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Draft Genome Sequence of a *Salmonella enterica* Serovar Typhi Strain Resistant to Fourth-Generation Cephalosporin and Fluoroquinolone Antibiotics

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ABSTRACT Typhoid is endemic in developing countries. We report here the first draft genome sequence of a *Salmonella enterica* serovar Typhi clinical isolate from Pakistan exhibiting resistance to cefepime (a fourth-generation cephalosporin) and fluoroquinolone antibiotics, two of the last-generation therapies against this pathogen. The genome is ~4.8 Mb, with two putative plasmids.

Typhoid fever is one of the leading causes of mortality due to infectious diseases in developing Asian and African countries. Resistance to third-generation cephalosporin and fluoroquinolone antibiotics has been increasing in recent years in *Salmonella* spp. (1, 2), but there have been few reports of resistance to fourth-generation cephalosporins and fluoroquinolones in *Salmonella enterica* serovar Typhi.

In this study, we performed whole-genome sequencing of a multidrug-resistant *Salmonella enterica* subsp. *enterica* serovar Typhi clinical isolate obtained from a patient admitted to a tertiary care hospital in Rawalpindi, Pakistan (3). The isolate was phenotypically resistant to a fourth-generation cephalosporin (cefepime) and several fluoroquinolones (ciprofloxacin, levofloxacin, and moxifloxacin). Whole-genome sequencing (WGS) was carried out to identify the resistance determinants carried by this *S*. Typhi isolate.

Genomic DNA from 10 colonies of the isolate was extracted with the bacteremia DNA kit (Mo Bio) and used as input for sequencing libraries using a modification of the Nextera XT kit (Illumina) (4). A total of 10,307,350 paired-end 150-bp reads were generated from an Illumina NextSeq 2500 and processed using the High-Throughput Computing Facility at the Washington University School of Medicine (St. Louis, MO, USA). Illumina adapters and potentially contaminating human reads were removed using Trimmomatic and deconseq (5, 6). The resulting 10,288,264 paired-end reads were assembled into scaffolds using SPAdes 3.10.0 (7). Contigs were analyzed by QUAST (8). The assembly produced 95 contigs >500 bp, with an N_{50} value of 144,739 bp. The longest contig was 393,347 bp. Contigs were annotated using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) (http://www.ncbi.nlm.nih.gov/genomes/static/Pipeline.html), which identified 4,975 coding sequences (CDSs), 69 tRNA genes, and 4 rRNA genes.

Further analysis identified putative plasmid sequences and antimicrobial resistance genes. PlasmidFinder 1.3 (https://cge.cbs.dtu.dk/services/PlasmidFinder/) (9) identified

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the presence of IncQ1 and IncY sequences. Antibiotic resistance genes (ARGs) were identified using ARDB (10), CARD (11), ResFinder (12), and ResFams (13). Annotation using these databases identified $bla_{CTX-M-15}$, bla_{TEM-1} , S83F *gyrA*, and *qnrS1*, which could potentially explain the observed resistance to cefepime and fluoroquinolones (ciprofloxacin, levofloxacin, and moxifloxacin).

The prevalence of $bla_{CTX-M-15}$ -positive *S*. Typhi isolates of Asian origin has been reported in Iraq, Kuwait, India, and Bangladesh (14–17). To the best of our knowledge, this study is the first draft genome sequence of a $bla_{CTX-M-15}$, bla_{TEM-1} , and *qnrS*-positive *S*. Typhi strain from Pakistan exhibiting resistance to cefepime and fluoro-quinolones.

Accession number(s). This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession number NIFP00000000. The version described in this paper is version NIFP01000000.

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REFERENCES

- Kumar S, Rizvi M, Berry N. 2008. Rising prevalence of enteric fever due to multidrug-resistant *Salmonella*: an epidemiological study. J Med Microbiol 57:1247–1250. https://doi.org/10.1099/jmm.0.2008/001719-0.
- Eng S-K, Pusparajah P, Ab Mutalib N, Ser H, Chan K, Lee L. 2015. Salmonella: a review on pathogenesis, epidemiology and antibiotic resistance. Front Life Sci 8:284–293. https://doi.org/10.1080/21553769 .2015.1051243.
- Munir T, Lodhi M, Ansari JK, Andleeb S, Ahmed M. 2016. Extended spectrum beta lactamase producing cephalosporin resistant *Salmonella* Typhi, reported from Rawalpindi, Pakistan. J Pak Med Assoc 66: 1035–1036.
- Baym M, Kryazhimskiy S, Lieberman TD, Chung H, Desai MM, Kishony R. 2015. Inexpensive multiplexed library preparation for megabase-sized genomes. PLoS One 10:e0128036. https://doi.org/10.1371/journal.pone .0128036.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- Schmieder R, Edwards R. 2011. Fast identification and removal of sequence contamination from genomic and metagenomic datasets. PLoS One 6:e17288. https://doi.org/10.1371/journal.pone.0017288.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. Bioinformatics 29:1072–1075. https:// doi.org/10.1093/bioinformatics/btt086.
- Carattoli A, Zankari E, García-Fernández A, Voldby Larsen M, Lund O, Villa L, Møller Aarestrup F, Hasman H. 2014. *In silico* detection and typing of plasmids using PlasmidFinder and plasmid multilocus sequence typing.

Antimicrob Agents Chemother 58:3895–3903. https://doi.org/10.1128/ AAC.02412-14.

- Liu B, Pop M. 2009. ARDB—Antibiotic Resistance Genes Database. Nucleic Acids Res 37:D443–D447. https://doi.org/10.1093/nar/gkn656.
- McArthur AG, Waglechner N, Nizam F, Yan A, Azad MA, Baylay AJ, Bhullar K, Canova MJ, De Pascale G, Ejim L, Kalan L, King AM, Koteva K, Morar M, Mulvey MR, O'Brien JS, Pawlowski AC, Piddock LJ, Spanogiannopoulos P, Sutherland AD, Tang I, Taylor PL, Thaker M, Wang W, Yan M, Yu T, Wright GD. 2013. The comprehensive antibiotic resistance database. Antimicrob Agents Chemother 57:3348–3357. https://doi.org/10.1128/AAC.00419-13.
- Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. J Antimicrob Chemother 67:2640–2644. https://doi .org/10.1093/jac/dks261.
- Gibson MK, Forsberg KJ, Dantas G. 2015. Improved annotation of antibiotic resistance determinants reveals microbial resistomes cluster by ecology. ISME J 9:207–216. https://doi.org/10.1038/ismej.2014.106.
- Rotimi VO, Jamal W, Pal T, Sovenned A, Albert MJ. 2008. Emergence of CTX-M-15 type extended-spectrum beta-lactamase-producing *Salmo-nella* spp. in Kuwait and the United Arab Emirates. J Med Microbiol 57:881–886. https://doi.org/10.1099/jmm.0.47509-0.
- Pfeifer Y, Matten J, Rabsch W. 2009. Salmonella enterica serovar Typhi with CTX-M beta-lactamase, Germany. Emerg Infect Dis 15:1533–1535. https://doi.org/10.3201/eid1509.090567.
- Ahmed D, Hoque A, Mazumder R, Nahar K, Islam N, Gazi SA, Hossain MA. 2012. Salmonella enterica serovar Typhi strain producing extendedspectrum beta-lactamases in Dhaka, Bangladesh. J Med Microbiol 61: 1032–1033. https://doi.org/10.1099/jmm.0.044065-0.
- González-López JJ, Piedra-Carrasco N, Salvador F, Rodríguez V, Sánchez-Montalvá A, Planes AM, Molina I, Larrosa MN. 2014. ESBL-producing Salmonella enterica serovar Typhi in traveler returning from Guatemala to Spain. Emerg Infect Dis 20:1918–1920. https://doi.org/10.3201/ eid2011.140525.