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#### NON-GENOMIC EFFECTS OF GLUCOCORTICOIDS: AN UPDATED VIEW

2

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#### 17 Keywords

18 Corticosteroids, glucocorticoid receptor, non-genomic, asthma, airway remodeling, smooth19 muscle.

20

#### 21 Abstract

Glucocorticoid (GC) anti-inflammatory effects generally require a prolonged onset of action and involve genomic processes. Because of the rapidity of some of GC effects, however, the concept that non-genomic actions may contribute to GC mechanisms of action has arisen. While the mechanisms have not been completely elucidated, the non-genomic effects may play a role in the management of inflammatory diseases. For instance, we recently reported that GC "rapidly" enhanced the effects of bronchodilators, agents used in the treatment of allergic asthma. In this
review, we will discuss i) the non-genomic effects of GCs on pathways relevant to the pathogenesis
of inflammatory diseases and ii) the putative role of membrane GC receptor. Since GC side effects
are often considered to be generated through its genomic actions, understanding GC non-genomic
effects will help design GCs with a better therapeutic index.

#### 32 Mechanism of action of glucocorticoids (GC).

GCs primarily mediate their effects by activating the ubiquitously expressed intracellular GC 33 receptor (GR) (see Glossary) [1]. In its inactive state, the GR resides in the cytoplasm and, upon 34 35 ligand activation, translocates to the cell nucleus to interact with GC response elements (GREs) thereby producing genomic effects that alter protein expression. Interestingly, evidence suggests 36 37 that GCs also manifest almost immediate **non-genomic actions** on several signaling processes [2]. 38 GC non-genomic effects involve non-specific interactions with the cell membrane, or specific 39 interactions with cytosolic GRs (cGR) or membrane-bound GRs (mGR) (Table 1). This report 40 summarizes the current knowledge on non-genomic effects of GCs, with a focus on GR-mediated 41 events and GR-associated signaling pathways. Where appropriate, potential links to inflammatory diseases will be highlighted in the main text and their potential impact will be discussed in Box 1 42 and 2. 43

44

#### 45 GCs exert rapid effects on levels of intracellular calcium.

46 Studies suggest that GC rapidly (within seconds) modulates basal intracellular calcium levels
47 and agonist-induced calcium mobilization (Tables 2 & 3).

48 Effects of GCs on intracellular calcium homeostasis. GCs can increase or decrease cytosolic calcium depending on the cell type. Evidence from non-immune cells, such as primary or 49 immortalized human bronchial epithelial cells, consistently demonstrate that acute exposure to 50 GC, and to a lesser extent to the mineralocorticoid (MC), aldosterone, reduces basal  $[Ca^{2+}]i[3, 4]$ . 51 Similarly, in rat thymocytes [5] and mouse neuroblastoma cells [6]  $[Ca^{2+}]i$  decreased following 52 53 acute exposure to GC, and in cichlid fish pituitary cells cortisol inhibited [Ca<sup>2+</sup>]i and reduced prolactin secretion [7]. However, in immune cells, it is unclear if GCs genuinely exert non-54 genomic effects on basal [Ca<sup>2+</sup>]i. For example, while acute exposure to GC was reported to 55 decrease [Ca<sup>2+</sup>]i in leukocytes, these leukocytes were obtained from donors who were treated with 56 oral prednisolone for 7 days [8], potentially confounding the results of the study. Similarly, studies 57 in human lymphoblasts show that cortisol markedly reduced basal [Ca<sup>2+</sup>]i only after 48 hrs of 58 treatment [9]. These data argue against a role for non-genomic effects of GC in altering basal 59  $[Ca^{2+}]i$  in immune cells. 60

61 With regard to the lungs, evidence supports that a variety of GCs differentially modulate basal [Ca<sup>2+</sup>]i upon immediate exposure. For instance, the acute inhibitory effects of dexamethasone 62 (within 30 seconds) on basal  $[Ca^{2+}]i$  in bronchial epithelial cells were comparable to triamcinolone 63 acetonide and hydrocortisone but not to budesonide [10]. Interestingly, the GR antagonist RU486 64 and the protein synthesis inhibitor cycloheximide failed to prevent these acute GC effects, 65 suggesting the involvement of GR-independent and non-genomic pathways. These observed 66 67 effects could be due to the various degrees of lipophilicity among GCs, as well as direct interactions of GCs with the cell membrane [10]. Non-genomic mechanisms have been proposed 68 mostly based on the use of pharmacological inhibitors. Urbach and colleagues found that the rapid 69 GC effects involved pathways regulated by the SERCA type  $Ca^{2+}$ -ATPase pump, adenylyl cyclase 70 71 and protein kinase A (PKA) but not protein kinase C (PKC) [10]. Collectively, these studies show the complexity of mechanisms involved in the rapid, GR-independent effect of GCs on [Ca<sup>2+</sup>]i, 72 which likely occurs through an adenylyl cyclase/PKA mediated stimulation of a thapsigargin 73 sensitive Ca<sup>2+</sup>-ATPase [10]. 74

Conversely, acute stimulatory effects of GC on basal calcium levels have been documented. A 75 brief exposure to GC can increase  $[Ca^{2+}]i$  in several cell types. For example, in mouse cortical 76 collecting duct cells, dexamethasone and aldosterone increased  $[Ca^{2+}]i$ . Interestingly, the effect of 77 aldosterone was mediated by a non-genomic activation of PKC pathway as evidenced by the 78 abolishment of its effect on basal  $[Ca^{2+}]i$  in the presence of the PKC inhibitor, chelerythrine 79 80 chloride, but not the mRNA synthesis inhibitor, actinomycin D [11]. Similarly, in rat vascular smooth muscle cells, GCs rapidly increased [Ca<sup>2+</sup>]i [12] potentially through GC-mediated 81 increases in inositol 1,4,5-triphosphate (IP3) levels associated with the translocation of the 82 calcium- and lipid-dependent PKC from the cytosolic to the membranous compartment [13]. In 83 84 these cells, while the administration of epinephrine by itself had little effect on IP3 levels, epinephrine potentiated the rapid response induced by cortisol [13]. Collectively, these findings 85 highlight a role of PKC in the rapid increase of basal  $[Ca^{2+}]i$  by GCs. 86

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*Effects of GCs on agonist-induced calcium mobilization.* The effects of GCs on agonistinduced calcium mobilization are variable depending on the agonist, the extra-cellular stimuli and the cell type. Evidence suggests that GCs rapidly inhibit, at least partially, the ability of adenosine triphosphate (ATP) to increase  $[Ca^{2+}]i$  in some cell types. In human bronchial epithelial cells for

92 example, 15 min exposure to dexame thas one (1 nM) markedly reduced ATP-induced increases in  $[Ca^{2+}]i$ . The ATP-induced  $Ca^{2+}$  response was independent of extracellular calcium but did involve 93 a Ca<sup>2+</sup>-mobilization from thapsigargin-sensitive intracellular stores [10]. Similarly, in murine HT4 94 neuroblastoma cells, acute (5 min) pre-incubation with corticosterone dose-dependently inhibited 95  $[Ca^{2+}]$  i signals induced by ATP [6]. Unlike in human bronchial epithelial cells, the  $Ca^{2+}$ -response 96 induced by ATP in these cells relies on  $Ca^{2+}$ -influx across the plasma membrane and  $Ca^{2+}$ -release 97 from intracellular stores [6]. Inhibition of PKA abrogated the inhibitory action of corticosterone 98 on ATP-induced Ca<sup>2+</sup>-elevation, whereas little influence was observed with respect to PKC 99 inhibition. Additional studies demonstrated that these GC inhibitory effects were unaffected by 100 GR blockade. These key findings obtained from studies in HT4 cells suggest that GC activates 101 102 membrane-initiated, non-genomic, PKA-dependent, PKC-independent pathways [6, 14]. In contrast, in rat B103 neuroblastoma cells, the inhibitory effects of corticosterone on serotonin-103 induced peak  $[Ca^{2+}]i$  were found to be PKC-dependent [15]. Together, these studies suggest that 104 the mechanisms mediating the acute non-genomic effects of GC on agonist-evoked calcium 105 106 mobilization are stimuli and cell type-dependent.

107 In contrast to human bronchial epithelial and murine HT4 neuroblastoma cells, pretreatment of guinea pig cochlear spiral ganglion neurons (SGN) with dexamethasone (10 min) 108 enhanced ATP-induced Ca<sup>2+</sup>-mobilization [16]. This effect was prevented in the presence of a GR 109 antagonist and mediated by rapid Ca<sup>2+</sup>-influx through activation of ionotropic purinergic P2X 110 receptors [16]. Of note, all P2X subtypes are expressed in SGN albeit to different extents [17, 18]. 111 Similarly, in rat hippocampal neurons, pretreatment with corticosterone or dexamethasone for 10-112 20 min prolonged N-methyl-D-aspartate (NMDA)-induced transient elevation in [Ca<sup>2+</sup>]i [19]. 113 Importantly, the steroid effect was reversed by the removal of corticosterone indicating that the 114 steroid effect was not due to irreversible impairment of Ca<sup>2+</sup>-extrusion from the neurons. 115 Thapsigargin and cyclohexamide had little effect on the potentiating effect of corticosterone, 116 excluding the involvement of a thapsigargin sensitive Ca<sup>2+</sup>-ATPase or *de novo* protein synthesis, 117 respectively. Interestingly, the GC effect was reproduced by the use of a membrane impermeable 118 BSA-conjugated cortisol, suggesting that mGR likely underlies the rapid non-genomic effects of 119 GC [19]. However, canonical genomic actions of GC can also alter Ca<sup>2+</sup> mobilization. In human 120 lymphoblasts, while cortisol reduced basal  $[Ca^{2+}]i$  (as indicated above),  $Ca^{2+}$ -mobilization induced 121

by platelet activating factor (PAF) is enhanced only by chronic treatment (48 hrs) with cortisol [9](Figure 1).

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### 125

#### GCs rapidly modulate skeletal and smooth muscle function.

126 Several studies have reported variable acute effects of GCs on **muscle reactivity** and tone. The specific example of airway smooth muscle cells in the pathogenesis of inflammatory diseases 127 is highlighted in **Box 1**. In mouse skeletal myotubes (C2C12 immortalized myoblasts), treatment 128 with dexamethasone (for less than 20 min) reduced glucose uptake induced by electrical pulse 129 stimulation (EPS)-mediated contraction, in a Ca<sup>2+</sup>/calmodulin protein kinase II (CaMKII) and 130 AMP activated protein kinase (AMPK) dependent fashion [20]. The effects were unaffected by 131 132 blockade of GR (RU486) or inhibition of protein synthesis (cyclohexamide), indicating a rapid non-genomic and GR-independent effect. In another study, cortisol synergized with isoprenaline 133 134 in reducing tracheal spasms in response to histamine [21]. The spasmolytic effect was fully prevented in the presence of RU486 (implicating a GR-dependent pathway), partially reduced by 135 PKC inhibition, but was unaffected by actinomycin D (excluding *de novo* RNA synthesis) again 136 suggesting a non-genomic, GR-mediated signaling pathway involving PKC [21]. 137

Other studies support a role for GCs in rapidly reducing airway smooth muscle (ASM) 138 tone. Pretreatment with budesonide (within 15 min) suppressed histamine-induced isometric 139 tension in guinea pig tracheal rings and shrinkage in individual tracheal ASM cells; effects that 140 were unaffected by cycloheximide (suggesting non-genomic actions by budesonide) [22]. Unlike 141 the findings by Wang and colleagues [21], these budesonide effects were insensitive to RU486, 142 excluding classic GR involvement [22]. Similarly, in murine ASM cells, exposure to 143 dexamethasone for 10 min decreased basal  $[Ca^{2+}]i$  and reduced peak elevations in  $[Ca^{2+}]i$  induced 144 145 by acetylcholine, effects that were insensitive to GR blockade and cycloheximide [23]. Consistently, studies using an *in vivo* guinea pig model of asthma, an established model to study 146 allergen-induced asthmatic reactions and airway hyperresponsiveness [24], revealed a beneficial 147 effect on ovalbumin-induced changes in lung resistance and compliance by acutely inhaled 148 budesonide. The protective effects of budesonide were evident within 10 minutes, suggesting a 149 150 non-genomic course of action [25]. In summary, GCs have acute spasmolytic actions in ASM that 151 can require both GR-dependent and -independent pathways, and potentially PKC-mediated152 signaling.

153 A recent study in rat vascular smooth muscle cells under conditions of lipopolysaccharide 154 (LPS)-induced septic shock showed that dexamethasone treatment for 10 min promotes norepinephrine (NE)-induced phosphorylation of key proteins associated with contraction [26]. 155 156 While no significant effect on myosin light chain 20 (MLC20) phosphorylation was observed after 157 exposure to either dexamethasone or NE alone, the combined treatment markedly enhanced 158 phospho-MLC20, an effect that was unaltered by GR blockade with RU486. Interestingly, inhibition of Rho-kinase with Y-27632 completely reversed the potentiating effects of 159 160 dexamethasone on NE-induced phospho-MLC20. Together, these findings could be of clinical 161 significance and indicate that the impaired vascular response to NE observed in septic shock may be restored by short-term exposure to dexamethasone through non-genomic activation of Rho-162 163 kinase activity [26].

164

# 165 GCs exert rapid effects on Reactive Oxygen Species (ROS)/Reactive Nitrogen Species 166 (RNS).

Studies demonstrated a rapid effect of GCs on ROS generation and the involvement of 167 nitric oxide (NO) in mediating some GC effects. An example of the role NO/ROS in the 168 pathogenesis of inflammatory disease is highlighted in Box 2. In breast cancer cells, cortisol 169 170 rapidly increased levels of ROS and RNS (as early as 15 min) and induced DNA damage. The GR antagonist (RU486) blocked the cortisol effect while L-NAME and 1400 W dihydrochloride 171 172 demonstrated the involvement of nitric oxide synthase (NOS) and inducible (i)NOS, respectively. 173 The pharmacological inhibition of Src by PP2 prevented GC-induced RNS elevation, suggesting 174 the ability of GC to rapidly stimulate Src- and iNOS-dependent release of damaging RNS levels [27]. 175

Rapid effects of GCs on endothelial NOS (eNOS), an important mediator of vascular
integrity with anti-inflammatory, anti-ischemic, and anti-atherogenic properties, have been
described as well [28-30]. Indeed, the treatment of human vascular endothelial cells with
dexamethasone rapidly enhanced (as early as 10 min), in a concentration-dependent manner, eNOS
activity, NO-production and NO-dependent vasorelaxation [31]. These GC effects were abrogated

by RU486, PI3-kinase inhibitors wortmannin and LY292002, or L-NAME, but not by the
transcriptional inhibitor actinomycin D.

Additional evidence supporting rapid effects of GCs on NOS/NO showed an augmented ATP-induced, NOS-dependent NO release in guinea pig type I spiral ganglion neurons by dexamethasone that was thought to be a consequence of ATP-induced  $[Ca^{2+}]i$  [16]. Similarly, GRmediated increases in  $[Ca^{2+}]i$ , eNOS phosphorylation, and NO production, were observed in human umbilical vein endothelial cells [32]. Interestingly, NO production increased  $[Ca^{2+}]i$ originating from intracellular and extracellular Ca<sup>2+</sup> sources [32].

The PI3K/Akt pathway is critical in the activation of NO signaling, e.g. phosphorylation 189 of eNOS [33], and the involvement of this pathway in the rapid effects of GCs has been 190 documented [33]. For example, dexamethasone rapidly increased (within 20 min), in a dose-191 dependent manner, GR-dependent phosphorylation and activation of PI3K as demonstrated by 192 phosphorylation of Akt and glycogen synthase kinase (GSK)-3, indicating that GCs can 193 functionally activate PI3K and downstream targets in human endothelial cells [31]. The potential 194 clinical relevance of these observations was confirmed in two different mouse models of ischemic 195 196 injury (i.e. transient myocardial ischemia and transient focal cerebral ischemia) where GC exerted rapid protective effects (within 30 min) via GR-dependent activation of PI3K and eNOS pathways 197 as evidenced by the administration of RU486, wortmannin and L-NAME, respectively [31, 32]. 198 Additional studies in COS-7 cells demonstrated a key role for GR in GC-induced activation of the 199 200 PI3K/Akt pathway. When cells were transfected with a dimerization-defective GR mutant (A458T, a construct that is unable to bind DNA and transactivate GC target genes), acute dexamethasone 201 202 stimulation still activated the PI3K/Akt pathway [34]. Together, these findings suggest the involvement of a non-transcriptional/non-genomic mechanism in the GR-dependent activation of 203 204 PI3K/Akt by GCs.

Since NO signaling plays a key role in chronic airway inflammatory diseases, such as asthma and COPD [35], we believe that the cross-talk between GC and NO signaling warrants further investigation to determine whether the rapid effects of GC on NO signaling would be beneficial or detrimental in disease pathogenesis.

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GCs exert acute effects on inflammatory and apoptotic pathways.

211 Evidence shows rapid non-transcriptional actions of GCs on inflammation both in 212 transformed cells and immune cells. In transformed cells, such as A549 adenocarcinoma cells, 213 acute exposure (as early as 1 min) to dexamethasone rapidly inhibited epidermal growth factor (EGF)-induced arachidonic acid (AA) release, an important mediator of inflammation [36]. This 214 inhibitory effect was due to hindering the recruitment of Grb2, p21ras and Raf to the EGF receptor 215 (EGFR) through a GR-dependent (RU486-sensitive) and transcription-independent (actinomycin 216 217 D-insensitive) mechanism. The inhibition of Grb2 recruitment was accompanied by lipocortin-1 recruitment to EGFR in the cell membrane. Subsequently, lipocortin-1 competitively inhibited 218 Grb2 binding to EGFR, thereby blocking the recruitment of critical signaling molecules necessary 219 for EGF actions [36]. 220

The acute effects of GCs on inflammatory pathways were also observed in immune cells, 221 such as human neutrophils, where acute exposure (5 min) to methylprednisolone or hydrocortisone 222 significantly inhibited N-formyl-methionyl-leucyl-phenylalanine (fMLP)-induced neutrophil 223 224 degranulation, effects that were not prevented by RU486 or cycloheximide treatments, suggesting the involvement of GR-independent and non-genomic pathways [37]. Also, in murine 225 226 macrophages acutely treated with dexamethasone (30 min), toll like receptor 9 (TLR9)-induced activation of different inflammatory signaling pathways, such as those involving NF-kB and 227 mitogen-activated protein kinases (MAPKs), was dramatically suppressed [38]. Following TLR-9 228 229 engagement, IL-1R-associated kinase 1 (IRAK1) is recruited to the cell membrane. A critical step in activating the TLR signaling cascade is the ubiquitination of IRAK1 through its physical 230 interaction with the E3 ligase,  $\beta$ -TrCP. Such ubiquitination and degradation of IRAK1 promotes 231 the trafficking of the "TNFR-associated factor 6 (TRAF6)- TAK1 adaptor proteins (TAB)-232 Transforming growth factor beta-activated kinase 1 (TAK1)" complex to the cytosol to 233 subsequently induce MAPK and NF-KB activation. Dexamethasone inhibition of IRAK1 234 ubiquitination did not occur in the presence of RU486 suggesting the involvement of GR-235 dependent mechanisms [38]. Further investigation of the molecular mechanisms revealed that by 236 237 physically interacting with IRAK1, GR interferes with the interaction between β-TrCP and IRAK1 thereby impeding its ubiquitination, a critical step in the activation of the TLR9-dependent 238 239 inflammatory cascade [38].

Rapid GC treatment can also exert pro-inflammatory action in other cell types. For example, in PC12 cells (cell line derived from rat adrenal gland), corticosterone induced rapid activation (within 15 min) of ERK1/2, p38, and JNK in a PKC-dependent manner [39, 40]. The
activation of MAPK pathways following GC treatment appears to be mediated by the putative
mGR, since corticosterone-BSA can rapidly (with 15 min) activate all MAPKs [39, 40]. Similarly,
in rat vascular smooth muscle cells, dexamethasone either alone or in combination with NE,
rapidly (within 10 min) induces ERK1/2 and p38 MAPK activities [26]. Thus, in certain cells, GCs
can activate MAPK in a non-genomic manner.

248 In CCRF-CEM cells, cell line derived from human T-cells (from pediatric ALL patients), sensitivity to acute dexamethasone induced cell death was determined in the presence and absence 249 250 of phosphodiesterase (PDE) inhibitors [41]. Non-specific PDE and specific PDE4 inhibition 251 reversed steroid resistance and markedly increased sensitivity to dexamethasone. This effect is 252 likely due to increased cAMP levels, consistent with abundant documentation on interactions between GR and cAMP pathways in the induction of apoptosis in lymphoid cells by both [42, 43]. 253 254 To date, the mechanisms of cAMP-induced apoptosis are unclear, but the presence of GR appears 255 to be required, even in the absence of GCs. For instance, in parental T-cells, elevation of cAMP, with either forskolin or dibutyryl cAMP, induced apoptotic cells death, whereas GR deficient cells 256 were insensitive to the apoptotic effects of cAMP elevation. When GR expression was 257 258 reconstituted by transfection, not only was GC sensitivity restored, but the sensitivity to cytolytic 259 effects induced by cAMP was promoted as well [42].

260 The effects of GCs on the mitochondrial control of cell metabolism and apoptosis have been extensively reviewed elsewhere [44-47]. For instance, Sekeris and colleagues were the first 261 to discover the presence of GR in mitochondria [48]. Through its acute non-genomic effects, GCs 262 promote mitochondrial apoptotic pathways resulting in the disruption of the mitochondrial 263 264 membrane-potential and the release of pro-apoptotic factors such as Cytochrome C [49]. Importantly, the translocation of GR from the cytoplasm to the mitochondria correlates with the 265 sensitivity of a given cell type to GC-induced apoptosis [50, 51]. In line with this, recent studies 266 267 in mouse thymocytes showed that short term treatment with GC induces a direct interaction of GR with the pro-apoptotic Bcl2 family member associated proteins such as Bim [52]. Such interaction 268 subsequently activates Bax decreasing thereby the mitochondrial membrane potential, 269 270 Cytochrome C release, and Caspase-9 activation. However, it important to note that the effects of GC on the mitochondria control of apoptosis involve also genomic pathways. For example, in 271 murine neuronal stem cells, dexamethasone was able to augment 2,3-methoxy-1,4-272

naphthoquinone-induced apoptosis where a large percentage of studied genes involved in the
mitochondrial respiratory chain and some encoding for anti-oxidant enzymes were downregulated
by long-term treatment with GC [53]. These events allowed GCs to increase cellular sensitivity to
oxidative stress promoting thereby neurotoxicity. This is clinically relevant as it can occur during
prenatal exposure of the fetal brain to excess GCs [53].

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#### Potential role of a putative mGR in mediating the rapid effects of GCs.

279 As previously described, the rapid non-genomic effects can, at least in part, be mediated through a putative mGR. Over the years, caveolin-1 (Cav-1), the major protein component of 280 281 caveolae, has been implicated as a scaffold for the organization of several cytoplasmic signal 282 complexes at the plasma membrane [54, 55]. In lung epithelial cells (A549), dexamethasone treatment leads to a rapid (within 2 min) phosphorylation of Cav-1 and protein kinase B (PKB)/Akt 283 in a Src-dependent fashion [56]. Subcellular fractionation revealed co-localization of GR and Src 284 285 to caveolin-containing membrane fractions [56]. Interfering with caveolae/caveolin (by disruption of lipid raft formation, impairment of function using dominant negative caveolin, down regulation 286 of Cav-1 using shRNA, or genetic ablation of Cav-1) prevented acute (within 2 min) GC-induced 287 PKB phosphorylation. Of note, caveolin down-regulation had little effect on GC-mediated 288 289 transactivation, supporting the existence of a putative mGR. Further functional studies in caveolin knockout cells revealed considerable inhibition of GC-mediated cell growth arrest, suggesting that 290 291 membrane-proximal signals acutely initiated by GC are required to mediate delayed effects (anti-292 proliferative effects) previously ascribed exclusively to the nuclear actions of GR [56]. Further 293 evidence supporting a role for caveolae in mGR function stems from studies of membrane nuclear 294 receptors such as estrogen receptor (ER) [57] showing requirement of Cav-1 in mediating acute cellular actions. Indeed, using epitope proximity ligation assays, Watson and colleagues 295 demonstrated interactions of ER $\alpha$  with Cav-1. Interestingly, the use of nystatin, which binds to 296 297 cholesterol and disrupts caveolar structures, blocked estrogen-induced rapid (5 min) ERK activation in pituitary tumor cells [57]. Together these findings indicate a critical role of Cav-1 in 298 299 acute nuclear receptor/steroid signaling.

While the expression of mGR has been demonstrated in a myriad of cell types [58], the colocalization and cross-talk between mGR and Cav-1 is variable and highly cell-specific. Indeed, in U2-OS and MCF-7 cells, double recognition proximity ligation assays demonstrated the physical association of Cav-1 with the mGR [58]. However, studies in human CD14<sup>+</sup> monocytes 304 showed that mGR and Cav-1 are not co-localized and overexpression of the recombinant Cav-1 transcript in human K562 chronic myelogenous leukemia cells did not affect mGR 305 306 expression/appearance suggesting that in these specific cell lines Cav-1 is not the limiting factor for mGR expression/appearance, without ruling out the possibility that it is a component of the 307 transport machinery of GR from the cytosol to the membrane [59]. Palmitoylation, a critical post-308 translational modification occurring through the addition of fatty acid (e.g. palmitic acid) on amino 309 acid residues of membrane proteins, plays a major role in the subcellular trafficking of proteins 310 between membrane compartments [60]. Interestingly, the involvement of palmitoylation in the 311 recruitment of other nuclear receptors, such as ER, to the plasma membrane has been reported 312 [61]. Recent studies investigated whether this process is necessary for the recruitment of GR to the 313 membrane and its co-localization with Cav-1 in COS-7 cells. Treatment of cells with the 314 palmitoylation inhibitor, 2-bromopalmitate, had little effect on membrane localization of GR and 315 its co-localization with Cav-1, and little influence on the acute effects of GC on MAPK signaling 316 pathways. In addition, human GRa did not undergo S-palmitoylation, rendering this process 317 unlikely to modulate membrane recruitment of GR [62]. Future studies on the mechanisms 318 319 underlying GR recruitment to caveolae rich parts and its potential association with Cav-1 are warranted, specifically in airway cells. 320

Several studies have reported an interaction of mGR with other membrane receptors, 321 particularly GPCRs [63]. Zhang and colleagues demonstrated the involvement of mGR and GPCR-322 323 dependent mechanisms in the rapid effect (as early as 1 min) of corticosterone on NMDA-evoked currents in hippocampal neurons [63] and further suggested that mGR may couple to multiple G 324 proteins, including  $G_s$  and  $G_{a/11}$ . Other studies indicate that mGR directly elicits the activation of 325 downstream intracellular signaling pathways. For instance, corticosterone might act via mGR to 326 327 rapidly elicit PKC-dependent activation of ERK1/2 MAPK pathway (with 15 min) in PC12 cells 328 [39]. Interestingly, proteomic analysis of the lymphoma cell line CCRF-CEM identified 128 proteins that were differentially regulated by the specific activation of mGR using BSA-conjugated 329 cortisol for a short-term period (5 and 15 min) [58]. These actions were unique to mGR, as no 330 activation of cGR target genes, such as GILZ, were observed. The majority of networks rapidly 331 332 activated by mGR were mainly involved in cellular growth and cancer (after 5 min treatment with 333 cortisol-BSA), cellular development, or hematological system development and function (after 15 334 min treatment with cortisol-BSA). Ingenuity pathway analysis provided strong evidence that mGR

is involved in pro-apoptotic, immune-modulatory, and metabolic pathways that are also regulated
by GCs through cGR, suggesting that acute mGR stimulation can trigger rapid early priming
events, ultimately paving the way for the slower genomic activities by GCs [58].

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#### Concluding Remarks and Future Perspectives.

Although we have some insight in how GCs regulate different signaling pathways in a non-340 genomic fashion, future in depth investigations are warranted to further unravel details of these 341 complex interactions. Indeed, key questions (see Outstanding Questions) still need careful 342 consideration and additional research must address several important issues: i) the differential 343 nature of non-genomic effects of GC in immune cells versus non-immune/structural cells; ii) 344 differences between non-genomic effects of various steroids based on their lipophilicity [10]; iii) 345 the fact that not all non-genomic effects are GR-mediated (RU486 insensitive) and may be due to 346 non-specific interactions of GC with the cell membrane [2]; iv) the possibility that non-genomic 347 and genomic effects are interconnected, where the acute non-genomic effects pave the way for the 348 slower genomic activities of GCs [58]; and v) the significant role of Cav-1, and possibly other 349 350 scaffolding/anchoring proteins, as a modulator of mGR activation, where the relative numbers of mGR associated with Cav-1 are critical in mediating non-genomic effects of GC [58, 64, 65]. Since 351 352 side effects associated with GC therapy are often generated through its genomic actions [66], 353 uncovering the non-genomic actions of GC with beneficial effects will likely lead to the 354 development of compounds that selectively activate non-genomic signaling and thus have improved therapeutic profiles. 355

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507 Text boxes.

508

#### 509 Box 1: Calcium regulation, ASM tone, and asthma pathogenesis.

Because airway smooth muscle (ASM) serves as the pivotal tissue regulating the bronchomotor 510 511 tone, changes in the pathways regulating ASM contractile properties may play an important role in the development of abnormal lung function in asthma. Abnormal G-protein-coupled receptor 512 (GPCR)-associated calcium homeostasis and ASM shortening may represent one of such 513 mechanisms. Changes could occurs that at different levels of the contraction cascade including i) 514 515 [Ca<sup>2+</sup>]i release from internal stores, ii) myosin light-chain kinase (MLCK) activity, iii) myosin light chain phosphorylation (pMLC), and iv) actin-myosin crossbridge cycling leading to cell 516 shortening. Changes in ASM shortening could also be due to changes in sensitivity of the 517 contractile apparatus to [Ca<sup>2+]</sup>i initiated by the small GTPase, RhoA, which activates Rho kinase 518 519 (ROCK) to inactivate myosin light chain phosphatase (MLCP). Decreased MLCP activity results in an increase in pMLC levels for a given level of  $[Ca^{2+}]i$ , and thus enhancing ASM contractility. 520 It is important to note that there are other parallel pathways where actin polymerization also 521 mediates agonist-induced ASM shortening independently from  $Ca^{2+}$  and pMLC but potentially 522 523 through the phosphorylation of other proteins such as vinculin. Collectively, this evidence suggests 524 that ASM contractile function can mediate airway hyperresponsiveness in chronic airway inflammatory diseases by involving, at least partially, changes in  $Ca^{2+}$ -regulatory pathways. 525

526

#### 527 Box 2. Role of NOS/NO signaling in asthma pathogenesis.

528 Altered NO production has been implicated in the development of both acute and chronic allergen-529 induced AHR. Production of NO occurs through the action of nitric oxide synthase (NOS), of which 3 isoforms have been identified thus far: two constitutive (c)NOS isoforms referred as 530 531 neuronal (n), endothelial (e) NOS, and one inducible isoform called (i) NOS. Upon activation, 532 cNOS isoforms produce relatively low amounts of NO, whereas iNOS can produce high and 533 potentially damaging levels of NO. Whereas NO generated by eNOS is associated with beneficial bronchodilatory effects in allergic asthma, iNOS-derived NO is generally considered detrimental, 534 as it has been linked to for instance epithelial damage, inflammatory cell infiltration, and mucus 535 hypersecretion. These detrimental effects are largely due to the accumulation of Reactive Nitrogen 536

537 Species (RNS), including peroxynitrite, which are reaction products of NO and superoxide anions.
538 Since NOS/NO signaling and RNS play key roles in chronic airway inflammatory diseases,
539 including asthma and COPD, an acute role for GC/GR signaling and (inducible and/or endothelial)
540 NOS activity can be envisioned. In depth studies are warranted to determine whether such
541 functional interaction exists and whether or not targeting it would provide any therapeutic benefit
542 for asthma.

546 Tables.

| GC effects                                    | Acute (simultaneous or within 30 min) | Chronic<br>(delayed) |
|---|---------------------------------------|----------------------|
| Genomic effects                               | -                                     | +                    |
| Inhibitory effects of CHX or<br>Actinomycin D | _                                     | +                    |
| GR involvement                                | - or +                                | +                    |
| Inhibitory effects of RU486                   | - or +                                | +                    |
| Type of GR involved                           | None, membrane GR or cytosolic GR     | Cytosolic GR         |
| GR-independent mechanisms                     | GC interaction with membrane          | None                 |

549 non-genomic effects of glucocorticoids.

**Table 1**: Various criteria (either alone or in combination) used to distinguish genomic effects from

|   | Cell types                                  | GCs                     | References |
|---|---|-------------------------|------------|
| • | Human bronchial epithelial cells            | Dexamethasone           | 3, 4, 10   |
|   |   | Triamcinolone           |            |
|   |   | Hydrocorticone          |            |
| • | Rat thymocytes                              | Methylprednisolone      | 5          |
| • | Mouse neuroblastoma cells                   | Corticosterone          | 6, 14      |
| • | Cichlid fish pituitary cells                | Cortisol                | 7          |
| • | Mouse cortical collecting duct cells        | Dexamethasone           | 11         |
|   |   | Aldosterone             |            |
| • | Rat vascular smooth muscle cells            | Aldosterone             | 12, 13, 25 |
|   |   | Cortisol                |            |
|   |   | Dexamethasone           |            |
| • | Rat B103 neuroblastoma cells                | Hydrocorticosone        | 15         |
| • | Guinea-pig cochlear spiral ganglion neurons | Dexamethasone           | 16         |
| • | Rat hippocampal neurons                     | Corticosterone          | 19         |
|   |   | Dexamethasone           |            |
|   |   | BSA-conjugated cortisol |            |
| • | Mouse skeletal C2C12 cells                  | Dexamethasone           | 20         |
| • | Guinea-pig tracheal tissues                 | Budesonide              | 22         |
| • | Murine airway smooth muscle cells           | Dexamethasone           | 23         |
| • | Guinea-pig mouse model of allergic asthma   | Budesonide              | 24         |
| • | Human vascular endothelial cells            | Dexamethasone           | 30         |

Table 2: Examples of the various cells types where GCs were reported to have non-genomic
effects due their rapid onset, insensitivity to GR blockade (RU486), and protein synthesis
inhibition (cycloheximide).

|   | Signaling pathways                            | Cell types  | GCs                          | References |
|---|---|---|------------------------------|------------|
| • | PKA<br>SERCA Ca2+-ATPases<br>Adenylyl cyclase | Human bronchial epithelial<br>cells   | Dexamethasone                | 10         |
| • | РКС   | Mouse cortical collecting duct cells  | Dexamethasone<br>Aldosterone | 11         |
| • | IP3 accumulation<br>PKC                       | Rat vascular smooth muscle<br>cells   | Dexamethasone<br>Aldosterone | 12         |
| • | РКА   | HT4 neuroblastoma cells   | Corticosterone               | 6          |
| • | РКС   | Rat B103 neuroblastoma cells  | Corticosterone               | 15         |
| • | СаМКІІ<br>АМРК                                | Mouse skeletal myotubes   | Dexamethasone                | 20         |
| • | РКС   | Tracheal smooth muscle tissues  | Cortisol                     | 21         |
| • | Rho kinase                                    | Rat vascular smooth muscle cells  | Dexamethasone                | 25         |
| • | ROS/RNS (NO<br>synthase)                      | Human breast cancer cells   | Cortisol                     | 26         |
| • | NO pathways                                   | Guinea-pig cochlear spiral<br>ganglion neurons<br>Human vascular endothelial<br>cells<br>Human umbilical endothelial<br>cells | Dexamethasone                | 16, 30, 33 |
| • | ERK1/2, P38MAPK,<br>JNK                       | PC12 cells<br>Rat vascular smooth muscle<br>cells   | Dexamethasone                | 25, 37     |

| • | Src tyrosine kinase | Human breast                        | Cortisol      | 26, 40 |
|---|---------------------|-------------------------------------|---------------|--------|
|   |                     | cancer cells A549                   | Dexamethasone |        |
|   |                     | cells                               |               |        |
| • | PI3K/Akt            | Human vascular endothelial<br>cells | Dexamethasone | 30, 33 |

**Table 3**: Examples of signaling pathways activated by GCs via nongenomic mechanisms that
were acute, sensitive (or insensitive) to GR blockade (RU486), and insensitive to protein
synthesis inhibition (cycloheximide).

561 Figure legends.

562

Figure 1. Acute non-genomic effects of GCs on basal and agonist-induced Ca<sup>2+</sup> responses. 563 GCs have been described to differentially affect basal intracellular  $Ca^{2+}$  ([ $Ca^{2+}$ ]i) homeostasis. 564 Depending on the cell type studied and GC applied, GCs can either reduce or augment basal 565  $[Ca^{2+}]i$ . (A) GCs may decrease  $[Ca^{2+}]i$  by activating AC/PKA mediated mechanisms, likely 566 through events taking place at the cell membrane level and independent of GR stimulation, 567 ultimately leading to SERCA activation (thapsigargin-sensitive Ca<sup>2+</sup>-ATPase). (B) Conversely, 568 GCs can activate PLC/IP3 and PKC dependent signaling cascades resulting in enhanced basal 569 [Ca<sup>2+</sup>]i; the involvement of GR in this process is currently unknown. (C) Agonist-induced 570 increases in [Ca<sup>2+</sup>]i can be counteracted by GC-mediated activation of AC/PKA-induced 571 stimulation of SERCA pumps as described in ATP stimulated cells. In contrast, a functional 572 role for PKC was determined in the effects of GC on serotonin-induced Ca<sup>2+</sup> responses, 573 suggesting that the acute inhibitory mechanisms of GCs are highly agonist specific. (D) Limited 574 studies are available on acute potentiating effects by GCs on agonist-induced Ca<sup>2+</sup> responses: 575 in neuronal cells it was suggested that these effects are mediated via the rapid activation of 576 Ca<sup>2+</sup>-influx through ionotropic ATP-gated purinergic 2X receptors. Whether glucocorticoid-577 mediated membrane receptors are involved in this pathway remains to be further investigated 578 (mGR?). These responses rely on the presence of external  $Ca^{2+}$ . Abbreviations: AC, adenylyl 579 cyclase; AR, agonist receptor; IP3, inositol 1, 4, 5-triphosphate; GC, glucocorticoid; mGR, 580 581 membrane glucocorticoid receptor; PKA, protein kinase A; PKC, protein kinase C; SERCA, sarco/endoplasmatic reticulum Ca2+ -ATPase. 582

584 Glossary.

585

586 Muscle reactivity: The ability of the muscle to respond to contractile agonists. It is impaired
587 during pathophysiological conditions such as asthma.

588 **Calcium mobilization:** Intracellular process triggered by external stimuli (e.g. contractile 589 agonists) where calcium is released to be engaged in different cellular functions such as increased 590 muscle reactivity and contraction. Calcium is usually acquired from extra-cellular sources 591 (calcium influx) or intracellular stores (e.g. endoplasmic reticulum).

592 **Genomic action:** Action that modulates the expression of genes. It involves transcriptional 593 processes where an activated transcriptional factor translocates to the nucleus and bind gene 594 promoters to modulate their expression. Such processes require certain time and are delayed.

**Glucocorticoid receptor (GR):** A nuclear receptor, which acts as a receptor and a transcriptional factor. It is primary located in the cytosol. Glucocorticoids, through their lipophilicity, diffuse across the cell membrane to bind GR in the cytosol. Such binding promotes the translocation of GR to the nuclear where it binds gene promoters to modulate their expression. As described in this article, several evidence demonstrate a membrane version of GR, not acting as a transcriptional factor, but rather as a membrane receptor modulating the acute non-genomic effects of GC.

Non-genomic Action: Action that does not modulate the expression of genes. It does not involve
 transcriptional processes or protein synthesis. Such action promotes rapid effects on events
 proximal to the cell membrane to activate certain signal transduction pathways.

Side effects of GC: Due to their wide range of actions that include effects on the immune system, metabolism, skeletal muscle, bone and eyes, to name just few, GC exert in addition to its intended effect, some harmful effects especially when used in high dose and in long-term like in asthma patients. Such effects usually require the genomic actions of GR.

608