

THE UNIVERSITY OF RHODE ISLAND

University of Rhode Island DigitalCommons@URI

Fisheries, Animal and Veterinary Sciences Faculty Publications

Fisheries, Animal and Veterinary Sciences

2018

Draft Genome Sequence of Loktanella Maritima Strain YPC211, a Commensal Bacterium of the American Lobster (*Homarus Americanus*)

Hilary J. Ranson *University of Rhode Island*, hranson@uri.edu

Jason LaPorte
University of Rhode Island

See next page for additional authors

Creative Commons License

Creative Commons License

This work is licensed under a Creative Commons Attribution 4.0 License.

Follow this and additional works at: https://digitalcommons.uri.edu/favs facpubs

Citation/Publisher Attribution

Ranson, H. J., LaPorte, J., Spinard, E., Gómez-chiarri, M., Nelson, D. R., Rowley, D. C. (2018). Draft genome sequence of Loktanella maritima strain YPC211, a commensal bacterium of the American lobster (*Homarus americanus*). *Genome Announcements*, 6(18), art. no. e00314-18. doi: 10.1128/genomeA.00314-18

Available at: http://dx.doi.org/10.1128/genomeA.00314-18

This Article is brought to you for free and open access by the Fisheries, Animal and Veterinary Sciences at DigitalCommons@URI. It has been accepted for inclusion in Fisheries, Animal and Veterinary Sciences Faculty Publications by an authorized administrator of DigitalCommons@URI. For more information, please contact digitalcommons@etal.uri.edu.









Draft Genome Sequence of Loktanella maritima Strain YPC211, a Commensal Bacterium of the American Lobster (Homarus americanus)

Hilary J. Ranson, a Jason LaPorte, b Edward Spinard, b Marta Gomez-Chiarri, c David R. Nelson, b David C. Rowleya

ABSTRACT Loktanella maritima strain YPC211 was isolated from the American lobster (*Homarus americanus*). We report here the draft genome sequence for *L. maritima* YPC211 and identify genes of potential importance to its role within the microbial community.

oktanella maritima is a Gram-negative aerobic bacterium that was originally isolated from shallow marine sediments in the Sea of Japan (1). Loktanella spp. appear to be ubiquitously distributed in the oceans and have been isolated from Antarctic to subtropical environments (2, 3). The genus belongs to the class Alphaproteobacteria, family Rhodobacterales, and order Rhodobacteraceae. Van Trappen et al. (2) originally proposed the genus to accommodate 3 species, but it has since been expanded and modified to contain 15 species (4–6). The strain Ha06YPC211 produces a beige/yellow pigment and was isolated from an egg mass of an American lobster (Homarus americanus) from Jamestown, RI.

L. maritima strain YPC211 was grown in artificial seawater (Instant Ocean) supplemented with yeast extract (1 g/liter) and peptone (5 g/liter) at 25°C on an elliptical shaker (New Brunswick) for 24 h. Genomic DNA was isolated using the Promega Wizard DNA purification kit, and DNA was resuspended in 2 mM Tris-HCl buffer (Bio Basic, Inc.). DNA was quantified using a NanoDrop 1000 spectrophotometer (ND-1000) and checked for quality on a 1% agarose gel stained with ethidium bromide. The DNA was sequenced on an Illumina MiSeq sequencer at the Genomics and Sequencing Center at University of Rhode Island. Reads were trimmed using CLC Genomic Workbench (version 8.5.1) for quality, ambiguous base pairs, adaptors, duplicates, and size, resulting in 4,491,164 paired-end reads. The draft genome was assembled using the de novo assembly algorithm of the CLC Genomic Workbench and SPAdes assembler (version 3.1.1). Contigs with a coverage of >76 reads were processed using the CLC Microbial Genome Finishing module. The completed draft genome is composed of 13 contigs, averaging 264,067 bp in size and 3 plasmids of 23,839, 6,277, and 215,371 bp in size (total genome, 3,678,360 bp), with an average G+C content of 53.5%. The draft genome was annotated using the Rapid Annotations using Subsystems Technology (RAST) server and resulted in 3,681 open reading frames (7).

The genome of *L. maritima* YPC211 encodes type I, II, III, and IV secretion systems. It has been reported that *Loktanella* spp. can survive temperature extremes and have been isolated from Antarctic mats (2). Accordingly, five putative cold shock proteins (including CspA, CspB, and CspC) were annotated. Additionally, the genome encodes four hemolysins, two metalloproteases, and one extracellular protease. Two iron ac-

Received 14 March 2018 **Accepted** 24 March 2018 **Published** 3 May 2018

Citation Ranson HJ, LaPorte J, Spinard E, Gomez-Chiarri M, Nelson DR, Rowley DC. 2018. Draft genome sequence of *Loktanella maritima* strain YPC211, a commensal bacterium of the American lobster (*Homarus americanus*). Genome Announc 6:e00314-18. https://doi .org/10.1128/genomeA.00314-18.

Copyright © 2018 Ranson et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to David C. Rowley, drowley@uri.edu.

^aDepartment of Biomedical and Pharmaceutical Sciences, University of Rhode Island, Kingston, Rhode Island, USA

^bDepartment of Cell and Molecular Biology, University of Rhode Island, Kingston, Rhode Island, USA

^cDepartment of Fisheries, Animal and Veterinary Sciences, University of Rhode Island, Kingston, Rhode Island, LUSA

quisition systems, TonB and hemin (HmuS), were annotated along with iron-specific hemin and ABC transporters. Ferric siderophore-related genes were also identified. The RAST-annotated draft genome was submitted to Antibiotics and Secondary Metabolite Analysis Shell (antiSMASH) for secondary metabolite biosynthesis gene cluster analysis (8), leading to the identification of four clusters, including a nonribosomal peptide synthetase, type I polyketide synthase, bacteriocin, and homoserine lactone biosynthetic gene clusters. L. maritima YPC211 inhibits the growth of the shrimp pathogen Vibrio parahaemolyticus when cocultivated on an agar surface, as determined by a zone of inhibition assay (9). Further investigation of these biosynthetic gene clusters may aid in characterization of compounds responsible for this antibiosis.

Accession number(s). This whole-genome shotgun project has been deposited in DDBJ/ENA/GenBank under the accession number PKFN00000000. The version described in this paper is the first version, PKFN01000000.

ACKNOWLEDGMENTS

This work was supported by funding from the Rhode Island Sea Grant (grant RISG18-R/F-1618-31-1. This research is based upon the work conducted using the Rhode Island Genomics and Sequencing Center, which is supported in part by the National Science Foundation under EPSCoR grants 0554548 and EPS-1004057.

REFERENCES

- 1. Tanaka N, Romanenko LA, Kurilenko VV, Svetashev VI, Kalinovskaya NI, Mikhailov VV. 2014. Loktanella maritima sp. nov. isolated from shallow marine sediments. Int J Syst Evol Microbiol 64:2370-2375. https://doi.org/ 10.1099/ijs.0.061747-0.
- 2. Van Trappen S, Mergaert J, Swings J. 2004. Loktanella salsilacus gen. nov., sp. nov., Loktanella fryxellensis sp. nov. and Loktanella vestfoldensis sp. nov., new members of the Rhodobacter group, isolated from microbial mats in Antarctic Lakes. Int J Syst Evol Microbiol 54:1263-1269. https:// doi.org/10.1099/iis.0.03006-0.
- 3. Lau SCK, Tsoi MMY, Li X, Plakhotnikova I, Wu M, Wong P-K, Qian P-Y. 2004. Loktanella hongkongensis sp. nov., a novel member of the α -Proteobacteria originating from the marine biofilms in Hong Kong waters. Int J Syst Evol Microbiol 54:2281-2284. https://doi.org/10.1099/ijs.0.63294-0.
- 4. Moon YG, Seo SH, Lee SD, Heo MS. 2010. Loktanella pyoseonensis sp. nov., isolated from beach sand, and emended description of the genus Loktanella. Int J Syst Evol Microbiol 60:785-789. https://doi.org/10.1099/ijs.0 .011072-0
- 5. Lee SD. 2012. Loktanella tamlensis sp. nov., isolated from seawater. Int ${\sf J}$ Syst Evol Microbiol 62:586-590. https://doi.org/10.1099/ijs.0.029462-0.
- 6. Tsubouchi T, Shimane Y, Mori K, Miyazaki M, Tame A, Uematsu K, Maruy-

- ama T, Hatada Y. 2013. Loktanella cinnabarina sp. nov., isolated from a deep subseafloor sediment, and emended description of the genus Loktanella. Int J Syst Evol Microbiol 63:1390-1395. https://doi.org/10.1099/ ijs.0.043174-0.
- 7. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O, 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. https://doi.org/10.1186/1471-2164-9-75.
- 8. Weber T, Blin K, Duddela S, Krug D, Kim HU, Bruccoleri R, Lee SY, Fischbach MA, Müller R, Wohlleben W, Breitling R, Takano E, Medema MH. 2015. antiSMASH 3.0—a comprehensive resource for the genome mining of biosynthetic gene clusters. Nucleic Acids Res 43:W237-W243. https:// doi.org/10.1093/nar/gkv437.
- 9. Zhao W, Dao C, Karim M, Gomez-Chiarri M, Rowley D, Nelson DR. 2016. Contributions of tropodithietic acid and biofilm formation to the probiotic activity of Phaeobacter inhibens. BMC Microbiol 16:1. https://doi.org/ 10.1186/s12866-015-0617-z.