University of Arkansas, Fayetteville ScholarWorks@UARK

Technical Reports

Arkansas Water Resources Center

1974

Effects of Mosquito Control Chemicals on Aquatic Fauna

J. L. Lancaster Jr. University of Arkansas, Fayetteville

M. V. Meisch University of Arkansas, Fayetteville

Follow this and additional works at: https://scholarworks.uark.edu/awrctr Part of the Entomology Commons, Fresh Water Studies Commons, Terrestrial and Aquatic Ecology Commons, and the Water Resource Management Commons

Recommended Citation

Lancaster, J. L. Jr. and Meisch, M. V. 1974. Effects of Mosquito Control Chemicals on Aquatic Fauna. Arkansas Water Resource Center, Fayetteville, AR. PUB023. 34

This Technical Report is brought to you for free and open access by the Arkansas Water Resources Center at ScholarWorks@UARK. It has been accepted for inclusion in Technical Reports by an authorized administrator of ScholarWorks@UARK. For more information, please contact scholar@uark.edu, ccmiddle@uark.edu.

PUB-0023

EFFECT OF MOSQUITO CONTROL CHEMICALS ON AQUATIC FAUNA

by

J.L. Lancaster, Jr.

M.V. Meisch



WATER RESOURCES RESEARCH CENTER

Technical Completion Report

Project A-018-ARK

UNIVERSITY OF ARKANSAS FAYETTEVILLE

PROJECT COMPLETION REPORT

OWRR Project No. A-018-ARK Starting Date: June, 1970 Agreement No. 14-31-0001-3804 Completion Date: June 1973

EFFECT OF MOSQUITO CONTROL CHEMICALS

ON AQUATIC FAUNA

J. L. Lancaster, Jr.

M. V. Meisch

Entomology Department

The work upon which this report is based was conducted in cooperation with and in part with funds provided by the Office of Water Resources Research, U.S. Department of the Interior, under Annual Allotment Agreement No. 14-31-0001-3204 as authorized under the Water Resources Research Act of 1964, P. L. 88-379 as amended.

INTRODUCTION

No mosquito abatement districts have ever been organized in Arkansas. Mosquito control efforts have been largely adulticiding operations by either aerial application or ground thermal fogging machines. Practically no chemical applications have been directed at the larval stage in residual water in ditches and depressions from which adult populations arise. Some larviciding with ethyl parathion has been done in ricefields. Although the treatment is very effective in mosquito reduction, voluntary treatment has not been completely successful.

Because relatively little insecticide has been used as a larvicide in Arkansas, it was possible to evaluate the effect of recommended larvicides on non-target organisms in the aquatic environment. A developing mosquito control demonstration program in the rice-producing area provided the study site.

METHODS AND MATERIALS

On the basis of the estimated flight range of the dark ricefield mosquito, four sections were mapped to receive the larviciding chemicals. An additional 1-mile radius was scheduled for treatment of all rice with ethyl parathion. This provided a distance of 2 miles from the center of the treatment area.

Sampling for aquatic fauna was carried out by taking 10 dips with an 8 oz dipper and concentrating the specimens by pouring them through a siphoning device (ADCAS- automatic device for collection of aquatic samples). Excess water was drained off carefully after siphoning was complete and the reduced sample was placed in a baby food jar, marked with date and sample site. At the end of the day, alcohol was added to preserve the specimens. Sampling was carried out before and after treatment on approximately a weekly basis. Control samples were taken at random from outside the treatment areas. Samples were stored and later analyzed in the laboratory, after mosquito-control operations had ceased and personnel were freed to perform this task.

Variable numbers of samples were taken, depending on the availability of standing water. In 1970, 588 samples were collected; in 1971, 277; and 1972, 427. The number of samples decreased as water sources dried up and increased during rainy periods when sample sites were filled. Generally, the same sites were sampled when water was present.

During the first season, treatment was applied almost on a weekly basis to standing water in ditches and to any source which could be found producing mosquito larvae. Later, treatment was applied to sites producing mosquito larvae rather than to all standing water.

The four larvicides used were (1) Abate^(R) in the NW section; (2) malathion in the NE section; (3) fenthion (Baytex) in the SE section; and (4) Flit MLO, a petroleum hydrocarbon, in the SW section. Application of these materials was by power sprayer to standing water. The rates used were (1) Abate^(R) 4 lb EC - 1.5 oz/gal finished spray; (2) malathion 57% EC - 3.0 oz/gal finished spray; (3) fenthion 4 lb EC - 1.5 oz/gal finished spray; and (4) Flit MLO used directly. Table I shows the dates of application and amounts of larviciding chemicals used; Table II shows the dates and amounts of adulticides used.

Data	Ab	NE ate	4EC	Fei	SE	n 4EC	Malat	NW hion	57% EC	SW Flit MLO 70 71 72				
Date	<u>/0</u>			<u>70</u>	/1	/2		/1	12	<u>/0</u>	/1			
6-01	-	-	-		-		-	-	-	48	-	35		
6-02	_	_	-	-	-	75	-	-	_	-	60	_		
6-03	-	-	-	-	30	-	25	-	-	-	-	-		
6-04	-	-	-	60	55	-	-	-		60	÷	-		
6-05	60	-	75	-	-	-	-	-	75	n -	-	-		
6-07	-	10	-	-	45	-	-	-	-	-	-	-		
6-08	-	40	-	-	-	-	-	-	-	-	-	-		
6-09	-	-	-	-	-	-	-	72	-	-	-	-		
6-11	-	70	-	-	-	-	-	-	-	-	-	-		
6-15	-	-	-	-	-	-	25	-	-	19	-	-		
6-16		-	-	75	40	-	-	-	-	-	-	-		
6-17	20	-	-	-	-	-	-		-	-	55	-		
6-22	-	-	-	-	-	-	-	٥5	-	-	13	-		
6-23	-	42	-	-	-	-	-		-	-	-	-		
6-25	-	-	-	-	80	-	-	-	-	-	-	-		
6-28		-	-	-	-	75	.+	-	-	-	-	-		
6-29	-	-	-	-	-	-	25	_	-	30	-	-		
6-30	25	-	-	55	-	_ **	-	-	75	-	-	-		
7-01	-	-	-	-	-	-	-	85		-	70	-		
7-05	-	-	75	-	-	-	-	-	-	-	-	35		
7-06	-	-	-	-	-	75	-	-	75	-	-	-		
7-07	-	-	-	-	69	-	-	-	-	-	-	-		
7-08	-	50	-	-	-	-	-	80	-	-	22	-		
7-09	-	-	-	-	-	-	67	-	-	63	-	-		
7-10	-	-	-	105	-	-	-	-	-	-	-	-		
7-13	37	-	-	-	8 	-	-	-	-	-	-	-		
7-16	-	-	-	-	-	-	35	-	-	· 70	-	-		
7-17	-	-	-	185	-	-	-	-	-		-	-		
7-20	70	-	-	-	-	-	-	-		-	-	-		
7-21	-	15	-	-	30	-	-	52	-	-	-	-		
7-27	-	-	-	-	62	-	-	15	-	-	-	-		
7-28	50	-	-	-	-	-	~	-	-	-	-	-		
7-29	-	-	-	-	-	-	-	70	-	-	-			
8-03	-	-	75	-	-	75	68	-	75	51	-	-		
8-04	30	-	-	92	-	-	-	-	-	-	-	-		
8-05	2.5	3 5	-	-	-	-	=	-	-		-	-		
8-09	-	-	-	-	-	-		-	-	-	17	-		
8-10	-	-	-	-	85	-	-	-	-	-	-	-		
8-11	150	-	-	-	-	-	-	-	-	-	-	-		
8-12	-	-	-	165	-	-	-	-	-	-	-	-		
8-13	-	-	-	-	-	-	-	-	-	49	-	-		

Table I.	Dates and amounts	(gal) of	larvicides applied i	n treatment area at
	Lonoke, Arkansas,	in 1970,	1971 and 1972.	

	Aerial Appli	cations	
		Fenthion 4EC	ж
Date	1970	<u>1971</u>	<u>1972</u>
6-19	15.00	-	15.00
6-25	-	15.00	-
7-02	15.00	-	- ,
7-07	-	-	15.00
7-10	15.00	-	-
7-17	15.00	-	-
8-14	15.00	-	-

Table II. Gallons of insecticide concentrates applied against adult mosquitoes in 1970, 1971, and 1972, Lonoke, Arkansas.

ULV Applications

		Malathion*	
Date	<u>1970</u>	<u>1971</u>	1972
6-02	-	-	3.00
6-04	2.00	-	-
6-06	-	-	3.00
6-09	4.00	-	-
6-12	2.50	-	-
6-15	1.75	7.00	3.00
6-16	-	-	3.00
6-17	4.00	-	3.00
6-18	-	8.00	3.00
6-22	3.50	2.00	-
6-23	-	-	3.00
6-24	3.00	-	-
6-26	2.50	-	3.00
6-28	-	-	3.00
6-30	-	4.00	-
7-01	-	4.00	3.00
7-02	-	3.13	-
7-05	-	3.20	-
7-08	3.50	3.25	-
7-09	-	15.00(Dibrom)	-
7-10	-	-	3.00
7-11	-	3.90	-
7-12	-	4.20	-
7-13	4.50	2.50(Dursban)	-
7-14	-	1.90(Dursban)	3.00
7-15	-	15.00(Dibrom)	-
7-16	-	1.90(Dursban)	-
7-19	-	4.00	-
7–20	1.30	2.50	-
7-21	-	2.00	3.00
7-26	-	3.50	-
7-27	-	3.00	-
7-28	-	4.00	-
8-02	-	4.50	-

Date	<u>1970</u>	<u>1971</u>	<u>1972</u>
8-09	-	_	3.00
8-11	1.50	-	
8-17	4.25	-	3.00
8-22	-	-	3.00
8–26	2.50	-	-

* Except where indicated.

LITERATURE REVIEW

As early as 1919, studies on <u>Anopheles quadrimaculatus</u> (Say) were being conducted in the area of Lonoke by Geiger <u>et al</u>. (1919), who stated that an infestation of this species was thought to originate from two ricefields a mile from the city. They also conducted flight range experiments with 4,000 marked mosquito adults liberated near the ricefields. Only 10 adults were recaptured. Nine were caught 0.75 mile from the release point and one was recaptured a mile away. Whitehead (1957) conducted further experiments in the area on the flight range of ricefield mosquitoes. These data indicated that if breeding of ricefield mosquitoes is prevented for a distance of 4 miles from a given area, less than 3% of the mosquitoes originating outside will fly into the area. Only about 10% were trapped at a distance of more than 1.7 miles from the nearest field in the study.

Kiker and Breedlove (1941) conducted flight dispersal studies in Tennessee. They showed that <u>Anopheles</u> mosquitoes traveled up to 1.25 miles from the breeding site, but average flight range was 1 mile.

In relation to aquatic arthropods other than mosquito larvae, Fales <u>et al</u>. (1968) eradicated a large population of <u>Chaoborus</u> spp. in a lake treated with Abate^(R) at a rate of 0.39 lb/acre. This concentration also proved toxic to nearly all other insects in the lake. Von Windeguth and Patterson (1966) studied the effects of an organophosphorous insecticide on aquatic biota. Field application of 0.25 lb of technical Abate per acre produced no noticeable mortality of Odonata, Chaoborus, Copepoda and Ostracoda. Porter and Gojmerac (1969) stated that Abate at 0.03 lb/acre was toxic to nymphs of Libellulidae and to

Cladocera (Crustacea). It was not toxic to mosquito pupae, Amphipoda, Isopoda, Ostracoda and Copepoda.

Kimball and Perruzzi (1970) applied Flit MLO at the rate of 2-3 gal/acre to breeding sources in an urban community. They concluded that Flit MLO killed both mosquito larvae and pupae when applied at the rate of 2-3 gal/acre and provided control of mosquitoes for a period of up to 14 days in Orange County, Calif. Felton (1944) stated that petroleum oils do not kill mosquito larvae and pupae by suffocation. The volatile components of the oil exert a direct toxic effect on the tissue of the larvae and pupae which result in death.

Mount et al. (1970), using a Leco cold aerosol nozzle connected to a modified Curtis 55,000 cold aerosol generator, showed that technical malathion (95%) was dispersed in droplets averaging 11μ in diameter. The nozzle was operated at 3.5 psi with a flow rate of 2.85 fl oz/min; vehicular speed was 10 mph. The dispensed droplet size corresponds to the optimum of $5-10\mu$ mass median diameter as suggested by Mount (1970). Caged female Aedes taeniorhynchus (Wiedemann) adults were used as the test animal. At distances of 150,300 and 600 ft percentage mortality after 18 hr was 93, 93 and 93 respectively. The LD_{50} , for malathion and fenthion were 0.025 and 0.072 lb/acre respectively, based on amount of active ingredient used per acre. Taylor and Schoof (1968) tested thermal and non-thermal fogs against adults of Aedes aegypti (Linnaeus), Anopheles albimanus (Wiedemann), Culex quinquefasciatus and Aedes taeniorhynchus. At distances of 150 and 300 ft malathion was found superior to Dursban, naled and Baygon^(R). The latter three were metered out at the rate of 2 oz/min/gal, whereas the malathion dose rate was 6 oz/min/gal. Dursban proved most effective at the 2 oz/min/gal level.

Paulini et al. (1962) demonstrated residual activity of fenthion

against <u>Culex fatigans</u> (Wiedemann) adults on various types of wood surfaces. Bare wood surfaces produced the best residual activity to the adult mosquitoes when fenthion was applied at $1.1 - 2.3 \text{ g/m}^2$. Residual activity of the chemical lasted from 3 to 6 months.

Lewallen and Wilder (1963) tested insecticides on mosquito larvae in polluted and pure tapwater. Polluted tapwater was created by mixing pure tapwater with manure. C. fatigans larvae were found to be more susceptible to DDT and fenthion in polluted tapwater than in pure tapwater. Malathion was equally toxic to the larvae in the two water conditions. Parathion was more effective in pure tapwater than in polluted tapwater. Using 1% fenthion, Stevens et al. (1963) produced 100 and 99.4% mortality at distances of 80 to 330 ft respectively from the release point. Burton (1964), using the World Health Organization Kit for larval resistance or susceptibility, found fenthion to be the most toxic of the chemicals tested. Diazinon was second, followed by malathion, BHC, dieldrin and DDT. Gahan and Noe (1955) studied control of mosquito larvae in ricefields with water soluble phosphorous insecticides. The study was conducted to determine the effectiveness of water soluble larvicides for control of ricefield mosquitoes. The chemicals used were Bayer L 13/59, Shell OS 2046, DDVP and parathion. All were applied with an automatic applicator to the water before it entered the field. Parathion at 0.1 ppm proved to be highly effective after flowing more than 0.5 mile through a canal and 400 ft into the ricefield. This kill rate was in relation to Psorophora confinnis (Lynch and Arribalzaga). None of the chemicals had residual activity against A. quadrimaculatus and Culex erraticus (Dyar and Knab).

Mulla <u>et al</u>. (1961a) found that parathion produced a high toxicity to <u>C</u>. <u>quinquefasciatus</u> with an LD₅₀ of 0.0045 ppm. Mulla (1961b) stated

that <u>C</u>. <u>quinquefasciatus</u> larvae were more susceptible to insecticides in the early larval instars than in the latter stages of larval development.

The 48-hr LC_{50} of Abate for stoneflies (<u>Pteronarcys californica</u>) and Amphipods (<u>Gammarus lacustris</u>) was 100 ppb and 1,500 ppb respectively (FWPCA, 1968). The 24-hr LC_{50} of Abate for the Amphipod (<u>G. lacustris</u>) was 960 ppb (Sanders, 1969).

Abate was one of the least toxic of eight chemicals tested against <u>Cyclops spartinus</u> (Ruber, 1963), and <u>Cypronotus incongruens</u> was remarkably insensitive to Abate (Ruber, 1963, 1965a, b).

The LC_{50} of malathion for various arthropods is shown in Table III (Pimentel, 1971).

The 48-hr EC₅₀ (immobilization value at 60°F) of malathion for waterfleas, <u>Simocephalus serrulatus</u> and <u>Daphnia pulex</u>, was 3.5 ppb and 1.8 ppb respectively (Sanders and Cope, 1966).

Crayfish in streams in a malathion-treated (2 lb/acre) watershed were unaffected by the treatment (Peterle and Giles, 1964). Other arthropod numbers decreased greatly but recovered soon after treatment.

The 48-hr EC₅₀ (immobilization value at 60°F) of fenthion for waterfleas, <u>Simocephalus serrulatus</u> and <u>Daphnia pulex</u>, was 0.92 ppb and 0.80 ppb respectively (Sanders and Cope, 1966). The LC₅₀ values of fenthion for various arthropods are shown in the Table IV (Pimentel, 1971)

Mortality has been observed in dytiscid adults and larvae and in hydrophilid and notonectid adults after application of a surface film of Flit MLO to their normal habitat. Dragonflies, damselflies, mayflies and caddisflies were not affected by Flit MLO (Exxon, 1973). The only significant hazard observed is to certain insect life with biological stages that occur at the air-water interface (Humble, 1972).

Arthropod Species	Exposure <u>Time (hr)</u>	LC ₅₀ (ppm)	Source
Amphipod			
(<u>Gammarus</u> <u>lacustris</u>)	24	0.0038	Sanders, 1969
Stonefly			
(<u>Pteronarcella</u> <u>badia</u>)	24	0.010	Sanders and Cope, 1966
Stonefly			
(Claassenia sabulosa)	24	0.013	n
Stonefly			
(Pteronarcys californica)	24	0.035	**
Hermit crab	24	0.118	Eisler, 1969
Grass shrimp	24	0.131	n
Sand shrimp	24	0.246	n
Waterflea	·		
(Daphnia pulex)	48	0.0018	FWPCA, 1968
Amphipod	• -	•	
(G. lacustris)	48	0.0018	11
Waterflea (D. pulex)	48	0.002	Cope, 1966
Stonefly	•		
(Simocephalus serrulatus)	4 8	0.003	11
Stonefly (P. badia)	<u>4</u> 8	0.006	FWPCA, 1968
Mavfly (Raetis sp.)	48	0.006	Cope, 1966
Stonefly			
(P. californicus [sic])	48	0.020	11
Red crawfish	48	20.0	Muncy and Oliver, 1963
	70	_0,0	inter, 1905

Table III. LC_{50} of malathion for various arthropods.

Table IV. LC_{50} of fenthion for various arthropods.

Arthropod Species	Exposure Time (hr)	LC50 (ppm)	Source
Amphipod			
(Gammarus lacustris)	24	0.015	Sanders, 1969
Stonefly			•
(Pteronarcys californica)	24	0.130	Sanders and Cope, 1966
Stonefly			
(Simocephalus serrulatus)	48	0.0031	FWPCA, 1968
Waterflea			
(Daphnia pulex)	48	0.004	n
Stonefly			
(P. californica)	48	0.039	n
Amphipod			
(<u>G. lacustris</u>)	48	0.070	12
Stonefly			
(P. californica)	48	0.130	n

RESULTS AND DISCUSSION

For the past three years the community of Lonoke, Arkansas, has been the site of a mosquito control demonstration. The project was to demonstrate that mosquito numbers could be reduced within the budget of a ricefield community, and was supported jointly by the City of Lonoke, the Lonoke Chamber of Commerce, and a grant from the Office of Water Resources Research. The project was supervised and directed by the Arkansas Agricultural Experiment Station and the Arkansas Cooperative Extension Service. This program is hoped to have altered a rather prevalent negative attitude toward mosquito control; i.e., mosquitoes have always been troublesome, they always will be, and nothing can be done about it.

As in the two previous years, the 1972 Lonoke program consisted of larviciding mosquito breeding sites within a 2-sq-mile area with Lonoke at the center. All ricefields within an additional 2-mile zone were larvicided; however, no other breeding sites were treated. A ULV cold aerosol generator was used against adult mosquitoes as a supplement to larviciding. Finally, aerial sprays were made when mosquito populations could not be controlled by the aforementioned techniques.

Two mosquito species (<u>Psorophora confinnis</u> and <u>Anopheles quadri-</u><u>maculatus</u>) collected from New Jersey light traps were of primary concern during the three year study (Fig. 1). <u>Psorophora confinnis</u> was most abundant. Peak numbers occurred near June 10 and 20 and July 1. After July 1, <u>P</u>. <u>confinnis</u> numbers decreased sharply. <u>Anopheles</u> <u>quadrimaculatus</u> gradually increased throughout the season and became

the dominant species after August 1. The rise in <u>Anopheles</u> numbers coupled with the decrease in <u>Psorophora</u> numbers served to maintain pestiferous populations of mosquitoes late in the summer. It should be noted that <u>A</u>. <u>quadrimaculatus</u> is not a species which is readily attracted to light traps; however, other observations including landing rate counts and larval surveys indicated that the population trend shown by light traps was correct.

Total numbers of mosquitoes collected from New Jersey light traps in 1970 and 1971 were averaged and compared to total numbers of mosquitoes collected in 1972 (Fig. 2). In 1970 and 1971, populations exceeded 2000/trap for much of June, whereas the maximum in 1972 was 412. These numbers are expressed as logarithms; numerically, a single trap in 1970 captured as many as 38,000 specimens, and in 1971 and 1972 the maximum numbers were 4000 and 1500 respectively. Five aerial sprayings were needed in 1970, one in 1971, and two in 1972. Control was not absolute, but mosquito numbers were reduced substantially.

The scope of the Lonoke project was restricted by financial resources. For more effective control the abatement area should be increased at least twofold. A mosquito abatement district is the ultimate solution. Even though the value of mosquito abatement has been proven, many local authorities remain skeptical and it appears necessary to demonstrate the practicality of mosquito control. The technology for successful mosquito control in Arkansas does exist; however, the best available recommendations at this time are primarily chemically oriented. The approach must become more integrated and additional research is needed for long-range control, because many mosquito abatement problems in Arkansas rice-growing areas are unique to Arkansas.

During the Lonoke study, 588 samples of aquatic fauna were analyzed from the 1970 collections; 277 and 427 were analyzed respectively from 1971 and 1972 collections. The numbers and identity of the species found are shown in Table V. A comparison of the average numbers by years and section is **shown** in Table VI. During the course of the investigation, reduction in numbers could be detected after treatment but recovery was also apparent. Another factor to be borne in mind is that when winged adults emerged, migration out of the aquatic habitat resulted. Comparison with control samples indicates considerable reduction in overall aquatic fauna in all sections. The fact that the highest percentage reduction occurred in mosquito larvae attests to the effectiveness of the chemicals for mosquito control.





- ·			1970						19/1						197	4		
Specimens	Total	Control	. NE	SE	NW	SW	Total	Control	NE	SE	NW	SW	Total	Control	NE	SE	NW	S
Gastropoda	2091	107	451	1028	125	380	29	12		1	12	4	2109	315	515	538	49 8	24
Arachnida	41				 ;		1844	523	231	506	344	240	59		5	19	16	1
Araneida	29	3	8	10	4	4	39	8	7	3	9	12	30	6	1	12	7	
Acarina	12	3	2	4	1	2	4	0	1			3	37	2	4	7	9	1
Jrustacea	553 (555)					610						3340		24	145	590	258
Ostracoda	256	112	6	60	71	7	189	102	31	7	42	7	359	107	3	31	47	17
Malocostraca	67 (69)						63						5		1			
Decapoda	10-		1		9		12	2	1	9			7	1	1	2		
Isopoda Jacuada (non a	13	10	1			2	34	5	9	2	10	8	8	7				
Isoboga (uou-a	quatic) 2		2						-	-	-							
Amphipoda	44	5		6	32	1	17	8	1	2		6						
Branchiopoda	36						51						3027		20	112	489	240
Cladocera	32						24						3027		20	112	489	240
Daphnidae	25						1	30 					3012		20	98	488	240
<u>Moina</u> brachi	ata 23	5	2		8	8	1	1*			% 		3015	3	20	98	488	240
Ceriodaphnia	<u>spp</u> . 2				1	1											***	
Diaptomidae	1						23						15			14	1	
Diaptomus sp	p. 1			1			23	2			19	2	<u>_</u> 16	1		14	₈ 1	
Inmature stage	в 6	1				5												
Conchostraca	L					4	88	27		18	26	17	14	14				

Table V.	Numbers and	† dentities	of species	found in	water	samples,	Lonoke,	Arkansas

			1970	C			Table V. (continued) 1971							1972					
Specimens	Total Co	ntrol	NE	SE	NW	SW	Total	Control	NE	SE	NW	SW	Total	Control	NE	SE	NW	SW	
Copopeda	194						307						163				2		
Eucopepoda	194			60 6			307							6 -0					
Calanoida	9	2		3	4		12	2		3	7								
Harpacticoida	15					a n 85 an	3				***								
Ergasilidae	15						3												
Ergasilus	15	11	1	2		1	3	1			1	1					4 00		
Cyclopoida	170				-		72			Q , 1 1149			54				54		
Cyclopidae	170																		
Cyclops	169	54	5	21	55	34	72	25	11	5	8	23	94	40			54		
Immature stag	ges 1					1	110	39	41	4	26		20	20			A		
Insecta 1	156(1268)						2380					4	2524		228	328	564	1404	
Psocoptera						<u></u>							1		1				
Collembola	6			جه _{موا} جه			129						22	3 	2	15	3	2	
Entomobryidae	4	-	2	1		1	66	21	13	32			22	11	2	6	2	1	
Sminthuridae	2	1				1	63	25	14		24	1	23	13		9	1		
Ephemeroptera	75						361			8 9 			763		6	54	34	669	
Baetidae	74						361						763		6	54	34	669	
Cloeon	74	37	5	3	9	20	361	116	18	7	24	196	840	77	6	54	34	669	
Immature stages	1					1													
Odonata	29						112						201	13	12	22	87	80	

	Table V. (continued) 1970 1971												1972						
Specimens	Total	Control	NE	SE	NW	SW	Total	Control	NE	SE	NW	SW	Total	Control	NE	SE	NW	SW	
Libellulidae	4						3						42		3	3		36	
Sympetrum	1	,		1			3					3	39			3		36	
Other	3			3	***														
Aeshnidae	1						1						2		••• ••••			2	
Aeshna	1						1					1	2					2	
Coenagrionidae	21						102												
Enallagma	17	9	5		1	2	76	37	9	5	3	22							
Neonura	1					1	12	2				10						~	
Ischnura	3		2			1	14	8	1			5							
Lestidae	2				*		4						127		8	19	76	29	
Lestes	2			2			4		1	1		2	127		8	19	76	2 9	
Gomphidae	1						2						16		1		11	4	
Gomphus	1	1					2				1	1	17	1	1		11	4	
Orthoptera	3						1						4		1	2	1		
Blattidae	1		1						45										
Tettigoniidae	2	1	1		***		1		1				4	1		2	1		
Plecoptera	1		1				2				2						_		
Dermaptera	2	2			***							au = 11.							
Hemiptera	39 (44))					47			***			60		2	21	11	26	
Corixidae	31	1	4	8	4	14	4	1	1		2		54	1	2	19	7	25	
Veliidae	1	1				4+ 8+a.	7	3		2		2	4	2		2			

	1970							able V. (1972								
Specimens	Total	Control	NE	SE	NW	SW	rotal	Control	NE	SE	NW	SW	Total	Control	NE	SE	NW	SW
Notonectidae	1				***		33						5				4	1
Buenoa	1						33	7	5	5	10	6	9	4			4	1
Belostomatidae	3																	
Lethocerus							3		1			2						
Haliplidae	1						4						14		4	1		9
Feltodytes	1	1					4				2	2	14		4	1		9
Nitidulidae	1		1			» ——	8	1	3		4		1	1		-		
Noteridae	6						1	1						er ===				
Hydrocanthus	6	5			1		1				1							
Curculionidae	29						28						28		3	8	8	9
<u>Lissorhoptru</u> s	29	6	4	2	4	13	28	9	5	1	5	8	35	8	2	8	8	9
Hydraenidae	1						1											
Hydraena	1				1		1		1									
Cantharidae	1			. 1														
Scarabaeidae	5	1		2	1	1	4	1	1	1	1		2	1		1		
Carabidae	3	1		1	1		13		2	1	10		22	1		1	16	L
Staphylinida e	1				1													
Other (non-aquatic)) 3	1	2		1													
Homoptera							33	7	2	7	5	11	110	4	33	9	51	13
Neuroptera							16						2					ر. د
Corydalidae				*			15						2					2

			1070						Table V. (continued)							1972			
				197	0					171						• 71			
=	Specimens	Total	Control	NE	SE	NW	SW	lotal	Control	NE.	SE	NW	SW	Total	Control	NE	SE	NW 	SW
	<u>Corydalus</u>							15	14	1				9	7				2
	Sialidae						4	1											
	Sialus							1		1									
(Coleoptera							361						310		32	45	76	157
	Hydrophilidae							92	€ = •• =					49		7	9	13	20
	<u>Hya</u> rophilus							13	9	1		3		53	5	6	9	13	20
	Berosus							8		1		1	6						
	<u>Tropisternu</u>		****					26	3	10	3	4	6		-				
	Helophorus							20	3	1	5	8	3		-5-		•		
	Paracymus							20	1	9			10						
21	Hydrochus							5		2		2	1						
	Neohyarophilusc	astus -						9		3		6							
	Gyrinidae							8	***	8									
	Helodidae							3				1	2	5		1	2	1	1
	Dytiscidae							199				***		123	***	8	13	3 ੪	64
	Dytiscus							10	5	2		1	2	125	3	7	13	38	64
	Desmopachria							18	5	1	9	3							
	Bidessus							2		2									-
	Laccophilus							6	1 9 * * 19	3	2	1							
	Hydrovatus							86	25	7		3	51						
	Acilus							76	17	17	11	23	8						

							•T.	able'V.	(cont	inued)		r.					
			19	70			1971					1972						
Specimens	Total C	ontrol	NE	SE	NW	SW	Total	Control	NE	SE	NW	SW	Total	Control	NE	SE	NW	SW
Suphiseltus							1				1							
Lepidoptera	4	1	2		1		4			3		1	2	1		1		
Diptera	608(635)						1315						1007		125	147	294	441
Chironomidae	72						131						489		68	72	256	93
Pupae	1	1																
Pentaneura	27	6.	1	5	7	8	63	22	1	30	1	9	15	14				1
Tendipes	25	14		3	4	4	68	8	54	5		1	499	11	68	72	256	92
Unidentified	19	1	3	2		13												
Sciomyzidae	1	1	÷- •				2			2			40	1	7	5	16	11
Culicidae	480	366	17	24	31	42	1085						372	18 3	12	3 8	40	199
Culicidae pupa	e 12	3			8	1	12		9	3			7	2	2	2		1
Psorophora con	finnis 432	35 0	10	23	28	21	598	306	88	24	17	163	720	328	29	9	39	315
Aedes vexans	19	10	2		6	1	429	313	14	36	27	39	204	162	16	26		
A. quadrimacula	atus 6	1	1	1	2	1	7	2		3	2		3	2	1			
<u>Culex</u> salinari	<u>us</u> 2				2		4			3		1	3		3			
<u>C. territans</u>	1	1					2	1	1				2	1				1
C. quinquefasc	iatus 3	1	2				26	5	3	17	1		5	3	1			1
Chaoborus	1		1				8	2	1	1	3	1	20	1	15	1	1	2
Unidentified la	ar va 4		1		3													
Mycetophillidae	1					1	4		1	1	2	1						
Ceratopogonidae	6						21						20				7	13

							, 1	Table V.	(cont	tinue	1)		•					
	_		197	70			1971					1 972						
Specimens	Total	Control	NE	SE	NW	SW	Total	Control	NE	SE	NW	SW	Total	Control	NE	SE	NW	SW
Culicoides	6	^ت 1	1		2	2	21		6	15				s				
Sciaridae	1		-			1	5	1	3		1		28	~~ ~		8	1	19
Simulidae	3		2			1	5	1	3		1		2	1				1
Tabanidae	***												1				1	
Otitidae	1	1					2			2			1	1				
Ephydridae	3	~~~			1	2	1		1						6 978			
Strationyidae	4						1						3				1	2
Eulalia	4	3				1	1			1			5	2			1	2
Ti pulidae	1						55						10	~ **		3	4	3
Tipula	1		1				55	7	4	22	16	6	14	4	-	3	4	3
Cecidomyiidae	1			1			5	1	3			1	2	1		-	1	
Other dipterous	larva 57	1	2	5	45	4												
Other dipterous	adults 4		1	2	1													
Hymenoptera	22		7	4	3	8	62	13	21	5	3	20	57	7	14	14	8	14
Annelida	1				_ 1		24	3		_ 4		17	16	2	1	6		7
Nematode	4	1		1		2	38	7	1	13	12	5	63	- 4	13	38	8	
Anura	48		13	3 2	1	2	120	16	16	29	20	39						
Osteichthyes	3	2	1				86	21	11	9	27	18			•••			
	Tot	al numbe	er of	' samp	les:	588	Tota	l number	of s	ample	s: 2	277	Tot	al numbe:	r of	samp]	les:	427

			Northea	st section			
Date	1970	1971	1972	Date	1970	1971	1972
May 19		7.75		July 8			
25			2.75**	9			
30			4.33**	10	2.00		
tune 2	0.62			11		2 ()04	4.25*
	0.02			12		3.00*	10 / 0*
4				1)	1 00		19.40*
6	2.25		10.67#	14	1.00		
- 7				16	0.50		
8		29.16		17			
9	0.66		7.33*	18			6.33 *
10	7.50	2.00		20			
11				21		1.00**	
12	2.25			22			
13			2.60*	23		1.75*	
15		*****	***	25			3.50*
16	11.00		8.00*	27	1.50		
17		26.66*		28	0.50		6 . 33**
18		······································		29		2.14*	
19	1 00			30			
20	1.00		12.17*	4			0.00**
21	0.83			August 2			9.20**
23	0.05	11 81++	9 00*	4		1.50**	
24		2.33*	7.00^	5	2 33	1 66*	
25	3,25	~•>>>		8	~• <i>))</i>		11.67*
26				9			
27			6.17×	10	1.33	16.25*	
28				11			
30			10 .71**	12	0.66	*****	
				13	1.00		
July 1		5.00**		14			
5			8 .33**	17			
6				18		3.00*	
7		7 .3 3**	6.17*	26	1.25		

Table VI. Comparison of average numbers of aquatic arthropods by years and sections, Lonoke, Arkansas.

* Post-treatment.

** Pre-treatment.

Date	1970	1971	1972	Date	1970	1971	1972
May 19		10.60		July 8		2.50	
25			14.00**	9			
30 -	() () () () () () () () () () () () () (49 . 50 **	10	1.00		
				11			• /2*
June 2	1.43			12			20 108
4				13	1 00		20.40*
5	1.50			14	1.00		
6	4.00		5.67*	15			
7		13.16		16	0.50	Ch 100 Car (1) Ch	
8	5.75	4.00*		17			2 (7 -
9			6.33*	18			3.0/*
10	10.50			20	1.33		
11				21		5.00*	
12	3.33			22	1.50		
13			8.33*	23	0.50	2.83*	
15				25			4.80*
16	3.75		18.80*	27		0.66 *	
17	0.50	6.66*		28	13.50		43.50**
18				29	0.50		
19	1.00	Arr		30	0.66	2.00*	
20	0.50	1.00*	16.74*				
21				August 2		3.00*	20.00*
22	0.14	-		4		1.00	
23	1.25	1.00**	6 .33 *	5			
24		4.00*		6			
25				8			14.00*
26	0.75			9			
27			1.40×	10	4.0 0	1.16**	
28		3.33*		11		2.40*	
30			5.00*	12			
				13	3.33		
July 1				14	0.50		
5		The set of the set of the	1.50**	17	1.00		
6	5.00	6.66**		18			
7	1.00	3.00**	2 . 17*	26	4.00		

Southeast section

* Post-treatment.

** Pre-treatment.

Table VI. (continued)

Date	1970	1971	1972	Date	1970	<u>1971</u>	1972
May 19		20.16		July 8	0.20	3.00*	
25			3.67**	9		2.33*	
30			5.24**	10			
	1/23		<i>,</i> ,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	11	<u>82</u>		9.80*
June 2	2.80			12			
4	3.00			13	5.00		104.80*
-5				14	8.00		
6	1.50		8.40*	15			
7				16	0.66		
8	3.00	1.00		17			
9	9.20		21.60*	18			8 .50 *
10	3.50	9.40		20	0.50		
11				21	0.50	2.00**	
12	1.00		÷ 241-0-15	22	0.50		
13			5.67*	23	1.00	1.60 *	
15		A		25			1.33*
16	9.00		35 . 50*	27	4.00		
17	1.00			28			7.50*
18				29	4.00	23 . 33**	
19	9.00			30	2.50	5.50*	
20	1.00		3.20*				
21		**		August 2		1.00*	7.40**
22	5.00			- 4	2.00	5.00	
23	2.66		2. 50 *	5	2.33		
24	1.66	8 .60 *		6	0.50	0 .75**	
25	0.33			8			9.50*
26	1.00			9			
27			4.00 *	10	4.00	12.80*	
28				11	(1)-(1)-(1)-(1)-(1)-		
30	6.50		5.60**	12			
				13			
July 1	55.00			14			
5			3.33**	17			
6				18			
7	1.50		106.60*	26	3.00		

Northwest section

* Post-treatment.

****** Pre-treatment.

					8			
Date	2	1970	1971	1972	Date	1970	1971	1972
May	19		10.66		July 8	1.50	1.50*	
	25			9 . 00 **	9		1.83*	
	30			5 . 25**	10	3.33		
-	•	• • •			11			3.40*
June	2	2.44			12			
	4	3.66			13	4.00		14.67*
	5	4.00			14	1.00		
	6	1.50		51 . 50 *	15			
	7			*****	16	2.00	*****	
	8	5.00	8 . 33*		17	0.66	47.00**	
	9	3.75		65.00*	18			7.50*
	10				20			
	11				21	4.00		
	12	≈2,00			22	1.00		*****
	13			323.50*	23	1.00	2.25*	
	15				25			3.00*
x.	16	2.33		14.17 ×	27	2.00	43 . 33**	
	17				28	0.50	27.40*	31.80*
	18		5.00*		29	8.50	22.00	
	19	1.00			30			
	20			5.00*				
	21			Group an 00 m	August 2		<u></u>	18.17*
	22	2.50			4	6.00	2 . 33*	
	23		*****	4.00*	5	5.66		
	24	10.50	2 . 33*		6	3.00		
	25	2.00			8			12.83*
	26	0.75			9		1.00**	
	27		4+m	52.43*	10	5.20	32.66*	*****
	28				11		1.00*	
	30∍		*****	8.40**	12			
7 7	4		•/ //		13			
. JATA	-		16.66*		14	0.50		**************************************
	2			· 9.00*	17	2.33		
	0				18		0.66*	
	- 7	5.00		10 . 17*	26	3.66	*	

Southwest section

* Post-treatment.

****** Pre-treatment,

Table VI. (continued)

		_	Con	trol			
Date	1970	1971	1972	Date	1970	1971	1972
May 19		14.66	14.66	July 8	2.50	34.50	18.50
25				9		11.00	11.00
30				10	42.0 0		42.00
June 2			*====	12		15.00	15.00
4	24.66		24.66	13			
5	38.14		38.14	14	1.00		1.00
6	3.00		3.00	15	2.00		2.00
7		24.00	24.00	16	3.50		3.50
8		6.00	6.00	s 17		14.50	14.50
9	11.00		11.00	18			
10		28.75	28.75	20	6.00		6.00
11				21	4.00	21.50	12.75
12	2.25		2.25	22	-		
13				23	36.00	37.00	36.50
15		32.50	32.50	25		400	
16	6.60		6.60	27	3.00	17.00	10.00
17	3.50	25.50	14.50	28		8.50	8.50
18		20.00	20.00	29	2.00	15.00	8.50
19				30	8.66	27.00	17.83
20		11.00	11.00				
21		11.00	11.00	August 2		32.00	32.00
22				4	8.00	10.00	9.00
23		31.50	31.50	5	0.50		0.50
24		28.00	28.00	6		25.33	25.33
25		5.00	5.00	8		****	
26				⇒ 9		6.50	6.50
27				10	1.00	11.00	6.00
28		13.50	13.50	11		29.50	29.50
30		*****		12			
July 1		14.66	14.66	14			
5	*****			17	1.00		1.00
6	30.00	8.50	19.25	18		12.50	12.50
7		22.55	22.55	26	1.33		1.33

CONCLUSION

The data collected demonstrate that community mosquito control, with presently available chemicals, can be achieved in the rice-producing area of Arkansas. Such a program requires that someone be responsible for having the necessary work carried out when and where it is needed. The data also demonstrate that mosquito control chemicals, used as recommended, did not create a biological desert although a reduction in numbers of aquatic arthropods did occur after treatment. Abate and Flit MLO were the least damaging to non-target organisms.

LITERATURE CITED

- Burton, G. J. 1964. Results of insecticide resistance tests against larvae of <u>Culex p. quinquefasciatus</u> Say in British Guiana. Mosq. News 23:329-331.
- Cope, O. B. 1966. Contamination of the freshwater ecosystem by pesticides. J. Appl. Ecol. (Supplement on pesticides in the environment and their effects on wildlife). 3:33-44.
- Eisler, R. 1969. Acute toxicities of insecticides to marine decapod crustaceans. Crustaceana 16:302-310.

Exxon. 1973. Flit MLO: Product and properties.

- Fales, J. H., P. J. Spangler, O. F. Bodenstein, G. D. Mills, Jr. and C. G. Curbin, Jr. 1968. Laboratory and field evaluation of Abate against a backswimmer <u>Notonecta undulata</u> Say (hemiptera: Notonectidae). Mosq. News 29:77-81.
- Felton, H. C. 1944. Sample field and laboratory tests of larvicides. Mosq. News 4:50-54.
- FWPCA. 1968. Water quality criteria. Report of the National Tech. Adm. Comm. to Secr. of the Interior. Fed. Water Pollution Control Adm. USDI 234p.
- Gahan, J. B. and J. R. Noe. 1955. Control of mosquito larvae in ricefields with water soluble phosphorous insecticides. J. Econ. Entomol. 48:665-667.
- Geiger, J. C. <u>et al</u>. 1919. Effective malaria control in a ricefield district with observation on experimental mosquito flights. J. Med. Assoc. 72:844-847.

Humble. 1972. Aerial application of Flit MLO.

- Kiker, C. C. and H. E. Breedlove. 1941. Mosquito-proofing for malaria control from the standpoint of construction costs. Amer. J. Hyg. 34C:95-101.
- Kimball, J. H. and N. D. Perruzzi. 1970. Flit MLO preferred for mosquito control in residential areas. Mosq. News 30:128-130.
- Lewallen, L. L. and W. H. Wilder. 1963. Laboratory tests of insecticides on mosquito larvae in polluted and pure tapwater. J. Econ. Entomol. 56:834-835.
- Mount, G. A., N. W. Pierce, C. S. Lofgren and J. B. Gahan. 1970. A new ultra-low volume cold aerosol nozzle for dispersal of insecticides against adult mosquitoes. Mosq. News 30:56-59.

- Mulla, M. S. <u>et al</u>. 1961a. Effectiveness of new insecticides against mosquito larvae. Mosq. News 21:216-224.
- Mulla, M. S. 1961b. Susceptibility of various larval instars of <u>Culex p. quinquefasciatus</u> Say to insecticides. Mosq. News. 21:320-323.
- Muncy, R. J. and A. D. Oliver. 1963. Toxicity of ten insecticides to the red crawfish, <u>Procambarus clarki</u> (Girard). Trans. Am. Fish. Soc. 92-428-431.
- Paulini, E. <u>et al</u>. 1962. Laboratory tests with <u>Culex fatigans</u> on the persistence of Baytex on various types of surfaces. Rev. Bras. Malariol. 14:403-414.
- Peterle, T. J. and R. H. Giles. 1964. New tracer techniques for evaluating the effects of an insecticide on the ecology of a forest fauna. Ohio State Univ. Res. Found. Rep. 435 p.
- Pimentel, David. 1971. Ecological effects of pesticides on non-target species. Cornell University, Ithaca, N.Y.
- Porter, C. H. and W. L. Gojmerac. 1969. Field observations with Abate and Bromophos: Their effect on mosquitoes and aquatic arthropods in a Wisconsin park. Mosq. News 29:617-620.
- Ruber, Ernest. 1963. The effect of certain mosquito larvicides on microcrustacean populations. 50th Ann. Mtg. N. J. Mosq. Exterm. Assoc. and the 19th Ann. Mtg. Amer. Mosq. Contr. Assoc. P. 256-262.
- Ruber, Ernest. 1965a. The effects of chemical and physical methods of mosquito control on salt marsh microcrustacea. Unpubl. Ph.D. Dissert., St. Univ., Rutgers. 262 p.
- Ruber, Ernest. 1965b. The effects of certain mosquito larvicides on cultures of microcrustaceans--1963. Proc. 52nd Ann. Mtg. N. J. Mosq. Exterm. Assoc. P. 207-210.
- Sanders, H. O. 1969. Toxicity of pesticides to the crustacean, <u>Gammarus</u> <u>lacustris</u>. Tech. Paper 25, Bur. Sport Fish. Widl., USDI 18 p.
- Sanders, H. O. and O. B. Cope. 1966. Toxicities of several pesticides to two species of Cladocerans. Trans. Am. Fish Soc. 95:165-169.
- Stevens, L. F. <u>et al</u>. 1963. Evaluation of Baytex as a thermal aerosol for adult mosquito control. Proc. New Jersey Mosq. Exterm. Assoc. 50:351-354.
- Taylor, R. T. and H. F. Schoof. 1968. Evaluation of thermal and non-thermal fogs against four species of mosquitoes. Mosq. News 28:8-11.
- von Windeguth, D. L. and R. S. Patterson. 1966. The effects of two
 organic phosphate insecticides on segments of the aquatic biota.
 Mosq. News 26:377-380.

Whitehead, F. E. 1957. Flight range of ricefield mosquitoes. Agric. Exp. Sta. Bull. 590. Ark.