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Ilia Rochlin

Howard S. Ginsberg University of Rhode Island, hginsberg@uri.edu

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Rochlin, I., Ginsberg, H. S., & Campbell, S. R. (2009). Distribution and Abundance of Host-seeking Culex Species at Three Proximate Locations with Different Levels of West Nile Virus Activity. *American Journal of Tropical Medicine and Hygiene*, 80(4), 661-668. doi: 10.4269/ajtmh.2009.80.661

Available at: https://doi.org/10.4269/ajtmh.2009.80.661

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]	Authors Ilia Rochlin, Howard S. Ginsberg, and Scott R. Campbell							

Distribution and Abundance of Host-seeking *Culex* Species at Three Proximate Locations with Different Levels of West Nile Virus Activity

Ilia Rochlin,* Howard S. Ginsberg, and Scott R. Campbell

Division of Vector Control, Suffolk County Department of Public Works, Yaphank, New York; U.S. Geological Survey Patuxent Wildlife Research Center, Coastal Field Station, University of Rhode Island, Kingston, Rhode Island; Arthropod-Borne Disease Laboratory, Suffolk County Department of Health Services, Yaphank, New York

Abstract. Culex species were monitored at three proximate sites with historically different West Nile virus (WNV) activities. The site with human WNV transmission (epidemic) had the lowest abundance of the putative bridge vectors, Culex pipiens and Cx. salinarius. The site with horse cases but not human cases (epizootic) had the highest percent composition of Cx. salinarius, whereas the site with WNV-positive birds only (enzootic) had the highest Cx. pipiens abundance and percent composition. A total of 29 WNV-positive Culex pools were collected at the enzootic site, 17 at the epidemic site, and 14 at the epizootic site. Published models of human risk using Cx. pipiens and Cx. salinarius as the primary bridge vectors did not explain WNV activity at our sites. Other variables, such as additional vector species, environmental components, and socioeconomic factors, need to be examined to explain the observed patterns of WNV epidemic activity.

INTRODUCTION

Mosquitoes of the genus Culex have been implicated as major vectors of West Nile virus (WNV) worldwide.1 In the northeastern United States, the virus enzootic transmission cycle is maintained primarily by ornithophilic Culex pipiens and Cx. restuans mosquitoes.²⁻⁴ Most wild and domestic mammals, as well as humans, appear to be dead end or incidental hosts.^{1,3} Several species of mosquito have been implicated as potential bridge vectors, including Cx. salinarius, Cx. pipiens, and various other Aedes and Culex species, but questions remain as to the importance of each species overall and at individual sites.^{3,5} Culex salinarius has been proposed to be the main bridge vector because of its abundance during the peak transmission season, indiscriminate feeding habits, vector competence, and a considerable number of WNV isolates.^{2,4,6–8} An alternative hypothesis implicates Cx. pipiens as the major epidemic vector on the basis of a risk assessment model, vector competence, high number of WNV isolates, and a shift in feeding behavior from avian to mammalian hosts in the late summer. 9,10 The two hypotheses for these Culex species enable additional species, such as Aedes vexans and Ae. japonicus, to be minor epidemic vectors.^{2,9,11,12} A third hypothesis is that Cx. pipiens and Cx. salinarius are responsible for equine and human WNV transmission, and several non-Culex species serve as occasional, or locally important, bridge vectors.13

Little is known about the relationships between mosquito populations and the incidence of WNV within and outside recognized WNV foci,² and studies that address population dynamics and abundance of mosquito vectors, specifically *Culex* species, in areas of WNV transmission are relatively scarce, although such knowledge is invaluable for disease management and vector control.¹⁴ Limited knowledge on geographic dimensions of WNV exists,¹⁵ especially on the subcounty level, thus obscuring risk patterns on a finer spatial scale relevant to vector ecology.¹⁶ Recent studies have

MATERIALS AND METHODS

Selection and characterization of study sites. Data on locally acquired WNV human and equine cases, positive mosquito pools, and positive birds were provided by Suffolk County and New York State agencies (Table 1). All spatial data were processed using ArcGIS 9.1 software (Environmental Systems Research Institute Inc., Redlands, CA). Spatial cluster detection SaTScanTM software²² was used to identify, rank, and determine the spatial extent of the geographic clusters of WNV human and equine cases that occurred from 1999 through 2004. SaTScanTM uses a circular window (set to the maximum radius of 10 km) to detect potential clusters using a pre-determined coordinate file (tracts of census centroids in this study). The larger spatial extent of the most probable human cluster (5.5 km, approximately 3 miles) was used to define the three study areas. These areas were ranked based on Centers for Disease Control and Prevention (CDC) (Atlanta, GA) WNV guidelines23 as either epidemic (high-to-outbreak human transmission risk, categories 4-5 with human and equine cases), epizootic (moderate-to-high human transmission risk, categories 3-4 with equine cases only), or enzootic (low-to-moderate human transmission risk, categories 2-3 with positive birds only). Area 1 (the most probable human cluster, Figure 1) was thus classified as epidemic, area 3 as epizootic (equine cluster only, no human cases), and area 2 as enzootic (no evidence of mammalian transmission). The human population density was calculated for each area using the 2000 U.S. Census Bureau block data.24

suggested highly focal WNV transmission patterns with fine-scale spatial and temporal dynamics. 16-21 To address this issue, we examined the three bridge vector hypotheses in terms of *Culex* species abundance, composition, and population dynamics within three proximate areas located inside the original WNV epicenter in Suffolk County, New York⁷ and characterized by different levels of historical WNV epidemic and epizootic activities. We selected sites and sizes of sample areas by analyzing historical data using geographic clustering software. We then compared actual data on mosquito distribution and WNV infection to predictions from current models of WNV transmission.

^{*} Address correspondence to Ilia Rochlin, Division of Vector Control, Suffolk County Department of Public Works, 335 Yaphank Avenue, Yaphank, NY 11980-9744. E-mail: ilia.rochlin@suffolkcountyny.gov

Table 1 Comparative analysis of West Nile virus (WNV) activity and human population data at three study areas (three-mile radius), Suffolk County, New York

		WNV activity*								
	Huma	Human cases		Mosqu	Mosquito pools		Birds			
Area	1999–2004‡	2005-2006§	1999–2004	1999–2004	2005–2006	1999–2004	2005–2006	Human population† Density (%)		
1	5	1¶	3	4	17	54	4	2,692 (5.4)		
2	None	None	None	None	32	64	16	2,707 (5.4)		
3	None	None	8	24	20	72	11	2,843 (5.7)		
County total	19	11	64	272	133	903	165	1,556		

- No. of human and equine cases, mosquito pools, and positive birds are shown.
- † No. of persons per square mile (density) and percentage of the total county population (%) are indicated. ‡ Pre-study, 1999–2004 (mosquito trap locations used for routine surveillance were different from those used for the study).
- \$ Study, 2005–2006 (enhanced surveillance with nine new trap locations used in this study).

 ¶ One additional human case occurred approximately 0.5 miles mile the south of area 1.

To select unbiased study sites (Sites 1–3), one-mile radius circles were delineated around the centroids of polygons formed by either interconnecting human and equine WNV cases within each of areas 1 and 3, or those outside area 2 (Figure 1). The resulting sites were characterized by land use/cover (LUC) using on-screen digitizing of 2001 aerial ortho-photography of Suffolk County,25 and validated by ground-truthing (Table 2). Three LUC types within each study site were monitored: residential, commercial, and natural (forested, open water, and wetlands). These areas were considered the most suitable for Culex spp. because of larval habitat (catch and retention basins, open water, wetlands), resting places (vegetation), and hosts (humans and wildlife). Residential areas were assumed the most likely locations where WNV human and equine infection could be acquired.26

Collection and identification of mosquitoes. Trapping was carried out weekly (n = 36 weeks total) from June 1 through September 30 in 2005 and 2006. One CDC miniature light

trap baited with dry ice and one CDC gravid trap baited with rabbit chow infusion were placed approximately 20 meters apart in residential, commercial, and natural areas at each of three sites specifically selected for enhanced surveillance (Figure 1). Mosquitoes were anesthetized and identified to species level except for Cx. pipiens/restuans and Cx. salinarius.²⁷ Routine identification of female mosquitoes of these three species can be difficult or not possible because of irregular morphology and damage during the collection process.²⁸⁻³¹ Thus, a rapid molecular identification method using a polymerase chain reaction (PCR) was used.²⁷ Briefly, legs from individual Culex mosquitoes treated with proteinase K were subjected to a multiplex PCR with species specific primers,29 and the products visualized after electrophoresis on a gel.

Mosquito processing for WNV testing. Culex and other species females were pooled in groups of 5 to 50 mosquitoes by date, site, LUC, and trap type (combined if < 5). The pools

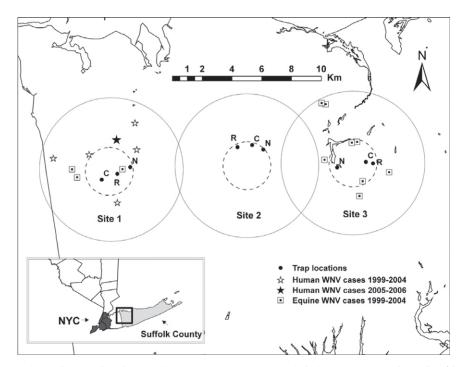


FIGURE 1. Study sites and trapping locations in Suffolk County, New York. Dotted circles show a one-mile radius (sites 1-3) and solid circles show a three-mile radius (areas 1-3). Human and equine West Nile virus cases and mosquito trap locations (each with one CDC light trap and one gravid trap approximately 20 meters apart) are shown. The land use/cover types are indicated for each mosquito trap location. C = commercial; N = natural; R = residential.

Table 2
Comparative analysis of percentage land use/cover at three study sites (one-mile radius), Suffolk County, New York

Land use, %*								
Site	Bar	Comm	For	Resd	Rec	Wet	Wat	
1	4.0	13.0	9.2	71.2	0.0	2.4	0.3	
2	11.2	8.7	21.5	48.9	8.9	0.5	0.3	
3	6.0	9.2	25.8	47.0	6.1	3.0	2.8	

*Bar = barren (open grassy or sandy areas); Comm = commercial (large stores and strip malls; For = forested (sylvan habitat > 80% tree cover; Resd = residential (mostly single-family homes and small apartment complexes); Rec = recreational (golf courses); Wet = wetland (natural [NewYork State Department of Environmental Conservation]²⁴ and human-made areas); Wat = open water (natural [lakes and ponds] and human-made [recharge basins]).

were submitted to New York State Department of Health Arboviruses Laboratories for WNV testing by reverse transcription–PCR (RT-PCR) as a part of routine WNV surveillance to ensure timely results prior to our molecular species identification carried out in the off-season. Historically, combined *Cx. pipiens/Cx. restuans* pools represented most (approximately 90%) of the total *Culex* spp. pools tested by New York State Department of Health, ³² with some pools likely containing *Cx. salinarius*. ³³ Weekly WNV minimum infection rate (MIR) was calculated using a Microsoft Excel add-in program supplied by CDC. ³⁴

Statistical analysis of Culex species abundance and composition. Data from 2005 and 2006 were combined to increase the power of the analysis. Abundance (mean weekly catch per light trap per night) and species percent composition (the proportion of each species in light traps) were calculated for Cx. pipiens, Cx. restuans, and Cx. salinarius for each site and LUC type using SPSS software (SPSS Inc, Chicago, IL). The mean weekly catch was $\log (x + 1)$ transformed and analyzed by multivariate analysis of variance (general linear models function in SPSS) for main and interaction effects of year, site, and LUC type. Dunnett's T3 post hoc test was used for pairwise comparisons due to unequal variances among the groups. Bonferroni adjustment was applied to the statistical significance at P < 0.05 for multiple comparisons. To compare Culex spp. composition among the study sites and LUC types, contingency tables were analyzed using Pearson's chi-square test. Significant results at P < 0.05 were interpreted using the standardized residuals considered significant if > 2.0 or < -2.0.

RESULTS

Characterization of study sites. Of 19 human WNV cases recorded in Suffolk County from 1999 through 2004 (prestudy), 5 (26%) occurred within area 1 (Table 1). Areas 1 and 3 had 3 (5%) and 8 (12.5%) of 64 WNV equine cases, respectively. Routine mosquito trapping at locations different from those established for this study resulted in 4 (1.5%) and

24 (9%) of 272 WNV-positive mosquito pools collected in areas 1 and 3, respectively. In contrast, area 2 had no indication of WNV activity except positive birds, whose distribution, as well as mosquito surveillance efforts and human population density were comparable among the three areas.

The three sites selected for this study (Figure 1) represented typical suburban environments with residential LUC occupying from approximately half (sites 2 and 3) to two-thirds (site 1) of the total area (Table 2). Site 3 had the highest natural LUC (approximately 32%), followed by site 2 (approximately 22%) and site 1 (approximately 12%). Commercial LUC occupied approximately 10% of each site.

Abundance, composition, and population dynamics of Culex species. More than two-thirds of 15,302 Culex spp. females were caught in the gravid traps (Table 3). In 2005, approximately 50% of these specimens, a representative sample from the weekly gravid traps, were identified by PCR and 98% of those were Cx. pipiens. In 2006, a randomly selected sample produced similar results. Accordingly, the gravid trap collection was considered primarily Cx. pipiens. More than 97% of all female Culex caught in CDC light traps were identified by PCR. Overall, Cx. pipiens was the predominant species with 55% in 2005 and 69% in 2006. Culex restuans percent composition decreased from 35% in 2005 to 22% in 2006, and that of Cx. salinarius remained at approximately 10% each year.

In a multivariate model with *Culex* species abundance as the dependent variable, the main effects of year, site, and LUC, as well as the interaction effect of site × LUC were significant (Table 4). Significantly fewer *Cx. restuans* were caught in 2006 than in 2005. The abundance of *Cx. pipiens* and *Cx. salinarius* differed among the sites, with significantly more *Cx. pipiens* collected at site 2 and significantly fewer *Cx. salinarius* caught at site 1 (Tables 4 and 5). *Culex pipiens* and *Cx. restuans* had significantly higher abundance in residential LUC, followed by commercial and natural areas (Tables 4 and 5).

In terms of *Culex* species composition, *Cx. pipiens* was significantly overrepresented at site 2, and underrepresented at sites 1 and 3, whereas *Cx. restuans* was significantly underrepresented at site 2 and overrepresented at site 1 (Table 5). *Culex salinarius* was significantly overrepresented at site 3, and underrepresented at sites 1 and 2. *Culex restuans* and *Cx. salinarius* were more prevalent in natural areas compared with *Cx. pipiens*, which had significantly higher percent composition in commercial areas. In addition, *Cx. restuans* was significantly overrepresented in residential areas.

Populations of *Cx. pipiens* were characterized by several spikes of approximately 15–20 of *Cx. pipiens* females per trap night. *Culex restuans* populations reached approximately 12–15 mosquitoes per trap night during the early summer, decreasing

Table 3

Combined *Culex* spp. collected in gravid (G) and Centers for Disease Control and Prevention light (L) traps, Suffolk County, New York, during the 2005–2006 study period*

Year	Trap	No. collected	No. tested (%)	No. identified (%)	PIP, no. (%)	RES, no. (%)	SAL, no. (%)
2005	G	5,887	2,898 (49.2)	2,782 (96.4)	2,729 (98.1)	44 (1.6)	9 (0.3)
2006	G	5,215	83 (1.6)	82 (98.8)	81 (98.8)	1 (1.2)	0
2005	L	2,124	2,095 (98.6)	2,023 (96.5)	1,117 (55.2)	696 (34.4)	210 (10.4)
2006	L	2,076	2,006 (96.6)	1,953 (97.4)	1,341 (68.7)	428 (21.9)	184 (9.4)

^{*} PIP = $Culex\ pipiens$; RES = $Cx.\ restuans$; SAL = $Cx.\ salinarius$.

TABLE 4

Multivariate analysis of variance test for variability in *Culex pipiens* (PIP), *Cx. restuans* (RES), and *Cx. salinarius* (SAL) abundance (dependent variables) as a function of years, site, and land use/cover (LUC) type (independent variables), Suffolk County, New York*

Source		Wilks λ	Hypothetical df	Error df	F	P
Intercept		0.23	3	304	343.23	0.001
Year		0.96	3	304	3.68	0.012
Site		0.80	6	608	12.21	< 0.001
LUC		0.70	6	608	19.71	< 0.001
Year × site		0.96	6	608	2.30	0.033
Year × LUC		0.98	6	608	0.82	0.551
Site \times LUC		0.82	12	805	5.18	< 0.001
Year × site ×	LUC	0.95	12	805	1.19	0.284
Between sub	jects					
Year	PIP	NA	1	306	0.01	0.918
	RES	NA	1	306	7.52	0.006
	SAL	NA	1	306	0.40	0.530
Site	PIP	NA	2	306	26.41	< 0.001
	RES	NA	2	306	0.20	0.816
	SAL	NA	2	306	9.39	< 0.001
LUC	PIP	NA	2	306	43.78	< 0.001
	RES	NA	2	306	23.48	< 0.001
	SAL	NA	2	306	0.27	0.766
Site * LUC	PIP	NA	4	306	6.46	< 0.001
	RES	NA	4	306	3.48	0.008
	SAL	NA	4	306	8.25	< 0.001

^{*}Statistically significant results are indicated in **bold** letters and numbers. df = degrees of freedom; NA = not applicable.

to fewer than 5 by the late summer (Figure 2). Culex salinarius populations gradually increased to approximately 5 per trap night by mid-August and then decreased. Analysis of the combined data by week showed some variation among the three sites (Figure 3). Site 2 was dominated by Cx. pipiens, whereas Culex restuans was more common at sites 1 and 3 especially in June and July. Culex salinarius was more abundant with higher percent composition at site 3, where it reached up to 40% of the total Culex species by late summer.

Activity of West Nile virus. In 2005, a total of 37 Culex WNV positive pools were collected from the study sites, with the first pool obtained on July 12. A single non-Culex positive pool containing Uranotaenia sapphirina was collected at site 3 (natural LUC). The Culex WNV-positive pool ratio (site 1/site 2/site 3) was 4/8/2 for light traps and 8/10/5 for gravid traps, respectively. The MIR curve for Culex species displayed two peaks, a smaller one in the late August and a higher one in mid-September (Figure 2). In 2006, a total of 23 Culex WNVpositive pools were collected, with the first pool obtained on July 11, with no non-Culex positive pools. The Culex WNVpositive pool ratio (site 1/site 2/site 3) was 2/5/3 for light traps and 3/6/4 for gravid traps. The MIR curve for Culex species displayed three peaks, a smaller peak in the late July, a higher peak in the late August, and the highest peak in mid-September (Figure 2). The total number of mosquito pools collected for the study and during routine surveillance and number of positive birds obtained in 2005–2006 are shown in Table 1. No equine cases occurred during the study period, most likely as a result of equine WNV vaccine, which was introduced in 2001. This intervention was followed by a steep decrease in equine WNV cases in Suffolk County to 4 in 2002, 3 in 2003, 1 in 2004, and 0 in 2005–2006. One human case occurred within area 1 in 2005 (Figure 1); an additional human case was reported in close proximity south of the area's boundary (approximately 0.5 miles). The closest human case to area 2 was that of area 1, whereas the closest human case to area 3 occurred approximately 2.5 miles south of the area's boundary.

DISCUSSION

In the northeastern United States, Cx. pipiens and Cx. restuans are commonly recognized as primary enzootic vectors of WNV, whereas Cx. pipiens and Cx. salinarius have been proposed as the main epidemic vectors of the virus.^{2,4,6,9,10,36} The risk of human infection from a bridge vector is directly associated with its relative or average abundance, fraction of blood meals taken from mammals, vector competence, and WNV prevalence.^{9,37} Similarly, the vectorial capacity of a bridge vector is postulated to increase with its population density and human blood index, i.e., the proportion of blood meals taken from humans. 38,39 Because the most important risk factor for acquiring WNV is exposure to infected mosquitoes,1 the human risk should increase proportionately to the species' vectorial capacity and thus abundance,38 as demonstrated for some arboviruses (including WNV) in the field. 21,40,41 For example, a Colorado study found that census tracts with increased Cx. tarsalis abundance also had higher WNV disease incidence.²¹ In this study, the level of WNV human risk at a particular location was defined by the CDC guidelines²³ as high (epidemic transmission with recurring human and equine cases, site 1), moderate (epizootic transmission, equine cases only, site 3), or low (enzootic transmission, positive birds only, site 2). The spatial extent of these areas was determined by spatial cluster detection software²² and represented a subcounty scale, which enables optimal characterization of the spatial variability of WNV risk.16 Accordingly, certain epidemiologic predictions could be examined in light of the two main hypotheses proposing a single Culex species, either Cx. pipiens, 10 or Cx. salinarius, 2,4,6,7 as the main WNV bridge vector (and thus WNV risk factor) in our region.

The probability that a species of mosquito will infect a human with WNV, i.e., human risk = $A \times F_m \times P \times C_v$, where A is the species proportion or percent composition (termed relative abundance by the authors), F_m is a fraction of mammalian blood meals, P is WNV prevalence, and C_v is vector competence. Given the study sites geographic proximity, F_m and C_v were assumed similar, and P did not differ significantly among the sites, and was, in fact, slightly higher at the enzootic site. Thus, the location with elevated human risk (site 1) is expected to have a higher proportion of the mosquito species serving as a bridge vector according to this model. This epidemiologic assumption was found to be incorrect for both Cx. pipiens and Cx. salinarius in this study. Because Cx. pipiens was the predominant mosquito at all three sites, a cursory analysis might implicate this species as the primary vector. However, its distribution was not correlated with WNV activity, and epidemic activity was greatest where this species was least abundant. Site 2, where Cx. pipiens was most abundant, also had the highest number of WNV isolations, mostly from Cx. pipiens in gravid traps. These findings are indicative of increased vectorial capacity of Cx. pipiens and together with WNV competence of this species' local populations42 and increased abundance in the residential areas would be expected to lead to a higher human risk of exposure to WNV.26 Additionally, if the same vector contributes to the mammalian and avian cycles, pronounced disease activity is likely where this species

Table 5

Culex pipiens, Cx. restuans, and Cx. salinarius abundance and composition by site and by land use/cover type (LUC), Suffolk County, New York*

			Site			LUC	
Species		1	2	3	Comm	Nat	Resd
Cx. pipiens	No. collected	517	1,264†	677	860†	357†	1,241†
• •	% within site/LUC	55	70	55	73	47	61
	Standardized residuals	-2.7	4.5	-3.1	5.1	-5.1	-0.7
Cx. restuans	No. collected	371	386	367	184	282	658†
	% within site/LUC	39	21	30	16	37	32
	Standardized residuals	6.4	-5.4	1.0	-8.1	4.6	3.3
Cx. salinarius	No. collected	55†	150	189	127	118	149
	% within site/LUC	6	9	15	11	16	7
	Standardized residuals	-4.0	-2.1	6.0	1.0	5.0	-3.8
Total	No. collected	943	1,800	1,233	1,171	757	2,048

^{*} No. collected = total numbers of collected specimens identified by polymerase chain reaction. To obtain mean trap night catch, each number should be divided by 36 (no. of trap weeks). Percent species composition within each site or LUC category and standardized residuals are shown. Results with statistically significant difference (standardized residuals > 2.0) for species percent composition within each site or LUC categories are indicated by **bold** standardized residuals. Comm = commercial areas; Nat = natural areas; Resd = residential areas. † Statistically significant differences (P < 0.05) in the abundance of each species among site or LUC categories by Dunnett's T3 post hoc test.

is abundant.³ However, of the three sites, site 2 had the lowest WNV activity among mammals. One possible explanation for this result is that host preferences of *Cx. pipiens* have been found to be highly ornithophilic in this region, ^{26,31,36,43–45} supporting *Cx. pipiens* as the main enzootic vector of WNV in Suffolk County.

Another potential bridge vector, *Cx. salinarius*, has exhibited WNV competence in the laboratory,⁸ often harbors the virus in Connecticut^{2,6} and New York,^{4,7} and is described as an indiscriminate feeder on a wide range of mammalian and avian hosts in this region.^{26,31,36,43,44} However, this species was also least abundant at the site with greatest WNV epidemic activity (site 1), and higher abundance and percent composition of *Cx. salinarius* were not associated with elevated risk of human infection at either sites 2 or 3. White-tailed deer, a large mammal, has been identified as the source of approximately two-thirds of *Cx. salinarius* mammalian blood meals.^{36,43,44}

Feeding preponderance of this species for large domestic animals in the vicinity of natural wetlands was also noted in some studies.⁴⁶ *Culex salinarius* was overrepresented at the study area (site 3) with numerous horse properties adjoining extensive and deer-free parkland (i.e., natural areas), which may account, in part, for the elevated number of WNV equine cases seen in that area before the vaccine was introduced. Conversely, this vector species may play a greater role in WNV epidemic transmission in coastal areas of the Suffolk County's south shore, where a second most probable cluster of WNV human cases was located. These areas also support extensive tidal salt marshes, the main *Cx. salinarius* larval habitat in Suffolk County, and therefore experience much higher adult abundance of this species.⁴⁷

Unlike *Cx. pipiens* and *Cx. salinarius*, the highest percent composition of *Cx. restuans* was observed at the epidemic site (site 1) and the lowest at the enzootic site (site 3) corresponding

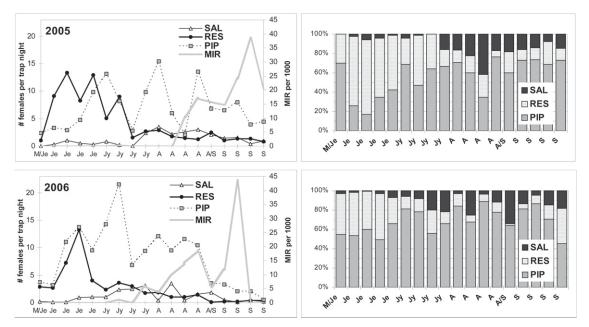


FIGURE 2. Left, Weekly average trap night catch and minimum infection rate (MIR) per 1,000 females for all sites combined in each year (2005 or 2006). Right, Percent species composition of *Culex pipiens* (PIP), *Cx. restuans* (RES), and *Cx. salinarius* (SAL) in CDC light traps in each year. M = May; Je = June; Jy = July; A = August; S = September.

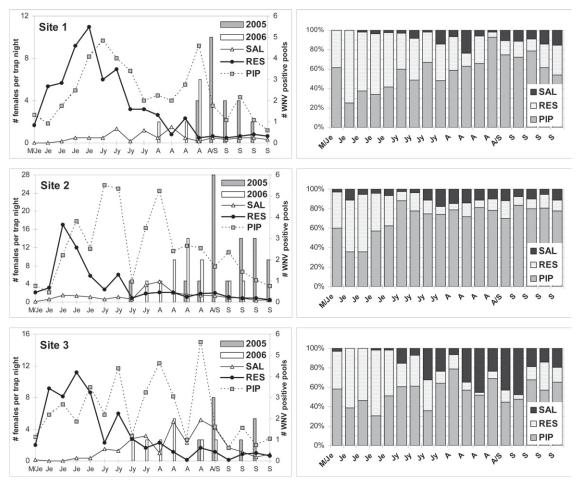


FIGURE 3. Left, CDC light trap weekly average trap night catch and number of West Nile virus (WNV)—positive pools (left). Right, Corresponding percent species composition by site for three *Culex* species: *Cx. pipiens* (PIP), *Cx. restuans* (RES), and *Cx. salinarius* (SAL). On the left, average trap night catch is indicated by lines, and number of WNV-positive pools by bars. Trap night catch and percent composition data were combined for 2005 and 2006. M = May; Je = June; Jy = July; A = August; S = September.

to the levels of human risk predicted by the model. Although much higher in June and July, Cx. restuans abundance was comparable with that of Cx. salinarius during the peak WNV transmission in August and September (Figure 3), especially at sites 1 and 2 with smaller natural areas and located further inland. Like Cx. pipiens, Cx. restuans was significantly more abundant in residential areas, where larvae of both species were frequently found in groundwater retention basins (Rochlin I, Campbell SR, unpublished data). The local populations of Cx. restuans were vector competent for WNV,42 and the virus has frequently been detected in field collected specimens from New York32 and Connecticut.2,6 However, this species is not generally considered a potential epidemic vector³⁶ because of its primarily early season activity and blood meal preferences. 26,36,43 The host preference results that suggest ornithophily are not unequivocal; mammalian feeding was observed in New York, 31,45 and mammalian blood was found in approximately 15-30% of the Cx. restuans specimens (a higher proportion than in Cx. pipiens) in the same studies. 26,43 The limited sample size of 10–40 blooded specimens in these studies raises the possibility of selection and technical (i.e., preferential amplification) biases. Additionally, Cx. restuans adults are often morphologically indistinguishable from those of Cx. pipiens, 30,31 which might have led to confusion between the two species in the past.^{30,31} Abundance data from this study, both spatial (high versus low human risk sites) and temporal (*Cx. restuans* was less numerous during 2006 when lower WNV activity was also recorded) clearly indicates a continuing need for more research on the role of this species in WNV transmission.

Our analysis focused on vector abundance and viral prevalence as the key components of vectorial capacity and human risk, and other factors, such as feeding preferences, local ecology, and human behavior, were assumed to have little variation among the three study sites caused by geographic proximity, similar LUC composition, and comparable human population density. The apparent lack of association between the risk of WNV epidemic transmission and the abundance of *Cx. pipiens* and *Cx. salinarius* at these three sites raises important questions about these species' role as the only primary epidemic vectors and suggests additional variables that might determine human risk.

One possibility is the presence of additional mosquito vectors, such as *Ae. vexans*, *Ae. japonicus*, *Ae. sollicitans*, *Ae. triseriatus*, and *Ae. trivittatus*, ^{9,48} (found in low numbers at our study sites), which therefore might transmit WNV from birds to humans under favorable environmental and demographic conditions.¹³ Environmental heterogeneity over

spatial micro-scales might have been another factor. Droughts were found to induce WNV amplification in Florida by bringing the hosts and the vectors together.⁴⁹ A similar process might have occurred in our study, where the epidemic site's small and isolated but well preserved natural areas with wetlands served as focal points for birds and mosquitoes. These wetland areas were also surrounded by heavily residential areas, potentially leading to greater human exposure, and more extensive and contagious parkland at the epizootic site or dry forested habitat at the enzootic site did not provide the same level of human exposure. Subtle socioeconomic, demographic, and behavioral differences not captured by human population density might also have contributed to the different transmission patterns.^{18,50}

A combination of unique environmental and demographic factors was more likely as an underlying cause of conditions appropriate for human transmission than was the presence of a particular bridge vector species. West Nile virus human infection occurs sporadically in Suffolk County despite high levels of enzootic viral activity, suggesting that conditions conducive to viral epidemic transmission are also intermittent, and that they may change through time. Epidemiologic and ecologic research on a subcounty level has been proposed as a priority for nationwide development to elucidate the spatial patterns of vector-borne disease risk. ¹⁸ Consequently, this study should contribute to our understanding of WNV risk factors and to develop strategies for disruption and prevention of WNV epidemic transmission.

Received May 13, 2008. Accepted for publication September 30, 2008.

Acknowledgments: We thank Melissa Zanini (site selection, mosquito collection/identification, and molecular species identification), Kerri Harding (geocoding, mosquito pool processing and database management), and Ralph Narain (molecular species identification) for excellent technical assistance; N. Petti, R. Chayes, C. Provenzano, and A. Culkin for assistance in processing mosquitoes; Ward Stone and the staff of the New York State Department of Environmental Conservation Wildlife Pathology Unit for bird processing and analysis; Laura Kramer and the staff of the New York State Department of Health Arboviruses Laboratories for positive bird and mosquito pool processing and analysis; and Richard Mayer (U.S. Department of Agriculture) for advice on the study.

Financial support: This study was partially supported by Specific Cooperative Agreement #58-5410-4-338 from the United States Department of Agriculture.

Authors' addresses: Ilia Rochlin, Division of Vector Control, Suffolk County Department of Public Works, 335 Yaphank Avenue, Yaphank, NY 11980-9744, E-mail: ilia.rochlin@suffolkcountyny.gov. Howard S. Ginsberg, U.S. Geological Survey Patuxent Wildlife Research Center, Coastal Field Station, Woodward Hall-PLS, University of Rhode Island, Kingston, RI 02881, E-mail: hginsberg@usgs.gov. Scott R. Campbell, Arthropod-Borne Disease Laboratory, Suffolk County Department of Health Services, 335 Yaphank Avenue, Yaphank, NY 11980-9744, E-mail: scott.campbell@suffolkcountyny.gov.

REFERENCES

- Hayes EB, Komar N, Nasci RS, Montgomery SP, O'Leary DR, Campbell GL, 2005. Epidemiology and transmission dynamics of West Nile virus disease. *Emerg Infect Dis* 11: 1167–1173.
- Andreadis TG, Anderson JF, Vossbrinck CR, Main AJ, 2004. Epidemiology of West Nile virus in Connecticut: a five-year analysis of mosquito data 1999–2003. Vector Borne Zoonotic Dis 4: 360–378.
- 3. Gould LH, Fikrig E, 2004. West Nile virus: a growing concern? *J Clin Invest 113*: 1102–1107.

- Kulasekera VL, Kramer L, Nasci RS, Mostashari F, Cherry B, Trock SC, Glaser C, Miller JR, 2001. West Nile virus infection in mosquitoes, birds, horses, and humans, Staten Island, New York, 2000. Emerg Infect Dis 7: 722–725.
- Kramer LD, Styer LM, Ebel GD, 2008. A global perspective on the epidemiology of West Nile virus. *Annu Rev Entomol* 53: 61–81.
- Andreadis TG, Anderson JF, Vossbrinck CR, 2001. Mosquito surveillance for West Nile virus in Connecticut, 2000: isolation from Culex pipiens, Cx. restuans, Cx. salinarius, and Culiseta melanura. Emerg Infect Dis 7: 670–674.
- Bernard KA, Maffei JG, Jones SA, Kauffman EB, Ebel G, Dupuis AP, Ngo KA, Nicholas DC, Young DM, Shi PY, Kulasekera VL, Eidson M, White DJ, Stone WB, Kramer LD, 2001. West Nile virus infection in birds and mosquitoes, New York State, 2000. Emerg Infect Dis 7: 679–685.
- Sardelis MR, Turell MJ, Dohm DJ, O'Guinn M, 2001. Vector competence of selected North American Culex and Coquillettidia mosquitoes for West Nile virus. Emerg Infect Dis 7: 1018–1022.
- 9. Kilpatrick AM, Kramer LD, Campbell SR, Alleyne EO, Dobson AP, Daszak P, 2005. West Nile virus risk assessment and the bridge vector paradigm. *Emerg Infect Dis* 11: 425–429.
- Kilpatrick AM, Kramer LD, Jones MJ, Marra PP, Daszak P, 2006. West Nile virus epidemics in North America are driven by shifts in mosquito feeding behavior. PLoS Biol 4: e82.
- Diuk-Wasser MA, Brown HE, Andreadis TG, Fish D, 2006. Modeling the spatial distribution of mosquito vectors for West Nile virus in Connecticut, USA. Vector Borne Zoonotic Dis 6: 283–295.
- Molaei G, Andreadis TG, 2006. Identification of avian- and mammalian-derived bloodmeals in *Aedes vexans* and *Culiseta melanura* (Diptera: Culicidae) and its implication for West Nile virus transmission in Connecticut, USA. *J Med Entomol 43*: 1088–1093.
- Gingrich JB, Williams GM, 2005. Host-feeding patterns of suspected West Nile virus mosquito vectors in Delaware, 2001–2002. J Am Mosq Control Assoc 21: 194–200.
- Kunkel KE, Novak RJ, Lampman RL, Gu W, 2006. Modeling the impact of variable climatic factors on the crossover of *Culex restuans* and *Culex pipiens* (Diptera: Culicidae), vectors of West Nile virus in Illinois. *Am J Trop Med Hyg 74*: 168–173.
- Yiannakoulias NW, Svenson LW, 2007. West Nile virus: strategies for predicting municipal-level infection. Ann NY Acad Sci 1102: 135–148
- Eisen L, Eisen RJ, 2007. Need for improved methods to collect and present spatial epidemiologic data for vectorborne diseases. *Emerg Infect Dis* 13: 1816–1820.
- 17. Bertolotti L, Kitron UD, Walker ED, Ruiz MO, Brawn JD, Loss SR, Hamer GL, Goldberg TL, 2008. Fine-scale genetic variation and evolution of West Nile Virus in a transmission "hot spot" in suburban Chicago, USA. *Virology 374:* 381–389.
- Eisen RJ, Eisen L, 2008. Spatial modeling of human risk of exposure to vector-borne pathogens based on epidemiological versus arthropod vector data. *J Med Entomol* 45: 181–192.
- Hamer GL, Walker ED, Brawn JD, Loss SR, Ruiz MO, Goldberg TL, Schotthoefer AM, Brown WM, Wheeler E, Kitron UD, 2008. Rapid amplification of West Nile virus: the role of hatchyear birds. Vector Borne Zoonotic Dis 8: 57–67.
- Ruiz MO, Walker ED, Foster ES, Haramis LD, Kitron UD, 2007.
 Association of West Nile virus illness and urban landscapes in Chicago and Detroit. Int J Health Geogr 6: 10–20.
- Winters AM, Bolling BG, Beaty BJ, Blair CD, Eisen RJ, Meyer AM, Pape WJ, Moore CG, Eisen L, 2008. Combining mosquito vector and human disease data for improved assessment of spatial West Nile virus disease risk. Am J Trop Med Hyg 78: 654–665.
- 22. Kulldorff M, 1997. A spatial scan statistic. Comm Statist Theory Methods 26: 1481–1496.
- 23. Centers for Disease Control and Prevention, 2003. *Epidemic/Epizootic West Nile Virus in the United States: Revised Guidelines for Surveillance, Prevention, and Control*. Available at: http://www.cdc.gov/ncidod/dvbid/westnile/surv&control.htm. Accessed March 11, 2008.
- Cornell University Geospatial Information Repository (database on the Internet). Available at: http://cugir.mannlib.cornell.edu. Accessed February 19, 2008.

- New York State Geographic Information Systems (database on the Internet). New York State GIS Clearinghouse. Suffolk County Direct Download (2001). Available at: http://www.nysgis.state. ny.us/gateway/mg/2001/Suffolk. Accessed February 19, 2008.
- 26. Apperson CS, Harrison BA, Unnasch TR, Hassan HK, Irby WS, Savage HM, Aspen SE, Watson DW, Rueda LM, Engber BR, Nasci RS, 2002. Host-feeding habits of *Culex* and other mosquitoes (Diptera: Culicidae) in the borough of Queens in New York City, with characters and techniques for identification of *Culex* mosquitoes. *J Med Entomol 39: 777–785*.
- Rochlin I, Santoriello MP, Mayer RT, Campbell SR, 2007. Improved high-throughput method for molecular identification of *Culex* mosquitoes. J Am Mosq Control Assoc 23: 488–491.
- 28. Corsaro BG, Munstermann LE, 1984. Identification by electrophoresis of *Culex* adults (Diptera: Culicidae) in light-trap samples. *J Med Entomol* 21: 648–655.
- Crabtree MB, Savage HM, Miller BR, 1995. Development of a species-diagnostic polymerase chain reaction assay for the identification of *Culex* vectors of St. Louis encephalitis virus based on interspecies variation in ribosomal DNA spacers. *Am J Trop Med Hyg 53*: 105–109.
- Harrington LC, Poulson RL, 2008. Considerations for accurate identification of adult *Culex restuans* (Diptera: Culicidae) in field studies. *J Med Entomol* 45: 1–8.
- Means RG, 1987. Mosquitoes of New York. Part II. Genera of Culicidae other than Aedes occurring in New York. New York State Mus Bull 430b: 1–180.
- Lukacik G, Anand M, Shusas EJ, Howard JJ, Oliver J, Chen H, Backenson PB, Kauffman EB, Bernard KA, Kramer LD, White DJ, 2006. West Nile virus surveillance in mosquitoes in New York State, 2000–2004. J Am Mosq Control Assoc 22: 264–271.
- 33. Bernard KA, Kramer LD, 2001. West Nile virus activity in the United States, 2001. *Viral Immunol 14*: 319–338.
- Biggerstaff B, Centers for Disease Control and Prevention, Software for Mosquito Surveillance. Available at: www.cdc.gov/ncidod/ dvbid/westnile/software.htm. Accessed February 19, 2008.
- 35. Lawal BH, 2003. Categorical Data Analysis with SAS and SPSS Applications. Mahwah: Lawrence Erlbaum Associates.
- Molaei G, Andreadis TG, Armstrong PM, Anderson JF, Vossbrinck CR, 2006. Host feeding patterns of *Culex* mosquitoes and West Nile virus transmission, northeastern United States. *Emerg Infect Dis* 12: 468–474.
- 37. Moncayo AC, Edman JD, 1999. Toward the incrimination of epidemic vectors of eastern equine encephalomyelitis virus in

- Massachusetts: abundance of mosquito populations at epidemic foci. *J Am Mosq Control Assoc 15*: 479–492.
- 38. Spielman A, 1999. The role of surveillance in interventions directed against vector-borne disease. *Ecosyst Health 5*: 141–145.
- Freier JE, 1989. Estimation of vectorial capacity: vector abundance in relation to man. Bull Soc Vector Ecol 14: 41–46.
- Hu W, Tong S, Mengersen K, Oldenburg B, Dale P, 2006. Mosquito species (Diptera: Culicidae) and the transmission of Ross River virus in Brisbane, Australia. *J Med Entomol* 43: 375–381.
- Ryan PA, Do KA, Kay BH, 1999. Spatial and temporal analysis of Ross River virus disease patterns at Maroochy Shire, Australia: association between human morbidity and mosquito (Diptera: Culicidae) abundance. *J Med Entomol* 36: 515–521.
- Ebel GD, Rochlin I, Longacker J, Kramer LD, 2005. Culex restuans (Diptera: Culicidae) relative abundance and vector competence for West Nile Virus. J Med Entomol 42: 838–843.
- 43. Apperson CS, Hassan HK, Harrison BA, Savage HM, Aspen SE, Farajollahi A, Crans W, Daniels TJ, Falco RC, Benedict M, Anderson M, McMillen L, Unnasch TR, 2004. Host feeding patterns of established and potential mosquito vectors of West Nile virus in the eastern United States. Vector Borne Zoonotic Dis 4: 71–82.
- Crans WJ, 1964. Continued host preference studies with New Jersey mosquitoes, 1963. Proc NJ Mosq Control Assoc 51: 50–58.
- Means RG, 1968. Host preferences of mosquitoes (Diptera: Culicidae) in Suffolk County, New York. Ann Entomol Soc Am 61: 116–120.
- LeDuc JW, Suyemoto W, Eldridge BF, Saugstad ES, 1972. Ecology of arboviruses in a Maryland freshwater swamp. II. Blood feeding patterns of potential mosquito vectors. Am J Epidemiol 96: 123–128.
- Rochlin I, Dempsey ME, Campbell SR, Ninivaggi DV, 2008. Salt marsh as *Culex salinarius* larval habitat in coastal New York. *J Am Mosq Control Assoc 24*: 359–367.
- Turell MJ, Dohm DJ, Sardelis MR, Oguinn ML, Andreadis TG, Blow JA, 2005. An update on the potential of North American mosquitoes (Diptera: Culicidae) to transmit West Nile virus. J Med Entomol 42: 57–62.
- 49. Shaman J, Day JF, Stieglitz M, 2005. Drought-induced amplification and epidemic transmission of West Nile virus in southern Florida. *J Med Entomol* 42: 134–141.
- Rios J, Hacker CS, Hailey CA, Parsons RE, 2006. Demographic and spatial analysis of West Nile virus and St. Louis encephalitis in Houston, Texas. J Am Mosq Control Assoc 22: 254–263.