#5232 **Extraction and Metabolism of NNK in the Isolated Perfused Lung System**



Laura A. Maertens¹, Stephen S. Hecht², and Cheryl L. Zimmerman^{1,2}

Department of Pharmaceutics¹ and the Cancer Center² University of Minnesota, 308 Harvard St. SE, Minneapolis, MN 55455

Purpose

To validate the use of the recirculating isolated perfused lung system for studying the lung tissue retention of NNK and its metabolites.

Introduction

4-(Methylnitrosamine)-1-(3-pyridyl)-1-butanone (NNK) is formed by nitrosation of nicotine during the smoking and curing processes of tobacco1. NNK is one of the most abundant and potent carcinogens found in cigarette smoke, and requires metabolic activation to elicit its carcinogenic effects^{2,3}. NNK and its major metabolite, 4-(methylnitrosamine)-1-(3-pyridyl)-1-butanol (NNAL), can be metabolized by α-hydroxylation pathways to form keto alcohol, keto acid, hydroxy acid, and diol (Figure 1). The α -hydroxylation metabolic pathways of NNK and NNAL have been shown to result in the formation of DNA adducts. However, NNK and NNAL can also be metabolized to their respective N-oxide metabolites, which are considered detoxification pathways for the elimination of NNK and NNAL3.

NNK induces pulmonary tumors in rodents regardless of the route of administration³. Doses as low as 8.7 µmol/kg have been shown to induce lung tumors, whereas doses of 3 mmol/kg or higher are required to induce the formation of liver tumors⁵. Thus it appears that the lung tissue is selectively sensitive to the carcinogenic effects of NNK. It has been previously reported that the liver clearance (6.9 ± 1.6 ml/min) of NNK is greater than lung clearance (2.1 ± 0.5 ml/min) in rats, and each organ produces a different metabolic profile6. Since in vivo studies of investigating the carcinogenicity effects of NNK on the lung are complicated by liver metabolism, it would be of interest to use the isolated lung perfusion system to investigate NNK metabolism and the retention of its metabolites in the lung.



Figure 1: Metabolic scheme of NNK

Tissue Sample Analysis



Methods

- · Anesthetized rats (Male Fisher 344 rats, 315 ± 15 g) with pentobarbital sodium (60 mg/kg ip)
- · Cannulated trachea (control lung inflation) and pulmonary artery
- · Excised lungs from chest cavity and rinsed of blood
- · Lungs inflated at constant pressure (4 cm H2O) and perfused with 50 ml of Ringers buffer (pH 7.4) at 8 ml/min
- Perfusate oxygenated with 95% O₂ and 5% CO₂, and maintained at 37°C
- 50 µCi bolus dose of H³-NNK administered to perfusate reservoir
- · Perfusate samples drawn at 1, 5, 10, 15, 20, 30, 45, 60, 75, 90, 105, and 120 minutes
- · Following perfusion the lungs were rinsed and flash frozen with liquid nitrogen

Results

Table 1: Metabolites extracted from the tissue (n=4) and perfusate (n=6) following a 120 min perfusion with 50 µCi of 3H-NNK in an isolated lung system. The metabolites are expressed as the mean percent (± standard deviation) of total radioactivity in the sample

	Tissue	Perfusate
	[% of total radioactivity]	[% of total radioactivity]
Hydroxy Acid	9 ± 0.5 %	< LOD*
Keto Acid	21 ± 4 %	12 ± 4 %
NNAL-N-Oxide	24 ± 3 %	5.5 ± 1 %
Diol	7 ± 0.5 %	2 ± 0.7 %
NNK-N-Oxide	10 ± 5 %	48.5 ± 7 %
Keto Alcohol	3 ± 0.8 %	14 ± 4 %
NNAL	5 ± 2 %	5 ± 1 %
NNK	6.5 ± 2 %	4.5 ± 4 %

Table 2: Estimated pharmacokinetic parameters for NNK in an isolated lung system expressed as mean ± standard deviation (n=5)

Clearance (ml/min)	1.63 ± 0.4
Extraction Ratio	0.21 ± 0.05
Terminal Elimination Half-Life (min)	24.8 ± 8.2





Figure 2: NNK and metabolites in the tissue and perfusate following a 120 min perfusion with 50 µCi of 3H-NNK. The metabolites are expressed as the mean percent (± standard deviation) of total radioactivity in the sample

Perfusate Sample Analysis



Conclusions

The perfusate metabolites measured in this study are similar those previously reported6. However, our ability to measure tissue metabolites indicates that perfusate data alone will a give an accurate reflection of metabolic profile and retenti by the lung tissue.

(S)-NNAL has been reported to be as carcinogenic as NNK a we hypothesize that its selective retention in the lung tiss may be the cause7. This study demonstrates the feasibility the use of the isolated perfused lung system to evalu formation and retention of NNK and NNAL metabolites in lung

References

- 1. Hoffmann D, Lavoie EJ, Hecht SS 1985. Nicotine: a precursor for carcinogens. Car Lett 26(1):67-75. Hecht SS, Hoffmann D 1988. Tobacco-specific nitrosamines, an important gr
- 2 carcinogens in tobacco and tobacco smoke. Carcinogenesis 9(6):875-884
- Hecht SS 1998. Biochemistry, biology, and carcinogenicity of tobacco-specific nitrosamines. Chem Res Toxicol 11(6):559-603. Schrader E. Hirsch-Ernst KL Scholz E. Kahl GE. Foth H 2000 Metabolism (Methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK) in primary cultures of
- alveolar type II cells. Drug Metab Dispos 28(2):180-185. Belinsky SA, Foley JF, White CM, Anderson MW, Maronpot RR 1990. Dose-respo relationship between O6-methylguanine formation in Clara cells and inductio pulmonary neoplasia in the rat by 4-(methylnitrosamino)-1-(3-pyridyl)-1-butar
- Cancer Res 50(12):3772-3780 Schrader E, Hirsch-Ernst KI, Richter E, Foth H 1998. Metabolism (methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK) in isolated rat lung and l Naunyn Schmiedebergs Arch Pharmacol 357(3):336-343.
- 7. Wu Z, Upadhyaya P, Carmella SG, Hecht SS, Zimmerman CL 2002. Disposition (methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK) and 4-(methylnitrosamino (3-pyridyl)-1-butanol (NNAL) in bile duct-cannulated rats: stereosele metabolism and tissue distribution. Carcinogenesis 23(1):171-179.

Acknowledgements

The authors wish to acknowledge the financial support of the Public Health Service CA-81301 and NCI/NIDA P50DA13333 grants to SSH) and 3M (3M Science Technology Fellowship to LAM). A special thanks is due to Dr. Douglas Wangen University of Minnesota, for demonstrating the surgery and perfusion setup

View metadata, citation and similar papers at core.ac.uk

