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Structural, Spectroscopic, and Electrochemical Properties of Nonheme Fe(II)-Hydroquinonate Complexes: Synthetic Models of Hydroquinone Dioxygenases

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Abstract:

Using the tris(3,5-diphenylpyrazol-1-yl)borate (Ph2Tp) supporting ligand, a series of mono- and dinuclear ferrous complexes containing hydroquinonate (HQate) ligands have been prepared and structurally characterized with X-ray crystallography. The monoiron(II) complexes serve as faithful mimics of the substrate-bound form of hydroquinone dioxygenases (HQDOs) – a family of nonheme Fe enzymes that catalyze the oxidative cleavage of 1,4-dihydroxybenzene units. Reflecting the variety of HQDO substrates, the synthetic complexes feature both mono- and bidentate HQate ligands. The bidentate HQates cleanly provide five-coordinate, high-spin Fe(II) complexes with the general formula $[Fe(^{Ph2}Tp)(HL^{X})]$ (**1X**), where HL^X is a HQate(1-) ligand substituted at the 2-position with a benzimidazolyl (1A), acetyl (1B and 1C), or methoxy (1D) group. In contrast, the monodentate ligand 2,6-dimethylhydroquinone (H_2L^F) exhibited a greater tendency to bridge between two Fe(II) centers, resulting in formation of $[Fe_2(Ph_2Tp)_2(\mu -$ L^F)(MeCN)] [2F(MeCN)]. However, addition of one equivalent of "free" pyrazole (Ph2pz) ligand provided the mononuclear complex, $[Fe(^{Ph2}Tp)(HL^{F})(^{Ph2}pz)]$ [**1F**(^{Ph2}pz)], which is stabilized by an intramolecular hydrogen bond between the HL^{F} and Ph2pz donors. Complex **1**F(Ph2pz)represents the first crystallographically-characterized example of a monoiron complex bound to an untethered HQate ligand. The geometric and electronic structures of the Fe/HQate complexes were further probed with spectroscopic (UV-vis absorption, ¹H NMR) and electrochemical methods. Cyclic voltammograms of complexes in the 1X series revealed an Fe-based oxidation between 0 and -300 mV (vs. Fc^{+/0}), in addition to irreversible oxidation(s) of the HQate ligand at higher potentials. The one-electron oxidized species (1Xox) were examined with UV-vis absorption and electron paramagnetic resonance (EPR) spectroscopies.

Introduction

The degradation of single- and multi-ring aromatic hydrocarbons by bacteria is a key component of the global carbon cycle and the basis of bioremediation technologies. In aerobic environments, the catabolism of aromatic compounds is dependent on nonheme iron dioxygenases that cleave aromatic rings with incorporation of both atoms of O_2 into the product.¹ Such transformations are challenging due to the intrinsic stability of aromatic systems and the high activation barrier to reaction with triplet dioxygen. With the notable exception of the intradiol catechol dioxygenases, the active sites of ring-cleaving dioxygenases overcome these obstacles by coordinating

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both substrate and O_2 to a single Fe(II) site.² The iron center is typically attached to the protein by a facial array of one carboxylate (Asp or Glu) and two His residues (Scheme 1),³ although variants of this 2-His-1-carboxylate motif have recently been reported.⁴



Scheme 1

While the well-studied extradiol catechol dioxygenases (ECDOs) are the prototypical ring-cleaving dioxygenases,^{2,5} members of this enzymatic family employ a remarkable variety of substrates, including protocatechuates,⁶ 2-aminophenols,⁷ and salicylates.⁸ Of particular relevance to this manuscript are dioxygenases that cleave hydroquinones (HQs = 1,4-dihydroxybenzene and its derivatives). The HQ-cleaving dioxygenases (HQDOs) can be grouped into two categories. The first class oxidizes substrates with carboxylate groups at the 2-position of the aromatic ring, namely, gentisate⁹ and homogentisate¹⁰ (2,5-dihydroxybenzoate and 2,5-dihydroxyphenylacetate, respectively; Scheme 1). In these enzymes, the substrate likely binds to iron in a bidentate manner via the

phenolate and carboxylate donors.¹¹ In the second class, the substrate coordinates in a monodentate fashion, since the halogenated or unsubstituted HQs lack a metal-binding moiety at the *ortho* position. Examples include 2,6-dichlorohydroquinone 1,2-dioxygenase (PcpA),¹² chlorohydroquinone dioxygenase (LinE),¹³ and hydroquinone 1,2-dioxygenase (MnpC).¹⁴

While there have been few mechanistic studies of the hydroguinone 1,2-dioxygenases, the proposed catalytic cycles largely follow the pattern derived from extensive studies of the ECDOs.^{12a} Coordination of the deprotonated HQ substrate to the Fe(II) center displaces some or all of the H_2O ligands found in the resting state (Scheme 1), thereby facilitating O_2 binding to the iron center. Formation of a short-lived ferric-superoxo intermediate is thought to trigger the transfer of one electron from the substrate ligand to iron, resulting in a bound *p*-benzosemiquinone radical. The existence of this putative intermediate would likely require deprotonation of the distal -OH group by a second-sphere residue, although it is not clear whether these three events (O_2 coordination, electron transfer, and proton transfer) occur in a stepwise or concerted manner. The degree of semiquinone character on the substrate ligand in the O₂-bound form of the enzyme is also uncertain; for instance, a recent computational study by Ye and Neese¹⁵ has cast doubt on the existence of a superoxo-Fe(II)-semiguinone intermediate in the ECDO (and, by extension, the HQDO) mechanism. While the nature of this intermediate remains disputed, it is well-established that the next step of the catalytic cycle involves generation of an Fe(II)-alkylperoxo species, which undergoes a Criegee rearrangement and hydrolysis to eventually yield the ring-opened product.¹⁵⁻¹⁶

Unanswered questions regarding the HQDOs can be answered, in part, through the development of synthetic complexes that replicate the structure and/or function of the enzyme active site. Remarkably, a survey of the literature found only a single example of a crystallographically-characterized monoiron(II)-hydroquinonate complex: Fe(L)₂, where is L is a deprotonated Schiff base of 2,5dihydroxybenzaldehyde.¹⁷ The dearth of reported Fe/HQ complexes is partly due to the ability of hydroquinonate (HQate) ligands to adopt a bridging position between metal centers, as demonstrated by structures of diiron(III)-porphyrin and -salen complexes with bridging

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HQate dianions.^{18,19} Recently, Machonkin and Holland described the formation and ¹H NMR characterization of a mononuclear iron(II)-2methylhydroquinonate complex supported by the 1,3,5tris(tolylideneimino)cyclohexane ligand;²⁰ however, this species is unstable and it was not possible to obtain crystals suitable for crystallographic analysis.

In this manuscript, we report the synthesis and X-ray structural characterization of several monoiron(II) complexes containing HQate ligands. Each complex features the tris(3,5-diphenylpyrazol-1yl)borate(1-) supporting ligand (Ph2Tp), as substituted Tp ligands are well-known to faithfully mimic the coordination environment of the 2-His-1-carboxylate facial triad.²¹ We found that inclusion of bulky phenyl groups at the 3-positions of the pyrazole rings generally discourages formation of the diiron(II)µ-hydroquinonate(2-) complexes, although dinuclear species were generated with certain HQs. As shown in Scheme 2, two types of HQ ligands were employed in this study: i) bidentate (or "tethered") ligands that feature an ortho substituent capable of metal coordination (H_2L^{A-E}) , and (ii) the monodentate (or "untethered") ligand 2,6-dimethylhydroquinone (H_2L^F) . These HQs were selected because they reflect the range of substrates oxidized by HQDOs, with the monodentate and bidentate ligands resembling (chloro)hydroguinones and (homo)gentisates, respectively.{NOTE: The series also includes 2-hydroxyacetophenone (H_2L^c) as a control to properly evaluate the role of the distal –OH group in tuning the structural and electronic properties of our HQDO models. Each of the resulting complexes was characterized with crystallographic, spectroscopic (UV-vis absorption, ¹H NMR), and electrochemical techniques. Indeed, we report here the first X-ray structure of a mononuclear Fe complex featuring an untethered hydroquinonate ligand. We also employed spectroscopic methods, including electron paramagnetic resonance (EPR), to examine the ferric species generated upon one-electron oxidation of the monoiron(II) complexes. These results lay the foundation for future studies that will explore the O_2 reactivity of complexes that mimic the enzyme-substrate intermediates of HQDOs.



Scheme 2

Results and Discussion

1. Fe(II) Complexes with Tethered Hydroquinonate Ligands – Synthesis and Solid State Structures

The mononuclear iron(II) complexes **1A–D** (Scheme 2) were prepared by mixing equimolar amounts of $K(^{Ph2}Tp)$ and FeX_2 (X = Cl or OTf) with the singly-deprotonated ligands, $^-HL^{A-D}$, in MeCN (or MeCN/CH₂Cl₂ solvent mixture). The resulting air-sensitive complexes dissolve easily in CH₂Cl₂, but are largely insoluble in more polar solvents like MeCN and MeOH. With the exception of **1B**, which contains a 2-acetylphenolate ligand, the FTIR spectrum of each complex exhibits a v(O-H) feature arising from the distal hydroxyl group, indicating that the HQ ligands are monoanionic and coordinated to a single Fe center.

Crystals of **1A–D** suitable for X-ray structure determination were obtained by layering concentrated CH_2Cl_2 solutions with either MeCN or pentane. Details concerning data collection and analysis are provided in Table 3, and selected bond distances and angles for **1A–D** are shown in Table 1. As illustrated in Figure 1, each complex features a five-coordinate (5C) Fe(II) center bound to a facially coordinating ^{Ph2}Tp ligand and bidentate HL^{A–D} group. The Fe-N_{Tp} bonds exhibit an average distance of 2.14 Å across the series, characteristic of highspin (*S* = 2) ferrous complexes.^{21d, 22} The Fe1-O1 distances, which range between 1.927(1) and 1.961(1) Å, are also typical for iron(II)phenolate units in 5C complexes.²³



Figure 1 Thermal ellipsoid plots (50% probability) derived from the X-ray structures of $1A \cdot CH_2Cl_2$ (top), $1B \cdot 2CH_2Cl_2$ (middle), and $1D \cdot [HNEt_3]OTf$ (bottom). Non-coordinating solvent molecules and most hydrogen atoms have been omitted for clarity, as well as the Ph-rings at the 5-positions of the ^{Ph2}Tp ligand. The HNEt₃⁺ counter cation in the $1D \cdot [HNEt_3]OTf$ structure is not shown.

	1A-CH ₂ Cl ₂	$1B {\boldsymbol{\cdot}} 2CH_2Cl_2$	1C	1D•[HNEt3]OTf	1F
Fe-O1	1.961(1)	1.937(4)	1.927(1)	1.931(3)	1.893(1)
Fe-N1	2.122(1)	2.131(5)	2.108(1)	2.130(4)	2.085(1)
Fe-N3	2.148(2)	2.089(5)	2.093(1)	2.129(4)	2.085(1)
Fe-N5	2.147(2)	2.185(5)	2.228(1)	2.186(4)	2.285(1)
Fe-L ^a	2.139(2)	2.079(4)	2.103(1)	2.317(3)	2.341(2)
O1-C46	1.341(2)	1.303(7)	1.304(2)	1.337(6)	1.338(2)
O2-C49	1.370(2)	1.370(7)		1.371(6)	1.390(2)
Ol-Fe-Nl	110.64(5)	129.5(2)	128.04(5)	128.5(2)	134.21(6)
O1-Fe-N3	153.34(5)	139.9(2)	140.01(5)	130.0(2)	131.98(6)
Ol-Fe-N5	94.78(5)	97.9(2)	100.61(4)	115.6(1)	101.45(5)
$O1\text{-}Fe\text{-}L_X$	88.90(5)	86.4(2)	85.28(4)	75.7(1)	89.04(5)
N1-Fe-N3	95.65(6)	90.6(2)	91.50(4)	94.2(1)	93.61(5)
N1-Fe-N5	92.11(5)	90.2(2)	88.25(4)	89.4(2)	85.41(5)
N3-Fe-N5	79.38(6)	81.2(2)	84.39(4)	85.6(2)	83.65(5)
L_X -Fe-Nl	90.69(6)	87.4(2)	91.32(4)	85.3(1)	89.01(5)
L _X -Fe-N3	95.37(6)	95.3(2)	88.47(4)	84.5(1)	87.53(5)
L _X -Fe-N5	174.27(6)	175.7(2)	172.83(4)	168.4(1)	169.24(5)
τ -value \underline{b}	0.35	0.60	0.55	0.64	0.58

Table 1 Selected Bond Distances (Å) and Bond Angles (deg) from the X-ray Structures of Monoiron(II) Hydroquinonate Complexes **1A–D** and **1F**. ^aL is the N or O atom of the pendant donor of the HQ anion.

^bFor a definition of the τ -value, see reference ²⁴. A five-coordinate complex with ideal square-pyramidal geometry would have a τ -value of 0.0, while those with ideal trigonal bipyramidal geometry would have a value of 1.0.

	1A • CH ₂ Cl ₂	1B • 2CH ₂ Cl ₂ ^a	1C	1D • [HNEt3]OTf	1F	2E+2Et ₂ O	[2F(MeCN)]+2DCE b
empirical formula	C ₆₀ H ₄₇ BCl ₂ FeN ₈ O ₂	C55H45BCl4FeN6O3	C53H41BFeN6O2	C59H57BF3FeN7O6S	C68H55BFeN8O2	C106H96B2Fe2N12O6	C ₁₀₄ H ₈₇ B ₂ Cl ₄ Fe ₂ N ₁₃ O ₂
formula weight	1049.62	1046.46	860.58	1115.84	1082.86	1767.27	1826.02
crystal system	Monoclinic	Orthorhombic	Monoclinic	Triclinic	Monoclinic	Trielinie	Monoclinic
space group	$P2_1/c$	P212121	$P2_1/n$	P -1	$P2_{1}/c$	P -1	Pn
a, Å	14.5859(5)	11.2362(4)	10.4129(2)	9.6832(4)	17.8034(7)	9.6359(10)	13.4895(3)
b, Å	13.6416(4)	17.7643(6)	30.8300(5)	9.8868(5)	22.3235(10)	13.4413(14)	10.0544(3)
c, Å	25.2090(8)	25.2782(8)	13.1758(2)	28.3671(15)	13.6123(6)	18.205(2)	32.9607(9)
a, deg	90	90	90	85.740(4)	90	95.633(9)	90
β, deg	93.779(3)	90	90.634(2)	86.952(4)	99.746(4)	105.298(10)	96.806(3)
γ, deg	90	90	90	81.003(4)	90	99.002(9)	90
V, Å ³	5005.0(3)	5045.6(3)	4229.6(1)	2672.5(2)	5331.9(4)	2222.1(4)	4438.9(2)
Ζ	4	4	4	2	4	1	2
D _{calc} , g/cm ³	1.393	1.351	1.351	1.387	1.349	1.321	1.342
λ, Å	1.5418	1.5418	0.7107	1.5418	0.7107	1.5418	1.5418
μ, mm ⁻¹	3.831	4.518	0.408	3.209	0.340	3.131	4.018
θ -range, deg	7 to 148	7 to 148	7 to 59	9 to 148	7 to 59	7 to 148	7 to 148
reflections collected	35336	19096	58359	15029	62124	15573	23318
independent reflections	9913 [$R_{int} = 0.0382$]	8984 [$R_{int} = 0.0435$]	10931 [$R_{int} = 0.0402$]	15029	13600 [$R_{int} = 0.0409$]	8687 [$R_{int} = 0.0622$]	12059 [$R_{int} = 0.0406$]
data / restraints / parameters	9913 / 0 / 672	8984/0/634	10931 /0 / 569	15029 / 0 / 709	13600 / 0 / 731	8687 / 24 / 569	12059 / 31 / 1132
GOF (on F^2)	1.041	1.103	1.027	1.074	1.058	1.026	1.024
$R1/wR2~(I>2\sigma(I)) \stackrel{\rm g}{=}$	0.0354 / 0.0891	0.0814 / 0.2410	0.0375 / 0.0891	0.0864 / 0.2278	0.0414 / 0.1005	0.0718 / 0.1807	0.0638 / 0.1662
R1/wR2 (all data) ^C	0.0449 / 0.0954	0.0838 / 0.2437	0.0462 / 0.0953	0.1008 / 0.2402	0.0552 / 0.1083	0.1152 / 0.2227	0.0821 / 0.1826

Table 3 Summary of X-ray Crystallographic Data Collection and StructureRefinement.

^a One of the solvate molecules in **1B** • $2CH_2Cl_2$ is only partially (77%) populated. ^b One of the solvate molecules in **[2F(MeCN)]** • 2DCE is only partially (68%) populated.

 ${}^{c}R1 = \Sigma ||F_{o}| - |F_{c}|| / \Sigma |F_{o}|; wR2 = [\Sigma w (F_{o}^{2} - F_{c}^{2})^{2} / \Sigma w (F_{o}^{2})^{2}]^{1/2}$

The coordination geometry of **1A** is intermediate between square pyramidal and trigonal bipyramidal ($\tau = 0.35^{24}$), and the HL^A ligand adopts a twisted conformation with a dihedral angle of 35° between the planes of the HQate and benzimidazolyl rings (Figure 1). This orientation is likely the result of π -stacking interactions between the benzimidazolyl moiety and a 3-phenyl substituent of the ^{Ph2}Tp ligand, in addition to steric repulsion between the HQate ring and a second phenyl group. Compared to **1A**, the structures of **1B** and **1C** lie much further towards the trigonal-bipyramidal limit ($\tau = 0.60$ and 0.55, respectively) with the acetyl group in an axial position *trans* to a pyrazole donor (N5). The metric parameters for **1B** and **1C** are nearly identical, suggesting that the structural effects of the *para* hydroxyl group are minimal. The O1-C46 distances in **1B** and **1C** are shorter than the corresponding distance in **1A** (1.303 vs. 1.341 Å; Table 1) due to delocalization of the negative charge onto the 2-acetyl group.

The O1-C46 bond of the acetophenone-derived ligands therefore acquires some double-bond character, whereas the twisted conformation of the HL^A ligand indicates a lack of electronic conjugation between the π -systems.

In contrast to the HL^{A-C} donors, the 2-methoxyhydroguinonate ligand (HL^D) in **1D** forms a five-membered ring chelate with the Fe(II) center. This fact, coupled with the intrinsically weak donating ability of methoxy substituents, results in a rather lengthy Fe1-O3 distance of 2.317(3) Å. Thus, in certain respects, **1D** can be considered to possess an intermediate coordination number between 4 and 5. As evidence, the O1-Fe1-N5 angle increases from an average of 97.8° in **1A-C** to 115.6° in **1D** (with a corresponding decrease in the O3-Fe1- N_{Tp} angles), as the HQate donor shifts out of the equatorial plane (Table 1). Thus, if the weakly-bound $-OCH_3$ group is ignored, **1D** appears to adopt a distorted trigonal pyramidal geometry with the O1 donor in the axial position. Notably, complex **1D** co-crystallizes with one equivalent of [HNEt₃]OTf salt, and the triflate anion participates in a hydrogenbonding interaction with the distal -OH group in the solid state (Figure 1; the $02\cdots04$ distance is 2.782(6)). This feature is reminiscent of acid/base interactions between HQ substrates and conserved secondsphere residues that have been proposed to play an important role in HODO catalysis.9b, 12a

The diiron(II) μ -L^X complexes were never observed in preparations of **1A–D**, and we initially attributed the lack of dinuclear side-products to the steric demands of the ^{Ph2}Tp ligand. To evaluate this hypothesis, we generated the compound 2,5dimethoxyhydroquinone (H_2L^E) , which is capable of coordinating two metal centers in a bidentate fashion. Interestingly, use of this ligand provides the diiron(II) complex **2E** as the only isolated product even when the reactants are mixed in equimolar ratios, thereby proving that the ^{Ph2}Tp framework is capable of supporting dinuclear complexes. The X-ray structure of **2E** is shown in Figure 2 and key metric parameters are listed in the caption. The complex is centrosymmetric with an Fe···Fe distance of 8.15 Å. The Fe-O/N distances of **2E** are nearly identical to those of the analogous monoiron(II) complex **1D**, although the position of the HQate ligand with respect to the N_{Tp} donors is somewhat different (e.g., $\angle O1$ -Fe1-N5 = 98.1(2)° and 115.6(1)° in **2E** and **1D**, respectively). The fact that the 2,5-dimethoxylhydroguinonate

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ligand exclusively yields **2E**, whereas ligands HL^{A-D} favor monomeric species, suggests that the thermodynamic benefit of bidentate chelation at *both* Fe(II) centers is able to overcome the steric barrier to dimerization.



Figure 2 Thermal ellipsoid plot (50% probability) derived from the X-ray structure of **2E**•CH₂Cl₂. Non-coordinating solvent molecules and hydrogen atoms have been omitted for clarity, in addition to Ph-rings at the 5-positions of the ^{Ph2}Tp ligand. Ellipsoids are not shown for four Ph rings due to disorder. Selected bond lengths (Å) and angles (°) [note: the complex is centrosymmetric]: Fe1-O1 1.904(3), Fe1-O2 2.328(3), Fe1-N1 2.107(4), Fe1-N3 2.108(3), Fe1-N5 2.213(4), O1-C46 1.329(5), O2-C47 1.388(6); O1-Fe1-O2 75.2(1), O1-Fe1-N1 131.9(2), O1-Fe1-N3 138.7(2), O1-Fe1-N5 98.9(1), O2-Fe1-N1 91.8(2), O2-Fe1-N3 97.9(1), O2-Fe1-N5 174.1(1), N1-Fe1-N3 88.3(1), N1-Fe1-N5 91.8(1), N3-Fe1-N5 86.9(1).

2. Fe(II) Complexes with an Untethered Hydroquinonate Ligand – Synthesis and Solid State Structures

As noted in the Introduction, several HODOs oxidize "untethered" HOs that lack additional metal-coordinating groups. To replicate the monodentate binding mode of these HQ substrates, we employed the ligand 2,6-dimethylhydroquinone (H_2L^F). Reaction of H_2L^F with equimolar amounts of K(Ph2Tp), FeCl₂, and NaOMe in MeCN generates a bright orange solid, which was recrystallized by slow diffusion of MeCN into a concentrated 1,2-dichloroethane (DCE) solution. X-ray analysis of the crystals revealed a diiron(II) structure with the formulation, $[Fe_2(Ph_2Tp)_2(\mu-L^F)(MeCN)]$ (**2F**(MeCN); Figure 3). Unlike **2E**, the Fe(II) centers in **2F**(MeCN) are not equivalent: Fe2 is 4C due to steric hindrance from the methyl substituents of the bridging L^{F} dianion, and Fe1 is 5C with an additional solvent MeCN ligand. The Fe1 center exhibits a distorted trigonal bipyramidal coordination geometry ($\tau =$ 0.58), while the Fe2 geometry is best described as trigonal pyramidal $(\angle O2$ -Fe2-N_{Tp} = 125 ± 5°). The low Fe2 coordination number leads to relatively short metal-ligand bond lengths, especially the Fe2-O2 distance of 1.784(6) Å (see Figure 3 caption for additional metric parameters). The high-spin Fe ions are separated by 8.72 Å. While the initial synthesis of **2F** employed equimolar amounts of reagents, the complex can also be prepared in greater yield by using only 0.5 equivalent of H_2F^F .



Figure 3 Thermal ellipsoid plot (50% probability) derived from the X-ray structure of **[2F(MeCN)]**•2DCE. Non-coordinating solvent molecules, hydrogen atoms, and Phrings at the 5-positions of the ^{Ph2}Tp ligand have been omitted for clarity. Selected bond lengths (Å): Fe1-O1 1.852(6), Fe1-N1 2.111(5), Fe1-N3 2.136(6), Fe1-N5 2.187(5), Fe1-N13 2.289(8), O1-C93 1.348(10), Fe2-O2 1.784(6), Fe2-N7 2.105(5), Fe2-N9 2.119(5), Fe2-N11 2.136(5), O2-C96 1.352(10).

In an effort to prevent formation of 2F(MeCN), one equivalent of 3,5-diphenylpyrazole (^{Ph2}pz) was included in the reaction mixture described above. Under these conditions, the reaction provided a yellow product that was recrystallized by DCE/pentane layering. X-ray diffraction analysis revealed that the crystals contain the 5C monoiron(II) complex, [Fe(^{Ph2}Tp)(HL^F)(^{Ph2}pz)] [**1F**(^{Ph2}pz)]. As shown in Figure 4, ,1F1F(^{Ph2}pz) features a trigonal bipyramidal coordination geometry ($\tau = 0.58$) with the HQate and ^{Ph2}pz donors in equatorial and axial positions, respectively. These two ligands form an intramolecular hydrogen-bond that closes a five-membered ring, as evident in the

O1…N8 distance of 2.840(2) Å and O1…H7 distance of 2.17(2) Å (the H2 and H7 atoms were found objectively and refined). The Fe1-O1 bond distance of 1.893(1) is shorter than the corresponding distances in the tethered complexes **1A–D**, whereas the axial ^{Ph2}pz ligand is weakly bound with an Fe1-N7 distance of 2.341(2) Å (Table 1). As expected, HL^F coordinates to the Fe(II) center via the more sterically-accessible O-atom at the 4-position of the HQ. Without the constraint of a pendant ligand, the HQate ring in **1F**(^{Ph2}pz) rotates away from the Fe center, as signified by the large Fe1-O1-C46 bond angle of 148.7(1)° (compared values of 125 ± 5° for **1A–1D**).



Figure 4 Thermal ellipsoid plot (50% probability) derived from the X-ray structure of **1F**. Hydrogen atoms and Ph-rings at the 5-positions of the ^{Ph2}Tp ligand have been omitted for clarity. Selected bond lengths and angles are provided in Table 1.

3. Spectroscopic and Electrochemical Properties of Fe(II)-Hydroquinonate Complexes

Electronic absorption spectra of complexes 1A-D and 1F in CH_2Cl_2 are shown in Figure 5. Complexes 1B and 1C are both brightly

colored due a weak absorption manifold ($\varepsilon \sim 0.7 \text{ mM}^{-1}\text{cm}^{-1}$) in the visible region and an intense peak in the near-UV ($\varepsilon \sim 5.5 \text{ mM}^{-1}\text{cm}^{-1}$). The application of time-dependent density functional theory (TD-DFT) to **2B** revealed that the lower-energy band arises from an Fe(II) \rightarrow HL^B MLCT transition in which the acceptor molecular orbital (MO) has primarily acetyl(C=O*) character. The higher-energy feature is assigned to a HL^B-based $n \rightarrow n^*$ transition (see ESI⁺ for details concerning the TD-DFT calculations). While **1A** does not exhibit visible-region MLCT transitions like **1B** and **1C**, a very intense ligand-based $n \rightarrow n^*$ band is observed with $\lambda_{max} = 369 \text{ nm}$ (Figure 5). In contrast, complexes containing ligands derived from methoxy- and alkyl-substituted HQs (H₂L^{D-F}) have pale yellow colors due to broad UV absorption features that tail into the visible region (Figures 5 and S1, ESI⁺).



Figure 5 Electronic absorption spectra of complexes **1A–D** and **1F** in CH₂Cl₂ at room temperature.

¹H NMR spectra of complexes **1A**–**1D** and **1F** in CD₂Cl₂ display paramagnetically-shifted signals characteristic of high-spin monoiron(II) centers (Figure S2, ESI⁺). Peaks arising from the ^{Ph2}Tp supporting ligand are easily assigned by comparison to earlier literature reports (e.g., the signal from the 4-pyrzole protons consistently appears near 55 ppm).^{21d, 22a} In each case, the resonance arising from the distal hydroxyl substituent was identified through H/D exchange with a small amount of added MeOH-*d*₄. These peaks appear downfield with chemical shifts of 23 ± 3 ppm, although the hydroxyl proton is observed at 59 ppm in the **1D** spectrum (Figure S2, ESI⁺). The observation of paramagnetically-shifted –OH resonances confirms that the HQate ligands do not adopt bridging positions in solution.

The electrochemical behavior of the monoiron(II) complexes **1A–D** and **1F** were studied by cyclic voltammetry in CH₂Cl₂ or THF solutions containing 100 mM $[NBu_4]PF_6$ as the supporting electrolyte. The cyclic voltammograms are displayed in Figure 6 and the results are summarized in Table 2. All redox potentials are referenced to the ferrocenium/ferrocene couple (Fc⁺/Fc). Complexes **1A–C** display guasi-reversible one-electron oxidation waves between -290 and -30 mV that correspond to the Fe(II/III) couple. The Fe redox potential of **1A** is significantly lower than those of **1B** and **1C**, reflecting the stronger donating ability of benzimidazolyl relative to acetyl groups. When the window is expanded to more positive potentials, both 1A and **1B** exhibit a highly irreversible wave that likely corresponds to oxidation of the respective HQate ligands. The irreversible nature of the hydroquinonate-based oxidation is probably due to subsequent loss of the distal –OH proton to the surrounding medium. Notably, **1C** is redox inactive are higher potentials, which is not surprising given that phenolates are intrinsically harder to oxidize than HQates.²⁵



Figure 6 Cyclic voltammograms of **1A–D** and **1F**. Data was collected in CH_2CI_2 (**1A–C**) or THF (**1D** and **1F**) with 100 mM (NBu₄)PF₆ as the supporting electrolyte and a scan rate of 100 mV/s. Each voltammogram was initiated by the anodic sweep.

complex	solvent	Redox Potentials (mV vs $Fc^{+/0}$) $\frac{b}{}$
1A	CH ₂ Cl ₂	$E_{1/2} (\Delta E) = -290 (110); E_{p,a} = +740 \text{ mV}$
1 B	CH ₂ Cl ₂	$E_{1/2} (\Delta E) = -110 (140); E_{p,a} = +1050 \text{ mV}$
1C	CH ₂ Cl ₂	$E_{1/2}(\Delta E) = -30(150)$
1 D	THF	$E_{p,a} = -50, +370, \text{ and } +780 \text{ mV}$
1F	THF	$E_{p,a} = -100, +580, and +850 mV$

Table 2 Redox Potentials of Complexes 1A-D and 1F.^a

^a Conditions: Solutions contained 100 mM (NBu₄)PF₆; scan rate of 100 mV s⁻¹ at room

temperature.

^b $E_{1/2}$ and ΔE values are provided for (quasi)reversible processes; $E_{p,a}$ values are given for irreversible oxidation events.

As shown in Figure 6, reversible electrochemical processes were not observed in the cyclic voltammograms of **1D** and **1F**; instead, each complex displays a weak anonic wave ($E_{p,a} = -50$ and -100 mV for **1D** and **1F**, respectively) that is assigned to one-electron oxidation of the Fe(II) center. The corresponding cathodic waves appear at much more negative potentials ($E_{p,c} \sim -600$ mV in both cases), suggesting an irreversible change following oxidation to Fe(III). Additional irreversible events arising from HQate-based oxidation are evident at higher potentials for **1D** and **1F** (Table 2; Figure 6). As expected, the potential of the first HQate-based oxidation shifts to more negative potentials as the HQate substituents become more electron-donating: $E(HL^D) < E(HL^F) < E(HL^A) < E(HL^B)$. The ill-defined electrochemical behavior of **1D** and **1F** is likely a result of the greater conformational flexibility of their HQate ligands, which are not locked into a stable six-membered ring chelate like the HL^{A-C} ligands.

Given that complexes **1A–1C** display reversible Fe(II/III) redox couples, we sought to examine the corresponding ferric species, **1X**_{ox}, with spectroscopic methods. As shown in Figure 7, treatment of the Fe(II) complexes with one equivalent of a one-electron oxidant, such as acetylferrocenium or $[N(C_6H_4Br-4)_3]^+$, yields chromophores with broad, intense absorption features centered near 480 nm. Based on literature precedents,²⁶ these bands are confidently assigned to HL^{A–} $^{C}\rightarrow$ Fe(III) LMCT transitions. The high intensities of the LMCT bands are indicative of strong Fe1-O1 covalency in the oxidized state, arising from overlap between the out-of-plane π -orbital of the phenolate ligand and the partially-occupied Fe(xy) orbital.²⁷ EPR spectra of the oxidized species **1A_{ox}-1C_{ox}** (Figure S3, ESI⁺) each reveal an intense derivative-shaped feature at g = 4.3 and a very weak peak near g =9.4, characteristic of rhombic high-spin Fe(III) centers.



Figure 7 Electronic absorption spectra of $1A_{ox}-1C_{ox}$ in CH_2Cl_2 at room temperature. The $1X_{ox}$ species were obtained by treating the Fe(II) precursors with one equivalent of acetylferrocenuium $(1A_{ox})$ or $[N(C_6H_4Br-4)_3]^+$ $(1B_{ox}$ and $1C_{ox})$.

Conclusions

We have reported the synthesis and X-ray structure analysis of a series of monoiron(II) hydroquinonate complexes (**1A–D** and **1F**) that represent the first crystallographically-characterized models of Fe/HQate interactions in HQDO active sites. The spectroscopic and electrochemical properties of the complexes were also described. The models employed bidentate ("tethered") and monodentate ("untethered") HQate ligands, since HQDOs oxidize both types of substrates. Although HQate ligands are known to bridge multiple metal centers, the tethered ligands (H₂L^{A-D}) cleanly provided 5C mononuclear complexes supported by the tridentate ^{Ph2}Tp framework. It was possible, though, to obtain the diiron(II) complex **2E** by inclusion of an additional donor substituent at the 5-position of the HQate ring. Compared to the bidentate HQates, the untethered ligand, H₂L^F, readily adopted a bridging position between Fe(II) centers, as evident in the facile formation of **2F**. Addition of one equivalent of free

pyrazole (^{Ph2}pz) to the reaction mixture, however, provided the complex **1F(^{Ph2}pz)** – the only structurally-characterized example of a monoiron(II) complex with an untethered HQate ligand reported to date. The stability of **1F(^{Ph2}pz)** is undoubtedly enhanced by an intramolecular hydrogen bond between the HQate and ^{Ph2}pz ligands (Figure 4). While crystallographic studies of substrate-bound HQDOs are not currently available, structures of ECDO:substrate complexes have revealed similar hydrogen-bonding interactions between the deprotonated O-atom of the catecholate ligand and second-sphere residues.^{2a, 28}

Thus, $\mathbf{1F}(^{Ph2}pz)$ replicates important aspects of the enzymatic coordination environment. The results presented here provide a basis for future modeling studies of the HQDOs. As noted in the introduction, the non-innocent nature of HQate ligands is thought to play an important role in the HQDO mechanism. Indeed, the cyclic voltammograms of $\mathbf{1A}$ and $\mathbf{1B}$ reveal an irreversible wave that likely corresponds to HQate oxidation coupled to loss of the distal –OH proton. Detailed studies of the electron- and proton-transfer capabilities of our mono- and dinuclear HQate complexes are currently underway with the aim of generating novel Fe benzo(semi)quinone species. In addition, we will perform O_2 reactivity studies to determine whether these excellent structural models also behave as functional models of the HQDOs.

Experimental Section

Materials and Methods

Unless otherwise noted, all reagents and solvents were purchased from commercial sources and used as received. Acetonitrile, dichloromethane, and tetrahydrofuran were purified and dried using a Vacuum Atmospheres solvent purification system. The synthesis and handling of air-sensitive materials were performed under inert atmosphere using a Vacuum Atmospheres Omni-Lab glovebox. The ligands $K(^{Ph2}Tp)^{29}$ and 2,5-dimethoxyhydroquinone $(H_2L^E)^{30}$ were prepared according to literature procedures.

Elemental analyses were performed at Midwest Microlab, LLC in Indianapolis, IN. UV-vis absorption spectra were obtained with an

Agilent 8453 diode array spectrometer equipped with a cryostat from Unisoku Scientific Instruments (Osaka, Japan) for temperature control. Fourier-transform infrared (FTIR) spectra of solid samples were measured with a Thermo Scientific Nicolet iS5 FTIR spectrometer equipped with the iD3 attenuated total reflectance accessory. ¹H spectra were collected at room temperature with a Varian 400 MHz spectrometer. EPR experiments were performed using a Bruker ELEXSYS E600 equipped with an ER4415DM cavity resonating at 9.63 GHz, an Oxford Instruments ITC503 temperature controller, and an ESR-900 He flow cryostat. Electrochemical measurements were conducted in the glovebox with an epsilon EC potentiostat (iBAS) at a scan rate of 100 mV/s with 100 mM (NBu₄)PF₆. A three-electrode cell containing a Ag/AgCl reference electrode, a platinum auxiliary electrode, and a glassy carbon working electrode was employed for cyclic voltammetric (CV) measurements. Under these conditions, the ferrocene/ferrocenium (Fc^{+/0}) couple has an $E_{1/2}$ value of +0.52 V in CH_2Cl_2 and +0.61 V in THF.

2-(1-methyl-1H-benzimidazol-2-yl)hydroquinone (H₂L^A) To 2,5dihydroxybenzealdehyde (690 mg, 5.0 mmol) dissolved in 25 mL of ethanol, *N*-methyl-1,2-benzenediamine (0.56 mL, 5.0 mmol) in 15 mL of ethanol was added dropwise over the course of 30 minutes. The mixture was then stirred at 50 °C for two days. After cooling, 30 mL of H₂O was added and the mixture was placed overnight in a freezer. The resulting brown precipitate was filtered and dried under vacuum to give the product (0.79 g, 66%). Anal. Calcd for C₁₄H₁₂N₂O₂ (MW = 240.26 g mol⁻¹): C, 69.99; H, 5.03; N, 11.66. Found: C, 69.84; H, 5.15; N, 11.70. ¹H NMR (δ, DMSO): 3.81 (s, 3H, NCH₃), 6.83 (m, 2H), 7.02 (d, 1 H), 7.27 (m, 2H), 7.63 (m, 2H), 9.10 (br s, 1H, –OH), 10.43 (br s, 1H, –OH). ¹³C{¹H} NMR (δ, DMSO): 31.6, 110.4, 116.0, 116.1 117.2, 118.5, 118.6, 121.9, 122.4, 135.8, 141.5, 149.1, 149.6, 151.9.

 $[Fe(^{Ph2}Tp)(HL^A)]$ (1A) 2-(1-methyl-1H-benzimidazol-2yl)hydroquinone (H₂L^A) (120 mg, 0.50 mmol) was deprotonated by reaction with one equivalent of NaOMe in 10 mL of MeCN. To this solution was added FeCl₂ (64.2 mg, 0.50 mmol) and K(^{Ph2}Tp) (350 mg, 0.49 mmol). The mixture was stirred for overnight and the solvent removed under vacuum to give a yellow-brown solid. The crude product was dissolved in CH₂Cl₂ and filtered; the resulting solution

yielded yellow crystals suitable for crystallographic analysis after standing for several days (0.37 g, 77%). Anal. Calcd for $C_{59}H_{45}BFeN_8O_2$ (MW = 964.70 g mol⁻¹): C, 73.56; H, 4.70; N, 11.62. Found: C, 73.18; H, 4.87; N, 11.72. UV-vis [λ_{max} , nm (ϵ , M⁻¹cm⁻¹) in CH₂Cl₂]: 369 (10,100). FTIR (cm⁻¹, solution): 3592 (OH), 3047, 2988, 2901, 2611 (BH), 1543, 1484, 1415, 1332, 1243, 1171, 1070, 1007, 963, 914, 818, 760, 692.

[$Fe(^{Ph_2}Tp)(HL^B)$] (1B) Under an inert atmosphere, 182 mg (1.20 mmol) of 2',5'-dihydroxyacetophenone (H₂L^B) was deprotonated by mixing with one equivalent of NaOMe in THF for 30 minutes, after which the solvent was removed to yield the Na(HL^B) salt as a white solid. To this compound was added anhydrous FeCl₂ (146 mg, 1.15 mmol) and K(^{Ph_2}Tp) (815 mg, 1.15 mmol) in 15 ml of MeCN. After stirring the reaction mixture overnight, the resulting solid was collected by vacuum filtration, dried, and redissolved in CH₂Cl₂. Layering with pentane provided reddish brown crystals suitable for X-ray diffraction (0.26 g, 26%). Anal. Calcd for C₅₃H₄₁BFeN₆O₃ (MW = 876.59 g mol⁻¹): C, 72.62; H, 4.71; N, 9.59. Found: C, 72.49; H, 4.79; N, 9.73. UV-vis [λ_{max} , nm (ϵ , M⁻¹cm⁻¹) in CH₂Cl₂]: 394 (5410), 485 (780), 527 (690). FTIR (cm⁻¹, solid): 3559 (OH), 3058, 2608 (BH), 1604 (CO_{acetyl}), 1547, 1475, 1462, 1430, 1411, 1359, 1340, 1327, 1299, 1197, 1164, 1062, 1006, 965, 917, 810, 759, 693.

[$Fe(^{Ph2}Tp)(HL^{C})$] (1C) The method of preparation was similar to the one described for **1B**, except that 2'-hydroxyacetophenone (H₂L^C) was substituted for H₂L^B. Orange crystals were obtained by layering a concentrated CH₂Cl₂ solution with MeCN. Yield = 24 %. Anal. Calcd for C₅₃H₄₁BFeN₆O₂ (MW = 860.59 g mol⁻¹): C, 73.97; H, 4.80; N, 9.77. Found: C, 74.15; H, 4.92; N, 9.83. UV-Vis [λ_{max} , nm (ϵ , M⁻¹cm⁻¹) in CH₂Cl₂]: 368 (5950), 441 (540), 485 (570). FTIR (cm⁻¹, solid): 3060, 2618 (BH), 1613 (CO_{acetyl}), 1529, 1479, 1463, 1432, 1414, 1361, 1346, 1331, 1225, 1167, 1063, 1010, 966, 912, 863, 804, 753, 692.

 $[Fe(^{Ph2}Tp)(HL^D)]$ (1D) This compound was prepared via two methods. **Method A**: 2-methoxyhydroquinone (H₂L^D, 151 mg, 1.1 mmol) and triethylamine (1.1 mmol) were stirred in MeCN, followed by addition of K(^{Ph2}Tp) (710 mg, 1.0 mmol) and Fe(OTf)₂ (372 mg, 1.05 mmol) dissolved in CH₂Cl₂ and MeCN, respectively. The mixture was stirred overnight, filtered, and the solvent removed under vacuum.

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The resulting solid was washed multiple times with MeCN to remove triflate salts and other impurities, then dried again. The solid was dissolved in CH₂Cl₂ and layered with hexane to yield a yellow crystalline powder (0.28 g, 33%). Anal. Calcd for C₅₂H₄₁BFeN₆O₃ (MW = 864.58 g mol⁻¹): C, 72.24; H, 4.78; N 9.72. Found: C, 69.69; H, 5.65; N 10.63 (the discrepancies indicate the presence of small amounts of impurities). UV-Vis [λ_{max} , nm (ϵ , M⁻¹cm⁻¹) in CH₂Cl₂]: 383 (1490). FTIR (cm⁻¹, solid): 3563 (OH), 3056, 2931, 2615 (BH), 1543, 1495, 1477, 1461, 1410, 1357, 1305, 1260, 1226, 1164, 1060, 1008, 913, 818, 754, 690. **Method B**: Equimolar amounts of the four reagents – Fe(OTf)₂, K(^{Ph2}Tp), H₂L^D, and NEt₃ – were mixed in CH₂Cl₂ and stirred overnight. The solution was filtered and the solvent removed under vacuum. The solid was taken up in CH₂Cl₂ and layered with pentane to yield yellow crystals suitable for X-ray diffraction.

[$Fe(^{Ph2}Tp)(HL^F)(^{Ph2}pz)$] (1F) 3,5-diphenylpyrazole (236 mg, 1.04 mmol), K(^{Ph2}Tp) (714 mg, 1.01 mmol), and 2,6-dimethylhydroquinone (H₂L^F, 164 mg, 1.19 mmol) were dissolved in a 3:1 mixture of DCE:MeCN. To this solution was added FeCl₂ (129 mg, 1.02 mmol) in MeCN and NaOMe (0.23 mL of 4.37 M MeOH solution, 1.00 mmol). The reaction was stirred overnight. The solvent was evaporated under vacuum to give a pale orange solid. The crude solid was taken up into DCE and filtered, providing bright yellow solution. Yellow crystals were obtained by layering this DCE solution with pentane. Anal. Calcd for C₆₈H₅₅BFeN₈O₂ (MW = 1082.88 g mol⁻¹): C, 75.42; H, 5.12; N 10.35. Found: C, 75.22; H, 5.00; N 10.21. UV-Vis [λ_{max} , nm (ϵ , M⁻¹cm⁻¹) in CH₂Cl₂]: 374 (2530). FTIR (cm⁻¹, solid): 3355 (OH), 3060, 3038, 2912, 2631 (BH), 1598, 1543, 1477, 1465, 1430, 1410, 1339, 1306, 1212, 1165, 1062, 1004, 967, 913, 851, 810, 754, 688.

 $[Fe_2(^{Ph_2}Tp)_2(\mu-L^E)]$ (2E) 2,5-dimethoxyhydroquinone (H₂L^E, 91 mg, 0.53 mmol) was first deprotonated by treatment with two equivalents of NaOMe in THF. After removal of the solvent, the resulting white solid Na₂(L^E) was mixed with FeCl₂ (131 mg, 1.03 mmol) and K(^{Ph_2}Tp) (715 mg, 1.01 mmol) in MeCN, and the solution was stirred overnight. After removal of the solvent under vacuum, the yellow solid was taken up CH₂Cl₂ and the solution filtered to remove unwanted salts. Vapor diffusion of Et₂O into this CH₂Cl₂ solution provided yellow-orange needles suitable for X-ray crystallography (0.11 g, 13%). Anal. Calcd for C₉₈H₇₆B₂Fe₂N₁₂O₄ (MW = 1619.07 g mol⁻¹): C, 72.70; H, 4.73; N

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10.38. Found: C, 72.45; H, 4.67; N, 10.36. UV-Vis [λ_{max} , nm (ϵ , M⁻¹cm⁻¹) in CH₂Cl₂]: 317 (9300), 370 (sh), 444 (sh). FTIR (cm⁻¹, solid): 3058, 2926, 2614 (BH), 1541, 1478, 1465, 1438, 1407, 1359, 1260, 1221, 1194, 1167, 1154, 1061, 1008, 888, 802, 756, 690.

 $[Fe_2(^{Ph_2}Tp)_2(\mu-L^F)(MeCN)]$ [2F(MeCN)] Anhydrous FeCl₂ (130 mg, 1.02 mmol) and K(^{Ph2}Tp) (715 mg, 1.01 mmol) were combined with 0.5 equivalent of 2,6-dimethylhydroquinone (H_2L^F , 70.0 mg, 0.51 mmol) in 10 mL of MeCN. To this mixture was added 0.23 mL of 4.37 M solution of NaOMe (1.01 mmol). The reaction was stirred overnight, and the solvent removed under vacuum. The resulting solid was dissolved in DCE, filtered, and then layered with MeCN to provide reddish-brown needles (0.21 g, 26%) suitable for crystallographic analysis. The X-ray structure revealed two uncoordinated DCE molecules in the asymmetric unit, and elemental analysis suggest that a small amount of solvent (~0.8 equiv.) remains even after drying. Anal. Calcd for $C_{100}H_{79}B_2Fe_2N_{13}O_2 \bullet 0.8DCE$ (MW = 1707.27 g mol⁻¹): C, 71.48; H, 4.85; N 10.67. Found: C, 71.47; H, 4.77; N, 10.37. UV-Vis $[\lambda_{max}, nm (\epsilon, M^{-1}cm^{-1}) in CH_2Cl_2]$: 288 (11,500), 377 (3600). FTIR (cm⁻¹, solid): 3052, 2925, 2608 (BH), 1542, 1465, 1477, 1431, 1412, 1358, 1242, 1162, 1065, 1029, 1009, 969, 916, 847, 810.

Crystallographic Studies

Each complex was characterized with X-ray crystallography; details concerning the data collection and analysis are summarized in Table 3. The X-ray diffraction data were collected at 100 K with an Oxford Diffraction SuperNova kappa-diffractometer equipped with dual microfocus Cu/Mo X-ray sources, X-ray mirror optics, Atlas CCD detector and a low-temperature Cryojet device. The data were processed with CrysAlisPro program package (Oxford Diffraction Ltd., 2010) typically using a numerical Gaussian absorption correction (based on the real shape of the crystal) followed by an empirical multiscan correction using SCALE3 ABSPACK routine. The structures were solved using the SHELXS program and refined with the SHELXL program³¹ within the Olex2 crystallographic package.³² All computations were performed on an Intel PC computer with Windows 7 OS. Some structures contain disorder that was detected in difference Fourier syntheses of electron density and accounted for using capabilities of the SHELX package. In most cases, hydrogen atoms

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were localized in difference syntheses of electron density but were refined using appropriate geometric restrictions on the corresponding bond lengths and bond angles within a riding/rotating model (torsion angles of methyl hydrogens were optimized to better fit the residual electron density).

Supplementary Material

Electronic Supporting Information(ESI) for:

Structural, Spectroscopic, and Electrochemical Properties of Nonheme Fe(II)-Hydroquinonate Complexes: Synthetic Models of Hydroquinone Dioxygenases

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Computational Details

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Figure S1. Electronic absorption spectra of **2E** and **2F(MeCN)** in CH2Cl2 at room temperature.



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Figure S2. ¹H NMR spectra of **1A-1D**, and **1F** in CD_2Cl_2 at ambient temperature. Peaks marked with an asterisk (*) disappeared upon addition of a small amount of MeOH-d₄ and are therefore assigned to the exchangeable proton of the distal –OH moiety. Resonances arising from protons at the 4-positions of the ^{Ph2}Tp pyrazole rings (4-pz) were identified on the basis of peak integrations.



Figure S3. X-band EPR spectra of $1A_{ox}-1C_{ox}$ in frozen CH2Cl2 solutions. The $1X_{ox}$ species were obtained by treating the Fe(II) precursors with one equivalent of acetylferrocenuium ($1A_{ox}$) or [N(C₆H₄Br-4)₃]+ ($1B_{ox}$ and $1C_{ox}$). The spectra were collected under the following conditions: frequency = 9.63 GHz; power = 2.0 mW; modulation = 12 G; temperature = 10 K.

Computational Details.

Density functional theory (DFT) calculations of complex **1B** were performed using the ORCA 2.0 software package developed by Dr. F. Neese.¹ Atomic coordinates were obtained from the corresponding Xray structure, although the 5-Ph groups of the ^{Ph2}Tp ligand were replaced by -CH3 groups in the computational model. The DFT calculations employed the Becke-Perdew (BP86) functional² and Ahlrichs' valence triple- ζ basis set (TZV) for all atoms, in conjunction with the TZV/J auxiliary basis set.³ Extra polarization functions were used on non-hydrogen atoms. Time-dependent DFT (TD-DFT) calculations⁴ provided absorption energies and intensities within the Tamm-Dancoff approximation.⁵ Forty excited states were calculated.

- Neese, F.; ORCA An ab initio, Density Functional and Semi-empirical Program Package, version 2.8; University of Bonn: Bonn, Germany, 2010.
- (a) Becke, A. D. J. Chem. Phys. **1986**, 84, 4524-4529. (b) Perdew, J. P. Phys. Rev. B **1986**, 33, 8822-8824.
- (a) Schafer, A.; Horn, H.; Ahlrichs, R. J. Chem. Phys. 1992, 97, 2571-2577. (b) Schafer, A.; Huber, C.; Ahlrichs, R. J. Chem. Phys. 1994, 100, 5829-5835.
- 4. (a) Stratmann, R. E.; Scuseria, G. E.; Frisch, M. J. J. Chem. Phys. 1998, 109, 8218-8224. (b) Casida, M. E.; Jamorski, C.; Casida, K. C.; Salahub, D. R. J. Chem. Phys. 1998, 108, 4439-4449. (c) Bauernschmitt, R.; Ahlrichs, R. Chem. Phys. Lett. 1996, 256, 454-464.
- 5. (a) Hirata, S.; Head-Gordon, M. *Chem. Phys. Lett.* **1999**, *314*, 291-299.
 (b) Hirata, S.; Head-Gordon, M. *Chem. Phys. Lett.* **1999**, *302*, 375-382.

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Notes and References

- 1. (a) Gibson DT, Parales RE. Curr Opin Biotechnol. 2000;11:236–243.
 (b) Parales R, Resnick SM. In: Biodegradation and Bioremediation. Singh A, Ward OP, editors. Springer; Heidelberg: 2004. pp. 175–196.
 (c) Furukawa K. Curr Opin Biotechnol. 2000;11:244–249.
- 2. (a) Vaillancourt FH, Bolin JT, Eltis LD. Crit Rev Biochem Mol Biol. 2006;41:241–267.
 (b) Costas M, Mehn MP, Jensen MP, Que L., Jr Chem Rev. 2004;104:939–986.
- 3. Koehntop KD, Emerson JP, Que L., Jr J Biol Inorg Chem. 2005;10:87-93.
- 4. (a) Straganz GD, Nidetzky B. Chembiochem. 2006;7:1536–1548.
 (b) Diebold AR, Neidig ML, Moran GR, Straganz GD, Solomon EI. Biochemistry. 2010;49:6945–6952.

5. (a) Lipscomb JD. Curr Opin Struct Biol. 2008;18:644–649.
(b) Bugg TDH, Ramaswamy S. Curr Opin Chem Biol. 2008;12:134–140.
(c) Bruijnincx PCA, van Koten G, Gebbink RJMK. Chem Soc Rev. 2008;37:2716–2744.
(d) Siegbahn PEM, Haeffner F. J Am Chem Soc. 2004;126:8919–8932.
(e) Solomon EI, Brunold TC, Davis MI, Kemsley JN, Lee SK, Lehnert N, Neese F, Skulan AJ, Yang YS, Zhou J. Chem Rev. 2000;100:235–349.

- Lipscomb JD, Orville AM. In: Met Ions Biol Syst. Sigel H, Sigel A, editors. Vol. 28. Marcel Dekker; New York: 1992. pp. 243–298.
- 7. (a) Li XW, Guo M, Fan J, Tang WY, Wang DQ, Ge HH, Rong H, Teng MK, Niu LW, Liu Q, Hao Q. Protein Sci. 2006;15:761–773.
 (b) Zhang Y, Colabroy KL, Begley TP, Ealick SE. Biochemistry. 2005;44:7632–7643.
 (c) Lendenmann U, Spain JC. J Bacteriol. 1996;178:6227–6232.
- 8. (a) Matera I, Ferraroni M, Burger S, Scozzafava A, Stolz A, Briganti F. J Mol Biol. 2008;380:856–868.
 (b) Hintner JP, Lechner C, Riegert U, Kuhm AE, Storm T, Reemtsma T, Stolz A. J Bacteriol. 2001;183:6936–6942.
 (c) Hintner JP, Remtsma T, Stolz A. J Biol Chem. 2004;279:37250– 37260.

Dalton Transactions, Vol. 41, No. 39 (October 2012): pg. 12244-12253. <u>DOI</u>. This article is © Royal Society of Chemistry and permission has been granted for this version to appear in <u>e-Publications@Marquette</u>. Royal Society of Chemistry does not grant permission for this article to be further copied/distributed or hosted elsewhere without the express permission from Royal Society of Chemistry.

- 9. (a) Harpel MR, Lipscomb JD. J Biol Chem. 1990;265:22187–22196.
 (b) Chen J, Li W, Wang MZ, Zhu GY, Liu DQ, Sun F, Hao N, Li XM, Rao ZH, Zhang XC. Protein Sci. 2008;17:1362–1373.
- 10. (a) Veldhuizen EJA, Vaillancourt FH, Whiting CJ, Hsiao MMY, Gingras G, Xiao YF, Tanguay RM, Boukouvalas J, Eltis LD. Biochem J. 2005;386:305–314.
 (b) Titus GP, Mueller HA, Burgner J, Córdoba SRd, Penalva MA, Timm DE. Nat Struct Biol. 2000;7:542–546.
- 11. Borowski T, Georgiev V, Siegbahn PEM. J Am Chem Soc. 2005;127:17303–17314.
- 12. (a) Machonkin TE, Doerner AE. Biochemistry. 2011;50:8899-8913.
 (b) Machonkin TE, Holland PL, Smith KN, Liberman JS, Dinescu A, Cundari TR, Rocks SS. J Biol Inorg Chem. 2010;15:291-301.
 (c) Xu L, Resing K, Lawson SL, Babbitt PC, Copley SD. Biochemistry. 1999;38:7659-7669.
 (d) Ohtsubo Y, Miyauchi K, Kanda K, Hatta T, Kiyohara H, Senda T, Nagata Y, Mitsui Y, Takagi M. FEBS Lett. 1999;459:395-398.
- 13. (a) Nagata Y, Endo R, Ito M, Ohtsubo Y, Tsuda M. Appl Microbiol Biotechnol. 2007;76:741–752.
 (b) Miyauchi K, Adachi Y, Nagata Y, Takagi M. J Bacteriol. 1999;181:6712–6719.
- 14. Yin Y, Zhou NY. Curr Microbiol. 2010;61:471–476.
- 15. Christian GJ, Ye SF, Neese F. Chem Sci. 2012;3:1600–1611.
- 16. (a) Kovaleva EG, Lipscomb JD. Science. 2007;316:453–457.(b) Bugg TDH, Lin G. Chem Commun. 2001;11:941–953.
- 17. Becker JM, Barker J, Clarkson GJ, van Gorkum R, Johal GK, Walton RI, Scott P. Dalton Trans. 2010;39:2309–2326.
- 18. (a) Heistand RH, II, Roe AL, Que L., Jr Inorg Chem. 1982;21:676–681.
 (b) Maroney MJ, Day RO, Psyris T, Fleury LM, Whitehead JP. Inorg Chem. 1989;28:173–175.
- 19. Rheingold AL, Miller J. Private communication to Cambridge Structural Database. 2003.
- 20. Rocks SS, Brennessel WW, Machonkin TE, Holland PL. Inorg Chem. 2010;49:10914–10929.

Dalton Transactions, Vol. 41, No. 39 (October 2012): pg. 12244-12253. <u>DOI</u>. This article is © Royal Society of Chemistry and permission has been granted for this version to appear in <u>e-Publications@Marquette</u>. Royal Society of Chemistry does not grant permission for this article to be further copied/distributed or hosted elsewhere without the express permission from Royal Society of Chemistry.

- 21. (a) Paria S, Que L, Paine TK. Angew Chem Intl Ed. 2011;50:11129–11132.
 (b) Mukherjee A, Cranswick MA, Chakrabarti M, Paine TK, Fujisawa K, Munck E, Que L. Inorg Chem. 2010;49:3618–3628.
 (c) Bruijnincx PCA, Lutz M, Spek AL, Hagen WR, Weckhuysen BM, vanKoten G, Gebbink RJMK. J Am Chem Soc. 2007;129:2275–2286.
 (d) Mehn MP, Fujisawa K, Hegg EL, Que L., Jr J Am Chem Soc. 2003;125:7828–7842.
 (e) Ogihara T, Hikichi S, Akita M, Morooka Y. Inorg Chem. 1998;37:2614–2615.
- 22. (a) Park H, Baus JS, Lindeman SV, Fiedler AT. Inorg Chem.
 2011;50:11978–11989.
 (b) Siewert I, Limberg C. Angew Chem Intl Ed. 2008;47:7953–7956.
- 23. (a) Paria S, Halder P, Chakraborty B, Paine TK. Indian J Chem, Sect A. 2011;50:420–426.
 (b) Fujisawa K, Tada N, Nishida Y, Miyashita Y, Okamoto K. Inorg Chem Commun. 2008;11:381–384.
- 24. Addison AW, Rao TN, Reedijk J, Vanrijn J, Verschoor GC. J Chem Soc, Dalton Trans. 1984:1349–1356.
- 25. Warren JJ, Tronic TA, Mayer JM. Chem Rev. 2010;110:6961-7001.
- 26. (a) Ito M, Amagai H, Fukui H, Kitajima N, MoroOka Y. Bull Chem Soc Jpn. 1996;69:1937–1945.
 (b) Pyrz JW, Roe AL, Stern LJ, Que L., Jr J Am Chem Soc. 1985;107:614–620.
- 27. Davis MI, Orville AM, Neese F, Zaleski JM, Lipscomb JD, Solomon EI. J Am Chem Soc. 2002;124:602–614.
- 28. (a) Sato N, Uragami Y, Nishizaki T, Takahashi Y, Sazaki G, Sugimoto K, Nonaka T, Masai E, Fukuda M, Senda T. J Mol Biol. 2002;321:621–636.
 (b) Vaillancourt FH, Barbosa CJ, Spiro TG, Bolin JT, Blades MW, Turner RFB, Eltis LD. J Am Chem Soc. 2002;124:2485–2496.
- Kitajima N, Fujisawa L, Fujimoto C, Moro-oka Y, Hashimoto S, Kitagawa T, Toriumi K, Tatsumi K, Nakamura A. J Am Chem Soc. 1992;114:1277– 1291.
- 30. Hanss D, Walther ME, Wenger OS. Chem Commun. 2010;46:7034-7036.
- 31. Sheldrick GM. Acta Crystallogr Sect A. 2008;64:112–122.

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32. Dolomanov OV, Bourhis LJ, Gildea RJ, Howard JAK, Puschmann H. J Appl Crystallogr. 2009;42:339–341.

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