

12-28-2018

Checkpoint inhibitors: What gastroenterologists need to know.

Monjur Ahmed

Thomas Jefferson University, monjur.ahmed@jefferson.edu

Let us know how access to this document benefits you

Follow this and additional works at: https://jdc.jefferson.edu/gastro_hepfpPart of the [Gastroenterology Commons](#), and the [Hepatology Commons](#)

Recommended Citation

Ahmed, Monjur, "Checkpoint inhibitors: What gastroenterologists need to know." (2018). *Division of Gastroenterology and Hepatology Faculty Papers*. Paper 54.https://jdc.jefferson.edu/gastro_hepfp/54

This Article is brought to you for free and open access by the Jefferson Digital Commons. The Jefferson Digital Commons is a service of Thomas Jefferson University's [Center for Teaching and Learning \(CTL\)](#). The Commons is a showcase for Jefferson books and journals, peer-reviewed scholarly publications, unique historical collections from the University archives, and teaching tools. The Jefferson Digital Commons allows researchers and interested readers anywhere in the world to learn about and keep up to date with Jefferson scholarship. This article has been accepted for inclusion in Division of Gastroenterology and Hepatology Faculty Papers by an authorized administrator of the Jefferson Digital Commons. For more information, please contact: JeffersonDigitalCommons@jefferson.edu.

World Journal of *Gastroenterology*

World J Gastroenterol 2018 December 28; 24(48): 5415-5536



**EDITORIAL**

- 5415 Role of cenicriviroc in the management of nonalcoholic fatty liver disease
Neokosmidis G, Tziomalos K

REVIEW

- 5418 Colorectal cancer vaccines: Tumor-associated antigens *vs* neoantigens
Wagner S, Mullins CS, Linnebacher M

MINIREVIEWS

- 5433 Checkpoint inhibitors: What gastroenterologists need to know
Ahmed M
- 5439 Virtual reality simulation in endoscopy training: Current evidence and future directions
Mahmood T, Scaffidi MA, Khan R, Grover SC
- 5446 Quality of life in patients with minimal hepatic encephalopathy
Ridola L, Nardelli S, Gioia S, Riggio O
- 5454 Post-translational modifications of prostaglandin-endoperoxide synthase 2 in colorectal cancer: An update
Jaén RI, Prieto P, Casado M, Martín-Sanz P, Boscá L

ORIGINAL ARTICLE**Basic Study**

- 5462 Counteraction of perforated cecum lesions in rats: Effects of pentadecapeptide BPC 157, L-NAME and L-arginine
Drmic D, Samara M, Vidovic T, Malekinusic D, Antunovic M, Vrdoljak B, Ruzman J, Milkovic Perisa M, Horvat Pavlov K, Jeyakumar J, Seiwerth S, Sikiric P
- 5477 Mismatched effects of receptor interacting protein kinase-3 on hepatic steatosis and inflammation in non-alcoholic fatty liver disease
Saeed WK, Jun DW, Jang K, Ahn SB, Oh JH, Chae YJ, Lee JS, Kang HT
- 5491 Near-infrared photoimmunotherapy of pancreatic cancer using an indocyanine green-labeled anti-tissue factor antibody
Aung W, Tsuji AB, Sugyo A, Takashima H, Yasunaga M, Matsumura Y, Higashi T
- 5505 Integrated metabolomic profiling for analysis of antilipidemic effects of *Polygonatum kingianum* extract on dyslipidemia in rats
Yang XX, Wei JD, Mu JK, Liu X, Dong JC, Zeng LX, Gu W, Li JP, Yu J



Retrospective Study

- 5288 Safety of hepatitis B virus core antigen-positive grafts in liver transplantation: A single-center experience in China

Lei M, Yan LN, Yang JY, Wen TF, Li B, Wang WT, Wu H, Xu MQ, Chen ZY, Wei YG

ABOUT COVER

Editorial board member of *World Journal of Gastroenterology*, Harald Peter Hoensch, MD, Emeritus Professor, Marien Hospital Medical Department, Private Practice in Internal Medicine and Gastroenterology, Darmstadt D-64285, Germany

AIMS AND SCOPE

World Journal of Gastroenterology (*World J Gastroenterol*, *WJG*, print ISSN 1007-9327, online ISSN 2219-2840, DOI: 10.3748) is a peer-reviewed open access journal. *WJG* was established on October 1, 1995. It is published weekly on the 7th, 14th, 21st, and 28th each month. The *WJG* Editorial Board consists of 642 experts in gastroenterology and hepatology from 59 countries.

The primary task of *WJG* is to rapidly publish high-quality original articles, reviews, and commentaries in the fields of gastroenterology, hepatology, gastrointestinal endoscopy, gastrointestinal surgery, hepatobiliary surgery, gastrointestinal oncology, gastrointestinal radiation oncology, gastrointestinal imaging, gastrointestinal interventional therapy, gastrointestinal infectious diseases, gastrointestinal pharmacology, gastrointestinal pathophysiology, gastrointestinal pathology, evidence-based medicine in gastroenterology, pancreatology, gastrointestinal laboratory medicine, gastrointestinal molecular biology, gastrointestinal immunology, gastrointestinal microbiology, gastrointestinal genetics, gastrointestinal translational medicine, gastrointestinal diagnostics, and gastrointestinal therapeutics. *WJG* is dedicated to become an influential and prestigious journal in gastroenterology and hepatology, to promote the development of above disciplines, and to improve the diagnostic and therapeutic skill and expertise of clinicians.

INDEXING/ABSTRACTING

World Journal of Gastroenterology (*WJG*) is now indexed in Current Contents®/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports®, Index Medicus, MEDLINE, PubMed, PubMed Central and Directory of Open Access Journals. The 2018 edition of Journal Citation Reports® cites the 2017 impact factor for *WJG* as 3.300 (5-year impact factor: 3.387), ranking *WJG* as 35th among 80 journals in gastroenterology and hepatology (quartile in category Q2).

EDITORS FOR THIS ISSUE

Responsible Assistant Editor: *Xiang Li*
Responsible Electronic Editor: *Shu-Yu Yin*
Proofing Editor-in-Chief: *Lian-Sheng Ma*

Responsible Science Editor: *Rao-Yu Ma*
Proofing Editorial Office Director: *Ze-Mao Gong*

NAME OF JOURNAL

World Journal of Gastroenterology

ISSN

ISSN 1007-9327 (print)
 ISSN 2219-2840 (online)

LAUNCH DATE

October 1, 1995

FREQUENCY

Weekly

EDITORS-IN-CHIEF

Andrzej S Tarnawski, MD, PhD, DSc (Med), Professor of Medicine, Chief Gastroenterology, VA Long Beach Health Care System, University of California, Irvine, CA, 5901 E. Seventh Str., Long Beach, CA 90822, United States

EDITORIAL BOARD MEMBERS

All editorial board members resources online at <https://www.wjgnet.com/1007-9327/editorialboard.htm>

EDITORIAL OFFICE

Ze-Mao Gong, Director
World Journal of Gastroenterology
 Baishideng Publishing Group Inc
 7901 Stoneridge Drive, Suite 501,
 Pleasanton, CA 94588, USA
 Telephone: +1-925-2238242
 Fax: +1-925-2238243
 E-mail: editorialoffice@wjgnet.com
 Help Desk: <https://www.f0publishing.com/helpdesk>
<https://www.wjgnet.com>

PUBLISHER

Baishideng Publishing Group Inc
 7901 Stoneridge Drive, Suite 501,
 Pleasanton, CA 94588, USA
 Telephone: +1-925-2238242
 Fax: +1-925-2238243
 E-mail: bpgoffice@wjgnet.com
 Help Desk: <https://www.f0publishing.com/helpdesk>
<https://www.wjgnet.com>

PUBLICATION DATE

December 28, 2018

COPYRIGHT

© 2018 Baishideng Publishing Group Inc. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT

All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

INSTRUCTIONS TO AUTHORS

Full instructions are available online at <https://www.wjgnet.com/bpg/gerinfo/204>

ONLINE SUBMISSION

<https://www.f0publishing.com>

Checkpoint inhibitors: What gastroenterologists need to know

Monjur Ahmed

Monjur Ahmed, Division of Gastroenterology and Hepatology, Department of Internal Medicine, Thomas Jefferson University, Philadelphia, PA 19107, United States

ORCID number: Monjur Ahmed (0000-0003-0515-9224).

Author contributions: Ahmed M finished this manuscript alone.

Conflict-of-interest statement: No conflict of interest exists.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Corresponding author: Monjur Ahmed, MD, FRCP, Division of Gastroenterology and Hepatology, Department of Internal Medicine, Thomas Jefferson University, 132 South 10th Street, Suite 468, Main Building, Philadelphia, PA 19107, United States. monjur.ahmed@jefferson.edu.
Telephone: +1-215-9521493
Fax: +1-215-7551850

Received: September 28, 2018

Peer-review started: September 28, 2018

First decision: October 26, 2018

Revised: November 7, 2018

Accepted: November 16, 2018

Article in press: November 16, 2018

Published online: December 28, 2018

Abstract

Checkpoint inhibitors are increasingly being used in clinical practice. They can cause various gastrointestinal, hepatic and pancreatic side effects. As these side effects

can be serious, appropriate management is essential. The different checkpoint inhibitors with their mechanisms of action and indications, as well as evaluation and management of gastrointestinal, hepatic and pancreatic side effects, are discussed in this article.

Key words: Checkpoint inhibitors; Immunotherapy; Gastrointestinal management; Hepatic and pancreatic side effects

© **The Author(s) 2018.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Checkpoint inhibitors are a kind of immunotherapy used in the treatment of various malignancies. Nevertheless, they carry the risk of causing different immune-related side effects. Physicians should be vigilant in recognizing and appropriately managing these side effects for a better outcome.

Ahmed M. Checkpoint inhibitors: what gastroenterologists need to know. *World J Gastroenterol* 2018; 24(48): 5433-5438
URL: <https://www.wjgnet.com/1007-9327/full/v24/i48/5433.htm>
DOI: <https://dx.doi.org/10.3748/wjg.v24.i48.5433>

INTRODUCTION

Checkpoint inhibitors have emerged as one of the most promising modalities of anti-cancer therapy^[1]. They are monoclonal antibodies that block the checkpoint proteins either on T cells or cancer cells to enhance immune response against tumor cells^[2]. Normally, when our body recognizes cancer cells or foreign bodies, our innate immune system (macrophages, dendritic cells, natural killer cells, mast cells, neutrophils, eosinophils and basophils) tries to eliminate them. Then, our adaptive immune system (B lymphocytes and T lymphocytes) starts working *via* antigen presenting cells. Checkpoint molecules are proteins that control specific cellular

processes to prevent errors. Some immune checkpoint proteins help the T cells to remain active, particularly in case of infection, whereas other immune checkpoint proteins regulate the immune system negatively by directing the T cells to switch off. Some cancer cells synthesize high levels of such immune checkpoint proteins, which can switch off the T cells and, as a result, the T cells can neither recognize nor kill the cancer cells.

Some of the common checkpoint proteins include: (1) Cytotoxic T lymphocyte-associated antigen 4 (CTLA-4) receptors on CD4 and CD8 T lymphocytes; (2) programmed cell death protein 1 (PD-1) receptors on the surface of T cells, B cells, natural killer (NK) cells, monocytes and dendritic cells; and (3) programmed cell death protein ligand 1 (PD-L1) and programmed cell death protein ligand 2 (PD-L2) proteins on healthy tissues, hematopoietic cells and tumor cells.

When interactions between the PD-1 receptors and PD-L1 (also known as B7-H1) or PD-L2 (also known as B7-H2) occurs, it promotes exhaustion of peripheral effector T cells, conversion of effector T cells to regulatory T (Treg) cells and inhibition of tumor cell apoptosis^[3]. Some cancer cells are able to produce PD-L1 and PD-L2 on their surfaces to prevent any immunological attack.

CTLA-4 becomes activated by binding to B7-1 (also known as CD80) and B7-2 (also known as CD86) on antigen presenting cells (APCs), and then inhibits T cell activation at a proximal step in the immune response. On the other hand, PD-1 limits effector T cell function by linking with PD-L1 or PD-L2 in the later stages of the immune response. In the process of carcinogenesis, these immunosuppressive molecules are overexpressed^[4]. Checkpoint inhibitors are monoclonal antibodies against PD-1, PD-L1 or CTLA-4 proteins. They act as a form of immunotherapy by blocking the immunosuppressive molecules that otherwise inhibit the immune system from attacking cancer cells. As a consequence, there is an immunological boost against cancer cells^[5]. As they target T cells instead of cancer cells, they can be used in various malignancies^[6]. A combination of checkpoint inhibitors may give a better anti-tumor response. There was a 23% response rate for metastatic non-small cell lung cancer after administration of durvalumab and tremelimumab^[7].

Few checkpoint molecules have been discovered recently. These include TIM-3, LAG3, TIGIT and BTLA.

T cell immunoglobulin and mucin domain 3 (TIM-3) is present on the surface of CD4 T cells, CD8 T cells, regulatory T cells and innate immune cells (dendritic cells, macrophages and natural killer cells). TIM-3 binds to specific ligands: galectin (Gal-9), phosphatidyl serine (PtdSer), high-mobility group box-1 protein (HMGB) and carcinoembryonic antigen-related cell adhesion molecule 1 (CEACAM1). These interactions generate a variety of effects, including effector T cell apoptosis, T cell suppression, suppression of the innate immune response against tumor cells, suppression of anti-tumor activity and promotion of tumor growth^[8]. TIM-3 is upregulated

in patients with malignancy. In pre-clinical studies, TIM-3 monoclonal antibody monotherapy showed modest anti-tumor activities^[9], but combinations of anti-TIM-3 and anti-PD-1/PD-L1 monoclonal antibodies produced significant anti-tumor responses against a variety of malignancies, including colon cancer, lung cancer, ovarian cancer, melanoma, lymphoma, acute myelogenous leukemia and sarcoma^[10].

LAG-3 (lymphocyte activation gene-3 protein) is an inhibitory receptor expressed on CD4-positive T-lymphocytes, CD8-positive T-lymphocytes, NK cells and B cells, as well as on plasmacytoid dendritic cells^[11-13]. LAG-3 inhibits both activation and proliferation of T cells^[14,15]. Anti-LAG3 monoclonal antibodies can bind to the LAG-3 present on tumor infiltrating lymphocytes (TILs), and prevent their binding to MHC (major histocompatibility complex) class II molecules expressed on tumor cells. This may lead to activation of antigen-specific T lymphocytes and cytotoxic T cell-mediated tumor lysis. Clinical trials were done with different types of LAG-3 monoclonal antibodies (IMP321) on various malignancies, such as metastatic renal cell cancer, breast cancer, unresectable pancreatic cancer, as well as advanced and unresectable melanoma^[16].

T cell immunoreceptors with Ig and ITIM domains (TIGIT) are inhibitory immunoreceptors present on some T cells (CD4, CD8), NK cells and Treg cells that contain Ig and immunoreceptor tyrosine-based inhibitory motif (ITIM) domains. TIGIT ligands include CD155 and CD112. In certain malignancies, CD155 and CD112 are highly expressed on macrophages and dendritic cells. TIGIT ligation leads to inhibition of T cell proliferation and suppression of the cytolytic function of NK cells^[17]. Anti-tumor activity is suppressed by TIGIT, primarily *via* Treg cells and not CD8-positive T cells^[18]. Anti-TIGIT monoclonal antibodies as a monotherapy or in combination with anti-PD-L1 antibodies have shown anti-tumor activity^[19] in phase I / II trials.

BTLA (a B and T lymphocyte attenuator, also known as CD272) is an inhibitory protein functionally and structurally similar to CTLA-4 and PD-1. It is mainly expressed on immune cells, NK cells, dendritic cells and splenic macrophages. BTLA acts as a ligand for tumor necrosis factor receptor superfamily member 14 (TNFRSF-14), also known as herpes virus entry mediator (HVEM).

BTLA/HVEM complex inhibits T cell activation and proliferation^[20]. BTLA is overexpressed in certain malignancies like leukemia and melanoma. In mouse models, BTLA neutralizing antibodies limited tumor growth^[21]. Anti-human BTLA monoclonal antibodies are currently in development.

CURRENT CHECKPOINT INHIBITORS

Current checkpoint inhibitors, with their indications and a schematic diagram (Figure 1), are mentioned below^[22-28], including CTLA-4 blockers, PD-1 inhibitors, and PD-L1

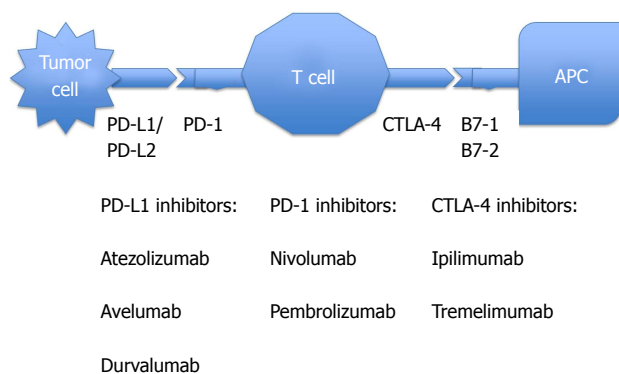


Figure 1 Schematic diagram of checkpoint inhibitors.

inhibitors.

CTLA-4 blockers

Ipilimumab: Indications include melanoma with lymph node involvement, advanced melanoma, non-small cell and small cell lung cancer, advanced renal cell cancer, and hormone refractory prostate cancer. Great success with durable clinical benefit was seen with nivolumab plus ipilimumab combination when given in DNA mismatch repair-deficient/microsatellite instability-high metastatic colorectal cancer^[29].

Tremelimumab: This drug is undergoing human clinical trials for the treatment of various malignancies, but is not yet approved by the United States Food and Drug Administration (FDA).

PD-1 inhibitors

Nivolumab: Indications are melanoma with lymph node involvement, unresectable or metastatic melanoma, advanced renal cell carcinoma, advanced or metastatic urothelial cancer, metastatic non-small cell lung cancer and small cell lung cancer with progression after platinum-based chemotherapy, refractory classical Hodgkin lymphoma, recurrent or metastatic squamous cell cancer of head and neck, microsatellite instability-high or mismatch repair-deficient colorectal cancer, and hepatocellular carcinoma.

Pembrolizumab: Indications include unresectable or metastatic melanoma, metastatic non-small cell lung cancer, advanced head and neck squamous cell carcinoma, advanced or metastatic gastric or gastroesophageal junction cancer, microsatellite instability-high cancer, locally advanced or metastatic urothelial cancer, recurrent or metastatic cervical cancer, refractory classical Hodgkin lymphoma, and refractory primary mediastinal large B cell lymphoma.

PD-L1 inhibitors

Atezolizumab: Indicated for advanced or metastatic non-small cell lung cancer and advanced or metastatic urothelial cancer.

Avelumab: Indicated for advanced or metastatic urothelial cancer and metastatic Merkel cell cancer.

Durvalumab: Indicated for advanced or metastatic urothelial cancer, as well as unresectable and stage III non-small lung cancer.

IMMUNE-RELATED ADVERSE EVENTS

Immune-related adverse events (IRAE) can occur due to the use of checkpoint inhibitors. Inflammatory side effects generally involve the skin, gastrointestinal tract, liver and endocrine glands. The cardiovascular, pulmonary, renal, hematological and musculoskeletal system are less commonly involved. IRAE are more severe following administration of CTLA-4 inhibitors in comparison to PD-1 or PD-L1 inhibitors. The time of onset of IRAE is generally 1-6 mo after administration of checkpoint inhibitors^[30]. Here, we will be mainly discussing the gastrointestinal, hepatic and pancreatic side effects of checkpoint inhibitors.

Immune-related adverse events are classified according to the National Cancer Institute's Common Terminology Criteria for Adverse Events (AE), version 3.0^[31]. Grade 1: Mild AE; Grade 2: Moderate AE; Grade 3: Severe AE; Grade 4: Life-threatening or disabling AE; Grade 5: Death related to AE.

Gastrointestinal

Diarrhea is the most common gastrointestinal side effect after administration of checkpoint inhibitors. Diarrhea occurs in 27%-31% of cases following CTLA-4 inhibitor therapy^[32] and less than 4% of cases following anti-PD-1 and anti-PD-L1 therapy^[33]. Diarrhea varies from mild to severe in intensity. Immune-mediated mild to severe colitis, colon perforation and even death can occur following checkpoint inhibitor therapy^[34,35]. Severe colitis can occur in 5% of cases following CTLA-4 inhibitor therapy and less than 2% of cases following anti-PD-1 therapy. Colitis may mimic Crohn's colitis or ulcerative colitis, and can be associated with intra-abdominal abscess, anal fissure and fistula^[36]. Pembrolizumab-induced collagenous colitis and lymphocytic colitis have also been reported in the literature^[37,38].

Management

Mild diarrhea or grade I diarrhea with stool frequency less than 4 times per day can be managed conservatively without discontinuing checkpoint inhibitors. Stool for ova, parasites, giardia antigen, stool culture and *C. difficile* toxin should be sent to evaluate for any underlying infection. Patient should be given adequate oral hydration and anti-diarrheal agents. If diarrhea still persists for 5-7 d or worsens, patients should be treated similar to cases of moderate diarrhea.

Moderate diarrhea or grade II diarrhea with stool frequency between 4-6 times per day should be managed by discontinuation of checkpoint inhibitors, by ruling out

infection by sending stool samples as mentioned above, by giving adequate oral hydration and by administering empiric treatment with oral corticosteroid (prednisone 1 mg/kg per day) with close clinical follow-up^[39]. Colonoscopy is not required if the patient responds to the above measures. After 2-5 d control of diarrhea, prednisone should be slowly tapered over a 1-2 mo period of time. Trimethoprim-sulfamethoxazole should be given as a prophylaxis against opportunistic infection during the tapering period^[40].

If the patient does not respond to the conservative measures or if the patient has severe diarrhea, *i.e.*, grade III or IV diarrhea, with stool frequency more than 6 times per day with severe and persistent abdominal pain, fever, rectal bleeding or ileus, intravenous hydration and intravenous steroid (methylprednisone 1-2 mg/kg per day) should be started^[41]. Antidiarrheal agents like loperamide and lomotil (diphenoxylate/atropine) should be avoided. Abdominal CT (computerized axial tomography) should be done to assess the severity and complications of colitis (perforation and peritonitis). Colonoscopy or flexible sigmoidoscopy should be done not only to evaluate the severity and extent of colitis, but also to take random and targeted colon biopsies to rule out underlying cytomegalovirus (CMV) infection^[35]. CMV colitis is diagnosed by characteristic histology (owl's eye intranuclear inclusion bodies), CMV biopsy PCR (polymerase chain reaction) or CMV biopsy viral culture^[42]. Colonoscopic findings may include loss of vascular markings, erythema, congestion, friability, ulcerations and spontaneous bleeding. The severity of diarrhea may not correlate with the colonoscopic or histological findings.

Treatment should be continued until there is significant improvement of diarrhea, *i.e.*, grade 0-1. If there is a clinical response to corticosteroids, it is recommended to continue treatment for a month and then slowly taper. If the patient's diarrhea is refractory to steroids, or colonoscopy shows severe colitis, multiple colon ulcers or pancolitis, anti-TNF therapy like Infliximab (5 mg/kg every 2 wk) or anti-integrin therapy like Vedolizumab (300 mg 0, 2, 6 wk) should be added^[43-45]. Concomitant bacterial, viral or *Clostridium difficile* infection should be treated simultaneously. Following resolution of symptoms, checkpoint inhibitors can be restarted if the benefits outweigh the risks, and if the daily dose of prednisone can be reduced to less than 10 mg per day without any other immunosuppressive medication.

Summary of diarrhea management

Diarrhea onset approximately 6 wk after checkpoint inhibitor therapy: assess severity of diarrhea and rule out infection by sending stool samples → Grade I diarrhea: conservative treatment with oral hydration and anti-diarrheal agents → Persistence of diarrhea after 5-7 d → Manage as grade II diarrhea. Grade II diarrhea: stop checkpoint inhibitor, start oral corticosteroid and continue oral hydration: (1) Clinical improvement → 2-5 d after

control of diarrhea, start tapering corticosteroid over 1-2 mo plus trimethoprim-sulfamethoxazole as prophylaxis; (2) no clinical improvement → Manage as grade III or IV diarrhea. Grade III or IV diarrhea: hospitalization, parenteral hydration, parenteral corticosteroid, abdominal CT and colonoscopy: (1) Clinical response: continue steroid for a month and taper; (2) no clinical response: anti-TNF therapy.

Hepatic

Checkpoint inhibitors can cause immune-mediated hepatitis in less than 5% of cases receiving these medications^[46]. Although this can occur anytime while the patient is on checkpoint inhibitor therapy, it occurs most commonly 6-7 wk after the onset of therapy^[47]. Most of the time, patients remain asymptomatic with elevated serum transaminases. Sometimes, the hepatitis can be more severe, with patients presenting with fever, malaise, fatigue, hepatomegaly and hyperbilirubinemia. Acute viral hepatitis (HAV, HBV, HCV, EBV, CMV) and autoimmune hepatitis need to be excluded by serology and liver biopsy^[48]. Predominant hepatic parenchymal injury with panlobular hepatitis or predominant bile duct injury with mononuclear cell infiltration around proliferated bile ductules can be seen after checkpoint inhibitor therapy^[49]. It is sometimes difficult to distinguish autoimmune hepatitis from drug-induced hepatitis. In autoimmune hepatitis, intra-acinar and portal plasma cells with rosette formation and emperipolesis are prominent, whereas neutrophilic infiltration is more commonly seen in drug-induced liver injury^[50].

Management^[51]

Grade 1 immune-mediated hepatitis: patient is asymptomatic or mildly symptomatic, but laboratory studies show AST/ALT: $< 2.5 \times \text{ULN}$ (upper limit of normal) and total bilirubin: $< 1.5 \times \text{ULN}$. Treatment: continue checkpoint inhibitor therapy, but monitor LFT.

Grade 2 immune-mediated hepatitis: patient is symptomatic (fever, malaise, fatigue) and laboratory studies show AST/ALT: $2.5-5 \times \text{ULN}$ and total bilirubin: $1.5-3 \times \text{ULN}$. Treatment: (1) hold checkpoint inhibitor therapy; (2) viral hepatitis (HAV, HBV, HCV, HDV, CMV, EBV, HSV, VZV), autoimmune hepatitis and drug-induced hepatitis need to be ruled out^[46]; (3) prednisone 1 mg/kg per day, taper the dose when patient's symptoms improve; and (4) if symptoms do not improve after 48 h, alternate immunosuppressive agents like tacrolimus, mycophenolate mofetil or cyclophosphamide need to be considered.

Grade 3 or 4 immune-mediated hepatitis: patient is symptomatic as mentioned above, and laboratory studies show: AST/ALT: $> 5 \times \text{ULN}$; total bilirubin: $> 3 \times \text{ULN}$. Treatment: (1) hold checkpoint inhibitors; (2) intravenous solumedrol 2-4 mg/kg per day. Taper the dose when patient's symptoms improve; and (3) if symptoms do not improve after 5-7 d, tacrolimus 0.10-0.15 mg/kg per day should be added. Alternative agents include

mycophenolate mofetil or cyclophosphamide.

Pancreatic

Immune-mediated pancreatitis with pancreatic insufficiency has been reported a few months after initiation of checkpoint inhibitor therapy^[52]. Asymptomatic elevations of amylase and lipase can occur without fulfilling the diagnostic criteria of acute pancreatitis. As the clinical significance of this lab abnormality is unknown, routine measurement of serum amylase and lipase is not recommended^[53]. However, if the patient is symptomatic with abdominal pain or nausea, immune-mediated pancreatitis should be considered by checking amylase and lipase levels and performing imaging studies.

Management

Intravenous methylprednisolone (1 mg/kg per day) for a few days, followed by oral prednisone (1 mg/kg per day). Taper the dose when patient symptoms improve.

Pancreatic enzyme supplementation should be given if there is evidence of pancreatic insufficiency (fecal elastase < 15 µg/g of feces).

CONCLUSION

Checkpoint inhibitors are novel forms of immunotherapy administered by oncologists. Although they are extremely useful in various advanced and metastatic malignancies, they can cause multiple side effects. Gastroenterologists need to be aware of the various gastrointestinal, hepatic and pancreatic side effects that can be fatal if not managed early. Prompt recognition of these side effects, administration of systemic immunosuppressive therapy and supportive care could improve the clinical outcome without affecting the benefit of checkpoint inhibitors. Multidisciplinary teams should be involved in the management of these side effects. As new checkpoints are being discovered and new checkpoint inhibitors are being developed, patients will be experiencing new IRAE. Management of those IRAE will improve as we gather more experience using new checkpoint inhibitors.

REFERENCES

- 1 **Chen Q**, Wang C, Chen G, Hu Q, Gu Z. Delivery Strategies for Immune Checkpoint Blockade. *Adv Healthc Mater* 2018; **7**: e1800424 [PMID: 29978565 DOI: 10.1002/adhm.201800424]
- 2 **Trivedi MS**, Hoffner B, Winkelmann JL, Abbott ME, Hamid O, Carvajal RD. Programmed death 1 immune checkpoint inhibitors. *Clin Adv Hematol Oncol* 2015; **13**: 858-868 [PMID: 27058852]
- 3 **Dine J**, Gordon R, Shames Y, Kasler MK, Barton-Burke M. Immune Checkpoint Inhibitors: An Innovation in Immunotherapy for the Treatment and Management of Patients with Cancer. *Asia Pac J Oncol Nurs* 2017; **4**: 127-135 [PMID: 28503645 DOI: 10.4103/apjon.apjon_4_17]
- 4 **Mellman I**, Coukos G, Dranoff G. Cancer immunotherapy comes of age. *Nature* 2011; **480**: 480-489 [PMID: 22193102 DOI: 10.1038/nature10673]
- 5 **Oiseth SJ**, Aziz MS. Cancer immunotherapy: a brief review of the history, possibilities, and challenges ahead. *J Cancer Met Tre* 2017; **3**: 250-261 [DOI: 10.20517/2394-4722.2017.41]
- 6 **Assarzagdean N**, Montgomery E, Anders RA. Immune checkpoint

- inhibitor colitis: the flip side of the wonder drugs. *Virchows Arch* 2018; **472**: 125-133 [PMID: 29143108 DOI: 10.1007/s00428-017-2267-z]
- 7 **Antonia S**, Goldberg SB, Balmanoukian A, Chaft JE, Sanborn RE, Gupta A, Narwal R, Steele K, Gu Y, Karakunnel JJ, Rizvi NA. Safety and antitumor activity of durvalumab plus tremelimumab in non-small cell lung cancer: a multicentre, phase 1b study. *Lancet Oncol* 2016; **17**: 299-308 [PMID: 26858122 DOI: 10.1016/S1470-2045(15)00544-6]
- 8 **Liu F**, Liu Y, Chen Z. Tim-3 expression and its role in hepatocellular carcinoma. *J Hematol Oncol* 2018; **11**: 126 [PMID: 30309387 DOI: 10.1186/s13045-018-0667-4]
- 9 **Ngio SF**, von Scheidt B, Akiba H, Yagita H, Teng MW, Smyth MJ. Anti-TIM3 antibody promotes T cell IFN-γ-mediated antitumor immunity and suppresses established tumors. *Cancer Res* 2011; **71**: 3540-3551 [PMID: 21430066 DOI: 10.1158/0008-5472.CAN-11-0096]
- 10 **Sakuishi K**, Apetoh L, Sullivan JM, Blazar BR, Kuchroo VK, Anderson AC. Targeting Tim-3 and PD-1 pathways to reverse T cell exhaustion and restore anti-tumor immunity. *J Exp Med* 2010; **207**: 2187-2194 [PMID: 20819927 DOI: 10.1084/jem.20100643]
- 11 **Triebel F**, Jitsukawa S, Baixeras E, Roman-Roman S, Genevee C, Viegas-Pequignot E, Hercend T. LAG-3, a novel lymphocyte activation gene closely related to CD4. *J Exp Med* 1990; **171**: 1393-1405 [PMID: 1692078]
- 12 **Kisielow M**, Kisielow J, Capoferri-Sollami G, Karjalainen K. Expression of lymphocyte activation gene 3 (LAG-3) on B cells is induced by T cells. *Eur J Immunol* 2005; **35**: 2081-2088 [PMID: 15971272 DOI: 10.1002/eji.200526090]
- 13 **Workman CJ**, Wang Y, El Kasmi KC, Pardoll DM, Murray PJ, Drake CG, Vignali DA. LAG-3 regulates plasmacytoid dendritic cell homeostasis. *J Immunol* 2009; **182**: 1885-1891 [PMID: 19201841 DOI: 10.4049/jimmunol.0800185]
- 14 **Blackburn SD**, Shin H, Haining WN, Zou T, Workman CJ, Polley A, Betts MR, Freeman GJ, Vignali DA, Wherry EJ. Coregulation of CD8+ T cell exhaustion by multiple inhibitory receptors during chronic viral infection. *Nat Immunol* 2009; **10**: 29-37 [PMID: 19043418 DOI: 10.1038/ni.1679]
- 15 **Huard B**, Prigent P, Tournier M, Bruniquel D, Triebel F. CD4/major histocompatibility complex class II interaction analyzed with CD4- and lymphocyte activation gene-3 (LAG-3)-Ig fusion proteins. *Eur J Immunol* 1995; **25**: 2718-2721 [PMID: 7589152 DOI: 10.1002/eji.1830250949]
- 16 **Andrews LP**, Marciscano AE, Drake CG, Vignali DA. LAG3 (CD223) as a cancer immunotherapy target. *Immunol Rev* 2017; **276**: 80-96 [PMID: 28258692 DOI: 10.1111/imr.12519]
- 17 **Johnston RJ**, Comps-Agrar L, Hackney J, Yu X, Huseni M, Yang Y, Park S, Javinal V, Chiu H, Irving B, Eaton DL, Grogan JL. The immunoreceptor TIGIT regulates antitumor and antiviral CD8(+) T cell effector function. *Cancer Cell* 2014; **26**: 923-937 [PMID: 25465800 DOI: 10.1016/j.ccell.2014.10.018]
- 18 **Kurtulus S**, Sakuishi K, Ngio SF, Joller N, Tan DJ, Teng MW, Smyth MJ, Kuchroo VK, Anderson AC. TIGIT predominantly regulates the immune response via regulatory T cells. *J Clin Invest* 2015; **125**: 4053-4062 [PMID: 26413872 DOI: 10.1172/JCI81187]
- 19 **Solomon BL**, Garrido-Laguna I. TIGIT: a novel immunotherapy target moving from bench to bedside. *Cancer Immunol Immunother* 2018; **67**: 1659-1667 [PMID: 30232519 DOI: 10.1007/s00262-018-2246-5]
- 20 **Zhang M**, Howard K, Winters A, Steavenson S, Anderson S, Smelt S, Doellgast G, Sheelo C, Stevens J, Kim H, Hamburger A, Sein A, Caughey DJ, Lee F, Hsu H, Siu G, Byrne FR. Monoclonal antibodies to B and T lymphocyte attenuator (BTLA) have no effect on in vitro B cell proliferation and act to inhibit in vitro T cell proliferation when presented in a cis, but not trans, format relative to the activating stimulus. *Clin Exp Immunol* 2011; **163**: 77-87 [PMID: 21078085 DOI: 10.1111/j.1365-2249.2010.04259.x]
- 21 **Sekar D**, Govene L, Del Rio ML, Sirait-Fischer E, Fink AF, Brüne B, Rodriguez-Barbosa JI, Weigert A. Downregulation of BTLA on NKT Cells Promotes Tumor Immune Control in a Mouse Model of Mammary Carcinoma. *Int J Mol Sci* 2018; **19** [PMID: 29518903 DOI: 10.3390/ijms19030752]

- 22 **Postow MA**, Sidlow R, Hellmann MD. Immune-Related Adverse Events Associated with Immune Checkpoint Blockade. *N Engl J Med* 2018; **378**: 158-168 [PMID: 29320654 DOI: 10.1056/NEJMra1703481]
- 23 Medscape. Ipilimumab (Rx). Available from: URL: <https://reference.medscape.com/drug/yervoy-ipilimumab-999636>
- 24 Medscape. Nivolumab (Rx). Available from: URL: <https://reference.medscape.com/drug/opdivo-nivolumab-999989>
- 25 Medscape. Pembrolizumab (Rx). Available from: URL: <https://reference.medscape.com/drug/keytruda-pembrolizumab-999962>
- 26 Medscape. Atezolizumab (Rx). Available from: URL: <https://reference.medscape.com/drug/tecentriq-atezolizumab-1000098>
- 27 Medscape. Avelumab (Rx). Available from: URL: <https://reference.medscape.com/drug/bavencio-avelumab-1000144>
- 28 Medscape. Durvalumab (Rx). Available from: URL: <https://reference.medscape.com/drug/imfinzi-durvalumab-1000145>
- 29 **Overman MJ**, Lonardi S, Wong KYM, Lenz HJ, Gelsomino F, Aglietta M, Morse MA, Van Cutsem E, McDermott R, Hill A, Sawyer MB, Hendlisz A, Neyns B, Svrcek R, Moss RA, Ledezne JM, Cao ZA, Kamble S, Kopetz S, André T. Durable Clinical Benefit With Nivolumab Plus Ipilimumab in DNA Mismatch Repair-Deficient/Microsatellite Instability-High Metastatic Colorectal Cancer. *J Clin Oncol* 2018; **36**: 773-779 [PMID: 29355075 DOI: 10.1200/JCO.2017.76.9901]
- 30 **Eigentler TK**, Hassel JC, Berking C, Aberle J, Bachmann O, Grünwald V, Kähler KC, Loquai C, Reinmuth N, Steins M, Zimmer L, Sendl A, Gutzmer R. Diagnosis, monitoring and management of immune-related adverse drug reactions of anti-PD-1 antibody therapy. *Cancer Treat Rev* 2016; **45**: 7-18 [PMID: 26922661 DOI: 10.1016/j.ctrv.2016.02.003]
- 31 Cancer Therapy Evaluation Program, Common Terminology Criteria for Adverse Events, Version 3.0. Available from: URL: https://ctep.cancer.gov/protocolDevelopment/electronic_applications/ctc.htm#ctc_40
- 32 **Hodi FS**, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, Gonzalez R, Robert C, Schadendorf D, Hassel JC, Akerley W, van den Eertwegh AJ, Lutzky J, Lorigan P, Vaubel JM, Linette GP, Hogg D, Ottensmeier CH, Lebbé C, Peschel C, Quirt I, Clark JI, Wolchok JD, Weber JS, Tian J, Yellin MJ, Nichol GM, Hoos A, Urba WJ. Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med* 2010; **363**: 711-723 [PMID: 20525992 DOI: 10.1056/NEJMoa1003466]
- 33 **Wang PF**, Chen Y, Song SY, Wang TJ, Ji WJ, Li SW, Liu N, Yan CX. Immune-Related Adverse Events Associated with Anti-PD-1/PD-L1 Treatment for Malignancies: A Meta-Analysis. *Front Pharmacol* 2017; **8**: 730 [PMID: 29093678 DOI: 10.3389/fphar.2017.00730]
- 34 **Celli R**, Kluger HM, Zhang X. Anti-PD-1 Therapy-Associated Perforating Colitis. *Case Rep Gastrointest Med* 2018; **2018**: 3406437 [PMID: 29955400 DOI: 10.1155/2018/3406437]
- 35 **Prieux-Klotz C**, Dior M, Damotte D, Dreanic J, Brieau B, Brezault C, Abitbol V, Chaussade S, Coriat R. Immune Checkpoint Inhibitor-Induced Colitis: Diagnosis and Management. *Target Oncol* 2017; **12**: 301-308 [PMID: 28540478 DOI: 10.1007/s11523-017-0495-4]
- 36 **Bertha M**, Bellaguara E, Kuzel T, Hanauer S. Checkpoint Inhibitor-Induced Colitis: A New Type of Inflammatory Bowel Disease? *ACG Case Rep J* 2017; **4**: e112 [PMID: 29043290 DOI: 10.14309/crj.2017.112]
- 37 **Baroudjian B**, Lourenco N, Pagès C, Chami I, Maillat M, Bertheau P, Bagot M, Gornet JM, Lebbé C, Allez M. Anti-PD1-induced collagenous colitis in a melanoma patient. *Melanoma Res* 2016; **26**: 308-311 [PMID: 26990271 DOI: 10.1097/CMR.0000000000000252]
- 38 **Ahmed M**, Francis G. Pembrolizumab-Induced Microscopic Colitis. *Am J Gastroenterol* 2018; **113**: 629-630 [PMID: 29610496 DOI: 10.1038/ajg.2018.8]
- 39 **Rudzki JD**. Management of adverse events related to checkpoint inhibition therapy. *Memo Eur Med Oncol* 2018; **11**: 132-137 [DOI: 10.1007/s12254-018-0416-y]
- 40 **Cooley L**, Dendle C, Wolf J, Teh BW, Chen SC, Boutlis C, Thursky KA. Consensus guidelines for diagnosis, prophylaxis and management of *Pneumocystis jirovecii* pneumonia in patients with haematological and solid malignancies, 2014. *Intern Med J* 2014; **44**: 1350-1363 [PMID: 25482745 DOI: 10.1111/imj.12599]
- 41 **Cheng R**, Cooper A, Kench J, Watson G, Bye W, McNeil C, Shackel N. Ipilimumab-induced toxicities and the gastroenterologist. *J Gastroenterol Hepatol* 2015; **30**: 657-666 [PMID: 25641691 DOI: 10.1111/jgh.12888]
- 42 **Goodman AL**, Murray CD, Watkins J, Griffiths PD, Webster DP. CMV in the gut: a critical review of CMV detection in the immunocompetent host with colitis. *Eur J Clin Microbiol Infect Dis* 2015; **34**: 13-18 [PMID: 25097085 DOI: 10.1007/s10096-014-2212-x]
- 43 **Geukes Foppen MH**, Rozeman EA, van Wilpe S, Postma C, Snaebjornsson P, van Thienen JV, van Leerdam ME, van den Heuvel M, Blank CU, van Dieren J, Haanen JBAG. Immune checkpoint inhibition-related colitis: symptoms, endoscopic features, histology and response to management. *ESMO Open* 2018; **3**: e000278 [PMID: 29387476 DOI: 10.1136/esmoopen-2017-000278]
- 44 **Hsieh AH**, Ferman M, Brown MP, Andrews JM. Vedolizumab: a novel treatment for ipilimumab-induced colitis. *BMJ Case Rep* 2016; **2016** [PMID: 27539137 DOI: 10.1136/bcr-2016-216641]
- 45 **Bergqvist V**, Hertervig E, Gedeon P, Kopljar M, Griph H, Kinhult S, Carneiro A, Marsal J. Vedolizumab treatment for immune checkpoint inhibitor-induced enterocolitis. *Cancer Immunol Immunother* 2017; **66**: 581-592 [PMID: 28204866 DOI: 10.1007/s00262-017-1962-6]
- 46 **Di Giacomo AM**, Biagioli M, Maio M. The emerging toxicity profiles of anti-CTLA-4 antibodies across clinical indications. *Semin Oncol* 2010; **37**: 499-507 [PMID: 21074065 DOI: 10.1053/j.seminoncol.2010.09.007]
- 47 **Weber JS**, Kähler KC, Hauschild A. Management of immune-related adverse events and kinetics of response with ipilimumab. *J Clin Oncol* 2012; **30**: 2691-2697 [PMID: 22614989 DOI: 10.1200/JCO.2012.41.6750]
- 48 **Suriawinata AA**, Thung SN. Acute and chronic hepatitis. *Semin Diagn Pathol* 2006; **23**: 132-148 [PMID: 17355087]
- 49 **Kim KW**, Ramaiya NH, Krajewski KM, Jagannathan JP, Tirumani SH, Srivastava A, Ibrahim N. Ipilimumab associated hepatitis: imaging and clinicopathologic findings. *Invest New Drugs* 2013; **31**: 1071-1077 [PMID: 23408334 DOI: 10.1007/s10637-013-9939-6]
- 50 **Suzuki A**, Brunt EM, Kleiner DE, Miquel R, Smyrk TC, Andrade RJ, Lucena MI, Castiella A, Lindor K, Björnsson E. The use of liver biopsy evaluation in discrimination of idiopathic autoimmune hepatitis versus drug-induced liver injury. *Hepatology* 2011; **54**: 931-939 [PMID: 21674554 DOI: 10.1002/hep.24481]
- 51 **Kumar V**, Chaudhary N, Garg M, Floudas CS, Soni P, Chandra AB. Corrigendum: Current Diagnosis and Management of Immune Related Adverse Events (irAEs) Induced by Immune Checkpoint Inhibitor Therapy. *Front Pharmacol* 2017; **8**: 311 [PMID: 28579959 DOI: 10.3389/fphar.2017.00311]
- 52 **Hofmann L**, Forschner A, Loquai C, Goldinger SM, Zimmer L, Ugurel S, Schmidgen MI, Gutzmer R, Utikal JS, Göpner D, Hassel JC, Meier F, Tietze JK, Thomas I, Weishaupt C, Leverkus M, Wahl R, Dietrich U, Garbe C, Kirchner MC, Eigentler T, Berking C, Gesierich A, Krackhardt AM, Schadendorf D, Schuler G, Dummer R, Heinzerling LM. Cutaneous, gastrointestinal, hepatic, endocrine, and renal side-effects of anti-PD-1 therapy. *Eur J Cancer* 2016; **60**: 190-209 [PMID: 27085692 DOI: 10.1016/j.ejca.2016.02.025]
- 53 **Postow MA**. Managing immune checkpoint-blocking antibody side effects. *Am Soc Clin Oncol Educ Book* 2015; 76-83 [PMID: 25993145 DOI: 10.14694/EdBook_AM.2015.35.76]

P- Reviewer: Aykan NF, Caboclo JF, Contini S, Lin JM, Linnebacher M, Tang Y, Wakao H

S- Editor: Wang XJ **L- Editor:** Filipodia **E- Editor:** Yin SY





Published by **Baishideng Publishing Group Inc**
7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA
Telephone: +1-925-223-8242
Fax: +1-925-223-8243
E-mail: bpgoffice@wjgnet.com
Help Desk: <https://www.f6publishing.com/helpdesk>
<https://www.wjgnet.com>



ISSN 1007-9327

