



THE AGA KHAN UNIVERSITY

eCommons@AKU

# Department of Biological & Biomedical Sciences

Medical College, Pakistan

9-2019

# Colostrum and mature breast milk analysis of serum irisin and sterol regulatory element-binding proteins-1c in gestational diabetes mellitus

Syeda Sadia Fatima Aga Khan University, sadia.fatima@aku.edu

Erum Khalid Hamdard University, Karachi, Pakistan

Asma Akbar Ladak Aga Khan University

Syed Adnan Ali University of Karachi, Karachi, Pakistan

Follow this and additional works at: https://ecommons.aku.edu/pakistan\_fhs\_mc\_bbs

Part of the Biology Commons, Endocrinology, Diabetes, and Metabolism Commons, Maternal and Child Health Commons, and the Obstetrics and Gynecology Commons

**Recommended Citation** 

# PROOF COVER SHEET

Author(s): Syeda Sadia Fatima, Erum Khalid, Asma Akbar Ladak, and Syed Adnan Ali

- Article title: Colostrum and mature breast milk analysis of serum irisin and sterol regulatory element-binding proteins-1c in gestational diabetes mellitus
- Article no: IJMF\_A\_1454422 Enclosures: 1) Query sheet 2) Article proofs

# Dear Author,

**1. Please check these proofs carefully.** It is the responsibility of the corresponding author to check these and approve or amend them. A second proof is not normally provided. Taylor & Francis cannot be held responsible for uncorrected errors, even if introduced during the production process. Once your corrections have been added to the article, it will be considered ready for publication.

Please limit changes at this stage to the correction of errors. You should not make trivial changes, improve prose style, add new material, or delete existing material at this stage. You may be charged if your corrections are excessive (we would not expect corrections to exceed 30 changes).

For detailed guidance on how to check your proofs, please paste this address into a new browser window: http://journalauthors.tandf.co.uk/production/checkingproofs.asp

Your PDF proof file has been enabled so that you can comment on the proof directly using Adobe Acrobat. If you wish to do this, please save the file to your hard disk first. For further information on marking corrections using Acrobat, please paste this address into a new browser window: http://journalauthors.tandf.co.uk/production/ acrobat.asp

2. Please review the table of contributors below and confirm that the first and last names are structured correctly and that the authors are listed in the correct order of contribution. This check is to ensure that your name will appear correctly online and when the article is indexed.

Sequence	Prefix	Given name(s)	Surname	Suffix
1		Syeda Sadia	Fatima	
2		Erum	Khalid	
3		Asma Akbar	Ladak	
4		Syed Adnan	Ali	

Queries are marked in the margins of the proofs, and you can also click the hyperlinks below.

# General points:

1. **Permissions:** You have warranted that you have secured the necessary written permission from the appropriate copyright owner for the reproduction of any text, illustration, or other material in your article. Please see http:// journalauthors.tandf.co.uk/permissions/usingThirdPartyMaterial.asp.

- 2. Third-party content: If there is third-party content in your article, please check that the rightsholder details for re-use are shown correctly.
- 3. Affiliation: The corresponding author is responsible for ensuring that address and email details are correct for all the co-authors. Affiliations given in the article should be the affiliation at the time the research was conducted. Please see http://journalauthors.tandf.co.uk/preparation/writing.asp.
- 4. **Funding:** Was your research for this article funded by a funding agency? If so, please insert `This work was supported by <insert the name of the funding agency in full>', followed by the grant number in square brackets `[grant number xxxx]'.
- 5. Supplemental data and underlying research materials: Do you wish to include the location of the underlying research materials (e.g. data, samples or models) for your article? If so, please insert this sentence before the reference section: 'The underlying research materials for this article can be accessed at <full link>/ description of location [author to complete]'. If your article includes supplemental data, the link will also be provided in this paragraph. See <http://journalauthors.tandf.co.uk/preparation/multimedia.asp> for further explanation of supplemental data and underlying research materials.
- 6. The **PubMed** (http://www.ncbi.nlm.nih.gov/pubmed) and **CrossRef databases** (www.crossref.org/) have been used to validate the references. Changes resulting from mismatches are tracked in red font.

# AUTHOR QUERIES

- Q1: Please check all authors names and affiliations are correct as present in the proofs. Also, Please provide the department name for affiliation 'd'.
- Q2: Please resupply the corresponding author details and email address if it is inaccurate.
- Q3: Please provide the significance of the indicator (\*) in Table 1.
- Q4: The ORCID details of the authors have been validated against ORCID registry. please check the ORCID ID details of the authors.
- Q5: Provide the location of the manufacturer with town and country names for "Kit Cat number 95512 and F9350, Glory Science Co Ltd".

# How to make corrections to your proofs using Adobe Acrobat/Reader

Taylor & Francis offers you a choice of options to help you make corrections to your proofs. Your PDF proof file has been enabled so that you can mark up the proof directly using Adobe Acrobat/Reader. This is the simplest and best way for you to ensure that your corrections will be incorporated. If you wish to do this, please follow these instructions:

1. Save the file to your hard disk.

2. Check which version of Adobe Acrobat/Reader you have on your computer. You can do this by clicking on the Help" tab, and then About".

If Adobe Reader is not installed, you can get the latest version free from http://get.adobe.com/reader/.

3. If you have Adobe Acrobat/Reader 10 or a later version, click on the Comment" link at the right-hand side to view the Comments pane.

4. You can then select any text and mark it up for deletion or replacement, or insert new text as needed. Please note that these will clearly be displayed in the Comments pane and secondary annotation is not needed to draw attention to your corrections. If you need to include new sections of text, it is also possible to add a comment to the proofs. To do this, use the Sticky Note tool in the task bar. Please also see our FAQs here: http://journalauthors.tandf.co. uk/production/index.asp.

5. Make sure that you save the file when you close the document before uploading it to CATS using the Upload File" button on the online correction form. If you have more than one file, please zip them together and then upload the zip file. If you prefer, you can make your corrections using the CATS online correction form.

# Troubleshooting

Acrobat help: http://helpx.adobe.com/acrobat.html Reader help: http://helpx.adobe.com/reader.html

Please note that full user guides for earlier versions of these programs are available from the Adobe Help pages by clicking on the link Previous versions" under the Help and tutorials" heading from the relevant link above. Commenting functionality is available from Adobe Reader 8.0 onwards and from Adobe Acrobat 7.0 onwards.

**Firefox users:** Firefox's inbuilt PDF Viewer is set to the default; please see the following for instructions on how to use this and download the PDF to your hard drive: http://support.mozilla.org/en-US/kb/view-pdf-files-firefox-without-downloading-them#w\_using-a-pdf-reader-plugin

# **ORIGINAL ARTICLE**



Check for updates

# Colostrum and mature breast milk analysis of serum irisin and sterol regulatory element-binding proteins-1c in gestational diabetes mellitus

# Syeda Sadia Fatima<sup>a</sup> 🝺, Erum Khalid<sup>b</sup>, Asma Akbar Ladak<sup>c</sup> and Syed Adnan Ali<sup>d</sup>

<sup>a</sup>Department of Biological and Biomedical Sciences, Aga Khan University, Karachi, Pakistan; <sup>b</sup>Department of Obstetrics and Gynecology, Hamdard University, Karachi, Pakistan; <sup>c</sup>Medical College, Aga Khan University, Karachi, Pakistan; <sup>d</sup>University of Karachi, Karachi, Pakistan

#### ABSTRACT

**Background:** We aimed to evaluate irisin and SREBP-1c levels in serum, colostrum and mature breast milk in women with and without gestational diabetes (GDM); and to relate them with maternal glucose, lipid profile and weight status of babies.

**Methods:** GDM positive women (n = 33) and normal glucose tolerant women (NGT) (n = 33) were recruited. Maternal blood samples were collected at 28th week of gestation and later at 6-week post-partum while breast milk samples of the lactating mothers were collected within 72 hours of birth (colostrum) and at 6 weeks post-partum (mature milk). Irisin and SREBP-1c levels were analyzed by commercially available ELISA kits for all maternal samples.

**Results:** Lower levels of irisin were seen in serum, colostrum and mature breast milk of GDM females (p < .01). SREBP-1c profile showed a similar trend of low serum levels in GDM, however, they were undetectable in colostrum and mature breast milk. Weak to moderate correlations of serum irisin with BMI (r = 0.439; p < .001), GTT 0 hours (r = 0.403; p = .01), HbA1c (r = -0.312; p = .011), Fasting blood glucose (r = 0.992; p = .008), and baby weight at birth (r = 0.486; p < .001). Colostrum and mature breast milk irisin showed positive associations with baby weight at 6 weeks (r = 0.325; p = .017; r = 0.296; p = .022, respectively). Serum SREBP-1c at 6 weeks correlated with the baby weight (r = 0.255; p = .039), random blood glucose (r = 0.318; p = .009), and HbA1c (r = -0.292; p = .011). All correlations were lost once we adjusted for maternal BMI. **Conclusions:** Low irisin and SREBP1-c levels may favor development of GDM in pregnant subjects. Further, low mature breast milk levels may act as a continued stressor from fetal to infant life as long as breast-feeding is continued. Further studies are required to identify the mechanistic relationship between these biomarkers and GDM.

#### **ARTICLE HISTORY**

Received 16 October 2017 Revised 29 November 2017 Accepted 15 March 2018

#### **KEYWORDS**

Mature breast milk; cytokine; obesity; gestational diabetes; irisin; SREBP

# Introduction

Gestational diabetes mellitus (GDM) is a growing pandemic in the world. At least 17.8% of women are being affected by this condition worldwide [1]. GDM is often accounted for by increased levels of maternal hormones during pregnancy ensuring adequate energy supply to the fetus like estrogen, progesterone, cortisol, human chorionic gonadotrophin, and prolactin. These hormones have been proven to work against insulin thereby increasing its resistance [2]. The perinatal nutritional environment is suggested to have a programing effect on an individual's susceptibility to develop obesity [1]. Breastfeeding is suggested to be protective against obesity which could partly be attributable to bioactive compounds present in mature breast milk [3]. However, the specific components of mature breast milk accounting for these effects have not yet been

identified. Since the discovery of the adipokines, many strong candidates for the development of metabolic syndrome (Mets) have been proposed [4].

Irisin is a recently discovered myokine which is secreted into the serum by the skeletal muscles post exercise [5,6]. Recent studies also show that low irisin levels are characteristic of and may perhaps be a vital factor contributing towards GDM. Irisin is involved in the conversion of brown to white adipose tissue which not only increase thermogenesis but also improves glucose tolerance, insulin sensitivity, reductions in body weight, decreased fat mass and reduced hepatic triglyceride content [7,8]. Decreased levels of irisin during pregnancy therefore may not only contribute to GDM but also towards obesity owing to poor lipid metabolism. Interestingly, some studies have reported significantly decreased serum irisin levels in GDM



CONTACT Syeda Sadia Fatima 🔊 sadia.fatima@aku.edu 🗈 Department of Biological and Biomedical Sciences, Aga Khan University, Stadium Road, Karachi, Pakistan

108

109

110

111

112

113

114

115

116

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

134

135

136

137

138

pregnancies compared to normal pregnancies [9–12], while others have reported little or no difference between the two groups [13–15]. Nevertheless, none fail to acknowledge the risks associated with GDM during and postpregnancy for both the mother and the child.

An important enzyme for the *de novo* lipid synthesis principally expressed in lipogenic tissues is fatty acid synthase (FAS) [16], and it is known to be expressed in placenta [17]. This expression is in turn controlled by "Sterol regulatory element-binding proteins-1c" (SREBP1c) levels. It is observed that whenever there are a decreased cellular sterols and/or poly-unsaturated fatty acid level; there is increase FAS gene transcription *via* actions of SREBP-1c. Expression of SREBP-1c is controlled by circulating hormones and nutrients within the body [18]. Recent reports reveal that in type II diabetes; the expression of SREBP-1c is decreased as a consequence of impaired insulin signaling. This will in turn cause irregular lipid synthesis especially during pregnancy.

However, whether these biomarkers remain elevated post-partum and released in mature breast milk to cause a continuous supply for the baby remains to be identified. Therefore, through this study we aim to estimate the levels of irisin and SREBP-1c during pregnancy and in mature breast milk in lactating women with and without GDM and relate them with maternal lipid profile and weight gain of the newborn babies.

# Materials and methods

This study recruited 66 pregnant women between the 139 ages of 20 and 35 years who visited the outpatient 140 clinic from July 2016 to August 2017 at the Taj 141 Medical Centre and Aga Khan University, Karachi. All 142 study subjects underwent an oral glucose tolerance 143 test at 24-28 weeks of gestation and were classified as 144 GDM (n = 33) or normal glucose tolerant women (NGT) 145 (n = 33) according to the International Association of 146 the Diabetes and Pregnancy Study (IADPSG) guidelines 147 [19]. The pregnant females were followed up from 148 24-28 weeks of gestation till 6 weeks' postdelivery. 149 Pregnant subjects with preexisting diabetes, hyperten-150 sion, twin pregnancies, and assisted pregnancies were 151 excluded from this study. After receiving ethical 152 approval from the institutional ethical review board 153 (Ref No. 4211-BBS-ERC-16) and written and informed 154 consent from all subjects, 5 ml of maternal blood 155 samples were collected twice; first at 28th week of 156 gestation and again at 6 weeks post-partum. 157

158Similarly, two breast milk samples of the lactating159mothers were collected; one within 72 hours of birth

(colostrum =3 ml) and second at 6 weeks post-partum (mature milk =10 ml) with a manual breast pump. The mothers were instructed to refrain from feeding for at least 2 hours (8 am till 10 am) before collecting the sample. These samples were aliquoted and were stored in the dark at -80°C until analysis. For biomarker analysis, milk samples were pretreated according to an established protocol [20]. Briefly, after thawing at room temperature the samples were vortexed and sonicated three times for a 5-s burst. This was followed by preparing skim milk by centrifugation of whole milk at 14,000 rpm, for 30 minutes at 4°C. The resulting translucent sample was used for analysis. All assays were carried out within 2 months of storage. Irisin and SREBP-1c levels were analyzed by commercially available ELISA kit (Kit Cat number 95512 and F9350, Glory Science Co Ltd).  $\bigcirc$ 

Additionally, a spike-and-recovery experiment was designed to assess the difference in assay response according to a set protocol [21]. Briefly, the following procedure and samples were used (i) sample diluent serving as blank, (ii) sample diluent spiked with known concentration of standard (to allow quantification of exogenous analyte), (iii) Two breast milk analytes without spiking (to allow guantification of endogenous analyte), (iv) Two breast milk analytes spiked with known concentration of standard (to allow quantification of both exogenous and endogenous analyte). Ten microliters of spike stock solution was added to the samples, calculated to yield 240 pg/ml spike concentration. Values used for breast milk spiked samples reflect subtraction of the endogenous (no-spike) sample value. The samples were run in duplicates and a mean score was used. The recovery rate for all samples was between 93 and 98% with intra-assay and interassay coefficients of variation <10% and <15%, respectively. A range between 80 and 120% recovery is considered acceptable for such experiments [22].

Weight of mothers was assessed at the time of first visit (i.e. 12–15 weeks of gestation), at the time of glucose tolerance test, i.e. 28 weeks of gestation and at 6 weeks' post-partum. Body mass index (BMI) was calculated by dividing weight by height squared (kg/m<sup>2</sup>). South Asian reference range for BMI was used to categorize study subjects [23]. Clinical data including maternal age, height, family history of diabetes mellitus, gender, and weight of newborn were recorded from the patient record card.

## Statistical analysis

Statistical analyses were conducted using the IBM Statistical Package for the Social Sciences (IBM SPSS 184

185

186

187

188

189

190

191

192

193

194

195

196

197

198

199

200

201

202

203

204

205

206

207

208

209

210

211

212

160

161

162

163

164

165

166

version 21; IBM Corp Inc, Armonk, NY, USA). Descriptive analysis of categorical data was presented in terms of freauencies and percentages, whereas that for continuous variables was expressed as mean ± standard deviation. Continuous variables were assessed by Mann-Whitney U test. Spearman's correlation between serum irisin, SREBP-1c and clinical parameters were also calculated. p value <.05 were regarded to be significant. 

# Results

 $\bigcirc$ 

The results are displayed in Tables 1–2. The study subjects were aged matched therefore we did not find any difference. Females with GDM had a higher body weight, fasting blood glucose (FBG), and random blood glucose (RBG) at 6 weeks postpartum (p < .01). Interestingly, the HbA1c levels at follow up were within normal range for the GDM subjects. The newborn data showed that babies of GDM positive mothers had a higher weight by scan at 28 weeks of gestation, at birth and at follow up 6 weeks after birth (Table 1).

Table 2 shows the biomarker levels in the study subjects. We observed a low serum irisin level in GDM females at 28, 6 weeks postpartum compared to normo-glycemic females (p < .01). Similarly, colostrum and mature breast milk irisin was also low in GDM subjects. SREBP1-c profile showed a similar trend of low serum levels in GDM, however, these levels reduced from a very low value to undetectable range in colostrum and mature breast milk respectively. However, we found no differences in the lipid profile post pregnancy in the study cohort. Furthermore, we found weak to moderate correlations of Serum irisin with BMI (r=0.439; p > .001) GTT 0 hours (r=0.403; p=.01), HbA1c (r=-0.312; r=.011), FBG (r=0.992; p=.008) and baby weight at birth (r=0.486; p < .001).

1. Maternal and	l newborn data.
<ol> <li>Maternal and</li> </ol>	newborn data

-

Colostrum and mature breast milk irisin showed positive associations with baby weight at 6 weeks (r = 0.325; p = .017; r = 0.296; p = .022, respectively). Serum SREBP-1c at 6 weeks correlated with the baby weight at 6 weeks (r = 0.255; p = .039), RBG at 6 weeks (r = 0.318; p = .009), and HbA1c (r = -0.292; p = .011). However all correlations were lost once we adjusted for maternal BMI. Therefore, these associations were dependent on the maternal weight and BMI.

### Discussion

The finding of our study showed lower levels of irisin in serum, colostrum and mature breast milk of GDM females. These low levels may increase the risk of developing diabetes mellitus type 2, hypoglycaemia, and respiratory distress syndrome [10,11]. Contrary to our finding, one study found irisin levels to increase from colostrum to transitional and mature milk in both normal glucose tolerant and GDM patients [10]. Low irisin levels in the serum, mature breast milk and colostrum acts as a continued stressor from fetal to infant life. This may continue to have adverse effects on the baby's health and lipid regulation as long as breast-feeding is continued. Irisin has proven to play a vital role in improving obesity and glucose homeostasis, It has been proposed that irisin increases adenosine monophosphate (AMP) activated protein kinase phosphorylation and increases glucose uptake; therefore plays an important role in glucose metabolism [24]. Irisin further has proven to inhibit atherosclerosis by endothelial proliferation [25], cholesterol synthesis in the hepatocytes [26] and promote osteoblast proliferation and differentiation [27]. Therefore, we believe that low irisin in the GDM population may have crucial effects on the mother's and baby's health, both during pregnancy and post-partum. It is also postulated that

	NGT	GDM	
	n = 33	n = 33	
Variables	$Mean \pm SD$	$Mean \pm SD$	p value
Maternal data			
Age (year)	$24.10 \pm 1.77$	$24.23 \pm 1.14$	.412
BMI (kg/m <sup>2</sup> )	$20.55 \pm 3.76$	$25.22 \pm 4.24^{*}$	<.001
GTT at 0 hours	$78.24 \pm 12.05$	103.27 ± 15.15	<.001
GTT at 2 hours	$120.48 \pm 10.47$	$162.66 \pm 44.68$	<.001
RBG —6 weeks postpartum (mg/dl)	$107.04 \pm 19.45$	$144.00 \pm 3.66^*$	<.001
HbA1c -6 weeks post-partum (mg/dl)	NA	$5.23 \pm 0.92$	-
New-born data			
Abdominal circumference by scan (cm) at 28 weeks of gestation	$26.80 \pm 3.98$	$28.51 \pm 5.09$	.135
Foetal weight by scan (kg) at 28 weeks of gestation	$1.98 \pm 0.54$	$2.28 \pm 0.59$	.008
Baby gender	Male 19 (57.57)	Male 16 (48.48)	.547
	Female 14 (42.42)	Female 17 (51.51)	
Baby weight at birth (kg)	$3.53 \pm 0.51$	$4.41 \pm 0.97$	<.001
Baby weight at 6 weeks (kg)	$4.35 \pm 0.40$	$5.53 \pm 0.15^{*}$	<.001

BMI: body mass index; RBG: random blood glucose. Values expressed as mean and standard deviation and absolute number with percentage in parenthesis.

321 322

323

324

325

326

327

328

329

330

331

332

333

355

356

357

358

359

360

361

362

363

364

365

366

367

368

369

370

371

Variables	NGT n = 33 Mean ± SD	GDM n = 33 Mean ± SD	p Valu
Irisin profile			
Serum Irisin at 28-week gestation (pg/ml)	72.94 ± 9.11	$42.09 \pm 3.21$	.01
Serum Irisin at 6 weeks post-partum (pg/ml)	264.97 ± 40.88	$138.32 \pm 6.82$	.003
Colostrum Irisin (pg/ml)	57.08 ± 8.28	$10.36 \pm 4.73$	<.001
Mature breast milk Irisin (pg/ml) SREBP-1c profile	$56.40\pm9.55$	$15.35 \pm 0.42$	<.001
Serum SREBP1-c at 28-week gestation (pg/ml)	73.94 ± 9.41	$43.62 \pm 7.33$	.013
Serum SREBP1-c at 6 post-partum (pg/ml)	38.33 ± 1.752	$20.92 \pm 1.755$	< .001
Colostrum SREBP1-c (pg/ml)	$9.39 \pm 4.07$	$4.09 \pm 1.65$	.233
Mature breast milk SREBP1-c (pg/ml) Lipid profile at 6 weeks post-partum	0.00	0.00	-
Cholesterol (mg/dl)	187.66 ± 31.50	$172.84 \pm 44.03$	.518
TAG (mg/dl)	133.09 ± 17.97	147.±11.00	.340
HDL (mg/dl)	$45.72 \pm 9.40$	$47.66 \pm 8.20$	.612
LDL (mg/dl)	$107.56 \pm 3.69$	$113.73 \pm 5.10$	.584

Table 2. Maternal serum, colostrum, and mature breast milk biomarker data

Values expressed as mean and standard deviation.

334 the levels affect intrauterine growth of the fetus. 335 However recent studies by Briana and group [28] 336 reported that no difference was observed in cord 337 blood levels of intra uterine growth retardation and 338 normal for gestational age babies. Similarly they 339 reported that irisin levels decrease with advancing ges-340 tational weeks and this could be attributed to 341 increased maternal fat deposition. That may lead to 342 insulin resistance in the latter half of pregnancy [29]. 343 These findings strongly support our hypothesis and 344 345 results. As of late, no validated normal range for irisin 346 have been reported, but low serum irisin levels were 347 reported in Japanese population  $(1.1 \pm 0.1 \text{ ng/ml} \text{ for})$ 348 males and  $1.2 \pm 0.2$  ng/ml for females) [30] and in 349 Tehran, Iran  $(0.81 \pm 0.41 \text{ ng/ml} \text{ in normal weight obese})$ 350 and 0.58±0.26 ng/ml controls) [31]. Perhaps these 351 differences in irisin levels are observed due to interpo-352 pulation or plasma versus serum sample differences, 353 yet our results are quite similar to the neighbor-354 ina countries.

Increased levels of FBG/RBG at 6 weeks post-partum were seen in our study. Similar to our findings, a positive association between GDM history, 2-hour glucose; and HbA1c during pregnancy and post-partum diabetes was seen within Chinese women [32]. Interestingly, in a recent study conducted by our group, we found that 14% of GDM positive females developed DM within 2 years of their last pregnancies [33]. Furthermore, we report a higher birth weight as well as infant weight 6 weeks post birth for GDM mothers, The incidence of fetal macrosomia ranges between 20 and 40% in GDM positive cases [34], but the baby weight normalizes with time [35]. However, our findings suggests that the constant stress in terms of low irisin though mature breast milk was a trigger for the deranged body fat accumulation and higher body weight of these infants.

We also report a significantly lower level of SREBP-1c in serum and colostrum of GDM positive females however, its level was undetectable in the mature breast milk of both the groups. Further we did not find any differences in maternal lipid profile among the groups. This may be due to the fact that GDM increases concentration of inflammatory cytokines which induces fatty acid synthase (FAS) expression but not the expression of SREBP protein [36]. A study found significant increase in expression of SREBP-2 and fatty acid synthase (FAS) in the placenta while both 3-hydroxy-3-methylglutaryl-CoA (HMG Co-A) lyase and SREBP-1 were not modified. Supposedly, in GDM as irisin levels tend to decrease, insulin resistance tend to increase which in turn may effect levels of SREBP-1 [37]. Since SREBP proteins are responsible for uptake and synthesis of fatty acids and cholesterol [38], our hypothesis suggested that there should be higher levels of lipid biomarkers in the maternal serum. Yet, no significant difference was observed in the lipid profile of GDM and non-GDM mothers. We assume that since the GDM inflicted mothers were kept on diet and drug regimens, that may have contributed to the within normal range levels as seen.

Lastly, we report a weak to moderate correlation of irisin with maternal BMI, GTT, baby weight at birth, and FBG. In addition, colostrum and mature breast milk irisin showed positive associations with baby weight at 6 weeks. However, all correlations were lost once we adjusted for maternal BMI. Therefore, we believe that these associations were dependent on the maternal weight and BMI. A similar finding was reported that irisin was positively related with BMI and FBG [39], yet another study reported that none of the anthropometric parameters to correlate with serum irisin levels [40]. Another study's findings suggested that irisin was negatively correlated with age and FBG while positively 409

410

411

412

413

414

415

416

417

418

419

420

421

422

423

424

372

373 374

375

376

377

correlated with HDL-c [41]. Interestingly, Sinanoglu and 42.5 group [40] proposed an ethnicity and maternal age 426 related link with the fat content of colostrum instead of 427 the commonly agreed notion of maternal pre preg-428 nancy BMI. They also suggest that this finding can 429 be used to design custom diets for mothers to 430 enhance the mature breast milk nutritional value and in 431 turn have a positive impact on new born health. 432 Furthermore, another recent study measured irisin lev-433 els in babies and compared them with growth rate and 434 insulin levels. They report that low irisin levels were 435 seen in intra uterine growth restricted babies and irisin 436 showed a weak positive correlation with the birth 437 weight of babies. In addition, a weak positive associ-438 ation was also seen for fetal irisin and insulin concentra-439 tions [42]. 440

We postulate that derangement of both irisin and SREBP in pregnancy may be-trigger abnormal fatty acid (FA) metabolism. Furthermore, impaired metabolism causes accumulation of reactive lipid species eventually altering the response to tissue insulin signaling. Irisin may induce gene expression changes by directly acting on the tissues or indirectly by inducing a "factor X". This has significant implication for irisin as a novel therapeutic target. However, further mechanistic research is required to validate and propose this effect.

Furthermore, in addition to being secreted from skeletal and cardiac muscle, irisin has also been detected in the brain (neurons and neuroglia), the skin (sebaceous glands), the testis, epididymis, liver, pancreas, spleen, stomach, and three major paired salivary glands (submandibular, sublingual, and parotid) [43-45]. It is removed from the body mainly through the hepato-biliary system and the kidneys [45-47]. It is possible that these molecules may act as trophic or mediating agents that sends signal to brain, gut, muscle, and adipose tissue during pregnancy or via breast milk post-partum. Therefore, it is plausible to comment that irisin may modulate energy expenditure or food intake through topical, paracrine or autocrine actions. This could potentially have postnatal effects on the baby via skin, tongue or gut to modify energy metabolism and stimulate appetite center in brain, tissue hyperplasia or hypertrophy in adipose tissue and liver [48-50]. The exact focal mechanism or function however is currently unknown, so future in vivo studies are required that might provide the conclusive results to consolidate this hypothesis.

# Conclusions

441

442

443

444

445

446

447

448

449

450

451

452

453

454

455

456

457

458

459

460

461

462

463

464

465

466

467

468

469

470

471

472

473

474

475

476

477

Low irisin and SREBP-1c levels in the serum, mature breast milk and colostrum may favor development of

525

526

527

528

529

530

GDM in pregnant subjects. Further, low mature breast milk irisin and <u>SREBP-1c</u> levels may act as a continued stressor from fetal to infant life till breast-feeding is continued. Further studies are required to identify the mechanistic relationship between these biomarkers and GDM.

## **Disclosure statement**

The authors declare that they have no conflict of interest.

# Funding

This work was supported by Department of Biological and Biomedical Sciences and AKU – Seed Money Funds.

# ORCID

Syeda Sadia Fatima D http://orcid.org/0000-0002-3164-0225

# References

- Fatima SS, Rehman R, Alam F, et al. Gestational diabetes mellitus and the predisposing factors. J Pak Med Assoc. 2017;67:261–265.
- [2] Lowe WL, Jr., Karban J. Genetics, genomics and metabolomics: new insights into maternal metabolism during pregnancy. Diabet Med. 2014;31:254–262.
- [3] Harder T, Bergmann R, Kallischnigg G, et al. Duration of breastfeeding and risk of overweight: a meta-analysis. Am J Epidemiol. 2005;162:397–403.
- [4] Hamosh M. Bioactive factors in human milk. Pediatr Clin North Am. 2001;48:69–86.
- [5] Boström PA, Fernández-Real JM, Mantzoros C. Irisin in humans: recent advances and questions for future research. Metab Clin Exp. 2014;63:178–180.
- [6] Boström P, Wu J, Jedrychowski MP, et al. A PGC1- $\alpha$ -dependent myokine that drives brown-fat-like development of white fat and thermogenesis. Nature. 2012;481:463–468.
- [7] Stanford KI, Middelbeek RJ, Townsend KL, et al. Brown adipose tissue regulates glucose homeostasis and insulin sensitivity. J Clin Invest. 2013;123:215–223.
- [8] So WY, Leung PS. Irisin ameliorates hepatic glucose/ lipid metabolism and enhances cell survival in insulinresistant human HepG2 cells through adenosine monophosphate-activated protein kinase signaling. Int J Biochem Cell Biol. 2016;78:237–247.
- [9] Garcés MF, Peralta JJ, Ruiz-Linares CE, et al. Irisin levels during pregnancy and changes associated with the development of preeclampsia. J Clin Endocrinol Metab. 2014;99:2113–2119.
- [10] Aydin S, Kuloglu T, Aydin S. Copeptin, adropin and irisin concentrations in breast milk and plasma of healthy women and those with gestational diabetes mellitus. Peptides. 2013;47:66–70.
- [11] Yuksel MA, Oncul M, Tuten A, et al. Maternal serum and fetal cord blood irisin levels in gestational

532

533

534

535

536

537

538

539

540

541

542

543

544

545

546

547

548

549

550

551

552

553

554

555

556

557

558

559

560

561

562

563

564

565

566

567

568

569

570

571

572

573

574

575

diabetes mellitus. Diabetes Res Clin Pract. 2014;104: 171–175.

- [12] Kuzmicki M, Telejko B, Lipinska D, et al. Serum irisin concentration in women with gestational diabetes. Gynecol Endocrinol. 2014;30:636–639.
- [13] Çağlar M, Göksu M, Isenlik BS, et al. Irisin in idiopathic foetal growth restriction. J Endocrinol Invest. 2014;37: 619–624.
  - [14] Piya MK, Harte AL, Sivakumar K, et al. The identification of irisin in human cerebrospinal fluid: influence of adiposity, metabolic markers, and gestational diabetes. Am J Physiol Endocrinol Metab. 2014;306: E512–E518.
    - [15] Ebert T, Stepan H, Schrey S, et al. Serum levels of irisin in gestational diabetes mellitus during pregnancy and after delivery. Cytokine. 2014;65:153–158.
    - [16] Griffin MJ, Sul HS. Insulin regulation of fatty acid synthase gene transcription: roles of USF and SREBP-1c. IUBMB Life. 2004;56:595–600.
    - [17] Urban G, Marinoni E, Di Iorio R, et al. New placental factors: between implantation and inflammatory reaction. Early Pregnancy. 2001;5:70–71.
  - [18] Janowski BA. The hypocholesterolemic agent LY295427 up-regulates INSIG-1, identifying the INSIG-1 protein as a mediator of cholesterol homeostasis through SREBP. Proc Natl Acad Sci USA. 2002;99: 12675–12680.
  - [19] International Association of Diabetes and Pregnancy Study Groups Consensus Panel, Metzger BE, Gabbe SG, et al. International association of diabetes and pregnancy study groups recommendations on the diagnosis and classification of hyperglycemia in pregnancy. Diabetes Care. 2010;33:676–682.
- [20] Brunner S, Schmid D, Zang K, et al. Breast milk leptin and adiponectin in relation to infant body composition up to 2 years. Pediatr Obes. 2015;10:67–73.
  - [21] Kragstrup TW, Vorup-Jensen T, Deleuran B, et al. A simple set of validation steps identifies and removes false results in a sandwich enzyme-linked immunosorbent assay caused by anti-animal IgG antibodies in plasma from arthritis patients. Springerplus. 2013;2:263.
- [22] Selby C. Interference in immunoassay. Ann Clin Biochem. 1999;36:704–721.
  - [23] Snehalatha C, Viswanathan V, Ramachandran A. Cutoff values for normal anthropometric variables in Asian Indian adults. Diabetes Care. 2003;26:1380–1384.
  - [24] Lee HJ, Lee JO, Kim N, et al. Irisin, a novel myokine, regulates glucose uptake in skeletal muscle cells via AMPK. Mol Endocrinol. 2015;29:873–881.
  - [25] Zhang Y, Song H, Zhang Y, et al. Irisin inhibits atherosclerosis by promoting endothelial proliferation through microRNA126-5p. J Am Heart Assoc. 2016;5: e004031.
- 576[26]Tang H, Yu R, Liu S, et al. Irisin inhibits hepatic choles-<br/>terol synthesis via AMPK-SREBP2 signaling. EBioMedicine.<br/>2016;6:139–148.5782016;6:139–148.
- [27]Qiao X, Nie Y, Ma Y, et al. Irisin promotes osteoblast<br/>proliferation and differentiation via activating the<br/>MAP kinase signaling pathways. Sci Rep. 2016;6:<br/>18732.
- 582[28]Briana DD, Boutsikou M, Boutsikou T, et al. Plasma583copeptin may not be a sensitive marker of perinatal

stress in healthy full-term growth-restricted fetuses. J Matern Fetal Neonatal Med. 2017;30:705–709.

- [29] Briana DD, Boutsikou M, Athanasopoulos N, et al. Implication of the myokine irisin in maternal energy homeostasis in pregnancies with abnormal fetal growth. J Matern Fetal Neonatal Med. 2016;29:3429–3433.
- [30] Fukushima Y, Kurose S, Shinno H, et al. Relationships between serum irisin levels and metabolic parameters in Japanese patients with obesity. Obes Sci Practice. 2016;2:203–209.
- [31] Mehrabian S, Taheri E, Karkhaneh M, et al. Association of circulating irisin levels with normal weight obesity, glycemic and lipid profile. J Diabetes Metab Disord. 2015;15:17.
- [32] Liu H, Zhang S, Wang L, et al. Fasting and 2-hour plasma glucose, and HbA1c in pregnancy and the postpartum risk of diabetes among Chinese women with gestational diabetes. Diabetes Res Clin Pract. 2016;112:30–36.
- [33] Aziz S, Munim TF, Fatima SS. Post-partum follow-up of women with gestational diabetes mellitus: effectiveness, determinants, and barriers. J Matern Fetal Neonatal Med. 2018;31:1607–1612.
- [34] Liu Z, Ao D, Yang H, et al. Gestational weight gain and risk of gestational diabetes mellitus among Chinese women. Chin Med J (Engl). 2014;127:1255–1260.
- [35] Liu F, Liu Y, Lai YP, et al. Fetal hemodynamics and fetal growth indices by ultrasound in late pregnancy and birth weight in gestational diabetes mellitus. Chin Med J (Engl). 2016;129:2109–2114.
- [36] Marseille-Tremblay C, Ethier-Chiasson M, Forest JC, et al. Impact of maternal circulating cholesterol and gestational diabetes mellitus on lipid metabolism in human term placenta. Mol Reprod Dev. 2008;75: 1054–1062.
- [37] Fukushima Y, Kurose S, Shinno H, et al. Effects of body weight reduction on serum irisin and metabolic parameters in obese subjects. Diabetes Metab J. 2016;40:386–395.
- [38] Jeon TI, Osborne TF. SREBPs: metabolic integrators in physiology and metabolism. Trends Endocrinol Metab. 2012;23:65–72.
- [39] Choi YK, Kim MK, Bae KH, et al. Serum irisin levels in new-onset type 2 diabetes. Diabetes Res Clin Pract. 2013;100:96–101.
- [40] Ural UM, Sahin SB, Tekin YB, et al. Alteration of maternal serum irisin levels in gestational diabetes mellitus. Ginekol Pol. 2016;87:395–398.
- [41] Zhao L, Li J, Li ZL, et al. Circulating irisin is lower in gestational diabetes mellitus. Endocr J. 2015;62: 921–926.
- [42] Baka S, Malamitsi-Puchner A, Boutsikou T, et al. Cord blood irisin at the extremes of fetal growth. Metabolism. 2015;64:1515–1520.
- [43] Aydin S, Kuloglu T, Aydin S, et al. A comprehensive immunohistochemical examination of the distribution of the fat-burning protein irisin in biological tissues. Peptides. 2014;61:130–136.
- [44] Huh JY, Panagiotou G, Mougios V, et al. FNDC5 and irisin in humans: I. Predictors of circulating concentrations in serum and plasma and II. mRNA expression and

[45]

[46]

[47]

circulating concentrations in response to weight loss

Aydin S, Aydin S, Kuloglu T, et al. Alterations of irisin

concentrations in saliva and serum of obese and nor-

mal-weight subjects, before and after 45min of a

Lv J, Pan Y, Li X, et al. Study on the distribution and elim-

ination of the new hormone irisin in vivo: new discov-

eries regarding irisin. Horm Metab Res. 2015;47:591-595.

Perakakis N, Triantafyllou GA, Fernández-Real JM,

et al. Physiology and role of irisin in glucose homeo-

Turkish bath or running. Peptides. 2013;50:13-18.

and exercise. Metabolism. 2012;61:1725-1738.

stasis. Nat Rev Endocrinol. 2017;13:324-337.

- [48] Wrann CD. FNDC5/Irisin-their role in the nervous system and as a mediator for beneficial effects of exercise on the brain. Brain Plast. 2015;1:55-61.
- [49] Kurdiova T, Balaz M, Vician M, et al. Effects of obesity, diabetes and exercise on Fndc5 gene expression and irisin release in human skeletal muscle and adipose tissue: in vivo and in vitro studies. J Physiol. 2014;592:1091-1107.

[50] Vergnes L, Chin RG, de Aguiar Vallim T, et al. SREBP-2deficient and hypomorphic mice reveal roles for SREBP-2 in embryonic development and SREBP-1c expression. J Lipid Res. 2016;57:410-421.