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# INTERRELATIONSHIPS OF COPPER, MOLYBDENUM AND SULFATE SULFUR IN BOVINE NUTRITION

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INTERRELATIONSHIPS OF COPPER,  
MOLYBDENUM AND SULFATE SULFUR  
IN BOVINE NUTRITION

BY

JOHN EDWARD VANDERVEEN

B. S., Rutgers University, 1956

A THESIS

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This thesis has been examined and approved.

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## INTRODUCTION

For over one hundred years it has been known that certain pastures in restricted areas of England caused a disease in cattle known locally as "teart". This disease was characterized by severe diarrhea, loss of body condition, and discoloration of the hair. In 1938 Ferguson, et al., attributed the cause of "teart" to a high molybdenum content of these pastures. Proof for this theory was provided when these investigators were able to produce identical symptoms of the disease by either drenching experimental animals with a molybdenum salt or by increasing the content of molybdenum in forage by adding molybdenum to the soil as a top-dressing.

Attempts to reduce the high molybdenum content of the forage were under way when Brouwer, et al., reported that a similar condition in Holland responded to large doses of inorganic copper. Although the "teart" pastures were shown to have what was considered an adequate level of copper, it was found that copper sulfate fed at the rate of 2 grams per day for cows and 1 gram per day for calves, or much smaller doses injected intravenously, would correct the toxic condition caused by the high molybdenum intake. Later it was found that the condition in Holland was not caused by high molybdenum levels but by inadequate copper intake by the animals.

In Australia about this same time investigators studying an enzootic disease of cattle, haematuria vesicalis,

observed that pastures on which these animals grazed, and the tissues of the animals, were abnormally high in molybdenum. Further analysis of the tissues of these animals revealed them to be lower than normal in copper. From this work in Australia and later work in New Zealand with a condition known as "peat scours" it was found that molybdenum has a profound effect on the metabolism of copper in the ruminant. This finding was further substantiated by the successful use of molybdenum salts in treatment of chronic copper poisoning. In effect these workers suggested that molybdenum toxicity is complicated copper deficiency.

Since the time that it was first shown that high molybdenum content of pastures was the cause of "teart" in England, molybdenum toxicity has been reported in Australia, New Zealand, the United States, Canada, Sweden and Japan. It was soon realized, however, that the conditions causing the toxicity in various locations differed greatly. The molybdenum content of the forage in the "teart" pastures of England was much higher than that of forage which caused the toxicity reported in Australia and New Zealand. In addition, the feeding of molybdenum salts was not successful in developing the symptoms of molybdenum toxicity unless it was carried out at a rate far in excess of the rates observed in forages in toxic areas.

These inconsistencies indicated that molybdenum toxicity was complicated by other unidentified conditions. Dick, in Australia, showed that molybdenum could reduce the copper level in the tissues of sheep only when a high level of sul-

fate also was present in the diet. Davis, in Florida, has shown that low phosphorus levels also will influence molybdenum toxicity. Even these findings did not explain the occurrence of molybdenum toxicity in some areas of the United States, England, and New Zealand where relatively low levels of molybdenum in forage are found. Still unexplained, also, is the critical question of how molybdenum exerts its toxic effects on the animal. The mechanism by which molybdenum and other complicating factors affect copper metabolism still is not understood. Whether molybdenum has toxic effects on the body in addition to its effect on copper metabolism is not known.

Losses in animal production in areas where molybdenum toxicity exists can be heavy. Although deficiency symptoms in most areas can be corrected by treating the animal with copper, or top-dressing the land with copper, such treatments are both time consuming and expensive. It was important that more of the answers to this problem be sought through research.

The staff of the U. S. Plant, Soil and Nutrition Laboratory, Ithaca, New York, has been studying conditions which cause poor growth and production of cattle in some western states. The symptoms found in these animals strongly suggested that either copper deficiency or molybdenum toxicity was involved. Examination of the forage in some of these areas revealed an adequate copper content and an above-normal molybdenum level but not high enough to suggest molybdenum toxicity. The presence of some complicating factor being the



cause of this problem was suspected.

As a result of the experience of the New Hampshire Agricultural Experiment Station in mineral metabolism problems and the unusually low trace mineral content of forage grown in New Hampshire, the Agricultural Research Service of the United States Department of Agriculture requested the New Hampshire Station to undertake a study of the molybdenum-copper-sulfate problem on a contract basis. This thesis represents the results of that study.

## LITERATURE REVIEW

## THE ABSORPTION AND METABOLISM OF SULFUR

For many years the absorption and metabolism of sulfur was considered to be important only because of its presence in certain essential amino acids. It was believed that sulfur, which represents 0.15 per cent of the body, could be utilized by the body only as part of the sulfur containing amino acids. In recent years, with the development of the practice of using urea as a protein replacement for ruminants, there has been renewed interest in the metabolism of sulfur.

Harris and Loosli (79) demonstrated that methionine added to the ration of lambs increased the utilization of urea. Later Loosli et al.(101) revealed that 10 essential amino acids were synthesized in the rumen by micro-organisms. This information indicated that, at least in the ruminant, the importance of sulfur in the diet is not confined to the organic sulfur found in amino acids consumed in the diet.

With the use of urea as a source of nitrogen for ruminants, diets could be formulated with a very low sulfur content. Feeding such diets were shown to cause sulfur deficiency. The symptoms of sulfur deficiency in ruminants were described by Starks et al.(148) and Thomas et al.(150) as poor appetite, loss of weight, weakness, excessive lachrymation, profuse salivation, cloudy eyes, emaciation and death.

It was surprising to note that sheep would continue to produce wool with normal sulfur content until death. Inorganic forms of sulfur such as sodium sulfate and elemental sulfur, as well as organic forms, were able to relieve the symptoms of deficiency, according to Starks et al (146,147,148). Ruminants receiving inorganic forms of sulfur added to sulfur deficient diets were shown by Thomas et al. (150), Starks et al. (146,148), Garrigus et al (70) and Lassiter et al. (94) to gain weight normally; and in sheep wool production was shown to increase.

Diets low in sulfur have caused a negative balance for both nitrogen and sulfur. The value of sulfur in increasing nitrogen retention in a low sulfur diet has been well established (70,94,146,147, 148,150). Efforts by Albert et al.(2), Lofgreen et al.(99) and Starks et al. (146,148) to increase nitrogen utilization by the addition of sulfur to diets already containing more than 0.2% sulfur, however, were not successful with lambs. Similar results were obtained by Jones et al (88) with dairy cattle. These workers found no increase in milk production or feed utilization when 1% sodium sulfate or 0.5% methionine were added to a ration containing 0.1% sulfur. For lambs, optimum levels of sulfur in the form of methionine, sulfate, and elemental sulfur have been suggested by Albert et al. (2) as  $0.640 \pm 0.048\%$ ,  $1.270 \pm 0.44\%$  and  $0.471 \pm 0.028\%$ , respectively. Loosli et al.(100) and Starks et al (146) have proposed a diet nitrogen to sulfur ratio of 15 to 1 and 20 to 1, respectively, for good nitrogen uti-

lization.

The ability of the ruminant to utilize cellulose as a source of energy also is influenced by the level of sulfur in the diet. In vitro studies by Hubbert et al.(87) and Trenkle et al.(151) indicated that there was no digestion of cellulose by micro-organisms when the level of sulfur was much below 10 p.p.m. Optimum cellulose digestion was obtained with sulfate sulfur levels between 10 and 20 p.p.m. and only slight depression of cellulose digestion was found when it was supplied at a rate of 1000 p.p.m. Sulfite sulfur gave optimum cellulose digestion when added to the medium in the range of 10 to 15 p.p.m. Levels above 30 p.p.m., however, inhibited cellulose digestion drastically. Organic forms of sulfur were required in much higher levels than inorganic sulfur for optimum cellulose digestion than inorganic forms indicating other beneficial properties than sulfur content. Also, high levels of sulfur, organically combined, did not impair cellulose digestion as was observed with inorganic sources forms of sulfur.

Studies on the fate of inorganic sulfur in the rumen by Anderson (8) and Lewis (97) indicate that sulfate and sulfite ions are reduced rapidly to sulfide by enzymes produced by the micro-organisms found in the rumen. Concentrations up to 97 micrograms per milliliter of sulfide were obtained by Anderson (8) when sulfate was fed at the rate equivalent to 2250 micrograms sulfur per milliliter. He found that sulfide administered as sodium sulfide disappeared from the rumen

very rapidly. It was found that with an initial level of 113 micrograms sulfide per milliliter, one half had disappeared in 30 minutes and all had been removed from the rumen in 2-1/2 hours. It was further shown that sulfide concentrations of the rumen were not affected by the administration of organic sulfur. According to Anderson (8) removal of the sulfide from the rumen is accomplished by direct absorption through the epithelium, and incorporation into amino acids by the bacteria of the rumen. Block et al. (16,17) demonstrated the synthesis of sulfur-containing amino acids by feeding sulfur-35 as sodium sulfate to cattle and goats. Radioactive sulfur labelled cystine and methionine were recovered from the milk of these animals as soon as three hours after administration of the sulfate. Experiments at Michigan by Emery et al. (58, 59) indicated inorganic sulfur is incorporated into organic form faster when added to a diet high in concentrates. In these experiments, bacteria incorporated labelled sulfur twice as rapidly into cystine as they did into methionine. Large amounts of glutathione also were recovered. Attempts by Emery et al. (58) to identify organisms which could utilize inorganic sulfur for amino acid synthesis, revealed that this ability is found in only a few strains and that even these strains would use organic sulfur more readily.

Evidence is now emerging that inorganic forms of sulfur may be used by an animal directly without first being synthesized into amino acids by rumen micro-organisms. Data on the removal of inorganic sulfur from the rumen strongly

indicates that large amounts are absorbed into the body in the inorganic form (8,52,92,99,158). Machlin et al.(103) reported finding radioactive sulfur incorporated into cystine in hens eggs after sulfur-35 was included in the diet as sodium sulfate. These investigators found 1.2% and 0.1% of the radioactive sulfur administered in the diet incorporated into cystine in the egg albumin and egg yolk, respectively. It was interesting to note that no radioactive sulfur was found in the methionine of the egg. In other work Machlin et al (104) reported that 60% of sulfur-35 injected as sulfate into a chick embryo was found incorporated into taurine ( $\text{NH}_2\text{CH}_2\text{-CH}_2\text{-SO}_3\text{H}$ ). These experiments indicate that higher animals have the ability to reduce inorganic forms of sulfur and incorporate them into organic form.

Recent advances in enzyme investigations reveal many new functions of sulfur in metabolism. Evidence that sulfhydryl groups are essential to the activity of many enzymes such as xanthine oxidase, homogentisate oxidase, 3 hydroxyanthranilic acid oxidase, p-hydroxyphenyl pyruvate oxidase and protocatechuic acid oxidase has been reported by Mason (114) and Harris (80). The existence of sulfhydryl groups in the metabolic regulators, glutathione and insulin; and the presence of sulfur in the vitamins, thiamine and biotin, have been known for some time.

The existence of inorganic sulfate in the blood has been regarded for many years only as the end product of sulfur

metabolism. As more sulfate bound compounds have been discovered in the body, the importance of this ion in body metabolism has been reevaluated. Many of these compounds such as aromatic sulfonates and steroid sulfates appear to be products of detoxification. Sulfate compounds like brain lipids and mucopolysaccharides, however, have been shown to be essential to body metabolism. The oxidation of organic sulfur in animals has been reviewed by Fromageot (67). The metabolism of sulfate has been reviewed recently by Gregory and Robbins (74). According to Weir and Rendig (158) serum inorganic sulfate levels reflect the total sulfur intake in sheep. When an animal has been fasted for a period of time, sulfate levels of the blood become high due to the metabolism of proteins for energy. Normal ranges of serum sulfate appear to be 2.0 to 5.0 mg. per 100 ml. of serum for sheep.

Although the excretion of sulfur occurs mostly in the urine, some is found normally in the feces. Recent balance studies by Muller et al. (127) indicate that half of the sulfur excreted by the bovine and two-thirds of that by sheep is in the form of inorganic sulfate in the urine. Dick (52), using sheep, found that 95 per cent of sulfur-35 fed as inorganic sulfate was excreted within 72 hours and only 1 to 3 per cent was found in the feces. Kulwich et al. (92) reported recovering 34% of the sulfur-35 in the feces and 52% in the urine in the four days following its administration as sodium sulfate. There appears to be no explanation for these differences. Balance studies by Knappen et al. (90) in Germany indicate that

the urinary sulfur cannot be used to obtain information on sulfur balance.

Sulfur in the diet as either organic sulfur or sulfate sulfur is not known to be toxic, even at very high levels. According to Anderson (8) the liver has the ability to oxidize sulfides at an estimated rate of .006 grams per hour per gram of liver. At this rate the production of sulfide in the rumen from inorganic sulfate contained in the ration at a level of 2% would not be harmful. The work of Starks et al. (146) indicates that the palatability of both inorganic sulfate and elemental sulfur are such that animals cannot be made to consume levels higher than 1.78% and 0.6% respectively.



## COPPER ABSORPTION AND METABOLISM

The presence of copper in animal tissues has been known for over a hundred years, according to Underwood (152). Like many other elements, little attention was given to its metabolism because it appeared in very low concentrations and was thought only to be a biological contaminant. In 1928 Hart et al.(81) created the first real interest in copper metabolism in mammals when they found that copper was required in addition to iron for the formation of hemoglobin. Details of the progress in copper metabolism in animal research have been the subject of many recent reviews by Underwood (152,153), Mason (113), and Singer et al.(144) and should be consulted for extensive details.

Since copper was first shown to be essential for several metabolic processes in animals, deficiencies of this element have been reported to occur naturally in many parts of the world (4,21,31,39,152). Ruminants, particularly cattle and sheep, are more sensitive to copper deficiency than the non-ruminant species. The symptoms most commonly described in ruminants are anemia, depressed growth, achromotrichia, gastrointestinal disturbances (diarrhea), poor bone formation, and ataxia. Alopecia has been described in cattle, and the wool in Merino sheep lacks the normal crimp.

The known functions of copper in the body have not explained satisfactorily the various symptoms which have been described for copper deficiency. Copper is now known to be

contained in the enzymes phenol oxidase, p-hydroxyphenylpyruvate oxidase, diphenylalanine (dopa) oxidase, butyrylcoenzyme A dehydrogenase, uricase, laccase, and cytochrome oxidase. Information on the properties of these enzymes has been reported by Mahler et al. (106,107,109), Singer et al. (144), and Waino (157). It is quite surprising that only the activity of cytochrome oxidase has been shown to be affected by experimentally induced copper deficiency. As early as 1939 it was noted by Schultze (140) that cytochrome oxidase activity was reduced during copper deficiency. Another enzyme, dopa oxidase, which is required to oxidize dihydroxyphenylalanine (dopa) and tyrosine to the pigment melanin is suspected of being reduced in activity during copper deficiency, thus explaining the loss of hair color.

It is interesting to note that the function of copper in hemoglobin formation, although it was the first evidence that copper was required in metabolism, still remains obscure. A recent review by Matrone (115) concerning the interrelationship of iron and copper states that only in the formation of hemoglobin is there an apparent interaction. Work done with calves by Matrone at North Carolina (115) indicates that iron absorption is not impaired by copper deficiency in contrast to the lower absorption of iron in copper deficient swine as reported by Gubler et al. (75). The only consistent finding with respect to the addition of copper to copper deficient animals is an increase in the per cent of reticulocytes, as reported by Matrone (115). This increase in per cent of

reticulocytes was demonstrated in dogs, rats, rabbits and swine, but no similar work involving ruminants has been reported.

Copper has been reported by McCutcheon (116) to be an additional co-factor in the exchange of formate with inosinic acid. The exact function copper has in this reaction is unknown.

Bone formation abnormalities have been noted in copper deficient sheep, cattle, pigs, dogs, and chickens by Underwood (152). Studies with both dogs, by Follis (66), and swine, by Bush (24), indicate a similarity between the changes in bone formation during copper deficiency and scurvy. In both cases there is a normal growth of cartilage but the deposition of bone in the cartilage matrix has been stopped. There is as yet no indication as to how copper deficiency interferes with the functional activity of the osteoblasts. Deficiencies of other elements generally stop both the osteoblastic and chondroblastic activities together, thus resulting in shorter bones but with normal bone conformation. The bone growth in copper deficiencies is of normal length but the conformation is altered due to the softer nature of the bone.

The symptom which is most readily observed in copper deficiency of ruminants is the change in hair and wool. According to Hayashi (82) and Cunningham (31) achromotricia and alopecia are commonly reported in cattle. The texture of such hair has been described by Cunningham (31) as starring and rough. In Merino sheep the crimp normally found in wool is absent and the physical properties are inferior. Burley et

al. (22) have shown that such wool has a greater number of sulfhydryl groups and fewer disulfide groups than normal wool. More recent evidence by Burley et al. (22) indicates that copper deficiency interferes with the arrangement of polypeptide chains in keratin. These workers have shown that wool from copper deficient sheep contains more N-terminal glycine and alanine and sometimes more N-terminal serine and glutamic acid than normal wool. Cunningham (36) has reported that the copper content of hair was not changed by the level of this element in the diet but that the copper content of hair varied with hair color. Copper-containing proteins which have been shown to catalyze the aerobic oxidation of L (-) cysteine, L (-) tyrosine, and  $\beta$ -3,4 dihydroxyphenyl alanine have been isolated from sheep's hide by Scaife (139).

Cardiac failure has been reported by Bennetts (14) in cattle grazing on pasture extremely low in copper in southwestern Australia. The condition was claimed to be caused by atrophy of the myocardium with replacement fibrosis. A similar condition has now been reported in pigs by Gubler et al (75). These workers also reported cardiac hypertrophy to be common in copper deficient swine and suggested its cause to be an attempt to compensate for the loss of tissue respiration which occurs as a consequence of reduced cytochrome oxidase activity. Cardiac failure is thought to result from stress on a depleted myocardium.

The assimilation of dietary copper in the ruminant is influenced by many conditions and generally amounts to only a

few per cent of that consumed. Comar et al.(25,26), feeding copper-64 as the nitrate salt, found that 75 per cent of the activity was recovered in the feces and 3% was found in urine after five days. In these experiments about 10 per cent of the dose was found in the tissues. In spite of the very low amounts of copper-64 absorbed, 0.15 per cent of it was found in the blood only 19 minutes after being administered orally. It is surprising that only limited research has been carried out in determining the availability of different forms of copper to the animal, and even less research has been undertaken to learn the mechanism of its absorption. Schultze et al.(141,142), using rats as an experimental animal, showed that copper in the form of copper glycine amide biuret, alanine amide biuret, hemocyanin, caseinate, aspartate, citrate, nucleinate, pyrophosphate, citrate, and cysteine cuprous mercaptide could be used by animals fed a copper deficient diet, whereas, copper hematoporphyrin and sulfide were not utilized. These investigators could not establish any information on the form in which such copper was absorbed because the copper was shown to be released from many of these compounds in dilute acid media. Recently Mills (123) has investigated the forms in which copper is found in plants and the ability of the animal to assimilate plant copper. These investigations showed that only 8 to 22 per cent of the copper in the plant was in a form which was soluble in water or organic solvents. The remaining copper was shown to be released from its organic form only in a solution with a pH lower than 2. This pH is far

lower than is found in the digestive tract of animals. In studies with rats, however, this investigator showed that plant-contained copper was more readily absorbed than was inorganic copper of the same concentration. This would indicate that copper can be transported through the intestinal mucosa as an organic complex as well as the free ion. Further support for this theory can be found in the observation of Mitchell and Tosie (125) that copper ions are very rapidly absorbed by rumen micro-organisms and incorporated into an organic form. Mills (123) states that copper in an organic complex would pass the membranes more rapidly because it would not be free to combine with complexing agents found in the membrane. It is interesting to note that the quantity of free copper in plants showed seasonal variations. Davis (40) recently reported isolating from grasses a complexing agent which renders copper completely unavailable to animals. This worker suggests such a complexing agent is responsible for the occurrence of copper deficiency in areas where seemingly adequate copper is present.

The total amount of copper found in the tissues of an animal is quite small. According to Underwood (152) mammals at birth generally contain from 3 to 7 p.p.m. copper but the quantity goes down in the adult to 2.0 to 1.5 p.p.m. Data on the levels of copper for sheep and cattle are not available, but it is expected that the copper contents of these animals are higher than for other mammals because the analysis of many individual tissues shows that they contain more copper. Although there is relatively little copper found in the body,

the distribution is very wide and there is good evidence to believe that copper can be found in every cell of the body. The liver of most animals, particularly sheep and the bovine, have high concentrations of copper. Normal ranges of copper in these species are reported by Underwood (152), and Marston (112), to be 100 to 400 p.p.m. Copper levels in the blood of sheep and cattle also have been found to vary over a wide range. Underwood (152) and Marston (112) have summarized values from many investigators and set the normal range at 80 to 120 micrograms per 100 ml. of blood. Values as low as 50 and as high as 180 micrograms per 100 ml. of blood, however, are commonly found. The level of copper in the blood has been shown to be affected greatly by many factors. Low levels of copper in the diet have been shown to cause decreased blood levels by Cunningham (31) and Bennetts et al. (14). In humans, pregnancy has lowered blood copper values, but no data on sheep or cattle are available. Heavy infestations of internal parasites have been shown by Bremner (19), Seekles (143) and Gibson (71) to lower both blood and liver concentrations of copper. The action of molybdenum in lowering blood copper levels will be discussed later.

Many balance studies have shown that the chief route of excretion of copper is through the intestinal tract. Comar et al. (25,27) reported that when copper-64 was injected intravenously into cattle only 3 per cent was excreted in the urine and an equal quantity in the feces. After reviewing the literature, Underwood (152) reported that 90 per cent of the dietary copper appears in the feces. Copper is excreted into

the intestinal tract by way of the bile.

The level of copper required in the diet for normal animal health is influenced greatly by many factors. In most areas of the world, levels of copper from 4 to 6 p.p.m. on a dry weight basis are adequate for normal health of ruminants, according to Cunningham (31), Marston (112), and Underwood (152). The form of copper in the diet has been discussed already in relation to assimilation and obviously is related to the level needed in the diet. The level of molybdenum in the diet, which will be discussed later, has a profound effect on the copper requirement. Recently, there has been growing evidence that zinc also has an effect on the copper requirement. High levels of zinc in the diet have been reported by Cox et al, (29) to require higher levels of copper. Zinc with lead has been shown by Gray and Ellis (73) to cause a lowering of hemoglobin values in rats but this could be overcome by copper supplementation. Earlier, Cunningham (31) reported zinc had no effect on copper metabolism. This investigator reported also that tungsten, manganese, vanadium, chromium, rhenium, uranium and tantalum have no effect on copper requirements.

Additional copper has been administered to both sheep and cattle successfully, both as copper salts added to the diet or as an injected cerate. Reports of copper being injected as a cerate indicate up to 90 per cent utilization in work reported by Branion (18) and Cunningham (31). Copper glycinate, given at the rate of 400 mg. for cattle and 150 mg. for sheep, has been used successfully for many years. A



recent report by Cunningham (35) on the use of cupric bis-8-hydroxyquinoline 5:7 disulfonic acid salt of tetraethylamine in cerate injections showed copper in this form was slowly absorbed, as desired, and caused no harmful effects to the tissues, but copper citrate caused great tissue damage when used as a cerate. Branion (18) reported that copper administered as a cerate must be given three times per year to maintain animal health. Dent et al. (42) and Cunningham (30) reported some success in meeting the requirements of cattle for copper by offering a free choice of a mineral mixture containing copper. Top-dressing of 10 to 20 pounds of copper sulfate per acre on forage has met with only moderate results in raising the copper content of the forages according to Branion (18) and Cunningham (33). Such a practice has proved to be very uneconomical. The feeding of copper sulfate at the rate of 3 grams for cattle and 0.5 grams for sheep has been effective but such treatment is required 3 times per week.

Copper salts taken in large doses have been known to be toxic for many hundreds of years, but only recently has the recognition of chronic copper poisoning under natural conditions occurred. Copper poisoning has been described by Dick (55) and Underwood (152) both in sheep and cattle grazing on pastures having a high copper content. In many cases of such poisoning the levels of copper in the forage were only 15 p.p.m. Kidder (89) demonstrated that hemolytic jaundice could be caused by large doses of copper over a long period of time. Branion (18) reported that feeding 3 g to 5 g per

day for a period of 3 years, however, was not harmful. Sheep are much more susceptible to copper poisoning than are any other species. Frequent accidental poisoning with orchard sprays and copper salts used for parasitic treatment has been reported by Allcroft (7) and Underwood (152). According to Dick and Bull (55) copper poisoning is characterized by sudden hemoglobinemia and hemoglobinuria usually accompanied by the occurrence of icterus which increases rapidly.

Excess copper intake may also cause reduction in feed utilization by ruminants. Studies carried on in vitro by McNaught et al. (117) indicated that levels of copper above 10 p.p.m. greatly reduced protein digestion. In other in vitro studies, Hubbert et al. (87) showed a level of 1.5 p.p.m. copper as the sulfate reduced cellulose digestion and 2.5 p.p.m. stopped digestion of this substance completely.

## THE ABSORPTION AND METABOLISM OF MOLYBDENUM

The discovery that molybdenum is part of xanthine oxidase was made by two groups of investigators, de Renzo et al. (43) and Richert and Westerfeld (134), working independently. Mahler et al. (110) in 1954 reported that molybdenum is a part of liver aldehyde oxidase. The third molybdenum enzyme, hydrogenase, was reported by Shug et al. (144) also in 1954. Although these investigators were able to show increased enzyme activity with increased molybdenum levels in the diet, the element was not considered essential because growth and purine metabolism were not affected by changes in dietary levels. Tungsten, as the tungstate, has been shown by de Renzo (45) to reduce the molybdenum level in the tissues, presumably in the role of an antagonist. Higgins et al. (85), feeding tungstate to rats, showed that the xanthine oxidase activity could be reduced by three-quarters without changing the uric acid excretion. In this experiment the xanthine oxidase production of the intestines was completely stopped. These same investigators showed that feeding sodium tungstate to chicks caused a 25 per cent decrease in growth in five weeks. This treatment reduced the secretion of uric acid which was replaced by xanthine and hypoxanthine. Molybdate added to the diet overcame all these effects. More recent work with rats by Richert et al. (135) showed that both xanthine oxidase and aldehyde oxidase activity could be reduced by feed-

ing tungstate, to a point where normal metabolism did not occur.

Claims that molybdenum is an essential element have gained support in recent years. Ried et al.(132), in 1957, was able to increase the growth of turkey poults by adding molybdenum to the diet. Leach and Norris (96) were able to show a similar growth response with chicks but were unable to confirm a response in turkeys. Ellis et al.(57) reported a beneficial effect in sheep when 1 and 2 p.p.m. of molybdenum were added to a basal diet containing 0.36 p.p.m. molybdenum. These sheep made significantly faster gains with the added molybdenum than were made on the basal diet. The addition of the molybdenum to this diet significantly increased cellulose digestion without altering digestion of other components of the ration. It appears now that low molybdenum levels can interfere with normal purine metabolism in sheep. Askew (10) has demonstrated that low molybdenum levels in the diet contribute to xanthine calculi formation in the kidneys of sheep.

Molybdenum is absorbed readily as the molybdate ion. Comar et al.(27), using molybdenum-99, found that only 34 per cent was recovered in the feces after being fed as the molybdate ion. These workers found that 45 per cent of the molybdenum-99 was excreted in the urine. These results are in good agreement with those of Dick (52). This investigator has demonstrated that molybdenum levels in the blood and tissues of the animal reflect the animal's intake of the ele-

ment. Molybdenum will increase to a certain level in the body, depending on intake, and then remain constant. If the molybdenum level in the diet is decreased the tissue concentration will decrease. The bone, kidney, spleen, liver, and blood show the greatest concentration of this element according to Anderson (8) and Comar et al.(27). Molybdenum excreted in the milk appears also to reflect the intake. Comar et al.(27) found that about 2 per cent of a dose of molybdenum-99 was excreted in the milk of a cow.

The molybdenum content of the forage is dependent on the molybdenum content of the soil, soil pH, and species of plants. According to Barshad (11) high pH favors greater accumulation of molybdenum in plants. Legume plants growing in the same field as grasses will contain many times as much molybdenum (11,18,64,84). In many cases legume plants, particularly white clover, have been shown to contain 160-260 p.p.m. molybdenum. Normal contents of plants, however, are assumed to be in the range of 1 to 2 p.p.m. throughout the world (11, 18,64,84). Ferguson (64) reported that 70 to 80 per cent of the molybdenum in grass, 40% of that in hay and 10% of that in dead winter herbage was water soluble.

Molybdenum was first found to be important in animal nutrition because of its toxic effect. Ferguson et al.(62) in 1938 reported that molybdenum was responsible for the sickness of cattle and sheep pastured on the "teart" areas of England. Since that time molybdenum toxicity has been reported by Cunningham (31) in New Zealand, by Dick (51) in

Australia, by Hayashi (82) in Japan, Hallgren et al. (77) in Sweden, Cunningham et al. (30) in Canada, and by Britton and Gross (20), Robinson and Dever (136), and Davis (39) in the United States. The symptoms of this toxicity in cattle and sheep have been described as follows: achromotricia, ataxia, unthriftiness, debilitating spring scouring, anemia, increased susceptibility to internal parasites, proneness to fracture of the bones, loss of reproductive capacity, low serum copper and low liver copper (4,30,31,52,82,112, 118,149).

Hair and wool of ruminants suffering from molybdenum toxicity have shown many changes. In adult cattle black hair has changed to red or gray, and alopecia has occurred around the eyes, according to Cunningham (31), Ferguson (63,64) and Hayashi (82). Hayashi (82) has reported that individual hairs of these animals have been shown to have alternate clear and pigmented areas and the clear areas show less strength than other areas. Cunningham (31) and Underwood (152) have reported that black wool of sheep on toxic levels of molybdenum shows loss of pigmentation. Such wool lacks quality, and because the normal crimp is absent, it has a stringy appearance. Ataxia generally occurs in lambs between the ages of 3 and 4 months, but occasionally young cattle have been seen with the disorder, according to Cunningham (31). Allcroft (4) and Underwood (152) report that the condition is caused by diffuse symmetrical demyelination of the central nervous system.

The loss of reproductive capacity has been observed experimentally in bulls, by Hayashi (82,83), and Thomas and

Moss (149). In these experiments spermatogenesis stopped entirely and the individuals completely lacked libido. Histological examinations of the testes showed that there was damage to the seminiferous tubules and inflammation of the interstitial tissue. Copper administration has been shown to correct all symptoms of the disease with the exception of advanced ataxia, but copper has been shown to prevent this condition from occurring.

THE INTERRELATIONSHIP OF MOLYBDENUM,  
COPPER AND SULFATE SULFUR

The original description of molybdenum toxicity by Ferguson (48) implied a relationship of this element with copper. Not only were many of the symptoms of this disorder the same as those described for copper deficiency, but the fact that copper sulfate was a successful treatment indicated a possible relationship. Dick and Bull (55), while studying copper metabolism, discovered that copper levels of sheep livers were below normal when a large amount of molybdenum was found in the diet. These workers used molybdenum to correct copper toxicity. Molybdenum toxicity, both in New Zealand (Cunningham, 31) and Florida (Davis, 39), was described as occurring with levels of molybdenum intake between 7 and 10 p.p.m. Davis (39) later reported that levels of molybdenum as low as 2 p.p.m. could cause molybdenum toxicity. It became apparent that the molybdenum toxicity in certain areas was caused by much lower levels of molybdenum than were originally described by Ferguson et al. (63). Furthermore, attempts to induce experimentally molybdenum toxicity required much higher levels than occurred naturally in toxicity areas.

According to Ferguson et al. (63) molybdenum toxicity in England was caused by very high levels of the element. Plants on the "teart" pastures contained from 20 to over 150 p.p.m. of molybdenum. Animals on these pastures are affected



by the high molybdenum content within a week, or two weeks, after the start of grazing. Most prominent symptoms include severe diarrhea, emaciation, poor production, and changes in hair color. Levels of molybdenum below the range indicated do not give these symptoms. Feeding molybdenum as molybdate at levels of 100 p.p.m. or more has been shown by Branion (18), Ferguson et al. (63), Hayashi (82), and Hallgren et al. (77) to cause the same symptoms.

Toxicity in New Zealand, Australia and Florida occurred on levels as low as 2 to 10 p.p.m. of molybdenum, according to Branion (18), Cunningham (31), Davis (39), Dick (52), Miller et al. (118), and Wynne and Glymont (159). The symptoms of the toxicity in these areas are not shown to occur quite as rapidly as those described in England but appear to be just as severe. Diarrhea, emaciation, anemia, and hair color changes are the first symptoms to be noticed. Analysis of the blood and liver for copper show levels to be below normal. Copper levels as low as 0.2 p.p.m. in the blood have been reported by Branion (18) and Cunningham (31). Molybdenum toxicity in these areas is most frequently associated with peat or muck soils. According to Cunningham (31), in New Zealand the problem was first encountered when a large program of reclaiming swamp land was carried out and cows and sheep were grazed on this new land.

Dick (49), while studying enzootic jaundice caused by high levels of copper, tried to reduce the copper in the animal with molybdenum and found that molybdenum was effective

in reducing copper with alfalfa but not with oat straw as forage. In later work this investigator indicated that the difference between the two diets which caused the effect was a high level of sulfate in the alfalfa hay. Dick (46,47,48, 49,50,51,52,53,55) further showed that supplements of neither sulfate nor molybdenum alone had any effect on copper metabolism separately, but together they caused all of the symptoms of molybdenum toxicity in sheep. Similar results with cattle have been reported by Cunningham (31) and Mylrea (128). It is interesting to note that studies by Kulwick et al. (93) using copper-64 in swine and rats, showed an increased copper deposition when molybdenum was added to the diet. These investigators suggest that copper may be unusable by the animal.

In studying the effects of sulfate on molybdenum metabolism, many investigators have become quite perplexed. In rats and other monogastric animals, sulfate was shown by Miller et al. (120,121), Reid et al. (131), and Van Reem (155, 156) to protect against the toxic effects of high molybdenum intakes in the diet. High levels of methionine were shown to have the same protective effect on molybdenum toxicity in rats, according to Gray and Daniel (72). Experiments with sheep by Dick (52) indicate that the molybdenum content of the blood and tissues can be reduced by the addition of inorganic sulfate to the diet. A single dose of sulfate was shown to increase greatly the level of molybdate excreted in the urine. Similar results have now been reported in cattle by Cunningham (37). Increases in the molybdenum content of both plasma and

blood cells were reported when only molybdenum was added to the diet but when sulfate was added with molybdenum the plasma alone showed an increase.

There is very little evidence to suggest that sulfate alone has any effect on copper metabolism. Mylrea (128) has reported that the liver copper of cattle was reduced from 144 p.p.m. to 63 p.p.m. by the addition of 1% gypsum to the diet. Similar experiments using other forms of sulfate have not shown any effect, according to Dick (52) and Cunningham (37). Singer et al. (144), in a review on copper enzymes, report that sulfate has an inhibitory effect on the enzyme laccase.

Indications of the phosphorus levels also adding to the severity of molybdenum toxicity have been reported by Cunningham (31) and Davis (39). In these situations normal phosphorus metabolism was shown to be altered, and bone fractures were frequent. In all cases, however, the phosphorus level of the diet was shown to be below what was considered an adequate level. Excess phosphorus has not been found to relieve molybdenum toxicity, according to Davis (39). Conflicting evidences to the effects of manganese in relieving toxicity caused by high molybdenum and sulfate intake has been found. Dick (51) has reported that manganese would counter the effects of high levels of sulfate and molybdenum. Mylrea (128) was unable to show similar results with cattle.

Despite the relations described above, certain areas

are known where animals show all the symptoms of molybdenum toxicity or copper deficiency, while in other areas identical conditions seem to exist without causing this malady. In England, Allcroft et al. (4) showed identical conditions of molybdenum, sulfate, and copper levels on two locations, but one had the symptoms of molybdenum toxicity in sheep and cattle and the other did not. In these areas copper was frequently as high as 24 p.p.m., molybdenum levels were about 7 p.p.m., and sulfate levels were as high as 1.1 per cent inorganic sulfate. Furthermore, even with high levels of sulfate, all of the symptoms of molybdenum toxicity could not be produced experimentally with levels of molybdenum below 50 to 100 p.p.m., as was shown by Branion (18) and Hayashi (82). A recent review of the conditions found in the Everglades area of Florida by Kretschmer and Beardsley (91) states that molybdenum has never been conclusively proven to be the causative agent for what was supposed to be molybdenum toxicity in this area. A response to copper supplementation, however, was well demonstrated. This author pointed out that many times the condition could be corrected by phenothiazine treatment, thus indicating that the problem was a copper deficiency brought on by internal parasites. Copper levels in the forage of this area are from 3 to 8 p.p.m. Such data as described here, indicate that there are still unidentified complicating factors involved in molybdenum toxicity.

That there exists a relationship among copper, molybdenum and sulfate has now been established by the work of Dick (52), Cunningham (37), and Mylrea (128) in the bovine

and sheep. The exact levels at which the interreaction affects animal health have not been established, nor has the exact mechanism by which this interrelationship operates been proven. Dick (52) has proposed that inorganic sulfate at high levels interferes with the transport of molybdenum across membranes. Such a membrane at which molybdenum is blocked would then interfere with the copper transport. Under this theory copper deficiency would occur due to reduced copper absorption from the alimentary tract. Further, this author suggests in the case of high levels of molybdenum, copper would be prevented from leaving the storage areas or the blood to enter the tissues where it is needed and thereby produce the symptoms of copper deficiency without lowering the copper levels of the blood or the liver. No details have been proposed by the proponent of this theory as to how sulfate stops molybdenum from passing through the membrane or how the latter in turn prevents copper from crossing a membrane. This theory, however, explains most of the observed conditions as to why blood and liver concentrations of copper vary so greatly when toxicity is caused by both high and low levels of molybdenum. Recent results of Cunningham (36) on the copper and molybdenum contents of hoof and hair of animals fed high levels of molybdenum and sulfate shows that molybdenum decreased the copper content of hair, wool and hoof, both with and without sulfate. The molybdenum content of the hair, wool and hoof was affected by the level of this element in the diet, but sulfate added to the diet decreased the level of molybdenum in these tissues.

This information would seem to contradict Dick's (52) theory because molybdenum alone seemed to have an effect on the movement of copper. It was interesting to note further that the copper level changes in the hair did not affect color, according to Cunningham (36).

A second theory on the copper, sulfate and molybdenum interrelationship has been proposed by Mills et al.(124) and has been extended by Halverson et al.(78). Mills et al.(124) reported that high molybdenum levels depressed the sulfide oxidase levels in the liver of the rat by 56 per cent. They then proposed that high sulfide levels may be produced from reduction of sulfate in the rumen. This sulfide would be toxic to the animal because of a limited ability to oxidize this compound. The depression of sulfide oxidase activity was explained on the basis of the fact that both copper and hypoxanthine are required as co-factors. Halverson et al. (78) reported that cystine did not reduce the toxic effects of molybdenum as methionine and inorganic sulfate were shown to do. These investigators reported that high levels of cystine were not oxidized by the rat because of reduced sulfide oxidase activity, and that this cystine formed a complex with copper rendering the latter unavailable to the animal. Halverson et al.(78) suggested that small amounts of sulfide in the tissues, even though undetectable, could form insoluble copper sulfide and thus render the copper useless to the animal. It is interesting to note that Van Reem (155) found that feeding potassium thiomolybdate ( $K_2MoS_4$ ) to

rats was extremely toxic. Singer et al. (144) have reported that hydrogen sulfide can be shown to inhibit the activity of copper enzymes by complexing with the copper.

The question of whether the interaction with copper is the only toxic effect molybdenum has on an animal has been raised several times, especially by Branion (18) and Miller et al. (118). This theory that molybdenum is toxic in some other system is strongly supported by the research of Branion (18) in which high levels of molybdenum caused the symptoms of toxicity to appear before any changes in copper levels of the blood and liver occurred. Further evidence that molybdenum per se is toxic is provided by the observation of Scaife (139) that the molybdate ion inhibited the action of three copper containing enzymes isolated from the skin of sheep. These enzymes, which were referred to earlier, catalyze the aerobic oxidation of L (-) cysteine, L (-) tyrosine, and dopa (3,4 dihydroxyphenylalanine).

## EXPERIMENTAL PROCEDURE

The experimental procedures used to study the inter-relationship among copper, molybdenum, and sulfate sulfur will be described as four separate experiments for the sake of clarity. These will be designated as Experiments I, II, III, and IV.

### EXPERIMENT I

The purpose of Experiment I was to investigate the effects of various levels of molybdenum supplementation on milking first calf heifers fed a diet relatively low in copper, molybdenum, and sulfate. Fifteen springing Holstein heifers purchased in the last summer of 1957 from dairy farms were brought to the Dairy Nutrition Barn at the University of New Hampshire Agriculture Experiment Station for this experiment. The experiment was designed with three replications, each having five animals on different treatments. The treatments consisted of a control and levels of 5, 10, 20, and 50 p.p.m. of molybdenum added to the total ration. Table 1 illustrates the experimental design. Each animal was placed on experiment at parturition and was assigned to a treatment at random. All animals were kept on experiment for four consecutive periods of 75 days each.

The animals were fed a 17 per cent crude protein concentrate mixture, which was low in copper, molybdenum, and sulfur, according to the Morrison Feeding Table (126) with



TABLE 1

## Design of Experiment I

Number of animals	Molybdenum fed	Sulfate sulfur	Other ions fed	Days spent in		
				Pole barn	Stanchion barn	Pasture
3	None	None	None	None	300	None
3	5 p.p.m.	None	None	None	300	None
3	10 p.p.m.	None	None	None	300	None
3	20 p.p.m.	None	None	None	300	None
3	50 p.p.m.	None	None	None	300	None

the adjustment for first calf heifers. The composition of this mixture is found in Table 2 of the Appendix. Hay consisting predominantly of grasses grown under high fertilization was fed ad libitum. Concentrate and hay weights were recorded, and refuse was weighed back three times per week and recorded. Molybdenum in the form of sodium molybdate ( $\text{Na}_2\text{MoO}_4 \cdot 2\text{H}_2\text{O}$ ) was used as the molybdenum supplement. The amount to be fed was calculated on a weekly basis using the previous week's total feed consumption as the basis for the calculation. The weekly allowance of the molybdate salt was weighed out on an analytical balance, diluted with cane sugar, and then divided into seven portions. Each portion was placed in a quarter-ounce gelatin capsule and one of these was administered each day. A plain salt block was accessible to each animal and individual automatic drinking cups provided water.

The animals were exercised for a few hours each day in a small lot when weather permitted. Heat periods were recorded and each animal was serviced artificially during the first heat occurring after 60 days post-partum. Pregnancy examinations were made once per month until an animal was found to be with calf. The cows were milked twice a day by machine and the milk weights were recorded. At the end of 300 days on experiment the animals were slaughtered and a section of liver was saved for analysis.

## EXPERIMENT II

The purpose of Experiment II was to study the effects of various levels of molybdenum supplementation on milking first calf heifers fed a diet high in sulfate sulfur and relatively low in copper and molybdenum. Fifteen springing first calf heifers were purchased for the experiment in the late summer of 1958 from dairy farms and brought to the Dairy Nutrition Barn. The experimental design of this experiment was planned to be identical with that of Experiment I except that 0.3% sulfate sulfur was added to the diet. As the experiment progressed, however, the following changes were made: At the end of 150 days on experiments two of the three animals receiving 5 p.p.m. of molybdenum showed positive reactions to a blood test for brucellosis and were removed from experiment and slaughtered to prevent further spread of the disease. The remaining 5 p.p.m. animal and the three 10 p.p.m. animals were carried on experiment for three consecutive periods of 75 days (275 days) and then divided into two new treatments with two animals receiving 100 p.p.m. and two getting 200 p.p.m. of supplemental molybdenum. These treatments were continued for one period of 75 days. At the end of four consecutive 75-day periods (300 days) on experiment, two animals from the control group and two animals from the 50 p.p.m. level were continued on treatment through the remainder of the first lactation, the ensuing dry period, and four consecutive 75-day periods during the second lactation. The

design of Experiment II is shown in Table 2.

The animals were placed on experiment and fed hay and concentrates of the same quality and in the manner as described for Experiment I except for the addition of potassium sulfate to the concentrate mixture. The feeding of sodium molybdate was carried out according to the same procedures as were used in Experiment I. The procedures followed with respect to milking, breeding, and housing were the same as for Experiment I with the exception of exercise. The animals in Experiment II were exercised for the first 150 days but this practice was discontinued in an attempt to prevent the spread of brucellosis.

### EXPERIMENT III

Experiment III was designed to investigate the effects of stress, pasture, and the addition of some ions which are found only in small amounts in New Hampshire grasses, on the development of toxicity in cattle fed moderately high levels of molybdenum and sulfate sulfur.

Fifteen first calf heifers were purchased in the late summer of 1959 and placed on experiment in the same way as in Experiments I and II. This experiment was designed with three replications of five different treatments. The treatments in each replication were as follows: 0.3 per cent sulfate sulfur - outdoors; 50 p.p.m. molybdenum - outdoors; 0.3% sulfate sulfur and 50 p.p.m. molybdenum - outdoors; 0.3% sulfate sulfur and 50 p.p.m. molybdenum - indoors; 50 p.p.m. molybdenum,

TABLE 2

Design of Experiment II

Number of animals	Molybdenum fed	Sulfate sulfur	Other ions fed	Days spent in		
				Pole barn	Stanchion barn	Pasture
3	None	0.3%	None	None	(1) 300 (2) 600	None 75 days
3	5 p.p.m.	0.3%	None	None	150	None
3	10 p.p.m.	0.3%	None	None	225	None
3	20 p.p.m.	0.3%	None	None	300	None
3	50 p.p.m.	0.3%	None	None	(1) 300 (1) 675 (1) 600	None None 75 days
2 <sup>**</sup>	100 p.p.m.	0.3%	None	None	75	None
2 <sup>**</sup>	200 p.p.m.	0.3%	None	None	75	None

\* Animals were either 5 p.p.m. or 10 p.p.m. Mo. plus 0.3% Sulfate before this treatment.

\*\* Number in parenthesis indicates number of animals.

0.3% sulfate sulfur, 5 p.p.m. arsenic, 5 p.p.m. lead, and 100 p.p.m. rubidium - indoors. The designation "outdoors" and "indoors" refer to the conditions under which the animals were housed other than during the milking period. The "outdoor" animals were kept in a lot and had access to an open pole barn. The "indoor" animals were housed in a station barn. After 225 days on experiment one animal from each treatment was selected at random and placed on pasture instead of receiving hay. With the exception of this change, all other conditions of the experiment remained the same for 300 days. The design of the experiment is shown in Table 3.

At the end of 300 days on Experiment III four animals which received 50 p.p.m. molybdenum and 0.3% sulfate sulfur and showed extensive hair color changes, were selected to receive additional copper. These animals were continued on their experimental diet but two received 10 p.p.m. copper and two received 20 p.p.m. copper added to the diet as copper sulfate. This treatment was then continued for 75 days at which time serum and liver samples were taken.

The animals received hay and concentrates of the same description as were used in Experiment I. The animals housed outdoors received hay in an open rack as a group and individual hay consumption data was not possible. Hay was fed to animals housed indoors in the same manner as described for Experiment I. All animals were milked and fed concentrates twice a day. The amount of sulfate required in the diet was supplied in the form of potassium sulfate mixed with the con-

TABLE 3

## Design of Experiment III

Number of animals	Molybdenum fed	Sulfate sulfur fed	Other ions fed	Days spent in		
				Pole barn	Stanchion barn	Pasture
3	None	0.3%	None	(2) <sup>**</sup> 300 (1) 225	None None	None 75
3	50 p.p.m.	None	None	(2) 300 (1) 225	None None	None 75
3	50 p.p.m.	0.3%	None	(2) 300 (1) 225	None None	None 75
3	50 p.p.m.	0.3%	None	None	(2) 300 (1) 225	None 75
3	50 p.p.m.	0.3%	5 p.p.m. Pb 5 p.p.m. As 100 p.p.m. Rb	None	(2) 300 (1) 225	None 75

<sup>\*\*</sup> Numbers in parenthesis indicate number of animals.

concentrates, as in Experiment II. Concentrate levels were determined by Morrison's Feeding Table modified for first calf heifers. The animals had free access to plain salt blocks and water.

Molybdenum as sodium molybdate, arsenic as sodium arsenate, lead as lead sulfate, and rubidium as rubidium chloride were used for administration to the animals. The amounts of each salt were calculated weekly using the previous week's feed consumption data as the basis for the calculations. For those animals which were fed in a group, estimates of hay consumption were made from data collected before the animals went on experiment. The weekly allowances of salts were weighed on an analytical balance, diluted with cane sugar and then portioned into seven gelatin capsules, one of which was administered daily.

Animals which were housed indoors received a few hours exercise each day in a small lot if weather permitted. Heat periods were recorded, and animals were bred during the first heat occurring after 60 days post-partum.

#### COLLECTION OF DATA ON EXPERIMENTS I, II, AND III

Body weights of all animals in Experiment I, II, and III were recorded immediately following parturition and once each week thereafter while on experiment. At the time of weighing, each animal was given a body condition rating which indicated her state of flesh. The ratings used were as



follows:

Excellent	-	very heavily fleshed
Very good	-	well fleshed
Good plus	-	relatively good flesh
Good	-	average working condition
Fairly good	-	relatively poor flesh
Fair	-	thin condition
Poor	-	very thin condition (emaciated)

The hair condition of each animal was examined weekly and a written description of the distribution, color, and texture was made. The bowel movements for all animals in the three experiments were observed and one of the following descriptions was recorded for each week:

Normal  
Slightly loose  
Loose  
Very loose  
Diarrhea

Bi-weekly hemoglobin determinations for all animals were made during the three experiments. The determinations were made by diluting 0.05 milliliters of heparinized blood to a volume of 10 milliliters with sodium bicarbonate and reading the optical density on a Klett-Summerson photoelectric colorimeter. A chart which was prepared with known standards was then used for converting the optical density readings into hemoglobin values.

Hematocrit values were determined on the blood samples drawn for hemoglobin determinations during the third and fourth periods of Experiment I, and bi-weekly during Experiments II and III. These values were obtained by a method described by Van Allen (154). Heparinized blood was pipetted into hematocrit tubes, and centrifuged in a 10-inch centrifuge

for 1-1/2 hours.

Serum protein percentages were determined three times during the last period of Experiment I and at the start of every 75 days thereafter during Experiments II and III. These values were obtained with the use of a hand protein refractometer. Inorganic phosphorus determinations were made each month on whole blood during the third and fourth periods of Experiments I and II and each month during Experiment III. These determinations were made by a method reported by Powers (130).

During the fourth period of Experiment III, sulfide levels of the rumen contents were determined. The sulfide determinations were carried out by a method reported by Fogo (65) with modifications to permit its use on rumen liquid and solid samples. Rumen samples were drawn by vacuum through a stomach tube into a clean 1000 milliliter flask. The samples then were centrifuged for 15 min. at 3000 r.p.m. to separate the liquid from the solid material. Samples of these fractions were diluted with distilled water and placed in a round bottom flask which had side arms connected through a stopcock. Glass tubing in a tight fitting rubber stopper permitted air to be bubbled through the sample and removed from the top of the flask. The gas removed from the flask in this manner was then bubbled through a basic solution of zinc acetate in a large test tube. A negative pressure, created by an aspirator, in the tube containing the zinc acetate, caused the flow of gases from the round bottom flask to the

receiving test tube. While the gas was flowing in this manner, concentrated hydrochloric acid was added to the sample of rumen material through the side arm of the flask. Any sulfide present in the sample was given off as hydrogen sulfide and collected in the zinc acetate solution. From this point the procedure used for determining hydrogen sulfide followed the method of Fogo (65).

After every seventy-five days on experiment a sample of liver obtained by biopsy, a blood serum sample, and a milk sample was taken from each animal and sent to the U. S. Plant, Soils and Nutrition Laboratory for analysis of copper and molybdenum. A set of samples also was taken at the beginning of Experiments II and III but not at the beginning of Experiment I.

The procedure for obtaining liver biopsies from the cattle was as follows:

1. The hair was clipped from an area over the two most posterior right ribs from the spinal column to a point 12 to 16 inches down the side of the animal.
2. The area was washed with soap and water and tincture of iodine was applied.
3. A small incision was made through the skin between the two most posterior ribs and about 6 inches from the spinal column.
4. An assembled trocar and cannula were forced through the muscle into the right lung cavity causing that lobe to collapse.
5. The trocar and cannula were then pushed through

the animal's diaphragm at which time the trocar was removed from the inside of the cannula.

6. The cannula, which was against the liver, was pushed into this organ cutting a small core.

7. The protruding end of the cannula was covered firmly with the thumb and removed from the animal.

8. A liver sample weighing from 0.1 to 0.8 grams was forced out of the cannula with the trocar onto filter paper.

9. The sample was blotted with filter paper to remove excess blood, placed in a small bottle, and dried in a vacuum oven.

10. The trocar and cannula were washed in redistilled water after each sample was taken and placed in a bath of redistilled ethyl alcohol to await further use.

Blood samples were taken in 50 milliliter centrifuge tubes and permitted to clot. These samples were placed in a centrifuge and spun at 3000 r.p.m. for 15 minutes. The serum was poured off into glass test tubes and frozen. Premilk removed from the udder by hand was collected in a glass jar and stored in the frozen state.

All the bottles and test tubes used for the milk, liver, and serum samples were boiled in nitric acid and rinsed in redistilled water. Both the redistilled water and ethyl alcohol were distilled the second time in a glass still.

The calves born to the four animals from Experiment II which were carried through a second lactation, were sacri-

ficed at birth. The liver, spleen, and a sample of blood serum was saved from these calves and sent to the U. S. Plant, Soil and Nutrition Laboratory in Ithaca, New York, for analysis for copper and molybdenum. Hemoglobin and hematocrit values also were determined on the blood of these calves.

#### EXPERIMENT IV

Six trials using copper-64 were carried out to observe the effects of adding molybdenum, sulfate sulfur, and molybdenum and sulfate sulfur on the copper absorption and metabolism of steers fed a diet low in copper, molybdenum, and sulfur.

The experimental design of these trials is found in Table 4. The animals were placed on the diet for each trial two months prior to the feeding of the radioactive sulfur. The day before feeding the isotope, the animals were placed in wooden metabolism stalls which were designed to collect urine and feces from steers. These stalls were isolated in a part of the barn so as not to be a health hazard when the radioisotope was in use.

Radioactive copper purchased from Union Carbide Corporation, at Oak Ridge, Tennessee, was prepared for administration behind lead bricks in the University of New Hampshire Central Counting Laboratory. In the first four trials 10 millicuries were purchased, and 20 millicuries were purchased for trials V and VI. The isotope was diluted and divided into two equal portions which were poured into two 10 oz. carbonated

TABLE 4

## Design of Radioactive Copper Experiment

Animal number	Age in months	Weight	Sulfate sulfur	Molybdenum
Trial I				
473	6	320	0.3%	50 p.p.m.
474	6	302	0.3%	None
Trial II				
473	8	415	0.3%	None
474	8	400	0.3%	50 p.p.m.
Trial III				
498	5-1/2	300	None	50 p.p.m.
499	5-1/2	330	None	None
Trial IV				
498	8	396	None	None
499	8	425	None	50 p.p.m.
Trial V				
473	16	781	0.3%	None
474	16	751	None	None
Trial VI				
498	13	665	None	None
499	13	707	0.3%	None

beverage bottles. These bottles were then equipped with two milking machine teat cup liners which fit firmly over the neck of the bottle. These liners when forced in the mouth of the animal while its head was held up as far as possible, delivered the solution contained in the bottle into the esophagus. Care was taken to make sure all of the solution was swallowed before the head was released.

Samples of blood, urine and feces were taken frequently for five days. Twenty-four hours after the administration of the isotope a liver biopsy was taken. Attempts to count the isotope with a windowless proportional flow counter model D-47 which was manufactured by Nuclear of Chicago Corporation for the first 24 hours of the first trial failed due to the weak  $\beta$  -radiation. After this period, all samples were counted by use of a gamma well scintillation counter model DS5-5 which was manufactured by Nuclear of Chicago Corporation.

Five milliliter samples of blood and urine were counted for 10 minutes. The samples of feces were wrapped in waxed paper and weighed on an analytical balance. Five minute counts were made on the feces samples because of high levels of the isotope. Liver samples also were wrapped in waxed paper and weighed on an analytical balance. These samples were counted for 20 minutes.

The animals were kept in the collection stalls for a period of 10 days and all feces and urine collected during this time were buried four feet below the surface of the

ground in accordance with Federal Register parts 20 and 30. When the six radioactive studies were completed, all four animals used in the trials were killed and buried according to regulations.



## RESULTS

### EXPERIMENT I

An average of the first three weekly body weights for each cow taken immediately after parturition and immediately after each seventy-five days on experiment are given in Table 4 in the Appendix. From these values, weight changes for each period were calculated. The changes for the cows grouped by treatments are found in Table 5. These data indicate greater weight gains for all experimental groups than for the controls. This indicates that increased molybdenum in the diet did not cause poorer gains as was expected. The greater weight gains shown for the molybdenum supplemented animals were not significant due to the wide variations within each treatment.

Weekly body condition ratings were averaged to give a single rating for each cow during each experimental period. The average ratings are summarized by treatments in Table 6. The differences observed between treatments are not significant although animals receiving 50 p.p.m. of molybdenum showed a lower group average.

The condition of the hair of all animals on all treatments was found to be normal, showing no changes in hair color, distribution, or texture. The weekly observations made on consistency of feces revealed no evidence of looseness or

**TABLE 5**  
**Weight Changes**  
**(Experiment I)**

Animals by treatment	Periods				Totals
	I	II	III	IV	
	Pounds				
<b>Control</b>					
282	-8	52	21	70	135
290	21	35	22	41	119
293	-71	11	20	76	36
<b>Totals</b>	<u>-58</u>	<u>98</u>	<u>63</u>	<u>187</u>	<u>290</u>
				Average	97
<b>Molybdenum 5 p.p.m.</b>					
285	-2	30	51	95	174
288	-37	43	33	62	101
292	22	-4	45	19	82
<b>Totals</b>	<u>17</u>	<u>69</u>	<u>129</u>	<u>176</u>	<u>357</u>
				Average	119
<b>Molybdenum 10 p.p.m.</b>					
283	30	76	19	47	172
287	-37	43	33	62	101
291	6	19	88	67	180
<b>Totals</b>	<u>-1</u>	<u>138</u>	<u>140</u>	<u>176</u>	<u>453</u>
				Average	151
<b>Molybdenum 20 p.p.m.</b>					
281	31	37	-7	55	116
289	39	50	57	79	225
295	34	23	51	82	190
<b>Totals</b>	<u>104</u>	<u>110</u>	<u>101</u>	<u>216</u>	<u>531</u>
				Average	177
<b>Molybdenum 50 p.p.m.</b>					
284	28	31	33	76	168
286	14	41	17	22	94
294	33	38	6	84	161
<b>Totals</b>	<u>75</u>	<u>110</u>	<u>56</u>	<u>182</u>	<u>423</u>
				Average	141

TABLE 6

Body Condition Ratings  
(Experiment I)

Animals by treatment	Periods				Average
	I	II	III	IV	
<b>Control</b>					
282	FG	FG	F	F	FG
290	G	G	G	G	G
293	G+	G+	G+	G+	G+
			Average		G
<b>Molybdenum 5 p.p.m.</b>					
285	G	G	G	G	G
288	FG	FG	FG	FG	FG
292	G	G	G	G	G
			Average		G
<b>Molybdenum 10 p.p.m.</b>					
283	FG	FG	FG	FG	FG
287	FG	FG	G	G	G
291	G	G	G+	G+	G+
			Average		G
<b>Molybdenum 20 p.p.m.</b>					
281	G	G	G+	G+	G+
289	FG	G	G	G	G
295	G	G	G+	G+	G+
			Average		G+
<b>Molybdenum 50 p.p.m.</b>					
284	FG	FG	FG	FG	FG
286	FG	FG	FG	FG	FG
294	G	G	G	G	G
			Average		FG

diarrhea for any animal, regardless of treatment.

Breeding records are given by treatments in Table 7. All but one animal had conceived successfully. The exception was 50 p.p.m. of molybdenum animal which was not pregnant after 5 services. A comparison of the number of days required to settle the other cows on this treatment with comparable data for the other treatments give no indication that the cause of breeding failure in the one cow was due to the treatment.

The average values for hemoglobin, hematocrit, inorganic phosphorus, and serum protein are recorded in Table 8 by treatments. The differences between treatments are not significant. This indicates among other things that molybdenum supplementation did not produce anemia or abnormal phosphorus metabolism.

The effects on appetite of added dietary molybdenum were studied by comparing each animal's total daily feed intake per hundred pounds of body weight. These values, as recorded in Table 9, show that there was greater variation among animals in each treatment than there was among treatments.

The milk and butterfat production records for each cow during each period of the experiment are given in Table 10. As might be expected there were large variations found among the animals on each treatment. The differences among treatments were not significant.

All of the animals in this experiment were considered

TABLE 7

Breeding Records  
(Experiment I)

Animals by treatment	Number of services	Number of days before conception
<b>Control</b>		
282	1	77
290	2	125
293	2	107
Total	<u>5</u>	<u>309</u>
Average	1.7	103
<b>Molybdenum 5 p.p.m.</b>		
285	1	74
288	2	162
292	4	199
Total	<u>7</u>	<u>435</u>
Average	2.3	145
<b>Molybdenum 10 p.p.m.</b>		
283	1	132
287	2	152
291	1	81
Total	<u>4</u>	<u>365</u>
Average	1.3	122
<b>Molybdenum 20 p.p.m.</b>		
281	1	142
287	1	83
295	1	142
Total	<u>3</u>	<u>367</u>
Average	1	122
<b>Molybdenum 50 p.p.m.</b>		
284	2	100
286	5+ (not preg.)	300
294	2	83
Total	<u>9+</u>	<u>483</u>
Average	3+	161

TABLE 8  
Average Composition of Blood  
(Experiment I)

Animals by treatment	Hemoglobin mg/100 ml	Hematocrits %	Phosphorus mg/100 ml	Serum protein %
<b>Control</b>				
282	9.64	30.7	6.9	7.5
290	10.18	33.7	6.1	7.3
293	8.86	30.7	6.0	7.1
<b>Totals</b>	<u>28.68</u>	<u>95.1</u>	<u>19.0</u>	<u>21.9</u>
<b>Average</b>	9.56	31.7	6.3	7.3
<b>Molybdenum 5 p.p.m.</b>				
285	9.70	31.2	6.8	7.5
288	10.06	33.6	7.3	6.6
292	10.27	34.4	6.2	8.0
<b>Totals</b>	<u>30.03</u>	<u>99.2</u>	<u>20.3</u>	<u>22.1</u>
<b>Average</b>	10.01	33.1	6.8	7.4
<b>Molybdenum 10 p.p.m.</b>				
283	9.21	29.9	7.0	8.1
287	10.19	33.8	7.0	8.0
291	9.72	33.6	6.4	7.4
<b>Totals</b>	<u>29.12</u>	<u>97.3</u>	<u>20.4</u>	<u>23.5</u>
<b>Average</b>	9.71	32.4	6.8	7.8
<b>Molybdenum 20 p.p.m.</b>				
281	9.99	32.4	6.2	7.3
289	9.97	30.6	4.8	7.1
295	9.34	29.3	6.6	7.7
<b>Totals</b>	<u>29.30</u>	<u>92.3</u>	<u>17.6</u>	<u>22.1</u>
<b>Average</b>	9.77	30.8	5.9	7.4
<b>Molybdenum 50 p.p.m.</b>				
284	9.90	31.9	6.4	7.5
286	9.16	29.8	8.0	7.7
294	9.33	30.5	6.8	7.8
<b>Totals</b>	<u>28.39</u>	<u>92.2</u>	<u>21.2</u>	<u>23.0</u>
<b>Average</b>	9.46	30.7	7.1	7.7

TABLE 9

Feed Consumed Daily per 100 Pounds Body Weight  
(Experiment I)

Animals by treatment	Periods				Total
	I	II	III	IV	
	Pounds				
<b>Control</b>					
282	3.50	3.52	3.36	2.95	13.33
290	3.18	3.42	3.22	3.20	13.02
293	3.59	3.56	3.24	2.98	13.37
				<b>Total</b>	<b>39.72</b>
				<b>Average</b>	<b>3.31</b>
<b>Molybdenum 5 p.p.m.</b>					
285	3.08	3.04	2.82	2.73	11.67
288	2.99	2.95	2.75	2.67	11.36
292	3.30	3.55	3.49	3.32	13.66
				<b>Total</b>	<b>36.69</b>
				<b>Average</b>	<b>3.06</b>
<b>Molybdenum 10 p.p.m.</b>					
283	3.17	3.29	3.17	3.09	12.72
287	3.56	3.04	3.76	3.37	14.53
291	3.51	3.75	3.51	2.92	13.69
				<b>Total</b>	<b>40.94</b>
				<b>Average</b>	<b>3.41</b>
<b>Molybdenum 20 p.p.m.</b>					
281	3.53	3.49	3.46	3.29	13.77
289	3.22	3.41	3.25	2.92	12.80
295	3.09	3.17	3.01	2.76	12.03
				<b>Total</b>	<b>38.60</b>
				<b>Average</b>	<b>3.22</b>
<b>Molybdenum 50 p.p.m.</b>					
284	3.17	3.59	3.62	3.41	13.79
286	3.23	3.61	3.64	3.42	13.90
294	3.50	3.27	2.87	2.57	12.21
				<b>Total</b>	<b>39.90</b>
				<b>Average</b>	<b>3.33</b>

TABLE 10  
Total Milk and Butterfat Production in Pounds  
(Experiment I)

Animals by treatment	Periods				Total milk	Total fat
	I	II	III	IV		
<b>Control</b>						
282 milk	2502.9	2118.6	1785.8	1524.4	7931.7	
fat	87.4	79.4	77.8	66.9		311.5
290 milk	2168.0	1772.0	1584.6	1627.8	7152.4	
fat	80.8	72.9	65.9	66.3		285.9
293 milk	2369.3	1841.6	1557.0	1098.2	6866.1	
fat	83.3	81.6	71.5	54.9		291.3
			<b>Total</b>		21950.2	888.7
			<b>Average</b>		7316.7	296.2
<b>Molybdenum 5 p.p.m.</b>						
285 milk	1562.6	1399.5	1183.3	985.8	5131.2	
fat	53.5	54.9	48.9	42.9		202.2
288milk	1471.9	1249.3	1114.0	1081.1	4916.3	
fat	43.8	39.6	38.9	42.3		163.6
292 milk	2436.0	2403.3	2235.5	2190.4	9265.2	
fat	81.3	83.2	67.3	72.8		304.6
			<b>Total</b>		19312.7	670.4
			<b>Average</b>		6437.6	223.5
<b>Molybdenum 10 p.p.m.</b>						
283 milk	1426.7	1285.9	1375.5	1022.7	5110.8	
fat	58.4	57.1	62.5	48.6		226.6
287 milk	2526.6	2292.1	2404.3	2231.6	9454.6	
fat	77.3	88.7	104.2	89.8		360.0
291 milk	2195.7	1986.9	1773.2	850.6	6806.4	
fat	72.2	76.4	63.2	35.3		247.1
			<b>Total</b>		21371.8	833.7
			<b>Average</b>		7123.9	277.9



TABLE 10 (Cont.)

Total Milk and Butterfat Production in Pounds  
(Experiment I)

Animals by treatment	Periods				Total milk	Total fat
	I	II	III	IV		
Molybdenum 20 p.p.m.						
281 milk	2909.8	2612.6	2154.7	1963.8	9640.9	411.8
fat	153.7	100.9	82.7	74.5		
289 milk	2106.9	2007.0	1770.7	1556.7	7441.3	272.8
fat	67.0	73.0	68.3	64.5		
295 milk	2405.1	2189.5	1871.7	1441.7	7908.2	237.1
fat	74.7	63.8	55.6	43.0		
			Total Average		24990.4 8330.1	921.7 307.2
Molybdenum 50 p.p.m.						
284 milk	1583.2	1365.9	1261.7	1192.1	5402.9	235.5
fat	62.9	63.2	56.2	53.2		
286 milk	2076.3	1951.9	1768.6	1769.6	7566.4	252.1
fat	61.9	71.5	64.4	54.3		
294 milk	2634.1	2208.8	1795.0	1176.0	7813.0	207.3
fat	86.8	75.5	58.1	49.9		
			Total Average		20783.2 6927.7	694.9 231.6

to have normal health except for mastitis. The incidence of this disease was quite high, but no treatment difference was noted. The reason for the unusually high mastitis infection was not discernible.

The copper analyses of the milk samples taken every 75 days during experiment are recorded in Table 10 of the Appendix. Although the levels of copper in these samples varied from 0.02 to 0.80 p.p.m., no treatment difference was found. All animals showed a significant change in the copper level of the milk as the lactation continued. The values for molybdenum analyses of the 75-day milk samples are recorded in Table 10 of the Appendix. The mean values calculated for each treatment indicate that the level of molybdenum in the milk increased with increased molybdenum in the diet. These values are plotted in Figure 1.

Copper and molybdenum levels of the blood serum are recorded in Table 11 of the Appendix. The mean values for each treatment calculated from these data are plotted in Figures 2 and 3. The mean levels of copper in the serum of all treatments decreased for the first 225 days of experiment. The decreases for the control group, however, was not as large as those for the treatments receiving molybdenum. After 225 days, the levels of serum copper for the controls increased to a mean level of 1.97 p.p.m. which was essentially the same as at the start of the experiment. The serum copper levels for the molybdenum treatments continued to decrease until at the end of 300 days on the experiment all molybdenum

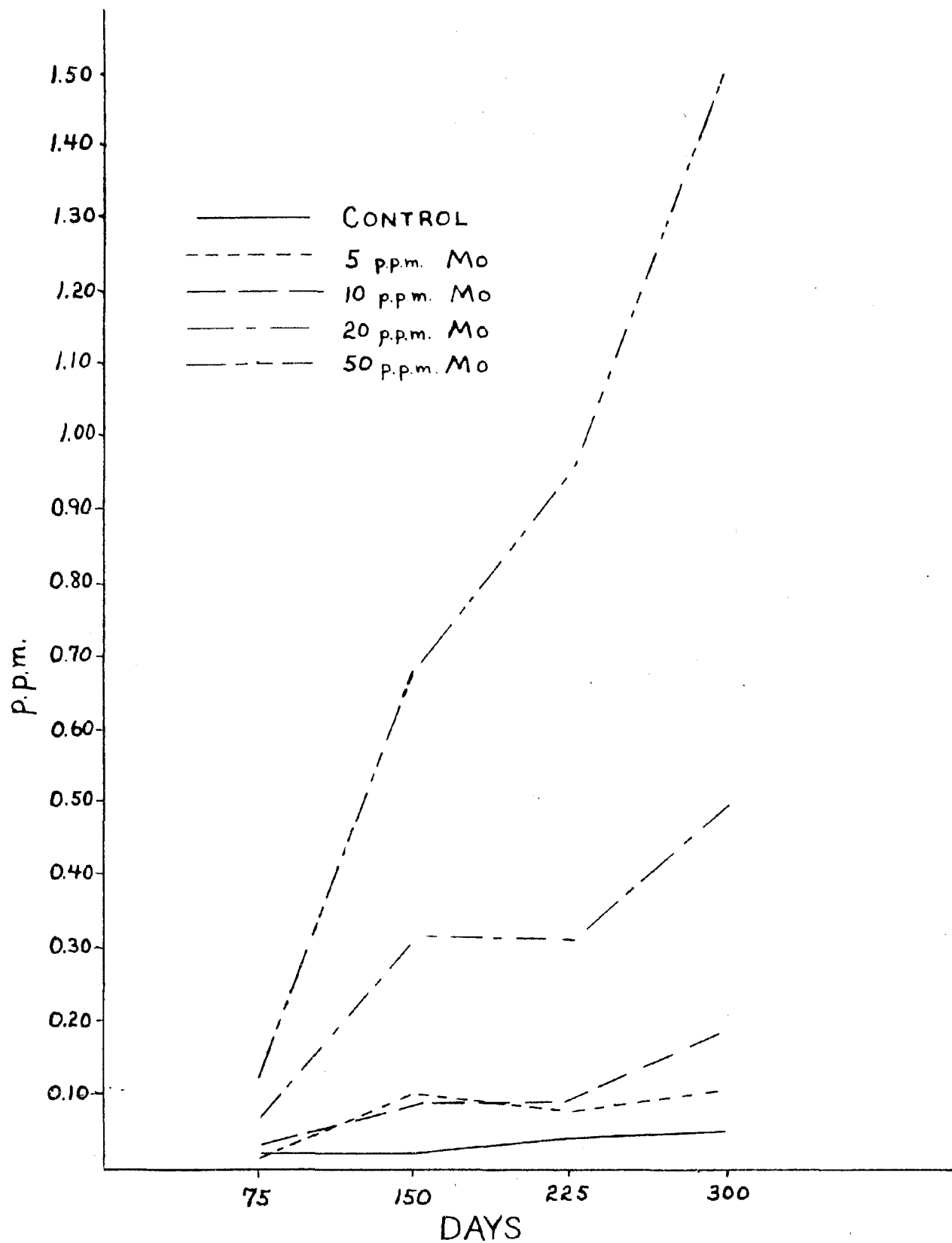


Fig. 1. Milk molybdenum levels for Experiment I.

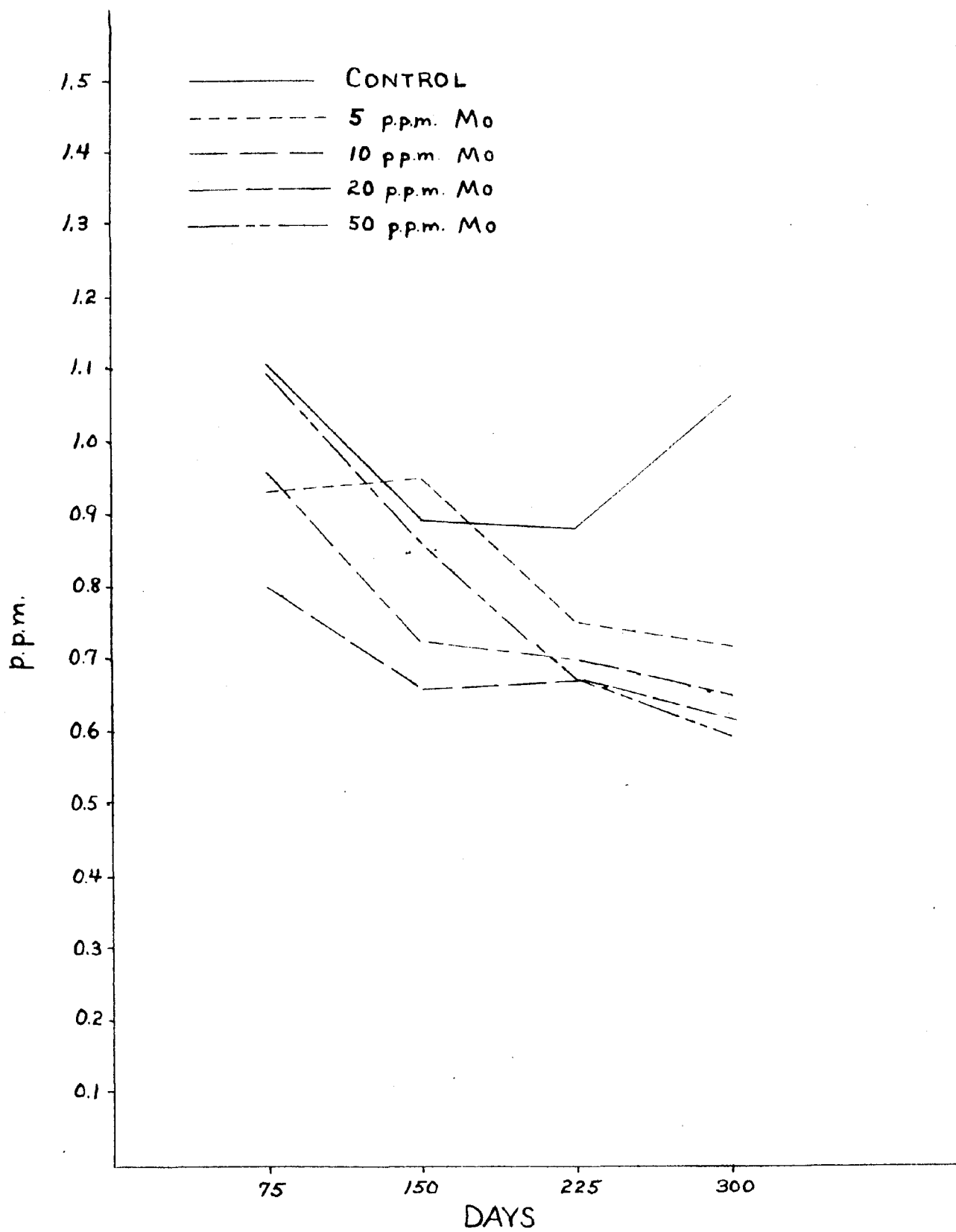


Fig. 2. Serum copper levels for Experiment I.

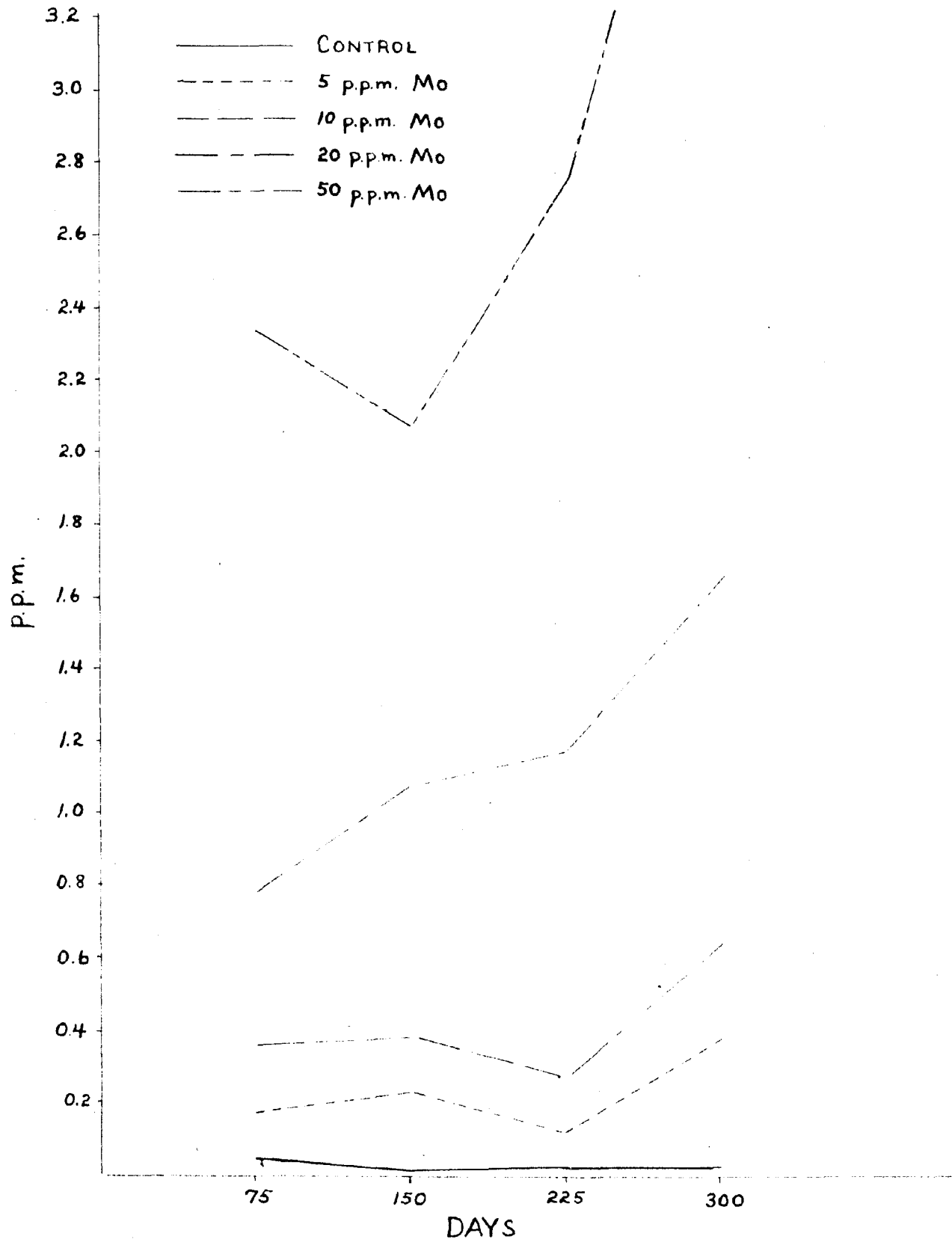


Fig. 3. Serum molybdenum levels for Experiment I.

treatments had serum copper levels from 0.6 to 0.72 p.p.m. The values, however, are still in the published normal range. Treatment means of serum molybdenum values indicate that the level of this element increased with increased molybdenum in the diet. These differences were significant at the 1% level.

Liver copper and molybdenum levels are recorded in Table 12. The mean liver copper values for each treatment are plotted in Figure 4. These values indicate that the liver copper levels for the animals which received molybdenum decreased during the period between 75 and 225 days and then showed an increase during the last 75 days of the experiment. The liver copper levels for the controls increased as the experiment continued. At the end of 300 days on experiment the liver copper levels for the control group were 60 p.p.m. greater than for the other treatments. The mean liver molybdenum values are shown in Figure 5. These data indicate that the molybdenum content of the liver increased with the level of molybdenum in the diet, but at each level of intake the liver seemed to stabilize after 150 days of experiment.

## EXPERIMENT II

Average body weights at the start of the experiment and at the end of each seventy-five days were calculated in the same manner as described for Experiment I. From these values, which are recorded in Table 15 in the Appendix, body weight changes for each period were calculated. A comparison of these weight changes, as found in Table 11, showed no

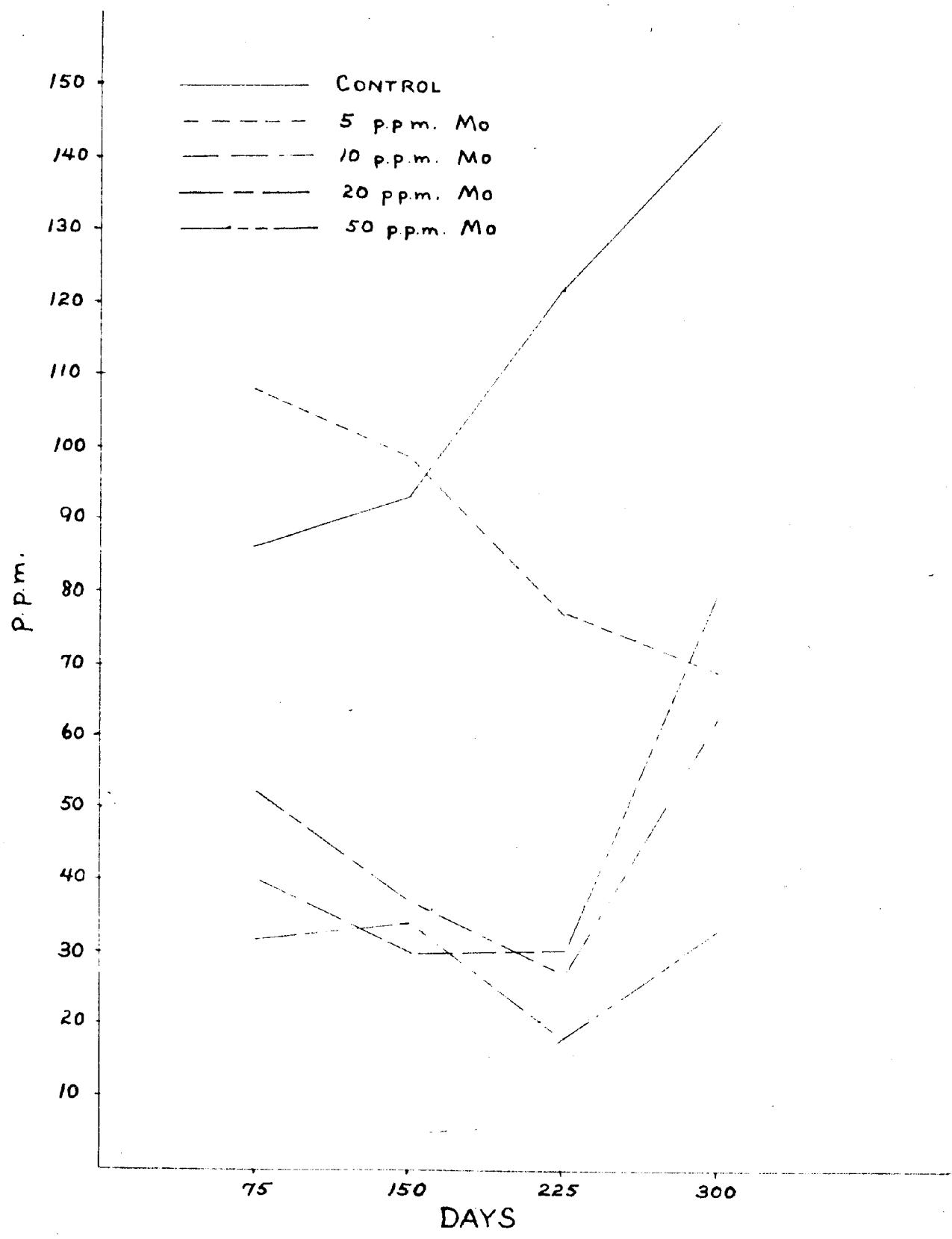


Fig. 4. Liver copper levels for Experiment I.

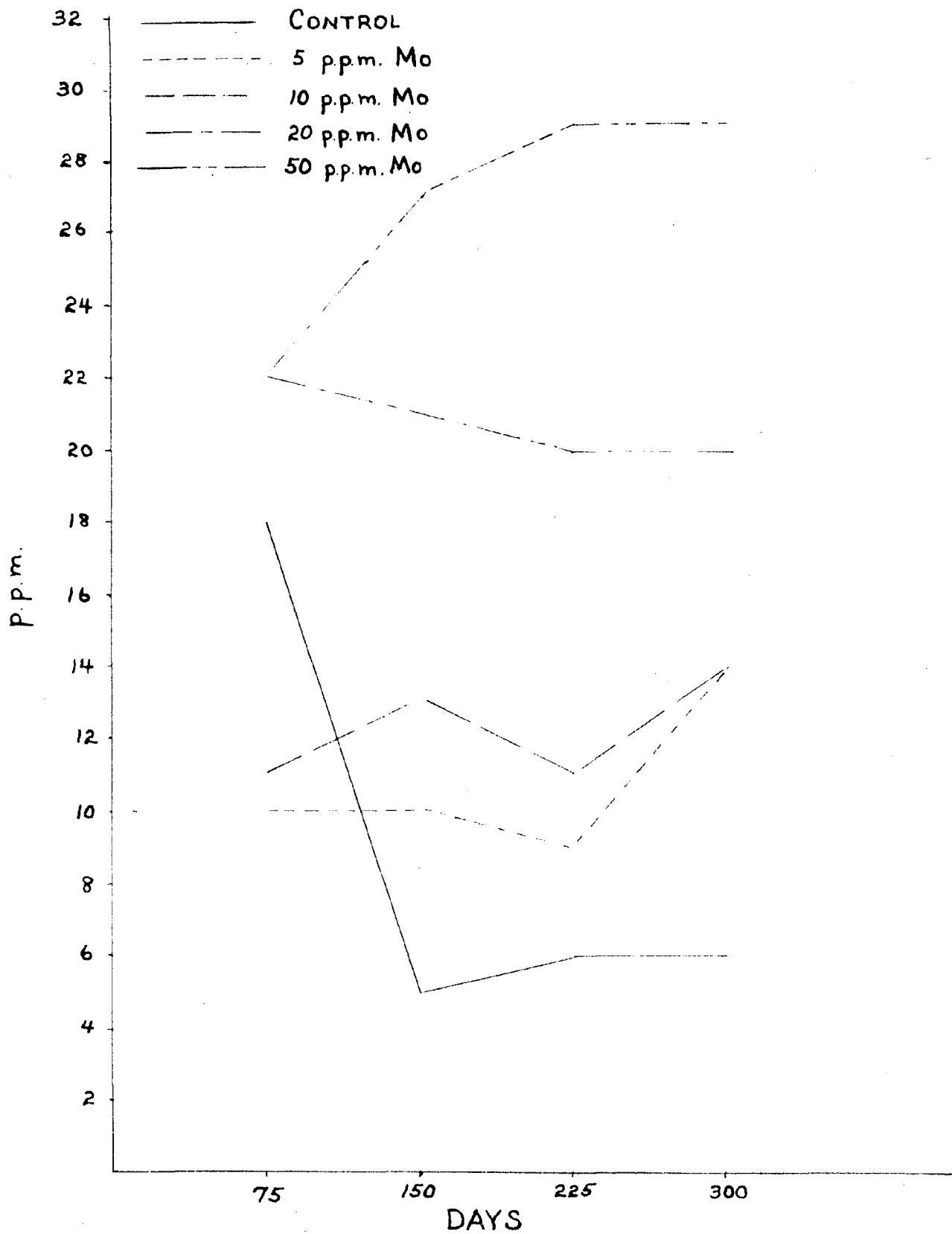


Fig. 5. Liver molybdenum levels for Experiment I.



TABLE 11  
Weight Changes  
(Experiment II)

Animals by treatment	Periods				Totals
	I	II	III	IV	
	Pounds				
<b>Sulfate sulfur 0.3%</b>					
386	3	-2	-24	69	46
395	-29	-5	5	66	37
398	-3	0	-20	-2	-25
Totals	-29	-7	-39	133	58
				Average	19
<b>Sulfate sulfur 0.3%, molybdenum 5 p.p.m.</b>					
390	-3	19			
392	34	-9			
399	-50	10			
Totals	-19	18			
<b>Sulfate sulfur 0.3%, molybdenum 10 p.p.m.</b>					
388	25	6	19		
391	10	-2	7		
396	-8	-32	2		
Totals	27	-28	28		
<b>Sulfate sulfur 0.3%, molybdenum 20 p.p.m.</b>					
389	7	-47	20	-38	-58
393	-3	6	74	76	153
397	31	16	-37	6	16
Totals	34	-25	57	44	111
				Average	37
<b>Sulfate sulfur 0.3%, molybdenum 50 p.p.m.</b>					
387	19	37	3	-3	56
394	38	-9	-11	39	57
400	20	-14	27	45	78
Totals	77	14	19	81	191
				Average	64

TABLE 11 (Cont.)

Weight Changes  
(Experiment II)

Animals by treatment	Periods				Totals
	I	II	III	IV	
	Pounds				
Sulfate sulfur 0.3%, molybdenum 100 p.p.m.					
388				-105	
392				-50	
Totals				-155	Average -78
Sulfate Sulfur 0.3%, molybdenum 200 p.p.m.					
391				-210	
396				-133	
Total				-343	Average -172

Animals Continued Through A Second Lactation

Sulfate sulfur 0.3%					
386	-32	-6	17	139	118
395	-31	20	-43	4	-50
Totals	-63	14	-26	143	68
				Average	34
Sulfate sulfur 0.3%, molybdenum 50 p.p.m.					
387	-5	0	-43	28	-20
394	-3	-4	41	-90	-56
Totals	-8	-4	-2	-62	-76
				Average	-38

effect from feeding molybdenum for 300 days at levels as high as 50 p.p.m. Although two cows which received 50 p.p.m. molybdenum and 0.3% sulfate sulfur showed a weight loss of 38 pounds, this had no significance because one of the two animals which received 0.3% sulfate sulfur showed a weight loss of 50 pounds during the second lactation. The animals which were fed 100 and 200 p.p.m. molybdenum for one period of seventy-five days, however, showed weight losses of 78 and 172 pounds respectively. These exceptionally high weight losses resulted from the feeding of molybdates.

Body condition ratings for each period are summarized in Table 12. Molybdenum levels as high as 50 p.p.m. caused no lowering of body condition, as is shown by these data. Similarly, the two cows retained on the treatment of 50 p.p.m. molybdenum and 0.3% sulfate sulfur for a second lactation showed no loss of body condition during the second lactation. The animals receiving 100 and 200 p.p.m. for seventy-five days, however, showed rapid deterioration of body condition.

A description of hair condition changes noted during this experiment is found in Table 13. No changes in hair color, texture, or distribution were noted for treatments containing 0.3% sulfate sulfur and up to 20 p.p.m. of molybdenum. After 225 days on experiment, animals receiving 0.3% sulfate sulfur and 50 p.p.m. molybdenum developed achromotrichia, particularly around the face and eyes, alopecia around the eyes and muzzle, and the hair texture became soft and lacked body. The two animals receiving this treatment which were continued

TABLE 12

Body Condition Ratings  
(Experiment II)

Animals by treatment	Periods				Average
	I	II	III	IV	
Sulfate sulfur 0.3%					
386	FG	FG	FG	FG	FG
395	G	G	G	G	G
398	FG	FG	FG	FG	FG
			Average		FG
Sulfate sulfur 0.3%, molybdenum 5 p.p.m.					
390	FG	G			
392	G	G+	G+		
399	FG	FG			
Sulfate sulfur 0.3%, molybdenum 10 p.p.m.					
388	G	G+	G+		
391	G	G	G		
396	G	G	G		
Sulfate sulfur, molybdenum 20 p.p.m.					
389	G+	G+	G+	G	G+
393	G	G+	G+	G+	G+
397	FG	G	FG	FG	FG
			Average		G
Sulfate sulfur 0.3%, molybdenum 50 p.p.m.					
387	G	G	G+	G	G
394	G+	G+	G+	G+	G+
400	G	G	FG	FG	FG
			Average		G
Sulfate sulfur 0.3%, molybdenum 100 p.p.m.					
388	FG				
392	G				
Sulfate sulfur 0.3%, molybdenum 200 p.p.m.					
391	F				
396	F				

Animals Continued Through A Second Lactation

Sulfate sulfur 0.3%					
386	G+	G	G	G+	G
395	G+	G	G	G	G
			Average		G
Sulfate sulfur 0.3%, molybdenum 50 p.p.m.					
387	G	G	G	G	G
394	G	G	G	G	G
			Average		G

TABLE 13  
Hair Condition  
(Experiment II)

Animals by treatment	Hair color change	Distribution	Hair texture
Sulfate sulfur 0.3%			
386	None	Normal	Normal
395	"	"	"
398	"	"	"
Sulfate sulfur 0.3%, molybdenum 5 p.p.m.			
390	None	Normal	Normal
392	"	"	"
399	"	"	"
Sulfate sulfur 0.3%, molybdenum 10 p.p.m.			
388	None	Normal	Normal
391	"	"	"
396	"	"	"
Sulfate sulfur 0.3%, molybdenum 20 p.p.m.			
389	None	Normal	Normal
393	"	"	"
397	"	"	"
Sulfate sulfur 0.3%, molybdenum 50 p.p.m.			
387	Black hair turned brown, around eyes & muzzle changed to gray after 250 days	Some alopecia around eyes	Hair very soft and fine
394	Black hair turned brown over most of the body & gray around eyes. White became silver after 225 days	Some alopecia around eyes	Very light and soft
400	Black hair turned brown & gray over most of body after 225 days. Black was completely changed to copper color at 300 days. White became silver	Alopecia around eyes	Hair soft and cotton-like

TABLE 13 (Cont.)

Hair Condition  
(Experiment II)

Animals by treatment	Hair color change	Distribution	Hair texture
Sulfate 388	sulfur 0.3%, molybdenum 100 p.p.m. Slight graying around eyes after 75 days	Normal	Normal
392	Slight graying around eyes after 75 days. White hair interspersed in black	Normal	Normal
Sulfate 391	sulfur 0.3%, molybdenum 200 p.p.m. White hair interspersed in black after 30 days. Gray around eyes	Normal	Normal
396	White & golden hair interspersed in black areas after 40 days. Gray around eyes	Normal	Normal
Animals continued Through A Second Lactation			
Sulfate 386	sulfur 0.3% None	Normal	Normal
395	None	Normal	Normal
Sulfate 387	sulfur 0.3%, molybdenum 50 p.p.m. Achromotrichis of black hair became more complete after parturition	Alopecia severe over face. Some on body	Hair very soft and lacked body
394	Achromotrichia of black hair became more complete after parturition. White hair remained white	Some alopecia around eyes	Hair soft and lacked body

for a second lactation developed these changes more completely during the second lactation. The animals which received 100 and 200 p.p.m. molybdenum developed a condition where white hairs were interspersed throughout the black hair on the body. This change was noted as early as 30 days after the high level treatments were started.

Weekly observations revealed that the feces were higher in water content for cows in this experiment than for Experiment I. These feces were described as being slightly loose to loose, but no differences were noted among treatments. The four animals which received 100 and 200 p.p.m. molybdenum developed severe diarrhea within two weeks from the start of the treatments. The two animals which received the 100 p.p.m. level tended to overcome the diarrhea condition when a decrease in feed consumption lowered appreciably the amount of molybdate administered. The animals on the 200 p.p.m. level, however, continued to have debilitating scours during the entire time of treatment.

The breeding records for this experiment are recorded by treatments in Table 14. The rate of conception was low. Seven animals, representing every treatment, did not conceive at the end of 300 days on experiment. For many of the animals heat periods were not observed by the herdsmen and thus were not bred. Part of this problem may have been caused by limiting the exercise to prevent the spread of brucellosis. It was noted further that heat periods were more frequently missed during the period from December through February and more fre-

TABLE 14

Breeding Records  
(Experiment II)

Animals by treatment	Number of services	Number of days before conception
Sulfate sulfur 0.3%		
386	2	210
395	1	77
398	1 (not preg.)	300
Total	4	587
Average	1.3	196
Sulfate sulfur 0.3%, molybdenum 5 p.p.m.		
390	3	106
392	2	110
399	0 (not preg.)	300
Total	5+	516
Average	1.7	166
Sulfate sulfur 0.3%, molybdenum 10 p.p.m.		
388	2	159
391	4 (not preg.)	300
396	1 (not preg.)	300
Total	7+	759
Average	2.3	253
Sulfate sulfur 0.3%, molybdenum 20 p.p.m.		
389	4+ (not preg.)	300
393	1	76
397	0 (not preg.)	300
Total	5+	676
Average	1.7	225
Sulfate sulfur 0.3%, molybdenum 50 p.p.m.		
387	2	180
394	1	71
400	1 (not preg.)	300
Total	4+	551
Average	1.3	164
Animals Continued Through A Second Lactation		
Sulfate sulfur 0.3%		
386	1	152
395	2	239
Total	3	391
Average	1.5	186
Sulfate sulfur 0.3%, molybdenum 50 p.p.m.		
387	5+ (not preg.)	375
394	4	279
Total	9+	654
Average	4.5	316



quently for those animals kept in the darker areas of the barn.

The average hemoglobin and hematocrit values for each period are found in Tables 15 and 16, respectively. These values showed more variation within each treatment than among treatments thus indicating no difference due to treatment. Because values were all in the normal range, there was no anemia. Values for serum protein determinations are recorded in Table 17. There are no differences noted among treatments for these values.

The effects of added molybdenum on appetite were studied on the basis of total feed consumption as in Experiment I. These values, which are recorded in Table 18 show no differences due to treatment except for animals which received 0.3% sulfate sulfur with 100 and 200 p.p.m. molybdenum. The four animals which were on these treatments went off feed after two weeks and showed poor appetite until the high levels of molybdate were removed.

The milk and butterfat production values for the animals on this experiment are shown in Table 19. There appears to be no differences due to treatment. Production levels varied greatly in all treatments. Such variations should be expected under the conditions by which the animals were selected for the experiment. The feeding of 100 and 200 p.p.m. molybdenum with 0.3% sulfate sulfur, however, produced an abrupt drop in milk production when toxicity symptoms were observed. Evidence that the high molybdenum levels

TABLE 15

Average Hemoglobin Values  
(Experiment II)

Animals by treatment	Periods				Totals
	I	II	III	IV	
	mg/100 ml of blood				
<b>Sulfate sulfur 0.3%</b>					
386	9.87	9.49	9.31	9.29	37.96
395	9.88	9.30	8.46	8.44	36.08
398	9.26	8.46	8.59	8.45	34.76
				Total	108.80
				Average	9.07
<b>Sulfate sulfur 0.3%, molybdenum 5 p.p.m.</b>					
390	10.19	9.37			19.56
392	10.38	9.81	9.73		29.92
399	9.78	9.41			19.19
				Total	68.67
				Average	9.81
<b>Sulfate sulfur 0.3%, molybdenum 10 p.p.m.</b>					
388	9.38	9.21	9.24		27.83
391	10.46	8.74	9.77		28.97
396	9.60	9.21	8.87		27.68
				Total	84.48
				Average	9.39
<b>Sulfate sulfur 0.3%, molybdenum 20 p.p.m.</b>					
389	10.10	9.87	9.91	10.07	39.95
393	10.53	9.53	10.09	9.10	39.25
397	9.21	8.53	8.18	8.18	34.10
				Total	113.30
				Average	9.44
<b>Sulfate sulfur 0.3%, molybdenum 50 p.p.m.</b>					
387	10.32	9.94	9.57	9.25	39.08
394	9.47	8.50	8.37	8.30	34.64
400	10.26	9.35	8.92	8.99	37.52
				Total	111.24
				Average	9.27
<b>Sulfate sulfur 0.3%, molybdenum 100 p.p.m.</b>					
388				9.00	
392				8.95	
				Total	17.95
				Average	8.96
<b>Sulfate sulfur 0.3%, molybdenum 200 p.p.m.</b>					
391				9.06	
396				8.44	
				Total	17.50
				Average	8.75

TABLE 15 (Cont.)

Average Hemoglobin Values  
(Experiment II)

Animals by treatment	Periods				Totals
	I	II	III	IV	
	mg/100 ml of blood				

Animals Continued Through A Second Lactation

Sulfate sulfur 0.3%

386	9.89	9.50	9.89	10.24	39.52
395	9.41	8.80	9.75	9.45	37.41
				Total	76.93
				Average	9.62

Sulfate sulfur 0.3%, molybdenum 50 p.p.m.

387	10.88	9.98	10.59	9.62	41.07
394	8.85	8.82	9.49	9.21	36.37
				Total	77.44
				Average	9.68

TABLE 16

Average Hematocrit Values  
(Experiment II)

Animals by treatment	Periods				Totals
	I	II	III	IV	
<b>Sulfate sulfur 0.3%</b>					
386	30.6	29.5	29.5	29.4	119.0
395	31.7	29.9	26.7	26.8	115.1
398	28.1	27.2	26.1	26.0	107.4
				Total	<u>341.5</u>
				Average	28.5
<b>Sulfate sulfur 0.3%, molybdenum 5 p.p.m.</b>					
390	33.0	32.1			65.1
392	34.0	32.6	33.4		100.0
399	32.4	31.5			63.9
				Total	<u>229.0</u>
				Average	32.7
<b>Sulfate sulfur 0.3%, molybdenum 10 p.p.m.</b>					
388	30.6	29.2	31.5		91.3
391	33.2	29.5	32.4		95.1
396	30.0	29.2	28.2		87.4
				Total	<u>273.8</u>
				Average	30.4
<b>Sulfate sulfur 0.3%, molybdenum 20 p.p.m.</b>					
389	32.2	31.8	32.8	32.0	128.8
393	33.2	31.8	34.1	30.2	129.3
397	29.6	26.5	27.0	23.8	106.9
				Total	<u>365.0</u>
				Average	30.4
<b>Sulfate sulfur 0.3%, molybdenum 50 p.p.m.</b>					
387	33.0	32.5	31.2	28.8	125.5
394	29.5	27.0	27.5	26.2	110.2
400	32.3	30.7	29.3	29.1	121.4
				Total	<u>357.1</u>
				Average	29.8
<b>Sulfate sulfur, 0.3%, molybdenum 100 p.p.m.</b>					
388				27.4	
392				30.8	
				Total	58.2
				Average	29.1
<b>Sulfate sulfur, 0.3%, molybdenum 200 p.p.m.</b>					
391				29.1	
396				25.5	
				Total	54.6
				Average	27.3

TABLE 16 (Cont.)

Average Hematocrit Values  
(Experiment II)

Animals by treatment	Periods				Totals
	I	II	III	IV	

## Animals continued Through A Second Lactation

## Sulfate sulfur 0.3%

386	29.5	29.3	30.3	31.8	120.9
395	30.0	30.3	30.0	29.6	119.9
				Total	240.8
				Average	30.10

## Sulfate sulfur 0.3%, molybdenum 50 p.p.m.

387	32.3	31.7	33.6	29.5	127.1
394	27.5	27.0	29.1	28.8	112.4
				Total	239.5
				Average	29.94

TABLE 17  
Serum Protein Values  
(Experiment II)

Animals by treatment	Days					Total
	0	75	150	225	300	
Sulfate sulfur 0.3%						
386	7.8	8.6	8.0	8.3	7.7	40.4
395	7.4	8.6	7.2	7.2	7.0	37.4
398	7.5	8.1	6.6	7.2	7.4	36.8
				Total		114.6
				Average		7.6
Sulfate sulfur 0.3%, molybdenum 5 p.p.m.						
390	6.7	7.1	7.9			21.7
392	6.6	7.8	7.5	7.0		28.9
399	7.3	8.2	7.4			22.9
				Total		73.5
				Average		7.4
Sulfate sulfur 0.3%, molybdenum 10 p.p.m.						
388	8.1	7.7	7.7	7.3		30.8
391	7.4	8.8	7.8	8.0		32.0
396	7.0	7.3	7.6	8.0		29.9
				Total		92.7
				Average		7.7
Sulfate sulfur 0.3%, molybdenum 20 p.p.m.						
389	6.0	6.2	7.4	7.0	6.7	33.3
393	6.4	7.0	7.7	7.3	7.0	35.4
397	7.0	7.4	7.8	8.0	7.3	37.5
				Total		106.2
				Average		7.1
Sulfate sulfur 0.3%, molybdenum 50 p.p.m.						
387	7.0	8.1	8.0	7.0	6.8	36.9
394	6.0	7.6	7.5	7.7	7.5	36.3
400	7.8	8.0	7.6	7.8	7.9	39.1
				Total		112.3
				Average		7.5

TABLE 17 (Cont.)

Serum Protein Values  
(Experiment II)

Animals by treatment	Days					Total
	0	75	150	225	300	
Sulfate sulfur 0.3%, molybdenum 100 p.p.m.						
388						7.0
392						6.8
					Total	13.8
					Average	6.9
Sulfate sulfur 0.3%, molybdenum 200 p.p.m.						
391						6.4
396						7.3
					Total	13.7
					Average	6.9

Animals Continued Through A Second Lactation

Sulfate sulfur 0.3%						
386	8.2	8.6	8.1	8.5	8.3	41.7
395	7.1	8.2	7.7	8.0	7.8	38.8
					Total	80.5
					Average	8.05
Sulfate sulfur 0.3%, molybdenum 50 p.p.m.						
387	7.0	7.9	7.9	8.1	7.2	38.1
394	7.6	7.4	7.4	8.0	9.0	39.4
					Total	77.5
					Average	7.75

TABLE 18

Feed Consumed Daily per 100 Pounds Body Weight  
(Experiment II)

Animals by treatment	Periods				Total
	I	II	III	IV	
	Pounds				
<b>Sulfate sulfur 0.3%</b>					
386	3.64	4.01	3.73	3.47	14.85
395	3.68	3.89	3.53	3.09	14.19
398	3.39	3.05	2.80	2.61	11.85
				Total	40.89
				Average	3.41
<b>Sulfate sulfur 0.3%, molybdenum 5 p.p.m.</b>					
390	4.07	3.79			7.86
392	3.51	3.77	3.57		10.85
399	3.84	3.44			7.28
				Total	25.99
				Average	3.71
<b>Sulfate sulfur 0.3%, molybdenum 10 p.p.m.</b>					
388	3.55	3.33	3.29		10.17
391	3.55	3.45	3.08		10.08
396	4.01	3.48	2.69		10.18
				Total	30.43
				Average	3.38
<b>Sulfate sulfur 0.3%, molybdenum 20 p.p.m.</b>					
389	3.65	3.87	3.47	3.09	14.08
393	3.77	3.65	3.28	2.77	13.47
397	3.25	3.05	2.68	2.29	11.27
				Total	38.82
				Average	3.24
<b>Sulfate sulfur 0.3%, molybdenum 50 p.p.m.</b>					
387	3.37	3.05	2.81	2.48	11.71
394	3.39	3.12	2.83	2.89	12.23
400	4.40	4.11	3.11	3.27	14.89
				Total	38.83
				Average	3.24
<b>Sulfate sulfur 0.3%, molybdenum 100 p.p.m.</b>					
388				2.39	
392				2.47	
				Total	4.86
				Average	2.43
<b>Sulfate sulfur 0.3%, molybdenum 200 p.p.m.</b>					
391				1.87	
396				2.47	
				Total	4.34
				Average	2.17



TABLE 19

Total Milk and Butterfat Production in Pounds  
(Experiment II)

Animals by treatment	Periods				Total milk	Total fat
	I	II	III	IV		
Sulfate sulfur 0.3%						
386 milk	2147.1	2747.4	1671.8	1480.4	8046.7	243.2
fat	65.0	65.9	60.6	51.7		
395 milk	2842.0	2476.4	1866.5	1221.3	8406.2	357.8
fat	115.9	107.4	73.9	60.6		
398 milk	1787.7	1365.5	1123.6	795.4	5072.2	199.0
fat	77.8	53.1	42.2	25.9		
Total					21325.1	800.0
Average					7175.0	266.6
Sulfate sulfur 0.3%, molybdenum 5 p.p.m.						
390 milk	1856.2	1457.1				
fat	76.7	59.6				
392 milk	2356.3	1843.1	1479.1			
fat	77.6	62.9	50.7			
399 milk	2811.0	2052.9				
fat	128.5	76.3				
Sulfate sulfur 0.3%, molybdenum 10 p.p.m.						
388 milk	1703.7	1429.0	1412.0			
fat	54.8	47.9	48.3			
391 milk	2038.6	1695.7	1526.4			
fat	76.76	65.0	53.5			
396 milk	2778.0	2181.0	1683.2			
fat	105.6	85.3	71.55			
Sulfate sulfur 0.3%, molybdenum 20 p.p.m.						
389 milk	2361.5	1945.1	1552.8	1090.4	6949.8	252.5
fat	81.2	69.0	59.7	42.6		
393 milk	1970.4	1667.0	1221.3	310.9	5169.6	170.3
fat	62.3	55.7	40.0	12.3		
397 milk	2268.3	1702.8	1210.9	1025.6	6207.6	193.0
fat	64.5	55.2	30.9	34.4		
Total					18327.0	615.8
Average					6109.0	205.3

TABLE 19 (Cont.)

Total Milk and Butterfat Production in Pounds  
(Experiment II)

Animals by treatment	Periods				Total milk	Total fat
	I	II	III	IV		
Sulfate sulfur 0.3%, molybdenum 50 p.p.m.						
387 milk	1763.4	1169.0	657.2	150.1	3739.7	
fat	55.2	39.4	25.8	12.9		133.3
394 milk	2766.3	2276.0	1811.3	1239.2	8092.8	
fat	92.5	85.9	60.1	46.5		285.0
400 milk	3494.2	2691.6	1801.4	1349.9	9337.1	
fat	107.7	78.2	54.0	37.8		277.1
			Total		21169.6	695.4
			Average		7056.6	231.8
Sulfate sulfur 0.3%, molybdenum 100 p.p.m.						
388 milk				651.5		
fat				24.1		
392 milk				474.0		
fat				19.4		
Sulfate sulfur 0.3%, molybdenum 200 p.p.m.						
391 milk				657.9		
fat				25.2		
396 milk				591.7		
fat				28.5		
Animals Continued Through A Second Lactation						
Sulfate sulfur 0.3%						
386 milk	3364.7	3217.9	2712.0	2209.8	11504.4	
fat	110.4	99.1	92.9	86.2		388.6
395 milk	3165.1	2308.6	2239.0	2199.5	9912.3	
fat	130.5	140.6	104.8	93.4		469.3
			Total		21416.7	855.9
			Average		10708.4	428.0
Sulfate sulfur 0.3%, molybdenum 50 p.p.m.						
387 milk	2681.7	1265.2	247.1		4194.0	
fat	95.8	58.4	13.6			167.8
394 milk	2716.9	2553.3	1747.3	1467.9	8485.4	
fat	97.6	93.0	70.9	62.5		324.0
			Total		12679.4	491.8
			Average		6339.7	245.9

caused this drop was provided by the recovery of animals when removed from these treatments. Although these animals were producing less than 2 pounds of milk per day, production levels returned to 16 and 18 pounds when the molybdenum feeding was discontinued.

The health of all the animals receiving levels of not over 50 p.p.m. molybdenum was considered normal, except for brucellosis and mastitis. The occurrence of brucellosis in two animals cannot be considered to have had any connection with the experimental treatment the animals received. An unusually high incidence of mastitis was not confined to any particular treatment and no adequate explanation could be found for the occurrence of the disease. Animals receiving 100 and 200 p.p.m. molybdenum with 0.3% sulfate sulfur exhibited a very toxic condition after three weeks on experiment. These animals became emaciated and appeared lethargic. On several occasions they lost nervous control of their rear quarters and help was required to move them to the scales for weighing. When the animals showed these conditions, molybdenum feeding was suspended for one week. Upon resumption of the treatment it appeared the animals adjusted somewhat to these high levels and nervous symptoms did not recur.

The health of the animals continued through a second lactation on a level of 50 p.p.m. molybdenum and 0.3% sulfate sulfur was not shown to be impaired. Except for changes in hair condition the two animals performed comparably to the two animals which received 0.3% sulfate sulfur. All four

animals showed no problems during parturition and gave birth to normal size calves. These calves appeared normal in every respect except that both calves from the 50 p.p.m. molybdenum animals had brown and white coats whereas the calves of the control animals had normal black and white coats.

Table 20

Composition of Serum, Liver, and Spleen of Calves  
Born to Cows Continued Through a Second Lactation

Calf by dam's treatment	Copper content			Molybdenum content		
	Serum	p.p.m. Liver	Spleen	Serum	p.p.m. Liver	Spleen
Sulfate sulfur 0.3%						
Calf of 386	0.30	118	0.60	0.05	0.0	0.0
Calf of 395	0.16	127	1.4	0.00	0.0	0.0
Sulfate sulfur 0.3% and 50 p.p.m. molybdenum						
Calf of 387	0.00	1.9	0.91	0.13	0.0	0.0
Calf of 394	0.05	1.70	0.83	0.46	0.0	0.0

The copper and molybdenum analyses of serum, liver, and spleen taken from these calves are recorded in Table 20. The serum molybdenum values for these calves indicate that a small amount of molybdenum was transferred across the fetal membranes in the cows receiving molybdenum. Levels of molybdenum in the liver and spleen of these calves were undetectable.

The copper level in the serum and liver of the calves born to the cows which received 50 p.p.m. molybdenum with 0.3% sulfate sulfur indicated these calves were extremely copper deficient. Although the serum copper levels of the two calves from the cows which received sulfate were below normal, these

calves had considerable higher levels than the calves born to the cows which received molybdenum and sulfate.

The values for the copper and molybdenum analyses of milk samples for this experiment are recorded in Table 20 of the Appendix. Milk copper levels were not affected by the level of molybdenum in the diet but were significantly lower in this experiment than in Experiment I. This difference between Experiments I and II indicated that sulfate had an effect of lowering the level of copper in the milk. The mean milk molybdenum levels are plotted in Figure 6 by treatments. In contrast to Experiment I, the molybdenum levels of the milk in this experiment showed only slight increases with increased dietary molybdenum. These data indicate that the addition of 0.3% sulfate sulfur to the diet decreased the molybdenum excretion in the milk.

Serum molybdenum and copper levels for Experiment II are recorded in Table 21 of the Appendix. The mean serum copper values are plotted in Figure 7. Animals receiving 5, 10, and 20 p.p.m. of molybdenum with 0.3% sulfate sulfur added to the diet showed similar results to the 5, 10, and 20 p.p.m. of molybdenum treatments in Experiment I. For these treatments the serum copper decreased as the experiment progressed. Animals which received 50 p.p.m. of molybdenum with 0.3% sulfate sulfur had an increase in serum copper for the first 150 days of experiment and then showed a sharp decrease during the last 150 days of experiment. The animals on the 0.3% sulfate sulfur treatment showed only slight changes in serum copper

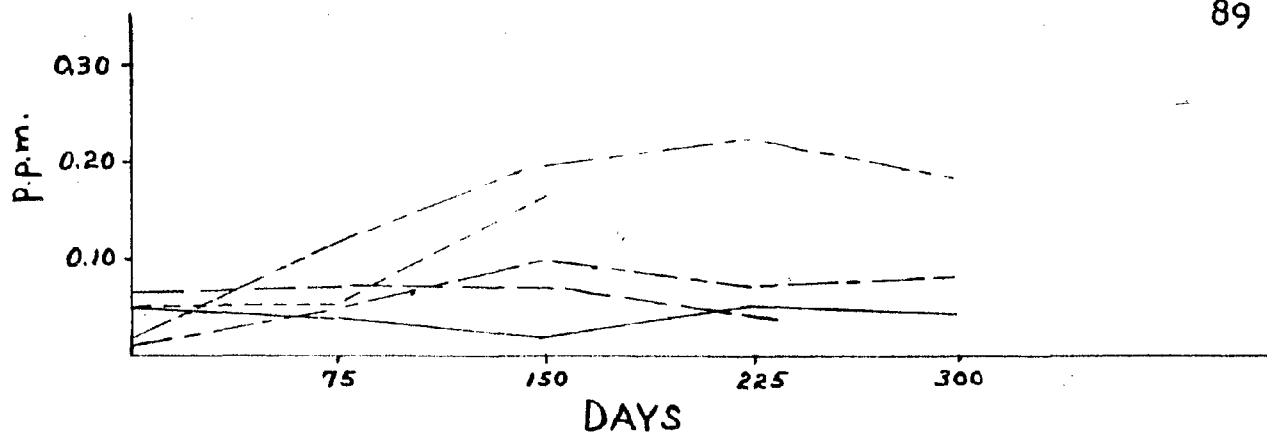


Fig. 6. Milk molybdenum levels for Experiment II.

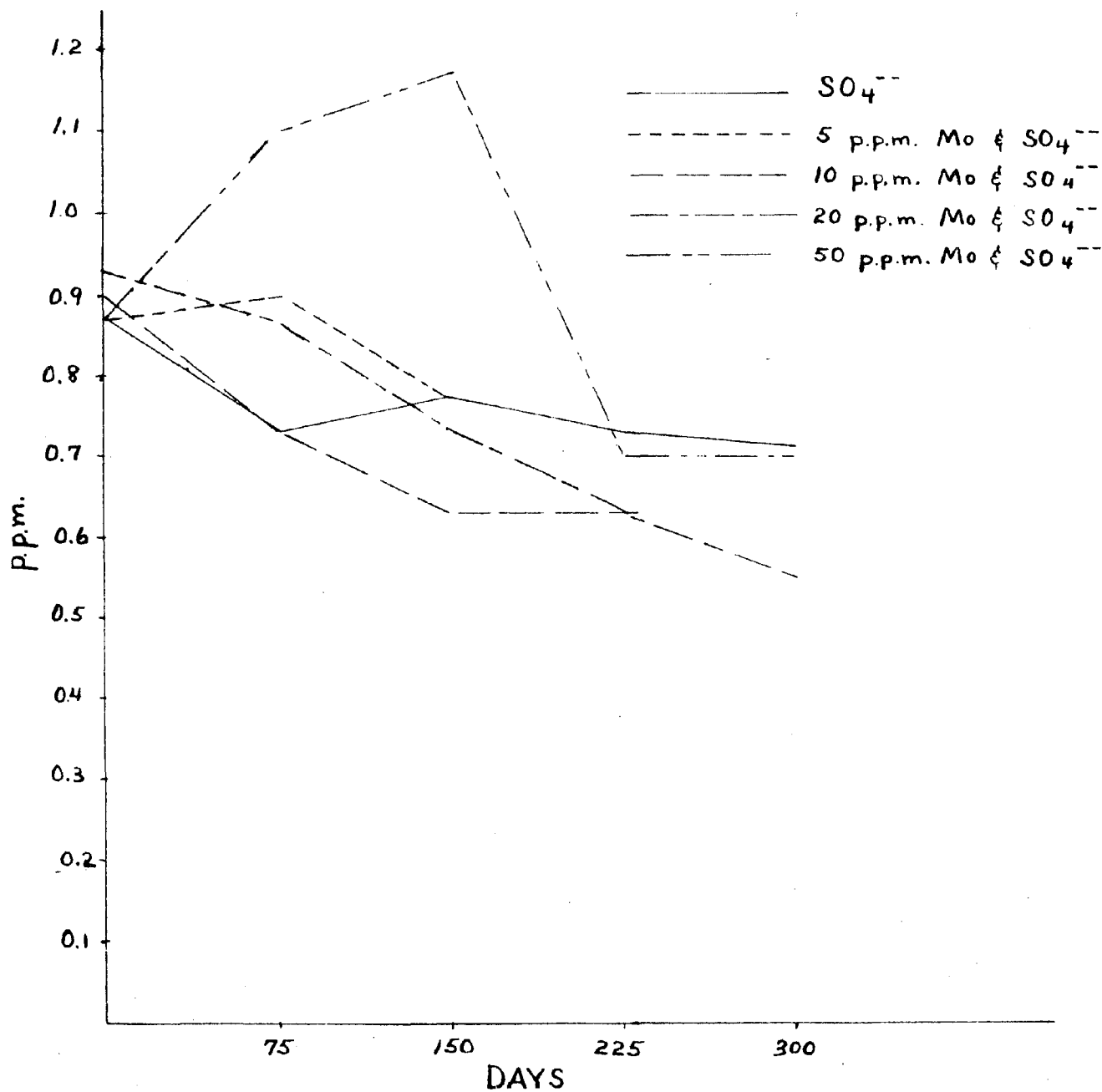


Fig. 7. Serum Copper levels for Experiment II

as the experiment progressed and did not differ significantly from the other treatments at the end of the experiment. The mean serum molybdenum values are plotted in Figure 8. These values increased with increased molybdenum in the diet. These data show, however, that the response to added dietary molybdenum was much less in this experiment than for Experiment I. This would indicate that the addition of sulfate sulfur diminished this response.

Liver copper and molybdenum levels for Experiment II are recorded in Table 22 of the Appendix. The mean liver copper values, which are plotted in Figure 9, show a rapid decrease during the first 225 days of experiment for all molybdenum treatments. After 225 days these values remained at this low level in contrast to Experiment I where these treatments showed an increase. The animals receiving 0.3% sulfate sulfur, without added molybdenum which had a lower value at the start of the experiment, showed a general increase in liver copper as the experiment progressed. The mean liver molybdenum values are plotted in Figure 10. In contrast to Experiment I, the liver molybdenum levels did not show consistent trends. It appears that the addition of 0.3% sulfate sulfur to the diet had an effect on the metabolism of molybdenum and that it was of a heterogeneous nature.

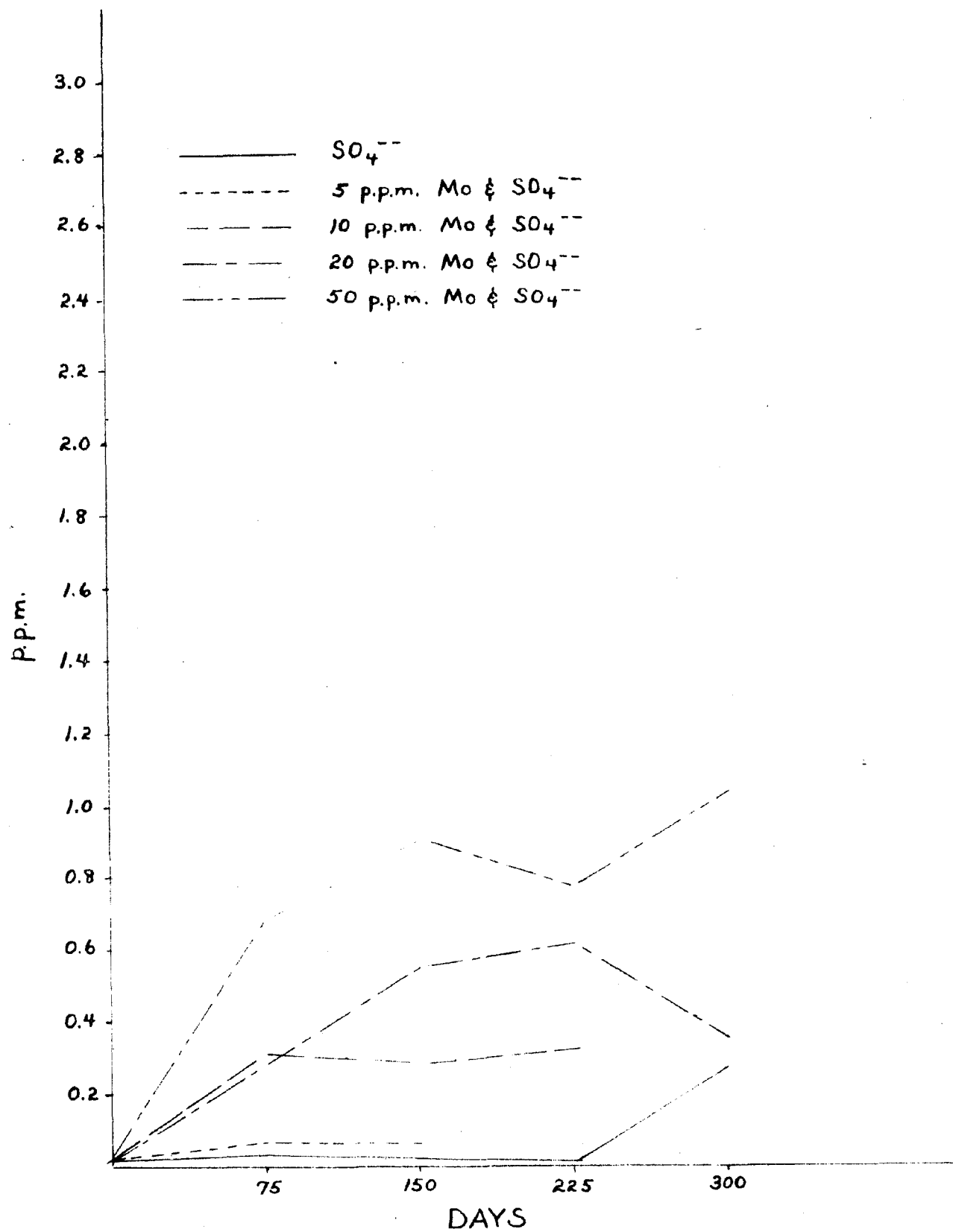


Fig. 8. Serum molybdenum levels for Experiment II.



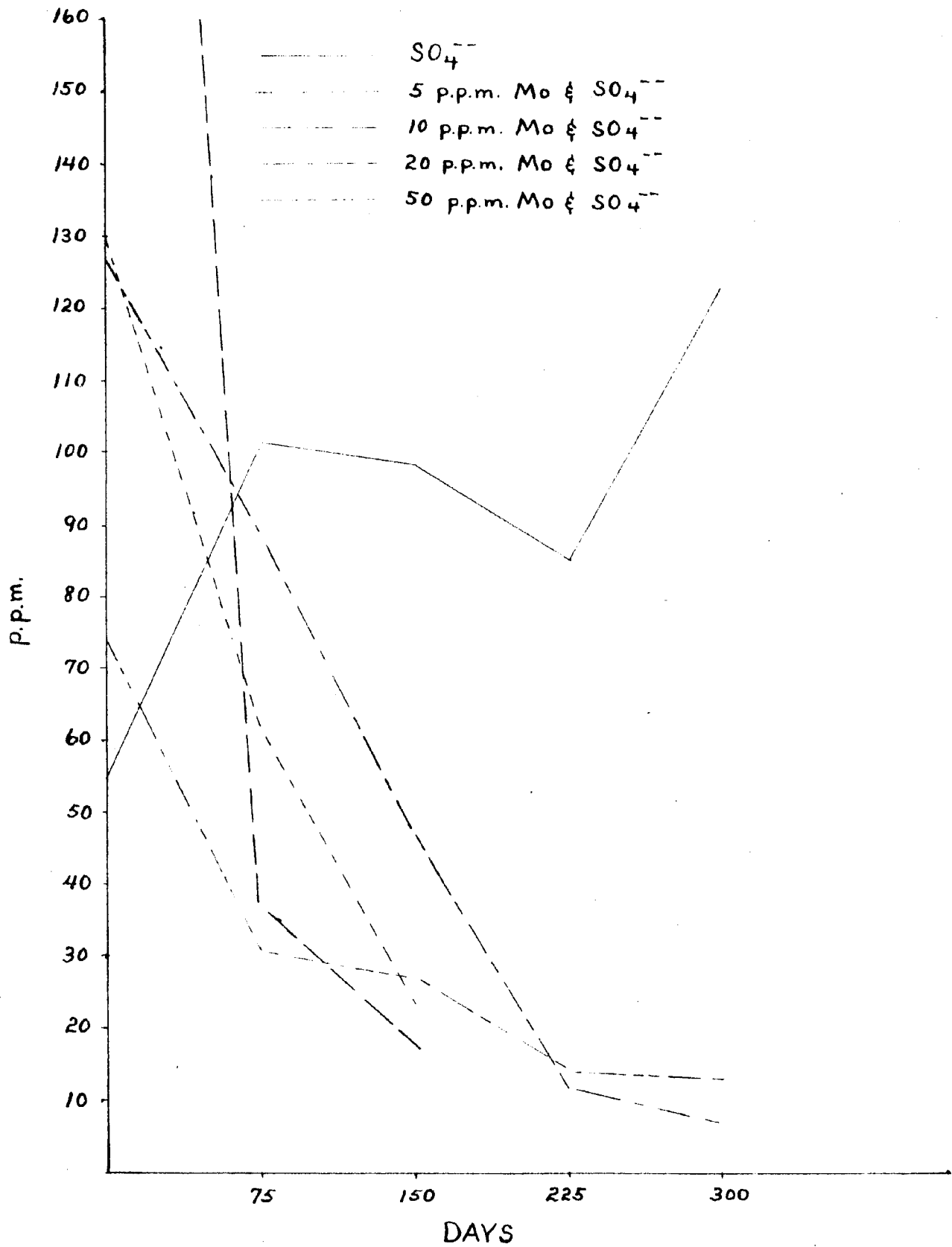


Fig. 9. Liver copper levels for Experiment II.

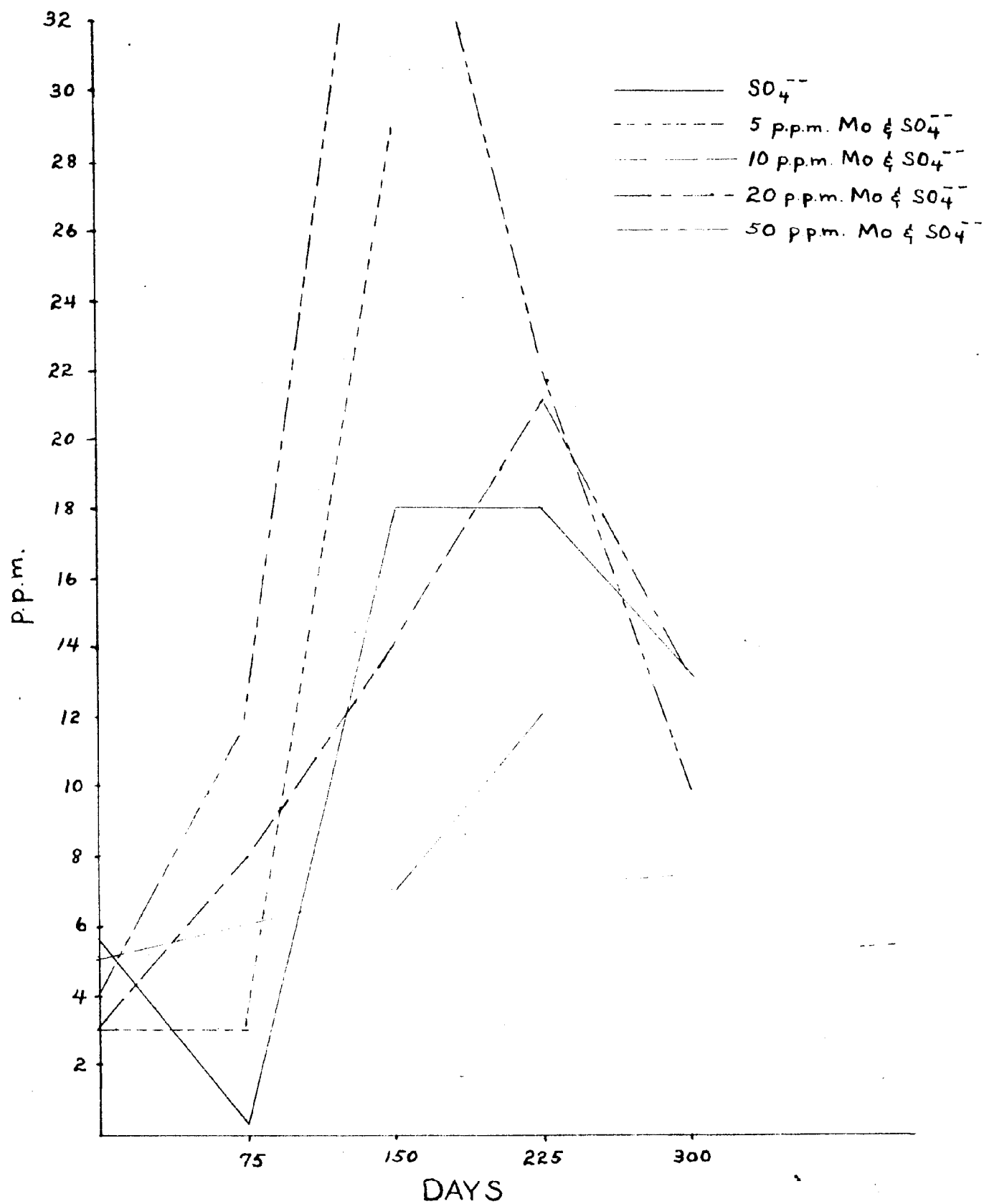


Fig. 10. Liver molybdenum levels for Experiment II.

## EXPERIMENT III

Body weights at the start of the experiment and at the end of each seventy-five day period were calculated in the same manner as described for Experiments I and II. These values are recorded in Table 25 in the Appendix. A summary of weight changes calculated from these values are recorded by treatments in Table 21. The average weight gains for the animals which received the treatments of 0.3% sulfate sulfur, outdoors, and 50 p.p.m. molybdenum, outdoors, were 148 pounds and 105 pounds, respectively. These average weight gains were considerably higher than the average gain of 11 pounds for the animals receiving the treatment of 50 p.p.m. molybdenum and 0.3% sulfate sulfur, outdoors. Both the treatment receiving 50 p.p.m. molybdenum and 0.3% sulfate sulfur, indoors, and the treatment receiving 50 p.p.m. molybdenum, 5 p.p.m. arsenic, 5 p.p.m. lead, 100 p.p.m. rubidium and 0.3% sulfate sulfur indoors, showed average weight losses during the experiment. The difference of the average weight loss for these treatments was 100 pounds which would indicate there was no effect on body weight from adding arsenic, lead, and rubidium to a diet high in molybdenum and sulfate. The difference found between the treatments outdoors and indoors are very large and appear to result from the difference of treatment.

Body condition ratings for each animal by periods are recorded in Table 22. Animals in the treatments 50 p.p.m.

TABLE 21

Weight Changes  
(Experiment III)

Animals by treatment	Periods				Totals
	I	II	III	IV	
	Pounds				
<b>Sulfate sulfur 0.3% (outdoors)</b>					
477	54	-77	71	35	83
482	45	64	77	29	215
488	15	33	57	42	147
Totals	114	20	205	106	445
				Average	148.3
<b>Molybdenum 50 p.p.m. (outdoors)</b>					
479	-3	14	32	67	110
481	7	66	12	50	135
489	-20	21	21	47	69
Totals	-16	101	65	164	314
				Average	104.7
<b>Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (outdoors)</b>					
480	15	12	9	12	48
483	-80	-28	51	24	-33
490	-38	-7	-20	82	17
Totals	-103	-23	40	118	32
				Average	10.7
<b>Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (indoors)</b>					
476	21	13	32	-29	37
485	14	-8	-7	-17	-18
487	0	-20	6	-97	-111
Totals	35	-15	31	-143	-92
				Average	-30.7
<b>Sulfate sulfur 0.3%, molybdenum 50 p.p.m., arsenic 5 p.p.m., rubidium 100 p.p.m., and lead 5 p.p.m. (indoors)</b>					
478	10	67	-101	62	38
484	-3	-39	-34	21	-55
486	2	-15	-3	-31	-47
Totals	9	13	-138	52	-64
				Average	-21.3

TABLE 22

Body Condition Ratings  
(Experiment III)

Animals by treatment	Periods				Average
	I	II	III	IV	
<b>Sulfate sulfur 0.3% (outdoors)</b>					
477	G	G+	G	G+	G+
482	G	G	G+	G+	G
488	G	G	G	G	G
	Treatment average				G
<b>Molybdenum 50 p.p.m. (outdoors)</b>					
479	G	G+	G+	G+	G+
481	G	G+	G+	VG	G+
489	G	G	G	G	G
	Treatment average				G+
<b>Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (outdoors)</b>					
480	G+	G+	G+	G+	G+
483	G+	G	G	FG	G
490	G	G	F	F	FG
	Treatment average				G
<b>Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (indoors)</b>					
476	F	F	FG	FG	F
485	G	G	G	G	G
487	FG	FG	F	P	F
	Treatment average				F
<b>Sulfate sulfur 0.3%, molybdenum 50 p.p.m., arsenic 5 p.p.m., rubidium 100 p.p.m. and lead 5 p.p.m. (indoors)</b>					
478	FG	G	FG	FG	FG
484	G+	G+	G	G	G
486	G	FG	FG	F	FG
	Treatment average				FG

molybdenum, outdoors, and 0.3% sulfate sulfur, outdoors, had higher ratings than the treatment 50 p.p.m. molybdenum with 0.3% sulfate sulfur, outdoors. All the treatments housed outdoors, however showed consistently higher body condition ratings than the treatments housed indoors. There was no advantage in body condition seen for either treatment indoors which indicated that the addition of arsenic, lead, and rubidium had no additional effect in lowering body condition.

A record of the hair condition ratings during Experiment III are found in Table 23. The treatments of 50 p.p.m. molybdenum, outdoors and 0.3% sulfate sulfur, outdoors, showed no changes in hair condition throughout the experiment. All other treatments, however, showed achromotrichia, alopecia, and hair texture changes which started after 150 days on experiment. It is evident from these observations that both a high level of molybdenum and a high level of inorganic sulfate were necessary to cause these hair changes. Four cows which received 50 p.p.m. molybdenum with 0.3% sulfate sulfur for 300 days, showed complete recovery of hair condition abnormalities after receiving 10 and 20 p.p.m. copper added to this diet. These data indicated a copper deficiency was the cause of these hair changes.

Weekly observations indicated that all animals receiving 0.3% sulfate sulfur in the diet had a higher water content in the feces than the three animals not receiving any sulfate sulfur. The feces of the animals which received 0.3% sulfate sulfur were described as being slightly loose to very

TABLE 23

Hair Condition  
(Experiment III)

Animals by treatment	Hair color change (outdoors)	Distribution	Hair texture
Sulfate sulfur 0.3% (outdoors)			
477	None	Normal	Normal
482	"	"	"
488	"	"	"
Molybdenum 50 p.p.m. (outdoors)			
479	None	Normal	Normal
481	"	"	"
489	"	"	"
Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (outdoors)			
480	White hair became silver white at 230 days on experiment	Alopecia around eyes	Hair lacked body and became very soft at 200 days
483	Achromotrichia of black hair & white hair became silver white at 160 days on experiment	Alopecia around eyes and under jaw	Hair very soft, lacked body after 160 days
490	Achromotrichia of black hair. White hair turned silver white at 125 days on experiment	Alopecia on face very pronounced, and moderate over body	Hair lacked body. Soft cotton texture after 150 days
Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (indoors)			
476	Achromotrichia of black hair at 150 days on experiment	Alopecia prominent around eyes, and to less degree over body	Hair very soft and loss of body
485	Black hair turned to brown at 200 days	Slight alopecia around eyes	Normal
487	Achromotrichia of black hair at 150 days	Wide spread alopecia over whole body	Hair very soft, having no body

TABLE 23 (Cont.)

Hair Condition  
(Experiment III)

Animals by treatment	Hair color change	Distribution	Hair texture
Sulfate sulfur 0.3%, molybdenum 50 p.p.m., arsenic 5 p.p.m., rubidium 100 p.p.m., and lead 5 p.p.m. (indoors)			
478	Moderate amount of black hair turning to brown. White hair became silver white	Slight alopecia around eyes	Normal
484	White hair turned to silver white	Wide-spread alopecia over face and body	Hair became softer with less body at 225 days
486	Achromotrichia of black hair. White hair became silver white at 170 days	Wide-spread alopecia over entire body	Hair soft cotton-like texture after 200 days



loose. Definite diarrhea was not observed for any animals while housed indoors or outdoors in the pole barn. The feces of the animals which were placed on pasture for 75 days showed an even greater water content. At times individual animals had bowel movements which approached a condition of diarrhea. At no time, however, was such a condition debilitating. Such looseness was present in all treatments of animals on pasture. It is interesting to note that the administration of copper sulfate at 10 and 20 p.p.m. to animals which had finished 300 days on experiment but were still receiving the assigned treatment showed a great lowering of the water content in the feces.

The breeding records are recorded in Table 24. All the animals housed outdoors conceived with little delay. One of these animals aborted at the end of 300 days on experiment, but tests for brucellosis and vibrio fetus were negative. Three animals from the treatments indoors were not pregnant at the end of 300 days. Two of these were bred more than four times, this indicating that heat periods were observed. The third animal was bred only twice, however, because heat periods were not observed. All the animals which did not conceive occupied stanchions in the darkest part of the barn.

The average hemoglobin and hematocrit values for each period are recorded in Tables 25 and 26, respectively. There is wider variation among animals in each treatment than there is among treatments, this indicating no treatment effect. Because all the animals, with one exception, maintained a normal range of values there was no evidence of anemia. The one ex-

TABLE 24

Breeding Records  
(Experiment III)

Animals by treatment	Number of services	Number of days before conception
Sulfate sulfur 0.3% (outdoors)		
477	2	269
482	2	158
488	1	112
	<hr style="width: 50px; margin: 0 auto;"/>	<hr style="width: 50px; margin: 0 auto;"/>
Total	5	539
Average	1.7	180
Molybdenum 50 p.p.m. (outdoors)		
479	1	180
481	1	117
489	4	206
	<hr style="width: 50px; margin: 0 auto;"/>	<hr style="width: 50px; margin: 0 auto;"/>
Total	6	503
Average	2	166
Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (outdoors)		
480	1 (Aborted)	162
483	1	110
490	1	129
	<hr style="width: 50px; margin: 0 auto;"/>	<hr style="width: 50px; margin: 0 auto;"/>
Total	3	401
Average	1	134
Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (indoors)		
476	4 (not preg.)	300
485	1	144
487	1	166
	<hr style="width: 50px; margin: 0 auto;"/>	<hr style="width: 50px; margin: 0 auto;"/>
Total	6	610
Average	2+	203
Sulfate sulfur 0.3%, molybdenum 50 p.p.m., arsenic 5 p.p.m., rubidium 100 p.p.m., and lead 5 p.p.m. (indoors)		
478	1	177
484	7 (not preg.)	300
486	2 (not preg.)	300
	<hr style="width: 50px; margin: 0 auto;"/>	<hr style="width: 50px; margin: 0 auto;"/>
Total	10	777
Average	3.3+	259

TABLE 25

**Average Hemoglobin Values  
(Experiment III)**

Animals by treatment	Periods				300 day averages
	I	II	III	IV	
	mg/100 ml of blood				
<b>Sulfate sulfur 0.3% (outdoors)</b>					
477	11.03	10.90	11.09	11.88	11.23
482	9.80	10.41	10.41	9.73	10.09
488	10.87	10.14	10.51	9.62	10.29
Totals	31.70	31.45	32.01	31.23	31.61
Averages	10.57	10.48	10.67	10.41	10.54
<b>Molybdenum 50 p.p.m. (outdoors)</b>					
479	8.80	8.56	8.78	8.52	8.67
481	9.92	10.88	10.48	10.25	10.38
489	10.21	11.10	11.02	9.25	10.40
Totals	28.93	30.54	30.28	28.02	29.45
Averages	9.64	10.18	10.09	9.34	9.82
<b>Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (outdoors)</b>					
480	10.12	9.65	9.56	10.32	9.91
483	9.46	9.43	9.03	8.87	9.20
490	11.04	11.22	10.16	9.50	10.48
Totals	30.62	30.30	28.75	28.69	29.59
Averages	10.21	10.10	9.58	9.56	9.86
<b>Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (indoors)</b>					
476	9.35	9.90	10.33	9.33	9.73
485	9.32	9.97	10.88	9.87	10.01
487	10.51	10.73	10.25	7.80	9.82
Totals	29.18	30.60	31.46	27.00	29.56
Averages	9.73	10.20	10.49	9.00	9.85
<b>Sulfate sulfur 0.3%, molybdenum 50 p.p.m., arsenic 5 p.p.m., rubidium 100 p.p.m., and lead 5 p.p.m.</b>					
478	9.70	10.01	10.43	10.82	10.24
484	10.40	10.54	11.17	9.73	10.46
486	9.63	9.96	9.52	8.76	9.47
Totals	29.73	30.51	31.12	29.31	30.17
Averages	9.91	10.17	10.37	9.77	10.06

TABLE 26

Average Hematocrit Values  
(Experiment III)

Animals by treatment	Periods				300 day Averages
	I	II	III %	IV	
Sulfate sulfur 0.3% (outdoors)					
477	34.0	33.8	35.4	38.8	35.50
482	29.6	29.8	33.4	31.9	31.17
488	33.7	33.1	32.6	31.4	32.70
Totals	97.3	96.7	101.4	102.1	99.37
Averages	32.4	32.2	33.8	34.0	33.12
Molybdenum 50 p.p.m. (outdoors)					
479	28.0	25.1	26.8	25.9	26.45
481	30.0	30.5	31.2	31.0	30.68
489	32.8	33.9	34.2	28.9	32.45
Totals	90.8	89.5	92.2	85.8	89.58
Averages	30.3	29.8	30.7	28.6	29.86
Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (outdoors)					
480	31.9	30.4	28.6	31.8	30.68
483	29.7	29.8	27.6	28.3	28.85
490	33.6	36.0	32.8	28.9	32.82
Totals	95.2	96.2	89.0	89.0	92.35
Averages	31.7	32.1	29.7	29.7	30.78
Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (indoors)					
476	29.0	31.7	33.4	29.5	30.90
485	30.7	31.8	32.8	33.0	32.08
487	32.7	32.2	30.6	24.4	29.98
Totals	92.4	95.7	96.8	86.9	92.96
Averages	30.8	31.9	32.3	29.0	30.99
Sulfate sulfur 0.3%, molybdenum 5 p.p.m., arsenic 5 p.p.m., rubidium 100 p.p.m., and lead 5 p.p.m. (indoors)					
478	30.9	31.9	35.4	38.8	34.25
484	34.2	35.2	35.6	31.9	34.23
486	31.0	31.3	30.7	27.6	30.15
Totals	96.1	98.4	101.7	98.3	98.63
Averages	32.0	32.8	33.9	32.8	32.88

ception, animal 487, received the treatment of 50 p.p.m. molybdenum and 0.3% sulfate sulfur indoors. This animal's hemoglobin and hematocrit values dropped from 10.48 to 6.42 and 29.5 to 20.0 respectively within a two month period. Serum protein values are recorded in Table 27. The values for all treatments are similar.

Milk and butterfat production are recorded in Table 28. The average milk production of the animals housed outdoors was found to greatly exceed the average production of the animals housed indoors. As might be expected the animals within each treatment vary in production due to the manner in which they were selected.

Outside of mastitis and flyspray burn, the health of all the animals with one exception was considered normal. The exception was animal 487, which received 50 p.p.m. molybdenum and 0.3% sulfate sulfur and was maintained indoors. This animal showed a loss of appetite during the third and fourth periods of experiment and on several occasions went off feed entirely. She continued to lose weight during period four and had a dull appearance. At 280 days on experiment, when anemia was noted, the animal was given 2 g. of copper sulfate per day. After two weeks with no sign of improvement the animal was slaughtered and the carcass was examined. The only abnormality found was that the liver had large white areas of infection interspersed throughout the tissues of the organ. The incidence of mastitis was unusually high, particularly among the animals housed indoors, but no treatment differences were seen. The animals housed outdoors were relatively free

TABLE 27

Serum Protein  
(Experiment III)

Animals by treatment	Days					Total
	0	75	150	225	300	
<b>Sulfate sulfur 0.3% (outdoors)</b>						
477	7.2	7.6	8.8	7.9	7.9	39.4
482	7.3	7.4	7.4	7.6	7.4	37.1
488	7.5	8.3	7.5	7.4	7.6	38.3
				Total		114.8
				Average		7.65
<b>Molybdenum 50 p.p.m. (outdoors)</b>						
479	6.4	7.4	6.8	7.0	7.7	35.3
481	7.9	8.2	8.6	8.1	8.4	41.2
489	7.5	8.0	7.2	7.4	7.2	37.3
				Total		113.8
				Average		7.59
<b>Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (outdoors)</b>						
480	7.9	8.0	7.4	7.4	7.8	38.5
483	6.4	7.2	7.1	7.0	7.8	35.5
490	7.9	8.0	6.9	6.6	7.4	36.8
				Total		110.8
				Average		7.39
<b>Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (indoors)</b>						
476	7.5	7.8	7.6	7.0	7.0	36.9
485	7.0	7.8	7.7	7.8	7.6	37.9
487	7.4	7.8	7.8	7.3	9.1	39.4
				Total		114.2
				Average		7.61
<b>Sulfate sulfur 0.3%, molybdenum 50 p.p.m., arsenic 5 p.p.m., rubidium 100 p.p.m., and lead 5 p.p.m. (indoors)</b>						
478	7.8	8.2	7.5	7.4	7.9	38.8
484	6.7	7.5	7.8	7.2	7.4	36.6
486	7.3	6.4	7.0	6.7	6.8	34.2
				Total		109.6
				Average		7.31

TABLE 28

Total Milk and Butterfat Production in Pounds  
(Experiment III)

Animals by treatment	Periods				Total milk	Total fat
	I	II	III	IV		
<b>Sulfate sulfur 0.3% (outdoors)</b>						
477 milk	3088.1	2877.0	2061.4	2386.8	10413.3	
fat	146.0	104.4	79.9	87.5		417.8
482 milk	1997.3	1993.1	1692.6	1282.9	6965.9	
fat	92.2	92.2	75.4	62.3		322.8
488 milk	2768.4	2541.8	1998.8	1512.3	8821.3	
fat	82.7	79.2	60.7	63.8		286.4
			<b>Total</b>		<b>26200.5</b>	<b>1027.0</b>
			<b>Average</b>		<b>8733.5</b>	<b>342.3</b>
<b>Molybdenum 50 p.p.m. (outdoors)</b>						
479 milk	2972.9	2513.7	1854.5	1543.9	8885.0	
fat	96.9	95.5	69.2	59.0		320.6
481 milk	2769.4	2213.5	1845.1	1329.2	8157.2	
fat	99.5	92.4	64.7	48.3		304.9
489 milk	3482.6	2722.9	2140.4	1593.3	9939.2	
fat	113.5	83.6	66.4	52.5		316.0
			<b>Total</b>		<b>26981.4</b>	<b>941.5</b>
			<b>Average</b>		<b>8994.0</b>	<b>313.8</b>
<b>Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (outdoors)</b>						
480 milk	2913.8	2462.4	1899.0	1414.5	8689.7	
fat	105.4	96.4	78.0	57.6		337.4
483 milk	3296.0	2473.1	1516.2	1455.1	8740.4	
fat	108.4	88.4	57.1	61.3		315.2
490 milk	2496.7	1279.3	678.2	189.1	4643.3	
fat	79.9	41.2	20.4	8.5		150.0
			<b>Total</b>		<b>22073.4</b>	<b>802.6</b>
			<b>Average</b>		<b>7357.8</b>	<b>267.5</b>

TABLE 28 (Cont.)

Total Milk and Butterfat Production in Pounds  
(Experiment III)

Animals by treatment	Periods				Total milk	Total fat
	I	II	III	IV		
Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (indoors)						
476 milk	1456.9	1746.6	1029.5	806.4	5039.4	
fat	55.9	70.7	44.5	31.1		202.2
485 milk	2668.5	2001.0	1257.7	1083.5	7010.1	
fat	85.3	79.3	42.8	41.1		248.5
487 milk	2468.0	1730.2	1050.3	466.4	5714.9	
fat	74.0	56.7	32.6	13.4		176.7
Totals					17764.4	627.4
Average					5921.5	209.1
Sulfate sulfur 0.3%, molybdenum 50 p.p.m., arsenic 5 p.p.m., rubidium 100 p.p.m., lead 5 p.p.m. (indoors)						
478 milk	2659.5	2391.0	1603.3	1997.5	8651.3	
fat	105.4	98.0	66.7	79.9		350.0
484 milk	2372.9	1796.9	1206.5	1117.8	6494.1	
fat	93.3	78.0	50.8	41.3		263.4
486 milk	2302.5	1639.7	1077.2	736.8	5756.2	
fat	56.7	49.0	37.7	28.1		171.5
Totals					20901.6	784.9
Average					6967.2	261.6



of the disease. During the summer, when all the animals were sprayed with an insecticide, several cows which received 50 p.p.m. molybdenum with 0.3% sulfate sulfur developed severe skin burns. These skin conditions healed very slowly. It was unusual that animals from other treatments were not effected.

The milk copper levels for Experiment III are recorded in Table 28 of the Appendix. Except for the values of the 150 day samples, the milk copper levels were in good agreement with those of Experiment I and II. The values for the 150 day samples for all animals appear to be inaccurate due to errors in analysis. The reason for these errors is not evident. The levels for milk molybdenum for this experiment are recorded in Table 28 of the Appendix. The means of these values for each treatment are plotted in Figure 11. The animals which received 50 p.p.m. of molybdenum with no sulfate had much higher milk molybdenum levels than those which received 50 p.p.m. of molybdenum with 0.3% sulfate sulfur. These results are in complete agreement with a comparison of Experiments I and II. It provides further evidence that increased sulfate sulfur in the diet reduces the excretion of molybdenum in the milk.

The values for serum copper and molybdenum for this experiment are recorded in Table 29 of the Appendix. The mean serum copper values, which are plotted in Figure 12, show that serum copper decreased during the course of the experiment for all treatments. Both of the treatments which received molybdenum with sulfate sulfur had serum copper levels which were subnormal at the end of the experiment. The mean serum

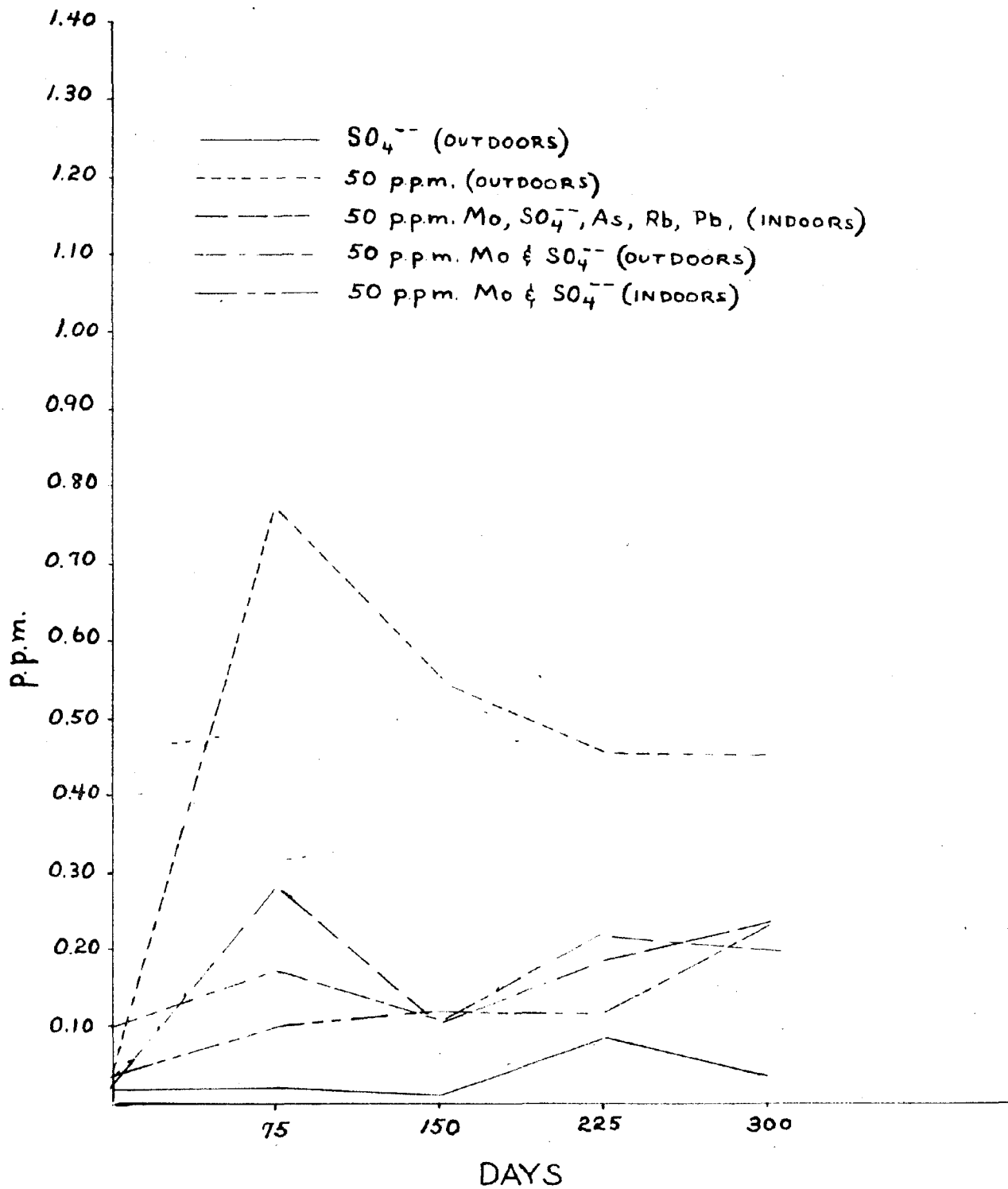


Fig. 11. Milk molybdenum levels for Experiment III.

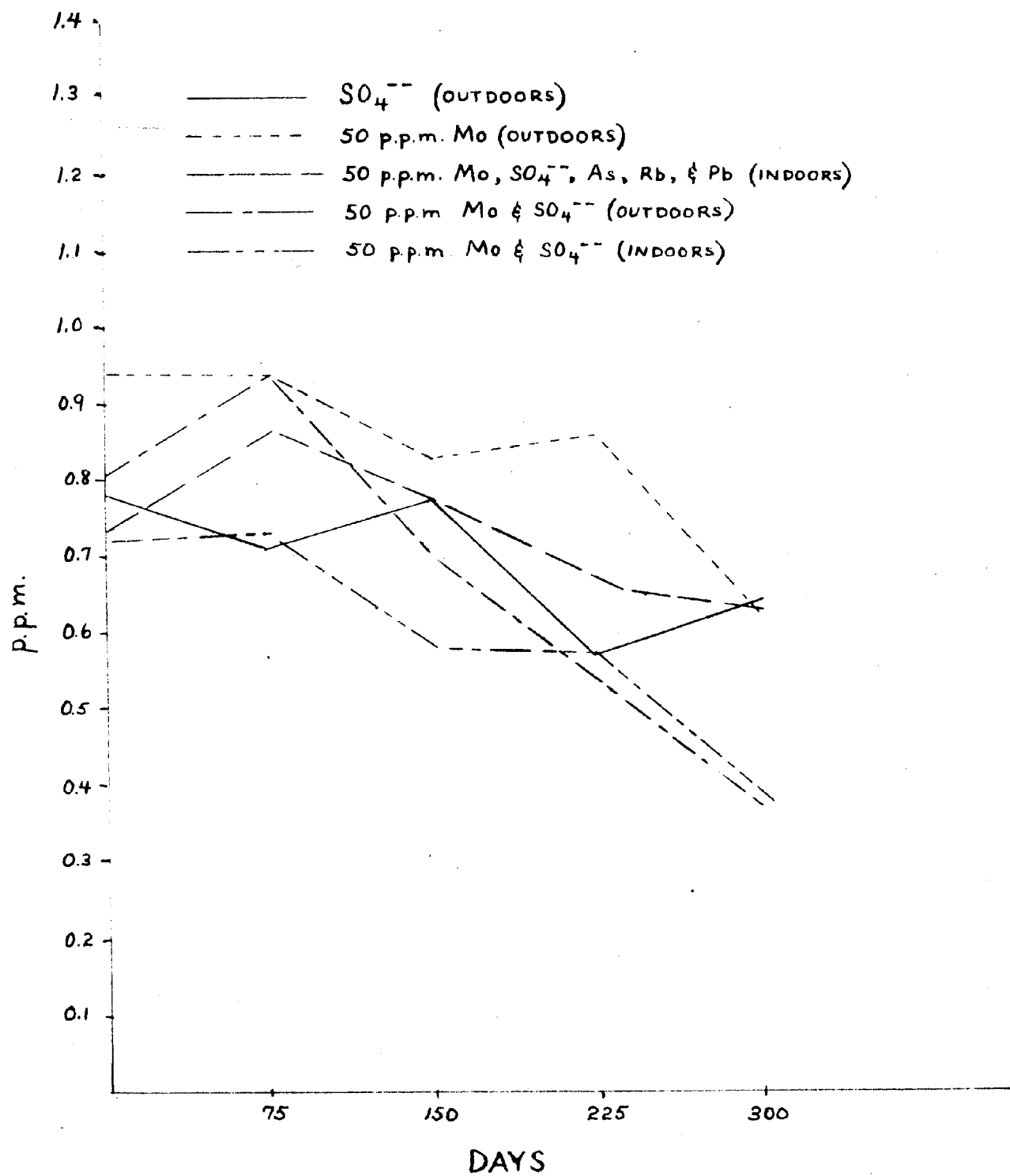


Fig. 12. Serum copper levels for Experiment III.

molybdenum levels are plotted in Figure 13. The level for the treatment which received 50 p.p.m. of molybdenum was higher than for the treatments which received 50 p.p.m. of molybdenum with 0.3% sulfate sulfur. These results are in agreement with the results of Experiments I and II, and indicate that the sulfate level of diet has an effect on the serum molybdenum level.

Liver molybdenum and copper levels are recorded in Table 30 of the Appendix. The mean liver copper levels are plotted in Figure 14. There was a general decrease in the levels of liver copper for all levels of molybdenum intake through the first 225 days of the experiment. During the last 75 days both outdoor treatments of molybdenum showed unexplained increases of liver copper. The treatment which received 0.3% sulfate sulfur showed an increase for the first 150 days of experiment and then a decrease for the remaining 150 days. The mean liver molybdenum levels, which are plotted in Figure 15, show that the 50 p.p.m. of molybdenum treatment had a higher liver molybdenum level than the treatments which received 50 p.p.m. of molybdenum with 0.3% sulfur. This comparison shows the depressing effect sulfate has on molybdenum deposition in the tissues. The level of molybdenum in the liver of the treatment which received 0.3% sulfate sulfur increased as the experiment progressed.

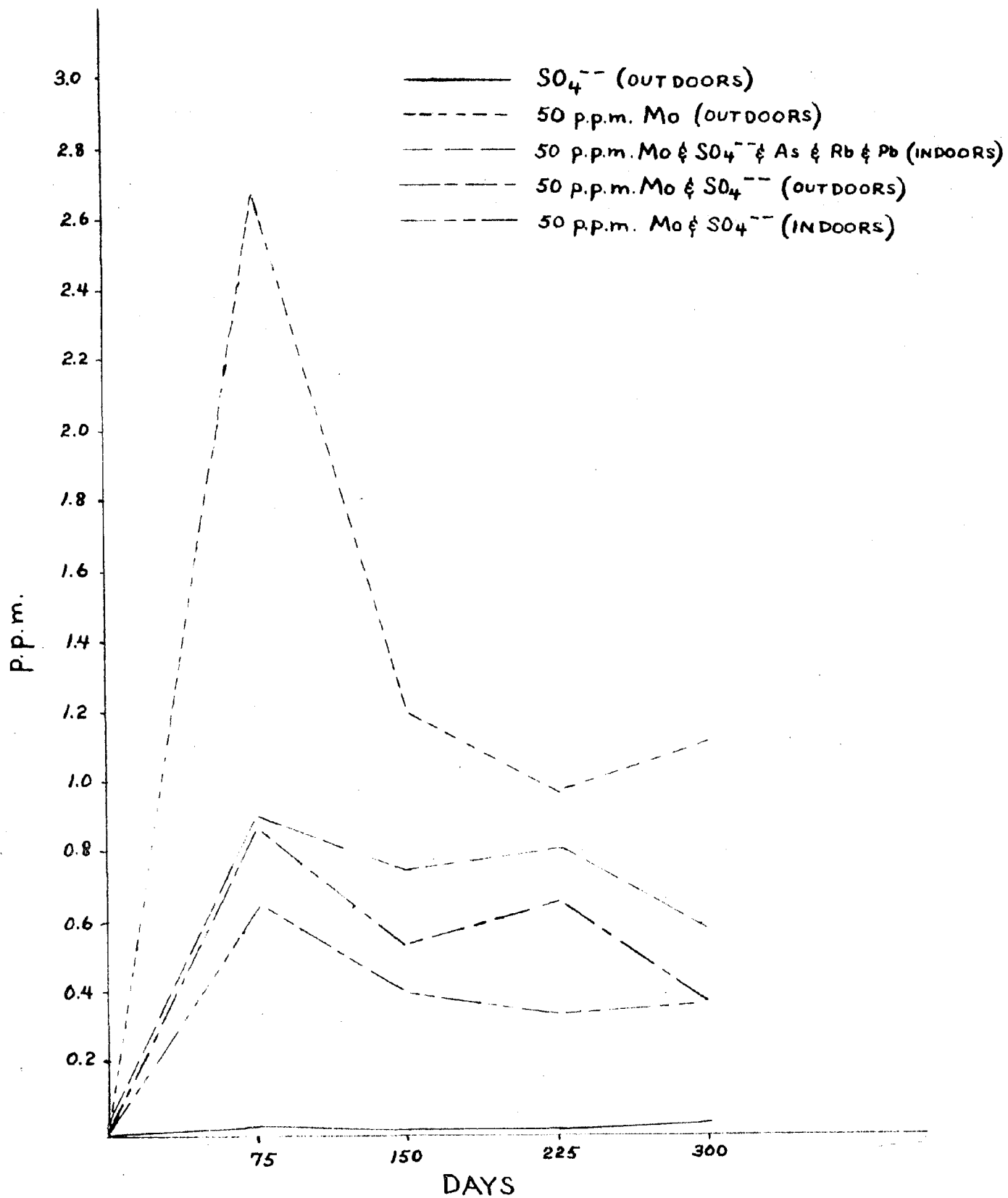


Fig. 13. Serum molybdenum levels for Experiment III.

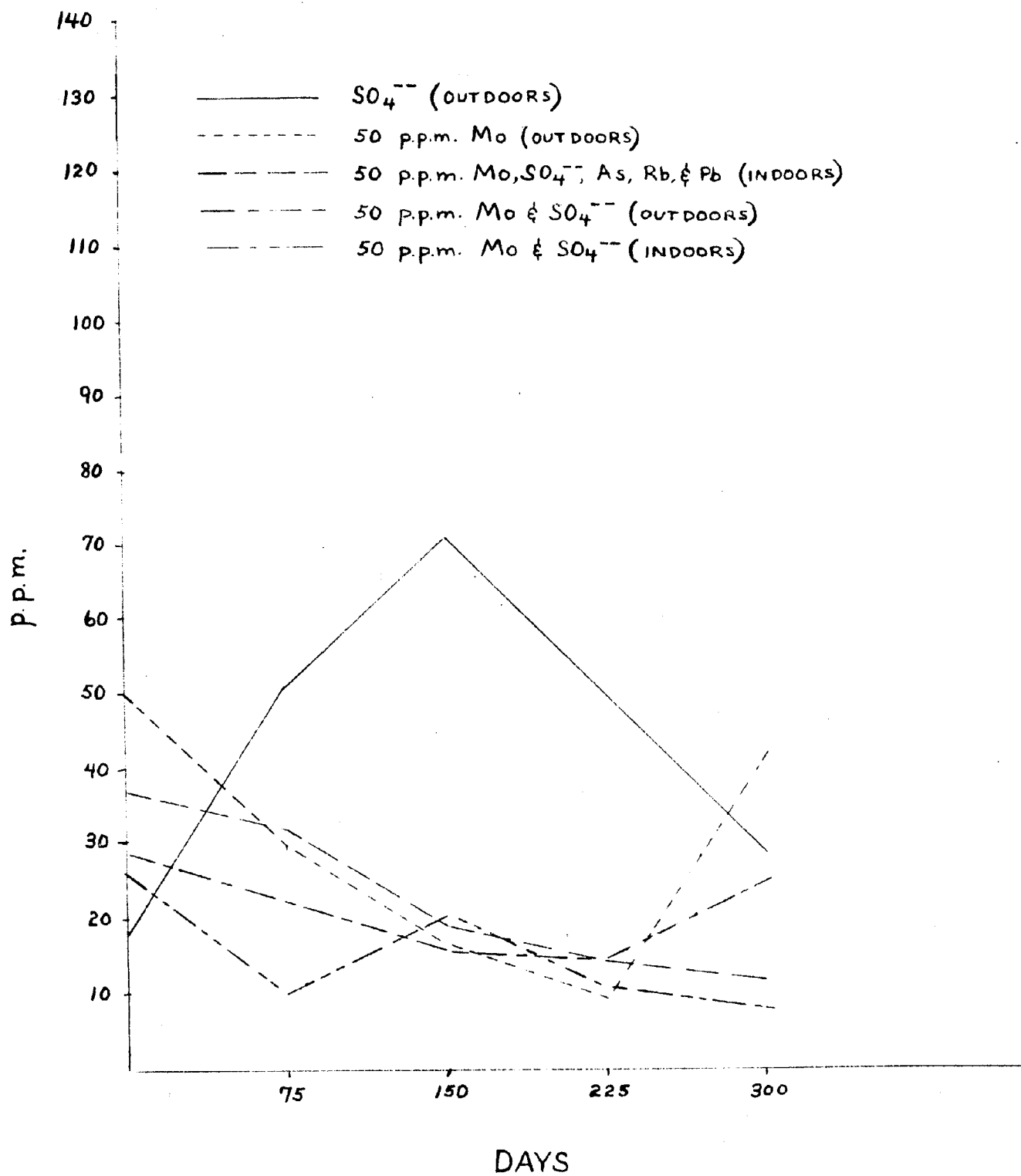


Fig. 14. Liver copper levels for Experiment III.

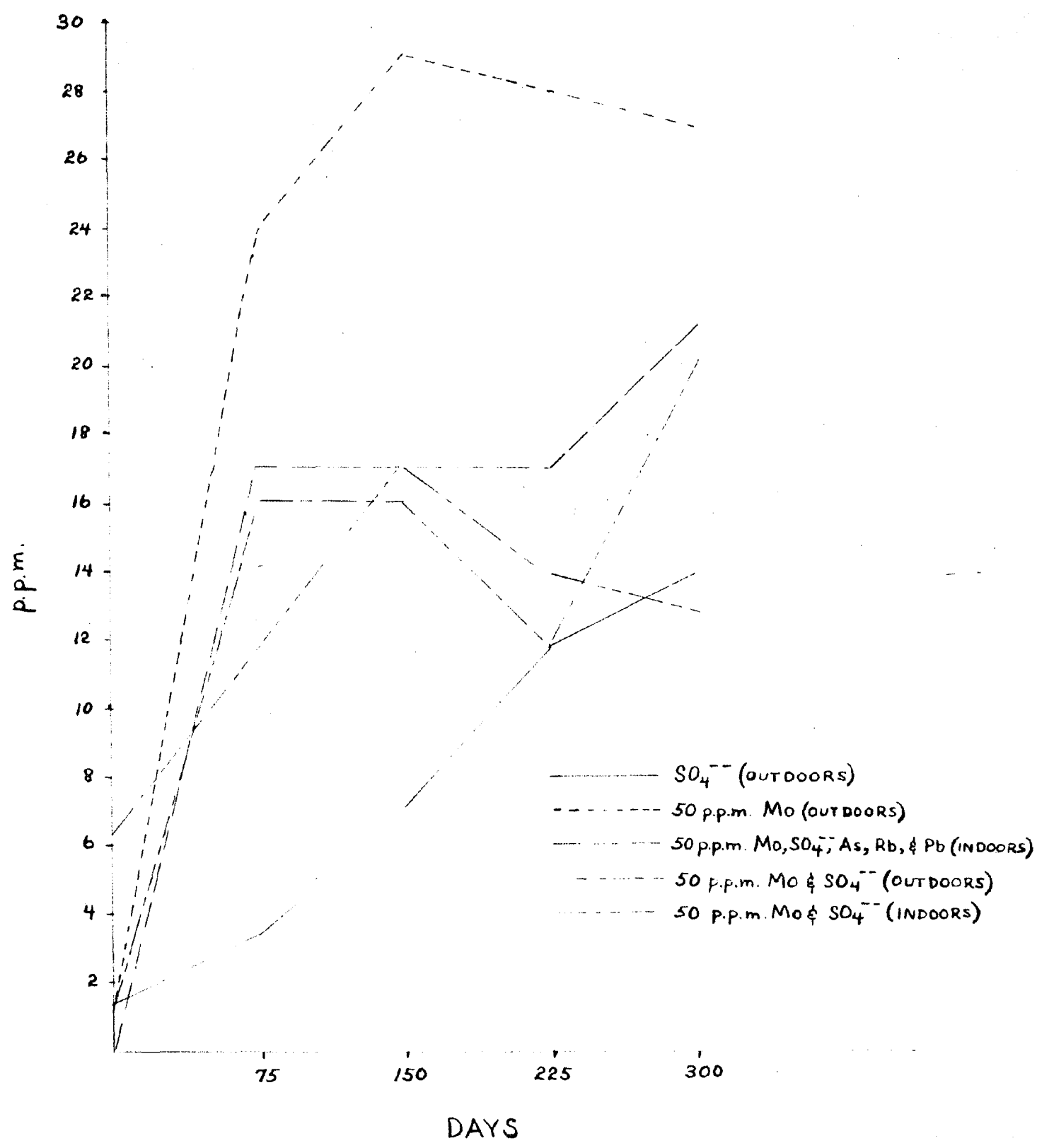


Fig. 15. Liver molybdenum levels for Experiment III.

## EXPERIMENT IV

Attempts to measure the absorption of copper-64 by the blood with a windowless proportional flow counter were not successful due to the low energy of the Beta radiation of this isotope. As a result accurate counts for the first 24 hours of experiment were not obtained during the first trial. The gamma well scintillation counter was used in all counting thereafter and was found to be very reliable. In Figure 16 the levels of copper-64 obtained from blood for Trials I and II are plotted. In both trials the steer which received 50 p.p.m. with 0.3% sulfate sulfur had a slightly higher level of copper-64 in the blood than was found for the steer which received 0.3% sulfate sulfur. The levels of copper-64 found in the blood for Trials III and IV are plotted in Figure 17. In Trial III steer 498 which received 50 p.p.m. molybdenum showed considerably higher levels of copper-64 in the blood than were found for the control steer 499. In Trial IV with the treatments reversed, both steers, 498 and 499, showed similar levels of copper-64 in the blood for the first 12 hours. After this time, however, steer 498, which received no molybdenum, showed higher levels than steer 499 which received 50 p.p.m. molybdenum. The levels of copper-64 found in the blood for Trials V and VI are plotted in Figure 18. No differences were found between the blood levels of the steer which received 0.3% sulfate sulfur and the control steer in either Trials V or VI.



Trial I

Trial II

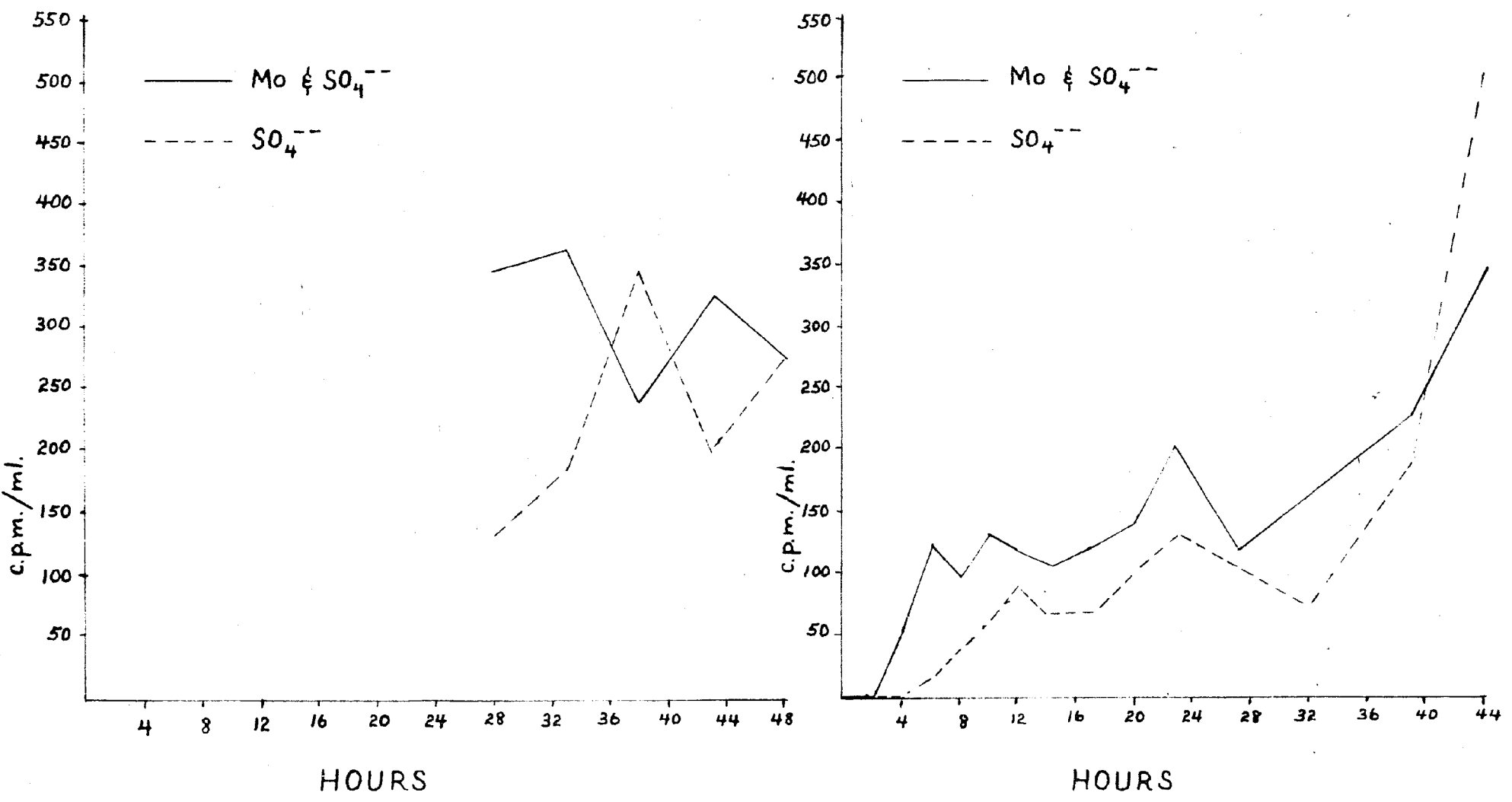
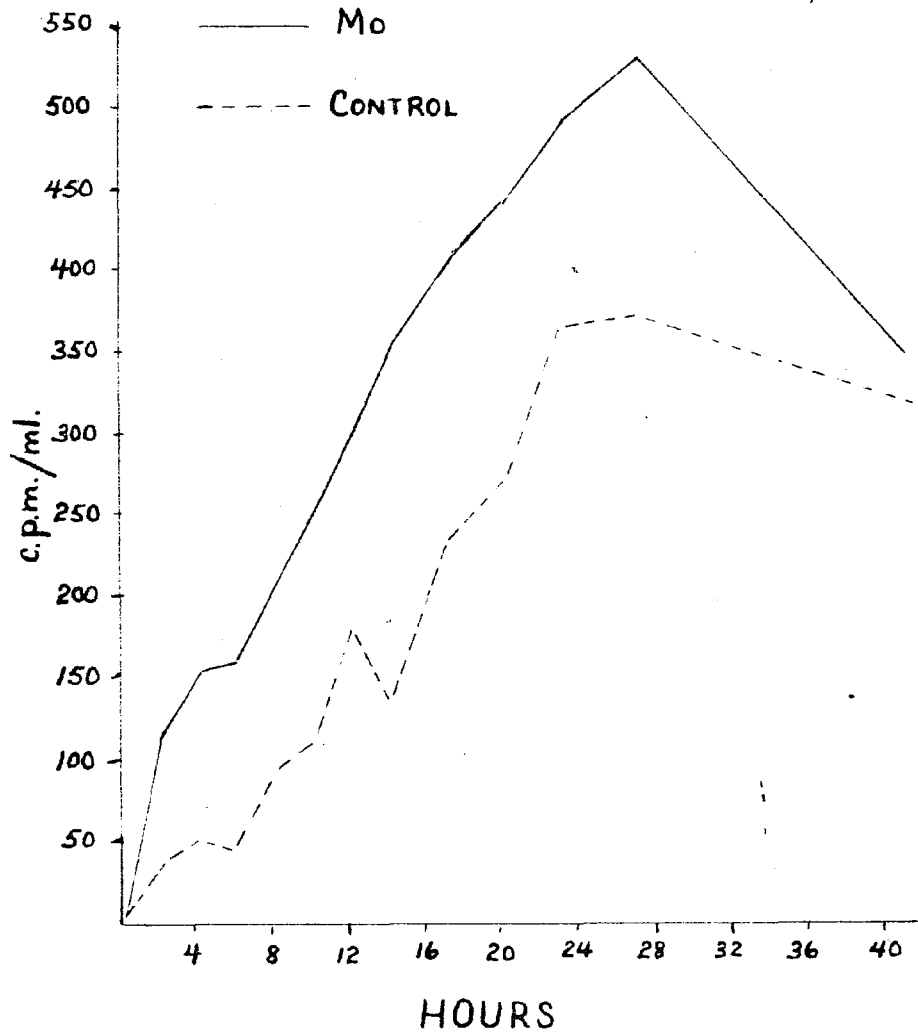


Fig. 16. Levels of copper-64 in blood for Trials I and II.

Trial III



Trial IV

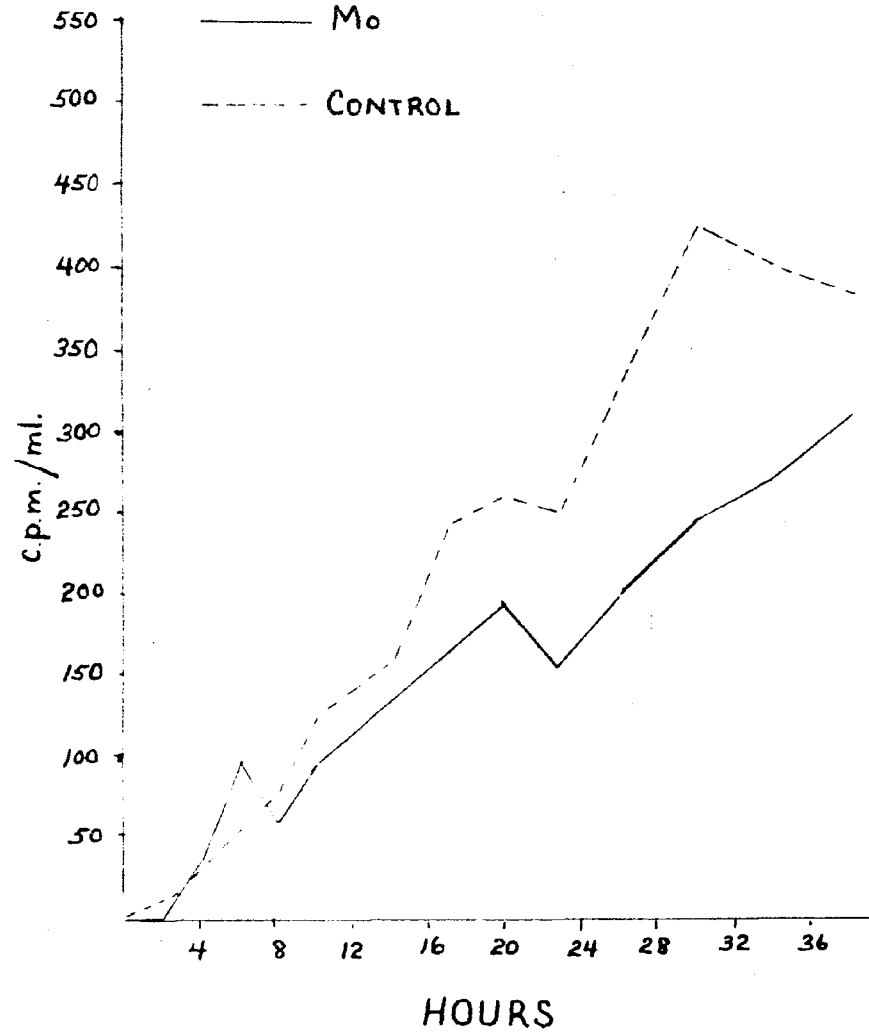


Fig. 17. Levels of copper-64 in Blood for Trials III and IV.

Trial V

Trial VI

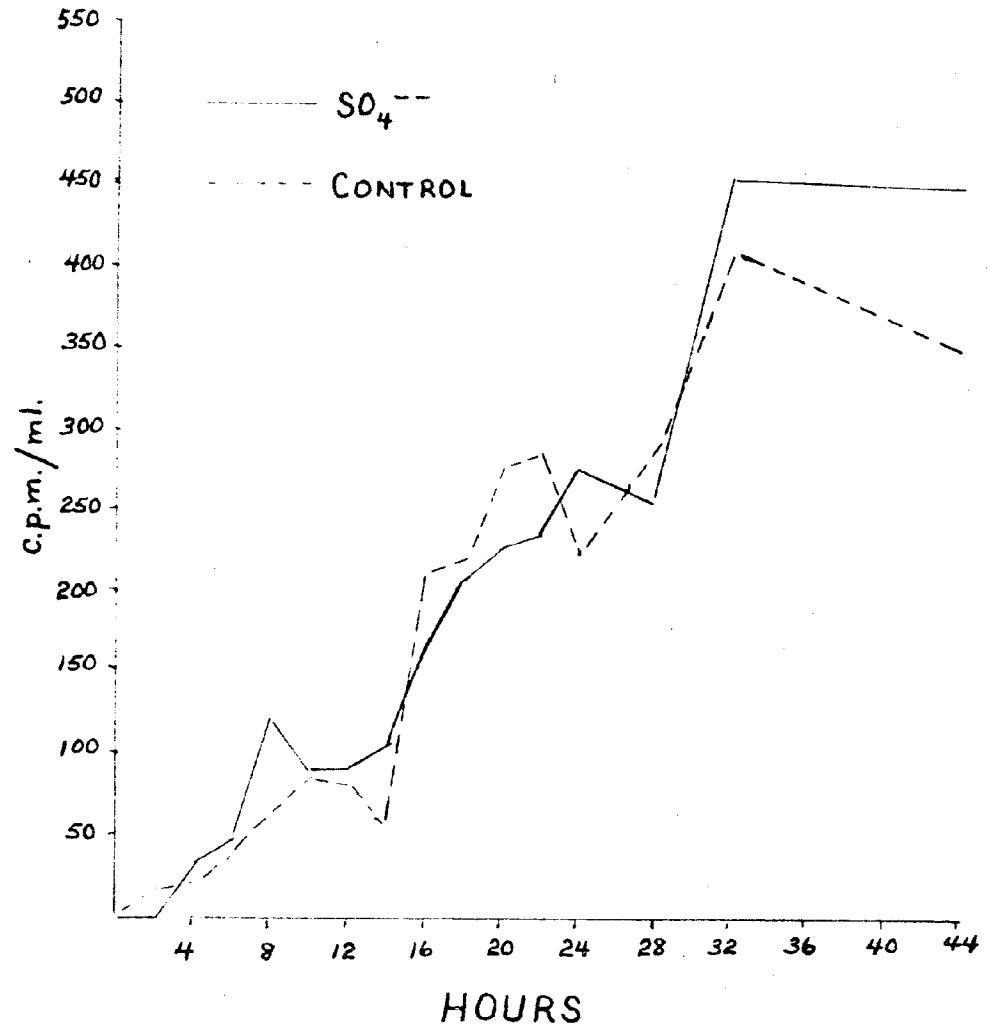
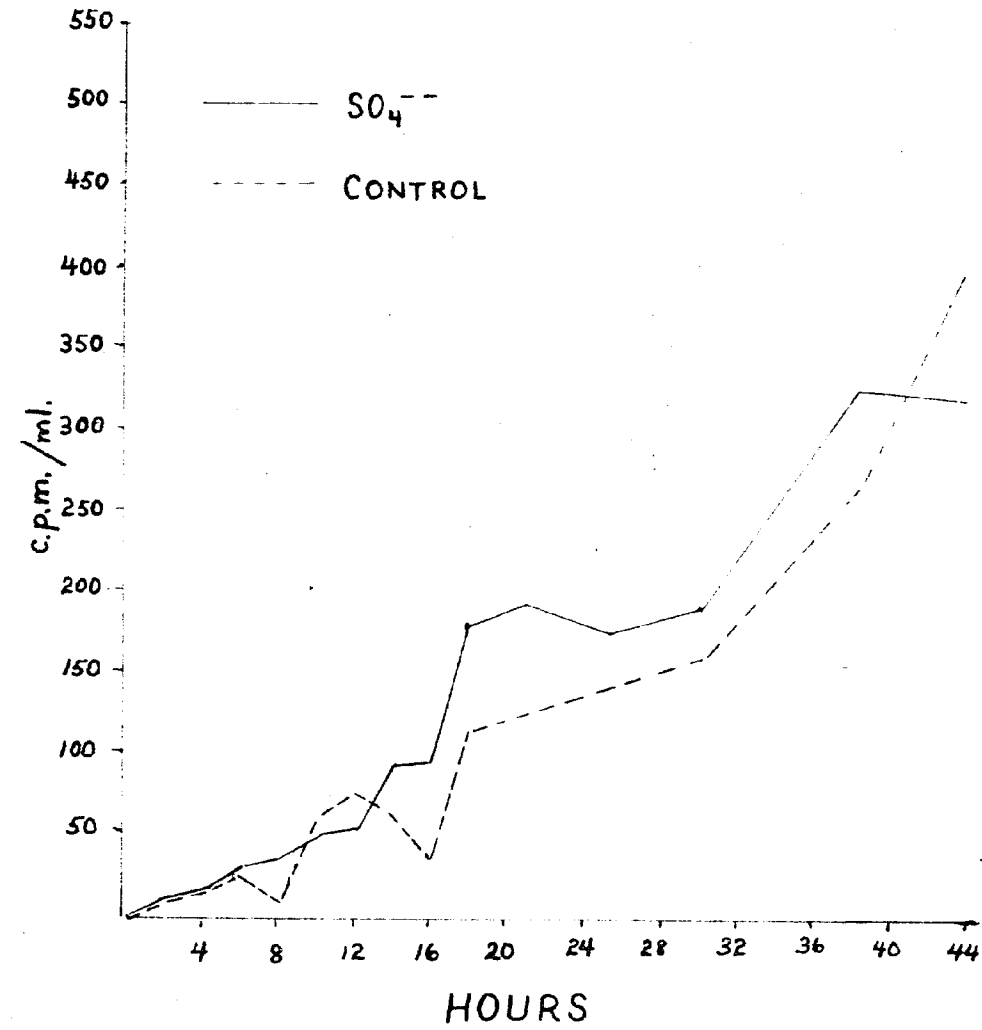


Fig. 18. Levels of copper-64 in blood for Trials V and VI.

A very surprising fact discovered was the unusually low absorption of the copper-64 by the steers. In all six trials, the recovery of copper-64 in the feces was greater than 96.5%. Copper-64 was recovered in the feces as early as six hours after administration and continued to be excreted for five days. The level of copper-64 found in the urine was not sufficient to count accurately without concentrating the sample. Results from measuring the activity of evaporated urine samples indicated less than 0.5% of the copper-64 administered was excreted by this pathway. Calculation of the level of copper-64 in the urine was hampered by feces contaminations on several occasions.

The copper-64 contents of liver samples obtained 24 hours after administration of the isotope are recorded by trials in Table 29. In Trial I the steer which received 50 p.p.m. molybdenum and 0.3% sulfate sulfur deposited only 30 per cent as much copper-64 in the liver as the steer which received 0.3% sulfate. Similar results were found in Trial II where the steer which received 50 p.p.m. molybdenum with 0.3% sulfate sulfur deposited 13 per cent of the level of copper -64 in the liver of the steer which received 0.3% sulfate sulfur. In Trials III and IV, where sulfate was removed from the ration, the deposition of copper-64 in the livers of the steers which received 50 p.p.m. molybdenum was respectively 21 and 6 per cent of the level found in the steers which did not receive molybdenum. In comparing the results of Trials V and VI, which was designed to test the

TABLE 29

Copper-64 levels in liver  
(Experiment IV)

Trial	Animal	Dose	Accumulation of copper-64 in liver during 24 hrs. counts per minute/gr.*	Treatment
I	473	5 mc.	1405	SO <sub>4</sub> <sup>=</sup> + Mo.
	474	5 mc.	4690	SO <sub>4</sub> <sup>=</sup>
II	473	5 mc.	5894	SO <sub>4</sub> <sup>=</sup>
	474	5 mc.	766	SO <sub>4</sub> <sup>=</sup> + Mo.
III	498	5 mc.	1887	Mo.
	499	5 mc.	9048	_____
IV	498	5 mc.	4945	_____
	499	5 mc.	315	Mo.
V	473	10 mc.	1440	SO <sub>4</sub> <sup>=</sup>
	474	10 mc.	1520	_____
VI	498	10 mc.	4306	_____
	499	10 mc.	4242	SO <sub>4</sub> <sup>=</sup>

\* Corrected to time when copper-64 level was equal to dose.

effect of adding sulfate to the diet on the deposition of copper-64 in the liver, differences of 5 and 1 per cent greater deposition occurred in the controls, respectively.

The data obtained from the analysis of serum, liver and milk were tested for statistical significance by Professor Owen B. Durgin, Agriculture Experiment Station Statistician. A summary of the results of this statistical analysis is found in the Appendix. These data were used as a guide for the reporting of results in this thesis.

## DISCUSSION

Although Davis (39), Cunningham (31), Miller (118), and Dick (52) reported that symptoms of molybdenum toxicity occurred when sheep and cattle were fed rations containing levels of from 6 to 36 p.p.m. of molybdenum, similar results were not obtained on this experiment. Levels of 5 to 50 p.p.m. were fed without developing the usual symptoms of toxicity such as loss of body weight, diarrhea, achromotrichia, anemia, and nervous disorders.

The analysis of milk, serum, and liver for copper and molybdenum, however, revealed some important changes which were the result of treatment. Molybdenum levels were shown to increase in the milk, serum and liver with increased feeding of sodium molybdate. These data indicate that molybdenum is absorbed at a rate which is dependent on the level of this element in the diet. The fact that all of the treatments showed stabilization of these molybdenum levels indicates that the excretion of this element is a function of its absorption. The data obtained from analysis of liver and serum are consistent with work reported by Dick (52) with sheep and Cunningham (37) with cattle. Similarly, the levels of molybdenum in the milk are in good agreement with work reported by Archibald (9) in which four cows were fed an estimated level of 10 p.p.m. molybdenum.

The analysis of milk, liver, and serum for copper is of even greater interest because molybdenum toxicity has been

described by Cunningham (31), Davis (39), and Dick (52) as complicated copper deficiency. In this experiment the copper levels of the milk showed a change due to time but not to treatment. Apparently, copper secretion in milk varies with stage of lactation. No supporting evidence of this fact could be found in the literature. The copper contents of both serum and liver were shown to decrease with an increase of molybdenum in the diet. The serum copper levels for animals which received from 5 to 50 p.p.m. of molybdenum were just under the normal range of 80 to 120 micrograms per 100 ml. of serum given by Underwood (152) and Marston (112). Adams et al. (1) have reported, however, finding levels of serum copper in cattle which were below this range with no apparent copper deficiency. It should be noted also that the level of copper found for these animals was from 10 to 20 micrograms per 100 ml. serum above the levels reported by Cunningham (31), Marston (112), and Davis (39) to be present in copper deficient animals.

At the end of Experiment I, several animals which received supplemental molybdenum had levels of liver copper as low as 9 to 12 p.p.m. This level of copper is far below the normal range of 100 to 400 p.p.m. reported by Underwood (152) and Marston (112). These levels are as low as those reported to occur in copper deficiency by Bennetts (14), Cunningham (31), Davis (39), and Marston (112). The existence of such low liver copper levels with no evidence of deficiency in these cows indicates that this species can tolerate



a wider range than indicated by the above investigators.

It is evident from these data that the feeding of molybdate lowered the liver copper and serum copper levels. These results are in good agreement with those reported by Cunningham (37) and Mylrea (128) in cattle. Dick (52) working with sheep reported that molybdenum had no effect on serum and liver copper levels unless sufficient inorganic sulfate was present in the diet. Analysis of hay fed during Experiment I revealed a copper content of 1.85 to 3.40 p.p.m., and a sulfur content of 0.16 to 0.26 per cent. The grain used in Experiment I had a copper content of 8.9 to 10.3 p.p.m., and a sulfur content of 0.28 to 0.34 per cent. The analysis of water showed only a trace of copper which was negligible in relation to the amount found in the feed. The overall estimates in the diet were for copper of 4.5 to 6.0 p.p.m. and for sulfur of 0.25 to 0.31 per cent. This level of copper was considered just adequate to maintain normal health in dairy cattle. The sulfur level was considered more than adequate but not unusually high. Unfortunately, the amount of sulfur as inorganic sulfate was not determined and therefore no comparison can be made with the work of Dick (52).

Because both liver and serum copper levels were lowered, it could be reasoned that sufficient inorganic sulfate was present to effect the molybdenum-copper interaction. If this were so, however, molybdenum toxicity symptoms should have been found in these cattle, but no such symptoms were observed. Cunningham (37) and Mylrea (128) reported similar

results from feeding levels up to 16 p.p.m. of molybdenum.

These data indicate that other unidentified conditions necessary for development of molybdenum toxicity were not present. It could be reasoned that the level of inorganic sulfate was not high enough to alter liver and serum copper to a point where toxicity symptoms could occur. A second possibility is that molybdenum was not in the proper form to cause the toxicity. Ferguson et al. (63) reported that 70 to 80 per cent of the molybdenum found in pasture plants was in the form of soluble molybdate. This information indicates that the molybdate salt should be equally as effective as plant molybdenum in causing the toxicity.

Experiment II was designed identically to Experiment I except that 0.3% sulfate sulfur was added to the basal diet. The development of achromotrichia and alopecia by the animals which received 50 p.p.m. molybdenum indicated that additional inorganic sulfate was a factor in producing molybdenum toxicity. Analysis of serum, liver, and milk for copper and molybdenum do not explain the role of this inorganic sulfate. Although the serum, liver and milk molybdenum levels of Experiment II were increased due to molybdenum feeding, these increases were small compared to those received in Experiment I. This comparison makes it evident that inorganic sulfate added to the diet alters molybdenum metabolism. Dick (52) has reported identical results from the analysis of serum and liver samples from sheep. Cunningham (37) also showed this effect of sulfate on the serum and liver molybdenum levels in cattle. Ac-

according to Dick (52), inorganic sulfate in the diet lowers the absorption of molybdenum, thus accounting for the lower levels of molybdenum found in the serum and liver.

The copper levels found in the serum and liver samples of this experiment are of even greater interest because achromotrichia and alopecia occurred in the 50 p.p.m. of molybdenum animals. Contrary to the expected results, the liver and serum copper levels of the animals in Experiment II were as high or higher than those of Experiment I. Further evidence that lower serum and liver copper levels are not responsible for the appearance of molybdenum toxicity symptoms is found by comparing the treatments in Experiment II. At the end of 300 days the levels of serum copper and liver copper are about the same for the 20 and 50 p.p.m. of molybdenum with 0.3% sulfate sulfur treatments, while only the 50 p.p.m. of molybdenum with sulfate showed achromotrichia and alopecia. Of even greater significance is the fact that both 100 and 200 p.p.m. of molybdenum treatments had high serum copper levels and not unusually low liver copper levels (10 to 47 p.p.m.) but showed all of the symptoms of severe molybdenum toxicity.

Except for the treatments of 100 and 200 p.p.m. of molybdenum, which are not typical of those found under natural conditions, only mild symptoms of molybdenum toxicity were observed and these occurred only in the 50 p.p.m. of molybdenum treatment. Evidence that additional time was not a factor in the lack of severity this toxicity was provided by

continuing two of the 50 p.p.m. of molybdenum with sulfate animals through a second lactation. These animals developed more marked changes in hair condition, but showed no further evidence of toxicity. These data indicate that factors other than inorganic sulfate are responsible for increasing the toxicity of molybdenum because the level of inorganic sulfate in this experiment far exceeds that found in plants.

In Experiment III the effect of stress and the addition of some ions not found in New Hampshire hays were tested. In many diseases stress is known to be an important factor in causing the onset of symptoms of the malady. Cows were subjected to extreme cold by housing them in a pole type barn and feeding them in an open lot. The results of this experiment indicated that this type of stress had no additional effect on molybdenum toxicity. All animals which received 50 p.p.m. of molybdenum with 0.3% sulfate sulfur developed achromotrichia and alopecia, while animals receiving either 50 p.p.m. of molybdenum or 0.3% sulfate sulfur did not. These results are in complete agreement with those found in Experiments I and II. The addition of lead, arsenic, and rubidium to the diet produced no further increase in molybdenum toxicity, thus indicating that none of these ions are complicating factors.

The development of toxicity symptoms other than achromotrichia and alopecia occurred in one 50 p.p.m. of molybdenum with 0.3% sulfate animal housed indoors. No good explanation for this occurrence can be found. This animal was

small and less rugged looking than the other animals at the start of the experiment, and it is believed that a lack of desirable constitution and vigor was the reason for the greater susceptibility to the experimental treatment. Evidence that the constitution of an animal can influence the incidence of toxicity is reported by Hayashi (82). This worker found that Holstein calves showed less susceptibility to molybdenum toxicity than the native cattle of Japan. The ability of some animals to withstand larger doses of molybdenum than others may account for part of the difference in the occurrence of molybdenum toxicity under natural conditions and experimental conditions. Frequently, molybdenum toxicity has occurred in range areas where less attention has been devoted to animal vigor, whereas animals selected for experimental purposes are generally of good constitution.

In trying to find a reason for differences in the level of molybdenum required to cause toxicity in these experiments and in areas where the malady occurs naturally, several other factors should be considered. One strong possibility is the effect of internal parasites. Bremner (19), Gibson (71), and Seekles (143) have reported a lowering of serum and liver copper by heavy infestations of internal parasites. In many of the areas where molybdenum toxicity has occurred, the climate also has been favorable for internal parasites. Kretschmer and Beardsley (91) reported that many cattle suspected of molybdenum toxicity in Florida responded to a treatment of phenothiazine. Animals in cooler climates and especially if confined in a barn are not normally infected

with parasites. An examination of all the animals on this experiment for internal parasites was negative.

The general state of nutrition may be another factor involved in molybdenum toxicity. Because molybdenum toxicity has occurred frequently in range areas where cattle are fed only grass, a shortage of protein may have been a complicating factor. Miller and Engel reported that the protein level of the diet greatly influenced the development of molybdenum toxicity in rats. The use of a 17% grain mixture in the present experiments provided more than enough protein to meet the animals' requirements. Recently, Davis (40) reported the isolation of an organic complexing agent from grass which reduced the availability of copper to the animal. It is possible that such a complexing agent could reduce the copper absorption and thereby increase molybdenum toxicity.

Extensive analysis of forage from areas where molybdenum toxicity occurs occasionally has shown high levels of zinc. Work with rats reported by Gray and Ellis (73) has shown that this element will cause anemia which can be corrected by copper administration. Because the zinc ion has a size and electronic structure similar to copper, it may act as a biological antagonist to copper. There also is a possibility that some other element or ion present in these forages increases the toxicity of molybdenum.

The question of whether molybdenum without the presence of any inorganic sulfate has an effect on copper metabolism is still not clear. Only Dick (52) has reported that molybdenum without inorganic sulfate does not change serum or liver copper

levels. Because it is very difficult to compound a ration with no inorganic sulfate, this question will be difficult to answer. That inorganic sulfate increases the toxicity of molybdenum has been shown by these experiments and by the work of Dick (52), Cunningham (37), and Mylrea (128). The mechanism by which sulfate brings about this effect is not clear.

The results of Experiment IV indicate that Dick's (52) theory of sulfate prohibiting the transport of molybdenum and copper across a membrane is in error. In four paired trials neither sulfate nor sulfate with molybdenum was shown to have any effect on the absorption of orally administered copper-64 as measured by the level of radioactivity in the blood. These trials indicated that molybdenum reduced the deposition of copper-64 in the liver.

The results of these trials also tend to disprove the theory proposed by Halverson et al (178), that large amounts of sulfide formed from sulfate reduction in the rumen combines with copper to form insoluble copper sulfide, which is unavailable to the animal. Because copper-64 levels of the blood were not affected by sulfate addition to the diet, it appears that the formation of insoluble copper sulfide must have been small. The results of sulfide determinations made on rumen samples during Experiment III indicated large amounts of the ion present in the rumen liquid for the first two hours following the feeding of sulfate. Treatments which received 0.3% sulfate sulfur had higher levels of sulfide

than the treatment which received no sulfate. Differences were not found to be due to addition of molybdenum to the diet, however. In all treatments sulfide in rumen liquid was undetectable a few hours after feeding, and only a trace of sulfide was detectable in the solid portion of rumen content.

It would seem quite probable that much of the copper found in plants would be released after two hours of digestion. If such were the case the formation of copper sulfide would not be appreciable. If this is the case a decrease in the copper level of liver would reflect a negative balance of copper. Using this line of reasoning many of the conditions found in this experiment can be explained. In Experiment I, the liver copper level for all molybdenum treatments was shown to increase during the last 75 days of experiment. These results are consistent because milk production was decreased and therefore less copper was excreted from the body. Similar results were obtained with the animals receiving 50 p.p.m. molybdenum in Experiment III. The liver's serving as a storage organ also would explain why serum copper levels did not drop as rapidly as the liver copper levels. It would not be until the liver had been depleted of all the copper stores that an appreciable decrease of serum copper would be observed.

If liver copper can serve as an indicator of the animal's balance of copper, it may help explain the action of molybdenum on copper. It does not seem probable that molyb-



denum acts as a biological antagonist to copper because chemically these two elements are quite different. Recent work reported by Scaife (139) indicates that molybdates inhibit the activity of certain copper containing enzymes. With such loss of enzyme activity it would seem probable that the body would increase the production of these copper enzymes and thus place a greater demand on the body for copper. The sudden increase in serum copper reported by Dick (52) when molybdenum was fed would indicate that such a body response was taking place. If these enzymes are blocked due to the action of molybdate they probably would be gradually excreted from the body and thus deplete the levels of copper in the body.

Recent reports by Singer (145) indicate a similar blocking of copper containing enzymes by both sulfide and sulfate ions. This action can be used to explain the effect inorganic sulfate has on increasing molybdenum toxicity. Because molybdenum has been shown by Mills et al. (124) to reduce the production of sulfide oxidase, abnormally high levels of sulfide ions could build up in the blood and tie up copper enzymes. The animal would respond to such a loss of enzyme activity by an increase in the production of these enzymes and thus create a greater demand for copper. If such an increased demand for copper placed the animal in a negative balance, the animal would develop copper deficiency.

This mechanism also could be used to explain why high levels of molybdenum cause the symptoms of molybdenum toxicity

to appear before the liver and serum copper levels are below normal. In such conditions the high concentration of both sulfide and molybdenum would tie up the copper enzymes in the body at a rate faster than the body could produce them and hence the symptoms of copper deficiency would appear. At the same time the excretion of these now useless copper containing enzymes may be slower than the rate of blocking and therefore serum and liver copper levels would still be normal or even above normal. The animals which received 100 and 200 p.p.m. of molybdenum in Experiment II were examples of animals showing molybdenum toxicity while having high serum and liver copper levels.

The appearance of achromotrichia and alopecia in animals on a diet of 50 p.p.m. molybdenum with 0.3% sulfate sulfur might be described as the first defense mechanism of the animals. Hair production being a less essential body process may be neglected by the body in preference to more important functions. This condition therefore presents a good indication of the stage of copper deficiency and may be a better index than the level of serum copper. It would appear that the low level of copper found in the liver and serum of calves born to the cows continued through a second lactation while receiving molybdenum and sulfate, was also an effort to conserve copper for vital body processes.

According to such an explanation, molybdenum toxicity causes an increase in rate of copper metabolism which results in a deficiency of this element. This scheme would account

for the fact that copper administered to animals suffering from this malady brings about complete recovery, as was shown in Experiment III.

Evidence against such a mechanism being responsible for the copper-molybdenum-sulfate interrelationship has been reported by Cox et al. (28). These workers fed high dietary levels of molybdenum to rats and calves and determined the effect of this treatment on liver and heart enzymes. The results of these experiments indicated no difference in liver tyrosine oxidase and heart cytochrome oxidase activity between control and experimental groups. It should be pointed out, however, that no sulfate was added to the diets of these animals and there were no toxicity symptoms recorded for the calves. Furthermore, the levels of liver copper in all experimental groups were extremely high, thus indicating that these animals were not suffering from copper deficiency.

In view of the results obtained by Scaife (139) and Mills et al. (124) with molybdenum inhibiting the activity of copper containing enzymes, more research on the effects of both molybdate and ions of sulfur on copper containing enzymes would seem most promising in explaining this interrelationship.

## SUMMARY AND CONCLUSIONS

Four experiments designed to investigate the copper-molybdenum-sulfate sulfur interrelationships in cattle were carried out. In Experiment I, fifteen Holstein first calf heifers were used to observe the effects of adding from 5 to 50 p.p.m. of molybdenum to a diet low in copper, molybdenum, and sulfur. At the end of 300 days on experiment, symptoms of molybdenum toxicity such as achromotrichia, alopecia, diarrhea, emaciation, and anemia were not found for any level of treatment. Animal production as measured by milk production and changes in body weight was not measurably affected by treatment. Analysis of serum, liver, and milk samples from these animals for copper and molybdenum indicated the following effects due to treatment.

1. The serum, liver, and milk molybdenum levels increased with increased molybdenum in the diet.
2. The feeding of molybdenum did not have any effect on the level of copper in the milk.
3. Treatments which received supplemental molybdenum showed decreased liver copper levels during the first 225 days of the experiment. During the last 75 days of experiment these animals showed increases in liver copper which were presumed caused by a lower molybdenum intake due to decreased feed consumption and less copper loss from the body due to decreased milk production.
4. Although the serum copper levels of all molybdenum

treatments decreased during the experimental period, all treatment means were still in the published normal range.

5. The control animals showed increases of serum and liver copper during the experiment.

In Experiment II fifteen first calf heifers were used to observe the effects of adding 0.3% sulfate sulfur to the treatments used in Experiment I. At the end of 225 days on experiment the cows receiving 50 p.p.m. molybdenum developed achromotrichia and alopecia. Continuation of the treatment for the remainder of the first lactation and for an additional lactation for two animals produced no further symptoms of molybdenum toxicity. The treatments receiving less than the 50 p.p.m. molybdenum level did not develop any molybdenum toxicity symptoms. The analysis of liver, serum, and milk for copper and molybdenum indicated the following effects due to adding sulfate to diets containing various levels of molybdenum.

1. The serum, liver, and milk molybdenum levels were lower for each treatment level of molybdenum than were observed in Experiment I. This indicated lower molybdenum absorption.

2. Milk copper levels were significantly lower in this experiment than in Experiment I, but there were no differences among molybdenum treatments.

3. Treatments receiving molybdenum continuously lost liver copper until they reached a low level and these low levels persisted until the end of the experiment. Because

no increase was observed during the latter part of the lactation as was observed in Experiment I, the addition of sulfate appeared to increase the copper deficiency caused by high levels of molybdenum.

4. Serum copper levels for all treatments receiving molybdenum decreased during the experiment, while the level for the treatment which did not receive molybdenum remained constant.

5. Four animals which were fed levels of 100 and 200 p.p.m. of molybdenum with 0.3% sulfate sulfur for 75 days developed achromotrichia, diarrhea, loss of nervous control, and emaciation, and milk production stopped. These symptoms occurred while serum copper levels were high and liver copper levels were not unusually low. These animals did not develop anemia.

6. Calves born to dams showing achromotrichia and alopecia had unusually low serum and liver copper levels. Although these calves appeared healthy, they had brown and white coats instead of black and white.

In Experiment III, fifteen Holstein first calf heifers were used to test the effect of stress and other possible complicating factors on the development of molybdenum toxicity. Nine animals were subjected to extreme cold by being housed in a pole barn and fed in an open lot for 300 days on experiment. The stress of extreme cold had no additional effect on the development of molybdenum toxicity. These nine animals, which were divided into the three treatment, 0.3% sulfate

sulfur, 50 p.p.m. molybdenum with 0.3% sulfate sulfur, and 50 p.p.m. molybdenum, showed identical results with those obtained from Experiment I and II. The results obtained from adding lead, arsenic, and rubidium to a diet containing 50 p.p.m. molybdenum with 0.3% sulfate sulfur indicated that these elements caused no additional effect on the development of molybdenum toxicity. During Experiment III one animal which received 50 p.p.m. molybdenum with 0.3% sulfate sulfur went off feed, became emaciated, and developed anemia after 275 days on experiment. This animal became progressively worse and, after showing no improvement when fed 20 p.p.m. copper for one week, was slaughtered. This occurrence was believed due to a lack of constitution and vigor in the animal. The analyses of serum, liver, and milk for copper and molybdenum were in good agreement with the results obtained from Experiments I and II.

In Experiment IV a series of six trials using copper-64 were carried out to study the effects of sulfate sulfur, molybdenum, and molybdenum with sulfate sulfur on copper metabolism. The data from these trials indicated the following:

1. The level of copper-64 absorbed into the blood is small and is not affected by the presence of high levels of sulfate sulfur, molybdenum, or molybdenum with sulfate sulfur in the diet.
2. The addition of molybdenum to the diet of steers reduced by 70 to 94% the amount of copper-64 deposited in the

in the liver.

3. The amount of copper-64 recovered in the feces of these steers was greater than 96.5% of that fed, while amounts found in the urine was less than 0.5%.

4. Copper-64 appeared in the feces as early as six hours after administration and continued to be excreted for five days.

Several factors are suggested as possible explanation as to why much higher levels of molybdenum are required to produce molybdenum toxicity experimentally than are found in areas where the toxicity occurs, under farm conditions. These factors are low levels of protein in the diet; internal parasites, high levels of zinc, and the presence of complexing agents which would render plant copper unavailable to the animal.

The theory that molybdenum toxicity in ruminants is a conditioned copper deficiency is quite well established. At this time, however, the mechanism by which molybdenum and sulfate sulfur cause this conditioned copper deficiency is not understood. Evidence that copper absorption is not affected by high levels of sulfate and molybdenum in the diet was obtained from the results of Experiment IV. This tends to disprove Dick's (52) theory that sulfate prohibits the transport of molybdenum and copper across membranes. Similarly, the theory of Halverson (78) that copper absorption is reduced by the formation of insoluble copper sulfide also appears to be in error.



Recent evidence by Mills et al. (124), Scaife (139), and Mason (113), indicating that copper-containing enzymes are inactivated by molybdate and sulfide ions, appears much more tenable. A theory based on the assumption of such blocking of copper enzymes by these ions appears to explain the observed facts of molybdenum toxicity.

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## APPENDIX

TABLE 1

Consumption of Sodium Molybdate\*  
(Experiment I)

Animals by treatment	Theoretical quantity to be fed in grams	Actual quantity fed in grams	Per cent error
<b>Control</b>			
282	0	0	0
290	0	0	0
293	0	0	0
<b>Molybdenum 5 p.p.m.</b>			
285	50.7	52.3	3.1
288	45.9	46.6	1.4
292	57.4	57.0	0.6
<b>Molybdenum 10 p.p.m.</b>			
283	100.8	104.1	3.3
287	128.9	129.9	0.8
291	116.6	115.7	0.8
<b>Molybdenum 20 p.p.m.</b>			
281	273.9	272.8	0.4
289	234.3	235.0	0.3
295	213.2	219.9	3.1
<b>Molybdenum 50 p.p.m.</b>			
284	559.3	551.5	1.4
286	551.3	555.6	0.8
294	500.2	497.8	0.5

\*Total for 300 days

TABLE 2

17% Cottonseed Ration  
(Experiments I, II, III, and IV)

800 lb. corn meal	20 lb. dicalcium phosphate
500 lb. crimped oats	20 lb. iodized salt
200 lb. molasses	1 lb. dry vitamin A (5000 units/g)
400 lb. cottonseed meal	1 gm. cobalt carbonate
200 lb. wheat bran	40 gm. irradiated yeast (142F)

TABLE 3

Body Weight in Pounds  
(Experiment I)

Week	Animal number														
	281	282	283	284	285	286	287	288	289	290	291	292	293	294	295
1	1083	871	850	862	948	835	960	913	979	934	928	915	853	985	1025
2	1128	900	852	873	957	825	953	924	934	914	928	940	828	957	1019
3	1087	862	800	880	959	818	919	918	983	930	932	955	792	928	999
4	1078	878	819	881	969	825	926	935	990	926	925	954	773	939	988
5	1095	877	825	900	965	824	929	930	995	924	920	964	779	898	973
6	1073	835	832	895	968	832	935	930	1000	919	919	952	792	927	977
7	1078	855	842	896	977	840	940	914	1005	924	893	960	791	908	949
8	1080	883	858	930	958	843	952	922	982	932	940	976	759	928	977
9	1122	870	853	909	960	829	967	889	974	939	960	948	744	930	980
10	1140	890	864	904	966	845	966	861	1008	958	930	973	788	911	960
11	1140	872	875	895	930	862	962	894	1029	945	916	970	789	923	1000
12	1132	883	869	872	984	863	984	878	1024	959	930	1002	789	937	982
13	1142	914	900	911	996	863	938	934	1034	961	949	965	780	925	996
14	1127	919	919	942	984	893	1006	916	1043	971	951	981	775	945	986
15	1138	915	910	915	992	890	983	904	1055	971	959	964	800	952	1000



TABLE 3 (Cont.)

Body Weight in Pounds  
(Experiment I)

Week	281	282	283	284	285	286	287	288	289	290	291	292	293	294	295
16	1171	919	919	926	1002	887	992	910	1049	973	961	985	773	949	986
17	1187	930	926	910	988	892	1011	917	1056	981	976	996	763	963	995
18	1150	938	923	944	983	896	1005	910	1049	1000	974	968	785	960	1005
19	1156	930	942	919	977	891	1037	923	1056	990	960	982	780	951	1032
20	1181	918	926	933	975	901	1007	925	1054	983	941	932	784	950	972
21	1176	951	943	934	974	893	1030	918	1045	990	962	950	792	965	1008
22	1169	920	952	934	991	895	1025	930	1064	972	959	977	793	971	1029
23	1187	924	945	934	1006	912	1039	916	1056	968	977	982	771	932	1023
24	1163	933	950	957	1004	907	1039	925	1057	990	996	970	770	947	1019
25	1153	943	938	942	990	905	1034	923	1074	991	1000	977	790	931	1045
26	1156	926	942	946	1010	921	1059	940	1074	1005	1012	971	780	961	1039
27	1165	951	961	962	1011	917	1047	946	1075	980	987	992	792	955	1033
28	1174	952	955	955	1030	913	982	947	1057	992	983	1004	802	937	1052
29	1196	941	975	957	995	891	1027	929	1080	1014	1009	1000	794	975	1065
30	1174	920	960	967	1006	911	1094	962	1084	1013	1012	1000	802	972	1065

TABLE 3 (Cont.)

Body Weight in Pounds  
(Experiment I)

Week	Animal number														
	281	282	283	284	285	286	287	288	289	290	291	292	293	294	295
31	1166	951	976	972	1038	919	1092	944	1095	1007	1044	1006	805	971	1061
32	1152	955	981	963	1054	908	1087	964	1130	1000	1031	995	803	952	1040
33	1172	973	919	975	1084	911	1104	963	1107	1006	1048	1009	807	981	1060
34	1188	996	930	977	1071	902	1092	970	1140	1008	1034	1007	832	955	1055
35	1185	985	925	1003	1084	911	1097	982	1106	1021	1045	983	824	971	1053
36	1207	983	914	994	1086	913	1105	968	1141	1008	1044	991	835	980	1072
37	1225	980	927	994	1080	918	1113	965	1117	1014	1062	995	854	922	1085
38	1211	976	935	1000	1085	909	1096	992	1168	1023	1061	985	835	1000	1093
39	1016	998	953	1013	1109	918	1139	1018	1148	1004	1090	982	861	994	1090
40	1206	993	967	1016	1115	912	1137	999	1141	1023	1086	1015	865	1009	1112
41	1222	979	1000	1029	1126	930	1162	1009	1189	1023	1075	1009	891	1039	1120
42	1235	1013	993	1033	1133	929	1155	1000	1168	1060	1126	1032	875	1032	1143
43	1200	1028	1011	1038	1124	935	1129	1015	1198	1053	1126		880	1067	1146
44	1229	1020	1013			940	1164	1042	1204	1080	1160		888	1056	
45						955	1147						867		

**TABLE 4**  
**Average Body Weights in Pounds**  
**(Experiment I)**

Animal Number	Days				
	0	75	150	225	300
281	1103	1134	1171	1164	1219
282	885	877	929	950	1020
283	834	864	940	959	1006
284	875	903	934	967	1033
285	954	952	982	1033	1128
286	831	845	896	913	935
287	958	965	1021	1094	1149
288	918	881	924	957	1019
289	965	1004	1054	1111	1190
290	926	947	982	1004	1045
291	929	935	954	1042	1109
292	937	959	955	1000	1019
293	845	774	785	805	881
294	957	924	962	968	1052
295	1014	980	1003	1054	1136

TABLE 5

Animals by treatment		Total Hay and Grain Consumption in Pounds (Experiment I)			
		Days			
		75	150	225	300
<b>Control</b>					
282	Hay	1342.6	1617.0	1605.7	1549.2
	Grain	941.8	821.7	771.1	655.3
290	Hay	1462.0	1723.9	1783.6	1797.6
	Grain	757.7	781.0	623.1	667.7
293	Hay	1206.3	1250.1	1254.5	1240.2
	Grain	924.0	823.4	672.1	677.2
<b>Molybdenum 5 p.p.m.</b>					
285	Hay	1634.2	1695.2	1722.9	1786.1
	Grain	582.6	554.2	422.1	461.3
288	Hay	1510.4	1602.8	1599.9	1608.9
	Grain	548.7	420.3	342.5	386.3
292	Hay	1496.0	1705.8	1737.0	1787.6
	Grain	863.9	878.2	848.8	704.9
<b>Molybdenum 10 p.p.m.</b>					
283	Hay	1423.2	1750.7	1722.3	1692.7
	Grain	582.9	522.5	547.2	534.0
287	Hay	1680.8	1975.2	1977.1	1933.9
	Grain	849.3	903.8	998.1	905.3
291	Hay	1661.6	1905.7	1967.9	1953.7
	Grain	775.5	789.6	684.1	413.3
<b>Molybdenum 20 p.p.m.</b>					
281	Hay	1849.5	1980.2	2151.2	2208.9
	Grain	1057.7	1044.0	886.7	767.3
289	Hay	1651.4	1920.9	1936.3	1890.0
	Grain	733.4	755.1	692.8	645.5
295	Hay	1386.6	1623.0	1693.0	1765.0
	Grain	902.5	752.5	667.7	509.4
<b>Molybdenum 50 p.p.m.</b>					
284	Hay	1509.9	1877.9	1999.7	2089.6
	Grain	610.5	602.0	592.1	479.5
286	Hay	1456.8	1715.7	1750.4	1747.4
	Grain	710.8	693.9	734.0	612.0
294	Hay	1423.4	1390.8	1376.2	1435.4
	Grain	1022.3	902.4	676.2	502.7

TABLE 6

Hemoglobins  
(Experiment I)

	Animal number														
	281	282	283	284	285	286	287	288	289	290	291	292	293	294	295
Biweekly values	mg/100 ml blood														
1	11.33	11.09	10.47	10.67	10.72	9.82	11.09	10.67	10.54	11.21	10.00	9.64	9.55	10.54	9.33
2	10.24	9.82	9.82	9.76	9.39	9.39	10.61	10.12	10.24	10.24	9.94	10.67	9.03	9.45	9.39
3	9.76	9.64	9.76	9.76	10.12	9.76	10.24	9.09	9.70	11.24	9.58	9.76	8.67	9.15	8.48
4	10.00	9.82	9.39	9.27	9.33	8.85	10.18	10.12	9.33	9.94	8.67	9.64	8.18	9.27	9.45
5	10.18	9.58	8.79	9.39	9.21	8.54	9.70	9.21	9.53	9.33	8.52	10.00	8.48	9.15	9.64
6	9.58	8.73	8.79	9.27	9.27	8.71	9.88	9.27	9.58	9.58	9.30	9.94	8.48	8.85	10.00
7	9.64	8.79	8.91	9.27	9.94	8.97	9.88	8.12	9.82	9.58	9.45	9.94	8.85	8.85	9.52
8	10.42	9.39	8.79	9.52	9.76	8.79	10.00	9.64	10.18	9.58	9.58	10.61	8.54	8.61	9.21
9	10.10	9.50	8.87	10.42	9.52	8.91	10.48	10.12	10.61	9.33	9.21	9.94	8.54	9.21	9.21
10	9.88	9.88	8.97	10.85	10.30	8.97	10.24	10.49	10.00	9.76	9.97	10.18	8.91	9.33	9.15
11	9.88	9.88	9.70	10.00	9.82	8.97	9.94	10.30	10.06	10.16	10.30	10.48	8.91	9.03	9.21
12	10.00	9.58	9.58	10.00	9.52	9.15	10.24	10.61	10.48	10.30	10.30	10.00	8.97	9.21	9.58
13	9.88	9.94	9.88	10.18	9.52	9.21	10.24	10.65	10.48	10.30	9.21	10.30	10.48	9.58	9.50
14	9.94	9.76	9.58	9.88	9.21	9.21	10.06	10.61	10.42	10.91	10.30	11.09	9.09	9.70	9.33
15	9.94	9.64	9.21	10.18	10.36	9.70	10.73	10.73	10.00	10.91	10.61	10.61	9.52	9.33	9.17
16	10.30	9.88	9.88	10.48	10.12	9.09	11.03	10.67	10.18	10.61	10.48	10.48	8.79	9.39	9.21
17	9.39	9.39	9.39	9.70	9.52	9.52	10.00	10.36	9.88	10.30	9.94	10.48	8.54	9.39	9.27
18	9.64	8.61	8.85	9.52	10.06	9.52	10.18	10.18	10.00	10.39	9.88	10.30	8.61	9.45	9.64
19	10.12	9.94	7.21	10.06	9.33	9.33	10.24	10.30	9.58	10.30	9.52	10.61	9.09	9.58	9.09
20	9.82	9.78	8.54	9.88	9.21	8.79	9.52	10.30	9.15	9.70	9.70	10.79	8.42	9.68	9.09
21	9.82	9.70	9.03	9.76	9.39	9.21	10.48	9.70	9.70	10.00	9.70	10.30	8.36	9.21	9.39

**TABLE 7**  
**Hematocrits**  
**(Experiment I)**

		Animal number														
		281	282	283	284	285	286	287	288	289	290	291	292	293	294	295
Biweekly values		%														
Period IV																
1	31.5	32.9	30.6	34.2	29.3	31.0	33.0	34.0	32.0	34.0	34.0	32.9	31.5	30.6	29.3	
2	34.0	29.8	29.0	32.3	34.0	30.0	36.5	37.4	29.8	32.0	33.0	33.0	30.5	30.0	29.0	
3	31.5	30.0	29.0	30.5	31.5	29.5	37.0	34.0	32.0	36.5	34.5	37.5	33.0	30.0	29.5	
4	31.5	31.0	30.0	31.0	31.5	29.0	34.0	33.0	32.0	33.0	33.0	35.0	31.0	30.0	29.0	
5	32.0	31.5	33.0	32.5	31.5	30.0	33.5	32.0	30.5	33.5	35.0	34.5	32.0	30.0	29.0	
6	32.5	30.0	29.0	31.0	30.5	30.0	31.0	32.0	29.5	33.5	33.0	34.0	30.0	31.5	29.5	
7	33.5	30.0	28.5	31.5	30.0	29.0	31.5	33.0	28.5	33.5	32.5	34.0	27.0	31.5	30.0	

**TABLE 8**  
**Serum Protein**  
**(Experiment I)**

		Animal number														
		281	282	283	284	285	286	287	288	289	290	291	292	293	294	295
Monthly values		%														
period IV																
1	7.8	7.3	7.0	7.8	7.4	8.0	8.0	6.4	7.2	7.2	7.2	7.9	7.0	7.9	7.8	
2	7.1	7.4	9.2	7.4	7.5	7.4	8.0	6.6	7.2	7.0	7.4	7.9	7.0	8.0	7.8	
3	7.1	7.8	8.2	7.3	7.5	7.7	8.0	6.7	7.0	7.6	7.5	8.2	7.4	7.6	7.6	

TABLE 9

Inorganic Phosphorous  
(Experiment I)

	Animal number														
	281	282	283	284	285	286	287	288	289	290	291	292	293	294	295
Biweekly values	mg/100 ml blood														
Periods III & IV															
1	5.2	6.3	6.8	5.4	5.8	7.8	6.2	6.7	3.2	5.3	4.5	4.5	5.4	7.0	6.3
2	6.0	6.8	6.8	6.0	7.2	8.8	6.4	7.1	5.0	5.5	6.1	7.5	6.0	6.3	5.8
3	6.4	6.3	6.5	6.0	7.3	7.8	7.0	7.1	5.0	5.7	6.2	7.0	6.5	6.2	6.0
4	5.8	6.3	6.7	6.0	7.0	7.3	6.9	6.1	4.5	5.8	6.3	6.1	6.8	5.8	6.0
5	6.2	6.4	6.3	6.7	6.8	7.4	7.3	7.5	5.2	5.9	6.8	6.4	6.0	5.7	6.5
6	6.8	7.2	6.3	8.0	7.4	8.0	7.2	8.0	5.6	7.8	7.0	5.8	5.8	7.0	6.8
7	6.2	7.4	8.0	6.4	6.2	8.2	7.4	7.6	5.8	6.2	6.8	6.6	6.4	8.4	7.4
8	6.8	8.2	8.2	6.4	7.0	8.8	7.2	8.2	4.0	6.4	7.6	6.0	5.4	8.0	7.6

TABLE 10

Composition of Milk  
(Experiment I)

		Milk copper				Milk molybdenum			
		75	150	225	300	75	150	225	300
Control									
	75								
	282	---	0.11	0.33	0.07	---	0.00	0.04	0.06
	290	0.38	0.18	0.08	0.04	0.05	0.02	0.04	0.04
	293	0.18	0.52	0.07	0.06	0.01	0.05	0.04	0.04
Molybdenum 5 p.p.m.									
	285	---	0.16	0.38	0.10	---	0.07	0.07	0.04
	288	0.17	0.20	0.80	0.05	0.02	0.17	0.10	0.17
	292	0.16	0.18	0.06	0.04	0.01	0.05	0.07	0.11
Molybdenum 10 p.p.m.									
	283	---	0.13	0.32	0.05	---	0.02	0.11	0.21
	287	0.16	0.12	0.80	0.04	0.02	0.13	0.06	0.10
	291	0.14	0.23	0.10	0.07	0.06	0.11	0.11	0.27
Molybdenum 20 p.p.m.									
	281	---	0.11	0.18	0.06	---	0.20	0.36	0.48
	289	0.17	0.14	0.07	0.04	0.05	0.21	0.20	0.18
	295	0.25	0.21	0.04	0.06	0.15	0.56	0.36	0.84
Molybdenum 50 p.p.m.									
	284	---	0.12	0.44	0.04	---	0.51	1.41	1.05
	286	0.12	0.16	0.79	0.02	0.18	0.72	0.50	0.66
	294	0.17	0.21	0.07	0.06	0.19	0.80	0.94	2.91



TABLE 11

Composition of Serum  
(Experiment I)

	Serum copper (p.p.m.)			Serum molybdenum (p.p.m.)		
	75	150	300	75	150	300
Control						
282	1.11	0.95	0.69	0.01	0.03	0.03
290	1.35	0.89	1.00	0.08	0.01	0.05
293	0.90	0.83	1.53	0.11	0.06	0.02
Molybdenum 5 p.p.m.						
285	1.24	1.10	0.76	0.24	0.12	0.21
288	0.63	0.85	0.66	0.16	0.45	0.65
292	0.93	0.89	0.82	0.12	0.13	0.29
Molybdenum 10 p.p.m.						
283	0.93	0.40	0.44	0.42	0.41	0.86
287	0.43	0.60	0.58	0.16	0.43	0.30
291	1.05	0.98	0.83	0.51	0.34	0.75
Molybdenum 20 p.p.m.						
281	1.11	0.73	0.49	0.69	0.50	1.96
289	0.83	0.70	0.69	0.42	0.72	1.00
295	0.95	0.78	0.78	1.23	1.98	1.96
Molybdenum 50 p.p.m.						
284	1.70	1.08	0.64	3.90	1.50	3.95
286	0.60	0.65	0.50	1.50	2.40	2.24
294	1.00	0.85	0.66	1.60	2.30	5.65

TABLE 12

Composition of Liver  
(Experiment I)

	Liver copper (p.p.m.)				Liver molybdenum (p.p.m.)			
Control	75	150	225	300	75	150	225	300
282	24	28	44	51	35	4	6	6
290	115	129	175	162	6	7	8	7
293	119	123	148	222	12	3	5	5
Molybdenum 5 p.p.m.								
285	206	197	165	158	15	9	8	5
288	102	79	55	29	5	13	10	9
292	17	21	12	21	10	7	10	29
Molybdenum 10 p.p.m.								
283	7	10	12	189	8	16	14	23
287	27	12	21	14	7	10	9	8
291	87	67	56	34	17	12	10	12
Molybdenum 20 p.p.m.								
281	25	17	20	22	26	18	21	23
289	26	12	--	9	20	19	--	13
295	106	81	33	157	19	25	19	24
Molybdenum 50 p.p.m.								
284	34	20	15	16	23	27	31	21
286	20	54	24	12	16	27	22	23
294	42	27	14	72	28	35	33	44

TABLE 13

**Administration of Sodium Molybdate  
(Experiment II)**

<b>Animals by treatment</b>	<b>Theoretical quantity to be fed in grams</b>	<b>Actual quantity fed in grams</b>	<b>Per cent error</b>
<b>Control</b>			
386	0	0	0
395	0	0	0
398	0	0	0
<b>Molybdenum 5 p.p.m.</b>			
390	29.2 **	27.7	5.0
392	48.7 ***	46.1	5.4
399	27.7 **	26.3	4.9
<b>Molybdenum 10 p.p.m.</b>			
388	80.3	78.5	2.2
391	80.0	77.2	3.4
396	86.1	88.5	2.8
<b>Molybdenum 20 p.p.m.</b>			
389	238.2	232.1	2.6
393	216.7	206.9	4.5
397	175.5	177.0	0.9
<b>Molybdenum 50 p.p.m.</b>			
387	486.6	490.9	0.9
394	631.5	614.7	2.7
400	611.5	625.6	2.3
<b>Molybdenum 100 p.p.m.</b>			
388	176.4*	162.6	7.8
392	210.7*	193.0	8.4
<b>Molybdenum 200 p.p.m.</b>			
391	278.8*	280.3	0.5
396	343.0*	410.3	19.6

\* one period  
 \*\* two periods  
 \*\*\* three periods

TABLE 14

Body Weight in Pounds  
(Experiment II)

	Animal number														
Week	386	387	388	389	390	391	392	393	394	395	396	397	398	399	400
1	1006	967	886	958	880	919	1025	900	1180	1060	1010	872	875	959	969
2	1014	955	883	1029	851	894	1011	891	1144	1055	1011	887	877	924	974
3	1048	979	900	1035	865	925	1034	915	1144	1028	981	874	865	891	946
4	1010	937	877	1010	851	915	1060	914	1151	1006	975	894	835	901	974
5	1022	958	900	1045	869	913	1042	920	1060	982	985	890	876	868	954
6	1030	954	910	1037	825	914	1150	906	1162	1000	996	888	866	883	979
7	998	928	910	1036	864	914	1018	881	1164	1030	974	889	875	865	941
8	1022	936	919	1034	882	909	1050	881	1155	1011	1010	911	852	860	975
9	926	943	908	918	857	904	1050	879	1198	1013	998	907	873	877	995
10	1048	950	923	1013	860	921	1053	885	1215	1005	1005	920	867	869	989
11	1050	959	900	1027	859	927	1053	898	1184	1044	987	909	885	876	953
12	979	936	921	1003	868	921	1066	915	1184	1007	988	906	856	882	1006
13	1052	994	932	1035	868	947	1023	872	1180	1001	994	927	875	868	950
14	1011	974	936	1010	870	916	1038	873	1138	1008	983	925	842	874	957
15	1050	991	932	1018	860	921	1046	896	1156	1017	981	945	868	886	959

TABLE 14 (Cont.)

Body Weight in Pounds  
(Experiment II)

	Animal number														
Week	386	387	388	389	390	391	392	393	394	395	396	397	398	399	400
16	1027	962	877	982	889	929	1030	886	1171	996	1003	955	855	877	977
17	1006	970	915	996	873	900	1035	890	1175	1012	962	921	847	867	947
18	1021	991	924	1004	848	918	1040	898	1183	991	993	950	835	888	985
19	1036	967	918	998	883	920	1044	900	1175	1022	991	931	858	903	955
20	1031	982	923	980	882	912	1047	902	1188	1007	950	940	862		970
21	1050	991	925	973	880	936	1050	911	1160	1020	952	920	873		977
22	991	982	915	947	881	916	1048	903	1206	1014	981	923	873		956
23	1033	969	918	975		927	1041	817	1189	1017	963	917	838		971
24	1011	986	910	953		920	1038	920	1198	1025	1008	919	867		939
25	1025	979	934	975		938	1058	937	1110	1051	971	927	833		916
26	988	968	926	963		953	1065	947	1227	1003	969	909	851		924
27	1002	996	954	985		943	1040	956	1224	1021	971	934	849		951
28	1046	1007	945	995		935	952	964	1198	1024	967	898	852		955
29	1035	1016	954	1010		940	1050	955	1223	998	930	892	857		940
30	983	972	931	984		930	1058	978	1161	982	959	882	870		950

**TABLE 14 (Cont.)**  
**Body Weight in Pounds**  
**(Experiment II)**

	Animal number														
Week	386	387	388	389	390	391	392	393	394	395	396	397	398	399	400
31	1026	998	943	997		917	1044	950	1198	1033	965	900	845		939
32	991	995	945	980		939	1042	1008	1213	1041	965	890	833		938
33	1001	950	918	952		955	1047	1013	1217	1011	945	915	864		933
34	1005	900	932	937		915	1060	966	1212	1040	861	883	858		935
35	1029	934	932	957		904	1051	1020	1230	1038	926	884	842		945
36	1020	932	914	940		769	983	1016	1210	1044	900	877	855		933
37	1022	924	909	967		815	845	1008	1185	1065	884	876	832		930
38	1035	949	845	957		839	998	1014	1192	1045	846	885	834		950
39	1048	955	832	963		785	1001	1029	1228	1058	848	866	835		922
40	1053	970	848	977		768	977	1038	1225	1095	842	864	833		991
41	1073	959	860	961		723	971	1063	1233	1072	818	904	862		970
42	1077	1007	840	965		697	1000	1036	1210	1075	826	891	845		990
43	1057	988	804	922		735	1024	1066	1197	1107	847	895			1000

TABLE 14 (Cont.)

Body Weight in Pounds  
Animals Continued Through a Second Lactation  
(Experiment II)

		Animal number							
Week	386	387	394	395	Week	386	387	394	395
1		1026	1158	1109	23	1145	982	1168	1085
2	1177	1004	1190	1109	24	1180	951	1209	1054
3	1154	1012	1218	1085	25	1129	975	1195	1072
4	1113	1022	1175	1092	26	1127	970	1178	1074
5	1127	1031	1187	1085	27	1127	979	1229	1099
6	1137	1017	1174	1102	28	1144	982	1190	1083
7	1140	1008	1182	1092	29	1123	967	1244	1109
8	1130	1010	1198	1067	30	1133	965	1214	1100
9	1156	1008	1195	1053	31	1141	938	1212	1009
10	1153	994	1208	1055	32	1197	994	1242	1032
11	1134	1027	1175	1069	33	1223	986	1218	1049
12	1114	1006	1176	1085	34	1200	986	1243	1071
13	1123	1032	1183	1036	35	1210	998	1158	1120
14	1143	1030	1183	1073	36	1230	988	1177	1129
15	1151	992	1160	1083	37	1235	1004	1198	1122
16	1200	1000	1205	1109	38	1236	1000	1200	1048
17	1129	998	1214	1045	39	1264	1011	1198	1066
18	1104	1003	1231	1070	40	1300	1006	1171	1074
19	1110	1006	1155	1074	41	1289	993	1149	1050
20	1129	1012	1163	1070	42	1300	989	1100	1052
21	1135	943	1210	1098	43	1300	1000	1149	1052
22	1155	1073	1174	1102					

**TABLE 15**  
**Average Body Weights in Pounds**  
**(Experiment II)**

Animal Number	Days				
	0	75	150	225	300
386	1023	1026	1024	1000	1069
387	967	948	985	988	985
388	890	915	921	940	835 *
389	1007	1014	967	987	949
390	865	862	881		
391	913	923	921	928	718 **
392	1023	1059	1048	1048	998 *
393	902	899	905	979	1055
394	1156	1194	1185	1174	1213
395	1048	1019	1014	1019	1085
396	1001	993	961	963	830 **
397	881	912	928	891	897
398	872	869	869	849	847
399	926	876	886		
400	963	983	969	942	987

\* Changed to 100 p.p.m.

\*\* Changed to 200 p.p.m.

**Animals Continued Through A Second Lactation**

386	1166	1134	1140	1157	1296
387	1014	1009	1009	966	994
394	1189	1186	1182	1223	1133
395	1101	1070	1090	1047	1051



TABLE 16

Total Hay and Grain Consumption in Pounds  
(Experiment II)

Animals by treatment	Days			
	75	150	225	300
<b>Control</b>				
386 Hay	2004.7	2335.3	2181.6	2187.3
Grain	768.7	742.9	659.6	520.4
395 Hay	1810.4	1928.5	1925.6	1954.1
Grain	1011.2	1018.1	778.0	509.6
398 Hay	1469.2	1559.4	1484.4	1474.9
Grain	734.5	410.4	303.5	178.1
<b>Molybdenum 5 p.p.m.</b>				
390 Hay	1769.0	1837.1		
Grain	853.3	643.5		
392 Hay	1864.5	2240.0	2249.3	
Grain	893.6	712.9	540.4	
399 Hay	1474.8	1462.6		
Grain	1083.9	809.4		
<b>Molybdenum 10 p.p.m.</b>				
388 Hay	1706.6	1770.4	1828.7	
Grain	694.1	528.8	483.2	
391 Hay	1594.1	1685.6	1595.1	
Grain	834.6	702.3	567.1	
396 Hay	1815.3	1761.5	1791.1	
Grain	1181.1	799.8	163.0	
<b>Molybdenum 20 p.p.m.</b>				
389 Hay	1870.0	2037.5	1918.6	1817.1
Grain	904.1	843.1	619.9	402.1
393 Hay	1811.5	1798.9	1880.4	1829.2
Grain	728.0	656.0	459.1	299.9
397 Hay	1349.9	1499.9	1423.3	1205.3
Grain	840.5	634.9	394.0	317.8
<b>Molybdenum 50 p.p.m.</b>				
387 Hay	1701.1	1801.7	1779.6	1721.8
Grain	703.2	437.5	286.0	70.0
394 Hay	1971.9	1929.2	1925.4	2152.0
Grain	956.9	867.4	758.3	470.4
400 Hay	1914.7	1905.7	1596.6	1827.1
	1283.0	1071.3	591.3	492.5

TABLE 16 (cont.)

Total Hay and Grain Consumption in Pounds  
(Experiment II)

Animals by treatment	Days			
	75	150	225	300
Molybdenum 100 p.p.m.				
388 Hay				1283.9
Grain				257.2
392 Hay				1610.7
Grain				229.8
Molybdenum 200 p.p.m.				
391 Hay				984.8
Grain				232.9
396 Hay				1298.1
Grain				200.0

TABLE 17

Hemoglobins  
(Experiment II)

	Animal number														
	386	387	388	389	390	391	392	393	394	395	396	397	398	399	400
Biweekly values	mg/100 ml blood														
1	10.97	10.85	9.76	10.97	10.91	10.91	10.73	10.91	10.06	10.36	10.24	10.12	10.18	10.48	10.82
2	10.61	10.51	9.76	10.32	10.00	10.79	10.24	10.61	9.45	9.21	9.39	9.94	9.33	10.00	10.54
3	9.52	10.18	8.97	10.30	9.21	10.00	9.64	10.42	9.21	9.94	10.00	9.45	8.79	9.39	10.00
4	8.91	10.00	9.39	9.76	9.76	9.94	10.48	10.91	9.09	10.48	9.58	8.42	9.03	9.03	10.00
5	9.32	10.06	9.03	9.15	11.03	10.18	10.79	9.82	9.52	9.39	8.79	8.12	8.97	10.00	9.94
6	10.36	10.36	9.94	10.79	9.58	9.27	10.12	9.82	9.21	9.33	9.15	8.97	8.79	9.09	9.94
7	9.15	9.82	8.91	9.82	10.00	8.91	9.82	9.76	8.24	9.82	9.82	9.09	7.88	8.79	8.79
8	9.82	9.88	8.91	9.52	9.58	8.97	10.12	9.76	8.76	9.76	9.82	7.21	8.67	9.52	9.09
9	9.09	9.82	9.09	9.45	9.09	8.67	10.06	9.52	8.73	8.42	8.18	8.73	8.79	9.70	9.82
10	9.03	9.82	9.21	9.76	8.61	7.88	8.91	8.79	7.58	9.15	9.09	8.67	8.18	9.94	9.09
11	8.79	9.27	8.00	9.09	9.52	9.70	9.09	9.64	8.73	9.39	9.27	7.94	8.00		9.09
12	9.27	9.58	9.15	9.76	9.88	9.70	9.39	10.00	8.18	8.48	8.91	8.54	8.48		8.91
13	9.39	9.76	9.09	9.82	9.70	10.30	9.58	9.58	8.48	8.48	8.48	8.67	8.67		9.70
14	9.09	9.70	9.15	10.00		9.76	9.76	10.24	8.18	8.30	8.97	8.42	8.79		8.97
15	9.39	9.27	9.39	10.00		9.64	10.00	10.61	8.48	8.54	8.91	8.06	8.73		8.54
16	9.40	9.52	9.40	10.00		9.45	9.94	10.00	8.54	8.48	9.09	8.06	8.30		8.48
17	9.64	9.76	9.52	10.42		8.85	9.45	9.76	8.79	8.00	8.67	8.18	8.18		9.03
18	9.76	9.82	9.39	10.52		8.67	9.15	9.21	8.00	8.88	8.48	8.00	8.12		8.91
19	9.52	9.58	9.39	10.00		8.97	8.79	9.39	8.24	8.48	7.88	8.12	8.42		8.91
20	9.21	8.73	8.54	9.64		9.21	8.67	8.30	8.18	8.00	9.09	8.00	8.61		9.09
21	8.30	8.36	8.18	9.76		9.60	8.71	8.85	8.30	8.85	8.06	8.61	8.92		9.03

TABLE 18

Hematocrits  
(Experiment II)

Biweekly values	Animal number																			
	386	387	388	389	390	391	392	393	394	395	396	397	398	399	400					
1	33.5	34.5	31.5	32.5	35.0	38.5	35.0	37.0	30.0	34.5	33.5	30.0	28.5	34.0	28.5	35.5				
2	30.5	34.5	30.0	31.5	31.5	33.5	35.0	33.5	30.0	31.5	30.0	30.0	28.5	34.0	28.5	35.5				
3	29.5	30.0	29.5	31.5	32.0	31.0	32.5	31.5	28.0	29.5	29.1	29.5	27.5	33.0	27.5	31.5				
4	27.0	30.0	29.0	31.5	31.0	30.8	32.0	32.0	30.5	32.8	29.5	27.5	29.0	30.5	29.0	30.5				
5	32.0	31.0	32.0	33.5	30.5	31.0	33.5	32.5	27.5	30.5	28.0	27.0	24.5	31.0	27.5	31.0				
6	29.0	33.0	29.0	32.0	32.5	30.0	33.0	31.5	27.0	31.5	30.0	25.0	27.5	29.0	27.5	31.5				
7	31.0	32.0	29.0	31.0	31.5	28.5	32.0	29.5	25.0	29.0	27.5	27.0	29.0	32.0	29.0	31.0				
8	28.0	32.0	28.5	31.0	32.5	28.0	30.0	32.5	27.5	29.0	29.0	27.0	28.5	32.0	28.5	30.5				
9	28.0	32.5	29.5	33.5	32.5	30.0	30.5	33.0	28.5	30.0	29.5	27.0	26.5	31.5	26.5	29.5				
10	31.0	32.5	29.5	34.0	33.0	30.0	33.5	33.0	28.5	29.5	29.5	27.0	26.5	32.5	26.5	29.5				
11	31.0	33.5	31.5	34.0	33.0	30.5	34.0	33.0	28.5	29.5	29.5	27.0	26.5	32.5	26.5	29.5				
12	30.5	33.5	31.5	34.0	34.0	32.5	35.0	33.0	27.5	28.5	29.5	27.0	26.5	32.5	26.5	29.5				
13	29.5	31.5	32.0	33.0	34.0	31.5	34.0	34.0	27.5	27.0	27.5	27.0	26.5	32.5	26.5	29.5				
14	29.5	30.5	32.0	32.0	32.0	33.0	33.0	34.5	27.5	26.5	27.5	27.0	26.5	32.5	26.5	29.5				
15	29.5	30.5	31.0	32.5	32.0	32.0	32.0	34.5	27.5	26.5	27.5	27.0	26.5	32.5	26.5	29.5				
16	29.5	30.5	31.0	32.5	32.0	32.0	32.0	34.5	27.5	26.5	27.5	27.0	26.5	32.5	26.5	29.5				
17	30.0	30.5	30.5	34.5	33.0	29.5	33.0	31.5	27.0	25.0	25.0	27.0	25.0	32.5	25.0	29.5				
18	29.5	30.5	28.5	31.5	30.5	30.5	30.5	31.5	26.0	26.5	26.5	27.0	25.0	32.5	26.5	29.5				
19	29.5	28.5	28.0	31.5	30.5	30.5	30.5	28.5	26.0	25.5	26.5	23.5	24.5	30.5	28.5	29.5				
20	29.5	26.5	25.0	31.5	31.5	30.0	31.5	28.5	26.0	25.5	26.5	23.5	24.5	30.5	28.5	29.5				
21	28.5	28.5	25.0	31.5	29.0	27.0	29.0	29.5	26.0	28.0	26.5	23.5	24.5	30.5	28.5	29.5				

TABLE 19  
 Animals Continued Through a Second Lactation  
 (Experiment II)

Biweekly values	Animal number				Hematocrits			
	386	387	394	395	386	387	394	395
	Hemoglobins mg/100 ml blood				%			
1	9.64	9.82	9.70	9.21	30.5	31.5	31.0	29.0
2	10.18	10.48	8.79	9.45	30.0	32.0	30.0	30.0
3	10.36	12.18	8.61	9.52	28.5	33.0	25.0	31.0
4	10.06	11.76	8.61	9.52	29.0	33.0	26.0	30.0
5	9.21	10.18	8.54	9.33	29.5	32.0	25.5	30.0
6	9.27	10.54	8.48	8.97	29.0	31.0	25.5	30.0
7	9.64	9.64	8.97	8.54	29.0	32.0	26.0	30.0
8	9.33	9.94	8.54	8.48	29.0	32.0	27.0	30.5
9	9.64	9.94	8.48	8.48	29.0	32.5	28.0	30.0
10	9.64	9.88	9.64	9.52	30.5	31.0	28.5	31.0
11	9.70	9.94	9.70	9.76	30.0	33.0	29.0	30.0
12	9.83	10.65	9.22	9.84	30.5	33.0	29.5	30.0
13	9.94	10.85	9.70	9.84	30.0	34.0	29.0	30.0
14	10.00	10.54	9.76	9.64	30.0	35.0	29.0	30.0
15	10.00	10.97	9.07	9.68	31.0	33.0	29.0	30.0
16	10.52	10.54	9.64	9.70	32.0	30.0	29.0	30.5
17	10.67	10.30	9.09	10.06	33.0	28.5	28.5	30.5
18	10.48	9.45	9.09	9.33	32.5	30.0	28.5	29.5
19	10.00	9.33	9.15	9.09	32.0	30.0	29.0	29.0
20	9.52	8.48	9.09	9.09	29.5	28.5	29.0	28.5

TABLE 20  
Composition of Milk  
(Experiment II)

Animals by treatment	Milk copper p.p.m.					Days	Milk molybdenum p.p.m.				
	0	75	150	225	300		0	75	150	225	300
<b>Sulfate sulfur 0.3%</b>											
386	0.04	0.01	0.02	0.05	0.04		0.10	0.00	0.01	0.07	0.01
395	0.06	0.02	0.06	0.03	0.04		0.03	0.08	0.04	0.03	0.02
398	0.13	0.05	0.05	0.02	0.02		0.02	0.05	0.00	0.04	0.09
<b>Sulfate sulfur 0.3%, molybdenum 5 p.p.m.</b>											
390	0.28	0.01	0.02				0.10	0.01	0.03		
392	0.09	0.04	0.03	0.05			0.07	0.07	0.04	0.02	
399	0.00	0.01	0.11				0.01	0.07	0.41		
<b>Sulfate sulfur 0.3%, molybdenum 10 p.p.m.</b>											
388	0.12	0.01	0.04	0.06			0.03	0.06	0.33	0.02	
391	0.18	0.02	0.01	0.05			0.12	0.05	0.04	0.08	
396	0.06	0.00	0.06	0.02			0.03	0.11	0.11	0.02	
<b>Sulfate sulfur 0.3%, molybdenum 20 p.p.m.</b>											
389	0.20	0.04	0.00	0.03	0.03		0.02	0.04	0.09	0.05	0.09
393	0.11	0.01	0.09	0.03	0.07		0.01	0.05	0.15	0.19	0.13
397	0.08	0.09	0.09	0.02	0.03		0.04	0.04	0.07	0.06	0.03
<b>Sulfate sulfur 0.3%, molybdenum 50 p.p.m.</b>											
387	0.08	0.02	0.13	0.05			0.03	0.14	0.17	0.20	
394	0.09	0.02	0.14	0.00	0.07			0.12	0.27	0.20	
400	0.11	0.14	0.09	0.03	0.04		0.03	0.06	0.14	0.25	0.18

TABLE 20 (Cont.)  
 Composition of Milk  
 (Experiment II)

Animals by treatment	Milk copper p.p.m.					Days	Milk molybdenum p.p.m.				
	0	75	150	225	300		0	75	150	225	300
Sulfate sulfur 0.3%, molybdenum 100 p.p.m.											
388					0.02						0.07
392					0.00						0.00
Sulfate sulfur 0.3%, molybdenum 200 p.p.m.											
391					0.05						0.33
396					0.06						0.91

Animals Continued Through A Second Lactation

Sulfate sulfur 0.3%											
386	0.01	0.00	0.05			0.00	0.02	0.05	0.13		
395	0.12	0.04	0.00	0.08	0.03	0.06	0.04	0.01	0.07	0.12	
Sulfate sulfur 0.3%, molybdenum 50 p.p.m.											
387	0.02		0.01	0.05		0.25		0.04	0.07		
394	0.04	0.02	0.02	0.06	0.07	0.10	0.44	0.25	0.32	0.06	

TABLE 21

Composition of Serum  
(Experiment II)

Animals by treatment	Serum copper p.p.m.					Days	Serum molybdenum p.p.m.				
	0	75	150	225	300		0	75	150	225	300
Sulfate sulfur 0.3%											
386	1.1	0.9	1.0	1.0	0.7		0.00	0.03	0.02	0.03	0.29
395	0.8	0.7	0.7	0.6	0.9		0.01	0.00	0.03	0.00	0.42
398	0.7	0.6	0.6	0.6	0.52		0.04	0.05	0.00	0.00	0.00
Sulfate sulfur 0.3%, molybdenum 5 p.p.m.											
390	1.0	0.9	0.8				0.01	0.09	0.02		
392	0.8	0.9	0.8	0.6			0.02	0.06	0.08	0.18	
399	0.8	0.9	0.7				0.02	0.05	0.07		
Sulfate sulfur 0.3%, molybdenum 10 p.p.m.											
388	0.6	0.5	0.4	0.6			0.02	0.26	0.05	0.22	
392	0.9	0.9	0.6	0.7			0.02	0.48	0.18	0.58	
399	1.2	0.8	0.9	0.6			0.01	0.18	0.61	0.15	
Sulfate sulfur 0.3%, molybdenum 20 p.p.m.											
389	1.1	0.8	0.6	0.7	0.8		0.02	0.30	0.28	0.41	0.22
393	0.9	1.1	1.1	0.7	0.6		0.02	0.30	0.88	1.2	0.67
397	0.8	0.7	0.5	0.5	0.24		0.03	0.20	0.48	0.23	0.18
Sulfate sulfur 0.3%, molybdenum 50 p.p.m.											
387	1.0	1.1	1.6	0.6	0.7		0.02	0.76	0.63	0.72	1.0
394	0.7	0.9	0.6	0.6	0.7		0.01	0.72	1.3	1.00	1.42
400	0.9	1.3	1.3	0.9	0.70		0.02	0.55	0.78	0.61	0.66



TABLE 21 (Cont.)

Composition of Serum  
(Experiment II)

Animals by treatment	Serum copper p.p.m.					Days	Serum molybdenum p.p.m.				
	0	75	150	225	300		0	75	150	225	300
Sulfate sulfur 0.3%, molybdenum 100 p.p.m.											
388		1.0					0.78				
392		0.9					1.70				
Sulfate sulfur 0.3%, molybdenum 200 p.p.m.											
391		0.9					1.3				
396		0.9					2.9				

Animals Continued Through A Second Lactation

Sulfate sulfur 0.3%											
386	0.74	0.74	0.96	1.11	1.50	0.02	0.00	0.00	0.00	0.00	0.00
395	0.75	0.72	0.71	0.68	0.78	0.0	0.00	0.01	0.03	0.00	0.00
Sulfate sulfur 0.3%, molybdenum 50 p.p.m.											
387	0.40		0.52	0.43	0.36	0.64		0.16	0.18		
394	0.52	0.68	0.68	0.48	0.60	0.39	1.34	0.71	1.47	0.33	
Calves of: Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (at birth)											
387			0.00					0.13			
394			0.05					0.46			
Sulfate sulfur 0.3% (at birth)											
386			0.30					0.05			
395			0.16					0.00			

TABLE 22

Composition of Liver  
(Experiment II)

Animals by treatment	Liver copper p.p.m.					Days	Liver molybdenum p.p.m.				
	0	75	150	225	300		0	75	150	225	300
Sulfate sulfur 0.3%											
386	111	198	158	120	124	4	1	2	40	8	
395	40	70	107	108	75	2	2	9	7	29	
398	13	36	28	26	170	11	1	44	7	2.5	
Sulfate sulfur 0.3%, molybdenum 5 p.p.m.											
390	148	100	30			7	3	6			
392	140	59	35	5		2	4	11	7		
399	104	24	8			0	2	70			
Sulfate sulfur 0.3%, molybdenum 10 p.p.m.											
388	10	9	11	2		10	4	3	7		
391	127	69	12	4		3	4	5	22		
396	1070	32	31	15		2	9	14	7		
Sulfate sulfur 0.3%, molybdenum 20 p.p.m.											
389	54	29	12	2	12	6	5	4	15	16	
393	257	179	92	18	9	2	9	24	40	13	
397	71	58	38	16	0	0	11	13	7	9	
Sulfate sulfur 0.3%, molybdenum 50 p.p.m.											
387	78	48	32	6	14	7	14	26	30	6	
394	58	11	29	16	24	4	6	56	27	9	
400	86	33	19	19	0	1	16	41	10	14	

TABLE 22 (Cont.)

Composition of Liver  
(Experiment II)

Animals by treatment	Liver copper p.p.m.					Days	Liver molybdenum p.p.m.				
	0	75	150	225	300		0	75	150	225	300
Sulfate sulfur 0.3%, molybdenum 100 p.p.m.											
388		10					14				
392		24					84				
Sulfate sulfur 0.3%, molybdenum 200 p.p.m.											
391		22					29				
396		47					36				
Animals Continued Through A Second Lactation											
Sulfate sulfur 0.3%											
386		126	133	83	36		8.2	6.6	5.5	2.1	
395	44	98		106	110	0	4.2		0	5.9	
Sulfate sulfur 0.3%, molybdenum 50 p.p.m.											
387	11	86	14	12	8	11	0.78	11	13	27	
394	7.4	10	20	27	15	23	31	31	35	24	
Calves of: Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (at birth)											
		Liver	Spleen			Liver		Spleen			
394		1.70	.83			0		0			
387		1.9	.91			0		0			
Sulfate sulfur 0.3% (at birth)											
386		118	.60			0		0			
395		127	1.40			0		0			

TABLE 23

Consumption of Trace Elements as Salts\*  
(Experiment III)

Animals by treatment	$\text{Na}_2\text{Mo}_4 \cdot 2\text{H}_2\text{O}$	$\text{Na}_2\text{HAsO}_4 \cdot 7\text{H}_2\text{O}$	$\text{PbSO}_4$	$\text{RbCl}$
Sulfate sulfur 0.3% (outdoors)				
477				
482				
488				
Molybdenum 50 p.p.m. (outdoors)				
479	609.4362			
481	583.9814			
489	628.5306			
Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (outdoors)				
480	662.9193			
483	664.2019			
490	520.8488			
Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (indoors)				
476	493.1749			
485	557.7292			
487	479.3851			
Sulfate sulfur 0.3%, molybdenum 50 p.p.m., rubidium 100 p.p.m., arsenic 5 p.p.m., and lead 5 p.p.m. (indoors)				
478	595.2067	72.1123	32.6528	613.2326
484	622.1376	77.9041	35.2567	685.4966
486	515.8651	65.9668	29.8400	589.6963

Animals Carried Through A Second Lactation

Sulfate sulfur 0.3%

386  
395

Sulfate sulfur 0.3%, molybdenum 50 p.p.m.

387      564.4218  
394      812.2045

\*Total for 300 days in grains.

TABLE 24

Body Weight in Pounds  
(Experiment III)

Week	Animal number														
	476	477	478	479	480	481	482	483	484	485	486	487	488	489	490
1	878	1011	973	1041	1063	1035	987	1230	1049	963	901	893	934	1109	992
2	900	977	952	992	1040	1000	997	1200	1046	937	875	835	912	1048	965
3	816	968	935	970	1017	1107	996	1111	1054	937	865	854	927	1020	963
4	857	995	873	967	1026	1009	1010	1088	1051	944	854	811	936	974	936
5	891	1011	886	976	1025	1025	1034	1100	1052	929	852	841	907	992	976
6	890	1022	912	1077	1044	1019	1010	1098	1068	967	872	846	919	1007	974
7	884	1007	913	982	1037	1031	1059	1114	1059	938	875	822	944	1016	963
8	874	1020	937	1014	1043	1064	1050	1100	1080	952	863	888	920	1025	974
9	887	1026	934	1022	1054	1020	1089	1067	1050	962	873	859	944	1012	977
10	904	1028	932	1020	1076	1053	1018	1083	1053	955	900	835	937	1033	923
11	860	1037	865	930	1075	1057	1062	1090	1041	960	881	886	922	1041	948
12	895	1052	957	1043	1015	1052	1079	1126	1048	965	866	862	957	1043	935
13	891	1029	929	947	1065	1091	1052	1084	1006	930	886	911	955	1034	953
14	903	983	940	1008	1059	1095	1024	1047	1058	985	883	856	963	1026	948
15	885	1047	957	1024	1077	1061	1072	1121	1046	986	899	866	929	1034	947

TABLE 24 (Cont.)

Body Weight in Pounds  
(Experiment III)

Week	Animal number														
	476	477	478	479	480	481	482	483	484	485	486	487	488	489	490
16	896	1032	988	1038	1052	1102	1059	1135	1055	1000	877	861	972	1044	920
17	886	1008	951	1026	1061	1092	1079	1096	1002	967	856	850	979	1023	915
18	874	979	922	1029	1096	1117	1095	1074	994	950	840	858	946	1042	913
19	886	1029	974	1026	1081	1104	1091	1079	1016	954	852	847	963	1054	926
20	908	987	966	1015	1066	1118	1105	1071	1022	945	872	834	930	1057	910
21	900	968	985	1034	1058	1132	1089	1047	1011	958	884	863	1000	1053	935
22	888	932	938	986	1078	1109	1112	1099	990	954	846	827	985	1069	939
23	899	959	925	1025	1069	1118	1130	1123	972	947	857	870	975	1048	889
24	919	1008	928	1016	1051	1095	1130	1083	980	953	845	856	996	1042	910
25	909	1010	946	1041	1055	1108	1154	1103	992	980	865	827	1010	1072	920
26	933	1028	930	1022	1068	1105	1136	1076	980	972	827	830	1000	1082	939
27	931	1035	931	1062	1057	1118	1151	1098	947	948	845	823	1015	1034	912
28	931	1024	926	1031	1071	1115	1159	1066	958	958	865	842	1032	1080	900
29	923	1019	930	1064	1074	1121	1155	1098	1000	953	874	800	1032	1077	893
30	939	1015	944	1022	1084	1139	1188	1112	952	930	858	855	1028	1077	895

TABLE 24 (Cont.)

Body Weight in Pounds  
(Experiment III)

Week	Animal number														
	476	477	478	479	480	481	482	483	484	485	486	487	488	489	490
31	929	1030	913	1057	1103	1145	1184	1133	977	943	867	829	1036	1062	905
32	926	1054	930	1054	1040	1111	1164	1133	993	962	869	857	1024	1105	923
33	920	1039	925	1059	1092	1109	1182	1163	939	934	849	858	1029	1041	922
34	896	1059	962	1070	1067	1117	1173	1131	984	927	854	860	1025	1080	932
35	904	1019	943	1094	1015	1126	1162	1075	1015	961	856	847	1025	1080	908
36	930	1014	912	1091	1112	1152	1196	1144	1020	958	838	829	1065	1085	905
37	915	1036	922	1090	1090	1188	1173	1145	1000	986	831	883	1052	1119	915
38	928	1044	955	1105	1091	1167	1179	1136	1022	948	834	792	1076	1105	943
39	886	1058	963	1112	1103	1183	1214	1130	992	935	843	759	1052	1122	946
40	930	1065	998	1140	1092	1164	1199	1142	998	923	872	750	1067	1122	951
41	922	1046	994	1111	1110	1167	1220	1139	1005	936	842	750	1070	1109	988
42	906	1096	988	1107	1087	1191	1203	1163	993	928	822		1092	1136	983
43	878	1063	992	1115	1068	1189	1200	1140	987	921	836		1052	1140	1000

**TABLE 25**  
**Average Body Weights in Pounds**  
**(Experiment III)**

Animal number	Days				
	0	75	150	225	300
476	865	886	899	931	902
477	985	1039	962	1033	1068
478	953	963	1030	929	991
479	1001	998	1012	1044	1111
480	1040	1055	1067	1076	1088
481	1047	1054	1120	1132	1182
482	993	1038	1102	1179	1208
483	1180	1100	1072	1123	1147
484	1050	1047	1008	974	995
485	946	960	952	945	928
486	880	882	867	864	833
487	861	861	841	847	750
488	924	939	972	1029	1071
489	1059	1039	1060	1081	1128
490	973	935	928	908	990



TABLE 26

Hemoglobins  
(Experiment III)

	Animal number														
	476	477	478	479	480	481	482	483	484	485	486	487	488	489	490
Biweekly values	mg/100 ml blood														
1	10.61	11.33	10.67	9.52	10.48	9.94	9.82	9.33	10.61	9.52	9.15	10.54	11.27	10.00	10.00
2	9.45	11.03	10.30	8.91	9.94	9.76	9.70	9.39	10.79	9.33	9.64	10.54	11.03	10.20	10.82
3	8.79	10.54	9.03	8.67	10.00	9.82	9.70	9.40	10.30	9.21	9.70	10.30	10.60	10.20	11.03
4	8.91	11.03	9.27	8.54	10.18	10.12	9.21	9.58	10.30	9.21	9.88	10.52	10.79	10.67	11.64
5	8.97	11.21	9.21	8.36	10.00	9.94	10.54	9.58	10.00	9.33	9.76	10.64	10.67	10.00	11.70
6	9.64	11.03	9.70	8.91	10.18	10.54	11.03	10.00	10.61	9.45	9.94	10.82	10.00	10.42	11.52
7	9.94	11.21	9.60	9.64	10.00	11.03	10.12	9.33	10.61	9.52	9.88	10.73	9.94	10.91	11.25
8	10.00	10.91	10.00	8.30	9.45	11.03	10.12	9.39	10.42	10.00	9.94	10.73	9.76	11.45	11.03
9	9.94	10.61	10.20	7.64	9.21	11.09	10.42	9.27	10.48	10.36	10.06	10.54	10.36	11.29	11.03
10	10.00	10.73	10.54	8.30	9.39	10.73	10.36	9.15	10.60	10.50	10.00	10.83	10.63	11.45	11.25
11	10.42	10.73	10.48	8.79	9.76	10.00	10.32	9.03	10.79	10.54	9.84	10.79	10.63	11.21	11.09
12	10.38	10.94	10.66	8.96	9.33	10.42	10.32	8.97	10.73	10.54	9.64	9.94	10.48	10.97	10.67
13	10.42	10.94	10.48	8.79	9.45	10.61	10.42	8.73	11.50	11.03	9.84	10.03	10.65	10.97	9.82
14	10.12	11.21	10.54	8.67	9.58	10.66	10.63	9.19	11.45	10.97	9.39	10.00	10.85	11.09	9.90
15	10.30	11.61	10.00	8.67	9.67	10.76	10.36	9.21	11.39	11.32	8.91	10.48	9.94	10.85	9.33
16	10.00	11.33	11.03	8.79	10.00	10.91	10.36	8.67	10.30	10.30	9.09	8.91	9.21	9.94	9.15
17	9.82	11.09	11.21	8.48	10.24	10.18	10.18	9.21	9.52	10.00	8.73	7.09	9.45	8.42	9.39
18	9.33	11.82	10.79	8.67	10.36	10.00	9.52	8.91	9.33	9.70	8.42	6.79	9.58	9.09	9.52
19	8.91	11.88	10.67	8.48	10.67	10.00	9.15	8.73	9.52	9.82	8.85	6.42	9.82	9.52	9.90
20	8.61	13.27	10.42	8.48		10.18	9.45	8.85	10.00	9.52	8.73		10.06	9.27	9.52

TABLE 27

Hematocrits  
(Experiment III)

Biweekly values	Animal number																			
	476	477	478	479	480	481	482	483	484	485	486	487	488	489	490					
1	32.0	31.5	34.0	29.0	33.0	30.0	30.0	29.5	34.0	32.0	32.0	36.5	36.0	32.0	34.0					
2	28.5	33.0	32.0	30.0	32.0	30.0	30.0	28.0	34.0	31.0	30.5	31.0	33.0	33.0	32.0					
3	29.0	34.0	28.5	27.5	31.0	30.0	30.0	30.0	35.0	30.5	30.5	32.0	33.0	33.0	33.0					
4	27.0	36.0	30.0	28.0	32.0	29.0	30.5	30.5	34.0	30.0	31.0	32.0	33.5	33.0	34.0					
5	28.5	35.0	31.0	24.5	31.0	29.0	30.5	30.5	34.0	30.0	31.0	32.0	33.0	33.0	35.0					
6	30.0	35.0	31.0	25.0	31.0	29.0	30.0	30.0	34.0	30.0	31.0	32.0	32.0	33.0	36.0					
7	31.0	33.0	32.0	24.0	30.0	29.0	30.0	30.0	35.0	30.0	31.0	32.0	32.0	33.0	36.0					
8	32.0	33.0	33.0	25.0	30.0	29.0	30.0	30.0	36.0	30.0	31.0	32.0	33.0	34.5	36.0					
9	33.0	34.0	31.0	25.5	30.0	29.0	30.0	30.0	36.0	32.0	33.0	32.5	33.0	35.0	36.0					
10	32.0	34.0	31.0	25.5	30.0	30.0	30.0	28.0	35.0	33.0	31.5	32.0	33.0	35.0	36.0					
11	33.0	35.0	31.0	26.0	28.0	32.0	28.0	28.0	36.0	34.0	32.0	32.0	33.0	35.0	35.0					
12	33.0	35.0	31.0	27.0	28.0	33.0	27.0	27.0	37.0	35.0	31.5	32.5	33.0	34.0	34.0					
13	33.0	35.0	32.0	28.0	29.0	34.0	27.5	27.5	36.0	32.0	31.5	30.0	32.0	34.0	33.0					
14	34.0	36.0	32.0	26.0	29.0	34.0	28.0	28.0	35.0	32.0	28.5	29.0	31.0	34.0	32.0					
15	34.0	36.0	32.0	26.5	29.0	34.0	27.0	26.5	33.0	31.0	28.5	29.0	30.5	34.0	29.5					
16	32.0	37.0	31.0	26.0	30.0	33.0	26.0	26.0	32.0	33.0	28.0	29.0	30.5	30.0	29.0					
17	31.0	37.0	31.0	25.0	31.0	31.0	29.0	29.0	32.0	34.0	28.0	27.0	30.5	26.0	29.0					
18	29.0	38.0	31.0	26.0	33.0	33.5	28.0	28.0	31.5	33.0	27.0	21.0	31.5	28.0	28.0					
19	28.0	40.0	30.0	26.0	33.0	33.0	28.0	28.0	32.0	32.0	27.0	20.0	32.0	30.0	28.0					
20	27.5	41.5	30.0	25.5	33.0	29.0	30.0	30.0	31.5	33.0	27.5	32.0	32.5	30.0	29.0					

TABLE 28

Composition of Milk  
(Experiment III)

Animals by treatment	Milk copper p.p.m.					Days	Milk molybdenum p.p.m.				
	0	75	150	225	300		0	75	150	225	300
Sulfate sulfur 0.3% (outdoors)											
477	0.08	0.04	0.01	0.00	0.08	---	0.02	0.01	0.01	0.05	
482	0.06	0.02	0.00	0.07	0.07	0.03	0.03	0.01	0.24	0.02	
488	0.08	0.02	0.00	0.05	0.06	0.03	0.00	0.00	0.03	0.06	
Molybdenum 50 p.p.m. (outdoors)											
479	0.06	0.02	0.00	0.00	---	0.02	0.60	0.79	0.55	0.88	
481	0.12	0.01	0.00	0.04	0.04	0.04	0.84	0.54	0.52	0.03	
489	0.07	0.02	0.00	0.02	0.01	0.00	0.86	0.33	0.30	---	
Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (outdoors)											
480	0.13	0.01	0.00	0.00	0.07	0.03	0.24	0.06	0.16	0.16	
483	0.08	0.01	0.00	0.01	0.03	0.03	0.16	0.14	0.11	0.25	
490	0.08	0.00	0.00	0.02	---	0.24	0.10	0.12	0.40	---	
Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (indoors)											
476	0.11	0.02	0.01	0.00	0.04	0.02	0.10	0.29	0.05	0.17	
485	0.10	0.01	0.00	0.06	0.02	0.06	0.10	0.04	0.17	0.14	
487	0.08	0.00	0.00	0.03	0.02	0.03	0.11	0.02	0.14	0.38	
Sulfate sulfur 0.3%, molybdenum 50 p.p.m., rubidium 100 p.p.m., arsenic 5 p.p.m., and lead 5 p.p.m. (indoors)											
478	0.11	0.01	0.00	0.00	0.06	0.02	0.33	0.18	0.22	0.05	
484	0.12	0.01	0.00	0.02	0.03	0.03	0.46	0.13	0.24	0.51	
486	0.06	0.00	0.00	0.05	0.09	0.02	0.05	0.01	0.10	0.15	

TABLE 29

Composition of Serum  
(Experiment III)

Animals by treatment	Serum copper p.p.m.					Serum molybdenum p.p.m.				
	0	75	150	225	300	0	75	150	225	300
Sulfate sulfur 0.3% (outdoors)										
477	0.80	0.95	0.98	0.59	0.60	0.0	0.01	0.0	0.0	0.0
482	0.40	0.56	0.55	0.55	0.59	0.0	0.02	0.01	0.03	0.05
488	1.14	0.62	0.78	0.57	0.73	0.0	0.0	0.02	0.0	0.03
Molybdenum 50 p.p.m. (outdoors)										
479	0.87	0.78	0.72	0.60	0.82	0.0	2.15	1.99	1.88	2.16
481	1.19	0.98	1.06	1.25	0.62	0.0	4.22	0.86	1.12	1.20
489	0.75	1.07	0.70	0.72	0.44	0.0	0.98	0.76	0.94	---
Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (outdoors)										
480	0.78	1.12	0.96	0.79	0.66	0.00	1.57	0.66	0.72	0.84
483	0.76	0.80	0.64	0.60	0.22	0.00	0.48	0.40	0.62	0.30
490	0.90	0.88	0.50	0.24	0.23	0.00	0.48	0.58	0.66	---
Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (indoors)										
476	0.60	0.68	0.70	0.55	0.71	0.00	0.42	0.73	0.29	0.59
485	0.88	0.84	0.62	0.56	0.16	0.00	0.54	0.31	---	0.10
487	0.68	0.68	0.42	0.60	0.26	0.00	1.00	0.20	0.75	0.45
Sulfate sulfur 0.3%, molybdenum 50 p.p.m., rubidium 100 p.p.m., arsenic 5 p.p.m., and lead 5 p.p.m. (indoors)										
478	0.80	0.90	0.76	0.64	0.68	0.02	1.07	1.16	1.24	0.44
484	0.69	1.09	0.80	0.72	0.61	0.00	1.00	0.78	0.93	1.04
486	0.69	0.78	0.74	0.66	0.56	0.00	0.64	0.32	0.27	0.30

TABLE 30

Composition of Liver  
(Experiment III)

Animals by treatment	Liver copper p.p.m.					Days	Liver molybdenum p.p.m.				
	0	75	150	225	300		0	75	150	225	300
Sulfate sulfur 0.3% (outdoors)											
477	6.4	53	75	38	46		2.3	2.4	6.2	20.0	6.0
482	0.0	23	49	48	14		0.0	4.3	7.0	7.8	7.8
488	48.0	78	90	65	27		1.9	3.2	8.2	8.0	28.0
Molybdenum 50 p.p.m. (outdoors)											
479	26.0	0	9.3	0	6.8		3.3	30	31	27	33
481	0.0	1.7	13.0	9.7	5.6		0.0	16	29	25	20
489	123.0	59.0	30.0	16.0	85.0		0.3	25	26	28	28
Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (outdoors)											
480	68	31	25	18	18		0.0	22	20	19	29
483	0	5.5	11	17.0	9.0		0.0	12	13	16	8
490	20	32	12	8.0	4.9		3.0	14	14	--	24
Sulfate sulfur 0.3%, molybdenum 50 p.p.m. (indoors)											
476	19.0	1.8	25	14	13.0		2.3	5.4	21	13	16
485	48.0	18.0	9.0	8.9	5.3		0.0	16	16	14	9.7
487	10.3	---	8.8	10	5.4		3.9	14	14	15	---
Sulfate sulfur 0.3%, molybdenum 50 p.p.m., rubidium 100 p.p.m., arsenic 5 p.p.m., and lead 5 p.p.m. (indoors)											
478	0.5	14	15	2.3	9.0		0.0	14	18	13	14
484	44	15	15	13	12		0.0	15	16	14	20
486	67	39	28	27	16.8		0.0	21	17	25	30



TABLE 31 (Cont.)

**Copper-64 Level in Blood  
(Experiment IV)**

Counts per minute of copper-64 per milliliter of blood\*

Hours after administration of copper-64	Trial V			Trial VI	
	Steer	Steer		Steer	Steer
	473	474		498	499
0	0	0	0	0	0
2	7	4	2	16	0
4	14	14	4	21	31
6	30	26	6	39	45
8	35	10	8	66	124
10	48	58	10	83	80
12	52	73	12	78	78
14	87	63	14	58	106
16	97	34	16	209	163
18	177	113	18	219	207
21	188	123	20	273	230
25	171	138	22	285	236
30	179	159	24	226	277
38	324	257	28	288	304
44	320	400	32	411	453
50	455	600	44	350	450

\* Corrected to time when copper level was 10 mc.











TABLE 35 (cont.)

Results of Statistical Analysis of Data  
for the Combined Experiments I and II

	Decreases in levels of copper			Increases in levels of molybdenum		
	Milk	Serum	Liver	Milk	Serum	Liver
----- Levels of significance -----						
<b>Effect due to molybdenum</b>						
75 days	>0.05	>0.05	>0.05	0.01	0.01	0.05
150 days	>0.05	>0.05	>0.05	0.01	0.01	>0.05
225 days	>0.05	>0.05	0.01	0.01	0.01	>0.05
300 days	----	>0.05	>0.05	>0.05	0.01	>0.05
<b>Effect due to sulfate</b>						
75 days	0.01	>0.05	>0.05	>0.05	0.05	0.01
150 days	0.01	>0.05	>0.05	0.05	0.01	>0.05
225 days	0.01	>0.05	>0.05	0.01	0.01	>0.05
300 days	----	>0.05	>0.05	0.05	0.01	>0.05
<b>Interactions of molybdenum and sulfate</b>						
75 days	>0.05	>0.05	>0.05	0.05	0.05	>0.05
150 days	>0.05	>0.05	>0.05	0.01	>0.05	>0.05
225 days	>0.05	>0.05	>0.05	0.01	0.01	>0.05
300 days	----	>0.05	>0.05	>0.05	0.01	>0.05