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Gabriella Gentile Virginia Commonwealth University

Lizette Carrasco Virginia Commonwealth University

Amy Stanford Virginia Commonwealth University

Kimberly Jefferson Virginia Commonwealth University

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Characterization of Protein Involved in Hemolysis Expressed by Sneathia amnii, a Pathogen of the Female Urogenital Tract

Gabriella L. Gentile, Lizette I. Carrasco, Amy Stanford, Kimberly K. Jefferson, PhD. Department of Microbiology and Immunology, School of Medicine, Virginia Commonwealth University, Richmond VA

Abstract

Sneathia amnii is a poorly characterized gram-negative anaerobe that commonly colonizes the vagina. It has been linked to many obstetric disorders, including preterm labor, preeclampsia, and chorioamnionitis. S. amnii lyses human red blood cells, and we aimed to identify the hemolysin. We identified two genes that appear to encode transporter and effector components of a two-partner secretion system. The putative effector, which we refer to as SaFHA, contains a domain with amino acid similarity to the filamentous hemagglutinin (FHA) of Bordetella pertussis and its predicted structure suggests it may form a transmembrane channel or pore. Thus, we hypothesized that SaFHA would be secreted by S. amnii and that it would play a role in hemoglobbin release. To test this, a portion of the gene encoding the SaFHA protein in S. amnii was expressed in *E. coli* and used as an immunogen in rabbits. Western analysis using anti-SaFHA revealed that the protein is secreted and localizes to the bacterial surface. Pre-treatment of S. amnii with anti-SaFHA blocked the hemolytic activity whereas antiserum against an irrelevant protein had no effect. We partially purified SaFHA from S. amnii using cation exchange chromatography and the partially purified protein mediated hemoglobin release from human RBC, supporting our hypothesis. Further characterization of SaFHA will help provide more insight on the virulence of *S. amnii*, and perhaps shed light on the etiology of Sneathia-associated vaginal conditions, as well as future treatment options.



treated bacteria for 2hr at 37°C. The cells were removed

by centrifugation and the absorbance of the supernatant

at 450 nm was measured.

Hypothesis

The FHA homologue of S. amnii plays a role in the release of hemoglobin from human RBC.



Fig. 1 (A) SEM micrograph of Sneathia amnii morphology. (B) S. amnii consists of long gram-negative rods and short rods and cocci.

Methods

Purification of SaFHA protein <u>Create an Isogen</u> saFHA mutant of *S. amnii* by homolgous recombination

Characterize the role of SaFHA in hemolysis and adherence



*SaFHA (~200 kDa) detected in lanes 3, 4, and 6. SaFHA ~250 kDA.

Purification Schematic of SaFHA Through Affinity Chromatography

SaFHA has a predicted pl of 9.4. Cation exchange was used to purify SaFHA from

Fig. 6 Purification of SaFHA using cation exchange. OM extracts were suspended in 20 mM Tris pH8 + 0.5% Triton-X, sonicated, and filter sterilized. The OM was applied to a cation exchange column, and passed through a series of wash steps. Western analysis was performed on the flow through from each wash step. 1). OM Before filtering, 2). OM After filtering, 3). OM cation flow through, 4). Tris pH 8, 5). Tris pH 8 + 100mM NaCl, 6). Tris pH8 + 200mM NaCl, 7). Tris pH 8 + 500mM, 8). Tris pH 8 + 1M. SaFHA ~250 kDA.

Fig. 7 Hemolytic activity of SaFHA. Purified SaFHA was added to human erythrocytes at increasing concentrations and allowed to incubate for 2 hrs. at 37°C. Remaining whole cells were removed by centrifugation and the amount of hemoglobin released into the supernatant was quantified by measuring absorbance at OD450nm.

Scheme for Production of Isogenic saFHA Deletion Mutant

In order to characterize the role of the SaFHA in the lysis of human red blood cells, an isogenic *saFHA* mutant of Sn35 will be constructed through homologous recombination. This mutant strain, in addition to purified protein (explained above), will be used in adherence and hemolysis assays to assess the role of



Fig. 8 Scheme for production of isogenic saFHA deletion mutant. A small portion of the upstream and downstream region of the gene remains intact, while the middle is knocked out and replaced with a CAT antibiotic

Fig. 9 Agarose gel of linearized DNA fragment, constructed through **SOE PCR.** 1). 1 kb DNA ladder, 2). SOE PCR product.

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