

Chapman University
Chapman University Digital Commons

Pharmacy Faculty Articles and Research

School of Pharmacy

7-1-2013

Tigecycline Induction of Phenol-Soluble Modulins by Invasive Methicillin-Resistant *Staphylococcus aureus* Strains

Jason Yamaki

Chapman University, yamaki@chapman.edu


Timothy Synold

City of Hope Medical Center

Annie Wong-Beringer

University of Southern California

Follow this and additional works at: http://digitalcommons.chapman.edu/pharmacy_articles

 Part of the [Amino Acids, Peptides, and Proteins Commons](#), [Bacteria Commons](#), and the [Other Pharmacy and Pharmaceutical Sciences Commons](#)

Recommended Citation

Yamaki J, Synold T, Wong-Beringer A. Tigecycline Induction of Phenol-Soluble Modulins by Invasive Methicillin-Resistant *Staphylococcus aureus* Strains. *Antimicrobial Agents and Chemotherapy*. 2013;57(9):4562-4565. doi:10.1128/AAC.00470-13.

This Article is brought to you for free and open access by the School of Pharmacy at Chapman University Digital Commons. It has been accepted for inclusion in Pharmacy Faculty Articles and Research by an authorized administrator of Chapman University Digital Commons. For more information, please contact laughtin@chapman.edu.

Tigecycline Induction of Phenol-Soluble Modulins by Invasive Methicillin-Resistant *Staphylococcus aureus* Strains

Comments

This article was originally published in *Antimicrobial Agents and Chemotherapy*, volume 57, issue 9, in 2013.
DOI: [10.1128/AAC.00470-13](https://doi.org/10.1128/AAC.00470-13)

Copyright

American Society for Microbiology

Tigecycline Induction of Phenol-Soluble Modulins by Invasive Methicillin-Resistant *Staphylococcus aureus* Strains

Jason Yamaki,^a Timothy Synold,^b Annie Wong-Beringer^{a,c}

University of Southern California, Los Angeles, California, USA^a; City of Hope Medical Center, Duarte, California, USA^b; Huntington Hospital, Pasadena, California, USA^c

We examined the effects of tigecycline on three types of exoproteins, α -type phenol-soluble modulins (PSM α_1 to PSM α_4), α -hemolysin, and protein A, in 13 methicillin-resistant *Staphylococcus aureus* isolates compared to those of clindamycin and linezolid. Paradoxical increases in PSM α s occurred in 77% of the isolates with tigecycline at 1/4 and 1/8 MICs and clindamycin at 1/8 MIC compared to only 23% of the isolates with linezolid at 1/8 MIC. Induction was specific to PSM α_1 to PSM α_4 , as protein A and α -hemolysin production was decreased under the same conditions by all of the antibiotics used.

Methicillin-resistant *Staphylococcus aureus* (MRSA) virulence in pneumonia and bacteremia has been attributed to exoproteins, specifically, α -hemolysin (Hla) and α -type phenol-soluble modulins (PSM α_1 to PSM α_4), which are produced by nearly all *S. aureus* strains and in excess by community-acquired MRSA strains (1). These exoproteins not only cause direct damage to target host cells but also exacerbate the host inflammatory response, contributing to acute lung injury. Additionally, recent reports have demonstrated the importance of protein A (Spa) in invasive diseases such as pneumonia (2, 3).

In light of the impressive arsenal of virulence factors contributing to the success of MRSA as a pathogen, it is of keen interest to determine if anti-MRSA agents belonging to the antibiotic class of protein synthesis inhibitors provide the added antivirulence benefit of exoprotein inhibition. In the present study, we investigated the antivirulence potential of tigecycline, linezolid, and clindamycin which have been proven efficacious in the treatment of MRSA infections. Our goal was to determine whether antivirulence effects can be generalized across different clinical isolates, different agents that inhibit protein synthesis, and different exoproteins. Specifically, we tested the effects of the above three antibiotics at subinhibitory concentrations on formylated PSM α_1 to PSM α_4 , Hla, and Spa production by invasive MRSA isolates.

Eleven invasive MRSA isolates were tested under the following conditions. A modified CLSI broth macrodilution assay was used to determine MICs after 24 h of incubation at 37°C and shaking at 250 rpm in tryptic soy broth. Supernatants were then analyzed by liquid chromatography-tandem mass spectrometry, and Hla and Spa were analyzed by Western blotting as previously described (4, 5). Measured exotoxin values were normalized to the cell optical density at 600 nm (OD₆₀₀) at the time the supernatant was harvested. PSM α concentrations under various test conditions were compared by analysis of variance with Dunn's correction using GraphPad Prism version 5.0 software (GraphPad, San Diego, CA).

Table 1 depicts the SCC_{mec} type, PVL status, and baseline PSM α production characteristics of the 13 isolates studied (11 clinical isolates and two control strains). The PSM α_1 to PSM α_4 peptides have been shown to cause concentration-dependent neutrophil lysis (6, 7). A PSM α_3 concentration of 5 μ g/ml has been shown to lyse nearly 50% of human polymorphonuclear neutrophils (PMNs), while 10 μ g/ml of PSM α_1 and PSM α_4 can cause 60% and 10% PMN lysis, respectively (8, 9). Thus, isolates were grouped on the basis of the amount of the most potent peptide,

TABLE 1 Characteristics of the 11 clinical isolates and two control strains used in this study

Isolate or strain	<i>mec</i>		Infection type	PSM ^b baseline	MIC (μ g/ml) ^c		
	type	PVL ^a			TYG	CL	LZ
1	IV	+	Blood	Low	0.125	0.188	3
2	IV	+	Blood	Medium	0.125	0.25	3
3	IV	+	Blood	Low	0.125	0.188	2
4	IV	+	Blood	Medium	0.125	0.25	2
5	IV	+	Blood	Medium	0.125	0.188	2
6	IV	+	Pneumonia	Low	0.188	0.125	2
7	IV	+	Pneumonia	Medium	0.125	0.188	3
8	IV	+	Necrotizing pneumonia	High	0.125	0.25	3
9	II	–	Pneumonia	Very low	0.125	>256	3
10	II	–	Pneumonia	Very low	0.125	>256	1
11	II	–	Pneumonia	Very low	0.125	0.188	2
Control strains							
USA300 (LAC)	IV	+	Wound	Medium	0.25	0.25	2
USA600	II	–	Blood	Low	0.125	>256	2

^a A plus or minus sign denotes the presence or absence, respectively, of the *lukE/S* gene, which encodes the Panton-Valentine leukocidin.

^b The PSM α_3 production level at the baseline was arbitrarily defined as very low (<1 μ g/ml), low (1 to 5 μ g/ml), medium (6 to 15 μ g/ml), or high (>15 μ g/ml).

^c TYG, tigecycline; CL, clindamycin; LZ, linezolid. Resistance is defined as a MIC of >256 μ g/ml (not tested for PSM α production at subinhibitory concentration).

PSM α_3 , produced at the baseline as very low to low (\leq 5 μ g/ml), medium (6 to 15 μ g/ml), or high (>15 μ g/ml) producers.

Bacterial growth at 1/2 MICs of all three antibiotics was altered in half of the isolates tested by as much as 50% of the final OD compared to a no-antibiotic control, while no significant difference in the growth of any isolates was observed at 1/4 and 1/8 MICs. Thus, subsequent discussions will focus on the effect of antibiotics at the latter subinhibitory concentrations that did not affect growth. Measured PSM α values were normalized to the OD₆₀₀ at the time the supernatant was harvested.

Received 6 March 2013 Returned for modification 23 April 2013

Accepted 22 June 2013

Published ahead of print 1 July 2013

Address correspondence to Annie Wong-Beringer, anniew@usc.edu.

Copyright © 2013, American Society for Microbiology. All Rights Reserved.

doi:10.1128/AAC.00470-13

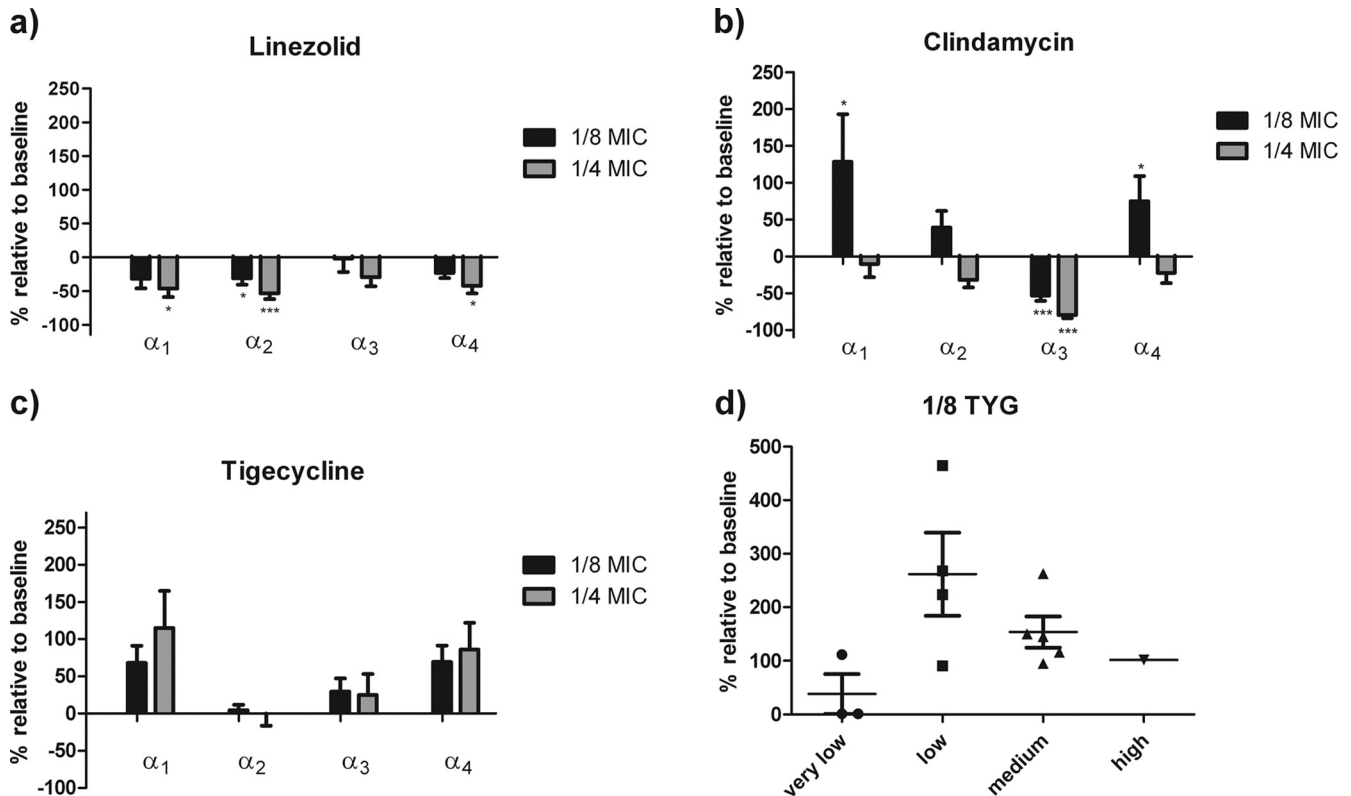


FIG 1 Overall effects of linezolid, clindamycin, and tigecycline on PSM α_1 to PSM α_4 production. (a to c) Effects of antibiotics on the production of PSM α_1 to PSM α_4 (α_1 , α_2 , α_3 , and α_4 , respectively). All measured PSM values ($\mu\text{g/ml}$) were normalized to OD₆₀₀ and then calculated as percentages relative to the baseline. Isolates ($n = 10$) included only those that produced measurable PSM α peptides at the baseline (***, $P < 0.0001$; **, $P < 0.001$; *, $P < 0.05$). (d) Strain-to-strain variability of PSM α_1 production at 1/8 MIC of tigecycline (TYG). Strains were grouped according to their baseline PSM α_1 production as very low ($<1 \mu\text{g/ml}$), low (1 to 5 $\mu\text{g/ml}$), medium (6 to 15 $\mu\text{g/ml}$), and high ($>15 \mu\text{g/ml}$) producers.

Tigecycline appears to have the least inhibitory potential overall, while linezolid has the greatest (Fig. 1a to c). Increased production of all four PSM α peptides was the primary response observed in the presence of tigecycline at 1/4 and 1/8 MICs, while with clindamycin this occurred only at 1/8 MIC (Fig. 1b and c). Notably, compared to clindamycin at 1/8 MIC, which significantly induced PSM production in seven isolates, linezolid at 1/8 MIC modestly induced PSM production in only three isolates (Fig. 1a).

Strain-specific responses to the presence of subinhibitory antibiotic concentrations in PSM α production were observed. Drug concentrations that induced PSM α production in some isolates did not do so in others, independently of the baseline production level. Specifically, with tigecycline at both 1/4 and 1/8 MICs (Fig. 1d), PSM α_1 was induced at least 1.5 times above the baseline in 77% (11/13) of the isolates, while induction did not occur in two isolates at any of the concentrations tested. Similar results were observed with clindamycin, where 54% (7/13) of the isolates were induced by greater than 150%; however, this was observed only at 1/8 MIC. In those isolates, PSM α_1 production was induced to greater than 10 $\mu\text{g/ml}$, which has been shown to cause significant PMN lysis (9). Linezolid at the same concentration resulted in increases in PSM α_1 in only 23% (3/13) of the isolates tested, was inhibitory in 5 isolates, and had no significant effects on the remaining isolates. Of additional interest is the observation that nonproducers at the baseline produced PSM α peptides in the presence of subinhibitory concentrations of both linezolid and

tigecycline. However, it is noteworthy that PSM α production that was induced in those two isolates did not exceed 4 $\mu\text{g/ml}$ and thus would not be expected to have a significant impact on host PMNs. In contrast to results observed with the PSM α_1 to PSM α_4 peptides, we found that the production of both Hla and Spa did not increase under any condition and was inhibited in a dose-dependent manner at the concentrations of all three antibiotics tested regardless of whether PSM α peptides were induced or suppressed in those isolates (Fig. 2).

To our knowledge, we are the first to investigate the effect of tigecycline on MRSA PSM α_1 to PSM α_4 peptide production. We also included clindamycin and linezolid for comparison and tested their antivirulence potential against two other key exotoxins, Hla and Spa. We found that at sub-MICs, agents in this class of antibiotics have pleiotropic effects on toxin production that are dependent on the drug, strain, and toxin tested.

All three of the antibiotics tested are known to have large volumes of distribution and would be expected to be present at high concentrations within tissues. However, the concentrations tested reflect clinical scenarios where sub-MICs might be achieved at sites of infection because of inadequate dosing, altered pharmacokinetic parameters of the patient, and differential drug distribution to various body sites. Specifically, compared to the concentrations achieved at other body sites, tigecycline achieves relatively low levels in serum (0.11 to 0.19 $\mu\text{g/ml}$) and lung epithelial lining fluid (0.11 to 0.31 $\mu\text{g/ml}$), which are relevant to bacteremia and

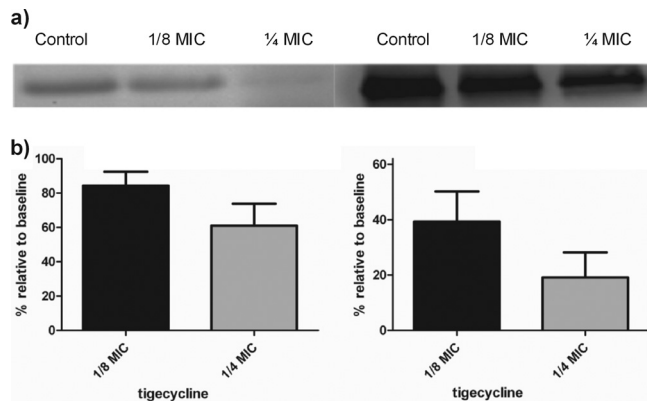


FIG 2 Effects of tigecycline on Hla and Spa production. (a) Representative images of Western blot assays of Hla and Spa, respectively. (b and c) Production of Hla and Spa in the presence of 1/8 and 1/4 MICs of tigecycline relative to the baseline. In no case was Hla or Spa production increased by subinhibitory concentrations of any of the antibiotics tested.

pneumonia, respectively, which are below the 0.5- μ g/ml FDA MIC breakpoint of *S. aureus* susceptibility (10–12).

Low concentrations (1/4 and 1/8 MICs) of the antibiotics tested were more likely to induce PSM α production overall, with tigecycline having the greatest induction potential, clindamycin having less, and linezolid even less than that. Furthermore, the magnitudes of induction and inhibition differed among isolates, where the same drug could induce PSM α production in one isolate while inhibiting production in another. The observation of PSM α induction by protein synthesis-inhibiting antibiotics (e.g., linezolid and clindamycin) at sub-MICs is contrary to published literature on other exotoxins, where studies have consistently found toxin suppression under this class of antibiotics (13–16). The production of both Hla and Spa was suppressed by antibiotics tested at subinhibitory concentrations among the study isolates, independently of whether PSM α was induced in the same isolates. Hence, our results indicate that the induction effects may be specific for PSM α peptides. In contrast to our findings where significant inhibition was observed, a recent article showed tigecycline to be minimally inhibitory of Hla and Spa production (17); however, this difference may be due to the different strains and testing methods used.

Our study has several limitations. First, while we accounted for changes in cell counts by normalizing the amount of toxin measured to CFU counts, other factors, such as initial slowing of growth, could also affect toxin production. In addition, our observations are strictly *in vitro* and may not reflect how MRSA strains express virulence in the host environment, where the effects of toxin induction may be mitigated by host defenses that are present.

Further investigation is needed to examine if any other major toxins are induced, as PSM α peptides did when exposed to subinhibitory concentrations of different antibiotics. Elucidating the mechanism that underlies the induction of PSM α peptides is of great interest and has implications for the future development of new antibiotics. In light of the recent evidence implicating tigecycline in excess deaths and treatment failure in severe infections (18), the enhanced toxin production observed in this study may have clinical relevance and deserves further study. For now, our data caution against the general assumption by clinicians that all

protein synthesis inhibitors possess inhibitory potential against all of the exotoxins produced by *S. aureus* and affirm the importance of adequate dosing.

ACKNOWLEDGMENTS

This study was supported by a research grant from Pfizer.

We thank Bixin Xi for his technical assistance in the development and optimization of the mass spectrometry assay for PSM α measurement. We also thank the Network on Antimicrobial Resistance in *Staphylococcus aureus* (NARSA) and The Los Angeles County Public Health Department for providing the control strains used.

REFERENCES

- Li M, Diep BA, Villaruz AE, Braughton KR, Jiang X, DeLeo FR, Chambers HF, Lu Y, Otto M. 2009. Evolution of virulence in epidemic community-associated methicillin-resistant *Staphylococcus aureus*. *Proc. Natl. Acad. Sci. U. S. A.* 106:5883–5888.
- Soong G, Martin FJ, Chun JR, Cohen TS, Ahn DS, Prince A. 2011. *Staphylococcus aureus* protein A mediates invasion across airway epithelial cells through activation of RhoA GTPase signaling and proteolytic activity. *J. Biol. Chem.* 286:35891–35898.
- Martin FJ, Gomez MI, Wetzel DM, Memmi G, O'Seaghda M, Soong G, Schindler C, Prince A. 2009. *Staphylococcus aureus* activates type I IFN signaling in mice and humans through the Xr repeated sequences of protein A. *J. Clin. Invest.* 119:1931–1939.
- Yamaki J, Synold T, Wong-Beringer A. 2011. Antivirulence potential of TR-700 and clindamycin on clinical isolates of *Staphylococcus aureus* producing phenol-soluble modulins. *Antimicrob. Agents Chemother.* 55:4432–4435.
- Montgomery CP, Boyle-Vavra S, Daum RS. 2009. The arginine catabolic mobile element is not associated with enhanced virulence in experimental invasive disease caused by the community-associated methicillin-resistant *Staphylococcus aureus* USA300 genetic background. *Infect. Immun.* 77:2650–2656.
- Wang R, Braughton KR, Kretschmer D, Bach THL, Queck SY, Li M, Kennedy AD, Dorward DW, Klebanoff SJ, Peschel A, DeLeo FR, Otto M. 2007. Identification of novel cytolytic peptides as key virulence determinants for community-associated MRSA. *Nat. Med.* 13:1510–1514.
- Loffler B, Hussain M, Grundmeier M, Bruck M, Holzinger D, Varga G, Roth J, Kahl BC, Proctor RA, Peters G. 2010. *Staphylococcus aureus* Panton-Valentine leukocidin is a very potent cytotoxic factor for human neutrophils. *PLoS Pathog.* 6(1):e1000715. doi:10.1371/journal.ppat.1000715.
- Hongo I, Baba T, Oishi K, Morimoto Y, Ito T, Hiramatsu K. 2009. Phenol-soluble modulin alpha 3 enhances the human neutrophil lysis mediated by Panton-Valentine leukocidin. *J. Infect. Dis.* 200:715–723.
- Gonzalez DJ, Okumura CY, Hollands A, Kersten R, Akong-Moore K, Pence MA, Malone CL, Derieux J, Moore BS, Horswill AR, Dixon JE, Dorrestein PC, Nizet V. 2012. Novel phenol-soluble modulin derivatives in community-associated methicillin-resistant *Staphylococcus aureus* identified through imaging mass spectrometry. *J. Biol. Chem.* 287:13889–13898.
- Rodvold KA, Gotfried MH, Cwik M, Korth-Bradley JM, Dukart G, Ellis-Grosse EJ. 2006. Serum, tissue and body fluid concentrations of tigecycline after a single 100 mg dose. *J. Antimicrob. Chemother.* 58:1221–1229.
- Brink AJ, Bizos D, Boffard KD, Feldman C, Grolman DC, Pretorius J, Richards GA, Senekal M, Steyn E, Welkovic N. 2010. Guideline: appropriate use of tigecycline. *S. Afr. Med. J.* 100:388–394.
- Conte JE, Golden JA, Kelly MG, Zurlinden E. 2005. Steady-state serum and intrapulmonary pharmacokinetics and pharmacodynamics of tigecycline. *Int. J. Antimicrob. Agents* 25:523–529.
- Dumitrescu O, Badiou C, Bes M, Reverdy ME, Vandenesch F, Etienne J, Lina G. 2008. Effect of antibiotics, alone and in combination, on Panton-Valentine leukocidin production by a *Staphylococcus aureus* reference strain. *Clin. Microbiol. Infect.* 14:384–388.
- Dumitrescu O, Boisset S, Badiou C, Bes M, Benito Y, Reverdy ME, Vandenesch F, Etienne J, Lina G. 2007. Effect of antibiotics on *Staphylococcus aureus* producing Panton-Valentine leukocidin. *Antimicrob. Agents Chemother.* 51:1515–1519.

15. Stevens DL, Ma Y, Salmi DB, McIndoo E, Wallace RJ, Bryant AE. 2007. Impact of antibiotics on expression of virulence-associated exotoxin genes in methicillin-sensitive and methicillin-resistant *Staphylococcus aureus*. *J. Infect. Dis.* **195**:202–211.
16. Ohlsen K, Ziebuhr W, Koller KP, Hell W, Wichelhaus TA, Hacker J. 1998. Effects of subinhibitory concentrations of antibiotics on alpha-toxin (*hla*) gene expression of methicillin-sensitive and methicillin-resistant *Staphylococcus aureus* isolates. *Antimicrob. Agents Chemother.* **42**:2817–2823.
17. Otto MP, Martin E, Badiou C, Lebrun S, Bes M, Vandenesch F, Etienne J, Lina G, Dumitrescu O. 2013. Effects of subinhibitory concentrations of antibiotics on virulence factor expression by community-acquired methicillin-resistant *Staphylococcus aureus*. *J. Antimicrob. Chemother.* **68**: 1524–1532.
18. Prasad P, Sun J, Danner RL, Natanson C. 2012. Excess deaths associated with tigecycline after approval based on noninferiority trials. *Clin. Infect. Dis.* **54**:1699–1709.