



NIH PUBLIC ACCESS

Author Manuscript

J Flow Chem. Author manuscript; available in PMC 2012 August 1.

Published in final edited form as:

J Flow Chem. 2011 August ; 1(1): 32–39. doi:10.1556/jfchem.2011.00008.

Accessing Stereochemically Rich Sultams via Microwave-Assisted, Continuous Flow Organic Synthesis (MACOS) Scale-out

Michael G. Organ^{*,[a],[c]}, Paul R. Hanson^{*,[b],[c]}, Alan Rolfe^{[b],[c]}, Thiwanka B. Samarakoon^{[b],[c]}, and Farman Ullah^{[a],[c]}

^[a]Department of Chemistry, York University, 4700 Keele Street, Toronto, ON, M3J 1P3, Canada

^[b]Department of Chemistry, University of Kansas, 1251 Wescoe Hall Drive, Lawrence, KS, 66045-7582, USA

^[c]The University of Kansas Center for Chemical Methodologies and Library Development (KU-CMLD), 2034 Becker Drive, Del Shankel Structural Biology Center, Lawrence, KS, 66047-3761, USA

Abstract

The generation of stereochemically-rich benzothioxazepine-1,1'-dioxides for enrichment of high-throughput screening collections is reported. Utilizing a microwave-assisted, continuous flow organic synthesis platform (MACOS), scale-out of core benzothioxazepine-1,1'-dioxide scaffolds has been achieved on multi-gram scale using an epoxide opening/ S_NAr cyclization protocol. Diversification of these sultam scaffolds was attained via a microwave-assisted intermolecular S_NAr reaction with a variety of amines. Overall, a facile, 2-step protocol generated a collection of benzothioxazepine-1,1'-dioxides possessing stereochemical complexity in rapid fashion, where all 8 stereoisomers were accessed from commercially available starting materials.

Keywords

MACOS; Sultam; S_NAr ; DOS; HTS

1. Introduction

The design, synthesis and production of molecular libraries for high-throughput biological screening (HTS) has been a critical aspect of drug discovery.ⁱ A recent survey of current compound collections for HTS has found that on average, they possess a higher sp^2 content as well as lower structural and stereochemical complexity when compared to currently available drugs.ⁱⁱ While enrichment of compound collections with natural products and their derivatives is one way to enhance complexity, the aforementioned dichotomy has warranted the development of efficient synthetic methods of increasing stereochemical complexity and sp^3 content in screening collections.^{iii,iv} We herein report a facile, 2-step protocol to generate a collection of benzothioxazepine-1,1'-dioxides possessing stereochemical complexity in rapid fashion. A microwave-assisted, continuous flow organic synthesis

ⁱM. Organ Tel: 416-736-5313. Facsimile: 416-736-5936, organ@yorku.ca (Mike Organ).

^{*}Authors listed alphabetically.

Supporting Information Available: Supplementary data (1H and ^{13}C spectra) associated with this article can be found in the online version at: doi:xxxxx.

(MACOS) platform is employed to produce all 8 stereoisomers of a benzothioxazepine-1,1-dioxide core scaffold via an epoxide opening/ S_NAr cyclization^v protocol, followed by intermolecular S_NAr diversification.

Diversity-oriented synthesis (DOS) has emerged in recent years as a powerful tool to enrich the diversity of molecules for HTS.^{vi} This approach has been led by the development of new methodologies, automated technologies, and collaborations within the area of chemical biology. In addition to structural diversity and complexity, the ability to rapidly access every potential stereoisomer of a structure, while functionalizing a core scaffold at every position, is a key concept at the heart of DOS that has the potential to benefit both early stage HTS and SAR, as well as downstream probe/drug development.^{iv,vi,vii}

It was envisioned that a collection of stereochemically-rich benzofused sultams could be rapidly generated where every stereoisomer is readily accessible from commercially available starting materials. In this regard, utilizing a combination of chiral 2° sulfonamides, epoxides, and amino alcohols, a two-step procedure was designed for the synthesis of core benzothioxazepine-1,1-dioxides via an epoxide opening/ S_NAr cyclization^v sequence, followed by intermolecular S_NAr diversification (Figure 1). Furthermore, the powerful combination of microwave heating and flow chemistry was seen to achieve this goal quickly through shortened reaction times and production at any scale necessary with no process re-optimization.

2. Results and Discussion

Preliminary investigations focused on addressing the ability to generate multi-gram quantities of core benzothioxazepine-1,1-dioxide scaffolds. Initial efforts in batch microwave were limited by reactor size and decreased yields resulting from increased by-product formation during scale-up.^{viii} To address these limitations, the MACOS platform was employed to scale-out, rather than scale-up, the scaffold production runs.^{ix,x} The scale-out approach to microwave transformations has been shown in recent years to be a highly efficient method for accessing large quantities of organic molecules.^{xi,xii}

At the outset, a batch microwave reactor was used to develop reaction times that would be suitable for adaptation to flow (Table 1). With some optimization, it was determined that benzothioxazepine-1,1-dioxides **2** could be generated in 70% yield (100% conversion based on **1**) utilizing 1.5 equiv. of ^tBuOK at 180 °C for 1 minute in the microwave (Table 1, entry 7). Such conditions are well tolerated in the MACOS reactor system where flow rates can be set to establish average residence times of approximately one minute for material in the irradiation zone of the microwave (where the sample is actually being irradiated and thus heated).

Taking these preliminary conditions into the MACOS platform, a variety of reactor parameters were investigated, namely flow rate, temperature and power to fine-tune the scale-out protocol (Table 2). When shorter production runs were performed (eg., to produce ~200 mg of **2**), the desired sultam was generated in 50% yield (100% conversion of **1**) utilizing a flow rate of 150 μ L/min at 190 °C and 220 Watts of power (Table 2, entry 7).^{xiii}

When the MACOS reaction was performed for longer times to increase product output, clogging of reaction capillaries was observed after approximately 3 mL of the reaction solution had been flowed. When isolated, this material was not soluble in an array of solvents, indicating that product decomposition under these conditions may have occurred. Suspecting that this problem might be related to the base, ^tBuOK was substituted with DBU. Re-optimization of reaction conditions in batch utilizing DBU (Table 1, entries 8–11) was carried out and the best conditions for the generation of **3** were attempted using the MACOS

platform leading to a gratifying 78% yield (Table 2, Entry 8). Utilizing 1.5 equiv of DBU at 180 °C with a flow rate of 100–200 μ L/min, benzothiazepine-1,1'-dioxides **2–5** were synthesised in gram quantities demonstrating the successful application of the MACOS platform for scale-out (Scheme 1). Building on these trial runs, benzothiazepine-1,1'-dioxides core scaffolds **6 – 13**, were prepared readily using optically-pure epoxides on the MACOS platform (Scheme 1).^{xiv}

With these benzothiazepine-1,1'-dioxide core scaffolds in hand, diversification via microwave-assisted S_NAr was investigated.^{xv} Utilizing the Anton Parr Synthos 3000® microwave platform, a collection of eight optically pure, stereoisomeric sultams (**14 – 21**) was produced demonstrating facile access to stereochemically-rich small-molecule collections for HTS (Scheme 2). Reactions were carried out on 100 mg scale in DMSO at 180 °C for 50 minutes, followed by dilution, filtration, and purification by column chromatography to produce the desired benzothiazepine-1,1'-dioxides **14 – 21** in high yield.

3. Conclusions

In conclusion, a method for the rapid access of stereochemically-rich benzothiazepine-1,1'-dioxides utilizing a two-step, microwave-accelerated protocol is reported. A MACOS platform was used for the scale-out production of core benzothiazepine-1,1'-dioxide scaffolds on multi-gram scale using a cascade epoxide opening/cyclization sequence. These core scaffolds were subsequently diversified by a microwave-assisted, intermolecular S_NAr reaction with chiral, non-racemic amino alcohols. Overall, this 2-step protocol efficiently generated all 8 stereoisomers of benzothiazepine-1,1'-dioxides in optically-pure form, bearing three stereogenic centers, using readily accessible starting materials. These compounds will be evaluated by HTS for potential biological activity by our collaborators, and these results will be reported in due course.

4. Experimental

Microwave irradiation experiments and Analysis of Compounds **2 – 13**

All MACOS experiments were performed in 1700 μ m (ID) borosilicate capillaries, using a single mode Biotage Smith Creator Synthesizer, operating at a frequency of 2.45 GHz with irradiation power from 0 to 350 W. The capillary was fed reactants from Hamilton gastight syringes attached to a Harvard 22 syringe pump preset to the desired flow rate. The system was connected to a sealed collection vial, where a pressurized air line (75 psi) was attached to create backpressure. The temperatures reported were measured off the surface of the capillaries by the IR sensor built into the microwave chamber. All reagents and solvents were purchased from commercial sources and used without additional purification. Column chromatography purifications were carried out using the flash technique on silica gel 60 (200–400 mesh). ¹H and ¹³C NMR spectroscopy was performed on a Bruker Avance 400 MHz instrument (at 400 MHz and 100 MHz respectively) or a Bruker DRX-500 spectrometer (at 500 MHz or 125 MHz respectively). All ¹H NMR spectra were calibrated to the signal from the residual proton of the deuterated chloroform solvent (7.26 ppm) while ¹³C NMR spectra were calibrated to the middle carbon signal of the triplet for deuterated chloroform (77.00 ppm). High-resolution mass spectrometry (HRMS) was recorded on a LCT Premier Spectrometer (Micromass UK Limited) operating the the ESI mode (MeOH). All compounds in this study have been isolated by silica gel/aluminum oxide chromatography for the purpose of spectroscopic identification

General Methods for compounds 14 - 21

All reactions were carried out in 1 dram vials using a reaction heating block in an Anton Paar © Synthos 3000 synthesizer. Parallel evaporations were performed using a GeneVac EZ-2 Plus evaporator. All reagents and solvents were purchased from commercial sources and used without additional purification. Flash column chromatography was performed on silica gel (30930M-25, Silica Gel 60A, 40-63 μ m) and thin layer chromatography was performed on silica gel 60F254 plates (EM-5717, Merck). Deuterated solvents were purchased from Cambridge Isotope laboratories. ^1H and ^{13}C NMR spectra were recorded in CDCl_3 (unless otherwise mentioned) on a Bruker DRX-500 spectrometer operating at 500 MHz, and 125 MHz, respectively, and calibrated to the solvent peak. High-resolution mass spectrometry (HRMS) was recorded on a LCT Premier Spectrometer (Micromass UK Limited) operating in the ESI mode (MeOH). Optical rotations at 589 nm, were measured using AUTOPOL IV Model automatic polarimeter. IR spectra were recorded on a Shimadzu FTIR-8400S instrument.

General procedure A: Scale-out synthesis of benzothiazepine-1,1-dioxide derivatives 2–13 utilizing MACOS flow-platform

A stock solution containing the sulfonamide (1.0 equiv.), epoxide (3 equiv.) and DBU (1.5 equiv.) in DMSO, (0.3-0.4 M) was prepared and loaded into a 10 mL Hamilton gastight syringe. The tubing was primed with DMSO and the syringe was connected to the reactor system with the aid of Microtight™ fittings. A sealed collection vial was connected to the system, where a pressurized airline (75 psi) was attached to create backpressure. A Harvard 22 syringe pump was set to deliver the reaction solution at a rate of 100-200 μ L/min (see Table 1 and 2 for specific conditions). The single mode microwave was programmed to heat constantly with the power level controlled manually so as to keep the temperature constant at the specified levels (see Scheme 1 and Table 2 for specific conditions). The effluent from the reactor was fed into a sealed vial and analyzed directly by ^1H NMR spectroscopy immediately after the reaction. The crude reaction mixture was collected and the product was purified by neutral aluminum oxide column chromatography.

7-Bromo-2-butyl-4-ethyl-1,2- benzoxathiazepine-1,1-dioxide (2)

Utilizing General Procedure A, sultam **2** was isolated in 60% yield (2.8 g, 7.76 mmol) as a yellow solid. **FTIR (neat)**: 2960, 2933, 1573, 1460, 1342, 1164, 1058 cm^{-1} ; **^1H NMR (300 MHz CDCl_3)** δ 7.70 (d, J = 8.1 Hz, 1H), 7.37-7.28 (m, 2H), 4.01-3.82 (m, 2H), 3.29-3.17 (m, 2H), 2.85-2.74 (m, 1H), 1.83-1.71 (m, 1H), 1.67-1.53(m, 3H), 1.42-1.27 (m, 2H), 1.17 (t, J = 7.2 Hz, 3H), 0.95 (t, J = 7.2 Hz, 3H) ppm; **^{13}C NMR (75 MHz CDCl_3)** δ 156.1, 133.5, 130.0, 127.4, 127.2, 123.8 80.9, 53.2, 48.2, 30.8, 26.4, 19.6, 13.7, 10.2 ppm; **DEPT ^{13}C NMR (75 MHz CDCl_3)** δ 130.1, 127.3, 126.5, 80.9, 53.2, 48.2, 30.8, 26.4, 19.6, 13.7, 10.2 ppm; **HRMS** calculated for $\text{C}_{14}\text{H}_{21}\text{BrNO}_3\text{S}$ ($\text{M}+\text{H}$) $^+$ 362.0426; found 362.0429 (TOF MS ES+).

4-(Butoxymethyl)-7-bromo-2-butyl-1,2-benzoxathiazepine-1,1-dioxide (3)

Utilizing General Procedure A, sultam **3** was isolated in 64% yield (3.45 g, 8.234 mmol) as a colorless viscous oil. **FTIR (neat)**: 2956, 2871, 1577, 1456, 1344, 1164, 1058, 931 cm^{-1} ; **^1H NMR (300 MHz CDCl_3)** δ 7.69 (d, J = 9.0 Hz, 1H), 7.37 (m, 2H), 4.22-4.15 (m, 1H), 3.96 (dd, J = 15, 10.5 Hz, 1H), 3.74-3.69 (m, 1H), 3.59-3.46 (m, 3H), 3.43 (d, J = 15 Hz, 1H), 3.26-3.16 (m, 1H), 2.82-2.61 (m, 1H), 1.61-1.52 (m, 4H), 1.51-1.72 (m, 4H), 0.98-0.83 (m, 6H) ppm; **^{13}C NMR (75 MHz CDCl_3)** δ 155.5, 133.34, 130.1, 127.4, 126.6, 77.8, 71.7, 70.6, 50.5, 48.0, 31.6, 30.7, 19.6, 19.3, 13.8, 13.6 ppm; **DEPT ^{13}C NMR (75 MHz CDCl_3)** δ 130.1, 127.5, 126.7, 77.8, 71.7, 70.6, 50.5, 48.0, 31.6, 30.7, 19.6, 19.3, 13.9,

13.6 ppm; **HRMS** calculated for $C_{17}H_{27}BrNO_4S$ (M+H)⁺ 420.0844; found 420.0842 (TOF MS ES+).

2-Butyl-4-ethyl-7-fluoro-1,2-benzoxathiazepine-1,1-dioxide (4)

Utilizing General Procedure A, sultam **4** was isolated in 54% yield (2.6 g, 8.64 mmol) as a colorless viscous oil. **FTIR** (neat): 2962, 2873, 1598, 1583, 1342, 1141, 1070, 995 cm^{-1} ; **¹H NMR** (300 MHz $CDCl_3$) δ 7.83 (dd, $J = 8.7, 6.3$ Hz, 1H), 6.94-6.83 (m, 2H), 3.98-3.81 (m, 2H), 3.28-3.18 (m, 2H), 2.81-2.74 (m, 1H), 1.79-1.72 (m, 1H), 1.64-1.54 (m, 3H), 1.38-1.31 (m, 2H), 1.5 (t, $J = 7.2$ Hz, 3H), 0.94 (t, $J = 7.2$ Hz, 3H); **¹³C NMR** (75 MHz $CDCl_3$) δ 167. (d, $J_{C-F} = 253.5$ Hz), 157.4 (d, $J_{C-F} = 12.0$ Hz), 130.9 (d, $J_{C-F} = 10.5$ Hz), 130.6 (d, $J_{C-F} = 3.7$ Hz), 111.4 (d, $J_{C-F} = 21.5$ Hz), 110.8 (d, $J_{C-F} = 23.2$ Hz), 80.9, 53.1, 48.1, 30.7, 26.3, 19.6, 13.6, 10.1 ppm; **DEPT ¹³C-NMR** (75 MHz $CDCl_3$) δ 130.9 (d, $J_{C-F} = 10.5$ Hz), 111.4 (d, $J_{C-F} = 21.8$ Hz), 110.8 (d, $J_{C-F} = 22.5$ Hz), 80.9, 53.1, 48.2, 30.8, 26.3, 19.6, 13.6, 10.2 ppm; **CHN analysis** calculated for $C_{14}H_{20}FNO_3S$: C, 55.79; H, 6.69; N, 4.65; S, 10.65; found C, 56.01; H, 6.42; N, 4.53; S, 10.72.

4-(Butoxymethyl)-2-butyl-7-fluoro-1,2-benzoxathiazepine-1,1-dioxide (5)

Utilizing General Procedure A, sultam **5** was isolated in 52% yield (3.0 g, 8.36 mmol) as a colorless viscous oil; **FTIR** (neat): 2958, 2933, 2871, 1598, 1585, 1423, 1344, 1118, 1070, 989 cm^{-1} ; **¹H NMR** (300 MHz $CDCl_3$) δ 7.82 (dd, $J = 8.7, 6.3$ Hz, 1H), 6.93 - 6.84 (m, 2H), 4.21 (m, 1H), 3.95 (d, $J = 15, 10.5$ Hz, 1H), 3.73 - 3.67 (m, 1H), 3.57 - 3.48 (m, 3H), 3.62 - 3.42 (dd, $J = 15, 1.3$ Hz, 1H), 3.17 (m, 1H), 2.77 - 2.72 (m, 1H), 1.59 - 1.52 (m, 4H), 1.41-1.32 (m, 4H), 0.94 - 0.87 (m, 6H), ppm; **¹³C NMR** (75 MHz $CDCl_3$) δ 167.1 (d, $J_{C-F} = 253.5$ Hz), 156.9 (d, $J_{C-F} = 12.0$ Hz), 130.9 (d, $J_{C-F} = 10.5$ Hz), 130.5 (d, $J_{C-F} = 3.0$ Hz), 111.6 (d, $J_{C-F} = 21.8$ Hz), 111.1 (d, $J_{C-F} = 23.3$ Hz), 77.7, 71.6, 70.6, 50.4, 47.9, 31.6, 30.6, 19.6, 19.2, 13.8, 13.6 ppm; **DEPT ¹³C NMR** (75 MHz $CDCl_3$) δ 130.9 (d, $J_{C-F} = 10.5$ Hz), 111.6 (d, $J_{C-F} = 22.5$ Hz), 111.1 (d, $J_{C-F} = 22.5$ Hz), 77.7, 71.6, 70.6, 50.4, 47.9, 31.6, 30.7, 19.6, 19.2, 13.9, 13.6 ppm; **CHN analysis** calculated for $C_{17}H_{26}FNO_4S$: C, 56.80; H, 7.29; N, 3.90; S, 8.92; found C, 57.18; H, 7.43; N, 3.96; S, 8.85.

((R)-4-(Benzyloxymethyl)-7-fluoro-2-((R)-1-pentylethyl)-1,2-benzoxathiazepine-1,1-dioxide (6)

Utilizing general procedure A, sultam **6** was isolated in 50% yield (3.01 g, 6.92 mmol) as a colorless viscous oil; $[\alpha]_D^{20} = +6.4^\circ$ ($c = 2.7$, $CHCl_3$); **FTIR** (neat): 2929, 2860, 1598, 1471, 1340, 1157, 1116, 1072 cm^{-1} ; **¹H NMR** (400 MHz $CDCl_3$) δ 7.84 (dd, $J = 6.4, 8.4$ Hz, 1H), 7.42 - 7.35 (m, 5H), 6.92 - 6.85 (m, 2H), 4.65 (s, 2H), 4.33 - 4.31 (m, 1H), 4.29 - 4.14 (m, 1H), 3.84 - 3.77 (m, 2H), 3.67-3.66 (m, 1H), 3.47 (dd, $J = 14.0, 1.7$ Hz, 1H), 1.60 (br s, 1H), 1.50 (m, 1H), 1.29 (br s, 6H), 1.0 (d, $J = 6.8$ Hz, 3H), 0.89 (t, $J = 6.8$ Hz, 3H); **¹³C NMR** (100 MHz $CDCl_3$) δ 166.2 (d, $J_{C-F} = 253.0$ Hz), 156.6 (d, $J_{C-F} = 12$ Hz), 137.4, 133.3 (d, $J_{C-F} = 3$ Hz), 128.8 (d, $J = 11$ Hz), 128.4, 127.9, 127.6, 111.1 (d, $J_{C-F} = 22$ Hz), 110.4 (d, $J_{C-F} = 23$ Hz), 80.7, 73.6, 70.1, 54.4, 44.8, 34.5, 31.2, 25.7, 22.4, 19.8, 13.9 ppm; **HRMS** calculated for $C_{23}H_{31}FNO_4S$ (M+H)⁺ 436.1958; found 436.1959 (TOF MS ES+).

((S)-4-(Benzyloxymethyl)-7-fluoro-2-((R)-1-pentylethyl)-1,2-benzoxathiazepine-1,1-dioxide (7)

Utilizing General Procedure A, sultam **7** was isolated in 51% yield (3.05 g, 7.01 mmol) as a colorless viscous oil; $[\alpha]_D^{20} = -10.3^\circ$ ($c = 4.7$, $CHCl_3$); **FTIR** (neat): 2929, 2861, 1561, 1470, 1340, 1156, 1116, 1073 cm^{-1} ; **¹H NMR** (300 MHz $CDCl_3$) δ 7.83 (dd, $J = 8.7, 6.3$ Hz, 1H), 7.41 - 7.34 (m, 5H), 6.89-6.79 (m, 2H), 4.77 - 4.73 (m, 1H), 4.66 (d, $J = 3.6$ Hz, 2H), 4.10 (m, 1H), 3.78 - 3.59 (m, 3H), 3.45 (dd, $J = 14.1, 3.0$ Hz, 1H), 1.43 - 1.26 (m, 8H),

1.10 (d, $J = 6.6$ Hz, 3H), 0.89 (t, $J = 6.6$ Hz, 3H) ppm; ^{13}C NMR (100 MHz CDCl_3) δ 166.0 (d, $J_{\text{C-F}} = 252.0$ Hz), 156.5 (d, $J_{\text{C-F}} = 13$ Hz), 137.5, 131.7, 129.1 (d, $J_{\text{C-F}} = 11$ Hz, 1H), 128.4, 127.9, 127.5, 110.6 (d, $J_{\text{C-F}} = 22$ Hz), 109.3 (d, $J_{\text{C-F}} = 24$ Hz), 80.6, 73.5, 69.9, 54.9, 44.6, 35.3, 31.2, 26.1, 22.4, 18.6, 13.9 ppm; **CHN analysis** calculated for $\text{C}_{23}\text{H}_{30}\text{FNO}_4\text{S}$: C, 63.42; H, 6.94; N, 3.22; S, 7.36; found C, 63.74; H, 6.80; N, 3.08; S, 7.18.

((R)-4-(Benzyloxymethyl)-7-fluoro-2-((S)-1-pentylethyl)-1,2-benzoxathiazepine-1,1-dioxide (8)

Utilizing General Procedure A, sultam **8** was isolated in 53% yield (3.15 g, 7.24 mmol) as a colorless viscous oil; $[\alpha]_{\text{D}}^{20} = +7.6^\circ$ ($c = 2.2$, CHCl_3); **FTIR (neat)**: 2929, 2858, 1600, 1473, 1336, 1157, 1116, 1076 cm^{-1} ; ^1H NMR (300 MHz CDCl_3) δ 7.83 (dd, $J = 8.7$, 6.3 Hz, 1H), 7.40 - 7.34 (m, 5H), 6.89 - 6.79 (m, 2H), 4.77 - 4.70 (m, 1H), 4.66 (d, $J = 3.6$ Hz, 2H), 4.10 (m, 1H), 3.78 - 3.59 (m, 3H), 3.45 (dd, $J = 13.8$, 2.8 Hz, 1H), 1.43 - 1.26 (m, 8H), 1.10 (d, $J = 6.6$ Hz, 3H), 0.89 (t, $J = 6.6$ Hz, 3H) ppm; ^{13}C NMR (75 MHz CDCl_3) δ 166.5 (d, $J_{\text{C-F}} = 252.0$ Hz), 156.5 (d, $J_{\text{C-F}} = 12.8$ Hz), 137.5, 131.8 (d, $J_{\text{C-F}} = 3$ Hz), 129.2 (d, $J_{\text{C-F}} = 10.5$ Hz), 128.5, 128.3, 127.9, 127.6, 110.8 (d, $J_{\text{C-F}} = 22.5$ Hz), 109.4 (d, $J_{\text{C-F}} = 24$ Hz), 80.7, 73.6, 69.9, 54.9, 44.6, 35.3, 31.3, 26.1, 22.5, 18.6, 14.0 ppm; **DEPT ^{13}C -NMR (75 MHz CDCl_3)** δ 129.3 (d, $J_{\text{C-F}} = 10.5$ Hz), 128.6, 128.0, 127.7, 110.8, (d, $J_{\text{C-F}} = 21.8$ Hz), 109.4 (d, $J_{\text{C-F}} = 23.33$ Hz), 80.7, 73.6, 70.0, 55.0, 44.7, 35.4, 31.3, 26.2, 22.5, 18.7, 14.0 ppm; **CHN analysis** calculated for $\text{C}_{23}\text{H}_{30}\text{FNO}_4\text{S}$: C, 63.42; H, 6.94; N, 3.22; S, 7.36; found C, 63.84; H, 6.92; N, 3.14; S, 7.14.

((S)-4-(Benzyloxymethyl)-7-fluoro-2-((S)-1-pentylethyl)-1,2-benzoxathiazepine-1,1-dioxide (9)

Utilizing General Procedure A, sultam **9** was isolated in 53% yield (3.15 g, 7.24 mmol) as a colorless viscous oil; $[\alpha]_{\text{D}}^{20} = +26.2^\circ$ ($c = 12.3$, CHCl_3); **FTIR (neat)**: 2929, 2858, 1596, 1475, 1341, 1157, 1115, 1072 cm^{-1} ; ^1H NMR (400 MHz CDCl_3) δ 7.84 (dd, $J = 6.4$, 8.4 Hz, 1H), 7.42 - 7.34 (m, 5H), 6.92 - 6.85 (m, 2H), 4.65 (s, 2H), 4.33 - 4.31 (m, 1H), 4.17 - 4.15 (m, 1H), 3.85 - 3.77 (m, 2H), 3.68 (dd, $J = 10.4$, 5.6 Hz, 1H), 3.47 (dd, $J = 13.6$, 1.7 Hz, 1H), 1.50 - 1.28 (m, 8H), 1.01 (d, $J = 6.8$ Hz, 3H), 0.89 (t, $J = 6.8$ Hz, 3H) ppm; ^{13}C NMR (100 MHz CDCl_3) δ 166.2 (d, $J_{\text{C-F}} = 253.0$ Hz), 156.6 (d, $J_{\text{C-F}} = 12.0$ Hz), 137.5, 133.3 (d, $J_{\text{C-F}} = 2.0$ Hz), 128.8 (d, $J_{\text{C-F}} = 11.0$ Hz), 128.4, 127.9, 127.6, 111.1 (d, $J_{\text{C-F}} = 22.0$ Hz), 110.4 (d, $J_{\text{C-F}} = 23.0$ Hz), 80.7, 73.6, 70.1, 54.4, 44.8, 34.5, 31.2, 25.7, 22.4, 19.8, 1.9 ppm; **DEPT ^{13}C -NMR (75 MHz CDCl_3)** δ 128.8 (d, $J_{\text{C-F}} = 10.5$ Hz), 128.6, 127.9, 127.8, 111.1 (d, $J_{\text{C-F}} = 21.8$ Hz), 110.4 (d, $J_{\text{C-F}} = 24.0$ Hz), 80.8, 73.7, 70.2, 54.4, 44.9, 34.6, 31.2, 25.8, 22.5, 19.9, 13.9 ppm; **CHN analysis** calculated for $\text{C}_{23}\text{H}_{30}\text{FNO}_4\text{S}$: C, 63.42; H, 6.94; N, 3.22; S, 7.36; found C, 63.86; H, 6.81; N, 3.19; S, 7.26.

((R)-4-(Benzyloxymethyl)-2-((S)-1-phenylethyl)-7-fluoro-1,2-benzoxathiazepine-1,1-dioxide (10)

Utilizing General Procedure A, sultam **10** was isolated in 48% yield (2.85 g, 6.463 mmol) as a colorless viscous oil; $[\alpha]_{\text{D}}^{20} = +5.8^\circ$ ($c = 3.7$, CHCl_3); **FTIR (neat)**: 3029, 2937, 2866, 1598, 1585, 1471, 1338, 1174, 1153, 1070 cm^{-1} ; ^1H NMR (300 MHz CDCl_3) δ 7.93 (dd, $J = 8.7$, 6.3 Hz, 1H), 7.43 - 7.31 (m, 10H), 6.98 - 6.87 (m, 2H), 5.30 - 5.46 (m, 1H), 4.57 (s, 2H), 4.38 - 4.44 (m, 1H), 3.78 - 3.70 (m, 2H), 3.69 - 3.51 (m, 1H), 3.37 (dd, $J = 15.0$, 1.8 Hz, 1H), 1.46 (d, $J = 6.3$ Hz, 3H) ppm; ^{13}C NMR (75 MHz CDCl_3) δ 166.9 (d, $J_{\text{C-F}} = 252.8$ Hz), 156.8 (d, $J_{\text{C-F}} = 12.0$ Hz), 139.4, 137.6, 133.3 (d, $J_{\text{C-F}} = 3.0$ Hz), 129.1 (d, $J_{\text{C-F}} = 10.5$ Hz), 128.6, 128.5, 128.4, 127.9, 127.8, 127.7, 127.6, 127.5, 111.4 (d, $J_{\text{C-F}} = 21.8$ Hz), 110.9 (d, $J_{\text{C-F}} = 23.3$ Hz), 81.1, 73.5, 70.1, 56.6, 46.1, 17.7 ppm; **DEPT ^{13}C -NMR (75 MHz CDCl_3)** δ 129.1 (d, $J_{\text{C-F}} = 10.5$ Hz), 128.6, 128.5, 128.0, 127.9, 127.6, 127.5, 111.5 (d, $J_{\text{C-F}} = 21.8$ Hz), 111.0 (d, $J_{\text{C-F}} = 24.0$ Hz), 81.1, 73.5, 70.2, 56.6, 46.1, 17.7 ppm; **CHN**

analysis calculated for C₂₄H₂₄FNO₄S: C, 65.29; H, 5.48; N, 3.17; S, 7.27: found C, 65.60; H, 5.38; N, 3.05; S, 7.21.

(S)-4-(Benzyloxymethyl)-2-((R)-1-phenylethyl)-7-fluoro-1,2-benzoxathiazepine-1,1-dioxide (11)

Utilizing General Procedure **A**, sultam **11** was isolated in 52% yield (3.10 g, 7.03 mmol) as a colorless viscous oil; $[\alpha]_D^{20} = +39.2^\circ$ ($c = 6$, CHCl₃); **FTIR (neat)**: 3020, 2939, 2866, 1596, 1584, 1474, 1338, 1171, 1153, 1070 cm⁻¹; **¹H NMR (300 MHz CDCl₃)** δ 7.94 (dd, $J = 8.7, 6.3$ Hz, 1H), 7.41 - 7.15 (m, 10H), 6.99 - 6.93 (m, 1H), 6.79 (dd, $J = 9.6, 2.4$ Hz, 1H), 5.41 (d, $J = 2.1$ Hz, 1H), 4.50 (d, $J = 2.1$ Hz, 2H), 3.72 (m, 2H), 3.57 - 3.47 (m, 2H), 3.39 (d, $J = 12.6$, Hz, 1H), 1.60 (d, $J = 6.9$ Hz, 3H) ppm; **¹³C NMR (75 MHz CDCl₃)** δ 166.8(d, $J_{C-F} = 253.5$ Hz), 156.7 (d, $J_{C-F} = 12.0$ Hz), 139.0, 137.6, 132.7 (d, $J_{C-F} = 3.0$ Hz), 129.3 (d, $J_{C-F} = 10.5$ Hz), 128.6, 128.4, 128.2, 127.8, 127.6, 127.4, 111.2 (d, $J_{C-F} = 22.5$ Hz), 110.4 (d, $J_{C-F} = 23.3$ Hz), 79.4, 73.2, 69.9, 56.6, 45.7, 17.5 ppm; **DEPT ¹³C NMR (75 MHz CDCl₃)** δ 129.3 (d, $J_{C-F} = 10.5$ Hz), 128.6, 128.4, 128.2, 127.8, 127.6, 127.4, 111.2 (d, $J_{C-F} = 21.8$ Hz), 110.5 (d, $J_{C-F} = 24.0$ Hz), 79.5, 73.3, 69.9, 56.6, 45.7, 17.5 ppm; **HRMS** calculated for C₂₄H₂₅FNO₄S (M+H)⁺ 442.1488; found 442.1490 (TOF MS ES+).

((R)-4-(Benzyloxymethyl)-2-((S)-1-phenylethyl)-7-fluoro-1,2-benzoxathiazepine-1,1-dioxide (12)

Utilizing General Procedure **A**, sultam **12** was isolated in 48% yield (2.84 g, 6.44 mmol) as a colorless viscous oil; $[\alpha]_D^{20} = -35.0^\circ$ ($c = 4.4$, CHCl₃); **FTIR (neat)**: 3029, 2935, 2866, 1598, 1585, 1473, 1336, 1155, 1116, 1070 cm⁻¹; **¹H NMR (300 MHz CDCl₃)** δ 7.96 (dd, $J = 8.7, 6.3$ Hz, 1H), 7.41 - 7.15 (m, 10H), 6.99 - 6.93 (m, 1H), 6.78 (dd, $J = 9.3, 2.7$ Hz, 1H), 5.42 - 5.38 (m, 1H), 4.57 (d, $J = 2.1$ Hz, 2H) 3.67 (s, 2H), 3.55 - 3.45 (m, 2H), 3.38 (d, $J = 12.6$ Hz, 1H), 1.59 (d, $J = 6.9$ Hz, 3H) ppm; **¹³C NMR (75 MHz CDCl₃)** δ 166.8(d, $J_{C-F} = 252.7$ Hz), 156.7 (d, $J_{C-F} = 12.0$ Hz), 139.0, 137.6, 132.6 (d, $J_{C-F} = 3.0$ Hz), 129.3 (d, $J_{C-F} = 10.5$ Hz), 128.6, 128.5, 128.2, 127.9, 127.6, 127.5, 111.2 (d, $J_{C-F} = 22.5$ Hz), 110.4 (d, $J_{C-F} = 23.3$ Hz), 79.4, 73.2, 69.8, 56.6, 45.7, 17.5 ppm; **CHN analysis** calculated for C₂₄H₂₄FNO₄S: C, 65.29; H, 5.48; N, 3.17; S, 7.27: found C, 65.58; H, 5.80; N, 3.12; S, 7.22.

((S)-4-(Benzyloxymethyl)-2-((S)-1-phenylethyl)-7-fluoro-1,2-benzoxathiazepine-1,1-dioxide (13)

Utilizing general procedure **A**, sultam **13** was isolated in 54% yield (3.20 g, 7.26 mmol) as a colorless viscous oil; $[\alpha]_D^{20} = -43.4^\circ$ ($c = 11.3$, CHCl₃); **FTIR (neat)**: 3028, 2934, 2865, 1597, 1585, 1471, 1339, 1171, 1152, 1070 cm⁻¹; **¹H NMR (300 MHz CDCl₃)** δ 7.92 (m, 1H), 7.42 - 7.30 (m, 10H), 6.98 - 6.85 (m, 2H), 5.52 - 5.45 (m, 1H), 4.57 (s, 2H), 4.38 - 4.33 (m, 1H), 3.77 (m, 2H), 3.56 - 3.51 (m, 1H), 3.40 (dd, $J = 16.8, 1.8$ Hz, 1H), 1.45 (d, $J = 6.9$ Hz, 3H) ppm; **¹³C NMR (75 MHz CDCl₃)** δ 166.9(d, $J_{C-F} = 253.5$ Hz), 156.7 (d, $J_{C-F} = 12$ Hz), 139.4, 137.6, 133.2 (d, $J_{C-F} = 3$ Hz), 129.1 (d, $J_{C-F} = 10.5$ Hz), 128.6, 128.5, 128.0, 127.9, 127.6, 127.5, 111.4 (d, $J_{C-F} = 22.5$ Hz), 110.9 (d, $J_{C-F} = 23.3$ Hz), 81.1, 73.5, 70.1, 56.6, 46.1, 17.7 ppm; **CHN analysis** calculated for C₂₄H₂₄FNO₄S: C, 65.29; H, 5.48; N, 3.17; S, 7.27: found C, 65.63; H, 5.83; N, 3.15; S, 7.26.

General procedure B: Microwave-assisted diversification of benzothiazepine-1,1-dioxides 2 -13 cores

Into a 1 dram vial was added benzothiazepine-1,1-dioxide **2 - 13** (100 mg, 0.23 mmol, 1 equiv.), dry DMSO (0.23 ml, 1M), DBU (3.4 μ L, 10 mol%) and the corresponding amine (5 equiv., 1.15 mmol). The reaction vessel was capped, placed in Anton Paar Synthos 3000 @ microwave and heated at 180 °C for 50 min [Power = 1200 W, 8 minute ramp then 50 min

hold]. After such time, the reaction was diluted in EtOAc, filtered through a SiO₂ SPE and concentrated. The resulting crude product was purified by flash chromatography (7:3, EtOAc:hexane) to afford the desired benzo-oxathiazepine 1,1-dioxide **14** – **21**.

(R)-4-((Benzyloxy)methyl)-7-((S)-2-(hydroxymethyl)pyrrolidin-1-yl)-2-((R)-1-phenylethyl)-3,4-dihydro-2H-benzo[b][1,4,5]oxathiazepine 1,1-dioxide (14)

Utilizing general procedure **B**, sultam **14** was isolated (92%, 110 mg, 0.21 mmol) as a colorless viscous oil; $[\alpha]_D^{20} = -21.4^\circ$ ($c = 2.3$, CHCl₃); **FTIR (neat)**: 3352, 2941, 1596, 1494, 1377, 1326, 1132, 1074 cm⁻¹; **¹H NMR (500 MHz, CDCl₃)** δ 7.66 (d, $J = 8.8$ Hz, 1H), 7.41 (d, $J = 7.3$ Hz, 2H), 7.39 – 7.33 (m, 4H), 7.32 (dd, $J = 8.0, 2.4$ Hz, 2H), 7.32 – 7.27 (m, 2H), 6.44 (dd, $J = 8.8, 2.4$ Hz, 1H), 6.36 (d, $J = 2.3$ Hz, 1H), 5.44 (q, $J = 7.1$ Hz, 1H), 4.56 (q, $J = 11.9$ Hz, 2H), 4.23 – 4.14 (m, 1H), 3.88 (dd, $J = 10.9, 7.1$ Hz, 1H), 3.74 (dd, $J = 10.2, 5.8$ Hz, 1H), 3.72 – 3.62 (m, 2H), 3.54 (dd, $J = 12.5, 5.0$ Hz, 1H), 3.49 (dt, $J = 9.6, 4.8$ Hz, 1H), 3.44 (dd, $J = 12.1, 5.2$ Hz, 1H), 3.21 (ddd, $J = 17.2, 11.7, 5.5$ Hz, 2H), 2.14 – 2.08 (m, 2H), 2.08 – 2.00 (m, 2H), 1.98 (d, $J = 8.4$ Hz, 1H), 1.37 (d, $J = 7.1$ Hz, 3H) ppm; **¹³C NMR (126 MHz, CDCl₃)** δ 156.4, 151.9, 140.2, 137.8, 128.5, 128.4, 127.8, 127.6, 127.5, 127.4, 123.5, 107.1, 105.4, 80.9, 73.4, 70.6, 62.6, 60.3, 56.1, 48.9, 46.3, 28.4, 23.3, 17.4 ppm; **HRMS** calculated for C₂₉H₃₅N₂O₅S (M+H)⁺ 523.2267; found 523.2259 (TOF MS ES+).

(S)-4-((Benzyloxy)methyl)-7-((S)-2-(hydroxymethyl)pyrrolidin-1-yl)-2-((R)-1-phenylethyl)-3,4-dihydro-2H-benzo[b][1,4,5]oxathiazepine 1,1-dioxide (15)

Utilizing general procedure **B**, sultam **15** was isolated (95%, 114 mg, 0.218 mmol) as a colorless viscous oil; $[\alpha]_D^{20} = -3.0^\circ$ ($c = 0.67$, CHCl₃); **FTIR (neat)**: 3352, 2939, 1598, 1496, 1377, 1326, 1130, 1074 cm⁻¹; **¹H NMR (500 MHz, CDCl₃)** δ 7.68 (d, $J = 8.8$ Hz, 1H), 7.36 – 7.31 (m, 2H), 7.31 – 7.27 (m, 1H), 7.25 (m, 2H), 7.20 (ddd, $J = 10.2, 6.9, 3.7$ Hz, 5H), 6.41 (dt, $J = 10.3, 5.1$ Hz, 1H), 6.22 (d, $J = 2.4$ Hz, 1H), 5.34 (q, $J = 7.0$ Hz, 1H), 4.45 (q, $J = 12.1$ Hz, 2H), 3.90 – 3.84 (m, 1H), 3.90 – 3.83 (m, 1H), 3.64 (dd, $J = 27.2, 12.0$ Hz, 2H), 3.53 (ddd, $J = 14.0, 13.2, 7.8$ Hz, 2H), 3.45 (dq, $J = 10.5, 5.3$ Hz, 2H), 3.27 (d, $J = 12.5$ Hz, 1H), 3.22 – 3.14 (m, 1H), 2.08 (dt, $J = 8.4, 4.3$ Hz, 2H), 2.06 – 1.98 (m, 2H), 1.64 (d, $J = 4.2$ Hz, 1H), 1.55 (d, $J = 7.0$ Hz, 3H) ppm; **¹³C NMR (126 MHz, CDCl₃)** δ 156.3, 151.6, 139.8, 137.8, 128.6, 128.4, 127.8, 127.7, 127.7, 127.4, 122.7, 107.0, 104.7, 79.0, 73.1, 70.2, 62.7, 59.9, 56.3, 48.9, 45.9, 28.4, 23.3, 17.7 ppm; **HRMS** calculated for C₂₉H₃₅N₂O₅S (M+H)⁺ 523.2267; found 523.2258 (TOF MS ES+).

(S)-4-((Benzyloxy)methyl)-7-((S)-2-(hydroxymethyl)pyrrolidin-1-yl)-2-((S)-1-phenylethyl)-3,4-dihydro-2H-benzo[b][1,4,5]oxathiazepine 1,1-dioxide (16)

Utilizing general procedure **B**, sultam **16** was isolated (90%, 108 mg, 0.207 mmol) as a colorless viscous oil; $[\alpha]_D^{20} = -27.0^\circ$ ($c = 0.51$, CHCl₃); **FTIR (neat)**: 3353, 2939, 1596, 1496, 1377, 1326, 1130, 1074 cm⁻¹; **¹H NMR (500 MHz, CDCl₃)** δ 7.66 (dd, $J = 8.7, 4.5$ Hz, 1H), 7.38 (d, $J = 7.1$ Hz, 2H), 7.36 – 7.31 (m, 4H), 7.31 – 7.25 (m, 4H), 6.40 (dd, $J = 8.8, 2.4$ Hz, 1H), 6.35 (d, $J = 2.4$ Hz, 1H), 5.42 (q, $J = 7.1$ Hz, 1H), 4.57 – 4.50 (m, 2H), 4.17 (ddd, $J = 9.4, 4.7, 3.3$ Hz, 1H), 3.89 (dt, $J = 11.3, 5.6$ Hz, 1H), 3.73 – 3.60 (m, 3H), 3.60 – 3.54 (m, 1H), 3.47 (dd, $J = 10.1, 6.2$ Hz, 2H), 3.22 – 3.16 (m, 2H), 2.09 (tt, $J = 10.4, 5.0$ Hz, 2H), 2.08 – 2.04 (m, 2H), 1.55 (t, $J = 5.2$ Hz, 1H), 1.36 (d, $J = 7.1$ Hz, 3H); **¹³C NMR (126 MHz, CDCl₃)** δ 156.4, 151.8, 140.1, 137.8, 128.5, 128.4, 127.8, 127.7, 127.6, 127.5, 123.7, 107.1, 105.5, 80.9, 73.4, 70.5, 62.8, 60.0, 56.1, 49.0, 46.3, 28.4, 23.3, 17.4 ppm; **HRMS** calculated for C₂₉H₃₅N₂O₅S (M+H)⁺ 523.2267; found 523.2260 (TOF MS ES+).

(R)-4-((Benzyloxy)methyl)-7-((S)-2-(hydroxymethyl)pyrrolidin-1-yl)-2-((S)-1-phenylethyl)-3,4-dihydro-2H-benzo[b][1,4,5]oxathiazepine 1,1-dioxide (17)

Utilizing general procedure **B**, sultam **17** was isolated (98%, 117 mg, 2.25 mmol) as a colorless viscous oil; $[\alpha]_D^{20} = -36.5^\circ$ ($c = 0.79$, CHCl_3); **FTIR** (neat): 3363, 2933, 1598, 1494, 1377, 1325, 1132, 1076 cm^{-1} ; **^1H NMR** (500 MHz, CDCl_3) δ 7.67 (t, $J = 6.6$ Hz, 1H), 7.36 – 7.32 (m, 2H), 7.31 – 7.27 (m, 1H), 7.24 (m, 2H), 7.23 – 7.20 (m, 3H), 7.20 – 7.16 (m, 2H), 6.42 (dt, $J = 15.3, 7.7$ Hz, 1H), 6.21 (d, $J = 2.4$ Hz, 1H), 5.32 (q, $J = 7.0$ Hz, 1H), 4.44 (q, $J = 12.1$ Hz, 2H), 3.90 – 3.83 (m, 1H), 3.68 – 3.57 (m, 3H), 3.53 (dq, $J = 10.5, 5.1$ Hz, 2H), 3.46 – 3.41 (m, 2H), 3.26 (d, $J = 12.5$ Hz, 1H), 3.20 – 3.13 (m, 1H), 2.12 – 1.96 (m, 4H), 1.64 (s, 1H), 1.55 (d, $J = 7.0$ Hz, 3H) ppm; **^{13}C NMR** (126 MHz, CDCl_3) δ 156.3, 151.6, 139.8, 137.9, 128.6, 128.5, 128.4, 127.8, 127.7, 127.4, 122.8, 107.0, 104.6, 78.9, 73.0, 70.3, 62.7, 60.1, 56.3, 48.9, 45.9, 28.4, 23.3, 17.6 ppm; **HRMS** calculated for $\text{C}_{29}\text{H}_{35}\text{N}_2\text{O}_5\text{S}$ ($\text{M}+\text{H}$)⁺ 523.2267; found 523.2261 (TOF MS ES+).

(R)-4-((Benzyloxy)methyl)-7-((R)-2-(hydroxymethyl)pyrrolidin-1-yl)-2-((R)-1-phenylethyl)-3,4-dihydro-2H-benzo[b][1,4,5]oxathiazepine 1,1-dioxide (18)

Utilizing general procedure **B**, sultam **18** was isolated (93%, 111 mg, 0.214 mmol) as a colorless viscous oil; $[\alpha]_D^{20} = +28.5^\circ$ ($c = 2.4$, CHCl_3); **FTIR** (neat): 3348, 2937, 2871, 1596, 1496, 1377, 1326, 1130, 1074 cm^{-1} ; **^1H NMR** (500 MHz, CDCl_3) δ 7.63 (d, $J = 8.8$ Hz, 1H), 7.38 (d, $J = 7.3$ Hz, 2H), 7.34 (ddd, $J = 7.4, 4.5, 2.0$ Hz, 4H), 7.31 – 7.28 (m, 2H), 7.28 – 7.25 (m, 2H), 6.39 (dt, $J = 10.3, 5.2$ Hz, 1H), 6.35 (d, $J = 2.3$ Hz, 1H), 5.42 (q, $J = 7.1$ Hz, 1H), 4.58 – 4.49 (m, 2H), 4.19 – 4.13 (m, 1H), 3.91 – 3.83 (m, 1H), 3.55 – 3.49 (m, 3H), 3.46 (td, $J = 9.7, 4.2$ Hz, 2H), 3.23 – 3.14 (m, 2H), 2.12 – 1.95 (m, 4H), 1.92 (s, 1H), 1.36 (d, $J = 7.1$ Hz, 3H) ppm; **^{13}C NMR** (126 MHz, CDCl_3) δ 156.4, 151.8, 140.2, 137.8, 128.5, 128.4, 127.8, 127.6, 127.5, 127.4, 123.5, 107.2, 105.5, 80.8, 73.4, 70.5, 62.6, 60.1, 56.1, 48.9, 46.3, 28.4, 23.3, 17.4 ppm; **HRMS** calculated for $\text{C}_{29}\text{H}_{35}\text{N}_2\text{O}_5\text{S}$ ($\text{M}+\text{H}$)⁺ 523.2267; found 523.2257 (TOF MS ES+).

(S)-4-((Benzyloxy)methyl)-7-((R)-2-(hydroxymethyl)pyrrolidin-1-yl)-2-((R)-1-phenylethyl)-3,4-dihydro-2H-benzo[b][1,4,5]oxathiazepine 1,1-dioxide (19)

Utilizing general procedure **B**, sultam **19** was isolated (94%, 112 mg, 0.216 mmol) as a colorless viscous oil; $[\alpha]_D^{20} = +38.6^\circ$ ($c = 1.95$, CHCl_3); **FTIR** (neat): 3369, 2931, 1598, 1494, 1377, 1325, 1132, 1076 cm^{-1} ; **^1H NMR** (500 MHz, CDCl_3) δ 7.68 – 7.65 (m, 1H), 7.36 – 7.31 (m, 2H), 7.31 – 7.28 (m, 1H), 7.28 – 7.23 (m, 3H), 7.23 – 7.18 (m, 3H), 7.17 (dd, $J = 7.3, 2.1$ Hz, 2H), 6.42 (dd, $J = 8.9, 2.4$ Hz, 1H), 6.21 (d, $J = 2.4$ Hz, 1H), 5.32 (q, $J = 7.0$ Hz, 1H), 4.48 – 4.39 (m, 2H), 3.85 (dd, $J = 10.5, 6.8$ Hz, 1H), 3.67 – 3.58 (m, 2H), 3.52 (dt, $J = 14.9, 7.7$ Hz, 2H), 3.45 – 3.39 (m, 2H), 3.30 – 3.23 (m, 1H), 3.15 (dt, $J = 15.8, 8.0$ Hz, 1H), 2.07 (dd, $J = 8.0, 5.2$ Hz, 2H), 2.04 – 1.96 (m, 2H), 1.91 (s, 1H), 1.55 (d, $J = 7.0$ Hz, 3H) ppm; **^{13}C NMR** (126 MHz, CDCl_3) δ 156.3, 151.7, 139.7, 137.9, 128.6, 128.5, 128.4, 127.9, 127.8, 127.7, 127.4, 122.6, 107.0, 104.6, 78.8, 73.0, 70.3, 62.6, 60.2, 56.3, 48.8, 45.9, 28.4, 23.3, 17.7 ppm; **HRMS** calculated for $\text{C}_{29}\text{H}_{35}\text{N}_2\text{O}_5\text{S}$ ($\text{M}+\text{H}$)⁺ 523.2267; found 523.2262 (TOF MS ES+).

(S)-4-((Benzyloxy)methyl)-7-((R)-2-(hydroxymethyl)pyrrolidin-1-yl)-2-((S)-1-phenylethyl)-3,4-dihydro-2H-benzo[b][1,4,5]oxathiazepine 1,1-dioxide (20)

Utilizing general procedure **B**, sultam **20** was isolated (97%, 116 mg, 0.223 mmol) as a colorless viscous oil; $[\alpha]_D^{20} = +21.7^\circ$ ($c = 2.6$, CHCl_3); **FTIR** (neat): 3352, 2941, 1596, 1494, 1377, 1326, 1132, 1074 cm^{-1} ; **^1H NMR** (500 MHz, CDCl_3) δ 7.64 (d, $J = 8.8$ Hz, 1H), 7.39 (m, 2H), 7.36 – 7.31 (m, 4H), 7.31 – 7.28 (m, 2H), 7.26 (m, 2H), 6.42 (dd, $J = 8.8, 2.4$ Hz, 1H), 6.34 (d, $J = 2.4$ Hz, 1H), 5.42 (q, $J = 7.1$ Hz, 1H), 4.53 (q, $J = 11.9$ Hz, 2H), 4.16 (ddd, $J = 10.4, 5.9, 4.6$ Hz, 1H), 3.86 (td, $J = 7.2, 4.1$ Hz, 1H), 3.72 (dt, $J = 10.4, 5.2$

Hz, 1H), 3.69 – 3.61 (m, 1H), 3.55 – 3.49 (m, 1H), 3.47 (dt, $J = 10.6, 5.3$ Hz, 1H), 3.42 (dd, $J = 12.3, 5.2$ Hz, 1H), 3.21 – 3.15 (m, 2H), 2.12 – 2.05 (m, 2H), 2.05 – 1.95 (m, 4H), 1.36 (d, $J = 7.1$ Hz, 3H) ppm; $^{13}\text{C NMR}$ (126 MHz CDCl_3) δ 156.4, 151.9, 140.2, 137.8, 128.5, 128.4, 127.8, 127.7, 127.6, 127.5, 123.5, 107.1, 105.4, 80.9, 73.4, 70.6, 62.6, 60.3, 56.1, 48.9, 46.3, 28.4, 23.3, 17.4 ppm; **HRMS** calculated for $\text{C}_{29}\text{H}_{35}\text{N}_2\text{O}_5\text{S}$ ($\text{M}+\text{H}$) $^+$ 523.2267; found 523.2261 (TOF MS ES+).

(R)-4-((Benzyloxy)methyl)-7-((R)-2-(hydroxymethyl)pyrrolidin-1-yl)-2-((S)-1-phenylethyl)-3,4-dihydro-2H-benzo[b][1,4,5]oxathiazepine 1,1-dioxide (21)

Utilizing general procedure **B**, sultam **21** was isolated (92%, 110 mg, 0.211 mmol) as a colorless viscous oil; $[\alpha]_{\text{D}}^{20} = +1.6^\circ$ ($c = 1.84, \text{CHCl}_3$); **FTIR** (neat): 3390, 2937, 1598, 1496, 1377, 1325, 1130, 1076 cm^{-1} ; $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.66 (dd, $J = 8.7, 4.3$ Hz, 1H), 7.36 – 7.32 (m, 2H), 7.31 – 7.28 (m, 1H), 7.27 – 7.23 (m, 2H), 7.23 – 7.16 (m, 5H), 6.41 (dt, $J = 10.7, 5.4$ Hz, 1H), 6.21 (d, $J = 2.4$ Hz, 1H), 5.33 (q, $J = 7.0$ Hz, 1H), 4.44 (q, $J = 12.1$ Hz, 2H), 3.88 – 3.83 (m, 1H), 3.69 – 3.57 (m, 3H), 3.56 – 3.49 (m, 2H), 3.47 – 3.41 (m, 2H), 3.28 (d, $J = 12.5$ Hz, 1H), 3.21 – 3.13 (m, 1H), 2.11 – 1.97 (m, 4H), 1.83 (s, 1H), 1.55 (d, $J = 7.0$ Hz, 3H) ppm; $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 156.3, 151.6, 139.7, 137.9, 128.6, 128.5, 128.4, 127.9, 127.8, 127.7, 127.4, 122.6, 107.1, 104.7, 78.9, 73.1, 70.3, 62.6, 60.0, 56.3, 48.9, 45.9, 28.4, 23.3, 17.7 ppm; **HRMS** calculated for $\text{C}_{29}\text{H}_{35}\text{N}_2\text{O}_5\text{S}$ ($\text{M}+\text{H}$) $^+$ 523.2267; found 523.2263 (TOF MS ES+).

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

This work was supported by the National Institute of General Medical Science (Center in Chemical Methodologies and Library Development at the University of Kansas, KU-CMLD, NIH P50 GM069663 and NIH P41-GM076302).

References Cited

- (i). Keseru G, Makara GM. *Nature Rev Drug Discov.* 2009; 8:203–212. [PubMed: 19247303]
- (ii). (a) Lovering F, Bikker J, Humblet C. *J Med Chem.* 2009; 52:6752–6756. [PubMed: 19827778] (b) Clemons PA, Bodycombe NR, Carrinski HA, Wilson JA, Shamji AF, Wagner BK, Koehler AN, Schreiber SL. *Proc Natl Acad Sci.* 2010; 107:18787–18792. [PubMed: 20956335]
- (iii). (a) Danishefsky SJ. *Nat Prod Rep.* 2010; 27:1114–1116. [PubMed: 20383353] (b) Ganesan A. *Curr Opin Chem Biol.* 2008; 12:306–317. [PubMed: 18423384]
- (iv). (a) Schreiber SL. *Science.* 2000; 287:1964–1969. [PubMed: 10720315] (b) Burke MD, Berger EM, Schreiber SL. *Science.* 2003; 302:613–618. [PubMed: 14576427] (c) Spring DR. *Org Biomol Chem.* 2003; 1:3867–3870. [PubMed: 14664374] (d) Tan DS. *Nat Chem Bio.* 2005; 1:74–84. [PubMed: 16408003] (e) Schreiber SL. *Nature.* 2009; 457:153–154. [PubMed: 19129834] (f) Damdapani S, Marcaurelle LA. *Curr Opin Chem Biol.* 2010; 14:362–370. [PubMed: 20409744] (g) Marcaurelle LA, Comer E, Dandapani S, Duvall JR, Gerard B, Kesavan S, Lee MD 4th, Liu H, Lowe JT, Marie J-C, Mulrooney CA, Pandya BA, Rowley A, Ryba TD, Suh B-C, Wie J, Young D, Akellam LB, Ross NT, Zhang Y-L, Fass DM, Reis SA, Zhao W-N, Haggarty SJ, Palmer M, Foley MA. *J Am Chem Soc.* 2010; 132:16962–16976. [PubMed: 21067169]
- (v). Rolfe A, Samarakoon TB, Hanson PR. *Org Lett.* 2010; 12:1216–1219. [PubMed: 20178346]
- (vi). (a) Schreiber SL. *Science.* 2000; 287:1964–1969. [PubMed: 10720315] (b) Burke MD, Berger EM, Schreiber SL. *Science.* 2003; 302:613–618. [PubMed: 14576427] (c) Spring DR. *Org Biomol Chem.* 2003; 1:3867–3870. [PubMed: 14664374] (d) Tan DS. *Nat Chem Bio.* 2005; 1:74–84. [PubMed: 16408003] (e) Schreiber SL. *Nature.* 2009; 457:153–154. [PubMed: 19129834] (f) Damdapani S, Marcaurelle LA. *Curr Opin Chem Biol.* 2010; 14:362–370.

[PubMed: 20409744] (g) Marcaurelle LA, Comer E, Dandapani S, Duvall JR, Gerard B, Kesavan S, Lee MD 4th, Liu H, Lowe JT, Marie J-C, Mulrooney CA, Pandya BA, Rowley A, Ryba TD, Suh B-C, Wie J, Young D, Akellam LB, Ross NT, Zhang Y-L, Fass DM, Reis SA, Zhao W-N, Haggarty SJ, Palmer M, Foley MA. *J Am Chem Soc.* 2010; 132:16962–16976. [PubMed: 21067169]

- (vii). (a) Zhang QS, Lu HJ, Curran DP. *J Am Chem Soc.* 2004; 126:36–37. [PubMed: 14709047] (b) Curran DP, Zhang Q, Richard C, Lu H, Gudipathi V, Wilcox CS. *J Am Chem Soc.* 2006; 128:9561–9573. [PubMed: 16848495]
- (viii). Batch scale-up (e.g., 1–2 g) was attempted previously, resulting in yields that were dramatically lower (20–60%, substrate specific), along with poorer percent conversion. In addition, the crude reaction mixtures in batch were far more complex, thus complicating purification. Preliminary investigation into this product loss in batch scale-up suggested a combination of decomposition and potential polymerization of reactive intermediate that did not occur on smaller scale.
- (ix). For a recent review on flow chemistry, see: Razzaq T, Kappe CO. *Chem Asian J.* 2010; 5:1274–1289. [PubMed: 20411525] and references therein.
- (x). (a) Achanta S, Liautard V, Paugh R, Organ MG. *Chem Eur J.* 2010; 16:12797–12800. [PubMed: 20931578] (b) Shore G, Yoo WJ, Li CJ, Organ MG. *Chem Eur J.* 2010; 16:126–133. [PubMed: 19950339] (c) Shore G, Organ MG. *Chem Eur J.* 2008; 14:9641–9646. [PubMed: 18816566] (d) Shore G, Organ MG. *Chem Commun.* 2008:838–840. (e) Shore G, Morin S, Mallik D, Organ MG. *Chem Eur J.* 2008; 14:1351–1356. [PubMed: 18080264]
- (xi). (a) Moseley JD, Woodman EK. *Org Process Res Dev.* 2008; 12:967–981. (b) Moseley JD, Lenden P, Lockwood M, Ruda K, Sherlock J-P, Thomson AD, Gilday JP. *Org Process Res Dev.* 2008; 12:31–40. (c) Bowman MD, Holcomb JL, Kormos CM, Leadbeater NE, Williams VA. *Org Process Res Dev.* 2008; 12:41–57. (d) Kreamsner JM, Stadler A, Kappe CO. *Top Curr Chem.* 2006; 266:233–278.
- (xii). Ullah F, Samarakoon TB, Rolfe A, Kurtz RD, Hanson PR, Organ MG. *Chem Eur J.* 2010:10959–10962. [PubMed: 20715214]
- (xiii). Measured temperatures are not exact values due to the problems associated with measuring the temperature inside a thin capillary tube with a standard IR sensor (designed to measure the temperature of 2–5 mL Pyrex vials).
- (xiv). Due to increased steric hindrance a flow rate of 100 $\mu\text{L}/\text{min}$ vs 200 $\mu\text{L}/\text{min}$ was utilized to maintain high conversion (100% vs 70–80%) for the synthesis of **6–13** on MACOS platform.
- (xv). Rolfe A, Samarakoon TB, Klimberg SV, Brzozowski M, Neuenswander B, Lushington G, Hanson PR. *J Comb Chem.* 2010; 12:850–854. [PubMed: 20879738]

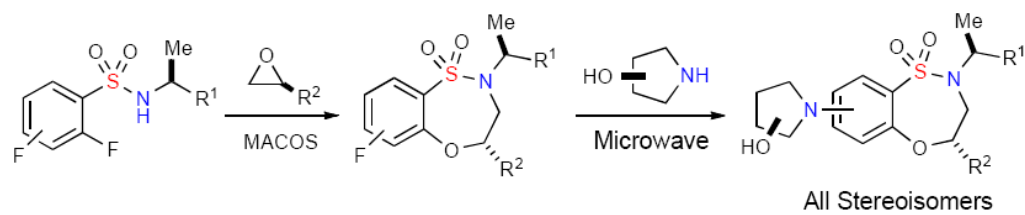
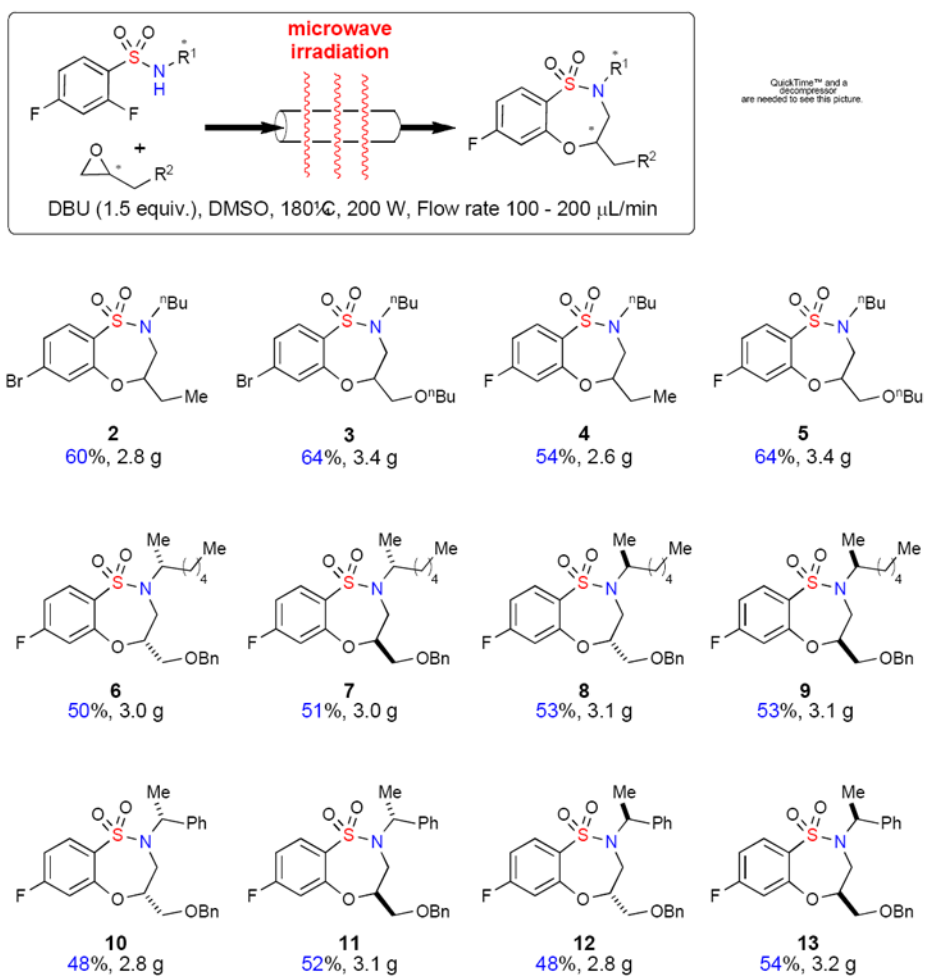
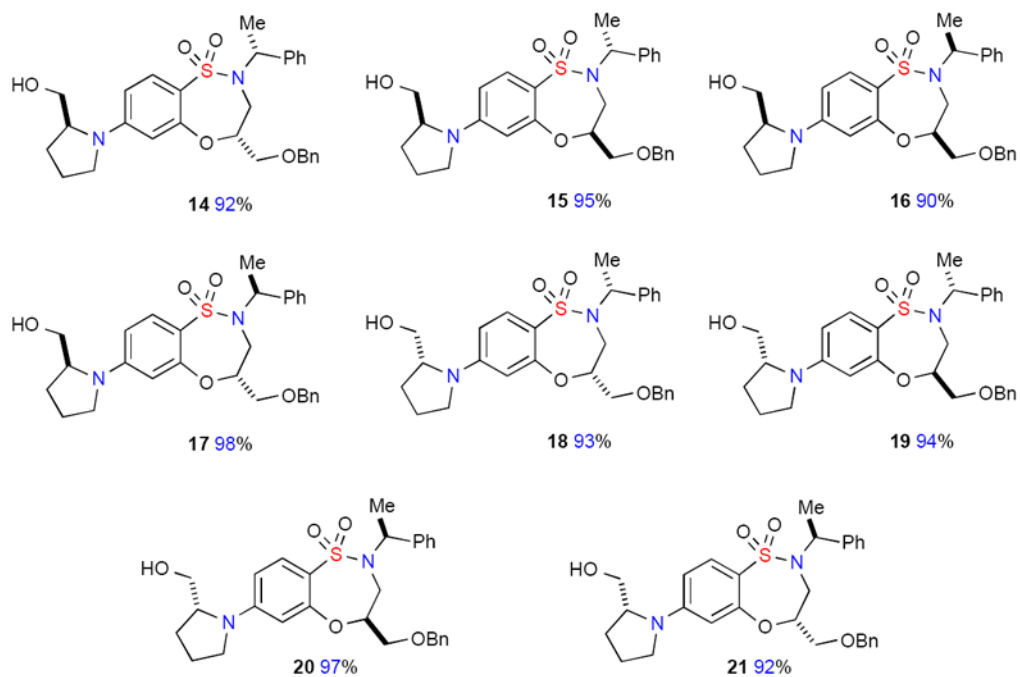
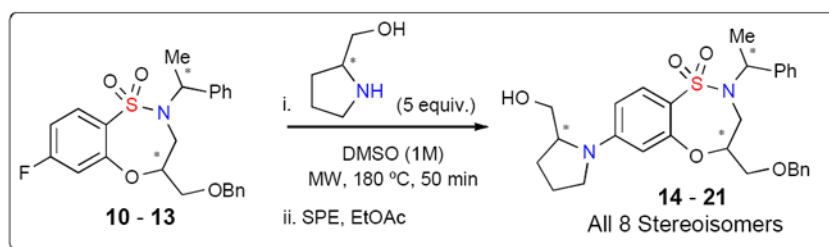


Figure 1.
Two-step synthesis of stereochemically-rich benzothiazepine-1,1-dioxides.



Scheme 1.
MACOS scale-out of optically-pure benzothiazepine-1,1'-dioxides **6-13**.



Scheme 2.
 Generation of optically-pure benzosultams **14 – 21** via microwave-assisted $\text{S}_{\text{N}}\text{Ar}$.

Table 1

Optimization studies of epoxide opening/cyclization sequence using batch MW.

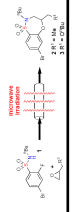
Entry	Temp (°C)	Time (min)	Base (equiv.)	Epoxy (equiv.)	% Con. (Yield)	Prod.
1 ^a	125	3	tBuOK (3)	1	Decomp.	2
2 ^a	110	3	tBuOK (2)	1	5	2
3 ^a	110	5	tBuOK (1.5)	1	30	2
4 ^a	110	20	tBuOK (1.5)	1.2	54	2
5 ^a	180	1	tBuOK (1.5)	1.2	61	2
6 ^a	180	1	tBuOK (1.5)	2	78	2
7 ^a	180	1	tBuOK (1.5)	3	100 (70)	2
<hr/>						
8 ^b	125	1	DBU (1.5)	3	25	3
9 ^b	150	1	DBU (1.5)	3	60	3
10 ^b	180	1	DBU (1.5)	3	100 (90)	3
11 ^b	RT	72 h	DBU (1.5)	3	26	3

[^a] Compound 2 generated with 2-ethylloxirane;

[^b] Compound 3 generated with 2-(butoxymethyl) oxirane.

Table 2

Optimization studies for MACOS scale-out of benzothiazoxazepine-1,1-dioxide **2** and **3**.



Entry ^a	Flow rate (μL/min)	Temp. (°C)	Power (W)	% Con. (Yield)	Prod.
1	50	145	380	90 (28)	2
2	50	150	240	34	2
3	50	200	215	100	2
4	75	195	230	100	2
5	100	195	230	100	2
6	125	190	225	100 (44)	2
7	150	190	220	100 (50)	2
8 ^b	150	190	220	100 (78)	3

[^a] **1** (1.0 equiv.), Epoxide (3.0 equiv.), ^tBuOK (1.5 equiv), DMSO (0.4 M);

[^b] **1** (1.0 equiv.), Epoxide (3.0 equiv.), DBU (1.5 equiv), DMSO (0.4 M).