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Ultrathin Gold Nanowire-Functionalized Carbon Nanotubes for Hybrid Molecular Sensing

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Abstract

Carbon nanotubes (CNTs) have shown great potential as sensing component in the electrochemical, field effect transistor and optical sensors, because of their extraordinary onedimensional electronic structure, thermal conductivity, tunable and stable near-infrared emission. However, the insolubility of CNTs due to strong van der Waals interactions limits their use in the field of nanotechnology. In this study, we demonstrate that non-covalent ultrathin gold nanowires functionalized multi-walled carbon nanotube (GNW-CNT) hybrid sensing agents show highly efficient and selective immune molecular sensing in electrochemical and near-infrared photoacoustic imaging methods. A detection limit of 0.01 ng/mL for the Alpha-Fetoprotein (AFP) antigen with high selectivity is shown. The extraordinary optical absorption, thermal and electric conductivity of hybrid GNW-CNTs presented in this study could be an effective tactic to integrate imaging, sensing and treatment functionalities.

Keywords

carbon nanotubes; gold nanowires; in-situ synthesis; electrochemical; photoacoustic imaging

Carbon nanotubes (CNTs) classified as single-walled CNTs (SWCNTs) and multi-walled CNTs (MWCNTs), have been attracting much interest since they were first introduced in 1990s, because of their excellent chemical stability, high thermal and electronic conductivity, and broad spectral optical absorption. Therefore, CNTs have been intensively investigated in many emerging applications, including field-effect emitters,^{1, 2} solar cells^{3–5} and hydrogen storage medium.^{6–8}

In recent years, CNTs have shown great potential in biomedical applications, such as bioimaging, sensing and drug delivery in cells and animals due to their strong optical absorption, high thermal conductivity and long-circulating characteristics.^{9–13} Particularly, CNTs absorb light in the near-infrared (NIR, such as700–900 nm), where tissue optical scattering and absorption reaches minima, thus they can image more deeply within tissues with high spatial resolution. CNTs have been used in various biomedical imaging applications, such as NIR photoluminescence imaging,¹⁴ Raman imaging,^{15, 16} magnetic resonance imaging (MRI)^{17, 18} and photoacoustic (PA) imaging.^{19–24} However, due to their

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Supporting Information Available. Description of particles synthesis, photoacoustic imaging instrument, morphology change of GNWs, experimental procedures. This material is available free of charge *via* the Internet at http://pubs.acs.org.

highly hydrophobic property, surface functionalization of CNTs should be performed to obtain high water solubility and biocompatibility before bioconjugation process. In the past, the chemical covalent or physical non-covalent approaches have been employed for surface functionalization.^{25–29} Chemical reactions are most widely studied to covalently functionalize CNTs by using harsh experimental conditions, such as strong acids or oxidants, but they usually create a number of defects on the sidewalls of CNTs and affect their electronic and optical properties.³⁰ Non-covalent interactions utilize the surfactant or polymers to coat on CNTs, which show promises to increase their water solubility, prevent the aggregation and enhance their biocompatibility.³¹ However, non-covalent interactions between CNTs and surfactant or polymers are not stable in the biological fluids, and simultaneously preserving the optical property of the CNTs remains as a central challenge.

In this study, we develop *in-situ* solution-phase synthesis of ultrathin gold nanowires (GNWs) incorporated MWCNTs as a model sensing system, in which non-covalent functionalization is shown to provide efficient electrochemical sensing and NIR PA imaging. The ultrahigh surface to volume ratio of the MWCNTs and *in-situ* GNWs synthesis allows for efficient inside and outside non-covalent functionalization. Since ultrathin gold nanowire surfaces exhibit excellent biocompatibility to incorporate additional surface functionalities,^{32, 33} the resulting water-soluble hybrids exhibit high optical absorbance while obtaining the abilities of bioconjugation, and also extraordinary thermal conductivity than that of MWCNTs, which is an important property for PA imaging. Additionally, we show the hybrid GNW-CNTs can be selective for detecting Alpha-Fetoprotein (AFP) antigen as the model system, using both electrochemical and NIR PA imaging techniques. AFP antigen is the early indicator of hepatocarcinogenesis and an important biomarker of hepatocellular carcinoma (HCC), which is the most common type of liver cancer.^{34–36} Our results demonstrate that the GNW-CNT hybrids can offer a promising alternative for non-invasive detection of AFP antigen.

In the following report, the structure of solution-phase GNW-CNTs will be firstly introduced. Moreover, we will show the PA imaging characteristics based on unique optical and thermal properties of the hybrid system. The GNW-CNTs hybrids for sensing the AFP antigen using both electrochemistry and PA imaging will also be presented.

RESULTS AND DISCUSSION

Figure 1(a) shows the schematic structure of the GNW-CNT hybrids. The GNW-CNTs are consisted of multi-walled carbon nanotube core with a diameter of 40~60 nm, which is noncovalently incorporated with ultrathin gold nanowires. In order to enhance water solubility of GNW-CNT, the cetyltrimethylammonium bromide (CTAB) and sodium citrate was used to transfer GNW-CNT hybrid phase from hexane to water. The well dispersed GNW-CNTs with relatively uniform size were highly water-soluble due to gold nanowire surface functionalization. Transmission electron microscopy (TEM) analysis of the GNW-CNTs (Fig. 1(b)) demonstrates that the gold nanowires with a diameter about 1 nm are incorporated outside and inside MWCNTs. Surface morphology of hybrid GNW-CNTs thin films by atomic force microscopy (AFM) confirms the fiber-like structure, shown in Fig. 1(c). The photoabsorption spectra of the GNW-CNTs (Fig. 1(d)) show a transverse plasmon absorption in the visible region (510~530 nm) and a wide longitudinal resonance peak in the IR region. Compared to the absorption spectrum of MWCNTs (Supplementary Fig.2), we attribute the IR absorption from high aspect ratio of ultrathin gold nanowires.³⁷ To confirm this localized surface plasmon resonance (LSPR), we study the refractive index effect of the solvent medium on the photoabsorption of GNW-CNT hybrids. Three different solvents have been applied: hexane, Carbon tetrachloride (CCl_4), and Carbon sulfide (CS_2) with refractive indices of 1.37, 1.46, and 1.63, respectively. The inset of Fig. 1(d) indicates that

the absorption band from GNW-CNTs redshifts with increasing refractive index, which confirms the LSPR feature of ultrathin gold nanowires incorporated CNTs, but not scattering or sample impurities.³⁸ The absorption of GNW-CNTs in hexane over 1600 nm incident light wavelength was not included in the figure, because the hexane shows extremely high intensity noise and strong interference in the signals from GNW-CNTs.

For PA applications, we explore the PA signal amplitudes of MWCNTs, GNW-CNT hybrids, MWCNTs mixed with GNWs, and GNWs, shown in Fig. 2. Figure 2(a) shows corresponding maximum amplitude projection (MAP) PA images. Figure 2(b) through 2(e) show the PA signals and the signal profile at the horizontal dashed line position in (a), respectively. The GNW-CNT hybrids exhibit significantly stronger PA signal amplitudes than that of MWCNTs and GNWs. Since the PA imaging is an absorption-based technique, the enhancement in PA amplitude are a result of the increase in optical absorption of GNW-CNT hybrids. Particularly, the PA amplitude of GNW-CNTs was also stronger than that of the randomly mixed GNWs and MWCNTs. The enhancement of GNW-CNT over CNT mixed with GNW was calculated to be about 25.66%. We attribute such PA signal amplitude increase to the enhanced absorption from NIR plasmonic resonance of GNWs under 820 nm laser wavelength used in the PA imaging. Therefore, GNWCNTs exhibit higher optical absorption than that of MWCNT, and leading to strong PA signals.

In addition, the GNWs are wrapped and incorporated into MWCNTs to facilitate the formation of percolation network and the interaction between GNW and MWCNT which will enhance thermal transfer and the overall thermal conductivity of the hybrids structure. In addition, the PA signal amplitude is dependent on the applied laser fluence and the MWCNT concentration. Figure 3(a) shows a nonlinear signal enhancement as the laser fluence increases at 820 nm laser wavelength. We postulate that the nonlinear PA signal increase is originated from the change of thermal expansion coefficient at the high laser fluence during the local temperature increase with thermal coupling.^{39–41} It is consistent with previous reports that the PA amplitude of MWCNTs exhibits a linear relationship as their concentration increases (Fig. 3(b)). However, the amplitude from the GNW-CNT hybrids shows a drastic enhancement starting from $\sim 1.5 \,\mu g/mL$ CNTs concentration compared to the MWCNTs. This result further confirms that the integrated structure of GNWs and MWCNTs can result in enhanced optical absorption and improved thermal properties, and thus, stronger PA signal amplitude. The morphological change of GNWs under long laser exposure was also observed. TEM images (Fig. 3(c) and 3(d)) show the melting behavior of GNWs on MWCNTs before and after high intensity laser energy, in which the GNWs were disconnected into rods slowly and into spheres eventually over time. A similar process has been reported for the gold nanowires under HRTEM analysis.⁴² Also, we found that the GNW-CNTs melted faster than those free GNWs which were not attached on the MWCNTs (see Supplementary Fig.3), which could be attributed to the high thermal conductivity of CNTs to transfer heat to the GNWs causing the melting behavior. This characteristic may have the potential as cancer drug carrier for the drug release into the targeting area under the laser irradiation for future drug delivery and cancer treatment.⁴³

For specific molecular sensing and targeting applications, we explore the GNW-CNT hybrids by functionalizing GNWs with the antibody, which is specific to the AFP antigen. Figure 4(a) shows the schematic of each step for the immune test using the electrochemistry method. In this test, GNWs on the surface of MWCNT can facilitate the Au-S interaction with the thiol groups of Cys residues in anti-AFP,⁴⁴ which enhances the affinity of anti-AFP to GNW-CNT. Figure 4(b) shows the redox peak current under different testing conditions. The redox peak current of the GNW-CNT modified electrode is significantly higher than that of bare electrodes without GNW-CNTs, suggesting that the GNW-CNTs film enhances electron transfer by providing a more conductive medium for the electron movement. After

the electrode was coated with anti- AFP antibody and bovine serum albumin (BSA) films, the peak currents of the electrode decreased significantly as a result of the low conductivity of anti-AFP/BSA films. In addition, the incubation of electrode with AFP antigen caused the peak current to decrease even more. Figure 4(c) shows that the peak current has a nonlinear decrease as AFP antigen concentration increases. The presented results were obtained from repeated experiments, and a small data spread range from the error bar showed an excellent reproducibility of this electrochemical sensing. As AFP concentration increases, the signal change to concentration ratio is decreased. This result can be understood by the consideration of the molecular size. As the concentration of AFP antigen increases, the incoming molecules were sterically hindered by the existed bioconjugated antigen. The result from the immune test using electrochemistry confirms the capability of GNW-CNTs for simple and robust bioconjugation. Based on the traditional protocol, the limit of detection (LOD) is estimated to the concentration corresponding to three times of the standard deviation of the noise. The standard deviation from our repeated experiments without AFP was shown 0.13 µA. The difference of signal amplitude with AFP at a concentration of 0.01 ng/mL, and without AFP is about 0.87 µA. It should be noted that 0.01 ng/mL AFP can be detected with this sensor by considering the standard deviation (0.39 μ A) lower than $0.87 \,\mu$ A.

To further investigate the feasibility of GNW-CNTs to detect AFP antigen, PA imaging immune test was conducted. Figure 5(a) shows a similar bioconjugation process for the PA imaging immune test. The biotinylated monoclonal antibody (BMA), which attached on the bottom of the microplates, was not well dispersed but accumulated on the corner as shown in Fig. 5(b). The PA imaging was performed on both treated wells and controlled wells. In Fig. 5(c) and 5(d), the weak PA signal amplitudes can be observed from the controlled wells. In the PA images of treated wells (Fig. 5(e)-5(h)), the bright area, showing the enhancement in the PA signals, indicates the existence of the GNW-CNTs and demonstrates that AFP antigen coupling to anti-AFP antibody conjugated GNW-CNTs can be monitored by the increase in the PA signal amplitude. In addition, the bright area matches very well with the location of the BMA. To quantify the change of PA amplitude after linking anti-AFP antibody conjugated GNW-CNTs with different concentrations of AFP, we calculated averaged PA amplitude enhancement in the approximate BMA immobilized region (Fig. 5(b)). Figure 5(i) shows the calculated PA amplitude as a function of AFP antigen concentration. The PA amplitude shows a linear enhancement until 5 ng/mL AFP antigen concentration, below which the antibody functionalized GNW-CNTs were gradually immobilized on the increasing number of AFP antigen, leading to the increased PA signal amplitude. However, when AFP antigen concentration is higher than 5 ng/mL, the PA amplitude reaches a saturation level, which agrees with the result from electrochemistry method, and can be attributed by the sterically hindering of large molecular size AFP antigen.

In order to demonstrate that GNW-CNT hybrids have better sensing sensitivity than GNW, the immune test using GNW as the sensor was performed. First, anti-AFP was immobilized on the surface of GNWs by mixing two solutions for 12 hours. Second, bovine serum albumin (BSA) was added to the resulting solution for blocking nonspecific absorber site on the GNWs for another 2 hours. Third, 25 μ L AFP serum was pipetted into the assigned biotinylated monoclonal antibody (BMA) coated microwells with different concentrations for incubating at room temperature and washed by PBS solution after 20 minutes in order to link the antibody to BMA. BMA were overdosed than AFP. Finally, 100 μ L of the anti-AFP antibody conjugated GNWs were added to each well and washed again by PBS solution after 20 minutes. The PA images were obtained on the treated microwells with different AFP concentrations as shown in figure 1. From the images, we can observe very weak PA signals from the center of the microwells at 25 ng/mL and 50 ng/mL AFP concentrations. At

75 ng/mL and 100 ng/mL AFP concentrations, PA signals become stronger because of the attachment of more GNWs on the microwells. However, we can observe very strong PA signals at 5 ng/mL AFP concentration with GNW-CNT as the sensor. Therefore, it is concluded that GNW-CNT hybrids have a much better sensitivity than GNWs.

CONCLUSIONS

In summary, we demonstrate solution-phase synthesized ultrathin gold nanowires incorporated carbon nanotube hybrids as both NIR photoacoustic imaging and efficient electrochemical sensing agents. Non-covalent ultrathin gold nanowire functionalized CNTs preserve the extraordinary optical and thermal properties, and more importantly, exhibit high water solubility and further biological molecule functionalization. The presented hybrid golden carbon nanotubes own high optical absorption, thermal and electric conductivity. We showed that high selectivity and sensing efficiency of hybrid GNW-CNTs by conjugating with targeted molecules. In addition, the morphology change under laser exposure and high thermal conductivity of CNTs also facilitates the disconnection process of GNWs, which could open up potential applications by releasing the drug conjugated on the GNWs into the targeted area under the laser illumination at certain intensity. Moreover, we showed that GNW-CNT conjugated with anti-AFP antigen has the ability to specifically link to the AFP antigen, and demonstrated a detection limit of 0.01 ng/mL for the AFP antigen with high selectivity, as one model detection system.

EXPERIMENTAL METHODS

A. In-situ synthesis of ultrathin GNW-CNTs

In a typical synthesis, MWCNTs were dispersed in hexane with 0.5 mg/mL concentration. Then, 100 μ L oleylamine (OA) mixed with 3 mg HAuCl₄ was added to the 2.5 ml MWCNTs and hexane mixture, followed by the addition of 150 μ L triisopropylsilane (TIPS) to form a solution. Then the reaction proceeded at room temperature without stirring for 24 hours. The final products were centrifuged, washed by ethanol and finally redispersed in hexane for further investigation.

In order to enhance water solubility of GNW-CNT, the following modification process has been performed. A mixture of 0.1 M cetyltrimethylammonium bromide (CTAB) and 0.01 mM sodium citrate was added to GNW-CNT solution for phase transfer from hexane to water. The transfer will be completed after at least 24 hours. GNW-CNT in water was used for immune test.

B. PA imaging system setup

The system schematic is show in Supplementary Figure 5. The system consists of an ultrasound transducer, a laser system, and receiving electronics. A Qswitched Nd:YAG laser (Surelite; Continuum, Santa Clara, CA) was used to pump a tunable OPO laser (Surelite OPO PLUS; Continuum, Santa Clara, CA) to obtain a 820 nm wavelength laser with a 10-Hz pulse repetition rate. The produced laser light forms a ring shape illumination after passing through several prisms and a conical lens and then is refocused inside the sample by an optical condenser. The subsequently generated ultrasonic waves are detected by a 5-MHz focused ultrasonic transducer (SU-108-013; Sonic Concepts, Bothell, WA) which was mounted in the middle of the condenser lens. The conical and condenser lenses were driven by a 3D translation stage to enable the transducer to mechanically scan the targeted region. Two prisms were used to enable the optical beam-folding for a 2D mechanical scanning. Prism Y could only move on the Y direction, whereas prism XY could move on both X and Y direction. The recorded photoacoustic signals by the transducer were amplified through a

pre- amplifier (5072PR; Olympus-NDT, Waltham, MA) and then collected by a PC through an A/D Scope Card (CS21G8-256MS; Gage, Lockport, IL) with a 250-MHz sampling rate. This design has been presented by Song *et al.*⁴⁵ for deep PAI, and proved to generate goodresolution photoacoustic images in deep regions inside soft tissue.⁴⁶

C. PA signals amplitude comparison among four nanoparticles

Four particles solutions dispersed in DI water were injected into tygon tubing with GNWs and MWCNTs concentrations of 0.35 mg/mL and 0.5 mg/mL for PA imaging. Laser wavelength and laser fluence used in this experiment was 820 nm and 14 mJ/cm², respectively.

D. Electrochemistry immune test

In electrochemistry assessment of the functionalization capability of GNW-CNTs, The patterned gold electrode was sputtered 100 nm using Sputter Coater on a Si substrate. First, 40 μ L GNW-CNTs were spin-coated on the cleaned electrode at 1800 r/min for 30s and the electrode was washed with DI water. Second, 30 μ L of anti-AFP antibody solution was dropped on the electrode for 12 h at 4 °C to anchor the antibody on the surface of the electrode. Third, 40 μ L bovine serum albumin (BSA) solution was dipped on the electrode for about 2 h at 4 °C in order to block possible remaining nonspecific bind of GNW-CNTs active sites. At last, 30 μ L AFP serum at different concentrations were dipped on the electrode was then washed carefully with DI water and stored at 4 °C before using. The schematic of the bioconjugation process is shown in Fig. 4(a). Cyclic voltammograms of the different steps of modified electrodes were then carried out in pH 7.5 PBS buffer containing 1.0 mmol/L Fe(CN)₆^{3-/4-} (Fig. 4(b)).

E. Photoacoustic immune test

Anti-alpha fetoprotein (AFP) antibody was immobilized on the surface of GNW-CNTs by mixing two solutions for 12 hours. Then, Bovine serum albumin (BSA) was added to the resulting solution for blocking nonspecific absorber site on the GNW-CNTs for another 2 hours. The anti-AFP antibody conjugated GNW-CNTs were ready to use. For the treated group, 25 μ L AFP serum was pipetted into the assigned biotinylated monoclonal antibody (BMA) coated microwells with different concentrations for incubating at room temperature and washed by PBS solution after 20 minutes in order to link the antibody to BMA. BMA were overdosed than AFP. Then, 100 μ L of the anti-AFP antibody conjugated GNW-CNTs were added to each well and washed again by PBS solution after 20 minutes. For controlled group 1, only anti-AFP antibody is added to the AFP-incubated well. For controlled group 2, bare GNW-CNTs, which were not conjugated with anti-AFP antibody but blocked by BSA, were added to the AFP-incubated well. The laser wavelength and fluence used in the immune test was 820 nm and 14 mJ/cm², respectively.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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REFERENCES AND NOTES

- Niemann DL, Ribaya BP, Gunther N, Rahman M, Leung J, Nguyen CV. Effects of Cathode Structure on the Field Emission Properties of Individual Multi-Walled Carbon Nanotube Emitters. Nanotechnology. 2007; 18 485702.
- 2. Park N, Han SW, Ihm J. Effects of Oxygen Adsorption on Carbon Nanotube Field Emitters. Physical Review B. 2001; 64 125401.
- Kongkanand A, Dominguez RM, Kamat PV. Single Wall Carbon Nanotube Scaffolds for Photoelectrochemical Solar Cells. Capture and Transport of Photogenerated Electrons. Nano Lett. 2007; 7:676–680. [PubMed: 17309316]
- Lu R, Christianson C, Kirkeminde A, Ren S, Wu J. Extraordinary Photocurrent Harvesting at Typeli Heterojunction Interfaces: Toward High Detectivity Carbon Nanotube Infrared Detectors. Nano Lett. 2012; 12:6244–6249. [PubMed: 23130570]
- Ruzicka BA, Wang R, Lohrman J, Ren S, Zhao H. Exciton Diffusion in Semiconducting Single-Walled Carbon Nanotubes Studied by Transient Absorption Microscopy. Physical Review B. 2012; 86 205417.
- Dillon AC, Jones KM, Bekkedahl TA, Kiang CH, Bethune DS, Heben MJ. Storage of Hydrogen in Single-Walled Carbon Nanotubes. Nature. 1997; 386:377–379.
- 7. Lee SM, Lee YH. Hydrogen Storage in Single-Walled Carbon Nanotubes. Appl. Phys. Lett. 2000; 76:2877–2879.
- Liu C, Fan YY, Liu M, Cong HT, Cheng HM, Dresselhaus MS. Hydrogen Storage in Single-Walled Carbon Nanotubes at Room Temperature. Science. 1999; 286:1127–1129. [PubMed: 10550044]
- de la Zerda A, Liu Z, Bodapati S, Teed R, Vaithilingam S, Khuri-Yakub BT, Chen X, Dai H, Gambhir SS. Ultrahigh Sensitivity Carbon Nanotube Agents for Photoacoustic Molecular Imaging in Living Mice. Nano Letters. 2010; 10:2168–2172. [PubMed: 20499887]
- De La Zerda A, Zavaleta C, Keren S, Vaithilingam S, Bodapati S, Liu Z, Levi J, Smith BR, Ma T-J, Oralkan O, et al. Carbon Nanotubes as Photoacoustic Molecular Imaging Agents in Living Mice. Nat. Nanotechnol. 2008; 3:557–562. [PubMed: 18772918]
- Pramanik M, Song KH, Swierczewska M, Green D, Sitharaman B, Wang LV. In Vivo Carbon Nanotube-Enhanced Non-Invasive Photoacoustic Mapping of the Sentinel Lymph Node. Physics in Medicine and Biology. 2009; 54:3291–3301. [PubMed: 19430111]
- Pramanik M, Swierczewska M, Green D, Sitharaman B, Wang LV. Single-Walled Carbon Nanotubes as a Multimodal-Thermoacoustic and Photoacoustic-Contrast Agent. J. Biomed. Opt. 2009; 14 034018.
- Xiang L, Yuan Y, Xing D, Ou Z, Yang S, Zhou F. Photoacoustic Molecular Imaging with Antibody-Functionalized Single-Walled Carbon Nanotubes for Early Diagnosis of Tumor. Journal of Biomedical Optics. 2009; 14 021008.
- Lefebvre J, Austing DG, Bond J, Finnie P. Photoluminescence Imaging of Suspended Single-Walled Carbon Nanotubes. Nano Lett. 2006; 6:1603–1608. [PubMed: 16895343]
- Doorn SK, Zheng LX, O'Connell MJ, Zhu YT, Huang SM, Liu J. Raman Spectroscopy and Imaging of Ultralong Carbon Nanotubes. J. Phys. Chem. B. 2005; 109:3751–3758. [PubMed: 16851421]
- 16. Mews A, Koberling F, Basche T, Philipp G, Duesberg GS, Roth S, Burghard M. Raman Imaging of Single Carbon Nanotubes. Adv. Mater. (Weinheim, Ger.). 2000; 12:1210–1214.
- Al Faraj A, Cieslar K, Lacroix G, Gaillard S, Canot-Soulas E, Cremillieux Y. In Vivo Imaging of Carbon Nanotube Biodistribution Using Magnetic Resonance Imaging. Nano Lett. 2009; 9:1023– 1027. [PubMed: 19199447]
- Wu H, Liu G, Wang X, Zhang J, Chen Y, Shi J, Yang H, Hu H, Yang S. Solvothermal Synthesis of Cobalt Ferrite Nanoparticles Loaded on Multiwalled Carbon Nanotubes for Magnetic Resonance Imaging and Drug Delivery. Acta Biomaterialia. 2011; 7:3496–3504. [PubMed: 21664499]
- 19. Xu MH, Wang LHV. Photoacoustic Imaging in Biomedicine. Rev. Sci. Instrum. 2006; 77 041101.
- Esenaliev RO, Karabutov AA, Oraevsky AA. Sensitivity of Laser Opto-Acoustic Imaging in Detection of Small Deeply Embedded Tumors. IEEE J. Sel. Top. Quantum Electron. 1999; 5:981– 988.

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- Hoelen CGA, de Mul FFM, Pongers R, Dekker A. Three-Dimensional Photoacoustic Imaging of Blood Vessels in Tissue. Opt. Lett. 1998; 23:648–650. [PubMed: 18084605]
- 22. Wang LV. Tutorial on Photoacoustic Microscopy and Computed Tomography. IEEE J. Sel. Top. Quantum Electron. 2008; 14:171–179.
- 23. Ku G, Wang LHV. Deeply Penetrating Photoacoustic Tomography in Biological Tissues Enhanced with an Optical Contrast Agent. Opt. Lett. 2005; 30:507–509. [PubMed: 15789718]
- 24. Song KH, Wang LV. Deep Reflection-Mode Photoacoustic Imaging of Biological Tissue. J. Biomed. Opt. 2007; 12 060503.
- 25. Liu JQ, Chou A, Rahmat W, Paddon-Row MN, Gooding JJ. Achieving Direct Electrical Connection to Glucose Oxidase Using Aligned Single Walled Carbon Nanotube Arrays. Electroanalysis. 2005; 17:38–46.
- Gooding JJ, Wibowo R, Liu JQ, Yang WR, Losic D, Orbons S, Mearns FJ, Shapter JG, Hibbert DB. Protein Electrochemistry Using Aligned Carbon Nanotube Arrays. J. Am. Chem. Soc. 2003; 125:9006–9007. [PubMed: 15369344]
- 27. Kim J, Baek J, Kim H, Lee K, Lee S. Integration of Enzyme Immobilized Single-Walled Carbon Nanotubes Mass into the Microfluidic Platform and Its Application for the Glucose-Detection. Sensors and Actuators a-Physical. 2006; 128:7–13.
- Wang H, Wang CC, Lei CX, Wu ZY, Shen GL, Yu RQ. A Novel Biosensing Interfacial Design Produced by Assembling Nano-Au Particles on Amine-Terminated Plasma-Polymerized Films. Anal. Bioanal. Chem. 2003; 377:632–638. [PubMed: 12937885]
- Wang Y, Joshi PP, Hobbs KL, Johnson MB, Schmidtke DW. Nanostructured Biosensors Built by Layer-by-Layer Electrostatic Assembly of Enzyme-Coated Single-Walled Carbon Nanotubes and Redox Polymers. Langmuir. 2006; 22:9776–9783. [PubMed: 17073511]
- Sui X-M, Giordani S, Prato M, Wagner HD. Effect of Carbon Nanotube Surface Modification on Dispersion and Structural Properties of Electrospun Fibers. Appl. Phys. Lett. 2009; 95 233113.
- Shi J, Cha T-G, Claussen JC, Diggs AR, Choi JH, Porterfield DM. Microbiosensors Based on DNA Modified Single-Walled Carbon Nanotube and Pt Black Nanocomposites. Analyst. 2011; 136:4916–4924. [PubMed: 21858297]
- Mukhopadhyay K, Phadtare S, Vinod VP, Kumar A, Rao M, Chaudhari RV, Sastry M. Gold Nanoparticles Assembled on Amine-Functionalized Na-Y Zeolite: A Biocompatible Surface for Enzyme Immobilization. Langmuir. 2003; 19:3858–3863.
- Eustis S, El-Sayed MA. Why Gold Nanoparticles Are More Precious Than Pretty Gold: Noble Metal Surface Plasmon Resonance and Its Enhancement of the Radiative and Nonradiative Properties of Nanocrystals of Different Shapes. Chem. Soc. Rev. 2006; 35:209–217. [PubMed: 16505915]
- Johnson PJ. The Role of Serum Alpha-Fetoprotein Estimation in the Diagnosis and Management of Hepatocellular Carcinoma. Clinics in liver disease. 2001; 5:145–159. [PubMed: 11218912]
- 35. Trevisani F, D'Intino PE, Morselli-Labate AM, Mazzella G, Accogli E, Caraceni P, Domenicali M, De Notariis S, Roda E, Bernardi M. Serum Alpha-Fetoprotein for Diagnosis of Hepatocellular Carcinoma in Patients with Chronic Liver Disease: Influence of Hbsag and Anti-Hcv Status. Journal of Hepatology. 2001; 34:570–575. [PubMed: 11394657]
- 36. Yamashita T, Forgues M, Wang W, Kim JW, Ye Q, Jia H, Budhu A, Zanetti KA, Chen Y, Qin L-X, et al. Epcam and Alpha-Fetoprotein Expression Defines Novel Prognostic Subtypes of Hepatocellular Carcinoma. Cancer Res. 2008; 68:1451–1461. [PubMed: 18316609]
- Guidez EB, Aikens CM. Theoretical Analysis of the Optical Excitation Spectra of Silver and Gold Nanowires. Nanoscale. 2012; 4:4190–4198. [PubMed: 22495627]
- Luther JM, Jain PK, Ewers T, Alivisatos AP. Localized Surface Plasmon Resonances Arising from Free Carriers in Doped Quantum Dots. Nat. Mater. 2011; 10:361–366. [PubMed: 21478881]
- Oshurko VB. Imaging Thermal Fields in Nonlinear Photoacoustics. Technical Physics Letters. 2006; 32:691–693.
- Egerev SV, Oraevsky AA. Optothermoacoustic Phenomena in Highly Diluted Suspensions of Gold Nanoparticles. Int. J. Thermophys. 2008; 29:2116–2125.

- 41. Sanchot A, Baffou G, Marty R, Arbouet A, Quidant R, Girard C, Dujardin E. Plasmonic Nanoparticle Networks for Light and Heat Concentration. ACS Nano. 2012; 6:3434–3440. [PubMed: 22394263]
- Pazos-Perez N, Baranov D, Irsen S, Hilgendorff M, Liz-Marzan LM, Giersig M. Synthesis of Flexible, Ultrathin Gold Nanowires in Organic Media. Langmuir. 2008; 24:9855–9860. [PubMed: 18652498]
- Wijaya A, Schaffer SB, Pallares IG, Hamad-Schifferli K. Selective Release of Multiple DNA Oligonucleotides from Gold Nanorods. ACS Nano. 2009; 3:80–86. [PubMed: 19206252]
- 44. Li N, Yuan R, Chai Y, Chen S, An H, Li W. New Antibody Immobilization Strategy Based on Gold Nanoparticles and Azure I/Multi-Walled Carbon Nanotube Composite Membranes for an Amperometric Enzyme Immunosensor. J. Phys. Chem. C. 2007; 111:8443–8450.
- 45. Song KH, Wang LV. Deep Reflection-Mode Photoacoustic Imaging of Biological Tissue. Journal of Biomedical Optics. 2007; 12
- 46. Zhang T, Cui HZ, Fang CY, Su LJ, Ren SQ, Chang HC, Yang XM, Forrest ML. Photoacoustic Contrast Imaging of Biological Tissues with Nanodiamonds Fabricated for High near-Infrared Absorbance. J. Biomed. Opt. 2013; 18 026018.

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Figure 1.

(a) Schematic of GNW-CNTs; (b) TEM image, scale bars: 50 nm. Inset scale bar: 20 nm; (c) AFM image, scale bars: 100 nm; (d) Absorption spectra of GNW-CNTs. The inset shows the red-shift in the LSPR maximum with increasing solvent refractive index.



Figure 2.

(a) PA MAP image of four particle solutions in tubings (from left to right: plain MWCNTs, MWCNTs wrapped with GNWs, MWCNTs mixed with GNWs, and pure GNWs); (b–e) PA signals from particle solutions; (f) Signal profile at the horizontal dashed line position in (a).

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(a) PA signals amplitudes from GNW-CNTs as a function of laser fluence; (b) PA signals amplitudes as a function of CNTs concentration. The error bars represent the standard deviation in five measurements, blue line: PA amplitude from MWCNTs, red line: PA amplitude from GNW-CNTs; (c–d) TEM images showing gold nanowires degradation upon long exposure to the high-energy electron beam during HRTEM analysis, scale bars: 20 nm.

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Figure 4.

(a) Schematic of the bioconjugation for the electrochemistry immune test. (b) Peak current amplitude in different steps of modified electrodes. (c) Peak current as a function of the AFP antigen concentration.

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Figure 5.

(a) Schematic of the controlled and treated methods of the immune tests using GNW-CNTs; (b) A schematic shows the location of the Biotinylated Monoclonal Antibody (BMA) on the bottom of the microplates. (c–d) PA images on controlled microplates; (e–h) PA images from treated microplates with different AFP concentrations as labeled. (i) PA signals amplitudes from GNW-CNTs as a function of AFP antigen concentration. (j) PA images from treated microplates with different AFP concentrations as labeled when only GNWs were used as sensor.