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# Enterobacteria secrete an inhibitor of *Pseudomonas* virulence during clinical bacteriuria

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***Escherichia coli* and other Enterobacteriaceae are among the most common pathogens of the human urinary tract. Among the genetic gains of function associated with urinary *E. coli* isolates is the *Yersinia* high pathogenicity island (HPI), which directs the biosynthesis of yersiniabactin (Ybt), a virulence-associated metallophore. Using a metabolomics approach, we found that *E. coli* and other Enterobacteriaceae expressing the *Yersinia* HPI also secrete escherichelin, a second metallophore whose chemical structure matches a known synthetic inhibitor of the virulence-associated pyochelin siderophore system in *Pseudomonas aeruginosa*. We detected escherichelin during clinical *E. coli* urinary tract infection (UTI) and experimental human colonization with a commensal, potentially probiotic *E. coli* bacteriuria strain. Escherichelin production by colonizing enterobacteria may help human hosts resist opportunistic infections by *Pseudomonas* and other pyochelin-expressing bacteria. This siderophore-based mechanism of microbial antagonism may be one of many elements contributing to the protective effects of the human microbiome. Future UTI-preventive probiotic strains may benefit by retaining the escherichelin biosynthetic capacity of the *Yersinia* HPI while eliminating the Ybt biosynthetic capacity.**

## Introduction

Urinary tract infections (UTIs) are among the most common infectious diseases treated by physicians and are increasingly associated with antibiotic-resistant bacteria (1, 2). Although bacteriuria, the presence of bacteria in the urine, is a key feature of UTI, it also occurs frequently in the absence of symptoms. Bacteriuria seeded from the intestinal microbiome may self-resolve, progress to UTI, or persist asymptomatically, in which case antibiotic treatment is not recommended (3). Indeed, recent studies support the existence of urinary tract microbiomes in otherwise healthy individuals (4). Multiple lines of evidence suggest that the microbiome can prevent infections. Paradoxically, antibiotic treatment has been observed to increase the risk of subsequent UTIs (5) and to increase the risk of UTIs caused by atypical uropathogens such as *Pseudomonas* (6). Members of the family Enterobacteriaceae, particularly *E. coli*, inhabit the intestinal microbiome and are most commonly associated with asymptomatic bacteriuria and UTI. The early bacteriuric state from which UTIs can develop is poorly understood and challenging to study, but it is known that UTI-associated *E. coli* isolates frequently possess nonconserved genetic systems related to virulence (7, 8).

Prominent among virulence-associated adaptations in uropathogenic *E. coli* (UPEC) isolates are siderophores, specialized metabolites able to bind extracellular Fe(III) ions for nutritional use in low-iron conditions (7, 9). High-affinity Fe(III) binding by

siderophores helps bacteria resist host-mediated iron deprivation, a prototypical example of nutritional immunity (10, 11). Siderophores are widespread among medically important bacteria, which often express chemically distinctive siderophore types. A recent quantitative metabolomics study of *E. coli* isolates from patients with UTI revealed that the resident intestinal strain in each patient that successfully infected the urinary tract preferentially secreted the siderophores yersiniabactin (Ybt) and salmochelin (12). These results suggest that the early stages of ascending UTI are competitive and favor isolates with specific siderophore profiles. In this and other studies, Ybt, a siderophore encoded by the *Yersinia* high pathogenicity island (HPI), was associated with urinary tract virulence (7, 13–17).

The *Yersinia* HPI is a complex, 4-operon mobile genetic element encoding proteins involved in Ybt biosynthesis, import, and transcriptional regulation. It is highly prevalent among clinical extraintestinal pathogenic *E. coli* isolates and is also found with varying frequencies in other Enterobacteriaceae, including the eponymous *Yersinia pestis* (18). *Yersinia* HPI genes are highly upregulated during UTI, and Ybt is detectable in patients' urine specimens (12, 14, 17). Ybt, the recognized product of *Yersinia* HPI-positive bacteria to date, has been distinguished from other *E. coli* siderophores by its ability to form stable Cu(II) complexes. This broad-spectrum metal-binding property enables Ybt to scavenge Fe(III) for nutritional use, detoxify copper, and catalyze superoxide dismutation (14, 15, 19). The multiple functions of Ybt illustrate the importance of diverse metal ion interactions during microbial pathogenesis.

A hybrid nonribosomal peptide synthetase/polyketide synthase (NRPS/PKS) pathway encoded on the *Yersinia* HPI directs the biosynthesis of Ybt. There is remarkable natural diversity in microbial

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NRPS/PKS systems, which are notable in infectious diseases as the source of antibiotics such as mupirocin, tetracyclines, and macrolides, but also the source of many virulence-associated siderophores such as pyochelin in *Pseudomonas aeruginosa* and mycobactin in *Mycobacterium tuberculosis* (20–22). NRPS and PKS pathways are characterized by their thiotemplating, assembly line mechanism, whereby all substrates and intermediates are covalently attached to phosphopantetheinylated carrier protein (CP) domains as they proceed through the catalytic domains of large, multimodular biosynthetic enzymes. The Ybt biosynthetic substrates are chorismate, 3 molecules of cysteine, malonate from malonyl-CoA, and methyl groups derived from S-adenosyl-methionine (SAM). During Ybt biosynthesis, YbtS converts chorismate to salicylate (23), which is activated and transferred onto the first NRPS, HMWP2 (*irp2*), by YbtE. After condensing salicylate and 2 molecules of cysteine, the last intermediate synthesized by HMWP2 is transferred onto the hybrid NRPS/PKS HMWP1 (*irp1*), where malonate, another molecule of cysteine, and methyl groups are incorporated. At this point, YbtU catalyzes an NADPH-dependent reduction of an HMWP1-bound intermediate. Once the molecule is complete, Ybt is released from the final CP domain of HMWP1 by a terminal thioesterase domain (20, 24). Although Ybt biosynthesis has been elegantly characterized *in vitro*, an untargeted survey of this pathway's products in bacterial culture has not been reported.

In this study, comparative metabolomic profiling revealed escherichelin, a newly appreciated cellular product of the Ybt biosynthetic pathway in *E. coli* and other urinary Enterobacteriaceae isolates. Structural characterization identified escherichelin as 2-(2-hydroxyphenyl)-4,5-dihydro-[2,4]bisthiazolyl-4-carboxylic acid, (abbreviated as HPTT-COOH from hydroxyphenyl-thiazolyl-thiazolyl-carboxylic acid), an Fe(III) chelator that was previously synthesized and found to inhibit pyochelin-mediated iron uptake in *P. aeruginosa* (25). Escherichelin purified from *E. coli* lacks the siderophoric activity of Ybt, but inhibits *P. aeruginosa* growth in a pyochelin-dependent culture condition. Consistent with this activity, we observed that *P. aeruginosa* minimizes escherichelin release compared with *E. coli*, despite generating the same biosynthetic precursors to escherichelin during pyochelin biosynthesis. This bioactivity and its production during human *E. coli* bacteriuria raises the possibility that escherichelin may help human hosts avoid developing symptomatic UTIs from pyochelin-producing organisms. Escherichelin was detectable in a human subject who was experimentally colonized with a derivative of the bacterial interference strain *E. coli* 83972 that has been studied in several human trials as a potential agent to prevent UTI (26, 27). These findings reveal how a second metallophore product of the *Yersinia* HPI biosynthetic system may participate in interspecies competition during polymicrobial bacteriuria.

## Results

**Metabolomic profiling identifies multiple *Yersinia* HPI-associated metabolites.** To identify secreted metabolites associated with the *Yersinia* HPI, we used liquid chromatography-mass spectrometry (LC-MS) to compare culture supernatants from UTI89, a Ybt-producing model UPEC strain (28), and its isogenic mutant UTI89 $\Delta$ *ybtS* that cannot perform the first committed step of Ybt biosynthesis, salicylate synthesis (12, 23). As a control, the

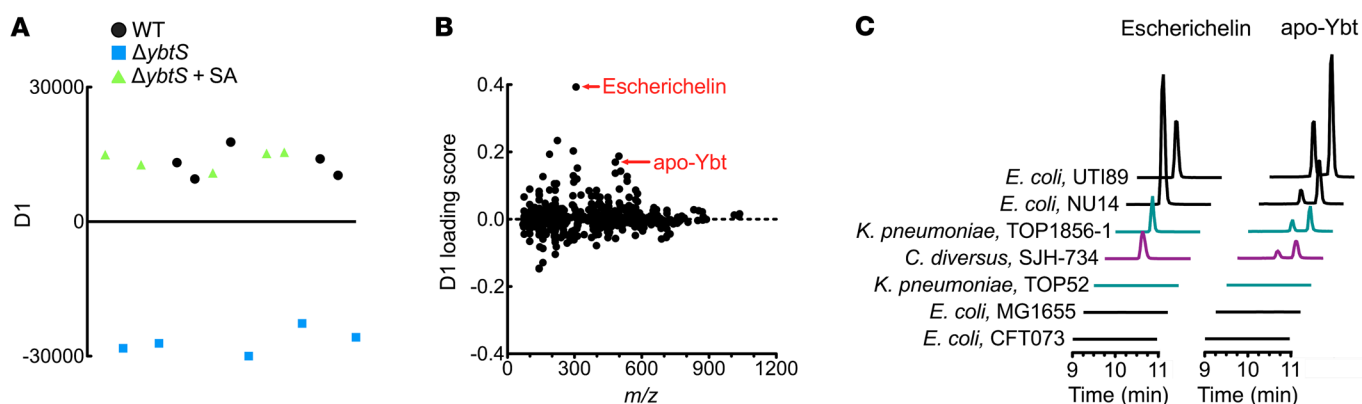
biosynthetic block in UTI89 $\Delta$ *ybtS* was experimentally overcome by chemical complementation with salicylate-supplemented medium. The metabolomic profiles from Ybt producers (WT and UTI89 $\Delta$ *ybtS* plus salicylate) and the Ybt nonproducer (UTI89 $\Delta$ *ybtS*) were compared using supervised principal component analysis-discriminant analysis (PCA-DA) (Figure 1A). Uniform separation of these 2 groups along the *y* axis of the PCA-DA plot confirmed that extracellular metabolites can differentiate Ybt-producing from nonproducing strains (Figure 1A).

To identify the specific extracellular metabolites that distinguish Ybt producers from nonproducers, we performed a loadings plot analysis (Figure 1B). In this plot, features with the highest D1 loading scores are most strongly associated with Ybt producers and are, therefore, candidates for follow-up analysis. Ybt in the metal-free form (apo) and bound to Al(III) were among the molecular features with the highest loading scores, as expected (Table 1) (29). We also observed dihydroaeruginic acid (Dha), a known metabolite of pyochelin-producing strains of *P. aeruginosa* (Table 1 and Supplemental Figure 1; supplemental material available online with this article; <https://doi.org/10.1172/JCI92464DS1>) (30, 31). Dha is the free form of an early phosphopantetheine-bound intermediate in both the pyochelin and Ybt biosynthetic pathways (20, 32). Unexpectedly, the molecular feature with the highest loading score was an ion with *m/z* 307, which is lower than apo-Ybt (*m/z* 482) (Figure 1B and Table 1). Like Ybt, the molecular feature at *m/z* 307, herein called escherichelin, was present in the WT and salicylate-complemented UTI89 $\Delta$ *ybtS* supernatants but entirely absent in  $\Delta$ *ybtS* supernatants. These results are consistent with the production of a previously unappreciated secreted metabolite by *E. coli* with an intact *Yersinia* HPI.

**Clinical Enterobacteriaceae isolates coproduce escherichelin and Ybt.** To determine whether escherichelin production is specific to UTI89 or is present in other bacteria encoding the *Yersinia* HPI, we measured its production in a series of clinical urinary isolates and model strains of the Enterobacteriaceae family (Figure 1C). The UPEC model strains UTI89 and NU14 produced both Ybt and escherichelin, whereas CFT073, a widely used UPEC model strain with known mutations in the *Yersinia* HPI, and the K12 strain MG1655 produced neither (33). *Klebsiella pneumoniae* and *Citrobacter diversus* isolates that produced Ybt also produced escherichelin (Figure 1C). These observations suggest that escherichelin production is pervasive among clinically significant Ybt-producing Enterobacteriaceae family members.

**Escherichelin production is associated with early Ybt biosynthetic enzymes.** The *Yersinia* HPI and other siderophore systems are regulated by Fur, an iron-responsive transcription factor, such that Ybt biosynthesis is downregulated in iron-rich conditions. To determine whether escherichelin is similarly regulated, we measured its secretion in a high-iron medium (complete M63 supplemented with 16.2 mg/l ferric chloride) (34). The high-iron medium abolished escherichelin production, suggesting that it is regulated by iron in a manner similar to that of Ybt (Figure 2A).

To better understand the relationship between escherichelin secretion and Ybt biosynthetic genes (Figure 2B), we screened a panel of *Yersinia* HPI gene deletion mutants for altered escherichelin production (Figure 2C). Mutants lacking the biosynthetic genes *ybtS*, *ybtE*, *irp1*, or *ybtU* do not produce Ybt (Supplemental



**Figure 1. Metabolomic analysis reveals escherichelin as an additional product of Ybt-producing *E. coli*.** (A) Two-group PCA-DA of LC-MS data demonstrate consistent variance in the secreted metabolomes of the Ybt producers, WT UTI89 and UTI89 $\Delta$ ybtS chemically complemented with salicylic acid (SA), and the nonproducer UTI89 $\Delta$ ybtS. Each point indicates the average of 3 technical replicates. Five biological replicates were analyzed from each experimental group. (B) The PCA-DA loadings plot, where each point is a molecular feature identified in the LC-MS analysis, reveals a molecular feature with  $m/z$  307 (escherichelin) that best distinguishes Ybt producers from nonproducers. (C) LC-MS/MS chromatograms of escherichelin (left) and apo-Ybt (right) from culture supernatants of Enterobacteriaceae isolates grown in iron-restricted medium. All of the strains that produced Ybt also produced escherichelin.

Figure 2) (23, 35–38). A mutant lacking the *Yersinia* HPI transcriptional regulator *ybtA* also produces no Ybt (38, 39). Deletion of *ybtT*, a putative editing enzyme, exhibits an intermediate phenotype with reduced Ybt levels (Supplemental Figure 2) (37, 38, 40). These isogenic mutants are not attenuated for growth in iron-restricted medium (38). Among Ybt-null strains, extracellular escherichelin was undetectable in the culture supernatants of UTI89 $\Delta$ ybtS,  $\Delta$ ybtE, and  $\Delta$ ybtA (the 2 early biosynthetic genes and transcriptional regulator, respectively). In contrast, escherichelin was detectable in supernatants from UTI89 $\Delta$ irp1 and  $\Delta$ ybtU, the late Ybt biosynthetic pathway deletion mutants (Figure 2C). Intracellular escherichelin levels followed a trend nearly identical to that of the extracellular levels between strains (Supplemental Figure 3). Overall, these patterns favor biosynthesis, rather than cellular export, as the major determinant of escherichelin secretion.

The requirement of early Ybt biosynthetic genes for escherichelin production is consistent with it being a *Yersinia* HPI product. The observation that late biosynthetic mutants (UTI89 $\Delta$ irp1 and  $\Delta$ ybtU) produce escherichelin, but not Ybt, further suggests that escherichelin is a de novo metabolite and not a Ybt breakdown product. The diminished, but not absent, escherichelin levels in late biosynthesis mutants (UTI89 $\Delta$ irp1 and  $\Delta$ ybtU, Figure 2C) mirror the diminished *irp2* expression observed with analogous *Y. pestis* mutants (36, 37). Although the precise reason for this altered gene expression in the mutants remains unclear, it has been proposed that feedback regulation from a late biosynthetic gene product is reduced, leading to the downregulation. The feedback regulation is believed to be mediated by YbtA, a member of the AraC transcription factor family whose members typically respond to allosteric activation by metabolites.

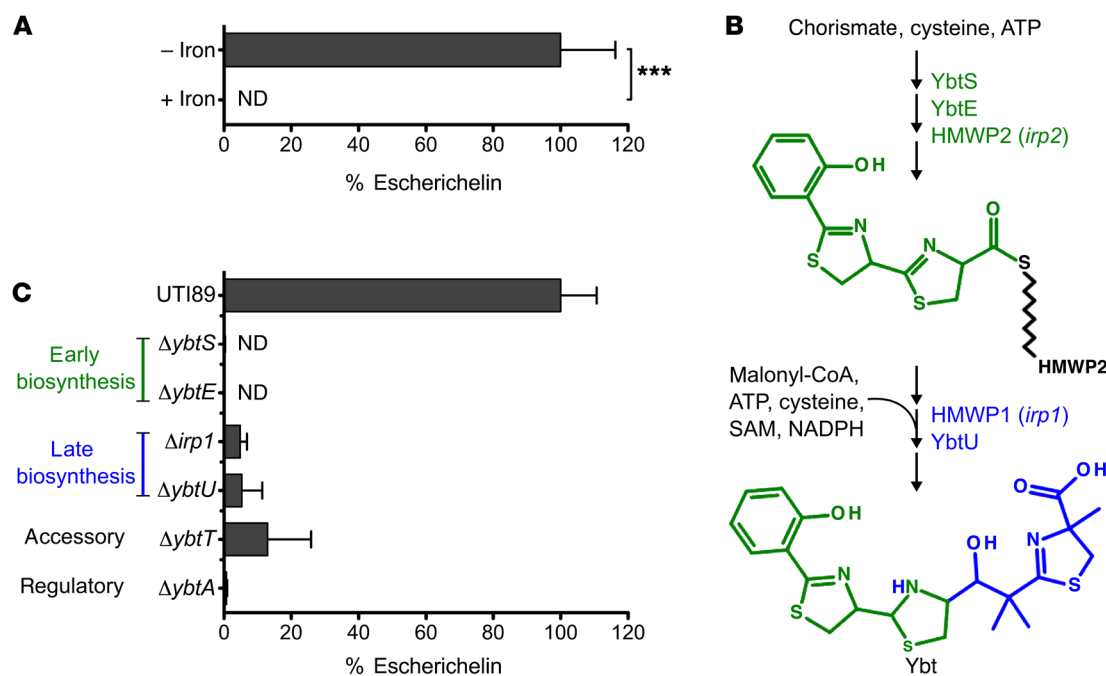
*Escherichelin is HPTT-COOH.* Multiple structural analyses were used to determine the chemical structure of escherichelin. These analyses were guided in part by previous in vitro investigations of Ybt biosynthesis that noted a product with a positive-ion  $m/z$  of 307 that was identified as 2-(2-hydroxyphenyl)-4,5-dihydro-[2,4] bithiazolyl-4-carboxylic acid (abbreviated as HPTT-COOH from hydroxyphenyl-thiazolyl-thiazolyl-carboxylic acid) (24, 41–44). HPTT-COOH is a truncated version of Ybt that arises when a biosynthetic intermediate (Figure 2B, top structure) derived from salicylate and 2 cysteines is released from the first NRPS protein, HMWP2 (*irp2*), and spontaneously oxidized. High-resolution MS of escherichelin yielded an exact mass of 307.0216 for the  $[M+H]^+$  ion, supporting an empiric formula of  $C_{13}H_{10}N_2O_3S_2$  (calculated mass: 307.0206 for  $C_{13}H_{11}N_2O_3S_2^+$ ). To determine whether escherichelin is synthesized from salicylate, we chemically complemented UTI89 $\Delta$ ybtS with  $^{13}C_6$ -salicylate and analyzed the supernatant by LC-MS. Upon  $^{13}C_6$ -salicylate complementation, escherichelin at  $m/z$  307 disappeared, and a new molecular feature at  $m/z$  313

**Table 1. Molecular features with the highest loading scores are candidate small molecules associated with Ybt biosynthesis**

Rank	Loading score <sup>A</sup>	$m/z$	Retention time (min)	Identification	Ref.
1	0.393	307.0	15.8	escherichelin	
2	0.234	224.1	12.8	Dha	30–32
3	0.203	295.0	17.1	apo-Ybt peak 2 <sup>B</sup>	14
4	0.193	188.1	4.9		
5	0.187	498.1	10.8		
6	0.169	482.1	17.1	apo-Ybt peak 2	14, 19
7	0.152	313.0	7.3		
8	0.143	506.1	11.6	Al(III)-Ybt	29
9	0.137	482.1	16.3	apo-Ybt peak 1	14, 19
10	0.129	295.1	16.4	apo-Ybt peak 1 <sup>B</sup>	14

<sup>A</sup>The loading scores come from the loading plot of the positive-ion LC-MS data (Figure 1B).

<sup>B</sup>Source-decay fragment of apo-Ybt.



**Figure 2. Escherichelin production is iron repressible and dependent on early Ybt biosynthetic enzymes.** Escherichelin levels in culture supernatants were quantified by LC-MS/MS relative to a  $^{13}\text{C}$ -IS. **(A)** UT189 grown in complete M63 medium with or without 16.2 mg/l  $\text{FeCl}_3$  (34). Data represent the mean of 3 replicates, with SD plotted.  $***P < 0.001$ , by unpaired  $t$  test. **(B)** Schematic of the Ybt biosynthetic pathway. Black zig-zag line designates a phosphopantetheine group covalently attached to the biosynthetic enzyme. Green designates chemical moieties built by early biosynthetic enzymes (YbtS, YbtE, and HMWP2), and blue designates chemical moieties built by late biosynthetic enzymes (HMWP1 and YbtU). **(C)** Isogenic mutants of UT189 in select genes of the *Yersinia* HPI grown in iron-restricted medium. Data represent the mean of 8 replicates, with SD plotted.

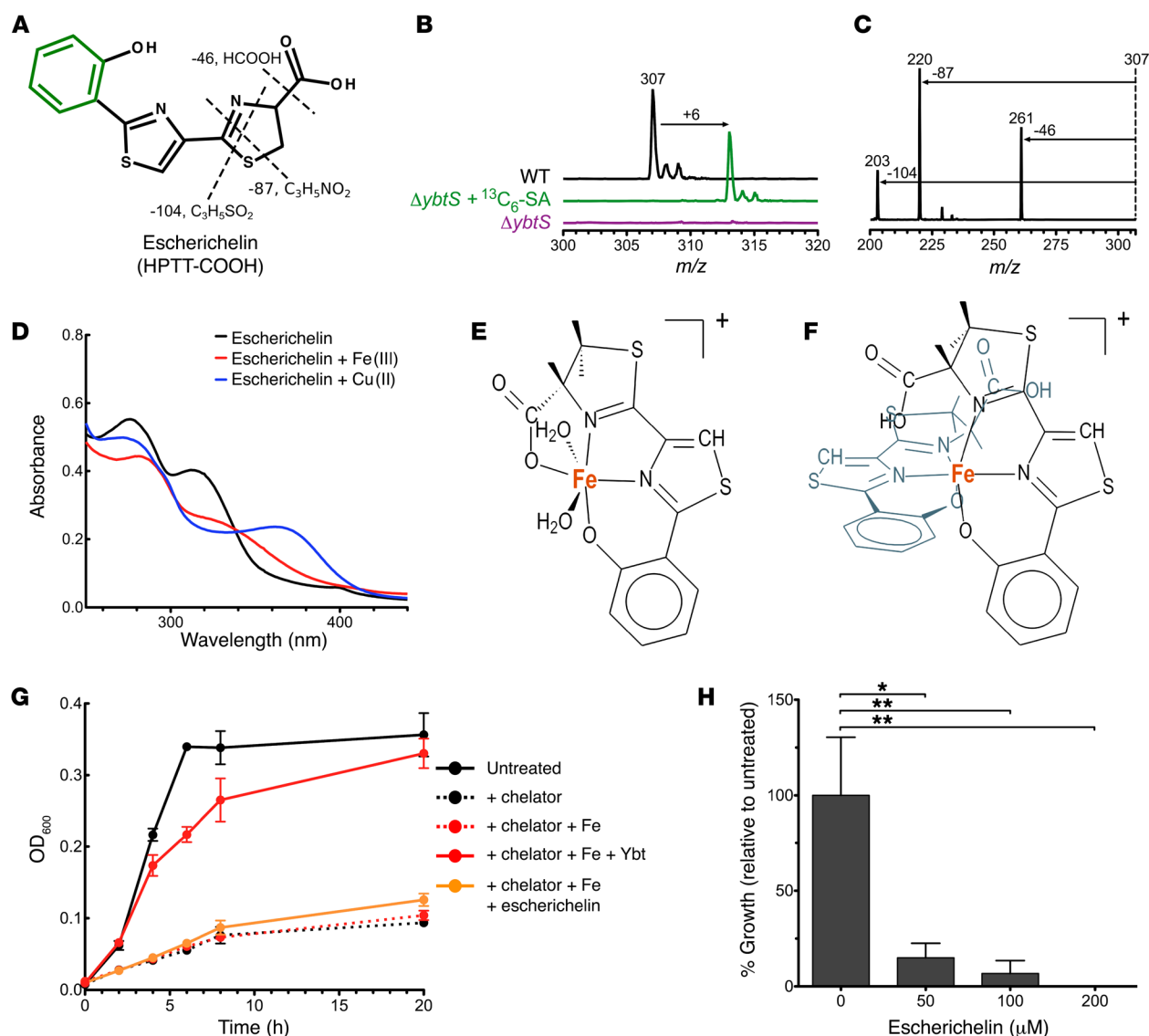
appeared at the same retention time, consistent with incorporation of the six  $^{13}\text{C}$  atoms (Figure 3, A and B). The empiric formula and heavy isotope labeling are consistent with a biosynthetic product derived from salicylate and 2 cysteines.

Information about the connectivity of escherichelin's atoms was obtained using tandem MS (MS/MS) and  $^1\text{H}$ -NMR. MS/MS of the escherichelin  $[\text{M}+\text{H}]^+$  ion yielded unique fragments arising from neutral losses of 46, 78, and 104 Da (Figure 3C). The 46-Da neutral loss is consistent with an equivalent loss of  $\text{HCOOH}$  (formic acid or sequential loss of  $\text{H}_2\text{O}$  and  $\text{CO}$ ) and supports the presence of a carboxylic acid. The atypical neutral losses of 78 and 104 Da are consistent with equivalent cleavages through the thiazoline ring to lose neutral fragments containing the carboxylic acid and either nitrogen  $[\text{M}-78]^+$  or sulfur  $[\text{M}-104]^+$  (Figure 3A). To further investigate the structure, we performed one-dimensional  $^1\text{H}$  and two-dimensional  $^1\text{H}$  correlation spectroscopy (COSY) NMR experiments. The chemical shifts and assignments were as follows:  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ), 8.15 (s, 1H,  $-\text{CH}=\text{}$ ); 6.97, 7.1, 7.4, and 7.63 (s, 1H, aromatic), 5.05 (m,  $\sim 1\text{H}$ ,  $\text{CH}-\text{COOH}$ ), and 3.2 (dd, 2H,  $-\text{CH}_2$ ). The 5.05 multiplet was partially obscured by a solvent impurity, preventing an accurate signal assignment. The chemical shifts at 6.97, 7.1, 7.4, and 7.63 ppm that each integrate to 1 proton and correspond to aromatic protons further support the presence of a salicylate-derived aromatic ring. A singlet at 8.15 ppm is consistent with the vinylic proton of the thiazole, and coupled resonances at 3.2 and 5.05 ppm are consistent with the presence of the methylene protons and the C-H proton of the thiazoline ring,

respectively (Supplemental Figure 4). Together, the molecular formula from high-resolution MS, the MS/MS fingerprint, and the  $^1\text{H}$ -NMR chemical shifts support HPTT-COOH as the escherichelin structure. Escherichelin corresponds to an oxidized form of a Ybt biosynthetic intermediate and is structurally similar to the *P. aeruginosa* siderophore pyochelin. Escherichelin and pyochelin differ in the oxidation state of their first heterocyclic ring, and pyochelin is also further modified by the reduction of the second thiazoline to a thiazole and by *N*-methylation (45).

**Escherichelin binds Fe(III).** Given that escherichelin is produced by a siderophore biosynthetic pathway, regulated by Fur, and retains some of the functional groups required for metal binding by Ybt and pyochelin, we assessed the ability of purified escherichelin to form an Fe(III) complex in aqueous solution with UV-visible spectroscopy. As reported in previous studies, Fe(III) addition caused notable shifts in the UV-visible absorption spectra of Ybt and pyochelin, consistent with the formation of Fe(III) complexes (19, 46, 47). The escherichelin UV-visible absorption spectrum was similarly shifted following exposure to equimolar Fe(III) (Figure 3D). Cu(II) addition caused a qualitatively distinct shift from Fe(III) (Figure 3D). These results demonstrate that escherichelin, like Ybt and pyochelin, is a bacterial metallophore that forms Fe(III) and Cu(II) complexes.

To determine whether escherichelin binds metal ions in a manner similar to Ybt and pyochelin, we conducted quantum-based calculations using density functional theory (DFT). These calculations predict stable 1:1 and 1:2 complexes of Fe(III)-escherichelin in a hexacoordinate octahedral geometry (Figure 3, E and F). In the 1:1 complex, Fe(III) is bound by the salicylate oxygen, the 2

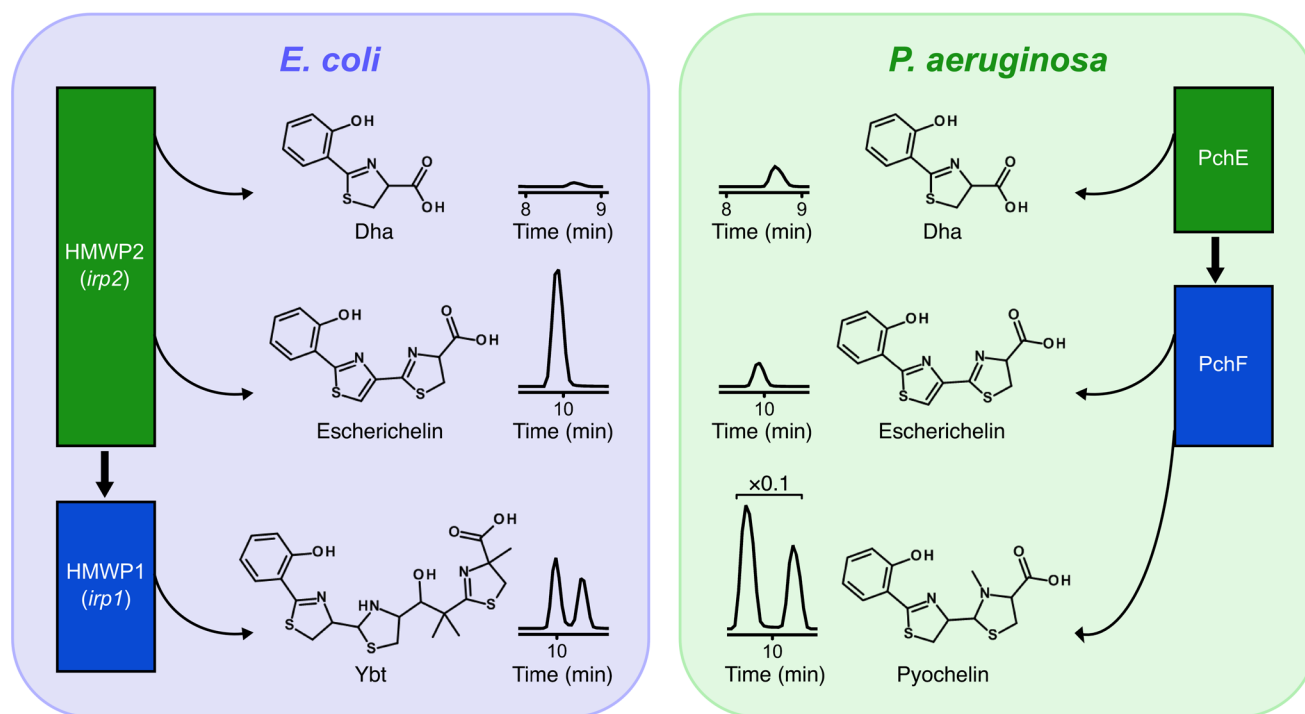


**Figure 3. Identification of escherichelin as HPTT-COOH, a metallophore inhibitor of *Pseudomonas* iron uptake.** (A) Multiple chemical characterizations identify escherichelin as HPTT-COOH, a compound shown to inhibit pyochelin-mediated iron uptake in *Pseudomonas*. The positions of  $^{13}\text{C}$  salicylate incorporation are indicated in green, while MS/MS neutral losses are indicated as dashed lines. (B) Mass spectra at the retention time for escherichelin from WT UT189, UT189 $\Delta$ ybtS, and  $^{13}\text{C}_6$ -SA-supplemented UT189 $\Delta$ ybtS culture supernatants revealed that 6 carbons in escherichelin are derived from salicylate. (C) MS/MS product ion spectrum of escherichelin. The 46-Da neutral loss indicates the presence of a carboxylic acid group, and the 87- and 104-Da losses correspond to fragmentation within the thiazoline ring. (D) UV-visible absorption spectra of purified escherichelin incubated with equimolar  $\text{FeCl}_3$  or  $\text{CuSO}_4$ . (E and F) DFT simulations of escherichelin bound to Fe(III) in a 1:1 (E) and 2:1 (F) complex. The simulations predict stable Fe(III)-escherichelin complexes with hexacoordinate octahedral geometry. (G) UT189 growth curve in the presence or absence of a ferric ion chelator (0.5 mM EDDHA) and/or ferric ion supplementation (1  $\mu\text{M}$   $\text{FeCl}_3$ ). Growth was monitored by OD at 600 nm ( $\text{OD}_{600}$ ). Data represent the mean of 3, with SD plotted. (H) PW5011 (*pvdA*-E02::ISlacZ/hah), a *P. aeruginosa* strain MPA01 transposon mutant (55, 56) expressing pyochelin as its sole siderophore, was grown in succinate medium for 20 hours in the presence of increasing escherichelin concentrations relative to vehicle control. Bacterial growth was quantified by enumerating CFU/ml and is expressed as a percentage of the vehicle control. Data represent the mean of 3, with the SEM plotted. \* $P < 0.01$  and \*\* $P < 0.001$ , by 1-way ANOVA with Dunnett's multiple comparisons test.

heterocyclic nitrogens, and the carboxylic acid group in the equatorial positions and by 2 water molecules in the axial positions. In the 1:2 complex, the carboxylic acids of the 2 escherichelin molecules and water do not participate as ligands. Instead, the nitrogen atoms and phenolic oxygen atoms of 2 the escherichelins occupy all of the liganding positions. Pyochelin binds Fe(III) in similar 1:1 and 1:2 complexes with the same ligands (48, 49). As with Ybt, escherichelin is also predicted to form a stable Cu(II) complex (Supplemental Figure 5). Overall, the calculated metal

ion coordination in the Fe(III)- and Cu(II)-escherichelin complexes is similar to that observed for metal-Ybt complexes and Fe(III)-Pch (15, 19, 48, 50). These results support the idea that escherichelin is an Fe(III)-binding molecule.

*Escherichelin does not support iron-dependent E. coli growth.* To determine whether escherichelin, like Ybt, can promote iron-dependent growth in *E. coli* possessing *Yersinia* HPI-encoded transport genes, we measured UT189 growth in a previously described siderophore-dependent growth assay (Figure 3G) (16, 19). Addition



**Figure 4. Escherichelin release is suppressed in *P. aeruginosa* relative to *E. coli*.** LC-MS/MS chromatograms of extracellular metabolites released from Ybt and pyochelin biosynthetic enzymes in the *E. coli* strain UTI89 (left) and the *P. aeruginosa* strain PAO1 (right) cultured in iron-deficient media are displayed in the center. Chromatograms are identically scaled, except for *P. aeruginosa* pyochelin, which is at  $\times 0.1$  magnification. The accompanying illustrations depict the sequential release of Dha, escherichelin, pyochelin, and apo-Ybt from multidomain NRPS/PKS enzymes associated with early (green rectangles) and late (blue rectangles) biosynthesis of Ybt (left) and pyochelin (right). Thick black arrows designate the biosynthetic steps at which metabolite intermediates transfer from early to late proteins.

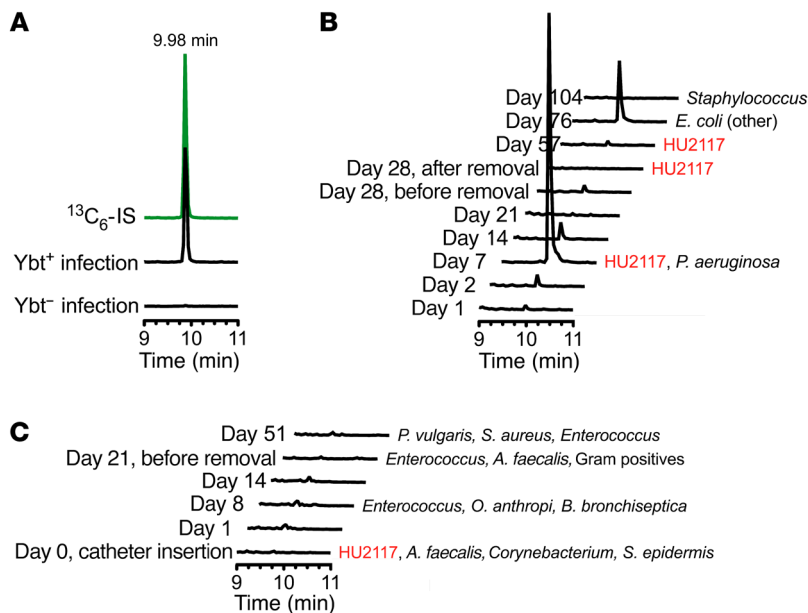
of the iron chelator EDDHA to the test medium causes substantial growth restriction. As previously observed, addition of Ybt that has been preincubated with equimolar Fe(III) restores growth to a density comparable to that of the unchelated medium. In contrast, addition of escherichelin that has been preincubated with equimolar Fe(III) resulted in no significant growth enhancement (Figure 3G). Although both escherichelin and Ybt bind Fe(III), these results show a nonequivalent ability to support iron-limited growth. This discrepancy may arise from differences in Fe(III) affinity or from differential recognition of the complexes by bacterial transport proteins.

***P. aeruginosa* growth inhibition by escherichelin.** The deficient siderophoric activity of escherichelin raises the possibility that this molecule performs an alternative function. In prior work by Mislin et al., escherichelin (previously designated HPTT-COOH) was synthesized as a pyochelin analog (51) and found to act as a competitive inhibitor of pyochelin-mediated iron import through the TonB-dependent outer membrane transporter FptA in *P. aeruginosa* (25, 51, 52). Escherichelin did not mediate iron uptake and inhibited Fe(III)-pyochelin transport through FptA with a  $K_i$  of 27 nM. In silico docking simulations could rationalize these results through occupancy of the Fe(III)-pyochelin binding site of FptA by a 1:2 Fe(III)-escherichelin complex [Fe(III):escherichelin] (25, 48). Given that escherichelin can bind Fe(III) in solution but does not possess canonical siderophoric activity, we hypothesized that it may be produced by UPEC to inhibit pyochelin uptake by competing *P. aeruginosa* strains in the urinary tract.

To determine whether escherichelin can inhibit ferric pyochelin uptake, we examined the effect of escherichelin on growth of the *P. aeruginosa* strain PAO1. Although pyochelin enhances *P. aeruginosa* growth during infections (53, 54), it is functionally redundant with pyoverdinin, a second siderophore, during in vitro growth in standard culture media. Escherichelin addition to a defined, iron-restricted minimal medium caused a dose-dependent reduction in growth of a pyoverdinin-deficient *P. aeruginosa* mutant strain, PW5011 (Figure 3H) (55–57). These results are consistent with the escherichelin-mediated inhibition of iron uptake in *P. aeruginosa* noted by Mislin et al. (25).

***P. aeruginosa* minimizes escherichelin production.** Production of a *P. aeruginosa* iron uptake inhibitor from the enzyme-bound precursor hydroxyphenyl-thiazolinyl-thiazolinyl-S-ppant (Figure 2B, top structure) presents a potential paradox, because this enzyme-bound species is also a biosynthetic intermediate of pyochelin (20). Escherichelin release from this intermediate in *P. aeruginosa* would be expected to autoantagonize pyochelin-mediated iron uptake. To determine whether *P. aeruginosa* minimizes escherichelin release relative to *E. coli*, we compared escherichelin production between UTI89 and PAO1 in low-iron medium. Despite comparable growth of both strains and substantial pyochelin production by PAO1, extracellular and intracellular levels of escherichelin were markedly lower in PAO1 than in UTI89 (Figure 4 and Supplemental Figures 6 and 7). This may reflect selective pressure on the *P. aeruginosa* pyochelin biosynthetic pathway to avoid





**Figure 5. Escherichelin is produced during clinical *E. coli* UTI and experimental human colonization with the asymptomatic bacteriuria strain *E. coli* HU2117.** (A) Representative LC-MS/MS chromatogram for escherichelin in urine from a patient with an uncomplicated UTI caused by a Ybt-producing UPEC strain (results from 18 urine samples are shown in Table 1). Escherichelin coeluted with the  $^{13}\text{C}_6$ -IS (green). Escherichelin was undetectable in urine from a patient with a UTI caused by a Ybt-nonproducing UPEC strain. (B and C) LC-MS/MS chromatograms for escherichelin in urine from subjects who were successfully (B) or unsuccessfully (C) colonized with *E. coli* HU2117. The days after insertion of the HU2117-coated study catheter are indicated on the left. Where microbiological culture data were available, the results are listed to the right of the chromatograms. The colonized subject (B, subject 9) was colonized with HU2117 for 57 days and then developed a febrile UTI caused by another strain of *E. coli* on day 76, after colonization had ended. On day 7, *P. aeruginosa* was also present in this subject's urine. Escherichelin was detectable throughout the colonization period and during the symptomatic *E. coli* UTI. The noncolonized subject (C) did not show a presence of HU2117 after day 0, when the study catheter was inserted, and escherichelin was not detected in the longitudinal samples. *P. vulgaris*, *Proteus vulgaris*; *A. faecalis*, *Alcaligenes faecalis*; *D. anthropi*, *Ochrobactrum anthropi*; *B. bronchiseptica*, *Bordetella bronchiseptica*; *S. epidermis*, *Staphylococcus epidermidis*.

inhibiting pyochelin-mediated iron uptake, as well as an opposing selective pressure on *E. coli* to suppress competing *Pseudomonas* strains in polymicrobial settings.

*Escherichelin* is produced during human *E. coli* cystitis. Enterobacterial bacteriuria may be a frequent occurrence in the human urinary tract and is frequently accompanied by other flora (58–60). When native enterobacteria in the microbiome are suppressed or eliminated by antibiotic treatment, *P. aeruginosa* UTIs become more frequent (6). Therefore, we hypothesize that escherichelin release during enterobacterial bacteriuria prevents *P. aeruginosa* UTI. To determine whether Enterobacteriaceae produce escherichelin in the human urinary tract, we used LC-MS/MS to detect escherichelin in urine from 18 patients with uncomplicated cystitis caused by Ybt-producing *E. coli*. Escherichelin was detectable in 8 of the 18 samples (Figure 5A and Table 2). In four escherichelin-positive specimens, Ybt was undetectable. As expected, escherichelin was undetectable in patients with cystitis caused by Gram-positive species (*Staphylococcus saprophyticus* or *Enterococcus* sp.) or Ybt-null *E. coli*. These results demonstrate that *E. coli* can produce escherichelin in the human bladder.

*Escherichelin* is produced during experimental bladder colonization with an *E. coli* 83972 strain. The concept that one bacterial strain or species can prevent infection by another — termed bacterial interference — has been of longstanding interest in infectious diseases (61–63). In this context, an avirulent strain or species that outcompetes more pathogenic strains may be useful as a probiotic to prevent infections. *E. coli* 83972, which asymptotically colonized a young Swedish girl for over 2 years (64), has been used in multiple UTI prevention studies (26, 27, 63, 65–70). Among these studies are anecdotal reports of protection against symptomatic *P. aeruginosa* UTIs, including 1 patient with asymptomatic cocolonization of 83972 and *P. aeruginosa* that progressed to UTI following loss of 83972 (26, 66). *E. coli* 83972 possesses the *Yersinia* HPI (71, 72) and was observed to produce escherichelin in iron-restricted medium (data not shown).

To determine whether 83972 secretes escherichelin in the human urinary tract, we analyzed urine specimens following experimental colonization of catheter-dependent patients with an 83972-derived *E. coli* strain lacking the virulence-associated *papG* adhesin (*E. coli* HU2117) (27). Bladder colonization was established by inserting urinary catheters that had been precoated with *E. coli* HU2117. Longitudinal urine specimens were available from 1 subject (104 days for subject 9) who became colonized with HU2117 and another subject who was unsuccessfully colonized (51 days for subject 10). Both subjects showed evidence of polymicrobial urinary tract colonization during the collection periods. In subject 9, escherichelin was detected throughout the 57 days of documented HU2117 colonization, reaching a maximum on day 7 after catheterization (Figure 5B). *P. aeruginosa* was detected by culture in the urine on

day 7 but was unassociated with symptoms and became undetectable in subsequent specimens (27). On day 76, HU2117 colonization had ended, and the subject developed a febrile UTI caused by another *E. coli* strain, which was accompanied by detectable escherichelin. Subject 10 was not colonized with HU2117 or any other *E. coli* strains after day 0, and escherichelin was undetectable in all specimens (Figure 5C). These results are consistent with escherichelin production by HU2117 in conditions in which *P. aeruginosa* is encountered, which may contribute to the ability of this strain to suppress symptomatic *P. aeruginosa* UTI.

## Discussion

In this study, we used isogenic mutants in the *Yersinia* HPI and untargeted LC-MS-based metabolomics to identify a previously unappreciated biosynthetic product of bacteria carrying the *Yersinia* HPI. Escherichelin had previously been observed during the in vitro reconstitution of Ybt biosynthesis (24), but we now identify it as a physiologically relevant product of the Ybt biosynthetic pathway. The name escherichelin was chosen to recognize its identification as a natural product from pathogenic *E. coli* (*escheri*-) and its

**Table 2. Escherichelin and Ybt prevalence in urine samples from patients with uncomplicated UTI**

Patient no.	Causative species	Ybt-producing in culture	Escherichelin in urine sample	Ybt in urine sample
3	<i>S. saprophyticus</i>	-	-	-
5	<i>Enterococcus</i> spp.	-	-	-
2	<i>E. coli</i>	+	-	-
4	<i>E. coli</i>	+	-	-
8	<i>E. coli</i>	+	-	-
12	<i>E. coli</i>	-	-	-
14	<i>E. coli</i>	+	-	-
15	<i>E. coli</i>	+	+	+
17	<i>E. coli</i>	+	-	-
24	<i>E. coli</i>	-	-	-
33	<i>E. coli</i>	+	-	-
39	<i>E. coli</i>	+	-	-
41	<i>E. coli</i>	+	-	-
43	<i>E. coli</i>	+	-	-
51	<i>E. coli</i>	+	+	-
54	<i>E. coli</i>	ND	+	+
56	<i>E. coli</i>	+	+	-
66	<i>E. coli</i>	+	+	-
74	<i>E. coli</i>	+	+	-
80	<i>E. coli</i>	+	+	+
83	<i>E. coli</i>	+	-	-
95	<i>E. coli</i>	+	+	+

ND, strain did not grow in complete M63, so Ybt production in culture could not be determined.

metal chelating properties (*-chelin*). Escherichelin may have eluded discovery in the 20 years since the identification of Ybt because its reverse-phase chromatographic and UV-visible spectroscopic characteristics are nearly identical to those of Ybt, making them difficult to resolve. Although escherichelin lacks the siderophoric activity of Ybt, the fortuitous synthesis of escherichelin prior to the present discovery (51) and the demonstration of its ability to inhibit pyochelin-mediated iron uptake by *P. aeruginosa* (25) suggests an alternative functional rationale for escherichelin's secretion by Enterobacteriaceae: to inhibit iron uptake in a competing species. This niche exclusion function is consistent with clinical observations regarding *Pseudomonas* UTI, asymptomatic bacteriuria, and protective influences of a healthy human microbiome.

Other microbes have demonstrated an ability to interfere with pyochelin function. *Pseudomonas fluorescens* can synthesize an enantiomeric form of pyochelin (enantiopyochelin), which preserves the coordinating properties of pyochelin while resisting FptA-mediated uptake by pyochelin-producing competitor strains. The fungal commensal and opportunistic pathogen *C. albicans* suppresses pyochelin biosynthesis, interfering with *P. aeruginosa* virulence, but not colonization, in a murine model (73). A detailed study by Mislin et al. (25) revealed that escherichelin acts as a drug-like inhibitor of pyochelin-mediated iron uptake. Specifically, escherichelin competitively inhibited radioactive <sup>55</sup>Fe iron uptake through the pyochelin outer membrane transporter FptA, with a  $K_i$  of 27 nM. A molecular docking simulation sup-

ports an inhibitory mechanism based on occupancy of the pyochelin-binding site with a 1:2 Fe(III)-escherichelin complex. Here, we experimentally confirmed that escherichelin forms an Fe(III) complex and found by quantum calculations that this complex favors the molecular geometry used in the docking model of Mislin et al. (25). We also experimentally confirmed that escherichelin purified from *E. coli* inhibits *P. aeruginosa* pyochelin-dependent growth. Overall, the results are consistent with a noncanonical siderophore function for escherichelin as an effector of interspecies bacterial antagonism. To our knowledge, this is the first report of a microbial natural product that is a direct inhibitor of a competing species' siderophore transport system.

Multiple studies have implicated the pyochelin siderophore system in virulence (74-76), including genetic abrogation of pyochelin-mediated iron uptake that diminished *P. aeruginosa* virulence in murine infection models (53, 54). In this context, escherichelin-producing Enterobacteriaceae at colonized devices or mucosal surfaces may prevent *P. aeruginosa* from progressing to symptomatic infection by chemically inhibiting pyochelin-mediated iron uptake. This scenario may occur at epithelial surfaces during transient or persistent colonization of the urinary tract. The apparent ability of asymptomatic bacteriuria *E. coli* strain 83972 (or its derivative HU2117) to produce escherichelin during experimental human colonization is consistent with a model in which *E. coli* in the urinary tract secretes an antivirulence compound against *P. aeruginosa* (26, 27). Expression of the virulence-associated *Yersinia* HPI in colonizing Enterobacteriaceae such as *E. coli* strain 83972 may thus confer both risk (from Ybt production) and benefit (escherichelin production) to patients (77). Engineering an *E. coli* strain that colonizes the bladder and retains escherichelin, but not Ybt, production may be a desirable bacterial interference strategy in the future. Lack of a recognized asymptomatic bacteriuria mouse model and mouse-specific bacterial tissue tropisms in the urinary tract suggest that these future engineered strains may be best evaluated in human studies.

Whereas the Ybt and pyochelin biosynthetic machineries both generate the immediate enzyme-bound precursor to escherichelin (Figure 2B, top structure), the pyochelin machinery in *P. aeruginosa* releases less escherichelin than does the Ybt machinery in *E. coli*. This is consistent with the evolutionary hypothesis that the pyochelin biosynthetic pathway evolved to avoid auto-inhibition of Fe(III)-pyochelin uptake. The architectures of these pathways provide a potential explanation for this difference in escherichelin production. Specifically, the escherichelin precursor in the Ybt pathway forms at the terminus of the first NRPS protein (HMWP2), where it must be transferred in *trans* to the second NRPS/PKS protein (HMWP1). In the pyochelin pathway, this same intermediate is instead passed in *cis* to a subsequent domain on the same protein (PchF). In the Ybt biosynthetic pathway, this handoff between proteins appears to be particularly susceptible to spontaneous hydrolysis, thereby allowing it to become a branch point in which the intermediate can proceed to form either escherichelin or Ybt. In contrast, this immediate precursor to escherichelin may be less vulnerable to hydrolysis off the pyochelin biosynthetic machinery, because it does not occur at a handoff between proteins. During pyochelin biosynthesis, Dha is passed in *trans* from PchE to PchF, and it is Dha, rather than escherichelin, that accu-

mutates in *P. aeruginosa*. Intriguingly, Dha has been shown to possess antimicrobial activity against fungal and bacterial species (31). Applied generally, this suggests that the NRPS/PKS pathways may evolve to segregate modules onto separate proteins at steps in the pathway where a “leak” yields a product that benefits the organism. Whether the branch point in Ybt biosynthesis that yields escherichelin and the similar branch points in other NRPS/PKS pathways reflect uncontrolled “leaks” or regulated processes is unclear. Further biochemical and genetic characterization of these pathways may address this possibility.

Advances in MS and the application of powerful computational and statistical analyses to the data have led to the fruitful application of metabolomics to secondary metabolism (78). MS-based metabolomics is enabling the rapid discovery of secondary metabolites from newly sequenced bacterial strains as well as novel products from well-studied pathways (79). This approach moves beyond the functional screens that were historically used to discover siderophores and other natural products. With this unbiased approach, one is open to discovering any molecule that is chemically compatible with the instrumentation, regardless of the molecule’s function. Continued application of LC-MS-based metabolomics to other siderophore biosynthetic pathways will further our understanding of iron acquisition by pathogens.

## Methods

**Bacterial strains and culture conditions.** The well-characterized, cystitis-derived model UPEC strain UTI89 was used for these studies (28). The creation of the isogenic mutants in *ybtS*, *ybtE*, *irp1*, *ybtU*, *ybtT*, and *ybtA* was previously described (12, 38). UTI89  $\Delta irp1/entB::kan$  was generated in the same manner, using the lambda red recombinase method (80, 81). For growth under iron-restricted conditions, strains were first grown in Luria-Bertani (LB) broth (BD Biosciences) overnight at 37 °C under shaking conditions. They were then back-diluted 1:1,000 into M63 minimal medium supplemented with 10 mg/ml niacin and 0.2% (v/v) glycerol (Sigma-Aldrich; complete M63). For salicylate complementation experiments, 50  $\mu$ M salicylate or  $^{13}C_6$ -salicylate (Sigma-Aldrich) was also added. To achieve iron-replete conditions, 16.2 mg/l  $FeCl_3$  (Sigma-Aldrich) was added to the complete M63. For comparison of *E. coli* and *P. aeruginosa* metabolites, the model *P. aeruginosa* strain PAO1 was used (82). In these experiments, UTI89 and PAO1 were grown in complete M63 supplemented with 1% (w/v) casamino acids (BD Biosciences). For *E. coli* siderophore-dependent growth assays, the iron-limited and nutrient-rich yeast extract-casamino acids (YESCA) medium was used (1% casamino acids and 0.12% yeast extract; BD Biosciences). All cultures were grown at 37°C under shaking conditions for approximately 18 hours. Other model *E. coli* strains (MG1655 [ref. 83], CFT073 [ref. 84], NU14 [ref. 85], and 83972 [ref. 64]) as well as clinical isolates of UPEC, *K. pneumoniae*, and *C. diversus* were grown in the same manner.

**Metabolomic analysis.** For metabolomics experiments, 5 biological replicate cultures were cultivated from 5 distinct single colonies under iron-restricted conditions, as described above. Cultures were centrifuged at 7,142 g for 10 minutes to pellet the cells. The spent supernatants were collected and filtered through 0.22- $\mu$ M filters (EMD Millipore). A quality control sample was generated by combining an equal volume of each sample. The samples were analyzed in technical triplicates and in a randomized order on a Shimadzu Prominence

UFLC-coupled AB Sciex 4000 QTrap mass spectrometer with a Turbo V electrospray ionization (ESI) source. The LC separation was performed on an Ascentis Express phenyl-hexyl column (100  $\times$  2.1 mm, 2.7  $\mu$ M; Sigma-Aldrich) with a 32-minute linear gradient from 2% A (HPLC-grade water plus 0.1% [v/v] formic acid; Sigma-Aldrich) to 98% B (90% acetonitrile plus 0.1% [v/v] formic acid; EMD Millipore) at 0.35 ml/min. The MS was operated in positive ion-enhanced MS mode, scanning from 50 to 1,200 *m/z*. The quality control sample was injected first to precondition the column and every 10 samples thereafter to assess instrument stability.

MarkerView, version 1.2.0 (Sciex) was used for the generation of an aligned peak list and statistical analysis. The triplicate runs were averaged, and the data were Pareto scaled for supervised PCA-DA with 2 groups: (a) UTI89 and  $\Delta ybtS$  plus salicylate; and (b)  $\Delta ybtS$ . Candidate molecular features were identified on the PCA-DA loading plots and then verified by visual inspection of the LC-MS data.

**Metabolite detection and quantification in bacterial culture.** Escherichelin, Dha, pyochelin, and Ybt were detected in culture supernatants and cellular extracts from UTI89, PAO1, and clinical Enterobacteriaceae strains by a multiple reaction monitoring (MRM) method on the Shimadzu Prominence UFLC-coupled AB Sciex QTrap 4000 with MS/MS transitions for escherichelin (307.0/219.9), Dha (224.0/178.0), pyochelin (325.0/190.0), apo-Ybt (482.2/295.1), and Fe-Ybt (535.6/349.5) and a collision energy of 37 V. The above column, mobile phases, and flow rate were used with the following LC gradient: solvent B was held constant at 5% for 2 minutes, increased to 56% over 8 minutes, and then increased to 98% over 2 minutes, all with a flow rate of 0.4 ml/min.

Stable isotope dilution was used to quantify escherichelin levels. A  $^{13}C_6$ -escherichelin internal standard (IS) was generated by culturing UTI89  $\Delta ybtS$  in complete M63 minimal medium supplemented with 50  $\mu$ M  $^{13}C_6$ -salicylate. For quantification of extracellular escherichelin, strains were grown under iron-restricted conditions, as described above, and 1 ml of culture supernatant was spiked with 25  $\mu$ l  $^{13}C_6$ -escherichelin IS and 6  $\mu$ l  $FeCl_3$ . The samples were incubated at room temperature for 10 minutes, centrifuged at 15,996 g for 3 minutes, and extracted on C18 solid-phase extraction 96-well plates (United Chemical Technologies) with 80% methanol (Sigma-Aldrich). Samples were then analyzed by the escherichelin-specific MRM, and escherichelin levels were assessed by taking the peak area ratio of the native isotope escherichelin to the IS  $^{13}C_6$ -escherichelin. Viable CFU were measured to ensure that all strains grew to an equal final density.

Intracellular escherichelin was extracted from 50-ml cultures grown under iron-restricted conditions, as described above. The cultures were centrifuged at 7,412 g for 10 minutes to pellet the cells. The supernatant was discarded, and the cells were washed 3 times with 25 ml PBS (Sigma-Aldrich). The pellets were then resuspended in 1.2 ml ice-cold 80% methanol, briefly vortexed, and incubated on dry ice for 30 minutes. After thawing, the samples were centrifuged at 15,996 g for 10 minutes to pellet the cell debris. The supernatant was retained, and 800  $\mu$ l acetonitrile was added. The mixture was incubated on ice for 15 minutes and then centrifuged at 15,996 g for 2 minutes to remove any remaining debris.  $^{13}C_6$ -escherichelin (10  $\mu$ l) IS was added to 250  $\mu$ l of each sample, and escherichelin levels were quantified by LC-MRM, as described above.

**MS for escherichelin identification.** A Shimadzu UFLC-coupled Agilent 6550 Q-TOF mass spectrometer with a Dual AJS ESI ion source was used to determine the accurate mass, and therefore molecular for-

mula, of escherichelin in UTI89 supernatant that had been prepared as described above. LC separation was performed on an Ascentis Express phenyl-hexyl column (100 × 2.1 mm, 2.7 μM) with the same mobile phases as described above. The gradient was as follows: solvent B was held constant at 2% for 1 minute, then increased to 50% over 6 minutes, then increased to 98% over 2 minutes, and then held constant at 98% for 1 minute, all with a flow rate of 0.6 ml/min. The MS was operated in positive-ion mode, scanning a mass range from 75 to 1,100 Da. The calculated mass for HPTT-COOH is  $[M+H]^+$  (C<sub>13</sub>H<sub>11</sub>N<sub>2</sub>O<sub>3</sub>S<sub>2</sub><sup>+</sup>) = 307.0206, and 307.0216 was found.

MS/MS was performed on the Shimadzu Prominence UFLC-coupled AB Sciex QTrap 4000. Supernatant from UTI89 grown under iron-restricted conditions was prepared as described above for detection and quantification. Data were collected in the positive-ion, enhanced product ion (EPI) mode, with the parent ion *m/z* set at 307 and the collision energy set at 37 V.

**Escherichelin purification.** Escherichelin was purified from UTI89Δ*irp1*/entB::kan grown under iron-restricted conditions, as described above, with the addition of 20 μM 2,2'-dipyridyl (Sigma-Aldrich) and 0.1% casamino acids (w/v) to the complete M63. Culture supernatant was applied to a methanol-conditioned C18 silica column (Sigma-Aldrich). The column was washed with water and then eluted with 80% methanol. The eluate was dried down with a lyophilizer (Labconco), resuspended in 20% methanol, and further purified on a Bio-Rad BioLogic DuoFlow 10 system equipped with a QuadTec UV-Vis detector and a Kromasil Eternity-5-phenylhexyl column (Akzo-Nobel). The column was run at 0.7 ml/min with water plus 0.1% formic acid (solvent A) and acetonitrile plus 0.1% formic acid (solvent B) and the following gradient: solvent B held steady at 0% for 2.5 ml, then increased to 30% over 2 ml, then increased to 70% over 20 ml, and finally increased to 100% over 2 ml. Fractions containing escherichelin were identified via UV-Vis detection at 318 nm, pooled together, and dried down by lyophilization. Purification was confirmed by LC-MS.

**NMR.** Purified escherichelin was resuspended in CDCl<sub>3</sub>. One-dimensional <sup>1</sup>H and two-dimensional <sup>1</sup>H COSY spectra were collected at 20°C on a Varian Unity Inova-500 MHz instrument.

**UV-Vis spectroscopy.** Purified escherichelin was reconstituted to 1 mM in 50% ethanol (Sigma-Aldrich). For experiments to monitor metal binding, escherichelin was diluted to 100 μM in water and combined with an equimolar amount of FeCl<sub>3</sub> or CuSO<sub>4</sub> (Sigma-Aldrich). The samples were incubated at room temperature for 2 hours, and then a wavelength scan was collected from 200 to 800 nm on a UV-Vis spectrophotometer (Beckman Coulter; DU800). For each escherichelin-plus-metal scan, the instrument was blanked with a metal-only control. For escherichelin alone, the instrument was blanked with 5% ethanol.

**E. coli siderophore-dependent growth assay.** The assay was adapted from a previously published Fe(III)-Ybt-dependent growth assay, and apo-Ybt was purified as previously described (16, 19). UTI89 was grown overnight in LB broth at 37°C under shaking conditions and then washed 3 times by centrifugation and resuspension in a volume of PBS equal to the culture volume. The washed bacteria were inoculated 1:100 into YESCA or YESCA supplemented with 0.5 mM ethylenediamine-*N,N'*-bis(2-hydroxyphenylacetic acid) (EDDHA; Complete Green Company). Prior to their addition, apo-Ybt and escherichelin were preincubated for 2 hours with an equimolar amount of FeCl<sub>3</sub> to allow for complex formation. The Fe(III) complexes were added at

a final concentration of 1 μM, and the cultures were grown at 37°C under shaking conditions. Bacterial growth was monitored over time by measuring the OD at 600 nm.

**Escherichelin activity assay on P. aeruginosa.** PW5011 (*pvda*-E02::IS-lacZ/hah) (55, 56) was grown in LB broth overnight at 37°C under shaking conditions and then back-diluted into succinate medium (25) at 10<sup>8</sup> CFU/ml. The growth experiments were performed in a final volume of 100 μl in 96-well plates. Escherichelin, solubilized in ethanol, was added to the cultures at increasing concentrations. The final ethanol (vehicle) concentration in all cultures was 5% (v/v). The cultures were grown at 37°C under shaking conditions for 20 hours, at which point they were serially diluted in PBS and plated on LB agar for CFU quantification.

**Theoretical calculations.** The theoretical calculations to identify thermodynamically stable escherichelin-metal complexes were performed with the DFT method as previously described (15, 19). This DFT approach was previously used to model a structure of Fe(III)-Ybt that matched the published crystal structure (50). The simulated Fe(III)-escherichelin complexes agree with a previously described model of Fe(III)-escherichelin bound to FptA (25).

**Escherichelin detection in urine.** The clinical uncomplicated cystitis isolates were grown under iron-restricted conditions, and their supernatants were analyzed for Ybt by LC-MRM. Seventeen of the nineteen selected isolates were Ybt producers. The 2 Ybt nonproducers were used as controls. Two additional strains (*Staphylococcus saprophyticus* and *Enterococcus* sp.) were selected from the cohort to serve as non-*E. coli*, Ybt-negative controls. After determining whether the causative strain was a Ybt producer, the corresponding urine samples were thawed on ice and then centrifuged at 15,996 g for 3 minutes to remove any debris. To each 500 μl of urine, 12.5 μl <sup>13</sup>C<sub>6</sub>-escherichelin IS and 3 μl 0.5 M FeCl<sub>3</sub> were added. The samples were incubated at room temperature for 15 minutes and then centrifuged at 15,996 g for 5 minutes. The supernatant was then extracted on 96-well C18 solid-phase extraction plates with 80% methanol. Escherichelin was detected by LC-MRM using the method described above. Escherichelin was detected in the bacterial interference trial specimens using the same extraction and LC-MRM methods.

**Statistics.** GraphPad Prism 4 (GraphPad Software) was used to perform statistical analysis and generate graphs for this study. All *P* values were determined by unpaired, 2-tailed *t* test or 1-way ANOVA with Dunnett's multiple comparisons test. A *P* value of 0.05 or less was considered statistically significant.

**Study approval.** The urine and bacterial specimens from patients with uncomplicated cystitis were collected with the approval of the Human Subjects Review Committee of the University of Washington. All participants provided written informed consent for the sample collection and following analysis, prior to inclusion in the study. Study design, inclusion and exclusion criteria, and sample preparation were previously described (14, 86). β-Hemolytic *E. coli* isolates from urine cultures with 10<sup>5</sup> or more CFU/ml were selected from the cohort for analysis.

The HU2117 experimental human colonization study protocol was approved by the FDA (investigational new drug [IND] 14007), the IRB of Baylor College of Medicine, and the Houston VA Research and Development committee and registered as a clinical trial (ClinicalTrials.gov identifier: NCT00554996). All participants provided written informed consent prior to inclusion in the study. The study design, inclusion and exclusion criteria, and specimen preparation were previously reported (27).

## Author contributions

SIO and JPH conceived and designed the experiments. SIO, DEG, and DAD performed the experiments. AES and BWT collected human specimens. SIO, DEG, DAD, and JPH analyzed the data. SIO and JPH wrote the manuscript.

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