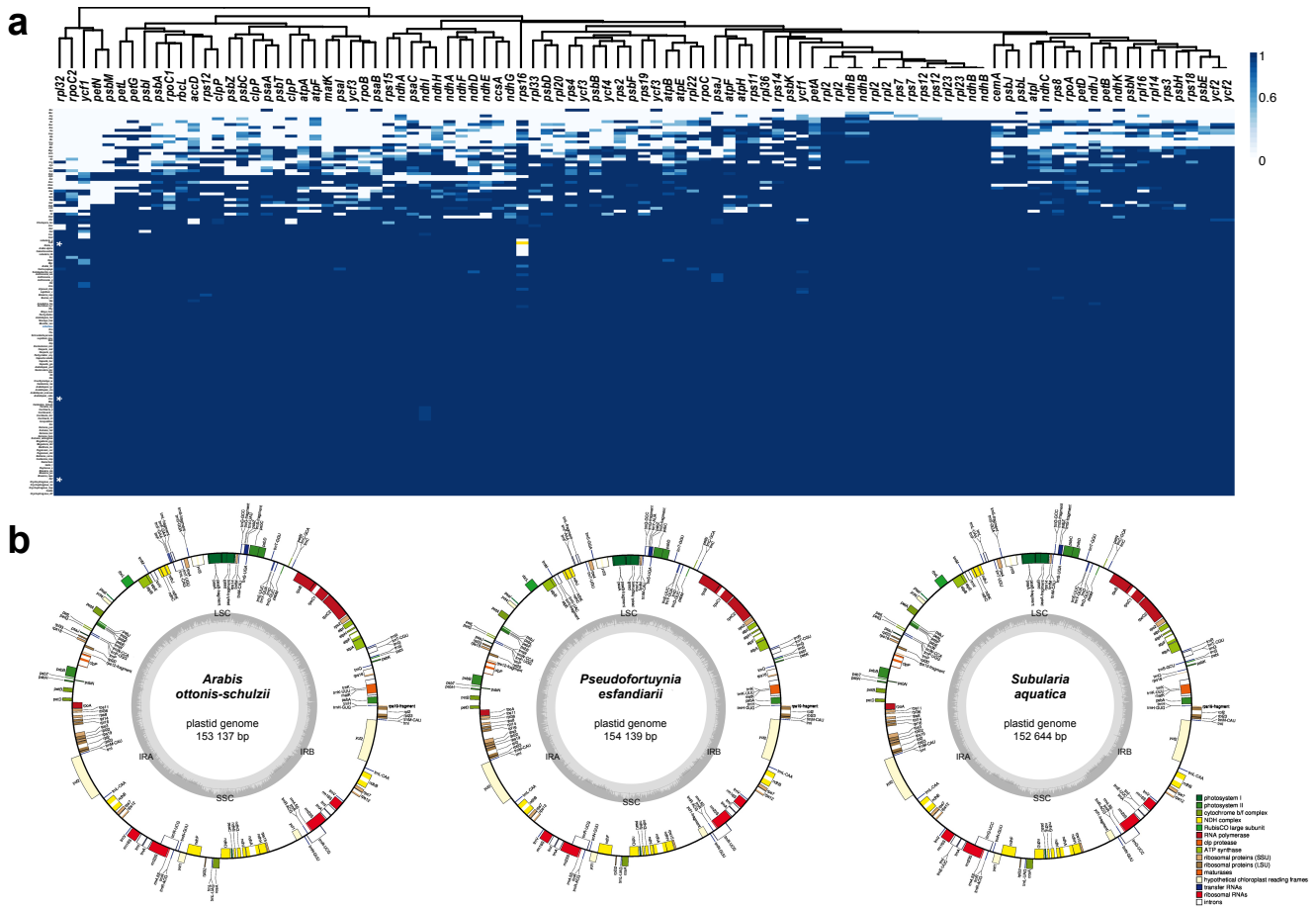


Figure S2. Plastid coding sequences and sequenced plastid genomes obtained from skimming. a, Coverage among species. Complete plastid genomes were assembled for several species; maps for three of these (asterisks) are shown below. Note that *rps16* (yellow bar) has been lost in *Subularia aquatica*. **b**, Annotated plastid genome maps for *Arabis ottonis-schulzii*, *Pseudofortuynia esfandiarrii*, and *Subularia aquatica*.

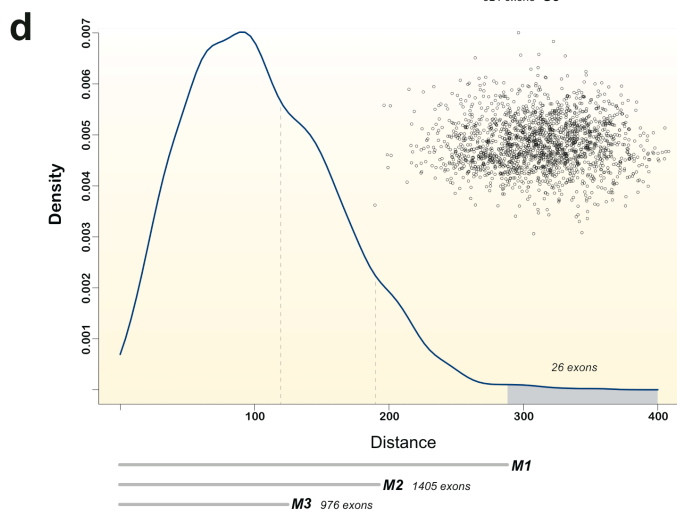
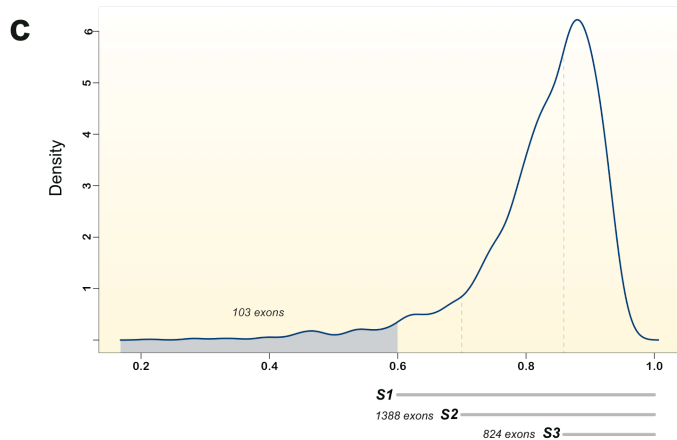
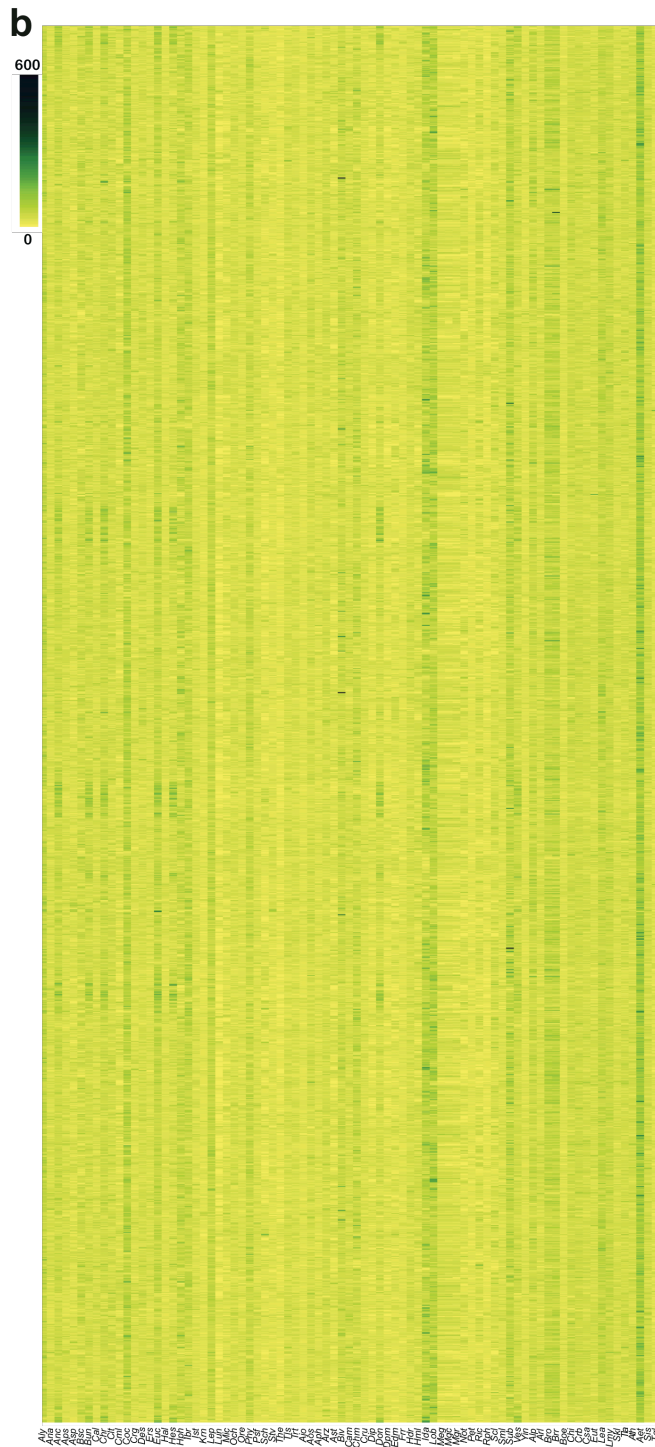
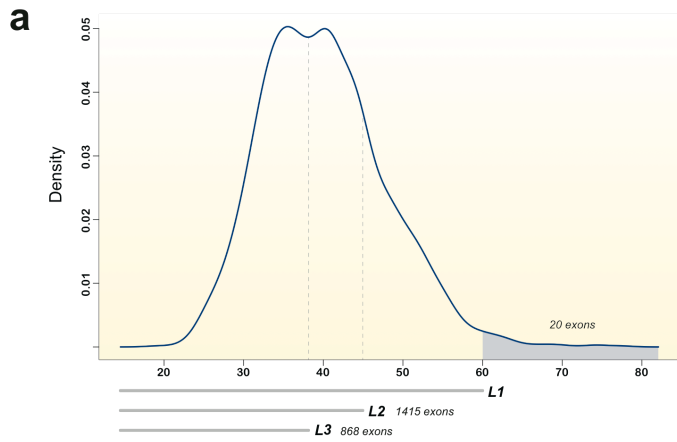


Figure S3. Density plots of metrics used to partition the exon set into 27 datasets after excluding the extreme outliers for each metric (grey shading in a, c, and d). a, Long-branch (*LB*) attraction score, *LB* upper quartile. Exons with *LB* score >60 were excluded, the rest were partitioned into three increasingly exclusive sets, L1, L2, and L3, with cutoffs 60, 45, and 37. **b,** Heat map of *LB* attraction score of exons across taxa. Species and exons with a higher score (shades of green) are prone to long-branch attraction. **c,** Exon-specific saturation score (*S*), saturation R^2 . Exons with *S* score <0.6 were excluded, the rest were partitioned into three increasingly exclusive sets, S1, S2, and S3, with cutoffs 0.6, 0.7, and 0.85. **d,** Tree distances measured as 'matching splits' (*MS*). Exons with *MS* score >250 were excluded, the rest were partitioned into three increasingly exclusive sets, M1, M2, and M3, with cutoffs 250, 190, and 120. The inset depicts the tree space used to derive the tree distances.

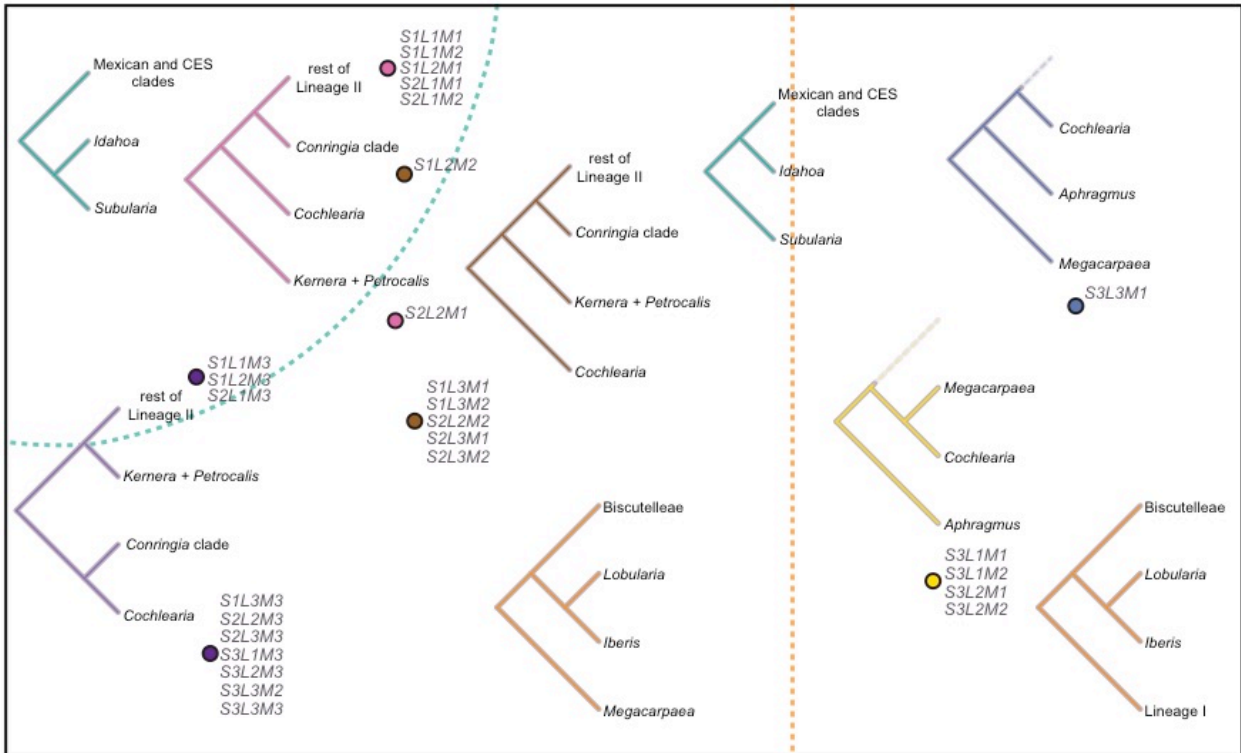
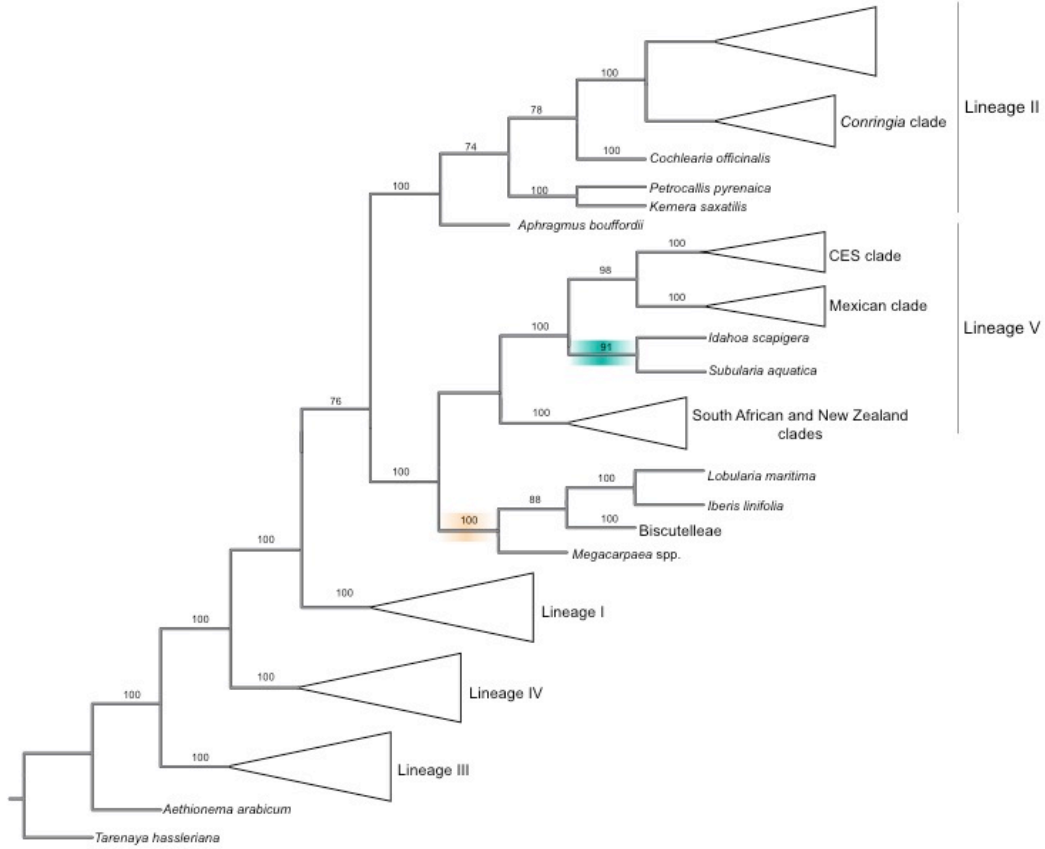


Figure S4. Summary of the concatenation results represented by the tree space of the eight unique topologies (circles) from the maximum likelihood analysis, and the topological differences among them. The summary tree on top is derived from the most inclusive dataset, *S1L1M1*, and color-coding marks the branches on the tree that are alternatively resolved with the other datasets. The tree space is divided into two areas (blue dotted line) based on the position of *Idahoa* and *Subularia* with respect to the Mexican (*Asta* + *Scoliaxon*) + CES (*Cremolobus* + *Schizopetalon* + *Brayopsis*) clade. It is further divided into two areas (orange dotted line) based on the position of the Biscutelleae + (*Lobularia* + *Iberis*) clade as sister to either *Megacarpaea* or Lineage I.

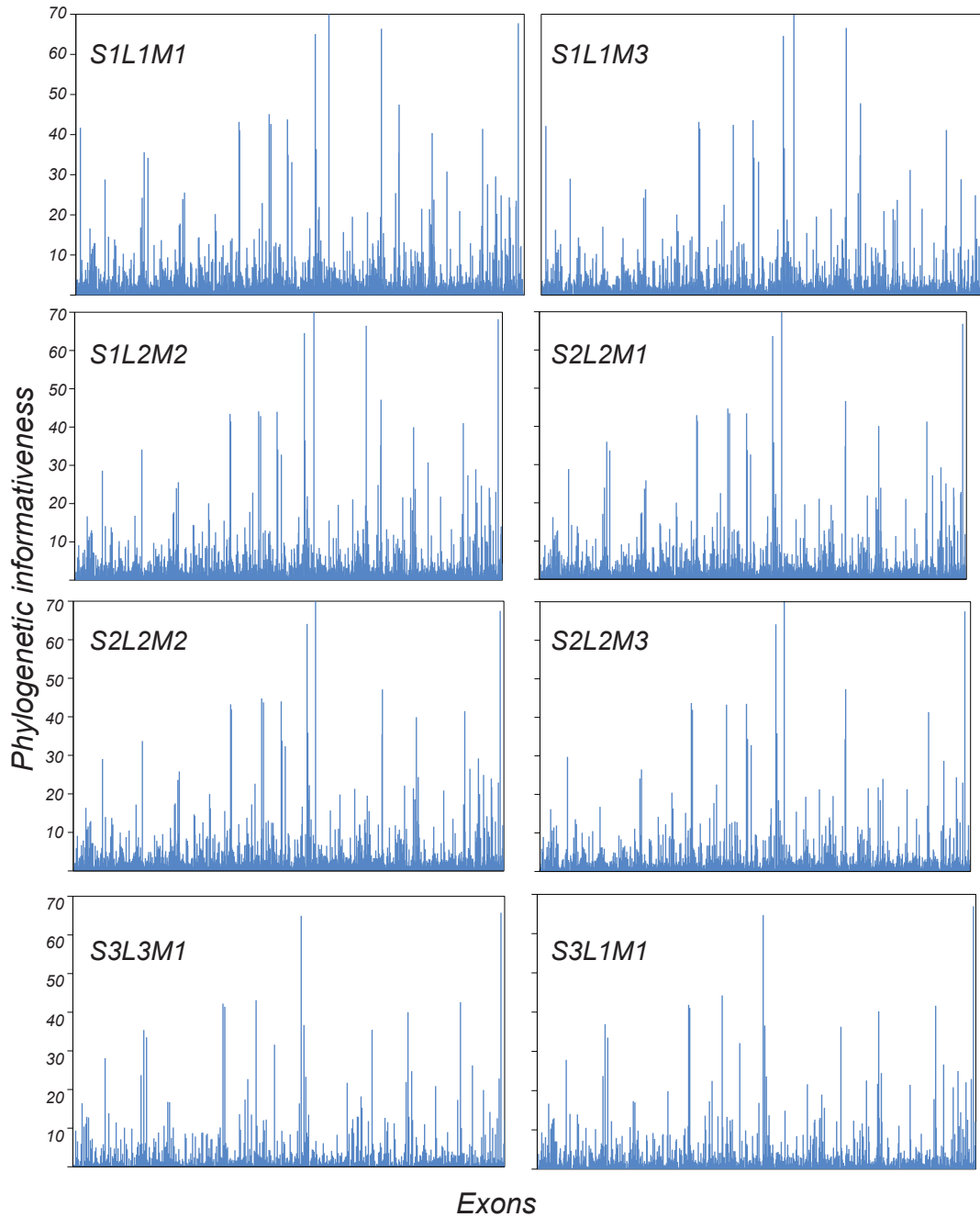


Figure S5. Phylogenetic informativeness of the loci from the most inclusive datasets that produced unique topologies (*S1L1M1*, *S1L1M3*, *S1L2M2*, *S1L3M1*, *S1L3M3*, *S2L2M1*, *S3L1M1*, *S3L3M1*). Exons with phylogenetic informativeness >10 deemed strong contributors were excluded from the subsequent concatenation analyses.

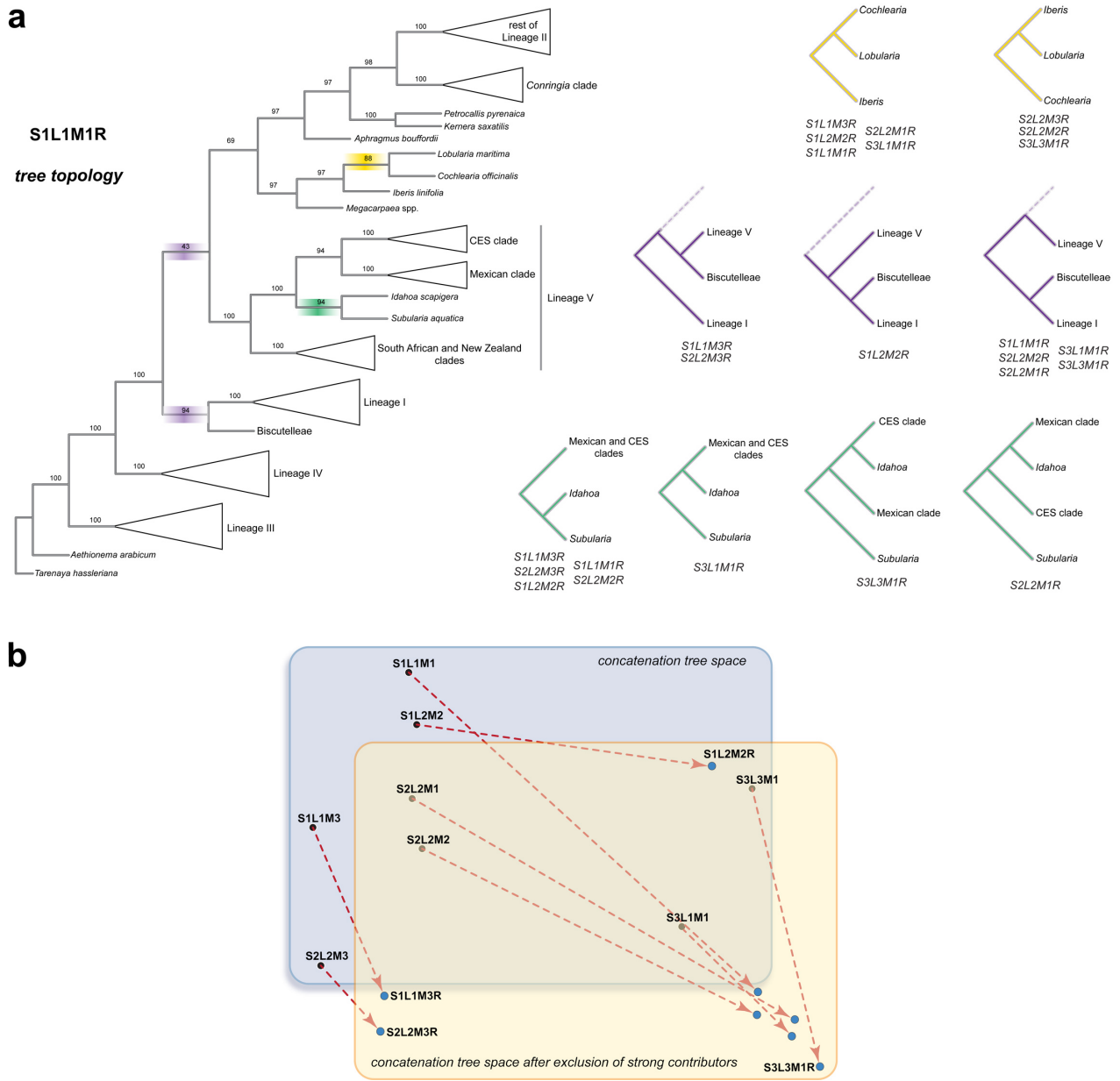


Figure S6. Summary of the concatenation results after excluding exons with phylogenetic informativeness score >10. **a**, The summary tree in the upper left corner is derived from the most inclusive dataset, *S1L1M1R*, and color-coding marks the branches on the tree that are alternatively resolved with the other datasets. **b**, Tree distances derived from the maximum likelihood analyses before (blue) and after (yellow) exclusion of the strong contributors from the phylogenetic informativeness test mapped onto two parallel planes.

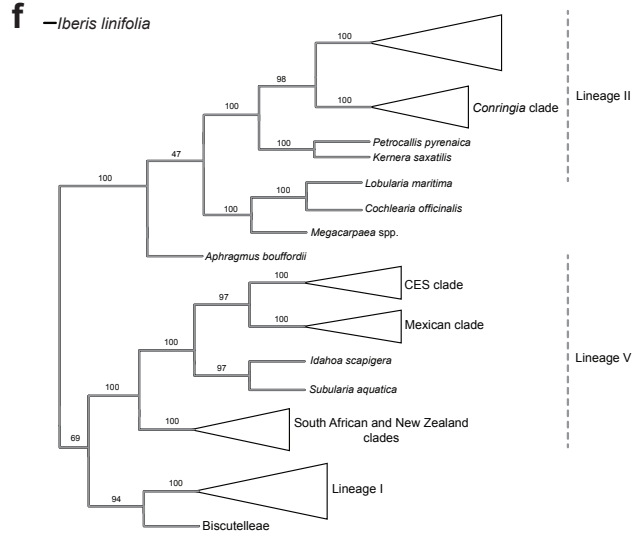
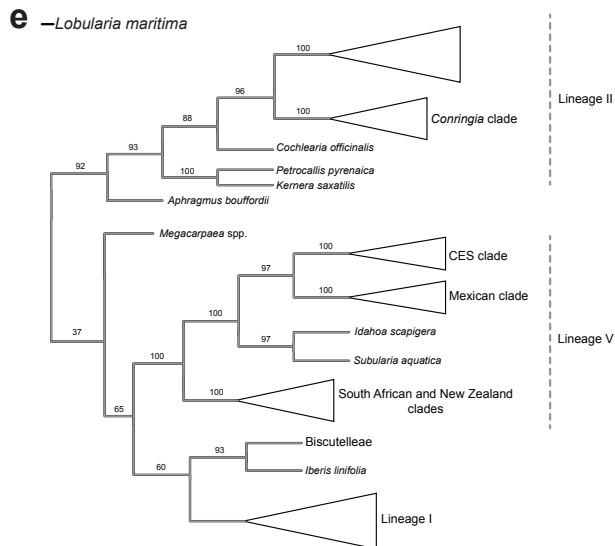
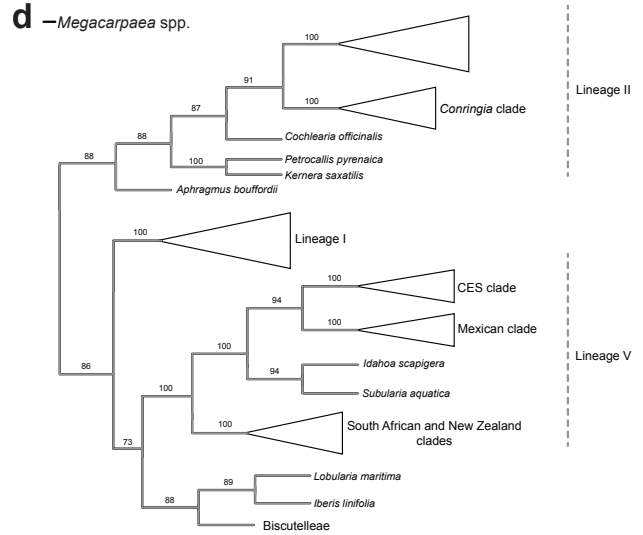
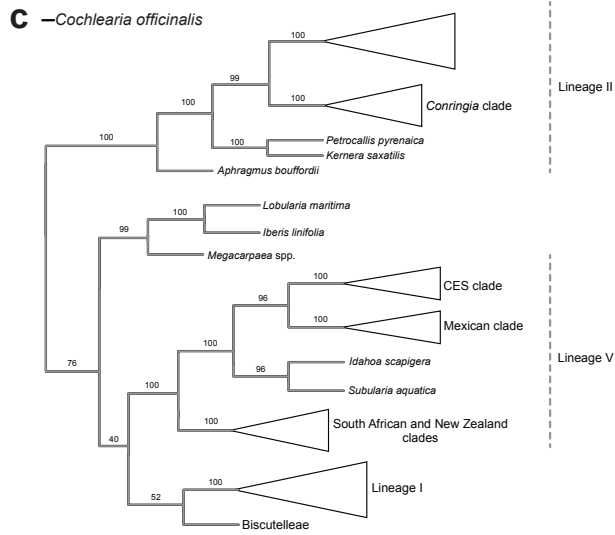
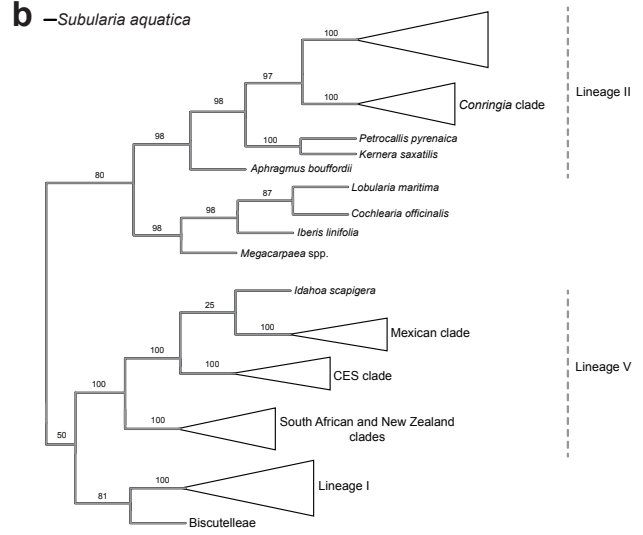
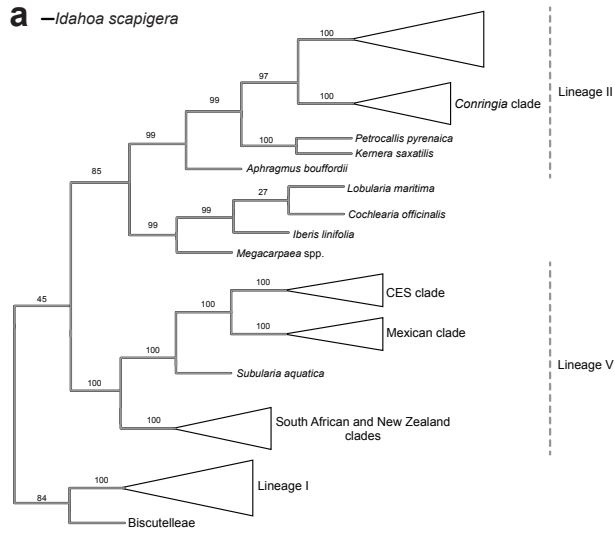


Figure S7. Pruning of taxa with unstable placement in different datasets and analyses. Relationships among *Aethionema*, Lineage III and Lineage IV did not vary among datasets and analyses and these are not represented. **a**, Excluding *Idahoa scapigera*. **b**, Excluding *Subularia aquatica*. **c**, Excluding *Cochlearia officinalis*. **d**, Excluding *Megacarpaea* spp. **e**, Excluding *Lobularia maritima*. **f**, Excluding *Iberis linifolia*.

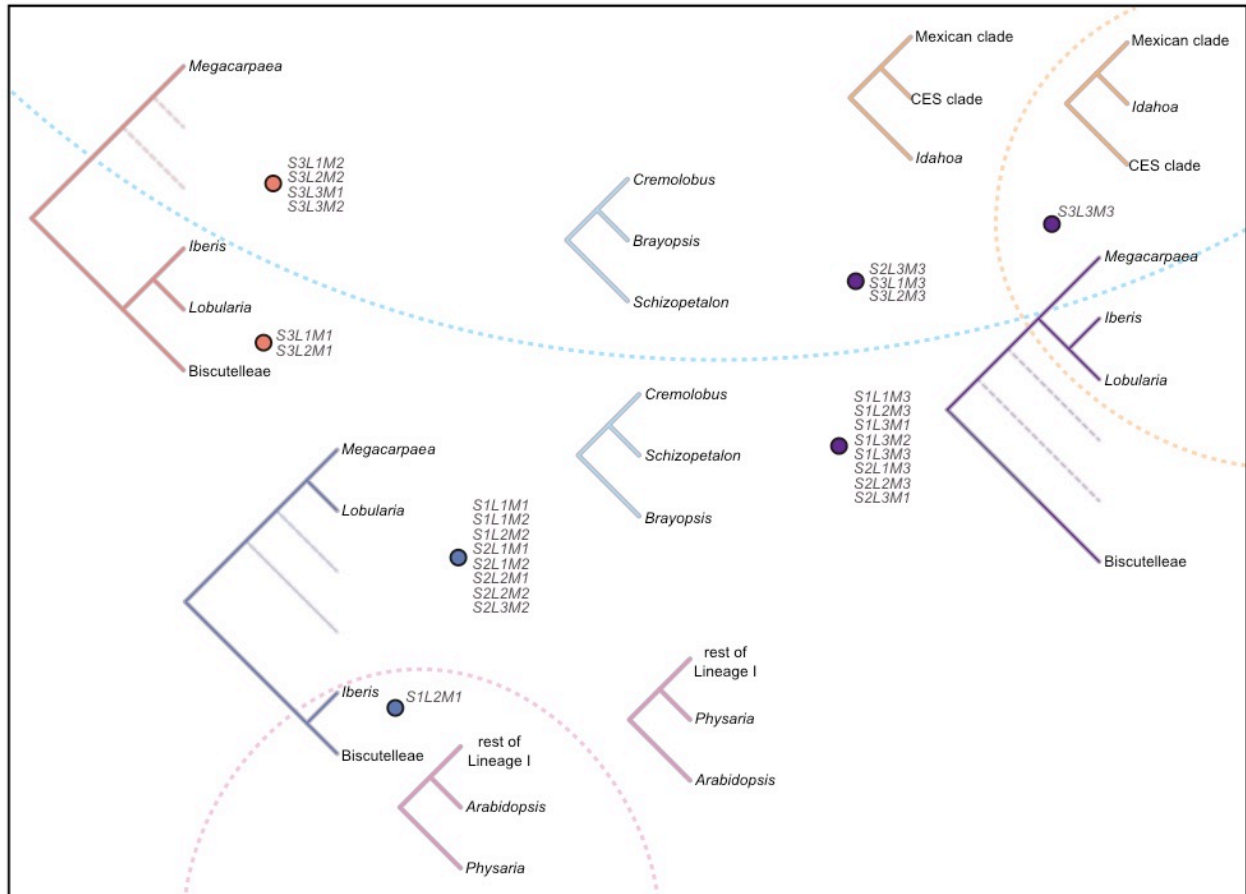
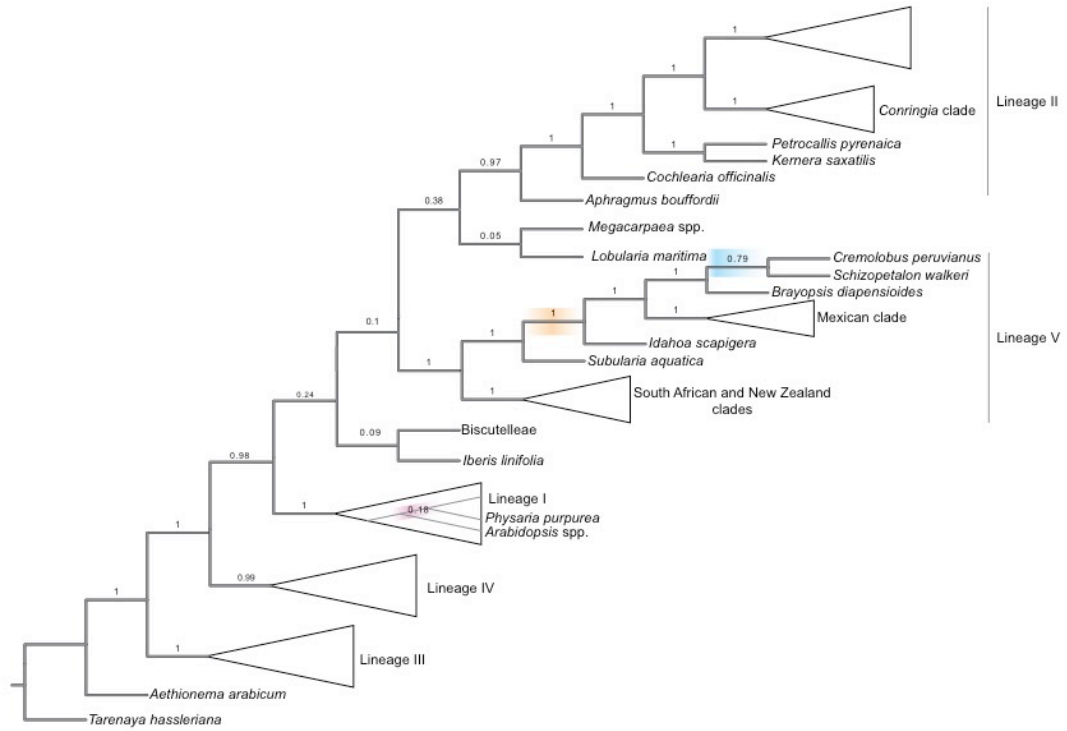


Figure S8. Summary of the ASTRAL-II results represented by the tree space of the seven unique topologies (circles), and the differences among them. The summary tree on top is derived from the most inclusive dataset, *S1L1M1*, and color-coding marks the branches on the tree that are alternatively resolved with the other datasets.

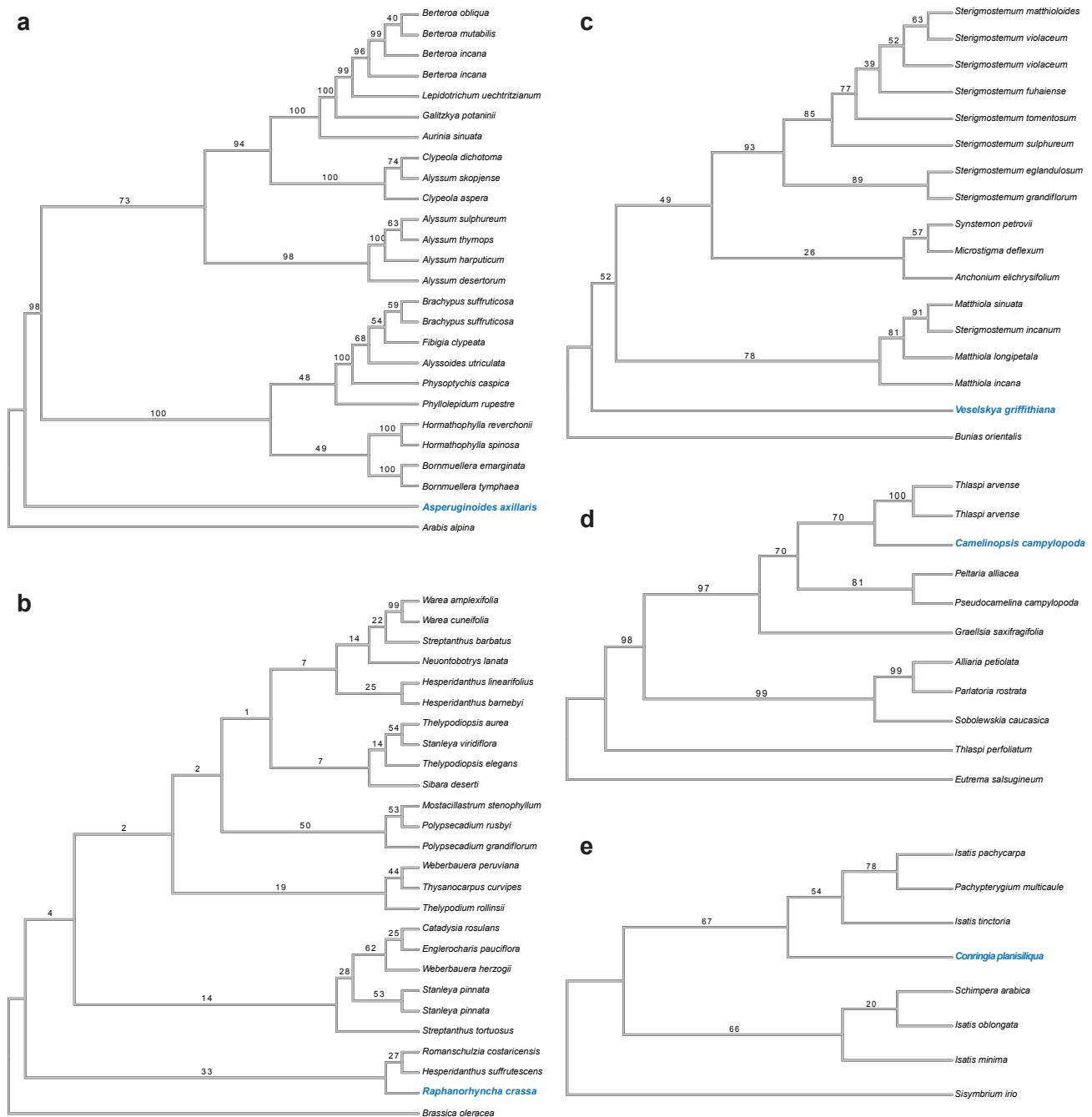
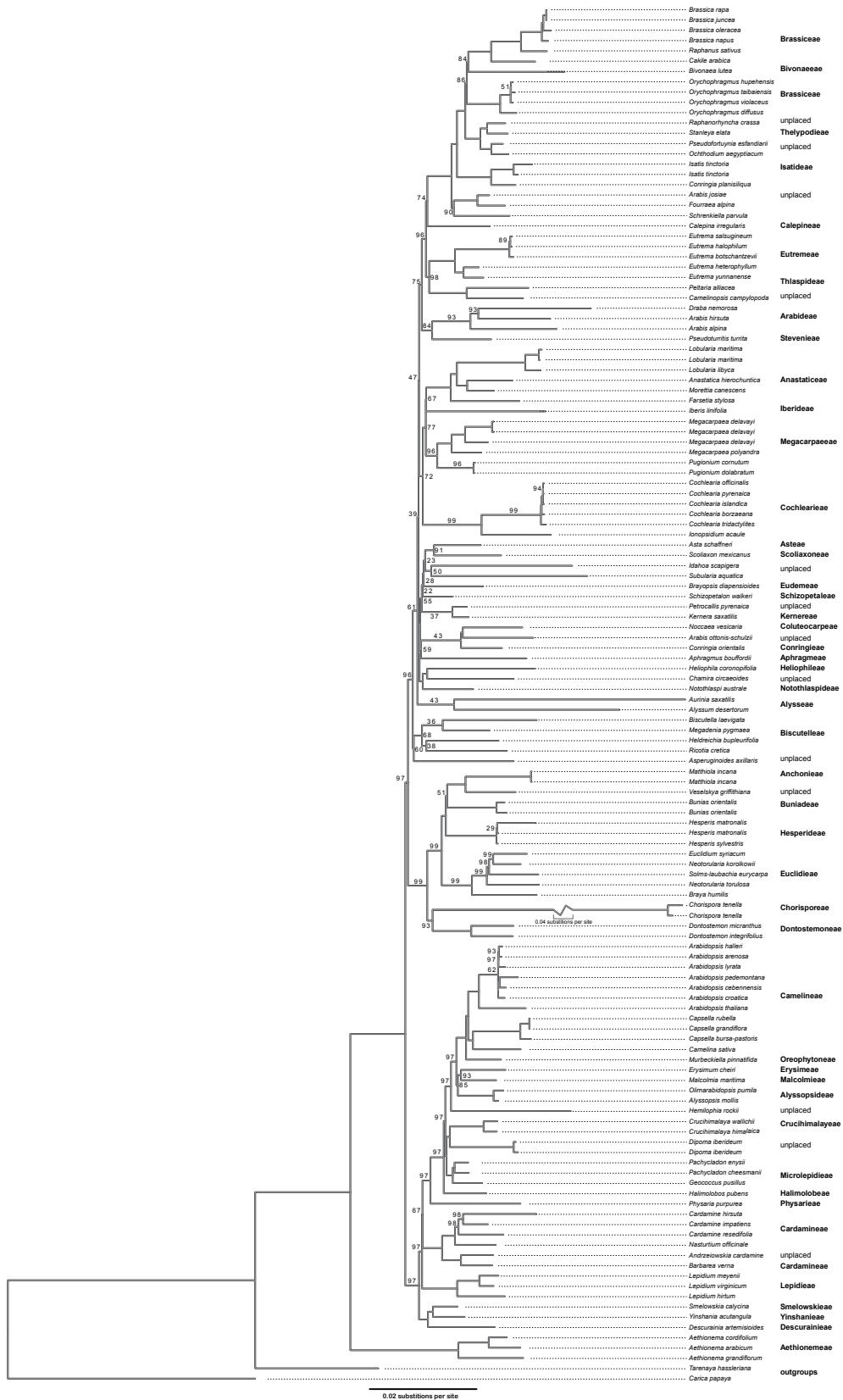


Figure S9. Placing unassigned to a tribe species with expanded taxon sampling of associated sister tribes. a, *Asperuginoides axillaris*. b, *Raphanorhyncha crassa*. c, *Veselskya griffithiana*. d, *Camelinopsis campylopoda*. e, *Conringia planisiliqua*.



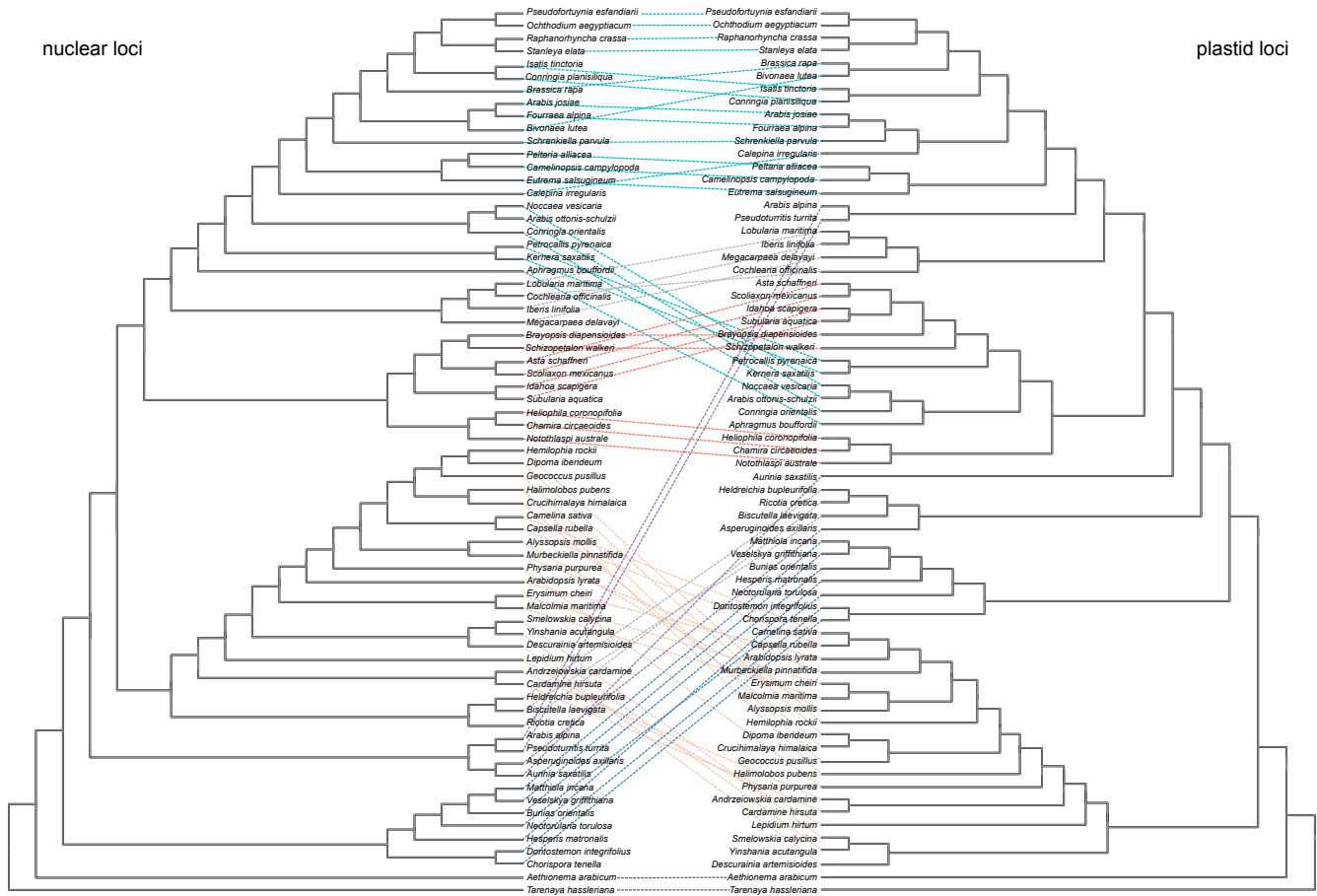


Figure S11. Comparison between tree topologies derived from nuclear and plastid loci. Unmatched taxa between the two datasets are excluded, and multispecies genera (except *Arabis* spp. and *Conringia* spp.) are trimmed to a single species.

Table S1. Taxon and voucher information for species with newly generated sequence data

<i>Name</i>	<i>Tribe</i>	<i>Figure code</i>	<i>Voucher information</i>
<i>Alyssopsis mollis</i> (Jacq.) O.E.Schulz	Alyssopsidae	Aps	B-2011-0216*, HEID
<i>Andrzeiowskia cardamine</i> Rchb.	unplaced	Arz	Gangale and Uzunov 11-2-17, B
<i>Aphragmus bouffordii</i> Al-Shehbaz	Aphragmeae	Aph	Boufford et al. 31446, A
<i>Arabis josiae</i> Jahand. & Maire	unplaced	Ajo	Nikolov 3829, BONN
<i>Arabis ottonis-schulzii</i> Bornm. & Gauba	unplaced	Aos	Nikolov 3830, BONN
<i>Asperuginoides axillaris</i> (Boiss. & Hohen.) Rauschert	unplaced	Asp	Freizi 36120, FUMH
<i>Asta schaffneri</i> (S.Watson) O.E.Schulz var. <i>pringlei</i> (O.E.Schulz) Rollins	Astaeae	Ast	Hinton et al. 20193, GH
<i>Aurinia saxatilis</i> (L.) Desv.	Alysseae	Aly	xx-0-BONN-29279*, BONN
<i>Biscutella laevigata</i> L.	Biscutelleae	Bsc	FR-0-BONN-36028*, BONN
<i>Bivonaea lutea</i> DC.	Bivonaeae	Biv	F. Wirtgen**, B100517564, B
<i>Brayopsis diapensioides</i> Gilg & Muschl.	Eudemeae	Edm	Fuentes et al. 12659, MOBOT
<i>Bunias orientalis</i> L.	Buniadeae	Bun	xx-0-BONN-3323*, BONN
<i>Calepina irregularis</i> (Asso) Thell.	Calepineae	Cal	xx-0-BONN-23168*, BONN
<i>Camelinopsis campylopoda</i> (Bornm. & Gauba) A.G. Mill. (syn. <i>Pseudocamelina campylopoda</i>)	Thlaspidaeae	Cam	Sorkh-e Hesar 12438, TUH
<i>Chamira circaeoides</i> (L. f.) Zahlbr.	unplaced	Chm	Helme 3533, K
<i>Chorispora tenella</i> (Pallas) DC.	Chorisporaeae	Chr	B-2006-0629*, HEID
<i>Cochlearia officinalis</i> L.	Cochlearieae	Coc	xx-0-BONN-19077*, BONN
<i>Conringia orientalis</i> (L.) C.Presl	Conringieae	Crg	xx-0-BONN-21509*, BONN
<i>Conringia planisiliqua</i> Fischer & C.A. Meyer	unplaced	Trt	B-2006-0697*, HEID; Nikolov 3818, BONN
<i>Cremolobus peruvianus</i> (Lam.) DC.	Cremolobeae	Cml	Woytkowski 8243, GH
<i>Crucihimalaya himalaica</i> (Edgeworth) Al-Shehbaz, O'Kane & Price	Crucihimalayaeae	Cru	Al-Shehbaz 9354, GH
<i>Descurainia artemisioides</i> Svent.	Descurainieae	Des	Nikolov 3814, BONN
<i>Dipoma iberideum</i> Franch.	unplaced	Dpm	Borosova et al. 10CS2304, K
<i>Dipoma iberideum</i> Franch.	unplaced	Dip	Rock 25136, GH
<i>Dontostemon integrifolius</i> (L.) C.A. Mey	Dontostemoneae	Don	Nikolov 3822, BONN
<i>Erysimum cheiri</i> (L.) Crantz	Erysimeae	Ers	xx-0-STGAL-1993/647*, BONN
<i>Fourraea alpina</i> (L.) Greuter & Burdet.	unplaced	Frr	Nikolov 3815
<i>Geococcus pusillus</i> J.Drumm. ex Harv.	Microlepidieae	Mic	B-2001-0352*, HEID
<i>Halimolobos pubens</i> (A.Gray) Al-Shehbaz & C.D.Bailey	Halimolobeae	Hal	Nikolov 3824, BONN
<i>Heldreichia bupleurifolia</i> Boiss.	Biscutelleae	Hdr	Döring et al. 6693, B
<i>Heliophila coronopifolia</i> L.	Heliophileae	Hph	Nikolov 3821, BONN
<i>Hemilophia rockii</i> O.E. Schulz	unplaced	Hml	Boufford et al. 33431, A
<i>Hesperis matronalis</i> L.	Hesperideae	Hes	DE-0-BONN-34419*, BONN
<i>Iberis linifolia</i> L. subsp. <i>boppardensis</i> (Jord.) Korneck	Iberideae	Ibr	DE-0-BONN-5874*, BONN
<i>Idahoa scapigera</i> (Hook.) A. Nels. & J.F. Macbr.	unplaced	Ida	Baum 364, A
<i>Isatis tinctoria</i> L.	Isatideae	Ist	GE-0-BONN-35893*, BONN
<i>Kernera saxatilis</i> (L.) Sweet	Kernereae	Krn	Nikolov 3825, BONN
<i>Lepidium hirtum</i> (L.) Sm.	Lepidieae	Lep	B-2014-0057*, HEID
<i>Lobularia maritima</i> (L.) Desv.	Anastaticheae	Lob	FR-0-BONN-32257*, BONN
<i>Lunaria rediviva</i> L.	Biscutelleae	Lun	xx-0-BONN-9709*, BONN

<i>Malcolmia maritima</i> (L.) W.T.Aiton	Malcolmieae	Ana	Nikolov 3817, BONN
<i>Matthiola incana</i> (L.) W.T.Aiton	Anchonieae	Anc	xx-0-BONN-28336*, BONN
<i>Megacarpaea delavayi</i> Franch.	Megacarpaeae	Mgc	Sino-American Botanical Exp. 1031, B
<i>Megacarpaea delavayi</i> Franch.	Megacarpaeae	Meg	Borosova et al. 10CS2307, K
<i>Megacarpaea polyandra</i> Benth.	Megacarpaeae	Mgr	Chase 565, K
<i>Murbeckiella pinnatifida</i> (Lam.) Rothm.	Oreophytoneae	Ore	B-2011-0222*, HEID
<i>Neotorularia torulosa</i> (Desf.) Hedge & J. Léonard	Euclidieae	Euc	Nikolov 3828, BONN
<i>Noccaea vesicaria</i> (L.) Al-Shehbaz	Coluteocarpeae	Clt	GE-0-BONN-35746*, BONN
<i>Notothlaspi australe</i> Hook.f.	Notothlaspidiae	Not	Heenan 2015, BONN
<i>Ochthodium aegyptiacum</i> (L.) DC.	unplaced	Och	Nikolov 3823, BONN
<i>Peltaria alliacea</i> Jacq.	Thlaspidiae	Tls	xx-0-BONN-3341*, BONN
<i>Petrocallis pyrenaica</i> (L.) W.T. Aiton.	unplaced	Pet	Nikolov 3820, BONN
<i>Physaria purpurea</i> (A.Gray) O'Kane & Al-Shehbaz	Physarieae	Phy	Nikolov 3826, BONN
<i>Pseudofortuynia esfandiarii</i> Hedge.	Sisymbrieae	Psf	Nikolov 3827, BONN
<i>Pseudoturritis turrita</i> (L.) Al-Shehbaz	Stevenieae	Stv	DE-0-BONN-14285*, BONN
<i>Raphanorhyncha crassa</i> Roll.	unplaced	Rph	Johnson et al. 10536, GH
<i>Ricotia cretica</i> Boiss. & Heldr.	Biscutelleae	Ric	Kamari and Turland 1322, B
<i>Schizopetalon walkeri</i> Sims.	Schizopetaleae	Sch	Nikolov 3819, BONN
<i>Scoliaxon mexicanus</i> (S.Watson) Payson	Scoliaxoneae	Scl	Henrickson 13702a, GH
<i>Smelowskia calycina</i> (Stephan ex Willd.) C.A.Mey.	Smelowskieae	Sml	B-2011-0226*, HEID
<i>Stanleya elata</i> M.E.Jones	Thelypodieae	The	Nikolov 3816, BONN
<i>Subularia aquatica</i> L.	unplaced	Sub	Štěpánek, B
<i>Turritis glabra</i> L.	Turritideae	Tgl	xx-0-BONN-15131*, BONN
<i>Veselskya griffithiana</i> (Boiss.) Opiz.	unplaced	Ves	Rechinger 33377, B
<i>Yinshania acutangula</i> (O.E.Schulz) Y.H.Zhang subsp. <i>acutangula</i>	Yinshanieae	Yin	Boufford et al. 36219, A

*cultivated specimen, collector name N/A

**collector and collection year unknown
(acquired by B in 1936)

Table S2. Accession information for species with sequenced nuclear genomes

<i>Name</i>	<i>Tribe</i>	<i>Figure code</i>	<i>Genome assembly</i>
<i>Aethionema arabicum</i> (L.) Andr. ex DC.	Aethionemeae	Aet	GCA_000411095.1
<i>Arabidopsis lyrata</i> (L.) O'Kane & Al-Shehbaz	Camelineae	Arl	GCA_000004255.1
<i>Arabidopsis thaliana</i> (L.) Heynh.	Camelineae	Ath	GCA_000001735.1
<i>Arabis alpina</i> L.	Arabideae	Alp	GCA_000733195.1
<i>Boechera stricta</i> (Graham) Al-Shehbaz	Boechereae	Boe	GCA_002079875.1
<i>Brassica oleracea</i> L.	Brassiceae	Bro	GCF_000695525.1
<i>Brassica rapa</i> L.	Brassiceae	Brr	GCF_000309985.1
<i>Camelina sativa</i> (L.) Crantz	Camelineae	Csa	GCA_000633955.1
<i>Capsella rubella</i> Reut.	Camelineae	Crb	GCF_000375325.1
<i>Cardamine hirsuta</i> L.	Cardamineae	Chi	chi_v1
<i>Eutrema salsugineum</i> (Pall.) Al-Shehbaz & S.I. Warwick	Eutremeae	Eut	GCA_000478725.1
<i>Leavenworthia alabamica</i> Rollins	Cardamineae	Lea	GCA_000411055.1
<i>Lepidium meyenii</i> Walp.	Lepidieae	Lmy	
<i>Carica papaya</i> L.	outgroup	Car	GCA_001310045.1
<i>Schrenkiella parvula</i> (Schrenk) D.A. German & Al-Shehbaz	unplaced	Skl	GCA_000218505.1
<i>Sisymbrium irio</i> L.	Sisymbrieae	Sis	GCA_000411075.1
<i>Tarenaya hassleriana</i> (Chodat) Iltis	outgroup	Tar	GCF_000463585.1
<i>Thlaspi arvense</i> L.	Thlaspideae	Tla	GCA_000956625.1

Table S3. Accession information for species with sequenced plastid genomes

<i>Name</i>	<i>Tribe</i>	<i>Accession number</i>
<i>Aethionema arabicum</i> (L.) Andr. ex DC.	Aethionemeae	KX886357
<i>Aethionema coridifolium</i> DC.	Aethionemeae	NC_009265.1
<i>Aethionema grandiflorum</i> Boiss. & Hohen.	Aethionemeae	NC_009266.1
<i>Alyssum desertorum</i> Stapf	Alysseae	NC_034299.1
<i>Anastatica hierochuntica</i> L.	Anastaticeae	KY912021.1
<i>Arabidopsis arenosa</i> (L.) Lawalrée	Camelineae	NC_029334.1
<i>Arabidopsis cebennensis</i> (DC.) O'Kane & Al-Shehbaz	Camelineae	NC_029335.1
<i>Arabidopsis croatica</i> (Schott) O'Kane & Al-Shehbaz	Camelineae	NC_030347.1
<i>Arabidopsis halleri</i> (L.) O'Kane & Al-Shehbaz	Camelineae	KX886356
<i>Arabidopsis lyrata</i> (L.) O'Kane & Al-Shehbaz	Camelineae	KX886355
<i>Arabidopsis pedemontana</i> (Boiss.) O'Kane & Al-Shehbaz	Camelineae	LN877384
<i>Arabidopsis thaliana</i> (L.) Heynh.	Camelineae	NC_000932.1
<i>Arabis alpina</i> L.	Arabideae	NC_023367.1
<i>Arabis hirsuta</i> (L.) Scop.	Arabideae	AP009369
<i>Barbarea verna</i> (Mill.) Asch.	Cardamineae	NC_009269.1
<i>Brassica juncea</i> (L.) Czern.	Brassiceae	KT581449
<i>Brassica napus</i> L.	Brassiceae	GQ861354
<i>Brassica oleracea</i> L.	Brassiceae	KR233156
<i>Brassica rapa</i> subsp. <i>pekinensis</i> Kitam.	Brassiceae	NC_015139.1
<i>Braya humilis</i> (C.A. Mey.) B.L. Rob.	Euclidieae	KY912032.1
<i>Bunias orientalis</i> L.	Buniadeae	KY912022.1
<i>Cakile arabica</i> Velen. & Bornm.	Brassiceae	NC_030775.1
<i>Camelina sativa</i> (L.) Crantz	Camelineae	NC_029337.1
<i>Capsella bursa-pastoris</i> (L.) Medik.	Camelineae	AP009371
<i>Capsella grandiflora</i> Boiss.	Camelineae	NC028517
<i>Capsella rubella</i> Reut.	Camelineae	NC_027693.1
<i>Cardamine hirsuta</i> L.	Cardamineae	chi_v1
<i>Cardamine impatiens</i> L.	Cardamineae	KJ136821
<i>Cardamine resedifolia</i> L.	Cardamineae	KJ136822
<i>Carica papaya</i> L.	outgroup	NC_010323.1
<i>Chorispora tenella</i> (Pall.) DC.	Chorisporaeae	KY912028.1
<i>Cochlearia borzaeana</i> Pobed.	Cochlearieae	LN866844
<i>Cochlearia islandica</i> Pobed.	Cochlearieae	NC_029254.1
<i>Cochlearia pyrenaica</i> DC.	Cochlearieae	NC_029331.1
<i>Cochlearia tridactylites</i> Banks ex DC.	Cochlearieae	LN866847
<i>Crucihimalaya wallichii</i> (Hook. f. & Thomson) Al-Shehbaz, O'Kane & R.A. Price	Crucihimalayaeae	NC_009271.1
<i>Draba nemorosa</i> L.	Arabideae	NC_009272.1
<i>Dontostemon micranthus</i> C.A. Mey.	Dontostemoneae	KY912023.1

<i>Euclidium syriacum</i> (L.) W.T. Aiton	Euclidiaceae	KY912024.1
<i>Eutrema botschantzevii</i> (D.A. German) Al-Shehbaz & S.I. Warwick	Eutremeaceae	KT962847
<i>Eutrema halophilum</i> (C.A. Mey.) Al-Shehbaz & S.I. Warwick	Eutremeaceae	NC_029378.1
<i>Eutrema heterophyllum</i> (W.W. Sm.) H. Hara	Eutremeaceae	KT270358
<i>Eutrema salsugineum</i> (Pall.) Al-Shehbaz & S.I. Warwick	Eutremeaceae	NC_028170.1
<i>Eutrema yunnanense</i> Franch.	Eutremeaceae	KT270357
<i>Farsetia stylosa</i> R. Br.	Anastatiaceae	KY912025.1
<i>Hesperis matronalis</i> L.	Hesperideae	KY912026.1
<i>Hesperis sylvestris</i> Crantz	Hesperideae	KY912027.1
<i>Ionopsidium acaule</i> (Desf.) DC. ex Rchb.	Cochleariaceae	NC_029333.1
<i>Isatis tinctoria</i> L.	Isatideae	NC_028415.1
<i>Lepidium meyenii</i> Walp.	Lepidieae	KX886353
<i>Lepidium virginicum</i> L.	Lepidieae	NC_009273.1
<i>Lobularia libyca</i> Webb & Berthel.	Anastatiaceae	KY912029.1
<i>Lobularia maritima</i> (L.) Desv.	Anastatiaceae	NC_009274.1
<i>Matthiola incana</i> (L.) W.T. Aiton	Anchonieae	KY912030.1
<i>Megacarpaea delavayi</i> Franch.	Megacarpaeaceae	KX886349
<i>Megadenia pygmaea</i> Maxim.	Biscutelleae	KX371593
<i>Morettia canescens</i> Boiss.	Anastatiaceae	KY912031.1
<i>Nasturtium officinale</i> W.T. Aiton	Cardamineae	NC_009275.1
<i>Neotorularia korolkowii</i> (Regel & Schmalh.) Hedge & J. Léonard	Euclidiaceae	KX886350
<i>Olimarabidopsis pumila</i> (Stephan) Al-Shehbaz, O'Kane & R.A. Price	Alyssopsideae	NC_009267.1
<i>Orychophragmus diffusus</i> Z.M. Tan & J.M. Xu	Brassicaceae	NC_033498.1
<i>Orychophragmus hupehensis</i> (Pamp.) Z.M. Tan & X.L. Zhang	Brassicaceae	NC_033500.1
<i>Orychophragmus taibaiensis</i> Z.M. Tan & B.X. Zhao	Brassicaceae	NC_033499.1
<i>Orychophragmus violaceus</i> (L.) O.E. Schulz	Brassicaceae	KX364399.1
<i>Pachycladon cheesmanii</i> Heenan & A.D.Mitch.	Microlepidieae	NC_021102.1
<i>Pachycladon ensyii</i> (Cheeseman) Heenan & A.D.Mitch.	Microlepidieae	JX205495
<i>Pugionium cornutum</i> (L.) Gaertn.	Megacarpaeaceae	KT844941
<i>Pugionium dolabratum</i> Maxim.	Megacarpaeaceae	KT844940
<i>Raphanus sativus</i> L.	Brassicaceae	NC_024469.1
<i>Schrenkiella parvula</i> (Schrenk) D.A. German & Al-Shehbaz	unplaced	NC_028726.1
<i>Solms-laubachia eurycarpa</i> (Maxim.) Botsch.	Euclidiaceae	KX886348
<i>Tarenaya hassleriana</i> (Chodat) Iltis	outgroup	KX886354.1

Table S4. Reads and contig statistics

Species	Initial reads	Trimmed reads	Deduplicated reads	<i>Arabidopsis</i> reference					<i>Aethionema</i> reference					<i>Sisymbrium</i> reference					Exons called
				Mapped reads	Contigs called	Contigs mapped	Positions called	Average coverage	Mapped reads	Contigs called	Contigs mapped	Positions called	Average coverage	Mapped reads	Contigs called	Contigs mapped	Positions called	Average coverage	
<i>Ajo</i>	21088980	20186973	7236479	1891793	2042	2023	1083822	124	1258004	3446	3265	941018	95	2001543	1937	1907	1099196	129	1827
<i>Aly</i>	7299616	7036025	4415819	1254300	2320	2232	1100259	104	816748	3331	3032	904721	82	1222755	2389	2283	1085227	103	1818
<i>Ana</i>	10655622	10279538	5928709	1839951	1893	1875	1180126	140	1161567	3006	2832	1010716	104	1706579	1989	1962	1155236	133	1827
<i>Anc</i>	8331770	7954100	5014105	1203472	2407	2305	1102574	100	832515	3317	3032	933537	81	1184682	2379	2284	1094053	99	1824
<i>Aos</i>	14443682	13866406	6173326	1524028	2136	2113	1071993	103	983006	3560	3334	900424	79	1549808	2096	2067	1077888	104	1827
<i>Aph</i>	27476918	26717159	9837212	1800303	2114	2097	1080586	119	1239315	3431	3252	943148	94	1794264	2140	2113	1080448	119	1826
<i>Aps</i>	10244638	9826460	5774737	1792444	1904	1899	1183882	137	1113256	3185	2985	1002864	100	1637668	2101	2065	1153692	128	1825
<i>Arz</i>	27245170	26479549	9925925	1767448	2037	2019	1086891	117	1131965	3549	3320	922742	88	1649043	2263	2214	1066861	111	1827
<i>Asp</i>	11604350	11183056	6879718	1885942	2059	2032	1134945	144	1297128	3148	2944	1008385	112	1860940	2111	2086	1131798	143	1827
<i>Ast</i>	22827346	22181923	8255994	1721665	2117	2089	1054222	114	1196654	3304	3110	931526	90	1701810	2127	2096	1049114	113	1822
<i>Biv</i>	24089978	23456084	2778699	203019	4118	3438	663874	19	130344	4412	3364	470228	17	224283	3892	3294	707115	19	1730
<i>Bsc</i>	9095296	8657698	6069868	1380331	2165	2109	1125202	109	925892	3207	2964	964155	86	1331769	2228	2173	1113970	106	1825
<i>Bun</i>	9627858	9252002	6705928	1103160	2303	2209	1111904	92	774915	3267	2963	948775	75	1084075	2350	2252	1105860	91	1823
<i>Cal</i>	11260696	10776155	5630398	1850394	2017	1985	1157675	143	1201405	3119	2910	989935	109	1907213	2002	1967	1166319	146	1826
<i>Cam</i>	22775172	22081638	6300269	1250434	2277	2234	1051793	84	801267	3660	3419	874431	65	1305118	2137	2101	1063475	87	1827
<i>Chm</i>	3068382	2971687	864985	286419	2637	2447	884373	23	203279	3503	3088	698273	20	279810	2699	2512	871658	23	1812
<i>Chr</i>	8702986	8356420	5554750	1316326	2169	2084	1113521	108	906290	3173	2912	946973	88	1296917	2164	2086	1109753	107	1824
<i>Clt</i>	8153628	7775634	5028631	1518686	2116	2086	1148628	120	969980	3312	3067	966928	91	1552980	2079	2042	1154761	121	1824
<i>Cml</i>	9270092	8821576	5876127	1257534	2091	2049	1122970	101	845775	3199	2970	962551	79	1246721	2086	2043	1117382	100	1826
<i>Coc</i>	11058294	10609462	7290312	1330638	2493	2339	1062263	111	848633	3419	3048	865035	87	1355098	2483	2347	1070912	112	1814
<i>Crg</i>	13994564	13468921	6503383	2062271	2029	2011	1173621	156	1377648	3144	2961	1023276	120	2092620	1981	1967	1175943	158	1827
<i>Cru</i>	22285532	21662204	7148442	1856950	1925	1918	1096952	122	1153525	3522	3340	924265	90	1681256	2266	2215	1068974	113	1827
<i>Des</i>	13865706	13276141	7144241	2135733	1917	1903	1178807	160	1336920	2985	2813	1014691	117	1991967	1959	1928	1158182	152	1827
<i>Dip</i>	21074238	20582565	6267172	1450491	1931	1920	1055628	95	924610	3432	3225	904428	71	1329803	2214	2178	1036324	89	1826
<i>Don</i>	22663108	22060167	10759260	1477916	2453	2379	1030529	102	1030574	3563	3305	892372	82	1455770	2518	2431	1025193	101	1825
<i>Dpm</i>	29627694	28969045	11134866	2069635	1911	1903	1069129	133	1320999	3378	3178	932769	99	1895321	2174	2148	1052792	125	1826
<i>Edm</i>	28324008	27291944	11639799	2040671	2118	2096	1052808	132	1375331	3475	3299	929694	102	2019272	2137	2111	1049883	131	1823
<i>Ers</i>	11130202	10722802	5913035	1862451	1920	1916	1182704	141	1175085	3147	2967	1015881	104	1729933	2061	2039	1159305	134	1827
<i>Euc</i>	8260374	7937133	4706539	1368147	2310	2239	1104463	113	931286	3288	3017	938790	90	1341920	2377	2266	1095646	111	1825
<i>Frr</i>	24129880	23383135	8047309	1826792	2112	2097	1085140	120	1205628	3578	3364	934140	92	1934503	1940	1927	1100428	125	1827
<i>Hal</i>	10782772	10395564	5340814	1888353	1906	1898	1186826	143	1178971	3118	2943	1010544	105	1738064	2069	2036	1160162	135	1827
<i>Hdr</i>	40420378	38345390	8259028	1847060	2145	2123	1072104	117	1212201	3587	3356	918394	90	1800576	2196	2167	1063494	115	1825
<i>Hes</i>	11543578	11057974	7504020	1296500	2214	2117	1114708	105	926774	3130	2868	971897	86	1276515	2269	2134	1108059	104	1823
<i>Hml</i>	46211756	44779475	11154122	2278628	1961	1939	1066695	145	1403384	3508	3263	909941	106	2035043	2325	2263	1040825	133	1820
<i>Hph</i>	13841056	13344045	7402019	1730349	2132	2060	1101726	136	1177614	3101	2860	949295	107	1701015	2125	2047	1096591	134	1824
<i>Ibr</i>	9517236	9148343	5686491	1382578	2227	2146	1105422	112	952928	3195	2930	944957	90	1343124	2270	2167	1093169	110	1821
<i>Ida</i>	26546786	25719931	5708715	723270	3010	2832	913203	54	506482	3871	3447	752332	45	703741	3131	2923	901049	53	1809
<i>Ist</i>	10731682	10267273	6798089	1819965	2087	2070	1161469	139	1181267	3261	3062	998169	106	1965420	1906	1895	1180819	148	1827
<i>Krn</i>	15878788	15241186	7275546	2273222	1982	1969	1178903	169	1542696	3049	2893	1041619	130	2296602	1983	1965	1181639	171	1827
<i>Lep</i>	10984084	10604616	5366464	1652499	2126	2042	1137596	132	1027973	3365	3057	938239	99	1500026	2309	2187	1100786	123	1812
<i>Lob</i>	22063064	21430221	9803717	1385031	2912	2770	983182	98	881449	3961	3559	793629	77	1376781	2915	2767	977162	98	1820
<i>Lun</i>	9974422	9582612	6452912	1656659	1968	1945	1143472	127	1154172	3004	2804	1028199	99	1652591	1926	1903	1140348	127	1827
<i>Meg</i>	30140858	28788167	11719016	767115	2245	2173	1007164	52	515863	3596	3312	841153	42	762102	2277	2206	1003845	52	1826
<i>Mgc</i>	19682190	19258960	9264817	1578842	2211	2179	1052854	106	1067926	3526	3309	912556	83	1569028	2206	2173	1050817	106	1825
<i>Mgr</i>	19032210	18559264	8111057	1621367	2172	2139	1054012	109	1101384	3473	3267	918076	85	1611596	2194	2158	1052640	108	1824

<i>Mic</i>	10357532	9891495	6137622	1867095	1908	1893	1155775	140	1145254	3153	2942	987618	102	1708765	2057	2033	1130697	132	1826
<i>Not</i>	27323496	26694964	9405503	1898037	2152	2119	1055521	126	1318405	3334	3163	928834	100	1873134	2202	2161	1052545	125	1817
<i>Och</i>	11585916	11170541	6225147	1795346	2094	2067	1159841	139	1145402	3328	3090	989664	104	1980367	1879	1865	1190111	149	1827
<i>Ore</i>	17103922	16410951	7411560	2404777	1901	1892	1190904	177	1497333	3137	2959	1026273	128	2207032	2103	2066	1165798	166	1824
<i>Pet</i>	30536488	29687163	9624017	2047407	2054	2041	1090575	133	1394726	3419	3220	956652	104	2065990	2028	2009	1092905	134	1827
<i>Phy</i>	15078646	14505604	11620874	1454884	2028	1978	1133130	115	892111	3221	2934	938728	85	1278086	2296	2189	1088993	105	1823
<i>Psf</i>	13159552	12707278	9448740	1515034	2038	2018	1160902	119	982985	3247	3027	993508	90	1649188	1857	1852	1187411	126	1826
<i>Ric</i>	27650286	26862269	7690382	1768835	2159	2123	1049782	116	1227475	3382	3206	929711	92	1732666	2241	2199	1043481	115	1827
<i>Rph</i>	37265244	36071593	14330040	2253336	2052	2030	1055533	145	1501622	3350	3160	942447	109	2442472	1875	1856	1068613	154	1827
<i>Sch</i>	8217830	7910301	5194347	1499933	2041	1993	1131982	118	1037514	3063	2849	992353	93	1487467	2020	1976	1126848	117	1819
<i>Scl</i>	21889346	21339856	8946367	1573955	2226	2181	1045297	106	1112560	3420	3201	919752	85	1552107	2283	2228	1040545	105	1822
<i>Sml</i>	24259666	22952366	10719799	1912307	1957	1948	1090851	123	1207591	3561	3365	932786	91	1784883	2183	2156	1072725	117	1823
<i>Stv</i>	9684514	9281572	5394934	1703160	1993	1974	1174003	131	1182923	3063	2900	1036245	103	1690466	2009	1992	1170980	131	1827
<i>Sub</i>	24228390	23583054	10038613	1394430	2689	2540	971816	100	905903	3806	3460	803384	78	1352415	2813	2660	958921	98	1815
<i>Tgl</i>	N/A	N/A	N/A	895903	1906	1814	1154105	95	574107	2952	2523	956714	74	841295	2175	1999	1129243	92	1781
<i>The</i>	11357328	10888548	8386347	1693010	2002	1982	1128794	130	1126429	3077	2882	1001507	99	1817078	1886	1854	1143942	137	1827
<i>Tls</i>	11616146	11145218	6135238	1860097	2113	2082	1161930	143	1189931	3330	3103	985666	108	1928674	2026	2003	1169640	147	1827
<i>Trt</i>	12394428	11966371	5975655	1879218	2086	2062	1165522	145	1203468	3251	3045	993659	109	2028961	1927	1903	1188849	153	1827
<i>Ves</i>	27649518	26872285	6274275	764037	2369	2256	1003110	52	528129	3554	3216	845236	43	749735	2388	2267	994395	52	1827
<i>Yin</i>	24328992	23671627	6661675	1745645	1946	1938	1091344	114	1135833	3501	3307	937555	87	1641831	2145	2115	1075556	109	1827

Table S5. Skimmed read and whole plastome statistics

Species	Mapped reads	Recovered genes	Plastome length, bp	LSC length, bp	IR length, bp	SSC length, bp	LSC average coverage	IR average coverage	SSC average coverage
<i>Ajo</i>	67834	77							
<i>Aly</i>	7734	9							
<i>Ana</i>	73735	97							
<i>Anc</i>	41302	95							
<i>Aos</i> *	254651	97	153138	82310	26354	18119	71	186	70
<i>Aph</i>	384416	84							
<i>Aps</i> *	66260	97	154839	83969	26497	17875	29	64	30
<i>Arz</i>	635876	85							
<i>Asp</i>	17750	52							
<i>Ast</i>	54810	36							
<i>Biv</i>	201720	10							
<i>Bsc</i>	7771	6							
<i>Bun</i> *	178742	97	153877	83598	26246	17786	82	167	93
<i>Cal</i>	28199	85							
<i>Cam</i>	81437	78							
<i>Chm</i>	81595	86							
<i>Chr</i>	639418	94							
<i>Clt</i>	30161	89							
<i>Coc</i>	348163	96							
<i>Crg</i>	20405	72							
<i>Cru</i>	44184	29							
<i>Des</i>	27616	85							
<i>Dip</i> *	362478	97	153804	82865	26502	17934	108	239	110
<i>Don</i>	76302	82							
<i>Dpm</i>	116263	97							
<i>Edm</i>	91717	82							
<i>Ers</i>	25159	46							
<i>Euc</i>	18935	54							
<i>Frr</i>	131664	97							
<i>Hal</i>	46096	96							
<i>Hdr</i>	98839	92							
<i>Hes</i>	18074	16							
<i>Hml</i>	178494	93							
<i>Hph</i>	29413	88							
<i>Ibr</i>	17583	43							
<i>Ida</i>	54263	55							
<i>Ist</i>	31213	93							
<i>Krn</i> *	61373	97	154832	84079	26426	17900	26	60	27
<i>Lep</i>	23612	69							
<i>Lob</i>	75841	90							
<i>Meg</i>	251510	97							
<i>Mgc</i>	198169	97							
<i>Mgr</i>	58369	71							
<i>Mic</i>	22618	68							
<i>Not</i>	96214	96							
<i>Och</i>	23607	72							

Ore *	80037	97	154918	84344	26460	17653	31	86	31
Pet	154123	97							
Phy	613614	97							
Psf *	1342274	97	154140	84032	26236	17635	635	1268	684
Ric	52321	37							
Rph	80327	86							
Sch	34029	88							
Scl	97717	88							
Sml	602344	96							
Stv	51868	97							
Sub *	205509	96	152645	82178	26347	17772	65	124	73
The *	801408	97	153669	83498	26258	17654	386	756	396
Tls	31247	92							
Trt	21847	75							
Ves	153925	97							
Yin	52177	49							

* species with complete plastomes

LSC, large single copy region

IR, inverted regions

SSC, small single copy region

Table S6. Dataset statistics

Dataset	Number of exons	Total sites, bp	Variable sites, bp	Variable sites, %	Missing data, %	Number of partitions
<i>S1L1M1</i>	1540	739071	463726	62.74%	2.57%	114
<i>S1L1M2</i>	1405	698340	434653	62.24%	2.60%	73
<i>S1L1M3</i>	976	540603	343343	63.51%	2.77%	63
<i>S1L2M1</i>	1415	691641	434639	62.84%	2.60%	90
<i>S1L2M2</i>	1300	655620	414212	63.18%	2.62%	97
<i>S1L2M3</i>	916	510426	326616	63.99%	2.79%	62
<i>S1L3M1</i>	868	478356	307797	64.34%	2.65%	47
<i>S1L3M2</i>	816	460569	297533	64.60%	2.67%	50
<i>S1L3M3</i>	588	366363	241174	65.83%	2.86%	46
<i>S2L1M1</i>	1388	672252	415974	61.88%	2.49%	79
<i>S2L1M2</i>	1260	633354	390995	61.73%	2.51%	49
<i>S2L1M3</i>	873	486444	307730	63.26%	2.68%	59
<i>S2L2M1</i>	1274	632925	390145	61.64%	2.52%	49
<i>S2L2M2</i>	1166	598737	374078	62.48%	2.54%	59
<i>S2L2M3</i>	822	463935	294973	63.58%	2.71%	56
<i>S2L3M1</i>	777	442203	283718	64.16%	2.58%	43
<i>S2L3M2</i>	726	424614	274830	64.72%	2.59%	47
<i>S2L3M3</i>	526	337437	221305	65.58%	2.77%	45
<i>S3L1M1</i>	824	412113	249405	60.52%	1.97%	43
<i>S3L1M2</i>	744	387561	234330	60.46%	1.96%	43
<i>S3L1M3</i>	508	293640	180128	61.34%	2.06%	39
<i>S3L2M1</i>	751	391251	238922	61.07%	1.99%	45
<i>S3L2M2</i>	686	370416	225825	60.97%	1.98%	44
<i>S3L2M3</i>	478	284811	176300	61.90%	2.07%	38
<i>S3L3M1</i>	469	289896	182959	63.11%	2.01%	39
<i>S3L3M2</i>	439	277785	174174	62.70%	1.99%	34
<i>S3L3M3</i>	319	220428	140436	63.71%	2.08%	36
<i>S1L1M1R</i>	1421	551271	335897	60.93%	2.48%	58
<i>S1L1M3R</i>	885	391167	245249	62.70%	2.68%	52
<i>S1L2M2R</i>	1191	483141	297630	61.60%	2.53%	48
<i>S2L2M1R</i>	1170	464361	283666	61.09%	2.42%	50
<i>S2L2M2R</i>	1068	436908	267883	61.31%	2.45%	44
<i>S2L2M3R</i>	744	331263	209055	63.11%	2.61%	55
<i>S3L1M1R</i>	750	288645	172071	59.61%	1.91%	39
<i>S3L3M1R</i>	407	186840	117218	62.74%	2.01%	33

Table S7. Approximate unbiased (AU) test for evaluating the support of a given dataset matrix for unique topologies.

<i>matrix / topology</i>	<i>S1L1M1</i>	<i>S1L1M3</i>	<i>S1L2M2</i>	<i>S1L3M1</i>	<i>S1L3M3</i>	<i>S2L2M1</i>	<i>S3L1M1</i>	<i>S3L3M1</i>
S1L1M1	0.885	0.165	0.337	0.052	0.015	0.065	3.00E-06	2.00E-04
S1L1M3	0.272	0.864	0.154	0.006	0.017	0.009	1.00E-70	1.00E-09
S1L2M2	0.543	0.418	0.642	0.449	0.268	0.352	3.00E-08	9.00E-06
S1L3M1	0.002	0.001	0.002	0.693	0.188	0.597	0.002	0.042
S1L3M3	0.001	0.007	0.002	0.115	0.931	0.143	4.00E-13	2.00E-41
S2L2M1	0.158	0.08	0.121	0.569	0.324	0.813	0.016	0.044
S3L1M1	0.051	0.021	0.052	0.196	0.043	0.118	0.845	0.343
S3L3M1	0.001	6.00E-05	4.00E-04	0.423	0.187	0.113	0.558	0.642

<i>matrix / topology</i>	<i>S1L1M1R</i>	<i>S1L1M3R</i>	<i>S1L2M2R</i>	<i>S2L2M1R</i>	<i>S2L2M2R</i>	<i>S2L2M3R</i>	<i>S3L1M1R</i>	<i>S3L3M1R</i>
S1L1M1	0.754	0.168	0.614	0.119	0.252	0.063	0.001	0.004
S1L1M3	0.095	0.815	0.367	0.004	0.053	0.247	2.00E-41	0.001
S1L2M2	0.522	0.193	0.724	0.244	0.531	0.17	0.067	0.02
S2L2M1	0.497	0.07	0.066	0.723	0.553	0.051	0.223	0.02
S2L2M2	0.509	0.151	0.077	0.501	0.775	0.129	0.268	0.013
S2L2M3	0.189	0.53	0.079	0.089	0.232	0.755	0.059	1.00E-04
S3L1M1	0.114	0.001	9.00E-05	0.499	0.677	0.007	0.524	0.197
S3L3M1	0.042	0.024	2.00E-05	0.371	0.14	0.051	0.09	0.835

Table S8. Leaf stability indices.

Taxon	Concatenated trees			ASTRAL trees		
	<i>lsDif</i>	<i>lsEnt</i>	<i>lsMax</i>	<i>lsDif</i>	<i>lsEnt</i>	<i>lsMax</i>
<i>Aet</i>	0.960575	0.954088	0.970431	0.991716	0.993077	0.993787
<i>Alp</i>	0.960575	0.954088	0.970431	0.991716	0.993077	0.993787
<i>Ajo</i>	0.966576	0.962266	0.974932	0.98816	0.99012	0.99112
<i>Aly</i>	0.960575	0.954088	0.970431	0.991716	0.993077	0.993787
<i>Arl</i>	0.972451	0.968075	0.979339	0.991039	0.991759	0.993279
<i>Ana</i>	0.972451	0.968075	0.979339	0.991716	0.993077	0.993787
<i>Anc</i>	0.960575	0.954088	0.970431	0.991716	0.993077	0.993787
<i>Aos</i>	0.943152	0.94352	0.957364	0.98816	0.99012	0.99112
<i>Aph</i>	0.957506	0.949003	0.96813	0.98816	0.99012	0.99112
<i>Aps</i>	0.972451	0.968075	0.979339	0.99162	0.992891	0.993715
<i>Arz</i>	0.972451	0.968075	0.979339	0.991716	0.993077	0.993787
<i>Asp</i>	0.960575	0.954088	0.970431	0.991716	0.993077	0.993787
<i>Ast</i>	0.94677	0.938001	0.960078	0.977302	0.980881	0.982977
<i>Ath</i>	0.972451	0.968075	0.979339	0.991039	0.991759	0.993279
<i>Biv</i>	0.966576	0.962266	0.974932	0.98816	0.99012	0.99112
<i>Bro</i>	0.966576	0.962266	0.974932	0.98816	0.99012	0.99112
<i>Brr</i>	0.966576	0.962266	0.974932	0.98816	0.99012	0.99112
<i>Bsc</i>	0.882394	0.862021	0.911795	0.971259	0.976076	0.978445
<i>Boe</i>	0.972451	0.968075	0.979339	0.99162	0.992891	0.993715
<i>Bun</i>	0.960575	0.954088	0.970431	0.991716	0.993077	0.993787
<i>Cal</i>	0.966576	0.962266	0.974932	0.98816	0.99012	0.99112
<i>Cam</i>	0.966576	0.962266	0.974932	0.98816	0.99012	0.99112
<i>Chi</i>	0.972451	0.968075	0.979339	0.991716	0.993077	0.993787
<i>Chm</i>	0.947317	0.938476	0.960488	0.977495	0.981256	0.983121
<i>Chr</i>	0.960575	0.954088	0.970431	0.991716	0.993077	0.993787
<i>Clt</i>	0.943152	0.94352	0.957364	0.98816	0.99012	0.99112
<i>Cml</i>	0.94677	0.938001	0.960078	0.976826	0.980504	0.98262
<i>Coc</i>	0.916828	0.913296	0.937621	0.98816	0.99012	0.99112
<i>Crg</i>	0.943152	0.94352	0.957364	0.98816	0.99012	0.99112
<i>Cru</i>	0.972451	0.968075	0.979339	0.99162	0.992891	0.993715
<i>Crb</i>	0.972451	0.968075	0.979339	0.99162	0.992891	0.993715
<i>Csa</i>	0.972451	0.968075	0.979339	0.99162	0.992891	0.993715
<i>Des</i>	0.972451	0.968075	0.979339	0.991716	0.993077	0.993787
<i>Dip</i>	0.972451	0.968075	0.979339	0.99162	0.992891	0.993715
<i>Don</i>	0.960575	0.954088	0.970431	0.991716	0.993077	0.993787
<i>Dpm</i>	0.972451	0.968075	0.979339	0.99162	0.992891	0.993715
<i>Edm</i>	0.94677	0.938001	0.960078	0.976826	0.980504	0.98262
<i>Ers</i>	0.972451	0.968075	0.979339	0.991716	0.993077	0.993787
<i>Eut</i>	0.966576	0.962266	0.974932	0.98816	0.99012	0.99112

<i>Euc</i>	0.960575	0.954088	0.970431	0.991716	0.993077	0.993787
<i>Frr</i>	0.966576	0.962266	0.974932	0.98816	0.99012	0.99112
<i>Hal</i>	0.972451	0.968075	0.979339	0.99162	0.992891	0.993715
<i>Hdr</i>	0.882394	0.862021	0.911795	0.971259	0.976076	0.978445
<i>Hes</i>	0.960575	0.954088	0.970431	0.991716	0.993077	0.993787
<i>Hml</i>	0.972451	0.968075	0.979339	0.99162	0.992891	0.993715
<i>Hph</i>	0.947317	0.938476	0.960488	0.977495	0.981256	0.983121
<i>Ibr</i>	0.882394	0.862021	0.911795	0.760208	0.82856	0.820156
<i>Ida</i>	0.944426	0.935964	0.95832	0.977104	0.980496	0.982828
<i>Ist</i>	0.966576	0.962266	0.974932	0.98816	0.99012	0.99112
<i>Krn</i>	0.937003	0.937075	0.952752	0.98816	0.99012	0.99112
<i>Lea</i>	0.972451	0.968075	0.979339	0.991716	0.993077	0.993787
<i>Lep</i>	0.972451	0.968075	0.979339	0.991716	0.993077	0.993787
<i>Lmy</i>	0.972451	0.968075	0.979339	0.991716	0.993077	0.993787
<i>Lob</i>	0.882394	0.862021	0.911795	0.871871	0.864523	0.903904
<i>Lun</i>	0.882394	0.862021	0.911795	0.971259	0.976076	0.978445
<i>Meg</i>	0.89856	0.880125	0.92392	0.970111	0.975121	0.977583
<i>Mgc</i>	0.89856	0.880125	0.92392	0.970111	0.975121	0.977583
<i>Mgr</i>	0.89856	0.880125	0.92392	0.970111	0.975121	0.977583
<i>Mic</i>	0.972451	0.968075	0.979339	0.99162	0.992891	0.993715
<i>Not</i>	0.947317	0.938476	0.960488	0.977495	0.981256	0.983121
<i>Och</i>	0.966576	0.962266	0.974932	0.98816	0.99012	0.99112
<i>Ore</i>	0.972451	0.968075	0.979339	0.99162	0.992891	0.993715
<i>Pet</i>	0.937003	0.937075	0.952752	0.98816	0.99012	0.99112
<i>Phy</i>	0.972451	0.968075	0.979339	0.990341	0.9904	0.992755
<i>Psf</i>	0.966576	0.962266	0.974932	0.98816	0.99012	0.99112
<i>Ric</i>	0.882394	0.862021	0.911795	0.971259	0.976076	0.978445
<i>Rph</i>	0.966576	0.962266	0.974932	0.98816	0.99012	0.99112
<i>Sch</i>	0.94677	0.938001	0.960078	0.976826	0.980504	0.98262
<i>Scl</i>	0.94677	0.938001	0.960078	0.977302	0.980881	0.982977
<i>Sis</i>	0.966576	0.962266	0.974932	0.98816	0.99012	0.99112
<i>Sml</i>	0.972451	0.968075	0.979339	0.991716	0.993077	0.993787
<i>SkI</i>	0.966576	0.962266	0.974932	0.98816	0.99012	0.99112
<i>Stv</i>	0.960575	0.954088	0.970431	0.991716	0.993077	0.993787
<i>Sub</i>	0.944426	0.935964	0.95832	0.977495	0.981256	0.983121
<i>Tla</i>	0.966576	0.962266	0.974932	0.98816	0.99012	0.99112
<i>Tgl</i>	0.972451	0.968075	0.979339	0.99162	0.992891	0.993715
<i>Tar</i>	0.960575	0.954088	0.970431	0.991716	0.993077	0.993787
<i>The</i>	0.966576	0.962266	0.974932	0.98816	0.99012	0.99112
<i>Tls</i>	0.966576	0.962266	0.974932	0.98816	0.99012	0.99112
<i>Trt</i>	0.966576	0.962266	0.974932	0.98816	0.99012	0.99112
<i>Ves</i>	0.960575	0.954088	0.970431	0.991716	0.993077	0.993787
<i>Yin</i>	0.972451	0.968075	0.979339	0.991716	0.993077	0.993787

Table S9. Accession information of marker genes used for infratribal placement

Alysseae phylogeny	<i>ndhF</i>
<i>Alyssoides utriculata</i>	KF022860.1
<i>Alyssum desertorum</i>	KY498535.1
<i>Alyssum harputicum</i>	KF022881.1
<i>Alyssum skopjense</i>	KF022923.1
<i>Alyssum sulphureum</i>	KF022927.1
<i>Alyssum thymops</i>	KF022929.1
<i>Arabis alpina</i> (outgroup: Arabideae)	DQ288731.1
<i>Asperuginoides axillaris</i>	this study
<i>Aurinia sinuata</i>	KF022952.1
<i>Berteroa incana</i>	LC090120.2
<i>Berteroa incana</i>	AY330097.1
<i>Berteroa mutabilis</i>	KF022956.1
<i>Berteroa obliqua</i>	KF022957.1
<i>Bornmuellera emarginata</i> , as <i>Leptoplax emarginata</i>	KF022989.1
<i>Bornmuellera tymphaea</i>	KF022963.1
<i>Brachypus suffruticosus</i> , as <i>Fibigia suffruticosa</i>	KF022978.1
<i>Clypeola aspera</i>	EU907360.1
<i>Clypeola dichotoma</i>	KF022968.1
<i>Fibigia clypeata</i>	KF022972.1
<i>Fibigia suffruticosa</i>	EU907361.1
<i>Galitzkya potaninii</i>	KF022983.1
<i>Hormathophylla reverchonii</i>	KP276171.1
<i>Hormathophylla spinosa</i>	KP276173.1
<i>Lepidotrichum uechtritzianum</i>	KF022988.1
<i>Phyllolepidum rupestre</i>	KF022993.1
<i>Physoptychis caspica</i>	KF022994.1

Anchonieae phylogeny	<i>rbcL</i>
<i>Anchonium elichrysofolium</i>	FN594834.1
<i>Bunias orientalis</i> (outgroup: Buniadeae)	KM360682.1
<i>Matthiola incana</i>	HM850161.1
<i>Matthiola longipetala</i>	KX282876.1
<i>Matthiola sinuata</i>	JN892935.1
<i>Microstigma deflexum</i>	KX527519.1
<i>Sterigmostemum eglandulosum</i>	KX434805.1
<i>Sterigmostemum grandiflorum</i>	KX434804.1
<i>Sterigmostemum incanum</i>	KX434806.1
<i>Sterigmostemum fuhaiense</i>	KX434802.1
<i>Sterigmostemum matthiolooides</i>	JN847817.1
<i>Sterigmostemum tomentosum</i>	KX434800.1
<i>Sterigmostemum sulphureum</i>	KX434803.1
<i>Sterigmostemum violaceum</i>	JN847818.1
<i>Sterigmostemum violaceum</i>	KX434801.1
<i>Systemon petrovii</i>	KX527518.1
<i>Veselskya griffithiana</i>	this study

Isatideae phylogeny	<i>rbcL</i>
<i>Conringia planisiliqua</i>	this study
<i>Isatis minima</i>	KJ765843.1
<i>Isatis oblongata</i>	KJ765844.1
<i>Isatis pachycarpa</i>	FN594830.1
<i>Isatis tinctoria</i>	JN847841.1
<i>Pachypterygium multicaule</i>	JN847842.1
<i>Schimpera arabica</i>	KX283017.1
<i>Sisymbrium irio</i> (outgroup: Sisymbrieae)	AY167982.1

Thelypodieae phylogeny	<i>ndhF</i>
<i>Brassica oleracea</i> (outgroup: Brassiceae)	DQ288742.1
<i>Catadysia rosulans</i>	EU718526.1
<i>Englerocharis pauciflora</i>	EU718543.1
<i>Hesperidanthus barnebyi</i>	EU718550.1
<i>Hesperidanthus linearifolius</i>	EU718553.1
<i>Hesperidanthus suffrutescens</i>	EU718554.1
<i>Mostacillastrum stenophyllum</i>	EU718558.1
<i>Neuontobotrys lanata</i>	EU718559.1
<i>Polypsecadium grandiflorum</i>	EU718560.1
<i>Polypsecadium rusbyi</i>	EU718562.1
<i>Raphanorhyncha crassa</i>	this study
<i>Romanschulzia costaricensis</i>	EU718564.1
<i>Sibara deserti</i>	EU718568.1
<i>Stanleya pinnata</i>	EU718574.1
<i>Stanleya pinnata</i>	EU718575.1
<i>Stanleya viridiflora</i>	EU718577.1
<i>Streptanthus barbatus</i>	EU718581.1
<i>Streptanthus tortuosus</i>	EU718598.1
<i>Thelypodopsis aurea</i>	EU718600.1
<i>Thelypodopsis elegans</i>	EU718602.1
<i>Thelypodium rollinsii</i>	EU718608.1
<i>Thysanocarpus curvipes</i>	EU718613.1
<i>Warea amplexifolia</i>	EU718614.1
<i>Warea cuneifolia</i>	EU718615.1
<i>Weberbaueria herzogii</i>	EU718616.1
<i>Weberbaueria peruviana</i>	EU718618.1

Thlaspideae phylogeny	<i>ndhF</i>
<i>Alliaria petiolata</i>	DQ288727.1
<i>Camelinopsis campylopoda</i>	this study
<i>Eutrema salsugineum</i> (outgroup: Eutremeae)	KR584659.1
<i>Graellsia saxifragifolia</i>	DQ288772.1
<i>Parlatoria rostrata</i>	DQ288809.1
<i>Peltaria alliacea</i>	KF023033.1
<i>Pseudocamelina campylopoda</i>	DQ288817.1
<i>Sobolewska caucasica</i>	KF023034.1
<i>Thlaspi arvense</i>	LC090145.2
<i>Thlaspi arvense</i>	DQ288839.1
<i>Thlaspi perfoliatum</i>	KF023022.1

Table S10. Tribal associations of unplaced taxa

Unplaced taxon	Recommended tribal assignments
<i>Andrzejowskia cardamine</i> Rchb.	Cardamineae
<i>Arabis josiae</i> Jahand. & Maire	new genus and tribe including <i>Fourraea alpina</i> , sister to Bivonaeae
<i>Arabis ottonis-schulzii</i> Bornm. & Gauba	new genus and tribe sister to Coluteocarpeae
<i>Asperuginoides axillaris</i> (Boiss. & Hohen.) Rauschert	sister to Alysseae, better sampling needed
<i>Camelinopsis campylopoda</i> (Bornm. & Gauba) A.G. Mill.*	Thlaspidiae
<i>Chamira circaeoides</i> (L. f.) Zahlbr.	sister to Heliophileae, better sampling needed
<i>Conringia planisiliqua</i> Fisch. & C.A. Mey.	new genus in Isatideae
<i>Dipoma iberideum</i> Franch.	new tribe with <i>Hemilophia</i> , sister to Microlepidieae
<i>Fourraea alpina</i> (L.) Greuter & Burdet.	new tribe with <i>Arabis josiae</i> , sister to Bivonaeae
<i>Heldreichia bupleurifolia</i> Boiss.	Biscutelleae**
<i>Hemilophia rockii</i> O.E. Schulz	new tribe with <i>Dipoma iberideum</i> , sister to Microlepidieae
<i>Idahoa scapigera</i> (Hook.) A. Nels. & J.F. Macbr.	near <i>Subularia aquatica</i> in the Mexican + South American (CES) clade, currently unresolved
<i>Lunaria rediviva</i> L.	Biscutelleae**
<i>Ochthodium aegyptiacum</i> (L.) DC.	Sisymbrieae
<i>Petrocallis pyrenaica</i> (L.) W.T. Aiton.	Kernereae
<i>Pseudofortuynia esfandiarii</i> Hedge.***	Sisymbrieae
<i>Raphanorhyncha crassa</i> Roll.	Thelypodieae
<i>Ricotia cretica</i> Boiss. & Heldr.	Biscutelleae**
<i>Schrenkiella parvula</i> (Schrenk) D.A. German & Al-Shehbaz	new monospecific tribe
<i>Subularia aquatica</i> L.	near <i>Idahoa scapigera</i> in the Mexican + South American (CES) clade, currently unresolved
<i>Veselskya griffithiana</i> (Boiss.) Opiz.	Anchonieae

* *Pseudocamelina campylopoda* (Bornm. & Gauba) Hadač & Chtrek, Esmailbegi *et al.*, 2018

** confirms placement by Özüdoğru *et al.*, 2015, 2017

*** *Sisymbrium leucocladum* (Boiss.) D.A. German & Al-Shehbaz

Table S11. Transcriptome assembly statistics

Species	Number of genes	All contigs				Only longest isoforms				Orthologous groups
		N50	Median	Average	Total length	N50	Median	Average	Total length	
<i>Aethionema arabicum</i>	66554	1672	601	1004.62	112438062	1390	426.5	795.35	52933457	35685
<i>Arabidopsis lyrata</i>	65605	1577	491	903.94	92706157	1397	355	738.29	48435496	29468
<i>Arabidopsis thaliana</i>	23003	2072	1204	1476.42	61020502	2021	1172	1403.13	32276268	16891
<i>Arabis alpina</i>	83281	1471	501	877.85	130812662	1205	347	682.73	56858816	34144
<i>Boechera stricta</i>	53468	1717	684	1062.73	110844442	1592	434	864.45	46220627	27368
<i>Capsella rubella</i>	66075	1685	564	985.39	103507838	1417	403	778.21	51419898	33615
<i>Cardamine hirsuta</i>	25273	2061	1178	1462.35	69214395	2012	1100	1361.6	34411777	17610
<i>Sisymbrium irio</i>	62074	1677	520	953.64	97873933	1520	353	768.17	47683330	26080

Table S12. Gene expression change in the core Brassicaceae

Locus	Gene Symbol	Full Name
AT1G03150		Acyl-CoA N-acyltransferases (NAT) superfamily protein
AT1G03750	<i>CHR9</i>	CHROMATIN REMODELING 9, SWITCH 2
AT1G08600	<i>ATRX</i>	P-loop containing nucleoside triphosphate hydrolases superfamily protein
AT1G09910		Rhamnogalacturonate lyase family protein
AT1G10130	<i>ECA3, ATECA3</i>	endoplasmic reticulum-type calcium-transporting ATPase 3
AT1G12450		SNARE associated Golgi protein family
AT1G26160		Metal-dependent phosphohydrolase
AT1G60000		RNA-binding (RRM/RBD/RNP motifs) family protein
AT1G62830	<i>LDL1, SWP1</i>	LSD1-like 1
AT1G64260		MuDR family transposase
AT1G65900		unknown protein
AT1G67280		Glyoxalase/Bleomycin resistance protein/Dioxygenase superfamily protein
AT1G77260		S-adenosyl-L-methionine-dependent methyltransferases superfamily protein
AT1G79120		Ubiquitin carboxyl-terminal hydrolase family protein
AT2G25010		Aminotransferase-like, plant mobile domain family protein
AT2G26280	<i>CID7</i>	CTC-interacting domain 7
AT2G32040		Major facilitator superfamily protein
AT2G33860	<i>ETT, ARF3</i>	Transcriptional factor B3 family protein / auxin-responsive factor AUX/IAA-related
AT2G42610	<i>LSH10</i>	LIGHT SENSITIVE HYPOCOTYLS 10 (DUF640)
AT2G47250		RNA helicase family protein
AT3G03950	<i>ECT1</i>	evolutionarily conserved C-terminal region 1
AT3G04610	<i>FLK</i>	RNA-binding KH domain-containing protein
AT3G07130	<i>ATPAP15, PAP15</i>	purple acid phosphatase 15
AT3G11230		Yippee family putative zinc-binding protein
AT3G12150		alpha/beta hydrolase family protein
AT3G14270	<i>FAB1B</i>	phosphatidylinositol-4-phosphate 5-kinase family protein
AT3G15000	<i>MORF8</i>	MULTIPLE ORGANELLAR RNA EDITING FACTOR 8
AT3G19910		RING/U-box superfamily protein
AT3G23150	<i>ETR2</i>	Signal transduction histidine kinase, hybrid-type, ethylene sensor
AT3G51480	<i>ATGLR3.6, GLR3.6</i>	glutamate receptor 3.6
AT3G53540		afadin
AT3G58030		RING/U-box superfamily protein
AT3G62600	<i>ATERDJ3B, ERDJ3B</i>	DNAJ heat shock family protein
AT3G66654		Cyclophilin-like peptidyl-prolyl cis-trans isomerase family protein
AT4G03270	<i>CYCD6;1</i>	Cyclin D6;1

AT4G04885	<i>PCFS4</i>	PCF11P-similar protein 4
AT4G12130		Glycine cleavage T-protein family
AT4G25030		Serine/Threonine-kinase
AT4G30000		Dihydropterin pyrophosphokinase / Dihydropteroate synthase
AT4G31430	<i>KAKU4</i>	(JAPANESE FOR NUCLEUS) 4
AT4G32620		Enhancer of polycomb-like transcription factor protein
AT4G34310		alpha/beta-Hydrolases superfamily protein
AT4G35870		early-responsive to dehydration stress protein (ERD4)
AT4G39530		Tetratricopeptide repeat (TPR)-like superfamily protein
AT5G04410	<i>NAC2, anac078</i>	NAC domain containing protein 2
AT5G04430	<i>BTR1, BTR1L</i>	binding to TOMV RNA 1L (long form)
AT5G04530	<i>KCS19</i>	3-ketoacyl-CoA synthase 19
AT5G11390	<i>WIT1</i>	WPP domain-interacting protein 1
AT5G12120		Ubiquitin-associated/translation elongation factor EF1B protein
AT5G12130	<i>PDE149, ATTERC</i>	integral membrane TerC family protein
AT5G15340		Pentatricopeptide repeat (PPR) superfamily protein
AT5G17900		microfibrillar-associated protein-related
AT5G21140	<i>emb1379</i>	embryo defective 1379
AT5G42450		Pentatricopeptide repeat (PPR) superfamily protein
AT5G46840		RNA-binding (RRM/RBD/RNP motifs) family protein
AT5G49530		SIN-like family protein *
AT5G53080		Tetratricopeptide repeat (TPR)-like superfamily protein
AT5G56730		Insulinase (Peptidase family M16) protein
AT5G56930	<i>emb1789</i>	CCCH-type zinc finger family protein
AT5G57870	<i>eIFiso4G1</i>	MIF4G domain-containing protein / MA3 domain-containing protein
AT5G59710	<i>VIP2, AtVIP2</i>	VIRE2 interacting protein 2
AT5G62710		Leucine-rich repeat protein kinase family protein
AT5G63940		Protein kinase protein with adenine nucleotide alpha hydrolases-like domain
AT5G64830		programmed cell death 2 C-terminal domain-containing protein

Table S13. Gene expression change leading to complex-leaved Brassicaceae

Locus	Gene Symbol	Full Name
AT1G05370		Sec14p-like phosphatidylinositol transfer family protein
AT1G09010		glycoside hydrolase family 2 protein
AT1G17930		Aminotransferase-like, plant mobile domain family protein
AT1G19485		Transducin/WD40 repeat-like superfamily protein
AT1G20510	<i>OPCL1</i>	OPC-8:0 CoA ligase1
AT1G26500		Pentatricopeptide repeat (PPR) superfamily protein
AT1G76990	<i>ACR3</i>	ACT domain repeat 3
AT1G77250		RING/FYVE/PHD-type zinc finger family protein
AT1G77930		Chaperone DnaJ-domain superfamily protein
AT2G01450	<i>ATMPK17, MPK17</i>	MAP kinase 17
AT2G14680	<i>MEE13</i>	myosin heavy chain-related
AT2G26570	<i>WEB1</i>	WEAK CHLOROPLAST MOVEMENT UNDER BLUE LIGHT-like protein (DUF827)
AT2G41520	<i>TPR15</i>	Heat shock protein DnaJ with tetratricopeptide repeat
AT2G42880	<i>ATMPK20, MPK20</i>	MAP kinase 20
AT3G01810		EEIG1/EHBP1 protein amino-terminal domain protein
AT3G05510		Phospholipid/glycerol acyltransferase family protein
AT3G11945	<i>PDS2, ATHST, HST</i>	homogentisate prenyltransferase
AT3G19720	<i>ARC5, DRP5B</i>	P-loop containing nucleoside triphosphate hydrolases superfamily protein
AT3G43540		initiation factor 4F subunit (DUF1350)
AT3G53320	<i>GPT2</i>	GROWING PLUS-END TRACKING PROTEIN 2
AT3G61360		Tetratricopeptide repeat (TPR)-like superfamily protein
AT4G01730		DHHC-type zinc finger family protein
AT4G18060	<i>SH3P3</i>	SH3 domain-containing protein
AT4G24530		O-fucosyltransferase family protein
AT4G29490		Metallopeptidase M24 family protein
AT4G38350	<i>ATNPC1-2</i>	Patched family protein
AT5G45030		Trypsin family protein
AT5G49530		SIN-like family protein
AT5G51740		Peptidase family M48 family protein

Table S14. Genes and protein residues under positive selection in complex-leaved species

Locus	Gene Symbol	Full Name	sites under selection (P > 95%)	sites under selection (P > 99%)
AT1G05790		lipase class 3 family protein		
AT1G08030	<i>TPST</i>	tyrosylprotein sulfotransferase	21M 28T	
AT1G08210		Eukaryotic aspartyl protease family protein	503F	
AT1G08600	<i>ATRX</i>	P-loop containing nucleoside triphosphate hydrolases superfamily protein	1V 6E 9S 56S 57S 129S 149S 169K 189S 202S 211S 225S 236L 280P 284N 301R 311S 379S 386T 396S	
AT1G13120	<i>ATGLE1, GLE1</i>	nucleoporin GLE1-like protein		
AT1G18750	<i>AGL65</i>	AGAMOUS-like 65	360-	325-
AT1G19010		unknown protein		
AT1G19485		Transducin/WD40 repeat-like superfamily protein	5- 6- 8- 13- 23- 28-	10-
AT1G20720		RAD3-like DNA-binding helicase protein	410M	
AT1G22310	<i>MBD8, ATMBD8</i>	methyl-CPG-binding domain 8	263V	112R
AT1G23360	<i>MENG</i>	S-adenosyl-L-methionine-dependent methyltransferases superfamily protein	1N 23L	22R
AT1G26330		DNA binding	19L	
AT1G27980	<i>DPL1, ATDPL1</i>	dihydrosphingosine phosphate lyase		
AT1G34355	<i>ATPS1, PSI</i>	forkhead-associated (FHA) domain-containing protein		
AT1G34360		translation initiation factor 3 (IF-3) family protein	53D	
AT1G59760		R helicase, ATP-dependent, SK12/DOB1 protein		
AT1G61667		serine protease (DUF538)		
AT1G67420		Zn-dependent exopeptidases superfamily protein	2- 4- 31E	18- 25A 30F
AT1G68890		magnesium ion binding; thiamin pyrophosphate binding; hydrolyases; 2-succinyl-5-enolpyruvyl-6-hydroxy-3-cyclo		
AT1G70570		anthranilate phosphoribosyltransferase, putative		
AT1G72416		Chaperone DJ-domain superfamily protein	138V	
AT1G72500		inter alpha-trypsin inhibitor, heavy chain-like protein		
AT1G73730	<i>EIL3, SLIM1, ATSLIM, AtEIL3</i>	ETHYLENE-INSENSITIVE3-like 3	569G	6P
AT1G73990	<i>SPPA, SPPA1</i>	sigI peptide peptidase	2N	
AT1G74700	<i>NUZ, TRZ1</i>	tRNase Z1	2V	
AT1G75100	<i>JAC1</i>	J-domain protein required for chloroplast accumulation response 1	267S	
AT1G75150		DNA ligase-like protein	2E 197T	
AT1G79440	<i>ALDH5F1, SSADH1, SSADH</i>	aldehyde dehydrogenase 5F1	11D 14S 17R	12L 15P 16R
AT1G79890		RAD3-like DNA-binding helicase protein	10S 117K	
AT2G04340		cytoplasmic dynein 2 light intermediate chain	155D 156V 162A 163R 167S	
AT2G13600		Pentatricopeptide repeat (PPR) superfamily protein		
AT2G19600	<i>ATKEA4, KEA4</i>	K+ efflux antiporter 4	14- 21-	15-
AT2G21370	<i>XK-1, XK1</i>	xylulose kise-1	8A 9S 10E	
AT2G24640	<i>UBP19</i>	ubiquitin-specific protease 19	147G 569D 625L 631D 633E 637A	558G 565V 596D 624P 636D
AT2G27760	<i>ATIPT2, IPPT, IPT2</i>	tRNA isopentenyltransferase 2		
AT2G27800		Tetratricopeptide repeat (TPR)-like superfamily protein		3-
AT2G27900		Protein of unknown function DUF2451		
AT2G30530	<i>MADA4</i>	zinc finger CCCH domain protein	29-	
AT2G32810	<i>BGAL9</i>	beta galactosidase 9	892L 894F	
AT2G32950	<i>COPI, DET340, FUS1, EMB168</i>	Transducin/WD40 repeat-like superfamily protein	3T	
AT2G33490		hydroxyproline-rich glycoprotein family protein		
AT2G35130		Tetratricopeptide repeat (TPR)-like superfamily protein	2I 4S 6C	8L
AT2G36960	<i>TKI1</i>	TSL-kise interacting protein 1		
AT2G38330		MATE efflux family protein		
AT2G39340		SAC3/GANP/Nin1/mts3/eIF-3 p25 family	115H	
AT2G43210		Ubiquitin-like superfamily protein		
AT2G45460		SMAD/FHA domain-containing protein		

AT2G45670		calcineurin B subunit-related		
AT3G03630	CS26	cysteine synthase 26	10A 12D	
AT3G06980		DEA(D/H)-box R helicase family protein		
AT3G07740	ADA2A	homolog of yeast ADA2 2A	4S 11D	
AT3G15130		Tetratricopeptide repeat (TPR)-like superfamily protein		
AT3G15290		3-hydroxyacyl-CoA dehydrogenase family protein	306Q	
AT3G16565		alanine-tRNA ligases		168V
AT3G18215		Protein of unknown function, DUF599		
AT3G25600		Calcium-binding EF-hand family protein	15Q	
AT3G45890	RUS1	Protein of unknown function, DUF647	95G	
AT3G49320		Metal-dependent protein hydrolase		4E
AT3G53920	SIGC, SIG3	Rpolymerase sigma-subunit C		
AT3G55080		SET domain-containing protein	89T	
AT3G59090		tobamovirus multiplication protein		397H
AT3G59680		Serine/Threonine-kinase	8F 15L 16S	27I 29F 30M 35T 36S 38A
AT3G59780		Rhodanese/Cell cycle control phosphatase superfamily protein	89R 109T	
AT3G61540		alpha/beta-Hydrolases superfamily protein	53G	
AT4G00980		zinc knuckle (CCHC-type) family protein		
AT4G03110	AtRBP-DR1, RBP-DR1	R-binding protein-defense related 1	407A 409Q 413N 414M 417G 421G 428Q 436G 443- 446S	399F 401S 402Y 403D 404S 410N 420L 430K 439-
AT4G03960		Phosphotyrosine protein phosphatases superfamily protein	6F 204Y	
AT4G11970		YTH family protein	408R	
AT4G16510		YbaK/aminoacyl-tRNA synthetase-associated domain	14K 23A	18I
AT4G16765		2-oxoglutarate (2OG) and Fe(II)-dependent oxygenase superfamily protein	7F	5S 6K 10E
AT4G18600	WAVE5, ATSCAR-LIKE, SCARL	SCAR family protein		
AT4G19020	CMT2	chromomethylase 2	788S	
AT4G24060		Dof-type zinc finger DNA-binding family protein	5G	
AT4G26660		kinesin-like protein	13- 16L 25L 89H 137L	
AT4G27510		2-isopropylmalate synthase		
AT4G27640		ARM repeat superfamily protein		
AT4G32190		Myosin heavy chain-related protein		
AT4G34530	CIB1	cryptochrome-interacting basic-helix-loop-helix 1	14L 15N	2F 7F 16P
AT4G35250		D(P)-binding Rossmann-fold superfamily protein	4K 7A 10V 11V	
AT4G39410	WRKY13, ATWRKY13	WRKY DNA-binding protein 13	2Q	
AT5G01030		Protein of unknown function (DUF3527)		
AT5G01950		Leucine-rich repeat protein kisse family protein		
AT5G06610		lipase (DUF620)	10E 372T	
AT5G08470	PEX1	peroxisome 1		
AT5G11040	TRS120, AtTRS120	TRS120		
AT5G11640		Thioredoxin superfamily protein	11F	
AT5G13410		FKBP-like peptidyl-prolyl cis-trans isomerase family protein		
AT5G14970		seed maturation-like protein	10-	8-
AT5G15410	DND1, ATCNGC2	Cyclic nucleotide-regulated ion channel family protein		
AT5G16715	EMB2247	ATP binding; valine-tRNA ligases; aminoacyl-tRNA ligases		
AT5G23050	AAE17	acyl-activating enzyme 17		
AT5G35930		AMP-dependent synthetase and ligase family protein	113S	
AT5G36740		Acyl-CoA N-acyltransferase with RING/FYVE/PHD-type zinc finger protein	609K	
AT5G45710	AT-HSFA4C, RHA1, HSFA4C	winged-helix DNA-binding transcription factor family protein		207N
AT5G46390		Peptidase S41 family protein		
AT5G47860		Gut esterase (DUF1350)	29K	
AT5G48030	GFA2	gametophytic factor 2		
AT5G51690	ACS12	1-amino-cyclopropane-1-carboxylate synthase 12		6S 7T
AT5G55760	SRT1	sirtuin 1	15P	
AT5G62770		membrane-associated kisse regulator, putative (DUF1645)	239E	
AT5G63990		Inositol monophosphatase family protein	11R	