An analytical method for simultaneously assessing biological and physical barrie of the rat intestinal mucosa

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Background

The preclinical development of drug candidates often requires laboratory animals to be orally administered a test compound in a solution or suspension containing a high concentration of excipient. However, recent in vitro studies suggest that commonly used excipients (e.g., polyethylene glycol) may affect intestinal firstpass elimination mechanisms such as P-gp and cytochrome P450 (CYP) (1-4). To evaluate the effects of excipients on rat intestinal P-gp and CYP, an in situ singlepass intestinal perfusion method will be employed using verapamil as a marker of transcellular metabolism/permeation and atenolol as a marker of paracellular permeation

Atenolol (pKa 9.4 (5); log P = 0.16 (6)) and verapamil (pKa 8.9 (7); log P = 3.8 (6)) will be experimentally administered in a 'cocktail' formulation and therefore simultaneous analysis of the resulting plasma and perfusate samples will be necessary. Both molecules are soluble in water or low concentrations of organic solvent (e.g., methanol), however, based on the greater lipophilicity of verapamil (higher logP) we expect different sample preparation (e.g., extraction) requirements from that of atenolol. Other labs have demonstrated reliable sample preparation methods followed by HPLC-UV and HPLC-MS/MS detection methods for either atenolol or verapmil (8-10). However, we are aware of no published method that describes the simultaneous extraction and quantitative measurement of both these drug molecules. Therefore, it is necessary to develop a suitably sensitive, precise and accurate method that allows for the simultaneous measurement of both molecules from the relevant experimental matrices, rat plasma and buffered lumenal perfusate.

Objectives

- To develop a sample preparation and analytical detection method to simultaneously measure atenolol and verapamil along with the N-demethylated metabolite, norverapamil, in rat plasma and aqueous buffered matrices
- To use this method in measuring levels of all three analytes in plasma and lumenal perfusate samples following in situ rat intestinal perfusion of an atenolol-verapamil drug cocktail formulation.

Methods

MS and MS/MS Scans: Each analyte was solubilized in 50% acetonitrile at a concentration of 20 µg/mL. Solutions were infused (5 µL/min) combined with a mobile phase flow path (50:50 acetonitrile:water, containing 0.1% formic acid) into a triple quadrapole mass spectrometer (QuatroMicro, Waters) Instrument settings include: cone voltage = 40-55V, source temperature = 100° C, desolvation temperature = 300-400° C, cone gas = 60 L/hr. MS and MS/MS spectra were collected and product-ions were selected for subsequent multiple reaction monitoring (MRM) analyses

Chromatography: Solutions of atenolol, verapamil, norverapamil (Sigma-Aldrich, St. Louis, MO) and metoprolo (internal standard, Sigma-Aldrich) in mobile phase were injected (20 µL) on a Alliance HPLC (Waters, Milford, MA) and chromatographed on a Zorbax Eclipse XDB-C18 column (2.1 x 50 mm, 3.5 mm, Agilent Technologies) using an isocratic acetontrile elution method. Following the testing of various mobile phase compositions, 30% acetontrile (containing 0.1% v/v formic acid), at 0.4 mL/min, was used in subsequent chromatography.

Sample preparation: Rat plasma or buffered perfusate solution were spiked with 1 µM each of atenolol, verapamil and norverapamil (AVN). Samples (200 μ L) of each spiked matrix were alkalinized with an equal volume of NaOH (5 M), to which 1 mL of extraction solvent was added. Samples were vigorously vortexed (10 min) and centrifuged (1500 rcf, 5 min). Organic solvent was transferred to clean tubes, dried, reconstituted in mobile phase, chromatographed and detected by MRM analyses (atenolol: $267 \rightarrow 145 \text{ m/z}$; verapamil: $455 \rightarrow 165 \text{ m/z}$; norverapamil: $441 \rightarrow 165 \text{ m/z}$; As a measure of extraction efficiencies, absolute peak areas were compared to those of 1 μ M AVN solutions in mobile phase. For subsequent studies matrix standards and experimental gut perfusion samples were extracted using tBME/iso-propanol (4:1) containing metoprolol (265 \Rightarrow) 145 m/z) as an internal standard



(each 20 µM) cocktail formulation.

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