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The Primary Structure of the Hemoglobins of the Adult Jaguar (*Panthera onco*, Carnivora)*

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Summary: The primary structure of the hemoglobins from Jaguar (*Panthera onco*) are presented. Electrophoretic separations without and with a dissociating agent revealed the presence of two hemoglobin components, $\alpha_2\beta_2^I$ and $\alpha_2\beta_2^{II}$. The separation of the hemoglobin components was achieved by ion-exchange chromatography. The globin chains were separated

by ion-exchange chromatography and also by reversed phase HPLC. The amino-acid sequences of the native chains and peptides were determined by liquid-phase and gas-phase sequencing. N-Acetylserine was detected by FAB-mass spectroscopy as N-terminal group of the βI chain. The sequences are compared with that of human hemoglobin (Hb A).

Die Primärstruktur der Hämoglobine des adulten Jaguars (*Panthera onco*, Carnivora)

Zusammenfassung: Die Primärstruktur der Hämoglobine des adulten Jaguar (*Panthera onco*) wird angegeben. Mit elektrophoretischen Methoden wurden zwei Hämoglobinkomponenten, $\alpha_2\beta_2^I$ und $\alpha_2\beta_2^{II}$, nachgewiesen. Die Isolierung der Komponenten und Ketten erfolgte mit Ionenaustauschchromatographie, die der Ketten auch mit „reversed phase“ HPLC. Die Amino-

säuresequenz wurde mit Flüssigphasen- und Gasphasensequenatoren bestimmt. Mit FAB-Massenpektroskopie wurde N-Acetylserin als N-terminale Gruppe der βI -Kette nachgewiesen. Die erarbeiteten Primärstrukturen wurden mit den α - und β -Ketten des menschlichen Hämoglobins (Hb A) verglichen.

Key words: Jaguar, hemoglobin, blocked chain, amino-acid sequence.

A large number of mammalian hemoglobins has been sequenced to date. We have undertaken a study on Jaguar hemoglobins to fill a gap in the existing sequences of Carnivora. Jaguar belongs to the order Carnivora, family Felidae, and genus *Panthera*. 2,3-Bisphosphoglycerate showed only minimal effect on the oxygen affinity of the hemoglobins within this family^[2]. The

presence of a blocked N-terminal chain was also reported for a hemoglobin of different members of the Felidae family^[3].

Materials and Methods

Blood from a Jaguar was collected in heparinised tubes (Zoological Garden, Berlin). Erythrocytes were washed four times with physiological saline and then lysed with

Enzyme:

Trypsin (EC 3.4.21.4).

Abbreviations:

Quadrol, *N,N,N',N'*-Tetrakis(2-hydroxypropyl)ethylenediamine; reagent IV, trisodium 7-(isothiocyanato)-naphthalene-1,3,5-trisulphonate; HPLC, high-performance liquid chromatography; TosPheCH₂Cl = (*N*-tosyl-L-phenylalanyl)chloromethane.

* 114th Communication on hemoglobins; for the 112th and 113th communication see ref. [1a,b].

twice distilled water for 1 h in the cold. Disc electrophoresis was done in 12% polyacrylamide gels in Tris/glycine buffer at pH 8.3^[4]. Triton electrophoresis^[5] was used to check the purity of the isolated chains. The hemoglobin components were separated by chromatography on a column of DEAE-Sephadex^[6].

Globin chains were separated on a column of CM-cellulose in the presence of 8M urea^[7]. In addition, HPLC on columns filled with reversed-phase material was used to separate the globin chains. A column of LiChrosorb-RP2 was equilibrated with 50mM ammonium acetate and 12% formic acid. The globin chains were eluted by applying a gradient with increasing concentrations of acetonitrile (35–60%) within 60 min. Globin chains were oxidized as described in ref.^[8].

Oxidized chains were digested with trypsin (TosPhe-CH₂-Cl-treated) for 3 h at 20 °C and pH 10.5 and 9.5 with an enzyme/substrate ratio of 5:100^[9]. Digested chains were centrifuged at 4000 rpm for 10 min and subjected to chromatography on a column of Sephadex-G25 (2.5 × 150 cm). The peptides were eluted with 0.1M acetic acid. Peaks prefractionated in this way were subjected to HPLC on a column with reversed-phase material^[10]. The peptides were eluted from the column of LiChrosorb-RP2 equilibrated with 50mM ammonium acetate by applying a linear gradient of 0–60% acetonitrile in 60 min.

The amino-acid composition of tryptic peptides was determined by the automatic amino-acid analyser

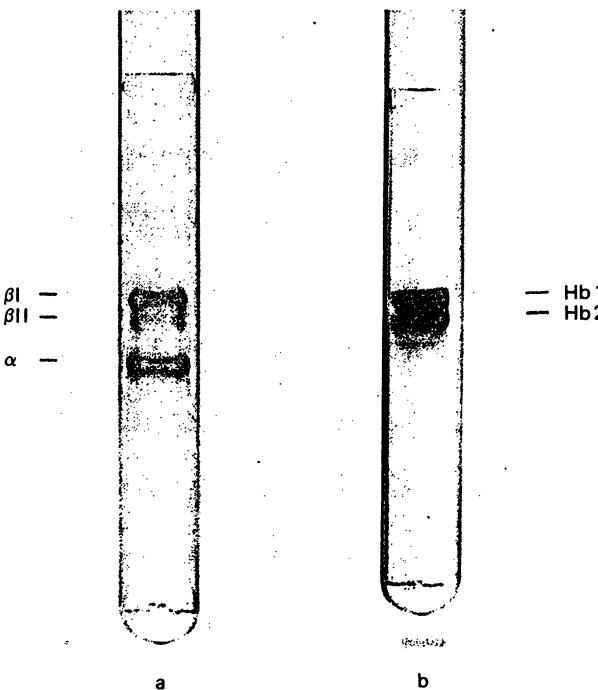


Fig. 1. Electrophoretic pattern of crude hemoglobin of Jaguar on polyacrylamide gel.

a) Under dissociating conditions, in 8M urea and Triton X-100.

b) Disc electrophoresis at pH 8.3.

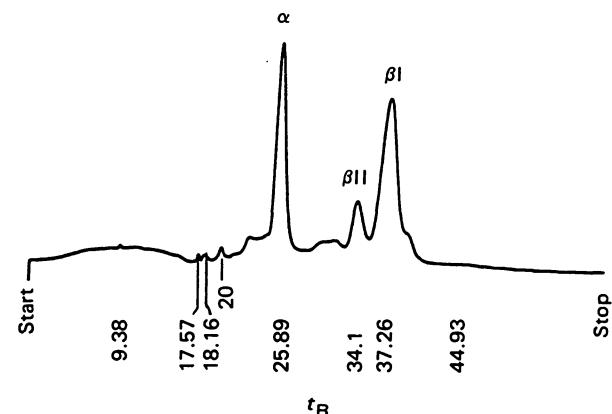


Fig. 2. Elution pattern of the Jaguar globin chains by reversed-phase HPLC on a LiChrosorb RP2 column.

Buffer: 12% formic acid/50mM ammonium acetate; gradient 0–35% acetonitrile in 2 min, followed by 35–60% acetonitrile in 60 min; the flow rate was 1 ml/min.

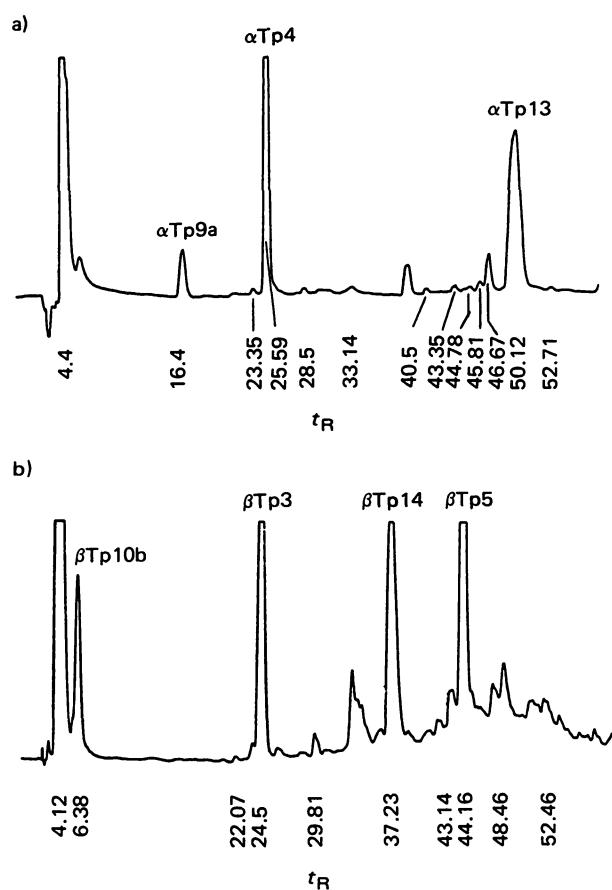


Fig. 3. Rechromatography of the prefractionated peaks of the tryptic peptides from Sephadex-G25 on a LiChrosorb RP2 column.

Buffer: 50mM ammonium acetate; gradient: 0–60% acetonitrile in 60 min; flow rate was 1 ml/min.

a) Peptides from globin α chain.

b) Peptides from globin β chain.

(Model LC 5000, Biotronic). Amino-acid sequences were determined by Edman degradation^[11] in liquid-phase sequencers (Models 890B and 890C, Beckman). A modified quadrol programme^[12] was used for the sequencing of the native chains and lysine peptides. The lysine peptides were sequenced after modification by reagent IV^[13]. A 3-(dimethylamino)propyne pro-

gramme^[14] was employed for arginine peptides. Some peptides were also sequenced by a gas-phase sequencer^[15]. The phenylthiohydantoin derivatives of amino acids were identified by HPLC^[16]. The sequence of a peptide with a blocked N-terminus from the β I chain (Tp 1) was established by FAB mass spectroscopy.

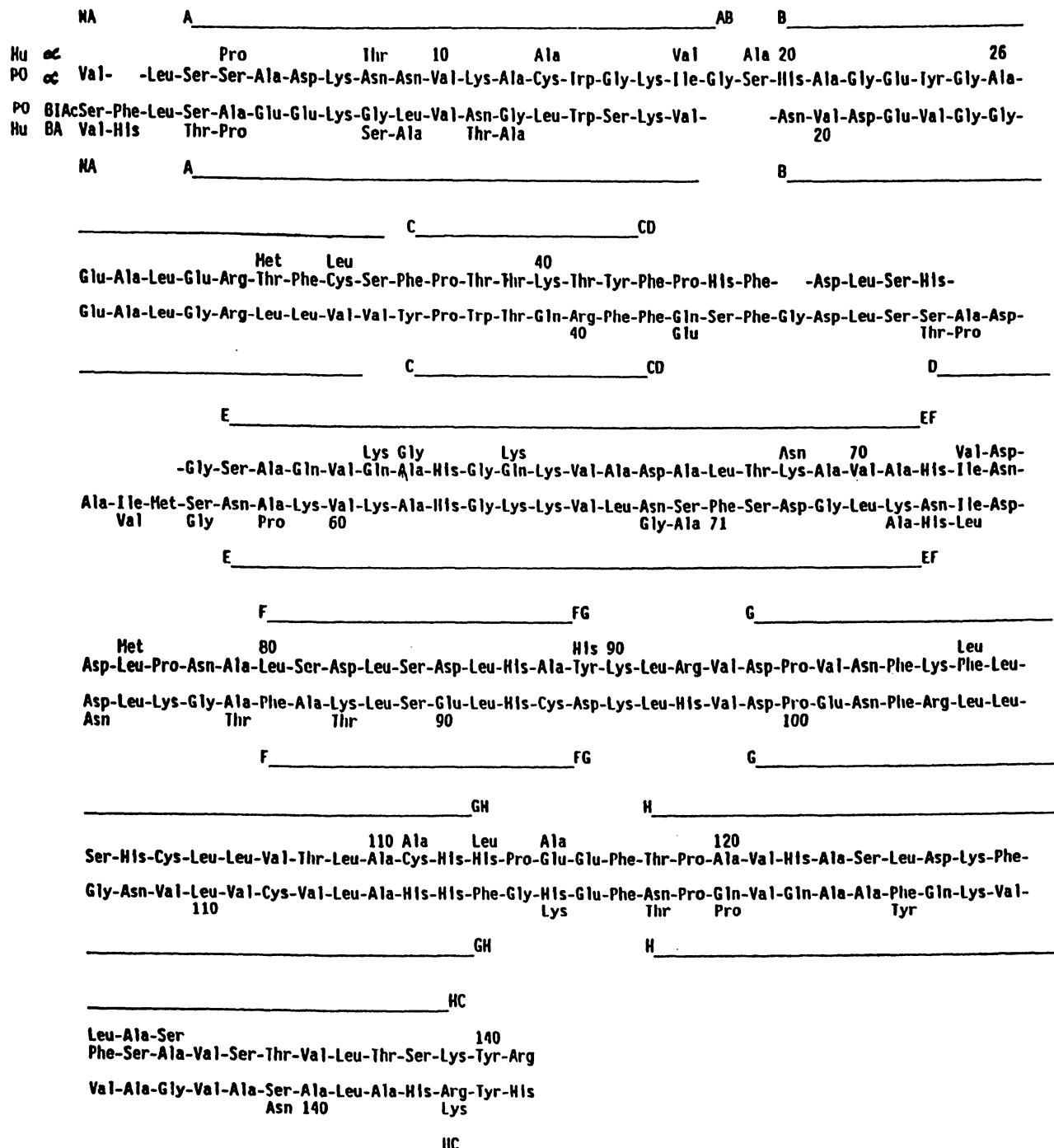


Fig. 4. Amino-acid sequences of Jaguar (PO) globin chains in alignment with the corresponding chains of human (Hu) globin.

In case of human hemoglobin only the exchanges are given. The Hb-II differs from the Hb-I at the following positions β I/ β II: β NA1 Ac-Ser/Gly, β HC1 Arg/Lys.

Results and Discussion

Electrophoresis on disc and triton gel showed two hemoglobin components of similar intensity containing three globin chains, namely α , β I, and β II (Fig. 1). Chromatography on DEAE-Sephacel revealed the presence of two hemoglobin components. Crude globin was separated by chromatography on a CM-Cellulose column to give three peaks. Triton electrophoresis of these peaks showed both, β I and β II, the latter being contaminated with the α chain. Reversed-phase HPLC resulted in the resolution of three purified chains (Fig. 2).

Prefractionation of the tryptic peptides on Sephadex-G25 gave some of the peptides in pure form. Rechromatography of the peptide mixture on reversed-phase HPLC led to pure peptides (Fig. 3), the amino-acid compositions of which are presented as Supplementary Material (Table 2-4). The amino-sequence of blocked β chain as determined by mass spectroscopy showed N-terminal acetylation.

The complete amino-acid sequence of all chains is presented in Fig. 4. The sequence comparison with human globin chains revealed 22 (15.6%) substitutions in the α , 28 (19.1%) in the β I and 27 (18.4%) in the β II chains. Between two the β chains differences were found at (β I/ β II): β NA1 Ac-Ser/Gly, β HCl Arg/Lys. The exchanges which are distributed over the entire length of the molecule result in the alteration of four α 1 β 1 contact points: α 34(B15)Leu/Cys, α 111-(G18)Ala/Cys, β 123(H1)Thr/Asn, β 125(H3)-Pro/Gln; one α 1 β 2 contact point at β 43(CD2) Glu/Gln and one heme contact point at β 70-(E14)Ala/Ser. Exchanges were also observed at binding sites for 2,3-bisphosphoglycerate at β 1(NA1)Val/Ac-Ser, and β 2(NA2)His/Phe. Substitution of the hydrophilic residue β NA2 (His) with a hydrophobic (Phe) should result in the alteration of the secondary structure^[17]. Exchanges at these phosphate-binding sites were also found in cat hemoglobin^[18] and in Leopard hemoglobin^[19]. The minimum numbers of

amino-acid differences found between the known sequences of the α and β I chains of three felidae are presented in Table 1.

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Literature

- 1a Kleinschmidt, T., Koop, B.F. & Braunitzer, G. (1987) *Biol. Chem. Hoppe-Seyler* **368**, 1197-1202.
- 1b Weber, R.E., Kleinschmidt, T. & Braunitzer, G. (1987) *Respir. Physiol.* **69**, 347-357.
- 2 Taketa, F. (1974) *Ann. N.Y. Acad. Sci.* **241**, 524-537.
- 3 Taketa, F., Attermeier, M.H. & Mauk, A.G. (1972) *J. Biol. Chem.* **247**, 33-35.
- 4 Davis, B.J. (1964) *Ann. N.Y. Acad. Sci.* **121**, 404-427.
- 5 Alter, B.P., Goff, S.C., Efremov, G.D., Gravely, M.E. & Huisman, T.H.J. (1980) *Br. J. Haematol.* **44**, 527-534.
- 6 Kleinschmidt, T., Sladic-Simic, D. & Braunitzer, G. (1982) *Blut* **45**, 283-286.
- 7 Clegg, G.B., Naughton, M.A. & Weatherall, D.J. (1966) *J. Mol. Biol.* **19**, 91-108.
- 8 Hirs, C.H.W., Stein, W.H. & Moore, S. (1956) *J. Biol. Chem.* **221**, 151-159.
- 9 Hirs, C.H.W. (1967) *Methods Enzymol.* **11**, 218-220.
- 10 Kratzin, H., Yang, C., Krusche, J.U. & Hilschman, N. (1980) *Hoppe-Seyler's Z. Physiol. Chem.* **361**, 1591-1598.
- 11 Edman, P. & Begg, G. (1967) *Eur. J. Biochem.* **1**, 80-91.
- 12 Braunitzer, G., Schrank, B. & Ruhfus, A. (1970) *Hoppe-Seyler's Z. Physiol. Chem.* **351**, 1589-1590.
- 13 Pfletschinger, J. & Braunitzer, G. (1980) *Hoppe-Seyler's Z. Physiol. Chem.* **361**, 925-931.
- 14 Braunitzer, G., Schrank, B., Stangl, A. & Scheithauer, U. (1978) *Hoppe-Seyler's Z. Physiol. Chem.* **359**, 137-146.
- 15 Begg, G., Rücknagel, P., Godovac-Zimmerman, J. & Braunitzer, G. (1986) *Biol. Chem. Hoppe-Seyler* **367**, 81-82.
- 16 Zimmerman, C.L. & Pisano, J.J. (1977) *Methods Enzymol.* **47**, 45-51.
- 17 Perutz, M.F. & Imai, K. (1980) *J. Mol. Biol.* **136**, 183-191.
- 18 Abbasi, A. & Braunitzer, G. (1985) *Biol. Chem. Hoppe-Seyler* **366**, 699-704.
- 19 Abbasi, A. & Braunitzer, G. (1985) *J. Protein Chem.* **4**, 57-67.

Table 1. The minimum amino-acid differences in α and β I chains of Felidae hemoglobins.

α Chains	Leopard	Cat	β Chains	Leopard	Cat
Jaguar	0	8	Jaguar	3	6
Leopard	-	8	Leopard	-	5

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Supplementary Material

Table 2. Amino-acid composition of peptides from α chain of Jaguar.

Pos.	Tp1 1-7	Tp2 8-11	Tp3 12-16	Tp4 17-31	Tp5 32-40	Tp6/7/8 41-61	Tp9a 62-68	Tp9b 69-90	Tp10 91-92	Tp11 93-99	Tp12 100-127	Tp13 128-139	Tp14 140-141
Asx	1.08	1.91	-	-	1.14	0.92	4.68	-	1.95	1.00	-	-	-
Thr	-	-	-	-	2.79	0.89	0.93	-	-	-	2.05	1.93	-
Ser	1.80	-	-	0.89	1.10	1.80	-	1.85	-	-	1.60	2.92	-
Glx	-	-	-	3.13	-	3.19	-	-	-	-	2.05	-	-
Pro	-	-	-	-	1.23	0.89	-	1.06	-	1.16	2.02	-	-
Gly	-	-	1.02	2.85	-	2.03	-	-	-	-	0.72	-	-
Ala	1.00	-	0.97	3.08	-	2.17	1.75	4.05	-	-	2.77	1.04	-
Cys*	-	-	0.97	-	0.95	-	-	-	-	-	2.30(2)	-	-
Val	0.96	0.98	-	-	0.98	0.91	1.04	-	1.91	1.75	2.05	-	-
Ile	-	-	-	0.96	-	-	-	0.90	-	-	-	-	-
Leu	1.08	-	-	1.04	-	1.10	1.45	4.20	1.02	-	4.00	1.05	-
Tyr	-	-	-	0.73	-	0.83	-	0.83	-	-	-	-	0.76
Phe	-	-	-	-	1.91	1.96	-	-	-	0.98	2.30(2)	1.97	-
His	-	-	-	1.26	-	3.09	-	2.31	-	-	4.11	-	-
Trp	-	-	0.81	-	-	-	-	-	-	-	-	-	-
Lys	1.06	1.10	1.03	-	1.00	0.98	1.02	1.02	-	0.99	0.88	0.98	-
Arg	-	-	-	1.01	-	-	-	-	1.2	-	-	-	0.81
Sum	7	4	5	15	9	21	7	22	2	7	28	12	2

* Determined after performic acid oxidation.

Values in brackets are taken from sequence data.

Table 3. Amino-acid composition of peptides from β I chain of Jaguar.

Pos.	Tp1 1-8	Tp2 9-17	Tp3 18-30	Tp4 31-40	Tp5 41-59	Tp6 60-61	Tp7 62-65	Tp8 66	Tp9a 67-76	Tp9b 77-82	Tp10a 83-87	Tp10b 88-95	Tp11 96-104	Tp12/13 105-132	Tp14 133-144	Tp15 145-146
Asx	-	1.28	2.04	-	3.04	-	-	-	2.07	3.07	-	1.05	2.03	2.10	-	-
Thr	-	-	-	1.00	-	-	-	-	-	-	-	-	-	-	-	-
Ser	1.82	1.00	-	-	3.68	-	-	-	1.77	-	-	0.89	-	-	0.98	-
Glx	2.20	-	2.16	1.03	1.18	-	-	-	-	-	1.07	1.14	4.38(4)	-	-	-
Pro	-	-	-	0.90	-	-	-	-	-	-	-	0.95	0.75	-	-	-
Gly	-	1.71	2.90	-	1.13	-	0.99	-	1.13	-	0.87	-	-	2.12	1.18	-
Ala	0.96	-	1.05	-	3.15	-	1.00	-	-	-	2.06	-	-	3.03	4.14	-
Cys*	-	-	-	-	-	-	-	-	-	-	-	1.02	-	1.04	-	-
Val	-	1.17	2.88	1.53(2)	-	1.09	-	-	1.00	-	-	-	0.96	3.89	2.64	-
Met*	-	-	-	-	1.01	-	-	-	-	-	-	-	-	-	-	-
Ile	-	-	-	-	0.81	-	-	-	-	0.95	-	-	-	-	-	-
Leu	1.02	2.07	1.00	2.13	1.03	-	-	-	2.05	0.98	-	1.99	1.05	3.99	1.15	-
Tyr	-	-	-	0.81	-	-	-	-	-	-	-	-	-	-	-	0.87
Phe	0.96	-	-	-	2.96	-	-	-	0.98	-	1.02	-	0.99	2.89	-	-
His	-	-	-	-	-	0.93	-	-	-	-	1.00	0.91	2.95	1.05	1.12	-
Trp	-	0.73	-	0.87	-	-	-	-	-	-	-	-	-	-	-	-
Lys	1.01	0.85	-	-	0.99	0.92	1.05	1.05	1.00	1.00	0.86	0.96	-	0.95	-	-
Arg	-	-	0.94	1.23	-	-	-	-	-	-	-	0.93	-	1.04	-	-
Sum	8	9	13	10	19	2	4	1	10	6	5	8	9	28	12	2

* Determined after performic acid oxidation.

Values in brackets are taken from sequence data.

Table 4. Amino-acid composition of peptides from β II chain of Jaguar.

Pos.	Tp1 1-8	Tp2 9-17	Tp3 18-30	Tp4 31-40	Tp5 41-59	Tp6 60-61	Tp7 62-65	Tp8 66	Tp9a 67-76	Tp9b 77-82	Tp10a 83-87	Tp10b 88-95	Tp11 96-104	Tp12/13 105-132	Tp14 133-144	Tp15 145-146
Asx	-	1.26	1.94	-	3.19	-	-	-	1.92	2.66	-	1.02	1.99	2.09	-	-
Thr	-	-	-	0.94	-	-	-	-	-	-	-	-	-	-	-	-
Ser	0.99	1.01	-	-	3.67	-	-	-	1.54(2)	-	-	0.91	-	-	1.08	-
Glx	1.94	-	2.10	1.03	1.04	-	-	-	-	-	1.14	1.05	3.93	-	-	-
Pro	-	-	-	0.85	-	-	-	-	-	-	-	0.78	1.06	-	-	-
Gly	1.04	1.89	2.73	-	1.24	-	1.04	-	1.33(1)	-	0.98	-	-	2.14	1.19	-
Ala	0.95	-	1.10	-	2.98	-	1.15	-	-	1.92	-	-	2.96	3.73	-	-
Cys*	-	-	-	-	-	-	-	-	-	-	0.85	-	0.96	-	-	-
Val	-	0.91	2.94	1.62	-	1.04	-	-	1.02	-	-	-	1.05	4.01	2.58	-
Met*	-	-	-	-	0.70	-	-	-	-	-	-	-	-	-	-	-
Ile	-	-	-	-	1.00	-	-	-	-	1.11	-	-	-	-	-	-
Leu	1.09	1.79	1.13	2.14	1.10	-	-	-	2.14	1.16	-	2.03	1.08	4.11	1.28	-
Tyr	-	-	-	0.90	-	-	-	-	-	-	-	-	-	-	-	0.87
Phe	0.96	-	-	2.89	-	-	-	-	1.03	-	1.01	-	0.96	2.75	-	-
His	-	-	-	-	-	1.05	-	-	-	-	-	1.01	1.02	2.96	1.12	1.12
Trp	-	0.82	-	0.79	-	-	-	-	-	-	-	-	-	-	-	-
Lys	0.89	1.18	-	-	1.16	0.98	0.80	1.01	0.98	1.05	1.00	1.00	-	0.98	1.15	-
Arg	-	-	1.02	1.10	-	-	-	-	-	-	-	-	0.92	-	-	-
Sum	8	9	13	10	19	2	4	1	10	6	5	8	9	28	12	2

* Determined after performic acid oxidation.

Values in brackets are taken from sequence data.