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Circadian Disruption and Metabolic Disease: Findings from Animal Models

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Metabolic comparison of one-anastomosis gastric bypass, single-anastomosis duodenal-switch, Roux-en-Y gastric bypass, and vertical sleeve gastrectomy in rat

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Highlights

- [Diet induced obese](#) rat models of OAGB, SADS, RYGB, and SG are suitable preclinical models. Weight loss and improved [glucose](#) dynamics are similar to those in the clinic.
- OAGB results in metabolic improvements that met or exceed RYGB.
- Each surgical procedure results in a specific metabolic signature – including differential changes in food intake, [insulin](#), GLP-1 response, iron regulation, and fecal microbiota.

Abstract

Background

One-anastomosis gastric bypass (OAGB) and single-anastomosis duodenal switch (SADS) have become increasingly popular weight loss strategies. However, data directly comparing the effectiveness of these procedures with Roux-en-Y gastric bypass (RYGB) and vertical sleeve gastrectomy (SG) are limited.

Objectives

To examine the metabolic outcomes of OAGB, SADS, RYGB, and SG in a controlled rodent model.

Setting

Academic research laboratory, United States.

Methods

Surgeries were performed in diet-induced obese Long-Evans rats, and metabolic outcomes were monitored before and for 15 weeks after surgery.

Results

All bariatric procedures induced weight loss compared with sham that lasted throughout the course of the study. The highest percent fat loss occurred after OAGB and RYGB. All bariatric procedures had improved glucose dynamics associated with an increase in insulin (notably OAGB and SADS) and/or glucagon-like protein-1 secretion. Circulating cholesterol was reduced in OAGB, SG, and RYGB. OAGB and SG additionally decreased circulating triglycerides. Liver triglycerides were most profoundly reduced after OAGB and RYGB. Circulating iron levels were decreased in all surgical groups, associated with a decreased hematocrit value and increased reticulocyte count. The fecal microbiome communities of

OAGB, SADS, and RYGB were significantly altered; however, SG exhibited no change in microbiome diversity or composition.

Conclusions

These data support the use of the rat for modeling bariatric surgical procedures and highlight the ability of the OAGB to meet or exceed the metabolic improvements of RYGB. These data point to the likelihood that each surgery accomplishes metabolic improvements through both overlapping and distinct mechanisms and warrants further research.

Keywords

One-anastomosis gastric bypass; Single-anastomosis duodenal switch; Roux-en-Y gastric bypass; Sleeve gastrectomy; Rodent; Animal model; Glucose regulation

[Roux-en-Y](#) gastric bypass (RYGB) and [sleeve gastrectomy](#) (SG) are the most commonly performed [bariatric](#) surgical procedures¹ and result in significant and sustained weight loss accompanied by dramatic changes in [glucose](#) metabolism.^{2,3} However, surgeons continue to explore other procedures that may carry advantages in terms of [surgical approach](#) or metabolic improvements. Among these procedures are the one-anastomosis gastric bypass (OAGB), also known as the mini bypass or the omega loop gastric bypass, and the single-anastomosis [duodenal switch](#) (SADS), both of which produce significant improvements in weight and glucose homeostasis.^{4,5}

Surgeons drawing observations from patients have dominated the development of these [bariatric](#) procedures. Nonetheless, the utilization of animal models is an important step in pinpointing the surgical effects on metabolism independent of genetic background, diet adherence, and environmental conditions. By allowing investigators to test a wide range of hypotheses while limiting variability compared with the human population, animal models enable the identification of mechanistic hypotheses to explain the potent metabolic effects of [bariatric surgery](#). Of note, the use of animal models in this study confers the following 2 important benefits over clinical studies: (1) when possible, we standardize gut manipulations to draw meaningful comparisons on how specific changes to gut physiology impact metabolism; and (2) as animal models do not follow clinician advice on diet and/or exercise, any observed outcomes are due entirely to the surgical procedure and its effect on the brain and overall physiology. An important validity measure of these animal models depends on the similarity in postsurgical responses compared with the patient population at the level of weight maintenance and metabolic outcome.

Indeed, the mechanisms by which [bariatric](#) surgical procedures result in weight loss and metabolic improvements are still not completely understood. A number of hypotheses have been forwarded to explain these metabolic improvements, including the involvement of glucose-regulating gut hormones, such as glucagon-like protein-1 (GLP-1),^{6,7} [bile acids](#),^{8,9} and alterations in the microbiome.^{10,11} By comparing different bariatric surgical procedures in obese rats, we can systemically investigate the physiologic components correlated with metabolic success and how they relate to specific physical changes of the surgery. Moreover, we can identify what surgical intervention may be more effective for a specific metabolic outcome in a controlled, nonvariable setting. Here, we present evidence that our rodent procedure similar to OAGB meets or exceeds the metabolic improvements of the comparative

gold standard bariatric surgical procedure, the RYGB. Moreover, each bariatric surgical procedure leads to differential effects on metabolism, and when viewed in light of the [surgical approach](#), suggests that a combination of overlapping and distinct mechanisms may contribute to the general success of [bariatric surgery](#).

Methods

Animals

Male Long-Evans rats (Envigo, Indianapolis, IN, USA) were maintained on a high-fat diet with butter fat (40% fat, 4.54 kcal/g; D03082706 Research Diets, New Brunswick, NJ, USA) to induce obesity before [bariatric surgery](#). Before surgery, all rats received baseline measurement of weight and body composition. After surgery, rats were maintained on the same high-fat diet. Weight, food intake, and body composition were measured periodically over 15 weeks. The number of animals per group that survived until the end of the study were as follows: sham (7/9), RYGB (7/13), OAGB (6/13), SG (12/13), and SADS (9/13). Rats were excluded from analysis for failure to recover from the surgery or for exhibiting signs of [intestinal obstruction](#) or infection. Rats were individually housed in temperature-controlled rooms with a 12:12-hour light cycle. All studies were approved by and performed according to the guidelines of the Institutional Animal Care and Use Committee of the University of Michigan.

Surgery

After induction of [isoflurane anesthesia, animals](#) received [buprenorphine](#) hydrochloride (.03 mg/kg), meloxicam (.5 mg/kg), gentamicin (8 mg/kg), and 10 mL isotonic saline, all by [subcutaneous injection](#). All surgeries began with a midline abdominal skin incision, followed by an incision in the underlying muscle wall. All [anastomoses](#) were performed with 8-0 [Prolene sutures](#) (Ethicon, Somerville, NJ, USA) and abdominal closures were performed in 2 layers with 4-0 Vicryl Rapide sutures (Ethicon). Postoperatively, animals received a liquid diet (Osmolite1 Cal, Abbott Nutrition, Abbott Laboratories; Columbus, OH, USA) for 4 days before being returned to the previously described high-fat diet. All animals received subcutaneous injection of buprenorphine hydrochloride twice daily and meloxicam and gentamicin once daily for 3 days.

Sham surgery

The stomach was exposed, gentle pressure applied with forceps, and the stomach was covered with a gauze pad soaked in sterile saline. The stomach was returned to the body cavity and the incision closed.

Roux-en-Y gastric bypass

The stomach and the intestines were exposed and draped on a gauze pad soaked in sterile saline. A triangular stomach pouch of reduced volume was created via 2 intersecting cuts using an [endoscopic stapler](#) (Endopath ETS-Flex 35 mm Endoscopic Articulating Linear Cutter; Ethicon [Endo-Surgery Inc.](#), Cincinnati, OH, USA) to separate the pouch from the remnant stomach. The [jejunum](#) was transected 30 cm distal to the [ligament of Treitz](#), and a gastrotomy incision was made in the stomach pouch. The transected end of the distal portion of the jejunum was anastomosed to the gastrotomy (forming the Roux limb) with interrupted sutures. The remaining transected end of the proximal portion of the intestine was then anastomosed with interrupted sutures to the Roux limb as an end-to-side jejuno-jejunosomy at 10 cm from the gastrojejunosomy.

One-anastomosis gastric bypass

The stomach and the intestines were exposed and draped on a gauze pad soaked in sterile saline. A stomach pouch of reduced volume was created as described for RYGB. The jejunum at 30 cm from the ligament of Treitz was brought up to contact the stomach pouch, and incisions of comparable length were made in both. The jejunum was anastomosed to the stomach pouch with interrupted sutures. Note, we have selected a limb length similar to our RYGB procedure to make meaningful comparisons about the effect of a single anastomosis on metabolic outcomes. Thus, this method deviates from clinical OAGB in that OAGB procedures traditionally have a longer limb length than that of RYGB procedures.

Sleeve gastrectomy

The stomach was exposed and transected to form a sleeve using an endoscopic stapler (ETS-Flex 35; Ethicon). Approximately 70% of the rat stomach (by weight) was removed, including the majority of the [fundus](#) and about 50% of the pyloric antrum.

Single-anastomosis duodenal switch

The stomach was exposed and covered with a gauze pad soaked in sterile saline. The [duodenum](#) was identified and transected proximally to where the [common bile duct](#) enters; the free end of the distal duodenum was sutured. The gauze was removed from the stomach, which was transected to form a sleeve as described for SG. The jejunum (at 30 cm from the ligament of Treitz) was mobilized to contact the proximal duodenal free end and an incision was made in the antimesenteric side of the former to match its length to the duodenal diameter. The jejunum was connected to the proximal duodenal segment by side-to-end anastomosis with interrupted sutures. As with the OAGB procedure, we have selected a limb length similar to our RYGB procedure to make meaningful comparisons about the effect of intestinal rerouting in the context of a reduced stomach size on metabolic outcomes. Thus, this method deviates from clinical SADS in that SADS procedures traditionally have a much longer limb length than that of RYGB procedures.

Metabolic outcome methods

Detailed methods on metabolic outcome measures, including body composition, mixed-meal [glucose tolerance tests](#), total GLP-1 measures, lipid analysis, iron regulation, and fecal microbiome can be found in supplementary methods.

Statistical analysis

Statistical analysis was performed using GraphPad Prism (GraphPad, La Jolla, USA). Unless otherwise specified, we used one-way analysis of variance and Tukey [post hoc tests](#) where necessary to determine significant differences between groups. We used a repeated-measures analysis of variance for analysis of weight and food intake over time with a Bonferroni's multiple comparison test when a significant interaction effect was found. All statistical analysis used a 2-tailed design and results were considered significant when $P < .05$.

Results

Surgical effect on body mass and body composition

In comparison to the control sham surgery, all [bariatric](#) surgical groups lost weight and maintained a lower weight for the duration of the 15-week study (106 d). The RYGB and OAGB groups lost the most weight, whereas SG and SADS groups lost an intermediate amount of weight ([Figs. 1A, 1B](#)). [Post hoc analysis](#) on total weight loss revealed that all groups lost more weight than sham ($P < .001$), and RYGB lost significantly more weight than SG ($P < .05$).

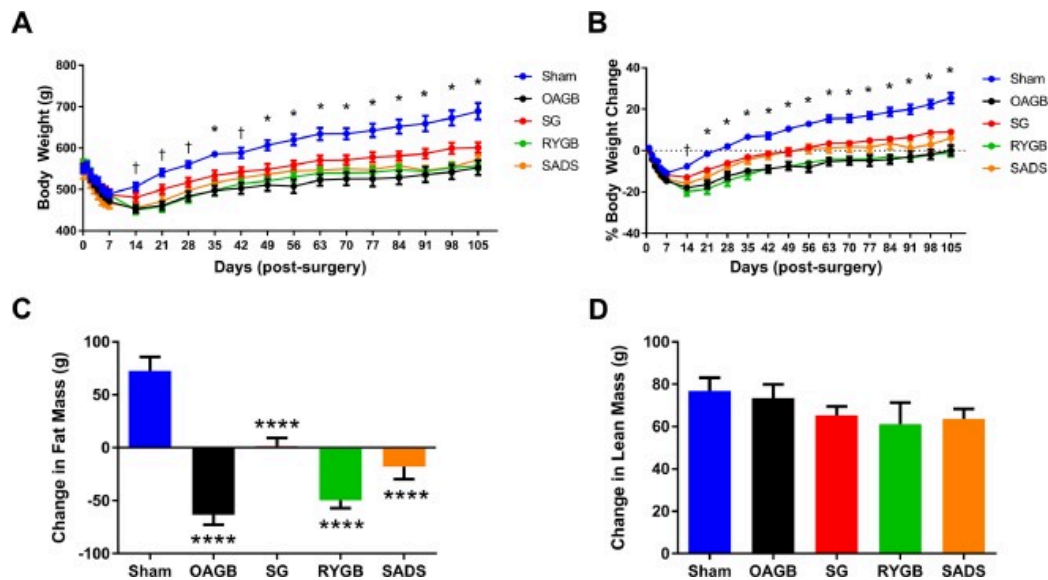


Fig. 1. Surgical effect on body mass and body composition. (A) Absolute weight changes (in g) after surgery. *All groups differ from sham; †one-anastomosis gastric bypass, [Roux-en-Y](#) gastric bypass, and single-anastomosis [duodenal switch](#) differ from sham. (B) Percent change from baseline weight after surgery. *All groups differ from sham; †one-anastomosis gastric bypass, Roux-en-Y gastric bypass, and single-anastomosis duodenal switch differ from sham. (C) Absolute change in fat mass 15 weeks postsurgery. ****Significant difference from Sham, $P < .0001$. (D) Absolute change in lean mass 15 weeks postsurgery.

Body fat composition was similarly altered 15 weeks after surgery, with all groups losing a significant amount of fat mass compared with sham ([Fig. 1C](#)). The largest amount of fat loss occurred after OAGB and RYGB, with SADS and SG exhibiting an intermediate amount of fat loss ([Fig. 1C](#)). Post hoc analysis revealed that OAGB lost significantly more fat mass than SG ($P < .001$) and SADS ($P < .05$). RYGB lost significantly more fat mass than SG ($P < .01$).

Surgery did not affect change in lean mass ([Fig. 1D](#)), and there were no significant differences in baseline fat or lean mass (Supplemental Fig. 1).

Surgical effect on food intake

Rats were maintained on a high-fat diet before and after surgery. Cumulative food intake, as measured from surgical recovery to 14 weeks (98 d) postsurgery, was reduced after the OAGB and SG compared with sham ([Fig. 2A](#)). Cumulative food intake was not reduced in RYGB or SADS compared with sham-operated animals ([Fig. 2A](#)). All surgical groups exhibited a transient decrease in food intake 2 weeks (SG, $P < .001$; OAGB, RYGB, SADS, $P < .0001$) and 3 weeks (SADS, $P < .01$; SG, $P < .001$; OAGB, RYGB, $P < .0001$)

after surgery compared with sham-operated controls (Fig. 2B). Notably, the OAGB and SG groups, which also had significant reductions in cumulative intake, exhibited longer transient reductions in food intake of approximately 6 weeks (42 d) in duration.

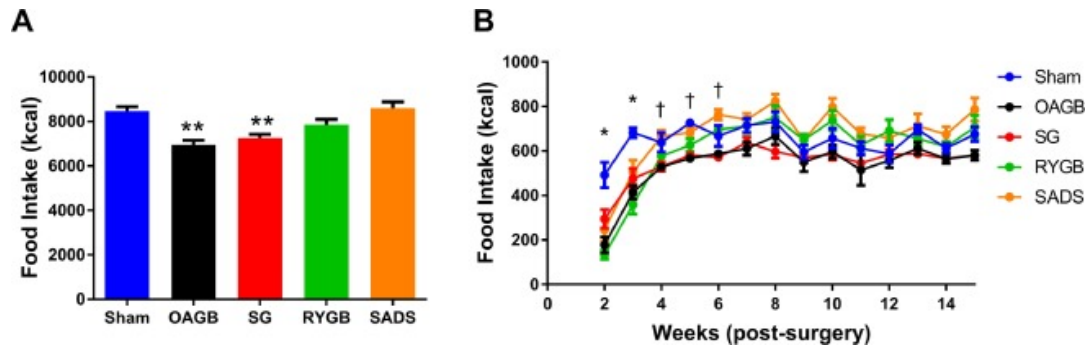


Fig. 2. Surgical effect on food intake. (A) Cumulative [caloric intake](#) from surgical recovery to 14 weeks postsurgery. **Significant difference from sham, $P < .01$ (B) Weekly caloric intakes of each surgical group; *All groups differ from sham; †one-anastomosis gastric bypass and [sleeve gastrectomy](#) differ from sham.

Surgical effect on glucose regulation

Fasting [glucose](#) levels were only decreased in the SG compared with sham controls (Fig. 3A). In response to an oral mixed-meal [glucose tolerance test](#), all surgical groups had improved glucose clearance and lower glucose levels at 30, 45, and 60 minutes after the oral load compared with sham-operated controls (Fig. 3B). The rise in blood glucose (e.g., the difference between the 15- min time point and the baseline) was significantly greater in OAGB ($P < .01$), RYGB ($P < .05$), and SADS ($P < .05$) compared with the glucose rise experienced by sham-operated controls (Fig. 3B).

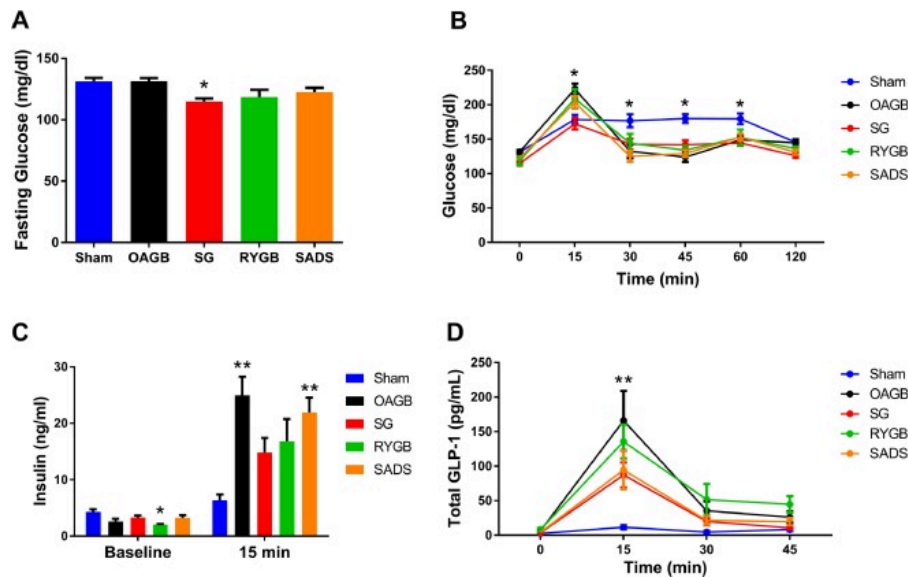


Fig. 3. Surgical effect on [glucose](#) regulation. (A) [Fasting blood glucose](#) levels. *Significant difference from sham, $P < .05$. (B) [Glucose response](#) to an oral mixed-meal tolerance test. *Significant difference from sham, $P < .05$. (C) Baseline/fasting plasma [insulin](#) levels and plasma insulin 15 minutes after an oral mixed meal. *Significant difference from sham at same time point, $P < .05$, ** $P < .01$, **** $P < .0001$. (D) Total glucagon-like protein-1 response to an oral mixed-meal tolerance test. *All groups significant difference from sham, $P < .05$.

Baseline [insulin](#) levels were only decreased in the RYGB compared with sham controls ([Fig. 3C](#)). Although all surgical groups exhibited an increase in insulin 15 minutes after an oral mixed-meal load compared with baseline, only OAGB and SADS had a glucose-stimulated [insulin response](#) greater than sham ([Fig. 3C](#)). Using a repeated-measures analysis of variance, total GLP-1 in plasma was significantly increased 15 minutes after an oral mixed-meal in all surgical groups compared with sham ($P < .01$; [Fig. 3D](#)). Moreover, post hoc analysis with Sidak's multiple comparison test revealed OAGB had increased total GLP-1 compared with SG ($P < .01$) and SADS ($P < .01$) at 15 minutes postgavage.

Surgical effect on lipid regulation

Circulating ad libitum plasma levels of cholesterol were significantly reduced in rats undergoing OAGB, SG, and RYGB compared with sham ([Fig. 4A](#)). No differences were observed in circulating levels of nonesterified fatty acids ([Fig. 4B](#)). Plasma triglyceride levels were reduced after OAGB and SG compared with sham ([Fig. 4C](#)). While there were no significant reductions in plasma triglycerides in the RYGB or SADS groups, there was a significant reduction in liver triglycerides ([Figs. 4C, 4D](#)). Indeed, liver triglycerides were significantly reduced in all bariatric surgical groups compared with sham ([Fig. 4D](#)). Moreover, post hoc analysis revealed that OAGB ($P < .01$), RYGB ($P < .001$), and SADS ($P < .05$) had lower liver triglycerides compared with SG.

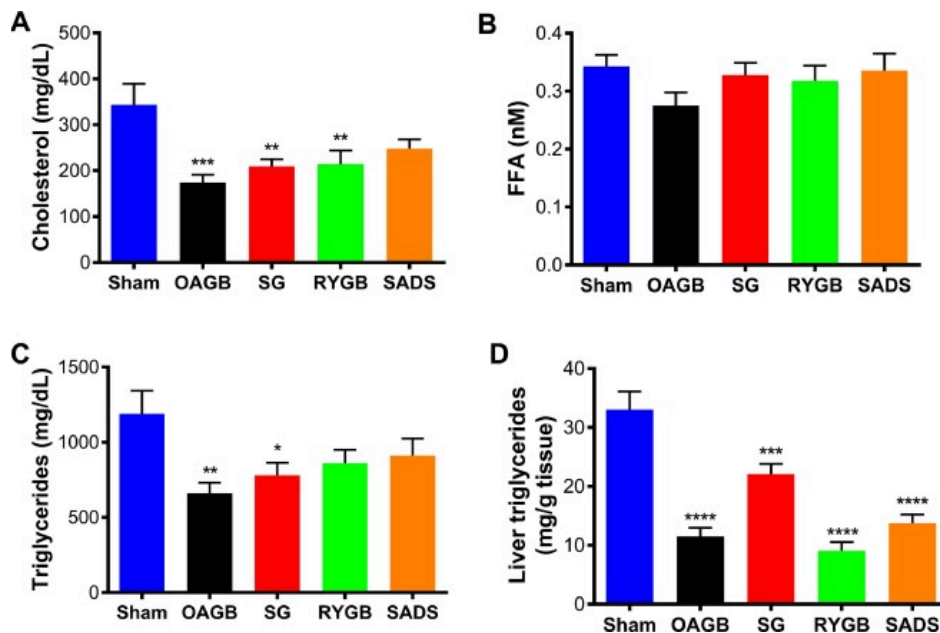


Fig. 4. Surgical effect on lipid regulation. (A) Plasma cholesterol levels. **Significant difference from sham, $P < .01$. (B) Plasma [free fatty acids](#) (FFA). (C) Plasma triglycerides. *Significant difference from sham, $P < .05$, ** $P < .01$. (D) Liver triglycerides expressed as milligram per gram of liver tissue. ***Significant difference from sham, $P < .001$, **** $P < .0001$.

Surgical effect on iron regulation

Compared with sham-operated controls, total circulating [iron levels](#) were reduced in all surgery groups except for RYGB ([Fig. 5A](#)). [Hematocrit](#) was reduced in rats undergoing SADS, RYGB, and OAGB compared with sham ([Fig. 5B](#)). Moreover, post hoc analysis revealed hematocrit of OAGB was significantly reduced compared with SG ($P > .001$). Although a trend was observed for an increase in reticulocyte count in all

groups, only SADS- and SG-operated rats had a significantly elevated reticulocyte count compared with sham (Fig. 5C).

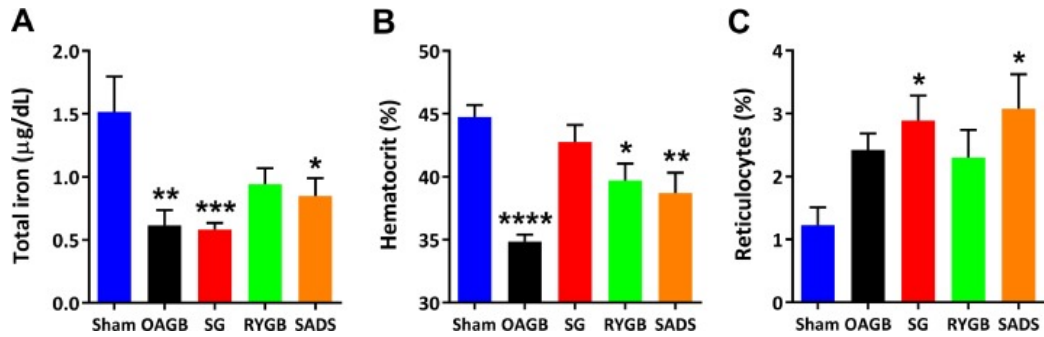


Fig. 5. Surgical effect on iron regulation. (A) Total circulating [iron levels](#). *Significant difference from sham, $P < .05$, ** $P < .01$, *** $P < .001$. (B) [Hematocrit](#) levels, *Significant difference from sham, $P < .05$, ** $P < .01$, **** $P < .0001$. (C) Reticulocyte count. *Significant difference from sham, $P < .05$.

Surgical effect on the fecal microbiome

θ_{VC} was used to determine the ability of the surgery to procedure distinct microbiome communities. θ_{VC} distances were significantly large for all surgical comparisons with sham except SG versus sham (Wilcoxon test $P < .05$), pointing to distinct microbiome communities occurring within all surgeries except SG (Fig. 6A, Table 1). Lack of significance in the SG group may be related to the variability in postsurgical weight. Specifically, axis 2 of θ_{VC} significantly predicted 15-week postsurgical weight within the SG group ($R^2 = .35$, $F_{1,10} = 5.32$, $P < .05$). In particular, 8 [operational taxonomic units](#) (OTUs) with an linear discriminant analysis (LDA) score >4 were identified as differently abundant across the surgical groups (Table 2). However, no significant differences were observed in microbiome diversity or richness as calculated using the inverse Simpson measure (Fig. 6B).

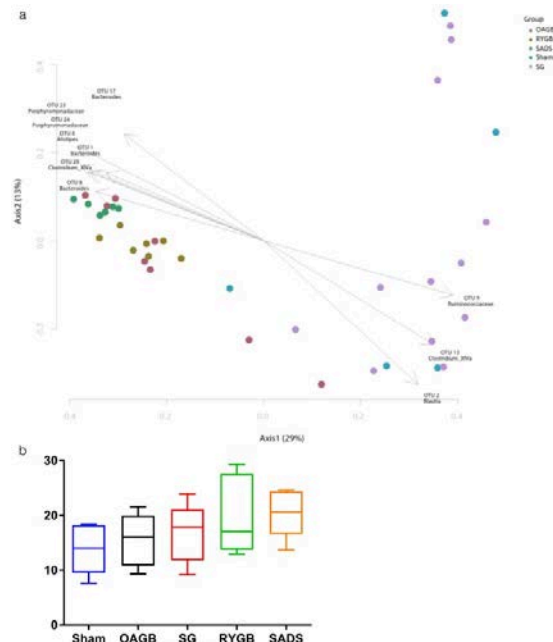


Fig. 6. Surgical effect on the fecal microbiome. (A) There were significant differences in the microbiota of most surgical groups, as calculated by measuring the distance between communities (θ_{VC}). Only [sleeve gastrectomy](#)

versus sham was found to be similar in microbiome population. (B) No significant differences were observed in microbiome diversity as calculated using the inverse Simpson measure.

Table 1. Surgical effect on the fecal microbiome population

	OAGB	SG	RYGB	SADS
Sham	<.001	.216	.002	.001
OAGB		<.001	.046	.029
SG			<.001	<.001
RYGB				.013

OAGB = one-anastomosis gastric bypass; SADS = single-anastomosis [duodenal switch](#); RYGB = [Roux-en-Y](#) gastric bypass; SG = [sleeve gastrectomy](#) (vertical).

P value comparison of θ_{vc} values across surgical groups.

Table 2. Surgical effect on the fecal microbiome population

LDA	P value	Group	Taxonomy
4.4893	1.01 ^{E-05}	RYGB	Bacteria; bacteroidetes; bacteroidia; bacteroidales; rikenellaceae; alistipes
4.55226	2.46 ^{E-06}	SG	Bacteria; firmicutes; clostridia; clostridiales; ruminococcaceae
4.35383	2.23 ^{E-05}	Sham	Bacteria; firmicutes; clostridia; clostridiales; lachnospiraceae; clostridium_XIVa
4.33306	4.59 ^{E-05}	SADS	Bacteria; firmicutes; clostridia; clostridiales; lachnospiraceae; clostridium_XIVa
4.19178	1.35 ^{E-05}	SADS	Bacteria; firmicutes; clostridia; clostridiales; lachnospiraceae; clostridium_XIVa
4.27848	6.72 ^{E-06}	SADS	Bacteria; bacteroidetes; bacteroidia; bacteroidales; porphyromonadaceae
4.05671	2.63 ^{E-05}	OAGB	Bacteria; bacteroidetes; bacteroidia; bacteroidales; porphyromonadaceae; barnesiella
4.26218	1.70 ^{E-06}	Sham	Bacteria; firmicutes; clostridia; clostridiales; lachnospiraceae; blautia

RYGB = [Roux-en-Y](#) gastric bypass; SG = [sleeve gastrectomy](#) (vertical); SADS = single-anastomosis [duodenal switch](#); OAGB = one-anastomosis gastric bypass; OTU = [operational taxonomic unit](#); LDA = linear discriminant analysis..

Differently abundant OTUs across surgical groups.

Discussion

Echoing the observation in humans, we found that rat models of [bariatric](#) surgical procedures led to significant and sustained weight loss and improvements in [glucose](#) metabolism. However, as summarized in [Fig. 7](#), SG, SADS, RYGB, and OAGB incorporated different [surgical approaches](#) and resulted in differential effects on metabolic outcomes. Using this controlled animal model, we observed that OAGB met or exceeded the metabolic outcomes of RYGB, including increased weight loss, lipid reduction, and increased glucose-stimulated [insulin](#) and total GLP-1 release.

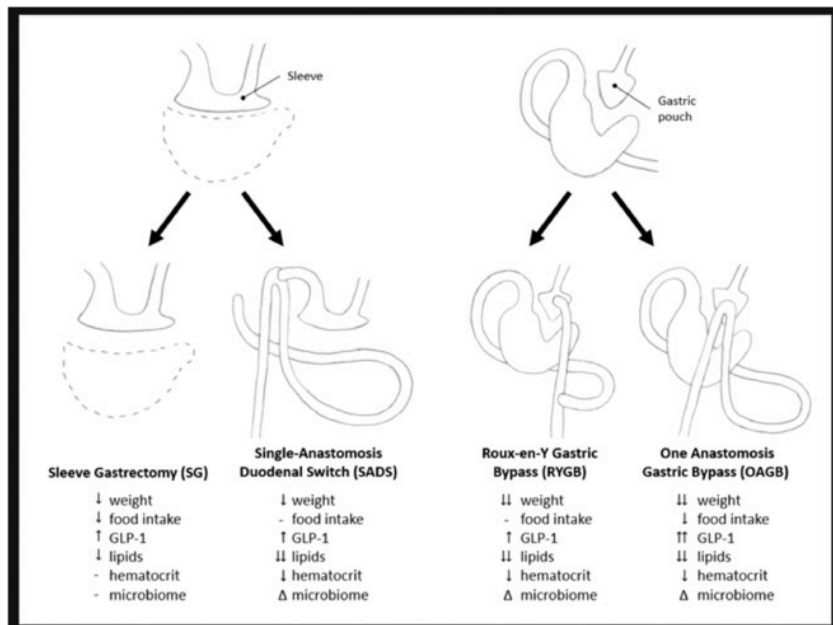


Fig. 7. Summary of surgical procedures and their major effects on metabolism.

We found that weight loss was significantly reduced in all surgical groups, with the most robust reductions in fat mass occurring in the OAGB and RYGB groups. Although a combined, clinical [meta-analysis](#) including all surgical procedures performed in this study is not available, separate clinical reports are available for comparison. In general, clinical and [randomized controlled trials](#) have indicated similar weight loss between RYGB and SG.^{12,13} However, some report greater fat mass loss in RYGB compared with SG,¹⁴ in agreement with our results. A randomized control trial found that OAGB produced weight loss comparative to SG,¹⁵ although OAGB was associated with better [glycemic](#) control.^{15,16} A recent report on the weight outcomes of SADS versus RYGB indicated no differences in weight loss or [diabetes](#) remission.¹⁷

OAGB and SG groups had decreased cumulative food intake compared with sham over the course of the experiment, contributing to weight loss. Although transient reductions in food intake were observed in all surgical groups, RYGB and SADS did not have any cumulative reduction in food intake, suggesting that these groups may experience an increase in [energy expenditure](#) or a decrease in [nutrient absorption](#) to account for the observed weight loss.

Notably, both OAGB and RYGB exhibit the most robust weight loss and leave the dissected stomach tissue in the body. Retaining the stomach tissue potentially allows gut hormones to be produced and pass readily into the circulation. OAGB also had the highest increases in GLP-1 levels in response to an

oral glucose load. GLP-1 has robust actions on glucose regulation¹⁸ and higher levels are associated with lowered glucose and insulin levels and improved metabolic health.¹⁹ Although improvement in [fasting blood glucose](#) was observed only in SG rats, glucose-induced insulin secretion was dramatically improved in all surgical groups.

An improved lipid profile was observed in all surgical groups, with the most profound decrease in plasma cholesterol and triglycerides observed in OAGB and SG, which could be attributed to the lower food intake also observed in these groups. Lower liver triglycerides reflect improved metabolism and lipid processing.^{20,21} Although all surgical groups experienced a significant decrease in liver triglycerides, the most profound decreases in liver triglycerides occurred in surgical groups where nutrients were rerouted from the original path (e.g., RYGB, OAGB, and SADS). These data are consistent with the clinical data that, in general, indicate that RYGB reduces triglycerides more profoundly than SG.^{22,23} Comparison studies examining triglyceride levels in OAGB and SADS are limited. One clinical study found that RYGB, OAGB, and SADS had similarly reduced triglyceride levels, consistent with the present data; however, there was no SG control to determine whether triglycerides were more robustly decreased in RYGB, OAGB, and SADS compared with SG.¹⁷ However, 2 recent studies comparing OAGB to SG found no significant differences in triglycerides.^{16,23} When comparing preclinical rodent data with human clinical data, it is important to point out that clinical studies use plasma triglycerides, and in the present study, we found that plasma triglycerides were only significantly reduced compared with sham after OAGB and SG. Therefore, in the rat model, liver triglycerides may be a more appropriate indicator of clinical [lipid metabolism](#) postsurgery.

[Iron deficiency](#) and related [anemia](#) are common phenomena after [bariatric surgery](#) and has primarily been attributed to bypassing the [duodenum](#).^{24,25,26,27} However, we find that except for RYGB, all surgical interventions reduced circulating [iron levels](#). To maintain sufficient circulating iron levels, [red blood cells](#) are degraded by the [macrophages](#) to free hemoglobin-bound iron. Accordingly, [hematocrit](#) levels are decreased compared with sham in all bariatric surgical groups except for SG. An increased reticulocyte count is indicative of increased [erythropoiesis](#) at the level of the bone marrow and, combined with a reduced hematocrit level, is indicative of an increased erythrocyte turnover rate. Altogether, these data suggest that the low hematocrit levels observed in OAGB, SG, and SADS are caused by an increase of erythrocyte degradation to compensate for disrupted intestinal iron [absorption](#).²⁸ These rodent data are consistent with the clinical data in which iron deficiency and related anemia are common phenomena and more prevalent after duodenal bypass interventions than after SG. Although not directly assessed in the present study, these data are complementary to the nutritional [malabsorption](#) observed after clinical OAGB and SADS procedures.^{5,29} Taken together, malabsorption is an important variable to consider when evaluating the metabolic outcome of a bariatric surgical procedure.

The fecal microbiome was significantly altered in OAGB, SADS, and RYGB surgical groups. These data point to the impact of rerouting nutrients and their effect on the microbiome independent of stomach tissue presence, [incretins](#), the magnitude of weight loss, or changes in food intake. Indeed, a number of studies have shown a shift in the microbiota after RYGB.^{10,30,31} Similarly, others have shown a significant increase in Bacteroidetes population, which is linked with decreased fat mass and [leptin](#) levels^{8,31,32} after SG. However, in this data set, we did not observe a significant difference in the microbiome of rats after SG compared with sham. Part of this may be due to high weight variability in the SG rats as a regression analysis indicated that axis 2 of θ_{VC} significantly predicted 15-week postsurgical weight in the SG group.

In the present study, we observe higher death rates after RYGB, OAGB, and SADS procedures compared with SG or sham, which is a potential limitation in our interpretations and conclusions. We suspect the loss to be due to the technical difficulty of performing the [anastomosis](#) technique in the relatively small rat model. Nevertheless, we have previously found that 6 to 7 animals per group is sufficient for robust metabolic analysis for animal studies of bariatric surgical procedures³³ and that the retention rate from the present study is in line with our previous observations.

In extrapolating our results to clinical practice, one key limitation of the present study is our surgical procedures for the OAGB and SADS do not exactly mirror those performed in humans. While we endeavored to closely recapitulate the clinical operations, we elected to standardize limb length across the RYGB, OAGB, and SADS procedures, deviating from clinical practice. This standardization is important as it allows us to draw conclusions about how a specific manipulation to the gut, such as a single anastomosis (e.g., OAGB) and/or rerouting of nutrients in the context of a sleeve (e.g., SADS), affects metabolism without the added confounding variable of limb length. While limb length may be a contributing factor to metabolic outcomes, previous research has not found limb length to be a factor in weight loss obtained after RYGB.³⁴ Therefore, while these data cannot be directly applied to humans due to the methodic deviation from clinical OAGB and SADS, these data increase our understanding of how systemic changes to the gut can alter metabolism, which has marked clinical implications.

Conclusions

The results from these studies reveal that OAGB, SADS, RYGB, and SG surgical procedures have overlapping effects on a variety of parameters, including body fat, [glucose](#) regulation, [insulin](#) secretion, GLP-1 secretion, iron regulation, and [lipid metabolism](#) (Fig. 7). However, there remain important differences among these procedures that may contribute to different clinical effects. The ability to perform carefully controlled studies to directly compare the effects of multiple procedures is a major advantage of rodent models. The present data demonstrate that OAGB meets or exceeds the metabolic improvements induced by RYGB.

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Disclosures

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Appendix. Supplementary materials

<https://ars.els-cdn.com/content/image/1-s2.0-S155072891830532X-mmc1.docx>

<https://ars.els-cdn.com/content/image/1-s2.0-S155072891830532X-mmc2.docx>

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